

Prospective Study

Helicobacter pylori and serum kynurenine-tryptophan ratio in patients with colorectal cancer

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Abstract

AIM: To evaluate how *Helicobacter pylori* (*H. pylori*) is able to evade the immune response and whether it enhances systemic immune tolerance against colorectal cancer.

METHODS: This prospective randomized study involved 97 consecutive colorectal cancer patients and 108 cancer-free patients with extra-digestive diseases. Colorectal cancer and cancer-free patients were assigned into subgroups according to *H. pylori* IgG seropositivity. Exposure to *H. pylori* was determined by IgG seropositivity which was detected by enzyme linked immunoassay (ELISA). Serum neopterin levels were measured by ELISA. Serum tryptophan, kynurenine, and urinary biopterin concentrations were measured by high performance liquid chromatography. Serum nitrite levels were detected spectrophotometrically. Serum indoleamine 2,3-dioxygenase activity was estimated by the kynurenine to tryptophan ratio and by assessing the correlation between serum neopterin concentrations and the kynurenine to tryptophan ratio. The frequencies of increased serum kynurenine to tryptophan ratio of *H. pylori* seronegative and seropositive colorectal cancer subgroups were estimated by comparing them with the average kynurenine to tryptophan ratio of *H. pylori* seronegative tumor-free patients.

RESULTS: Compared with respective controls, in both *H. pylori* seronegative and seropositive colorectal cancer patients, while serum tryptophan levels were decreased (controls vs patients; seronegative: 20.37 ± 0.89 $\mu\text{mol/L}$ vs 15.71 ± 1.16 $\mu\text{mol/L}$, $P < 0.05$; seropositive: 20.71 ± 0.81 $\mu\text{mol/L}$ vs 14.97 ± 0.79 $\mu\text{mol/L}$, $P < 0.01$) the kynurenine to tryptophan ratio was significantly increased (controls vs patients; seronegative: 52.85

$\pm 11.85 \mu\text{mol}/\text{mmol}$ vs $78.91 \pm 8.68 \mu\text{mol}/\text{mmol}$, $P < 0.01$, seropositive: $47.31 \pm 5.93 \mu\text{mol}/\text{mmol}$ vs $109.65 \pm 11.50 \mu\text{mol}/\text{mmol}$, $P < 0.01$). Neopterin concentrations in cancer patients were significantly elevated compared with controls ($P < 0.05$). There was a significant correlation between serum neopterin levels and kynurenine/tryptophan in control and colorectal cancer patients groups ($r_s = 0.494$, $P = 0.0001$ and $r_s = 0.293$, $P = 0.004$, respectively). Serum nitrite levels of *H. pylori* seropositive cancer cases were significantly decreased compared with seropositive controls (controls vs patients; $26.04 \pm 2.39 \mu\text{mol}/\text{L}$ vs $20.41 \pm 1.48 \mu\text{mol}/\text{L}$, $P < 0.05$). The decrease in the nitrite levels of *H. pylori* seropositive cancer patients may be attributed to excessive formation of peroxynitrite and other reactive nitrogen species.

CONCLUSION: A significantly high kynurenine/tryptophan suggested that *H. pylori* may support the immune tolerance leading to cancer development, even without an apparent upper gastrointestinal tract disease.

Key words: Colorectal cancer; Kynurenine/tryptophan; Immune tolerance; *Helicobacter pylori*; Oxidative stress

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Core tip: Persistent inflammation of the stomach induced by *Helicobacter pylori* (*H. pylori*) can have consequences on the rest of the body. Despite the vigorous innate and adaptive immune response against the bacterium, *H. pylori* escape and evade host responses by a variety of mechanisms. Low tryptophan levels and increased concentrations of its degradation product, kynurenine, may be directly involved in diminished T-cell responsiveness to antigenic stimulation in cancer. *H. pylori* seropositive colorectal cancer patients with significantly higher kynurenine/tryptophan and reduced nitric oxide suggested that *H. pylori* might support immune tolerance leading to cancer development, even in patients without an apparent upper gastrointestinal tract disease.

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INTRODUCTION

Persistent inflammation of the stomach induced by *Helicobacter pylori* (*H. pylori*) can have consequences on the rest of the body. Recent studies showed that the immunological response against *H. pylori* is not only

a locally but also a systemically evoked phenomena in the host. Particularly in the last few years, many studies have been performed on the role of *H. pylori* in the pathogenesis of extra-gastric diseases^[1]. It is now known that exposure to *H. pylori* and subsequent seropositivity is associated with an increased number of cardiovascular, respiratory, extra-gastrointestinal digestive, neurological and miscellaneous autoimmune disorders^[2].

It has previously been shown that the risk of colon adenomas is increased in *H. pylori*-infected subjects. Two recent studies from Japan, based on a large number of patients, added important evidence for the association between *H. pylori* infection and colorectal neoplasia. Previously, Fujimori *et al.*^[3] showed a positive relationship between *H. pylori* infection and the risk of adenoma and carcinoma, especially in women [odds ratio (OR); 1.68 and 2.09, respectively]. Later, Mizuno *et al.*^[4] found that *H. pylori* infection was associated with the presence of colon adenomatous polyps. A very significant increase in the incidence of adenomas was observed in the seropositive group compared with seronegative controls (44.3% vs 18.9%, $P < 0.0001$).

In the evaluation of the relationship between *H. pylori* and risk of colorectal cancer, the estimated OR showed a small increase in the risk of colorectal cancer development because of *H. pylori* infection. Recently, in one of two different meta-analyses, an OR of 1.49 (95%CI: 1.17-1.91) was found for the association between *H. pylori* infection and colorectal cancer. In another study, serological investigation demonstrated an OR of 1.56 (95%CI: 1.14-2.14) for the association between immunoglobulin G (IgG) antibody and colorectal cancer risk^[5,6]. Likewise, in our previous study, we found a 2.2-fold increase in the risk of colorectal carcinoma in patients with *H. pylori* IgG seropositivity^[7]. Very recently, a significant correlation was found by Popović and colleagues between *H. pylori* seropositivity and colon cancer ($P = 0.002$) in a series of 142 patients^[8].

Although the human host mounts a vigorous innate and adaptive immune response against the bacterium, *H. pylori* can escape and evade host responses by a variety of mechanisms that lead to persistent colonization and chronic active inflammation^[9]. Surprisingly, most people infected with *H. pylori* are asymptomatic, which suggests that additional factors are necessary for the development of *H. pylori*-associated diseases^[10].

Thomas and Stocker proposed a sequential defense mechanism for the immune response, in which indoleamine 2,3-dioxygenase (IDO) activity is the first-line of defense against invading cells^[11]. Interferon-gamma (IFN- γ)-induced IDO activity mediates an antimicrobial effect. During the first phase of infection, IDO-mediated tryptophan (Trp) depletion is predominantly antimicrobial whereas in the further stage, it is an inhibitor of T-cell growth^[12].

Despite the phenomenon of immune activation against cancer cells, low Trp levels and increased concentrations of its degradation products may be directly involved in diminished T-cell responsiveness to antigenic stimulation in cancer patients^[13]. This new mechanism proposed for tumoral immune resistance involves the expression of IDO by tumor cells. It rapidly degrades Trp, and resultant Trp depletion causes a strong inhibitory effect on the development of immune responses^[14].

These evidences raise the question of how *H. pylori* is consistently able to evade these cellular and humoral immune responses. Furthermore, it is not known whether *H. pylori* enhance systemic immune tolerance against colorectal cancer or not. Regarding the mentioned assumptions, the present study was designed to determine the effects of the serum kynurenine (Kyn)/Trp, serum Trp, Kyn and neopterin levels, which are sustained in persisting *H. pylori* seropositivity in colorectal cancer patients.

MATERIALS AND METHODS

This prospective randomized study involved 97 consecutive colorectal cancer patients; 61.7 ± 1.3 (mean \pm SE) years of age, and body mass index (BMI) of 25.5 ± 0.4 kg/m², and 108 cancer-free patients with extra-digestive diseases; 55.5 ± 1.3 years of age, and BMI of 27.1 ± 0.5 kg/m², referred to Gazi University, Faculty of Medicine, Department of General Surgery for surgical evaluation. Diagnosis was made by colonoscopy and histological examination of tumor biopsies in all colorectal cancer patients, and stratified according to the TNM classification of the American Joint Committee on Cancer Staging. From laboratory findings and clinical staging, the primary disease of all cancer patients was found to be suitable for surgical intervention. Subsequent confirmation of the preoperative diagnosis was made by routine histopathological examination of postoperative specimens regarding the presence of lymphatic invasion, lymph node involvement, peritumoral lymphoid cell infiltration, and tumor grade. However, these histological findings were ignored in our series of patients because of the high seroprevalence of *H. pylori* in the general population and prevalent asymptomatic infection makes the interpretation of the definite role of *H. pylori* difficult. Therefore, cancer patients and cancer-free cases were divided simply into two subgroups, based on the presence or absence of *H. pylori* IgG seropositivity. Exposure to *H. pylori* in each patient was determined by an IgG seropositivity test (ELISA) according to the manufacturer's instructions (*H. pylori* IgG ELISA, Demeditec, Germany). Considering the ELISA kit manual, individuals with an *H. pylori* IgG titer below 8 U/mL were accepted as *H. pylori* seronegative, while a value above 12 U/mL was interpreted as *H. pylori*

seropositive. The patients whose titers were between 8 and 12 U/mL appeared in the grey zone and were not included in the study. The 13C-urea breath test (UBT) is an accurate, non-invasive test to diagnose gastric viable *H. pylori* colonization in adults and it is also used to monitor the outcome of eradication therapy in patients^[15]. However, findings suggest that infections with *H. pylori* may have a long-lasting impact on the cell-mediated immune system even after viable bacteria are eradicated. Therefore, we did not perform UBT in our series of patients but determined *H. pylori* IgG seropositivity in our study. There were 37 *H. pylori* IgG seronegative and 71 *H. pylori* IgG seropositive control patients, while 19 of the colorectal cancer patients were *H. pylori* IgG seronegative and 78 were *H. pylori* IgG seropositive.

Cancer-free individuals were selected from the patient population admitted to the hospital for hernioraphy, hemorrhoidectomy or breast biopsies. These patients had undergone diagnostic endoscopic evaluation whenever their complaints were suggestive of digestive diseases. Patients who did not have any pathological finding in either the upper or lower gastrointestinal system were included in the cancer-free control group. No patient had cardio-pulmonary or metabolic risks that could be an obstacle for surgery.

The exclusion criteria were immune system disorders, not able to receive surgical intervention or treatment with neoadjuvant chemotherapy because of late stage carcinoma, or having malnutrition, autoimmune diseases, systemic inflammatory response syndrome, chronic granulomatosis, collagen tissue or neurodegenerative diseases. None of the patients who received traditional triple eradication therapy for *H. pylori* infection within the last two years were included in the study groups.

All participants' rights were protected and informed consents were obtained according to the Helsinki Declaration. Gazi University, Local Ethics Committee approved the study protocol.

Peripheral venous blood samples from each individual was collected and used for serum separation. Urine specimens were collected coincidentally. All samples were obtained in the early morning, and kept from direct light at -20 °C until assay.

Biopterin and creatinine levels in urine were analyzed by high performance liquid chromatography (HPLC), as previously described^[16]. Serum neopterin concentrations were determined according to the manufacturer's instructions by a commercially available enzyme immunoassay kit (ELISA, Tani Medical Laboratories, Ankara, Turkey). The optical density was measured at 450 nm. Trp and Kyn concentrations in serum were determined simultaneously by reversed-phase HPLC. In order to estimate Trp degradation, the Kyn to Trp ratio (Kyn/Trp) was calculated by dividing Kyn concentrations (μ mol/L) by Trp concentrations (mmol/L)^[17]. The frequency of increased serum Kyn/Trp of *H. pylori* seronegative and seropositive colorectal

Table 1 Comparison of neopterin, tryptophan, kynurenine, nitrite, biopterin levels (mean \pm SE) in controls without malignancy and in gastric cancer patients

	Control group (n = 108)	Colorectal cancer group (n = 97)	P value
Serum neopterin (nmol/L)	7.68 \pm 0.56	21.18 \pm 2.45	0.021 ^a
Tryptophan (nmol/L)	20.6 \pm 0.61	15.38 \pm 0.64	0.000 ^a
Kynurenine (nmol/L)	0.89 \pm 0.09	1.24 \pm 0.09	0.000 ^a
Kynurenine/tryptophan (μ mol/mmol)	49.10 \pm 5.52	99.93 \pm 8.70	0.000 ^a
Nitrite (μ mol/L)	24.60 \pm 1.69	23.05 \pm 1.74	0.107
Urinary biopterin/ creatinine (μ mol/mol)	121.65 \pm 7.79	122.70 \pm 7.69	0.926

^aP < 0.05, control vs cancer patients.**Table 2** Comparison of neopterin, tryptophan, kynurenine, nitrite, biopterin levels (mean \pm SE) in *Helicobacter pylori* seronegative controls and *Helicobacter pylori* seronegative colorectal cancer patients

	<i>H. pylori</i> seronegative control group (n = 37)	<i>H. pylori</i> seronegative colorectal cancer group (n = 19)	P value
Serum neopterin (nmol/L)	8.10 \pm 1.11	20.89 \pm 6.27	0.010 ^a
Tryptophan (nmol/L)	20.37 \pm 0.89	15.71 \pm 1.16	0.003 ^a
Kynurenine (nmol/L)	0.89 \pm 0.12	1.22 \pm 0.18	0.007 ^a
Kynurenine/tryptophan (mmol/mmol)	52.85 \pm 11.85	78.91 \pm 8.68	0.000 ^a
Nitrite (mmol/L)	21.86 \pm 1.81	25.51 \pm 4.34	0.938
Urinary biopterin/ creatinine (mmol/mol)	116.20 \pm 12.87	119.1 \pm 11.6	0.573

^aP < 0.05, *Helicobacter pylori* (*H. pylori*) seronegative control vs *H. pylori* seronegative cancer patients.

cancer groups was estimated by comparing each of them with the mean of seronegative cancer-free controls. Serum nitrite concentrations were measured by the Griess method^[18]. Serum nitrate measurements were not included in this study, because nitrates are not only released as the final products of nitric oxide (NO) oxidation via nitrites, but could also be produced from peroxynitrites formed during the reaction of NO with the free oxygen radicals. A conclusive profile of NO concentration is reflected by serum nitrite level^[19]. Measurement of serum creatinine levels was performed by an auto analyzer.

Statistical analysis

Data were analyzed using SPSS, version 13.0 (SPSS Inc., Chicago, IL, United States). All results are expressed as mean \pm SE. After checking the data with the Kolmogorov-Smirnov test, non-parametric data of two independent groups were compared with the Mann-Whitney *U* test and *P* < 0.05 was considered statistically significant. Correlations were assessed using the Spearman rank test.

Table 3 Comparison of neopterin, tryptophan, kynurenine, nitrite, biopterin levels (mean \pm SE) in *Helicobacter pylori* seropositive controls and *Helicobacter pylori* seropositive colorectal cancer patients

	<i>H. pylori</i> seropositive control group (n = 71)	<i>H. pylori</i> seropositive colorectal cancer group (n = 78)	P value
Serum neopterin (nmol/L)	7.44 \pm 0.61	21.8 \pm 3.05	0.0001 ^a
Tryptophan (nmol/L)	20.71 \pm 0.81	14.97 \pm 0.79	0.0001 ^a
Kynurenine (nmol/L)	0.89 \pm 0.13	1.28 \pm 0.11	0.0001 ^a
Kynurenine/tryptophan (μ mol/mmol)	47.31 \pm 5.93	109.65 \pm 11.50	0.0001 ^a
Nitrite (μ mol/L)	26.04 \pm 2.39	20.41 \pm 1.48	0.017 ^a
Urinary biopterin/ creatinine (μ mol/mol)	124.65 \pm 9.84	120.69 \pm 9.98	0.607

^aP < 0.05, *Helicobacter pylori* (*H. pylori*) seropositive control vs *H. pylori* seropositive cancer patients.

RESULTS

Mean serum neopterin levels of the cancer patients were significantly lower than in the non-tumor group (Table 1). In the evaluation of subgroups, there was a significant increase in serum neopterin in both *H. pylori* seronegative (Table 2) and seropositive (Table 3) cancer patients (*u* = 202, *P* = 0.01 and *u* = 1265, *P* = 0.0001, respectively) compared with their matched controls. However, the highly significant increase in serum neopterin levels in *H. pylori* seropositive cancer patients suggested that *H. pylori* seropositivity enhanced the immune response of macrophages against colorectal cancer. Considering all colorectal cancer patients, although the reduction in serum nitrite concentrations was not significant (*u* = 5538.5, *P* = 0.107) (Table 1), serum nitrite levels of *H. pylori* seropositive cancer patients were significantly lower compared with *H. pylori* seropositive cancer-free matched controls (2110, *P* = 0.01) (Table 3). Serum nitrite levels did not differ between the *H. pylori* seronegative cancer group and their matched controls (*u* = 347, *P* = 0.938) (Table 2). These findings may be consistent with the evidence that the released reactive oxygen and nitrogen species are produced by the phagocytic leukocytes which are recruited to the colorectal tumor site and by *H. pylori* itself. However, it was not investigated whether *H. pylori* colonization is still positive or not in 78 seropositive cases with colorectal cancer. Unchanged urinary biopterin, oxidized product of tetrahydrobiopterine (H4-bip) in *H. pylori* seropositive colorectal cancer patients (1385.5, *P* = 0.607) (Table 3) showed that despite the increased consumption of NO, supportive NO synthesis in these patients was not evident.

Trp concentrations in cancer patients were significantly lower than that of cancer-free controls (*u* = 2876, *P* = 0.0001) (Table 1). As a result of increased

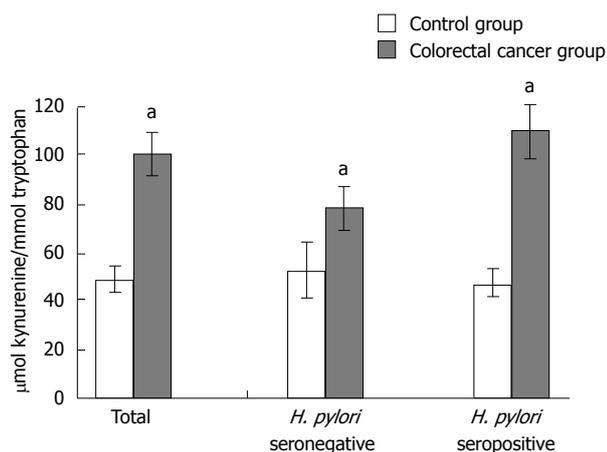


Figure 1 Comparison between kynurenine to tryptophan ratios in controls and colorectal cancer patients, in *Helicobacter pylori* seronegative or seropositive groups. ^a $P < 0.05$, controls vs colorectal cancer patients, *Helicobacter pylori* (*H. pylori*) seronegative controls vs *H. pylori* seronegative colorectal cancer patients, *H. pylori* seropositive controls vs *H. pylori* seropositive colorectal cancer patients, statistically significant.

IDO activity, levels of Kyn, the toxic product of the Trp degradation pathway, significantly increased in colorectal cancer cases ($u = 2931$, $P = 0.0001$). Thus, colorectal cancer patients showed a highly significant rise in Kyn/Trp ($u = 2272$, $P = 0.0001$) (Table 1, Figure 1). To elucidate the contribution of *H. pylori* seropositivity to the frequency of increased Kyn/Trp, the data of *H. pylori* seronegative or seropositive subgroups were evaluated. *H. pylori* seronegative or seropositive colorectal cancer patients had significantly higher Kyn/Trp compared with their matched cancer-free controls. Although no statistical difference was found between IDO activities of *H. pylori* seronegative and seropositive cancer patients, the frequency of increased Kyn/Trp was estimated as 46% and 57% for seronegative and seropositive subgroups, respectively. This means that *H. pylori* seropositivity might add 11% to the frequency of increased Kyn/Trp in the colorectal cancer group and enhance immune tolerance against cancer cells. There was a significant correlation between the serum neopterin levels and Kyn/Trp in control and colorectal cancer patients groups ($r_s = 0.494$, $P = 0.0001$ and $r_s = 0.293$, $P = 0.004$, respectively), while a positive correlation existed in both *H. pylori* seronegative and seropositive control individuals ($r_s = 0.652$, $P = 0.0001$ and $r_s = 0.381$, $P = 0.002$, respectively). Similarly, the correlation of the same parameters in *H. pylori* seropositive cancer patients was also significant ($r_s = 0.374$, $P = 0.002$, respectively).

DISCUSSION

Neopterin is mainly synthesized by activated monocytes/macrophages in response to induction by IFN- γ . Measurement of neopterin concentrations in body fluids provides information about T helper

cell 1 (Th1)-derived cellular immune activation^[20]. An increase in neopterin concentrations during cancer growth indicates a chronic cellular immune response; however, it is not specific for malignant cell proliferation^[21]. The mean serum neopterin level in the non-tumor group was below the standard cut-off value, 10 nmol/L^[22], while there was a significant increase in serum neopterin in cancer patients. Thus in our study, a highly significant increase in serum neopterin concentration of *H. pylori* seropositive colorectal cancer patients suggested that *H. pylori* seropositivity induced activation of cell-mediated immunity, in addition to cancer-induced chronic cellular immune response.

On the other hand, chronic stimulation of Th1-mediated immunity may also cause enhanced IDO activity in malignant diseases^[23]. IDO is an enzyme ubiquitously distributed in mammalian cells, and converts Trp to N-formylkynurenine. This substance is further catabolized to Kyn^[13]. Trp depletion as well as the accumulation of its metabolites results in a strong inhibitory effect on the development of immune responses^[24]. IDO-induced Trp depletion from the tumor microenvironment could be the result of enhanced activation of the enzyme and augmented Trp consumption by both tumor cells and antigen-presenting cells of the host^[25]. Recent data obtained from tumor models demonstrated that IDO inhibition could significantly enhance the antitumor activity of various chemotherapeutic and immunotherapeutic agents. These results were consistent with data showing that increased IDO expression was an independent prognostic variable for reduced overall survival in cancer patients^[24]. In colorectal cancer patients, significantly accelerated degradation of Trp, with lowered serum concentrations of Trp and increased Kyn, as well as an increased Kyn/Trp has previously been reported^[26]. Indeed, in our study, the highly significant correlation between neopterin concentrations with increased Kyn/Trp clearly indicated that the formation of Kyn is related to IDO activity by IFN- γ stimulation. It is postulated that IDO limits immune cell proliferation by depleting locally available Trp and/or producing its cytotoxic metabolites^[27]. However, enhanced IDO activity and further serum Trp degradation due to *H. pylori*-seropositivity in colorectal cancer patients was demonstrated for the first time in our study. In addition to reduced serum Trp concentrations and raised serum Kyn levels in the *H. pylori* seropositive colorectal cancer group, a significant increase in serum Kyn/Trp ($u = 1026$, $P = 0.0001$) and highly significant correlation between the serum neopterin and Kyn/Trp suggested that IDO activities may be induced by *H. pylori* seropositivity. A frequency of increased Kyn/Trp of 57% indicated that enhanced IDO activity may be an important additional factor in the development of *H. pylori*-associated colorectal cancer. As a result, *H. pylori* seropositivity may have an effect on the enhancement of immune tolerance against cancer cells. The number of advanced cancer

cases in the *H. pylori* seropositive group may support this statement.

In our study, while serum nitrite levels were significantly lower in *H. pylori* seropositive cancer patients, no difference was found between *H. pylori* seronegative cancer patients and their *H. pylori* seronegative matched controls. Thus, the nitrite-NO pathway may be viewed as a backup system, to ensure sufficient NO generation along the entire physiological oxygen gradient^[28]. Actually, NO regulates IDO activity biphasically, in a dose-dependent manner such that, while high NO production inactivates IDO enzyme and favors the immune response, low concentrations of NO increase IDO activity, resulting in immune tolerance^[29]. A decrease in serum NO in *H. pylori* seropositive cancer patients may be attributed to the relatively effective scavenging of the radicals by NO. NO and superoxide can antagonize each other's biological actions regarding the NO/superoxide balance in cytoplasmic fractions^[30]. The relative NO production rates have an impact on the NO-mediated toxic vs protective effects^[31]. Peroxynitrite formed *in vivo* from superoxide and NO can mediate oxidative nitration or nitrosation reactions, leading to tissue injury. Consequently, a reduction in NO may be due to the formation of peroxynitrite and other reactive nitrogen species (RNS). It was demonstrated that excessive reactive oxygen species (ROS)/RNS production in the *H. pylori*-infected stomach by activated neutrophils and *H. pylori* itself can damage DNA in gastric epithelial cells, implying its involvement in gastric carcinogenesis^[32]. A negative correlation between the amount of superoxide radicals and nitrites suggests that NO has antioxidative effects at the site of injury^[33]. However, the vast majority of the studies have employed serological surveillance rather than isolation of *H. pylori* from the target disease site, as serological testing for indirectly detecting *H. pylori* is quick, relatively cheap and specific. Moreover, it is well known that there is an ongoing risk of developing cancer even after the eradication of *H. pylori* infection^[34]. These data claimed that *H. pylori* may induce oxidative stress by different mechanisms. Most probably NO combines with reactive oxygen metabolites to form RNS, such as nitrogen dioxide and peroxynitrite, whereby NO bioavailability was decreased.

As we could not find any difference between the serum nitrite levels of *H. pylori* seronegative colorectal cancer cases and their matched controls; it seems that the presence of *H. pylori* may have caused a significant increase in NO consumption by formation of redox active radicals in cancer cases. H4-bip is an indispensable cofactor for NO generation by inducible nitric oxide synthase (iNOS)^[35]. The quantity of urinary biopterin excretion is a determinant of the amount of intracellular H4-bip which is critical for iNOS-dependent generation of NO^[36]. However, we did not observe an increase in urinary biopterin excretion of *H. pylori* seropositive cancer group.

In our study, a significant increase in serum neopterin concentrations suggested that IFN- γ -induced guanosine triphosphate (GTP) cyclohydrolase I activities largely supported neopterin synthesis in either *H. pylori* seronegative or *H. pylori* seropositive colorectal carcinoma patients. GTP cyclohydrolase I activities correlate with the sum of neopterin plus biopterin rather than with neopterin or biopterin alone^[37]. As a conclusion, *H. pylori* seropositive colorectal cancer patients with significantly higher Kyn/Trp and reduced NO suggest that *H. pylori* may support immune tolerance leading to cancer development, even in patients without an apparent upper gastrointestinal tract disease. There is little in the literature related to this subject, thus further studies in larger populations are warranted in order to support these findings.

COMMENTS

Background

Persistent inflammation of the stomach induced by *Helicobacter pylori* (*H. pylori*) can have consequences on the rest of the body. Despite the vigorous innate and adaptive immune response against the bacteria, it is still not known how *H. pylori* escape and evade host responses. Low tryptophan levels and increased concentrations of its degradation product, kynurenine, may be directly involved in diminished T-cell responsiveness to antigenic stimulation in cancer.

Research frontiers

An increased kynurenine to tryptophan ratio that is correlated with neopterin concentrations may be directly involved in T-cell unresponsiveness to cancer cells. The present study was designed to determine the effects of serum kynurenine/tryptophan, serum tryptophan, kynurenine and neopterin levels with persistent *H. pylori* seropositivity in colorectal cancer patients.

Innovations and breakthroughs

H. pylori seropositive colorectal cancer patients with significantly higher kynurenine/tryptophan and reduced nitric oxide suggested that *H. pylori* may support immune tolerance leading to cancer development, even in patients without an apparent upper gastrointestinal tract disease.

Applications

H. pylori seropositivity may have consequences not only in the stomach but also in the colon and rectum, and this may lead to the increased incidence of colorectal cancer. Significantly higher kynurenine/tryptophan and reduced nitric oxide may be an indicator of immune tolerance related to *H. pylori* seropositivity that supports colorectal cancer development.

Terminology

The presence of *H. pylori* immunoglobulin G is associated with previous infection. Indolamine-2,3-dioxygenase is an enzyme, ubiquitously distributed in mammalian cells, which converts tryptophan into kynurenine. The correlation of kynurenine/tryptophan with neopterin indicates indolamine-2,3-dioxygenase activity. Thus, tryptophan depletion as well as the accumulation of its metabolites results in a strong inhibitory effect on the development of immune responses against cancer cells.

Peer-review

This is a good descriptive study in which the authors analyzed consequences of *H. pylori* on the body other than stomach. The results are interesting and suggest that increased kynurenine/tryptophan- may be directly involved in diminished T-cell responsiveness to antigenic stimulation in cancer and *H. pylori* might support immune tolerance leading to cancer development, even in patients without an apparent upper gastrointestinal tract disease.

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