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**Gastrointestinal disease in children with autism spectrum disorders: Etiology or consequence?**

Krigsman A *et al.* GI disease in children with ASD

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**Abstract**

Chronic gastrointestinal (GI) symptoms and disorders are common in children with autism spectrum disorder and have been shown to be significantly correlated with the degree of behavioral and cognitive impairment. In this unique population, GI symptoms often arise very early in development, during infancy or toddlerhood, and may be misdiagnosed - or not diagnosed at all – due in part to the challenges associated with recognition of symptoms in a minimally or non-communicative child. Evidence demonstrating that the gut-brain-axis can communicate gut dysbiosis and systemic immune dysregulation in a bidirectional manner raises the question as to whether an untreated gastrointestinal disorder can directly impact neurodevelopment or, conversely, whether having a neurodevelopmental disorder predisposes a child to chronic GI issues. From the data presented in this mini review, we conclude that the preponderance of available evidence would suggest the former scenario is more strongly supported.

**Key Words:** Autism spectrum disorder; Gastrointestinal disease; Immune dysregulation; Neurodevelopment

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**Core Tip:** Children with autism spectrum disorder are at a significantly increased risk for chronic gastrointestinal issues from an early age. Clinicians and caregivers should be made aware of this association and provided with the tools necessary for recognition of gastrointestinal (GI) symptoms in these children. Because the first several years is a critical developmental window in the life of a child, and because of the demonstrated correlation between GI symptom severity and severity of autism spectrum disorder behaviors and cognition, it is plausible that a neurodevelopmental diagnosis may result from an untreated, persistent state of GI/immune dysregulation.

**INTRODUCTION**

Children with autism spectrum disorder (ASD) display gastrointestinal (GI) signs (*i.e.,* abnormalities noted by an observer) more frequently than their neurotypical counterparts[1]. These signs may present as constipation, diarrhea, abdominal pain, vomiting, food refusal, gastroesophageal reflux, malodorous macroscopic malabsorption, failure to thrive, or weight loss and although nearly all children experience gastrointestinal issues at some point during childhood, the reasons for this increased prevalence of GI signs in children with ASD has not been clearly established. This raises the question as to whether autism renders children more vulnerable to chronic GI issues, or whether chronic GI issues, especially in infancy and early childhood, increases the risk for developmental delay and/or ASD.

Verbally (or non-verbally) communicative children with ASD who can express what they are feeling are known to experience GI symptoms (the self-expressed feeling of an abnormality such as pain) at a greater frequency than their neurotypical counterparts, whereas children with ASD who have an underlying GI disorder, but who are unable to express what they are feeling, are known to manifest GI symptoms as extremes of behaviors, such as aggression, self-injury, and excessive self-repetitive behaviors[2]. It is this group of (treatable) children that is most at risk for not receiving a GI diagnosis and not receiving the proper clinical care.

Finally, core ASD features are more pronounced in GI symptomatic children with ASD, and the severity of GI signs and symptoms have been shown correlate with the severity of ASD features[3]. The constellation of GI sign/symptom presentations in ASD, as well as the GI-behavioral association, has been reviewed elsewhere[4] and will not be the focus of this review. It is worth noting here, however, that the frequent anecdotal reports of improvement (from modest to remarkable) in core ASD features following successful treatment of an underlying GI disorder provides compelling evidence that the two are interrelated.

In this mini review, the fundamental conundrum we wish to explore is: (1) Does chronic gastrointestinal dysfunction, especially at a very young age, predispose a child to developmental delay or disorder (etiology); or (2) Does having a developmental delay or disorder increases the risk of gastrointestinal dysfunction in children (consequence).

**IMPORTANCE TO THE MENTAL HEALTH PROFESSIONAL**

The relevance and importance of this information to the mental health professional cannot be overstated. Although it is clear that all core ASD features or their extremes cannot be explained solely by an underlying GI pathology, the documented relationship between these two demands vigilance on the part of those who interact with and care for the child with ASD. If a caregiver or medical professional suspects and/or is made aware that GI signs/symptoms exist, then appropriate pediatric gastroenterology referral is warranted. The logical consequence of acknowledging this relationship between GI signs/symptoms and ASD behaviors (documented[5], though not yet formally addressed in most of the medical literature) is that it will allow for accurate identification and treatment of GI pathology to commence and, when treatment is successful, will ultimately decrease the unfortunate dependence on often unnecessary behavior modifying medications and their associated adverse effect profiles. Most importantly, this strategy directly targets the underlying pathology as opposed to simply modifying the resultant disease related behaviors.

**HOW DOES gastrointestinal DYSFUNCTION INFLUENCE BEHAVIOR IN autism spectrum disorder?**

The vast autism-GI literature provides an additional layer of fundamental mechanistic questions regarding the notion that central nervous system (CNS) pathology alone is responsible for ASD behaviors. If GI signs/symptoms correlate with the degree of ASD features in children with ASD, and if they are in some cases the sole presenting indication of an underlying cognitive/behavioral disorder, what might the mechanism be? Although it is tempting to attribute behaviors as directly resulting from unattended pain (an “acting out” of sorts), abundant anecdotal experience with communicative children with ASD reveals that they experience intense compulsions, thoughts, and feelings during active GI symptoms, even in the absence of pain. This question of an underlying mechanism to explain how GI dysfunction can impact ASD behaviors has led to speculation that a “gut-brain-axis”, in which the products of one or more pathological GI processes contributes to CNS dysfunction, is the nexus.

The obligatory first step then in answering this pathophysiologic question lies in the clinical approach taken by the gastroenterologist when confronted with these signs/symptoms in the child with ASD. The standard of care for these children is to address their GI signs/symptoms in identical fashion as one would the child who is neurotypical, while at the same time being mindful of the unique behaviors that may be the only sign of underlying GI pathology. GI evaluation is therefore sign/symptom driven, and when standard non-invasive diagnostic tests (blood, stool, urine, imaging studies) are unrevealing, empiric finite trials of medication (*e.g.,* H2 blockers, laxatives, *etc.*) may be warranted. If unsuccessful, the standard of care at that point is usually endoscopy with biopsy.

The most important clues as to the mechanism of the unidirectional GI-CNS linkage (beyond the simple behavioral response to pain) would be expected to lie within the GI mucosa, and in our experience treating > 1500 GI-symptomatic children with ASD, the most common pathologic finding at endoscopy in such cases is non-specific inflammation. Therefore, the putative mechanism that will be described in the following sections differs from existing mechanistic hypotheses that have been put forth thus far in the literature in that it is based solely on human GI mucosal pathology, identified specifically (and often uniquely) in children with ASD, that has been published over the past twenty years. A mechanistic theory will then be proposed that incorporates these findings into the published systemic and CNS pathology of ASD.

**GASTROINTESTINAL INFLAMMATION IN autism spectrum disorder**

Kanner[6]’s seminal paper that describes 11 children with autism mentions the finding of GI disturbances in many of them, but it was not until more than 50 years later, in 1998, that the first histologic evaluation and description of GI inflammation of GI symptomatic children with ASD was published. The ongoing and evolving story of ASD related GI inflammation might appear to be better suited for a gastroenterology journal, however since GI disease is being evaluated in the context of ASD, journals focused on autism or psychiatry (ASD is diagnosed as a psychiatric illness, after all) are equally well suited. Regardless of the venue, the story itself is illuminating in so many respects that academic fidelity dictates that if we are to provide meaningful context, any discussion of this topic must necessarily include the chronologic appearances in the literature of evidence of gastrointestinal inflammation.

In 1998, led by world renowned pediatric gastroenterologist John Walker-Smith, the inflammatory bowel disease (IBD) study group at the Royal Free Hospital in London published a case report in the lancet about 12 children with regressive pervasive developmental disorder, characteristic of autism, who underwent diagnostic endoscopy for evaluation of chronic GI symptoms including diarrhea, abdominal pain, abdominal distention, and food intolerance[7]. Chronic colitis (the excessive presence of lymphocytes - primarily plasma cells - in the lamina propria) was found in 11 of the12 children, and active colitis (neutrophilic infiltration of the epithelium and lamina propria) was found in 5. Intraepithelial lymphocytosis of the cecum was observed in 1 patient. Importantly, and unlike the “classic” inflammatory bowel diseases Crohn’s disease (CD) and ulcerative colitis (UC), the chronic inflammatory changes in these children were present in areas of the colon that appeared endoscopically normal. The publication of this case series created a media frenzy at the time because of questions it raised regarding pediatric vaccination safety, ultimately prompting a “retraction of an interpretation” six years later in 2004[8]. Full retraction of the paper by the lancet occurred in 2010 following a four-year investigation of the paper’s findings by England’s General Medical Council (GMC), a body that concluded patients were not in fact consecutively referred and that the Institutional Review Board authorization was not appropriately obtained. But, as will be described next, by the time the GMC findings had been rendered, the histologic findings had already been replicated and expanded upon by these and other investigators, at other institutions, and using additional methodologies.

The following year, in 1999, Horvath *et al*[9] published a clinical report describing the histologic (but not endoscopic) findings in 36 children with ASD and GI symptoms of diarrhea, flatulence, nighttime awakening, unexplained irritability, and abdominal distention. In that study it was found that 25 (69.4%) of the children had histologic gastroesophageal reflux, almost all of whom had unexplained irritability as a key symptom. Additionally, Paneth cell hyperplasia, a measure of increased crypt mitotic activity secondary to inflammatory mucosal damage, was significantly increased. The authors do not discuss therapeutic interventions.

Before proceeding further through this brief review of the ASD-GI inflammation literature, it is important to note that not all “inflammation” is equal. For example, the cellular infiltrate of acid related disease is composed of different cellular lineages and intensities than, for example, CD. Even within the classification of classic IBD (CD and UC), the types of inflammatory cells differ between the differing IBD diagnoses. Using tissue immunostaining, Furlano *et al*[10] demonstrated not only a significant non-specific lymphocytic inflammatory plasma cell, CD3, and CD8 T-cell elevation in the colon of GI symptomatic children with ASD compared to healthy controls, but that there was also an excess γδ-T-cell presence that distinguishes autistic colitis from classic IBD.

Attention then turned to the upper gastrointestinal tract where, in 2002, the use of immunohistochemical staining again showed the non-specific increased CD3 and CD8 T-cell inflammatory infiltrate in the mucosa with the additional finding of a thickened basement membrane, a feature often found in the intestine of patients with systemic autoimmune disease such as diabetes, rheumatoid arthritis, and gluten sensitive enteropathy (celiac disease)[11]. However, what distinguished the duodenal inflammation from all previously reported forms of childhood enteropathy was the presence of a unique feature of autoimmunity, namely the presence of IgG colocalizing with complement c1q on the basolateral enterocyte membrane.

Next, in 2003, Ashwood *et al*[12] reproduced the findings of Furlano and Torrente using the newer technique of flow cytometry (as opposed to immunostaining). This group demonstrated elevated CD3 and CD8 cellular infiltrate in the mucosa (duodenum, terminal ileum, and colon) of 52 GI symptomatic children with ASD when compared to inflamed and non-inflamed mucosal tissues from 54 non ASD controls. They also reported statistically similar degrees of infiltrate compared to inflamed controls having other diagnoses (*e.g.,* IBD, indeterminate colitis, food allergy, celiac disease, *etc.*). Additionally, intraepithelial lymphocytosis and B-cell infiltration were present in GI symptomatic children with ASD but not in healthy children. The authors noted that this study confirmed and expanded upon the previously described unique nature of the combined inflammatory components in GI symptomatic children with ASD. Importantly, it was the first time that this pathology was shown to exist simultaneously in the small and large intestine.

In 2004, Torrente *et al*[13] described unique inflammatory pathology in the stomach of 20 (out of 25; 80%) GI symptomatic children with ASD when compared to 30 non-ASD controls. This pathology profile, like the one described by Torrente in the small bowel, was unique and unlike the focal enhanced gastritis seen in CD. Specifically, it consisted of a markedly elevated CD8 T-cell infiltration, marked presence of intraepithelial lymphocytes, and had the same subepithelial basement membrane IgG deposition colocalization with complement c1q seen in the small bowel.

In 2005, Balzola *et al*[14] reported similar chronic active colitis and gastritis (with gastric pseudopolyps) in a 28-year-old patient with regressive autism. Importantly, capsule endoscopy was also performed and revealed jejunal and ileal erosions and ulcerations. This remains the only published report of capsule endoscopic findings in a patient with autism. The authors suggested that their findings of small bowel disease beyond the limits of the duodenum and terminal ileum “demonstrate the potential for involvement of the entire bowel in this inflammatory disease”.

A 2008 case report of two GI-symptomatic patients (one 18-year-old male; one 19-year-old female) with autism describes the precise histopathologic findings previously reported [*i.e.,* non-specific gastritis, focal active colitis, and lymphonodular hyperplasia (LNH) of the terminal ileum] and reported clinical GI symptom improvement in the male patient following a course of steroids after a failed trial of high dose 5-ASA[15]. In their discussion of the findings, the authors commented that “although the idea of a shared pathophysiology between GI disease and autism remains controversial, the evidence presented so far warrants further exploration at the very least” and they advocated for a heightened awareness and lower threshold for work-up and management on this population.

In 2010, Krigsman *et al*[16] reported on the retrospective histologic findings (on routine H&E staining) as reported by the hospital pathologists, of 143 consecutive children with ASD and chronic GI symptoms. It remains the largest study of GI symptomatic ASD mucosal biopsy tissue to date. Based on H&E staining, the authors found ileitis in one third (35%), and ileal or colonic chronic active inflammation in nearly three fourths (74%) of the children. LNH, a feature of inflammation that may be observed at any site while not by itself meeting the definition for inflammation, was present in nearly three fourths (73.2%) of the patients, and for this patient series the presence of LNH predicted the presence of inflammation. Regarding LNH in general, and specifically in children with ASD, this finding represented an important observation because although the prominent presence of LNH in GI-symptomatic children with ASD had been noted (and vigorously debated) in the literature, this case series, due to the large size of the cohort, was the first to report a statistically significant link between LNH and actual mucosal inflammation. Regressive or plateau autism was more strongly linked to ileocolitis than early onset autism. This case series did not seek to identify inflammatory characteristics unique to ASD.

In 2013, Walker *et al*[17] expanded upon the uniqueness of the ASD-associated enterocolitis with characterization of the infiltrates at the molecular level. This case-control study utilized mRNA transcriptome profiling of inflamed ileal and colonic tissue from GI-symptomatic children with ASD and from controls consisting of anatomically matched (ileal and colonic) GI biopsy tissues from GI-symptomatic non-ASD control patients with either: (1) No histologic evidence of pathology; (2) CD; or (3) Ulcerative colitis. Comparison of differentially expressed transcripts between the ASD samples and each of the non-ASD groups showed a clear distinction between the ASD-inflamed mucosa and all other groups. Of particular interest, while there was some degree of overlap between gene expression in inflamed tissue from GI-symptomatic children with ASD and the control samples from non-ASD children with IBD (both CD and UC), no significant overlap existed between ASD and non-diseased controls. The authors noted that this correlates well with the similarities of GI symptom expression and response to therapy as seen in their clinical experience with these patients. Nearly identical molecular profile patterns emerged from both the ileal and colonic mucosal specimens from GI-symptomatic children with ASD which served to confirm the notion of unique ASD inflammatory patterns in both the small and large intestine. The differentially expressed mucosal transcripts unique to ASD that were identified in this study were subsequently evaluated in a follow up 2016 study that compared mucosal gene expression to blood gene expression simultaneously obtained in the same patients, to identify which of these biomarkers might serve as a molecular signature (*i.e.,* a “blood test”) for the presence of ASD-specific inflammatory bowel disease[18].

A subsequent 2016 report by Kushak *et al*[19] evaluated intestinal function in GI-symptomatic children with autism by measuring duodenal disaccharidase activity (lactase, sucrase, and palmitase), intestinal permeability (endoscopic infusion of rhamnose and lactulose followed by measurements in urine over the next 5 h), and fecal markers of intestinal inflammation (stool calprotectin). Results suggested that intestinal permeability, brush border enzyme activity, and the frequency of inflammation did not differ between children with or without autism. Routine H&E staining showed no significant difference in the frequency of gastric or intestinal inflammation between GI symptomatic ASD and non-ASD children and although 52% of children in the ASD group had some inflammation in the gastrointestinal tract, “it was generally mild and nondiagnostic”. The authors concluded that children with autism who have symptoms of GI disorders exhibit objective findings that are similar to those in children without autism.

Lastly, Alessandria *et al*[20] investigated the distribution of human leukocyte antigen-DQ2/DQ8 typing in GI-symptomatic patients with ASD, together with its correlation with duodenal histology and response to a gluten free casein-free diet. The study found unexplained duodenal intraepithelial lymphocytosis in 37% (56/151) of GI symptomatic ASD children, none of which were found to have celiac disease serologic markers. The authors concluded that their results suggest that GI-symptomatic children with ASD seem to have a high prevalence of duodenal histologic inflammation that is not linked to celiac disease.

**INTESTINAL MUCOSAL PRO-INFLAMMATORY CYTOKINE ACTIVITY IN autism spectrum disorder**

Evaluation and comparison of biopsy obtained mucosal tissue from the terminal ileum, ascending, and descending colon from GI-symptomatic children with pervasive developmental disorder (PDD) with age matched controls was first reported in 2003 by DeFelice *et al*[21]. Study subjects consisted of 6 children with PDD (3 with a specific diagnosis of autism or ASD) and 9 neurotypical controls. Using an ELISA kit assay of organ culture supernatants, the authors found no statistically significant differences in the concentration of IL-6, IL-8, and IL-1β between PDD and controls. Histologic findings for all patients, based on H&E staining, were nonpathological. The authors concluded that the data failed to support an association between autism and GI inflammation.

In 2004, Ashwood *et al*[22] prospectively tested the hypothesis that GI-symptomatic children with ASD demonstrated increased numbers of pro-inflammatory cytokine producing CD3+ lymphocytes. As compared to the previously cited 2003 study, this prospective and larger study of 21 children with ASD and 65 neurotypical pediatric controls focused on the intracellular content of pro-inflammatory cytokines as opposed to the cellular supernatant and evaluated the number of pro-inflammatory cytokine containing CD3+ lymphocytes using multicolor flow cytometry. Significant findings included elevated pro-inflammatory cytokine containing lymphocyte counts in children with ASD *vs* controls, particularly lymphocytes containing TNF-α, IL-2, IL-4, and IFN-γ. Significantly, the counterregulatory IL-10 was reduced in ASD as compared to controls. These trends were found in both the duodenum and colon, as well as in both the epithelium and lamina propria. The authors concluded that that there is a consistent profile of CD3+ lymphocyte cytokines in the small and large intestinal mucosa of GI-symptomatic children with ASD, providing further evidence of a diffuse mucosal immunopathology that may respond to dietary and immunomodulatory therapeutic approaches.

In 2006 Ashwood *et al*[23] using a different prospective patient cohort consisting of GI-symptomatic children with ASD (*n* = 18), and typically developing controls (*n* = 27), including non-inflamed controls (NIC), and inflamed GI control children with CD, further elaborated on their 2004 findings. As in the earlier study, the group used flow cytometry to assess the CD3+ lymphocyte intracellular pro-inflammatory and counterregulatory cytokine profiles, this time in mucosal tissue from the terminal ileum. In this study, the CD3+ lymphocyte cytokine profiles were also measured in peripheral blood and correlated to those from the mucosa. In both the peripheral blood and mucosa, CD3+ TNFα+ and CD3+ IFNγ+ were increased in children with ASD compared to NIC, reaching levels similar to those seen in CD. In contrast, peripheral and mucosal counterregulatory CD3+ IL-10+ were markedly lower in GI-symptomatic children with ASD compared to NIC and CD controls. The authors concluded that there was a unique pattern of peripheral blood and mucosal CD3+ lymphocyte intracellular cytokines, consistent with significant immune dysregulation, in that ASD cohort. This 2006 paper marks the last published investigation of mucosal inflammatory cytokine findings in children with ASD and chronic gastrointestinal disease.

In the years between 2006 and 2018, as the incidence of autism continued to rise (according to official reporting from the CDC) and the interest in identifying biomarkers for ASD was high, there was a renewed focus (and a concomitant rise in peer-reviewed publication activity) on peripheral cytokine activity in children with autism. A 2019 meta-analysis on all circulating (blood, serum, or plasma) pro-inflammatory cytokine related literature in ASD identified 38 studies, with a total of 2487 participants (1393 with ASD and 1094 non-ASD controls) and found evidence for higher concentrations of IFN-γ, IL-1β, IL-6 and TNF-α in participants with autism[24]. A second meta-analysis by the same group, also published in 2019, examined levels of circulating anti-inflammatory cytokines in ASD[25]. This report summarized data from 25 studies with a total of 1754 participants (1022 patients with ASD and 732 controls) and found lower concentrations of IL-10 and IL-1Ra in ASD subjects compared to controls.

**MUCOSAL BRUSH BORDER DIGESTIVE ENZYMES**

Disaccharidases, the enzymes that break down dietary carbohydrates, reside in the brush border lining of the small bowel. Deficiencies in the level of these enzymes and/or their activity can lead to unabsorbed carbohydrates reaching the colon where they are fermented by colonic bacteria. Aside from causing symptoms of diarrhea and gaseousness, these fermentation products have been hypothesized to play a role in exacerbating behavioral symptoms in children with ASD, possibly through abnormal absorption in the permeable GI mucosa. Due to the accessibility of the duodenum *via* endoscopic biopsy, the levels and activity of these enzymes in the mucosa can be measured relatively easily and as a result, several studies have been able to explore disaccharidase activity deficiencies in children with ASD.

For example, in 1999, Horvath *et al*[9] evaluated the structure and function of the upper GI tract in a group of 36 GI-symptomatic children with autism and found that 58% (21/36) had deficiencies in at least one brush border disaccharidase enzyme, most often lactase (14/21). Kushak *et al*[26] measured disaccharidase activity in duodenal biopsies from 199 GI-symptomatic individuals with autism and reported a lactase deficiency in 58% of autistic children < 5 years old and 65% in older patients. Finally, Williams *et al*[27] in a small case control study that investigated impaired carbohydrate digestion and transport in the GI tract of children with autism and GI disease compared to controls (GI disease alone), found deficient ileal mRNA transcripts encoding intestinal disaccharidases and hexose transporters in the children with ASD. This approach, though novel, did not ultimately measure actual enzyme activity, but the finding of deficient transcript levels provided additional support for the findings reported by Horvath *et al*[9] and Kushak *et al*[19].

Of interest is the case-control 2016 study by Kushak *et al*[19] which again found a similarly high frequency of lactase and other brush border enzyme deficiencies in GI symptomatic children with ASD, but also a similarly high frequency of disaccharidase deficiency in GI symptomatic neurotypical children undergoing diagnostic endoscopy. While disaccharidase deficiency is by no means a unique feature in autism, this finding in persons with ASD suggests it may play an important role in cognitive/behavioral function in conjunction with numerous other pathophysiologic GI activity known to occur in ASD.

**INTESTINAL MUCOSAL PERMEABILITY**

Intestinal mucosal permeability, or “leaky gut”, has been linked to developmental and behavioral changes in children. For example, a study published in 2010 that evaluated alterations in the intestinal barrier in patients with ASD and their first-degree relatives found a high percentage of abnormal intestinal permeability among patients (36.7%) and their relatives (21.2%) as compared to normal subjects (4.8%)[28]. The authors concluded that their results provided support for the leaky gut hypothesis.

A recent 2019 review of the concept of ‘leaky gut’ discusses how intestinal barrier integrity is commonly measured, what clinical impact a compromised intestinal barrier can have in humans, and sheds light on the nuances of this of this complex topic[29]. Although the review was not specifically about the leaky gut hypothesis in ASD-associated GI disease, the author’s conclusion that “clinicians should be aware of the potential of barrier dysfunction in GI diseases and of the barrier as a target for future therapy” is highly relevant to this discussion.

Between 1996-2016 at least five investigations into the frequency and extent of intestinal permeability in children with ASD were published[19,30-33], but the heterogeneity in study design and the exclusion of GI symptomatic children in some of the studies makes it difficult to draw meaningful conclusions as to the contribution of an excessively permeable membrane to cognitive and behavioral dysfunction in children with ASD and inflammatory GI disease. However, taken together, these studies report an overall greater frequency of increased intestinal permeability in ASD as compared to non-ASD controls, though it appears that this finding does not reach statistical significance in the more rigorous studies.

Lastly, in a study investigating blood-brain barrier and intestinal epithelial barrier function, Fiorentino *et al*[34] using duodenal mucosal biopsies from GI-symptomatic children with ASD (*n* = 12) and healthy controls (*n* = 9), found that 75% of the ASD samples had reduced expression of barrier-forming tight junction proteins, suggesting an impaired gut barrier integrity.

**INFLUENCE OF THE MICROBIOTA**

Human health and well-being are inextricably linked to a balanced microbiome and it is well established that intestinal dysbiosis can alter GI physiology, immune function, and even behavior in children with ASD, a population in which microbial gut dysbiosis is common[35]. In the context of the gut-brain-axis in ASD, the influence of the microbiome is an area of intense interest and research activity. Although published studies report imbalances in intestinal microbiome composition in children with ASD[36-38] and have investigated the relationship between dysbiosis and ASD-related cognition, behavior, and GI symptomatology[39], the mechanisms by which the microbiota are thought to alter GI function and behavior are still actively being investigated. A more comprehensive understanding of mechanisms by which dysbiosis occurs, and how it impacts host function, will accelerate the development of clinically relevant mitigation and therapeutic strategies[40].

**CLINICAL ENDOSCOPIC AND LABORATORY OBSERVATIONS/EXPERIENCE OF THE AUTHORS**

***Serum antibodies to luminal bacterial and fungal microbes***

In clinical gastroenterology, the presence of serological antibodies to normal intestinal flora and their association with IBD has been well established. Though not used clinically as a diagnostic tool, the presence of these antibodies distinguishes one form of IBD from another and emphasizes the prominent role of enteric bacteria and fungi as antigenic stimuli in IBD; only rarely are they present in the general and disease control populations. Their presence does not inform specific treatment strategies, but they do suggest an important microbiome related pathogenic pathway for development of intestinal mucosa inflammation in IBD.

To date, there have not been published studies that have investigated the frequency of these serologic markers in ASD-associated IBD. It has been the experience of the authors that, although the frequency with which they are present in ASD-IBD is somewhat less than in CD[41,42], they occur with much greater frequency than in the general population. As with CD, there is age-related appearance of these markers such that the anti-flagellan antibodies are prominent in children under 10 years of age, whereas the anti-glycan antibodies (*e.g.,* ASCA) are more dominant in the teenage and young adult patients[43].

***Focal patchy lymphangiectasia***

Intestinal lymphangiectasia (the presence of lipid and protein-rich fluid within dilated small intestinal villi) is another endoscopic finding commonly encountered by the authors in teenage and young adult patients with ASD and enteritis who have a long-standing history of untreated gastrointestinal symptoms. Although lymphangiectasia may be found in any number of disease states that prevent the free lymphatic flow away from the intestinal lumen (*e.g.,* elevated central venous pressure secondary to right sided cardiac failure, lymphoma of draining lymph nodes causing distal obstruction, *etc.*), it has also been noted in intestinal inflammatory states such as CD where the obstruction to outward lymph flow occurs in the intestinal wall layers as it does in ASD-enteritis[44]. Examples of intestinal lymphangiectasia in ASD enteritis co-occurring in the same region as classic IBD aphthous ulcers, and in adjacent mucosa in the same patient is presented in Figure 1. To date, the authors have not encountered protein-losing enteropathy resulting from lymphangiectasia.

In the experience of the authors, a third type of endoscopic lesion frequently seen in younger ASD-GI children (less than 10 years of age) is that of white spots, either singular or in clusters, that are prominent in the proximal small bowel and present, but much less frequent, in the distal small bowel (Figure 2). They are often, but not always, seen in conjunction with mildly erythematous mucosa, and inflammatory infiltrate is typically not noted on routine H&E staining. Upon histologic multiple sectioning of the biopsy specimen, truncated villi are noted (single or contiguous) that demonstrate a protective fibrous plume emanating from the core of the decapitated villi. Villous decapitation allows luminal contents to evade the highly evolved system of selective absorption and results in increased permeability (Figure 3). In these cases, capsule endoscopy does not reveal the typical lipid filled villi, but rather a white spot on the surface of the mucosa not within a villous structure.

**POTENTIAL MECHANISMS OF CNS INJURY SECONDARY TO UNIQELY INFLAMED GI MUCOSA**

Based on the cumulative published literature discussed to this point detailing what is known about the pathology of the inflamed GI mucosa in GI-symptomatic children with ASD, a unidirectional (bowel to brain) mechanism of enterogenous CNS injury in ASD is proposed (Figure 4).

For reasons that are not yet entirely clear - but that include environmental, epigenetic, maternal-fetal, and perhaps genetic factors - the immune response in ASD is primed towards over-responsiveness. This over-responsiveness manifests as a heavy lamina propria B and T-cell presence, IgG colocalization with complement c1q on the basolateral enterocyte membrane[11], and intraepithelial lymphocytosis[12,20]. Excessive levels of pro-inflammatory cytokines produced by these cell lineages, as well as deficiencies in counter-regulatory IL-10, are consistently present[22,23]. Marked LNH, a persistently occurring feature of GI immunopathology, is typically present and predicts the presence of cellular inflammation[16]. The combination of hypertrophied reactive LNH and the thickened fibrous basement membrane[11] may serve to obstruct, in whole or in part, lymphatic flow away from the mucosa, resulting in lymphangiectasia. The gene transcriptional profile of the inflammatory infiltrate is unique to children with autism and overlaps significantly with CD[17]. Luminal bacteria are antigens known to be targeted by the mucosal antibodies (Krigsman, unpublished observation).

This persistent inflammatory presence in the intestinal mucosa eventually results in mucosal and epithelial architectural damage. Villous destruction occurs (truncated decapitated villi) with attempts at self-reparation by villous production of a fibrinous covering. Inflammatory villous damage and destruction results, as always, in the loss of resident brush border enzymes[9,26] and increased intestinal permeability[28,30,34]. The clinical implications of the latter two findings are that luminal contents (food, degradation products of resident flora, medications) can more easily evade the highly developed mechanisms for selective absorption and enter the mucosa and capillaries of the GI tract as macro, not micro, molecules, thus exposing the local and systemic immune system to unrecognized antigens. The result of this immunologic activity at the level of the intestinal mucosa is the production of activated B and T cells, as well as numerous pro-inflammatory cytokines, antibodies, and antigen-antibody complexes, that may then enter the peripheral circulation. Once in the peripheral circulation, the effects of such a wide array of active inflammatory activity is largely unknown, but would help explain the observation by Jyonouchi[45] who in 2005 described an increase in GI symptoms and systemic pro-inflammatory cytokines after exposure to specific dietary antigens in GI symptomatic ASD children.

The presence of autoantibodies to the GI epithelium, *i.e.,* autoimmunity, often heralds the formation of autoantibodies directed against other organs as well[46] and thus, the inflammatory activity emanating from the bowel may contribute to the development of the numerous CNS autoantibodies described in the literature[47-49]. Additionally, cytokines produced locally in the GI mucosa travel peripherally and can cross the blood brain barrier[50].

Of note, the mechanisms of brain inflammation originating in the bowel may be entirely independent of immune dysregulation of peripheral origin that results in production of anti-CNS antibodies or intracranial cellular and molecular mediators of inflammation. The tendency of autoimmune processes to co-exist in the body and even to augment each other has been discussed elsewhere[46].

Finally, although a number of studies have investigated the role of immunomodulating therapy, targeting inflammatory pathways, as a treatment for autism have appeared over the years (reviewed in 51), the focus of the current review is to understand how the existence of a unique inflammatory bowel disease might impact the brain, and, by extension, how treatment of this IBD might ameliorate core symptoms of ASD. Although “classic” IBD such as CD is not known to affect cognition and behavior in the manner described in ASD, the difference may lie in the unique inflammatory features of ASD-associated enteritis that distinguishes it from CD (as already discussed), and the early age of onset of the ASD enteritis, a time when the brain is most susceptible to toxic insult.

**CONCLUSION**

We have presented a mechanistic concept, based on published pathologic immunologic processes occurring in the gastrointestinal tract of GI-symptomatic children with autism, describing how such pathology may be contributing to the cognitive and behavioral deficits seen in ASD. The logical next step is to conduct a prospective treatment study to determine whether standard immunotherapy for IBD results in improvement in GI symptoms and histologic disease, and if this improvement correlates with improvement in the ASD cognitive and behavioral domains as well.

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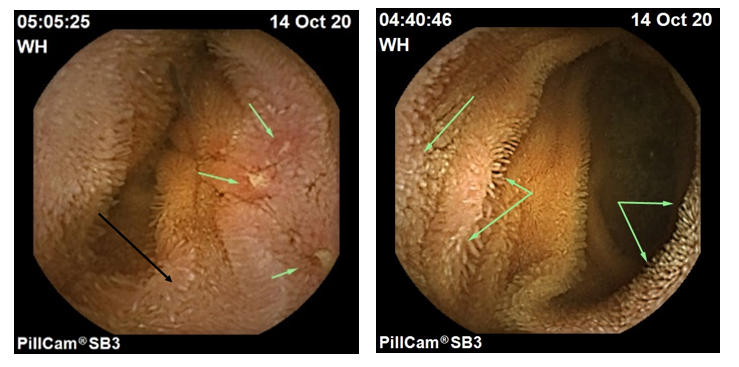
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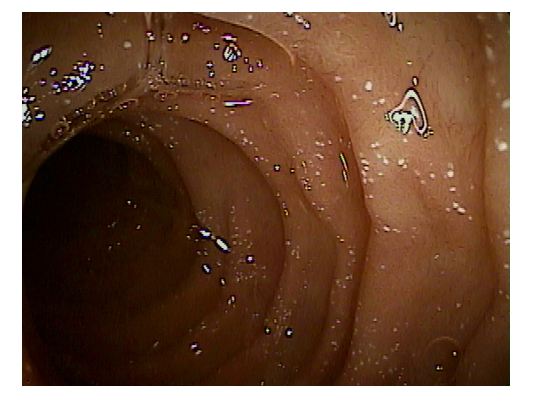
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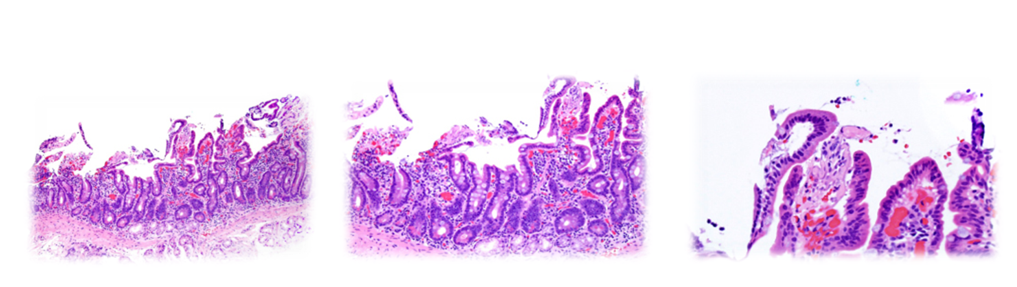
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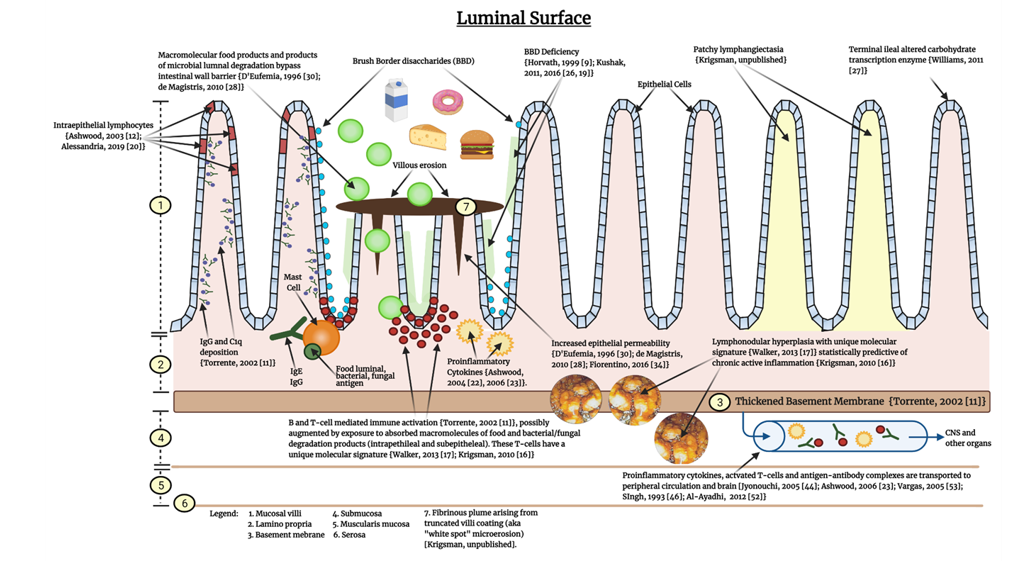
**Figure 1 Pillcam images from the small bowel of a gastrointestinal-symptomatic patient with** **autism spectrum disorder.** Lymphangiectasia in patient with autism spectrum disorder-associated enteritis (left panel, black arrow); the patient also exhibited aphthous ulcerations (green arrows); Jejunal lymphangiectasia (green arrows) in adjacent mucosa of same patient (right panel).



**Figure 2** **Endoscopic image of white spot lesions of the proximal small** **bowel in a gastrointestinal-symptomatic child with autism spectrum disorder.**



**Figure 3** **Histologic staining of a mucosal biopsy specimen from the proximal small bowel of a patient with autism spectrum disorder-associated enteritis**. White spot lesions (truncated villi) can be seen in low (left); medium (middle); and high magnification (right) of the H&E-stained images.



**Figure 4 Schematic diagram depicting pathological findings in a portion of the gastrointestinal tract of a representative gastrointestinal-symptomatic child with autism spectrum disorder[9,11,12,16,17,19,20,22,23,26,27,28,30,34,46,52,53].** Key landmarks are identified in the legend (#1-7) and key findings are described throughout the diagram.



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