# World Journal of *Critical Care Medicine*

World J Crit Care Med 2022 July 9; 11(4): 201-310





Published by Baishideng Publishing Group Inc

World Journal of C C M Critical Care Medicine

### Contents

### **Bimonthly Volume 11 Number 4 July 9, 2022**

### **REVIEW**

201 Effects of nutrients on immunomodulation in patients with severe COVID-19: Current knowledge

Costa BTD, Araújo GRL, da Silva Júnior RT, Santos LKS, Lima de Souza Gonçalves V, Lima DBA, Cuzzuol BR, Santos Apolonio J, de Carvalho LS, Marques HS, Silva CS, Barcelos IS, Oliveira MV, Freire de Melo F

### **MINIREVIEWS**

219 Challenges in hyperglycemia management in critically ill patients with COVID-19 Kethireddy R, Gandhi D, Kichloo A, Patel L

228 Medicinal nicotine in COVID-19 acute respiratory distress syndrome, the new corticosteroid

Ahmad F

236 Health-related quality-of-life and health-utility reporting in critical care

> Lau VI, Johnson JA, Bagshaw SM, Rewa OG, Basmaji J, Lewis KA, Wilcox ME, Barrett K, Lamontagne F, Lauzier F, Ferguson ND, Oczkowski SJW, Fiest KM, Niven DJ, Stelfox HT, Alhazzani W, Herridge M, Fowler R, Cook DJ, Rochwerg B, Xie F

### **ORIGINAL ARTICLE**

### **Observational Study**

Septic shock 3.0 criteria application in severe COVID-19 patients: An unattended sepsis population with 246 high mortality risk

Cidade JP, Coelho L, Costa V, Morais R, Moniz P, Morais L, Fidalgo P, Tralhão A, Paulino C, Nora D, Valério B, Mendes V, Tapadinhas C, Povoa P

255 Development and pilot implementation of a patient-oriented discharge summary for critically Ill patients

Shahid A, Sept B, Kupsch S, Brundin-Mather R, Piskulic D, Soo A, Grant C, Leigh JP, Fiest KM, Stelfox HT

### SYSTEMATIC REVIEWS

269 Immunomodulatory therapy for the management of critically ill patients with COVID-19: A narrative review

Andaluz-Ojeda D, Vidal-Cortes P, Aparisi Sanz Á, Suberviola B, Del Río Carbajo L, Nogales Martín L, Prol Silva E, Nieto del Olmo J, Barberán J, Cusacovich I

### **META-ANALYSIS**

298 Association between early viral lower respiratory tract infections and subsequent asthma development

Kenmoe S, Atenguena Okobalemba E, Takuissu GR, Ebogo-Belobo JT, Oyono MG, Magoudjou-Pekam JN, Kame-Ngasse GI, Taya-Fokou JB, Mbongue Mikangue CA, Kenfack-Momo R, Mbaga DS, Bowo-Ngandji A, Kengne-Ndé C, Esemu SN, Njouom R, Ndip L



### Contents

**Bimonthly Volume 11 Number 4 July 9, 2022** 

### **ABOUT COVER**

Peer Reviewer of World Journal of Critical Care Medicine, Zhong-Heng Zhang, Doctor, MD, MSc, Chief Physician, Doctor, Professor, Department of Emergency Medicine, Sir Run Run Shaw Hospital, Hangzhou 310016, Zhejiang Province, China. zh\_zhang1984@zju.edu.cn

### **AIMS AND SCOPE**

The primary aim of the World Journal of Critical Care Medicine (WJCCM, World J Crit Care Med) is to provide scholars and readers from various fields of critical care medicine with a platform to publish high-quality basic and clinical research articles and communicate their research findings online.

WJCCM mainly publishes articles reporting research results and findings obtained in the field of critical care medicine and covering a wide range of topics including acute kidney failure, acute respiratory distress syndrome and mechanical ventilation, application of bronchofiberscopy in critically ill patients, cardiopulmonary cerebral resuscitation, coagulant dysfunction, continuous renal replacement therapy, fluid resuscitation and tissue perfusion, hemodynamic monitoring and circulatory support, ICU management and treatment control, sedation and analgesia, severe infection, etc.

### **INDEXING/ABSTRACTING**

The WJCCM is now abstracted and indexed in PubMed, PubMed Central, Reference Citation Analysis, China National Knowledge Infrastructure, China Science and Technology Journal Database, and Superstar Journals Database.

### **RESPONSIBLE EDITORS FOR THIS ISSUE**

Production Editor: Yi-Xuan Cai; Production Department Director: Xiang Li; Editorial Office Director: Li-Li Wang.

NAME OF JOURNAL	INSTRUCTIONS TO AUTHORS
World Journal of Critical Care Medicine	https://www.wjgnet.com/bpg/gerinfo/204
ISSN	GUIDELINES FOR ETHICS DOCUMENTS
ISSN 2220-3141 (online)	https://www.wjgnet.com/bpg/GerInfo/287
<b>LAUNCH DATE</b>	GUIDELINES FOR NON-NATIVE SPEAKERS OF ENGLISH
February 4, 2012	https://www.wjgnet.com/bpg/gerinfo/240
FREQUENCY	PUBLICATION ETHICS
Bimonthly	https://www.wignet.com/bpg/GerInfo/288
EDITORS-IN-CHIEF	PUBLICATION MISCONDUCT
Hua-Dong Wang	https://www.wignet.com/bpg/gerinfo/208
EDITORIAL BOARD MEMBERS	ARTICLE PROCESSING CHARGE
https://www.wjgnet.com/2220-3141/editorialboard.htm	https://www.wignet.com/bpg/gerinfo/242
PUBLICATION DATE	STEPS FOR SUBMITTING MANUSCRIPTS
July 9, 2022	https://www.wjgnet.com/bpg/GerInfo/239
COPYRIGHT	ONLINE SUBMISSION
© 2022 Baishideng Publishing Group Inc	https://www.f6publishing.com

© 2022 Baishideng Publishing Group Inc. All rights reserved. 7041 Koll Center Parkway, Suite 160, Pleasanton, CA 94566, USA E-mail: bpgoffice@wjgnet.com https://www.wjgnet.com



World Journal of C C M Critical Care Medicine

World J Crit Care Med 2022 July 9; 11(4): 228-235

DOI: 10.5492/wiccm.v11.i4.228

Submit a Manuscript: https://www.f6publishing.com

ISSN 2220-3141 (online)

MINIREVIEWS

# Medicinal nicotine in COVID-19 acute respiratory distress syndrome, the new corticosteroid

### Farrukh Ahmad

Specialty type: Critical care medicine

Provenance and peer review: Unsolicited article; Externally peer reviewed.

Peer-review model: Single blind

### Peer-review report's scientific quality classification

Grade A (Excellent): 0 Grade B (Very good): 0 Grade C (Good): C Grade D (Fair): 0 Grade E (Poor): 0

P-Reviewer: Juneja D, India

Received: November 14, 2021 Peer-review started: November 14, 2021 First decision: April 7, 2022 Revised: April 23, 2022 Accepted: June 18, 2022 Article in press: June 18, 2022 Published online: July 9, 2022



Farrukh Ahmad, Department of Internal Medicine, Saint Vincent Hospital, Worcester, MA 01608, United States

Corresponding author: Farrukh Ahmad, MD, Staff Physician, Department of Internal Medicine, Saint Vincent Hospital, 123, Summer Street, Worcester, MA 01608, United States. farrukh2.ahmad@stvincenthospital.com

### Abstract

The cholinergic anti-inflammatory pathway (CAP) refers to the anti-inflammatory effects mediated by the parasympathetic nervous system. Existence of this pathway was first demonstrated when acetylcholinesterase inhibitors showed benefits in animal models of sepsis. CAP functions *via* the vagus nerve. The systemic antiinflammatory effects of CAP converges on the a7 nicotinic acetylcholine receptor on splenic macrophages, leading to suppression of pro-inflammatory cytokines and simultaneous stimulation of anti-inflammatory cytokines, including interleukin 10. CAP offers a novel mechanism to mitigate inflammation. Electrical vagal nerve stimulation has shown benefits in patients suffering from rheumatoid arthritis. Direct agonists like nicotine and GTS-1 have also demonstrated antiinflammatory properties in models of sepsis and acute respiratory distress syndrome, as have acetylcholinesterase inhibitors like Galantamine and Physostigmine. Experience with coronavirus disease 2019 (COVID-19) induced acute respiratory distress syndrome indicates that immunomodulators have a protective role in patient outcomes. Dexamethasone is the only medication currently in use that has shown to improve clinical outcomes. This is likely due to the suppression of what is referred to as a cytokine storm, which is implicated in the lethality of viral pneumonia. Nicotine transdermal patch activates CAP and harvests its anti-inflammatory potential by means of an easily administered depot delivery mechanism. It could prove to be a promising, safe and inexpensive additional tool in the currently limited armamentarium at our disposal for management of COVID-19 induced acute hypoxic respiratory failure.

Key Words: COVID-19; Acute respiratory distress syndrome; Medicinal nicotine; Cholinergic anti-inflammatory pathway; Corticosteroid

©The Author(s) 2022. Published by Baishideng Publishing Group Inc. All rights reserved.



WJCCM | https://www.wjgnet.com

Core Tip: Cholinergic anti-inflammatory pathway is novel pathway of the inflammatory reflex. Activation of this pathway can suppress maladaptive inflammatory response seen in coronavirus disease 2019 (COVID-19) acute respiratory distress syndrome (ARDS). Nicotine is a potent activator of this pathway and may offer benefits in the management of COVID-19 ARDS, via immune suppressive effects similar to dexamethasone.

Citation: Ahmad F. Medicinal nicotine in COVID-19 acute respiratory distress syndrome, the new corticosteroid. World J Crit Care Med 2022; 11(4): 228-235 URL: https://www.wjgnet.com/2220-3141/full/v11/i4/228.htm

DOI: https://dx.doi.org/10.5492/wjccm.v11.i4.228

### INTRODUCTION

A dramatic inflammatory response is a common manifestation of severe coronavirus disease 2019 (COVID-19) infection[1]. The purpose of such an inflammatory surge, under normal conditions, is to allow the body to attack, constrain, and kill invading organisms. However, that same inflammatory cascade has negative downstream consequences which can cause direct damage to the host.

Sepsis is the consequence of this hyperactive immune state, most commonly due to a poorly controlled infection or significant tissue injury<sup>[2]</sup>. The unbalanced immune reaction perpetuates further injury. Neutrophils are recruited and infiltrate the lungs where they undergo apoptosis, further causing tissue damage leading to the development of shock and acute respiratory distress syndrome (ARDS)[3]. These cells and the molecules they release are a potent force designed to neutralise pathogens, but cause significant collateral damage in the process. Another casualty of this inflammatory dysregulation is vasodilatation and microvascular thrombi that lead to poor tissue perfusion, further perpetuating the cycle of destruction. This self-perpetuating cycle of tissue damage and release of pro-inflammatory cytokines[4,5] causes further dysregulation of the immune system.

Cytokine is a term given to molecules that carry out inflammatory responses of the immune system, each having their respective receptors distributed across the body. They orchestrate most, if not all, of the consequences of sepsis. This phenomenon is now dubbed a 'cytokine storm'[6] and has been particularly devastating in the current pandemic of COVID-19 infection [7,8].

In recent years many immune modulators have been administered to mitigate sepsis and shock but with limited success in changing the disease course, morbidity, and mortality outcomes. Tocilizumab was used widely during the initial phase of the COVID-19 pandemic in ICUs across the world. But it failed to demonstrate mortality benefits[9]. The reason could partly be explained by the fact that it has a narrow scope of action, only blocking the interleukin (IL)-6 receptor. Upregulation of alternate pathways of inflammation likely are at play. A mechanism to reduce the global immune response is required to suppress collectively the molecules perpetuating inflammation. Corticosteroids are touted as one of the strongest tools in our arsenal to achieve such a goal. Dexamethasone is the only drug we have at our disposal that has shown mortality benefits during the COVID-19 pandemic[10]. Although corticosteroids are considered to globally suppress inflammation, patients are still succumbing to this coronavirus infection despite high doses administered over several days. Other medications for global suppression of inflammation are needed.

One potential pathway that may hold promise in achieving global suppression of the immune system is the cholinergic anti-inflammatory pathway (CAP). CAP is a component of the inflammatory reflex, mediated by the cholinergic nervous system and augmenting its tone has been shown to decrease inflammation in both human and animal models. The first evidence of the cholinergic system having immunomodulatory properties dates back to 1987. Zabrodskii[11] showed that Armin, an irreversible acetylcholinesterase inhibitor reduces mortality in animal models of sepsis. It was first recognized in humans when patients with Rheumatoid Arthritis and drug-resistant epilepsy underwent Vagal Nerve stimulation to ameliorate their recurrent seizures. After initiation of Vagal Nerve stimulation, patients incidentally reported improvement in joint pains[12].

### INFLAMMATORY REFLEX

The inflammatory reflex [13] is a central nervous system mediated reflex arc that modulates the immune system. Like other prototypical reflexes, it has an incoming and outgoing arm. Instead of a sensory input that begets a motor response, this circuit senses inflammation and responds with appropriate inflammatory inhibition to reestablish homeostasis. The afferent arm is activated by the products of sterile or infectious inflammation.



WJCCM https://www.wjgnet.com

The efferent arm is termed the CAP which, through diverse mechanisms, suppresses inflammation [14]. Both the afferent and efferent limbs of the reflex are transmitted predominantly by the vagus nerves. Tracey KJ team[15,16] has conducted extensive research in the potential therapeutic application of vagal stimulation in modulating the immune system, thereby providing initial major contributions to mapping this pathway (Figure 1)[17,18].

### THE AFFERENT LIMB

We are more familiar with the afferent limb of this pathway[19], which plays a role in triggering the mammalian febrile response. Disrupting the afferent arm, for example with a subdiaphragmatic vagotomy, prevented IL-1 $\beta$  induced fever in mice[20]. The afferent limb is activated by pro-inflammatory cytokines like tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ) and IL-1 $\beta$ , neuropeptide Y and prostaglandins. Vagal fibers innervating visceral organs like the lungs and gastrointestinal tract demonstrate sensitivity to IL-1 $\beta$ . Furthermore, the nodose ganglion expresses Toll-like receptors[18] which are directly stimulated by pathogen associated molecular patterns such as those found on bacterial cell walls[21]. Area postrema directly expresses proinflammatory cytokine receptors[22]. The afferent limb converges on the nucleus tractus solitarius (NTS), the primary central vagal afferent nucleus. Interneurons connect the NTS to the dorsal motor nucleus of vagus (DMV), which are the primary efferent nuclei of the vagus nerve (Figure 2).

### THE EFFERENT LIMB/CAP

The systemic anti-inflammatory effects of CAP are thought to exert its effects *via* the spleen[23,24]. The efferent limb originates at the DMV, the motor nuclei of the vagus nerve. Motor signals are transmitted *via* cholinergic fibers down the vagus nerve to mount an anti-inflammatory response, reestablishing homeostasis. The vagus nerve does not directly innervate the spleen like it does with other visceral organs such as the heart, intestines and liver. So to realize a response from splenic lymphocytes and macrophages, the splenic nerve functions as an intermediary. The efferent pathway is as follows: Cholinergic fibers from the vagus nerve innervate the celiac ganglion; Noradrenergic neurons from the celiac ganglion, *via* the splenic nerve, innervate the spleen, and by releasing norepinephrine stimulate  $\beta$  -2 adrenergic receptors on choline-acetyltransferase positive T cells that reside in the spleen; Activation of the  $\beta$ -2 adrenergic receptors with norepinephrine induces the release of acetylcholine (ACh) from these splenic T cells; ACh then activates  $\alpha$ -7 nicotinic acetylcholinergic receptor ( $\alpha$ 7nAChr) on the splenic macrophages; Activation of  $\alpha$ 7nAChr causes downstream inhibition of the NF-Kappa  $\beta$  pathway and subsequent suppression of pro-inflammatory cytokines. It also induces the release of anti-inflammatory molecules by activating the JAK2-STAT3 pathway[13,14].

Iatrogenic activation of the efferent limb of the inflammatory reflex, irrespective of the modality, has demonstrated anti-inflammatory effects in diverse pathological conditions[15] (Figure 3).

### HARVESTING THE POTENTIAL OF CAP

Augmenting the CAP offers an effective tool in controlling maladaptive inflammatory responses[25,26]. Modulating the cholinergic tone, irrespective of the modality used, has been shown to suppress inflammation[27]. Direct electrical stimulation of the vagus nerve aims to trigger an action potential that consequently activate this pathway downstream. Vagal nerve stimulation has been shown to suppress inflammation and decrease serum levels of TNF, IL-1 $\beta$  and IL-6[28-32]. Pharmacological modalities to increase the activity of CAP have also yielded similar results. Direct agonists of  $\alpha$ 7nAChr like the pharmacological agent nicotine have demonstrated anti-inflammatory properties[33-39]. Ongoing trials using GTS-1, a specific  $\alpha$ 7nAChr agonist, are being conducted in human models of sepsis[40,41]. Another feasible pharmacological strategy is to use inhibitors of acetylcholinesterase to delay degradation of ACh and, thus, enhance the tone of this pathway [42-47]. It must be noted that acetylcholinesterase inhibitors require a functional vagal pathway and fail to demonstrate anti-inflammatory effects in vagotomized animals[48].

Practical modalities for bedside manipulation of CAP is limited. Vagal nerve stimulation has limited feasibility for critically ill septic patients. GTS-1, an  $\alpha$ 7nAChr agonist, is in an experimental phase acetylcholinesterase inhibitors like physostigmine increase cholinergic tone systemically and cause undesirable muscarinic side effects. That currently leaves nicotine as the only feasible and medically available potentiator of CAP as an agonist of  $\alpha$ 7nAChr. As such, it has demonstrated anti-inflammatory properties in ulcerative colitis and models of human sepsis[33,34].

Zaishideng® WJCCM | https://www.wjgnet.com

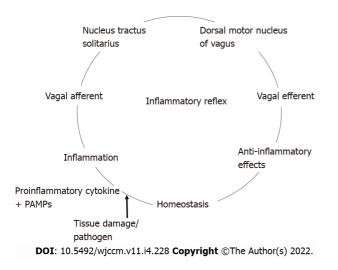


Figure 1 The inflammatory reflex. The above graphic demonstrates the inflammatory reflex. The afferent limb is activated by pro-inflammatory cytokines like tumor necrosis factor and interleukin 1βor by pathogen-associated molecular patterns via Toll-like receptors. The afferent limb connects to the nucleus tractus solitarius (NTS), the primary vagal afferent nuclei. The mammalian febrile response is initiated at the NTS. Interneurons connect NTS to dorsal motor nucleus of vagus (DMV) incoming signals. The DMV is the primary efferent nuclei of the vagus nerve. This efferent signal initiates an anti-inflammatory effect, reestablishing homeostasis. PAMPS: pathogen-associated molecular patterns.

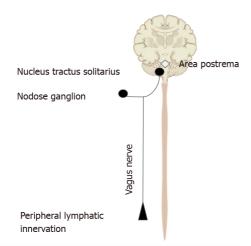




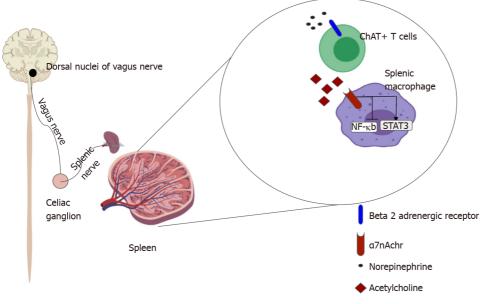
Figure 2 Afferent limb of the inflammatory reflex. This figure demonstrates the mechanisms by which the vagus nerve senses inflammation. Vagal sensory neurons directly express receptors for various pro-inflammatory cytokines such as, tumor necrosis factor, interleukin 1β, neuropeptide Y and prostaglandins. Vagal fibers innervating the lymphatic system demonstrate sensitivity to interleukin-1β. In addition, the nodose ganglion has been shown to express Toll-like receptors. Area postrema directly expresses proinflammatory cytokine receptors[22]. The signal is transmitted via the vagal afferents to the bilateral nucleus tractus solitarius, the primary vagal afferent nucleus[19].

### NICOTINE

Humans have been using nicotine since prehistoric times[49], mostly in the form of tobacco. Even though it is widely acknowledged that smoking or chewing tobacco is unequivocally injurious to health, nicotine by itself has not been shown to be harmful. Medicinal nicotine has demonstrated potent antiinflammatory properties while being safe and possessing a low side-effect profile in short term administration. Nicotine administration in animal models of ARDS and sepsis have shown improved survival with lower serum inflammatory markers and reduced migration of neutrophils[36-38]. Human models of lipopolysaccharide (LPS) induced sepsis show faster resolution of sepsis[33]. Nicotine has also shown anti-inflammatory effects in patients with ulcerative colitis[34,35].

Nicotine patches are well suited as a modality for increasing nicotinic cholinergic receptor activity, and possess the following advantages: Nicotine does not have any underlying muscarinic effects and, therefore, lack concerns of increasing airway secretions that occur with acetylcholinesterase inhibitors like galantamine or physostigmine; Using a nicotine patch achieves therapeutic levels of nicotine in the blood within 4-6 h, offering a rapid drug onset profile[50]; The active drug nicotine has a short half-life of 2 h. Its metabolite, cotinine, has minimal biological activity [51]. This allows for rapid withdrawal of

WJCCM https://www.wjgnet.com



DOI: 10.5492/wjccm.v11.i4.228 Copyright ©The Author(s) 2022.

Figure 3 Efferent limb of the inflammatory reflex. Signal from the dorsal nuclei of vagus is transmitted via cholinergic fibers of the vagus nerve to the celiac ganglion. Noradrenergic neurons from the celiac ganglion via the splenic nerve innervate the spleen. Choline-acetyltransferase positive T cells that reside in the spleen express β-2 adrenergic receptors. Activation of this receptor causes the release of Acetylcholine which binds to the α-7 nicotinic acetylcholinergic receptor on splenic macrophages causing the inhibition of NF-kappa β pathway and upregulation of STAT3, ultimately suppressing inflammation[16,23].

> treatment if necessary. Most acetylcholinesterase inhibitors have a much longer half-life; The depot mechanism of drug delivery for the nicotine patch allows for a rapid onset, prolonged drug delivery during the duration of application, with a quick withdrawal time; The 24-h depot administration avoids repeated administrations and minimized nursing exposure for delivery of the medication; Ease of administration; Nicotine transdermal patches are widely used as clinical medication for nicotine replacement therapy in both the hospital and outpatient settings; There are minimal drug-drug interactions<sup>[52]</sup>.

### IN-HOSPITAL SAFETY DATA ON NICOTINE REPLACEMENT THERAPY

The data on the safety of nicotine on non-smoking patients in an inpatient setting is limited.

Safety data on current or former smokers receiving nicotine replacement therapy in ICU settings and hospital settings fail to demonstrate an increase in adverse events [53-58]. Potential side effects of medicinal nicotine administration are few. They may include hypertension and tachyarrhythmias. Rash at the site of the nicotine patch application has been described. Patients with end stage renal disease have a decreased rate of nicotine metabolism so the safety profile for patients on dialysis is uncertain[59, 60].

### CONCLUSION

The current ongoing pandemic of severe acute respiratory syndrome coronavirus 2 proves a new challenge for the medical community. Owing to the tremendous ingenuity and grit demonstrated by teams across the globe, we now have several promising vaccines which demonstrate remarkable efficacy. However, we are yet to develop a similarly promising tool for management of severe infection which is still very prevalent. Consequently, patients continue to succumb in ICUs across the world to the COVID-19 acute hypoxic respiratory failure and septic shock. Several touted treatment modalities during this pandemic have emerged only to quickly fall out of favour due to lack of documented benefit, including Hydroxychloroquine, Tocilizumab, and transfusion of convalescent plasma. Management for COVID-19 pneumonia, at present, comprises two parallel approaches. Remdesivir or other upcoming potential antivirals, to control viral replication and immunomodulators like dexamethasone to control the maladaptive immune response. Dexamethasone has shown utility in reducing mortality in patients with COVID-19 induced acute hypoxic respiratory failure. However, despite its use early in the course of the disease, many still deteriorate, requiring increased levels of oxygen support or even mechanical ventilation. Patients continue to die even with dexamethasone as



part of their pharmacological regimen. Better modalities are needed to further improve patient outcomes. The hope is bringing to the attention of the medical community a fairly well studied, yet paradoxically unknown pathway of global immune modulation.

CAP is a part of a neural reflex termed the inflammatory reflex. It plays a central role in the neural control of inflammation. Inflammatory reflex has an afferent limb that senses systemic inflammation via the vagus nerve. This signal is relayed to the NTS, the sensory vagal nucleus in the central nervous system. Interneurons then communicate to the DMV, which is the primary motor nucleus of the vagus nerve. The efferent limb of the inflammatory reflex originates from the DMV via motor vagal fibers and trigger various anti-inflammatory mechanisms, reestablishing homeostasis. The systemic anti-inflammatory effects of CAP is thought to be due to suppression of pro-inflammatory cytokines from splenic macrophages. Nicotinic ACh receptors on these splenic macrophages are the point of convergence of this pathway's systemic anti-inflammatory effect. This translates to survival benefits with lower levels of serum TNF-a, and IL-6, along with reduced migration of neutrophils in models of sepsis. The potential of augmenting this pathway to mitigate inflammation has been demonstrated in several animal and human studies.

Nicotine is a commonly used molecule that is a potent activator of α7nAChr, with demonstrated antiinflammatory effects. Animal models of sepsis show improved survival with nicotine administration. Nicotine patch has been studied in the human model of LPS induced sepsis and demonstrated faster resolution of inflammation compared to controls. Nicotine transdermal patch has been used for decades as a means of nicotine delivery for nicotine replacement therapy in active tobacco users and has demonstrated a favorable safety profile. Thus, nicotine transdermal patch may offer a readily available tool with significant benefit-to-risk ratio in the setting of COVID-19 induced acute hypoxic respiratory failure

With patients suffering daily across the globe with COVID ARDS, there is little downside to the administration of this relatively inexpensive, widely available medication with a high safety. There is presently a lack of literature regarding the use of nicotine in COVID-19 ARDS patients and it must be further studied first before being applied routinely.

### FOOTNOTES

Author contributions: Ahmad F contributed hypothesis generation, evidence gathering, drafting and editing of manuscript.

**Conflict-of-interest statement:** The author reports no relevant conflicts of interest for this article.

**Open-Access:** This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is noncommercial. See: https://creativecommons.org/Licenses/by-nc/4.0/

Country/Territory of origin: United States

ORCID number: Farrukh Ahmad 0000-0003-0004-441X.

S-Editor: Gao CC L-Editor: A P-Editor: Gao CC

### REFERENCES

- 1 Hojyo S, Uchida M, Tanaka K, Hasebe R, Tanaka Y, Murakami M, Hirano T. How COVID-19 induces cytokine storm with high mortality. Inflamm Regen 2020; 40: 37 [PMID: 33014208 DOI: 10.1186/s41232-020-00146-3]
- Delano MJ, Ward PA. The immune system's role in sepsis progression, resolution, and long-term outcome. Immunol Rev 2 2016; 274: 330-353 [PMID: 27782333 DOI: 10.1111/imr.12499]
- Yang SC, Tsai YF, Pan YL, Hwang TL. Understanding the role of neutrophils in acute respiratory distress syndrome. 3 Biomed J 2021; 44: 439-446 [PMID: 33087299 DOI: 10.1016/j.bj.2020.09.001]
- 4 Chaudhry H, Zhou J, Zhong Y, Ali MM, McGuire F, Nagarkatti PS, Nagarkatti M. Role of cytokines as a double-edged sword in sepsis. In Vivo 2013; 27: 669-684 [PMID: 24292568]
- Schulte W, Bernhagen J, Bucala R. Cytokines in sepsis: potent immunoregulators and potential therapeutic targets--an updated view. Mediators Inflamm 2013; 2013: 165974 [PMID: 23853427 DOI: 10.1155/2013/165974]
- Tisoncik JR, Korth MJ, Simmons CP, Farrar J, Martin TR, Katze MG. Into the eye of the cytokine storm. Microbiol Mol Biol Rev 2012; 76: 16-32 [PMID: 22390970 DOI: 10.1128/MMBR.05015-11]



- 7 Caricchio R, Gallucci M, Dass C, Zhang X, Gallucci S, Fleece D, Bromberg M, Criner GJ; Temple University COVID-19 Research Group. Preliminary predictive criteria for COVID-19 cytokine storm. Ann Rheum Dis 2021; 80: 88-95 [PMID: 32978237 DOI: 10.1136/annrheumdis-2020-218323]
- Tay MZ, Poh CM, Rénia L, MacAry PA, Ng LFP. The trinity of COVID-19: immunity, inflammation and intervention. Nat Rev Immunol 2020; 20: 363-374 [PMID: 32346093 DOI: 10.1038/s41577-020-0311-8]
- Huang E, Jordan SC. Tocilizumab for Covid-19 The Ongoing Search for Effective Therapies. N Engl J Med 2020; 383: 9 2387-2388 [PMID: 33296566 DOI: 10.1056/NEJMe2032071]
- 10 Matthay MA, Thompson BT. Dexamethasone in hospitalised patients with COVID-19: addressing uncertainties. Lancet Respir Med 2020; 8: 1170-1172 [PMID: 33129421 DOI: 10.1016/S2213-2600(20)30503-8]
- Zabrodskii PF. [Effect of armin on nonspecific resistance factors of the body and on the primary humoral immune 11 response]. Farmakol Toksikol 1987; 50: 57-60 [PMID: 3549354]
- Bonaz B, Picq C, Sinniger V, Mayol JF, Clarençon D. Vagus nerve stimulation: from epilepsy to the cholinergic anti-12 inflammatory pathway. Neurogastroenterol Motil 2013; 25: 208-221 [PMID: 23360102 DOI: 10.1111/nmo.12076]
- 13 Tracey KJ. The inflammatory reflex. Nature 2002; 420: 853-859 [PMID: 12490958 DOI: 10.1038/nature01321]
- Tracey KJ. Reflex control of immunity. Nat Rev Immunol 2009; 9: 418-428 [PMID: 19461672 DOI: 10.1038/nri2566]
- 15 Pavlov VA, Wang H, Czura CJ, Friedman SG, Tracey KJ. The cholinergic anti-inflammatory pathway: a missing link in neuroimmunomodulation. Mol Med 2003; 9: 125-134 [PMID: 14571320]
- Pavlov VA, Tracey KJ. Neural circuitry and immunity. Immunol Res 2015; 63: 38-57 [PMID: 26512000 DOI: 16 10.1007/s12026-015-8718-1]
- 17 Rosas-Ballina M, Ochani M, Parrish WR, Ochani K, Harris YT, Huston JM, Chavan S, Tracey KJ. Splenic nerve is required for cholinergic antiinflammatory pathway control of TNF in endotoxemia. Proc Natl Acad Sci USA 2008; 105: 11008-11013 [PMID: 18669662 DOI: 10.1073/pnas.0803237105]
- 18 Satoh T, Akira S. Toll-Like Receptor Signaling and Its Inducible Proteins. Microbiol Spectr 2016; 4 [PMID: 28084212 DOI: 10.1128/microbiolspec.MCHD-0040-2016]
- Goehler LE, Gaykema RP, Hansen MK, Anderson K, Maier SF, Watkins LR. Vagal immune-to-brain communication: a 19 visceral chemosensory pathway. Auton Neurosci 2000; 85: 49-59 [PMID: 11189026 DOI: 10.1016/S1566-0702(00)00219-8
- 20 Simons CT, Kulchitsky VA, Sugimoto N, Homer LD, Székely M, Romanovsky AA. Signaling the brain in systemic inflammation: which vagal branch is involved in fever genesis? Am J Physiol 1998; 275: R63-R68 [PMID: 9688961 DOI: 10.1152/ajpregu.1998.275.1.R63]
- Hosoi T, Okuma Y, Matsuda T, Nomura Y. Novel pathway for LPS-induced afferent vagus nerve activation: possible role 21 of nodose ganglion. Auton Neurosci 2005; 120: 104-107 [PMID: 15919243 DOI: 10.1016/j.autneu.2004.11.012]
- Goehler LE, Erisir A, Gaykema RP. Neural-immune interface in the rat area postrema. Neuroscience 2006; 140: 1415-22 1434 [PMID: 16650942 DOI: 10.1016/j.neuroscience.2006.03.048]
- 23 Rosas-Ballina M, Tracey KJ. Cholinergic control of inflammation. J Intern Med 2009; 265: 663-679 [PMID: 19493060 DOI: 10.1111/j.1365-2796.2009.02098.x]
- 24 Huston JM, Ochani M, Rosas-Ballina M, Liao H, Ochani K, Pavlov VA, Gallowitsch-Puerta M, Ashok M, Czura CJ, Foxwell B, Tracey KJ, Ulloa L. Splenectomy inactivates the cholinergic antiinflammatory pathway during lethal endotoxemia and polymicrobial sepsis. J Exp Med 2006; 203: 1623-1628 [PMID: 16785311 DOI: 10.1084/jem.20052362]
- Hoover DB. Cholinergic modulation of the immune system presents new approaches for treating inflammation. Pharmacol 25 *Ther* 2017; **179**: 1-16 [PMID: 28529069 DOI: 10.1016/j.pharmthera.2017.05.002]
- 26 Ren C, Tong YL, Li JC, Lu ZQ, Yao YM. The Protective Effect of Alpha 7 Nicotinic Acetylcholine Receptor Activation on Critical Illness and Its Mechanism. Int J Biol Sci 2017; 13: 46-56 [PMID: 28123345 DOI: 10.7150/ijbs.16404]
- Kanashiro A, Sônego F, Ferreira RG, Castanheira FV, Leite CA, Borges VF, Nascimento DC, Cólon DF, Alves-Filho JC, 27 Ulloa L, Cunha FQ. Therapeutic potential and limitations of cholinergic anti-inflammatory pathway in sepsis. Pharmacol Res 2017; 117: 1-8 [PMID: 27979692 DOI: 10.1016/j.phrs.2016.12.014]
- 28 Koopman FA, Chavan SS, Miljko S, Grazio S, Sokolovic S, Schuurman PR, Mehta AD, Levine YA, Faltys M, Zitnik R, Tracey KJ, Tak PP. Vagus nerve stimulation inhibits cytokine production and attenuates disease severity in rheumatoid arthritis. Proc Natl Acad Sci U S A 2016; 113: 8284-8289 [PMID: 27382171 DOI: 10.1073/pnas.1605635113]
- 29 Genovese MC, Gaylis NB, Sikes D, KivitzA, Horowitz DL, Peterfy C, Glass EV, Levine YA, Chernoff D. Safety and efficacy of neurostimulation with a miniaturised Vagus Nerve stimulation device in patients with multidrug-refractory rheumatoid arthritis: A two-stage multicentre, randomised pilot study. Lancet Rheumatol 2020; 2: E527-E538 [DOI: 10.1016/S2665-9913(20)30172-7
- Borovikova LV, Ivanova S, Zhang M, Yang H, Botchkina GI, Watkins LR, Wang H, Abumrad N, Eaton JW, Tracey KJ. 30 Vagus nerve stimulation attenuates the systemic inflammatory response to endotoxin. Nature 2000; 405: 458-462 [PMID: 10839541 DOI: 10.1038/35013070]
- Kohoutova M, Horak J, Jarkovska D, Martinkova V, Tegl V, Nalos L, Vistejnova L, Benes J, Sviglerova J, Kuncova J, 31 Matejovic M, Stengl M. Vagus Nerve Stimulation Attenuates Multiple Organ Dysfunction in Resuscitated Porcine Progressive Sepsis. Crit Care Med 2019; 47: e461-e469 [PMID: 30908312 DOI: 10.1097/CCM.00000000003714]
- 32 Hilderman M, Bruchfeld A. The cholinergic anti-inflammatory pathway in chronic kidney disease-review and vagus nerve stimulation clinical pilot study. Nephrol Dial Transplant 2020; 35: 1840-1852 [PMID: 33151338 DOI: 10.1093/ndt/gfaa200]
- Wittebole X, Hahm S, Coyle SM, Kumar A, Calvano SE, Lowry SF. Nicotine exposure alters in vivo human responses to 33 endotoxin. Clin Exp Immunol 2007; 147: 28-34 [PMID: 17177960 DOI: 10.1111/j.1365-2249.2006.03248.x]
- 34 Guslandi M. Nicotine treatment for ulcerative colitis. Br J Clin Pharmacol 1999; 48: 481-484 [PMID: 10583016 DOI: 10.1046/j.1365-2125.1999.00039.x
- Guslandi M, Tittobello A. Outcome of ulcerative colitis after treatment with transdermal nicotine. Eur J Gastroenterol 35 Hepatol 1998; 10: 513-515 [PMID: 9855069 DOI: 10.1097/00042737-199806000-00014]
- Özdemir-Kumral ZN, Özbeyli D, Özdemir AF, Karaaslan BM, Kaytaz K, Kara MF, Tok OE, Ercan F, Yegen BÇ. 36



Protective Effect of Nicotine on Sepsis-Induced Oxidative Multiorgan Damage: Role of Neutrophils. Nicotine Tob Res 2017; 19: 859-864 [PMID: 27613897 DOI: 10.1093/ntr/ntw198]

- 37 Mabley J, Gordon S, Pacher P. Nicotine exerts an anti-inflammatory effect in a murine model of acute lung injury. Inflammation 2011; 34: 231-237 [PMID: 20625922 DOI: 10.1007/s10753-010-9228-x]
- 38 Kloc M, Ghobrial RM, Kubiak JZ. How nicotine can inhibit cytokine storm in the lungs and prevent or lessen the severity of COVID-19 infection? Immunol Lett 2020; 224: 28-29 [PMID: 32522666 DOI: 10.1016/j.imlet.2020.06.002]
- Gonzalez-Rubio J, Navarro-Lopez C, Lopez-Najera E, Lopez-Najera A, Jimenez-Diaz L, Navarro-Lopez JD, Najera A. 39 Cytokine Release Syndrome (CRS) and Nicotine in COVID-19 Patients: Trying to Calm the Storm. Front Immunol 2020; 11: 1359 [PMID: 32595653 DOI: 10.3389/fimmu.2020.01359]
- Kox M, Pompe JC, Gordinou de Gouberville MC, van der Hoeven JG, Hoedemaekers CW, Pickkers P. Effects of the  $\alpha$ 7 40 nicotinic acetylcholine receptor agonist GTS-21 on the innate immune response in humans. Shock 2011; 36: 5-11 [PMID: 21368716 DOI: 10.1097/SHK.0b013e3182168d56]
- 41 Garg BK, Loring RH. GTS-21 has cell-specific anti-inflammatory effects independent of a7 nicotinic acetylcholine receptors. PLoS One 2019; 14: e0214942 [PMID: 30947238 DOI: 10.1371/journal.pone.0214942]
- 42 Ahmad F. COVID-19 induced ARDS, and the use of galantamine to activate the cholinergic anti-inflammatory pathway. Med Hypotheses 2020; 145: 110331 [PMID: 33038588 DOI: 10.1016/j.mehy.2020.110331]
- 43 Pinder N, Bruckner T, Lehmann M, Motsch J, Brenner T, Larmann J, Knebel P, Hoppe-Tichy T, Swoboda S, Weigand MA, Hofer S, Zimmermann JB. Effect of physostigmine on recovery from septic shock following intra-abdominal infection - Results from a randomized, double-blind, placebo-controlled, monocentric pilot trial (Anticholium® per Se). J Crit Care 2019; **52**: 126-135 [PMID: 31035187 DOI: 10.1016/j.jcrc.2019.04.012]
- Li G, Zhou CL, Zhou QS, Zou HD. Galantamine protects against lipopolysaccharide-induced acute lung injury in rats. Braz J Med Biol Res 2016; 49: e5008 [PMID: 26648090 DOI: 10.1590/1414-431X20155008]
- 45 Liu ZH, Ma YF, Wu JS, Gan JX, Xu SW, Jiang GY. Effect of cholinesterase inhibitor galanthamine on circulating tumor necrosis factor alpha in rats with lipopolysaccharide-induced peritonitis. Chin Med J (Engl) 2010; 123: 1727-1730 [PMID: 20819637]
- 46 Yoshiyama Y, Kojima A, Ishikawa C, Arai K. Anti-inflammatory action of donepezil ameliorates tau pathology, synaptic loss, and neurodegeneration in a tauopathy mouse model. J Alzheimers Dis 2010; 22: 295-306 [PMID: 20847440 DOI: 10.3233/JAD-2010-1006811
- 47 Consolim-Colombo FM, Sangaleti CT, Costa FO, Morais TL, Lopes HF, Motta JM, Irigoyen MC, Bortoloto LA, Rochitte CE, Harris YT, Satapathy SK, Olofsson PS, Akerman M, Chavan SS, MacKay M, Barnaby DP, Lesser ML, Roth J, Tracey KJ, Pavlov VA. Galantamine alleviates inflammation and insulin resistance in patients with metabolic syndrome in a randomized trial. JCI Insight 2017; 2 [PMID: 28724799 DOI: 10.1172/jci.insight.93340]
- Pavlov VA, Parrish WR, Rosas-Ballina M, Ochani M, Puerta M, Ochani K, Chavan S, Al-Abed Y, Tracey KJ. Brain 48 acetylcholinesterase activity controls systemic cytokine levels through the cholinergic anti-inflammatory pathway. Brain Behav Immun 2009; 23: 41-45 [PMID: 18639629 DOI: 10.1016/j.bbi.2008.06.011]
- Musk AW, de Klerk NH. History of tobacco and health. Respirology 2003; 8: 286-290 [PMID: 14528877 DOI: 49 10.1046/j.1440-1843.2003.00483.x]
- 50 DeVeaugh-Geiss AM, Chen LH, Kotler ML, Ramsay LR, Durcan MJ. Pharmacokinetic comparison of two nicotine transdermal systems, a 21-mg/24-hour patch and a 25-mg/16-hour patch: a randomized, open-label, single-dose, two-way crossover study in adult smokers. Clin Ther 2010; 32: 1140-1148 [PMID: 20637967 DOI: 10.1016/j.clinthera.2010.06.008]
- 51 Moran VE. Cotinine: Beyond that Expected, More than a Biomarker of Tobacco Consumption. Front Pharmacol 2012; 3: 173 [PMID: 23087643 DOI: 10.3389/fphar.2012.00173]
- Hukkanen J, Jacob P 3rd, Peng M, Dempsey D, Benowitz NL. Effect of nicotine on cytochrome P450 1A2 activity. Br J 52 Clin Pharmacol 2011; 72: 836-838 [PMID: 21599724 DOI: 10.1111/j.1365-2125.2011.04023.x]
- Stefan MS, Pack Q, Shieh MS, Pekow PS, Bernstein SL, Raghunathan K, Nason KS, Lindenauer PK. The Association of 53 Nicotine Replacement Therapy With Outcomes Among Smokers Hospitalized for a Major Surgical Procedure. Chest 2020; 157: 1354-1361 [PMID: 31790653 DOI: 10.1016/j.chest.2019.10.054]
- 54 de Jong B, Schuppers AS, Kruisdijk-Gerritsen A, Arbouw MEL, van den Oever HLA, van Zanten ARH. The safety and efficacy of nicotine replacement therapy in the intensive care unit: a randomised controlled pilot study. Ann Intensive Care 2018; 8: 70 [PMID: 29881956 DOI: 10.1186/s13613-018-0399-1]
- 55 Pathak V, Rendon IS, Lupu R, Tactuk N, Olutade T, Durham C, Stumacher R. Outcome of nicotine replacement therapy in patients admitted to ICU: a randomized controlled double-blind prospective pilot study. Respir Care 2013; 58: 1625-1629 [PMID: 23533293 DOI: 10.4187/respcare.01791]
- 56 Cartin-Ceba R, Warner DO, Hays JT, Afessa B. Nicotine replacement therapy in critically ill patients: a prospective observational cohort study. Crit Care Med 2011; 39: 1635-1640 [PMID: 21494111 DOI: 10.1097/CCM.0b013e31821867b8]
- Pack QR, Priya A, Lagu TC, Pekow PS, Atreya A, Rigotti NA, Lindenauer PK. Short-Term Safety of Nicotine 57 Replacement in Smokers Hospitalized With Coronary Heart Disease. J Am Heart Assoc 2018; 7: e009424 [PMID: 30371184 DOI: 10.1161/JAHA.118.009424]
- Ng KT, Gillies M, Griffith DM. Effect of nicotine replacement therapy on mortality, delirium, and duration of therapy in 58 critically ill smokers: a systematic review and meta-analysis. Anaesth Intensive Care 2017; 45: 556-561 [PMID: 28911284 DOI: 10.1177/0310057X1704500505]
- Molander L, Hansson A, Lunell E, Alainentalo L, Hoffmann M, Larsson R. Pharmacokinetics of nicotine in kidney failure. Clin Pharmacol Ther 2000; 68: 250-260 [PMID: 11014406 DOI: 10.1067/mcp.2000.109006]
- 60 Perry RJ, Griffiths W, Dextraze P, Solomon RJ, Trebbin WM. Elevated nicotine levels in patients undergoing hemodialysis. A role in cardiovascular mortality and morbidity? Am J Med 1984; 76: 241-246 [PMID: 6695947 DOI: 10.1016/0002-9343(84)90780-0]

WJCCM | https://www.wjgnet.com



## Published by Baishideng Publishing Group Inc 7041 Koll Center Parkway, Suite 160, Pleasanton, CA 94566, USA Telephone: +1-925-3991568 E-mail: bpgoffice@wjgnet.com Help Desk: https://www.f6publishing.com/helpdesk https://www.wjgnet.com

