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***Basic Study***

**Contrast-enhanced ultrasound for quantitative assessment of portal pressure in canine liver fibrosis**

Zhai L *et al*. Portal pressure in canine liver fibrosis

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**Abstract**

**AIM:** To explore the feasibility of noninvasive quantitative estimation of portal venous pressure by contrast-enhanced ultrasound (CEUS) in a canine model.

**METHODS:** Liver fibrosis was established in adult canines (Beagles; *n* = 14) by subcutaneous injection of carbon tetrachloride (CCl4). CEUS parameters including the area under the time-intensity curve and intensity at portal/arterial phases (Qp/Qa and Ip/Ia, respectively) were used to quantitatively assess the blood flow ratio of the portal vein/hepatic artery at multiple time points. The free portal venous pressures (FPP) were measured by a multi-channel baroreceptor using a percutaneous approach at baseline and 8, 16, and 24 wk after CCl4 injections in each canine. Liver biopsies were obtained at the end of 8, 16, and 24 wk from each animal, and the stage of the fibrosis was assessed according to the Metavir scoring system. A Pearson correlation test was performed to compare the FPP with Qp/Qa and Ip/Ia.

**RESULTS:** Pathologic examination of 42 biopsies from the 14 canines at weeks 8, 16, and 24 revealed that liver fibrosis was induced by CCl4, and furthermore, represented various stages of liver fibrosis including F0 (*n* = 3), F1 (*n* = 12), F2 (*n* = 14), F3 (*n* = 11), and F4 (*n* = 2). There were significant differences in the measurements of Qp/Qa (19.85 ± 3.30 *vs* 10.43 ± 1.21, 9.63 ± 1.03, and 8.77 ± 0.96) and Ip/Ia (1.77 ± 0.37 *vs* 1.03 ± 0.12, 0.83 ± 0.10, and 0.69 ± 0.13) between control and canine fibrosis at 8, 16, and 24 wk, respectively (all *P* < 0.001). There were statistically significant negative correlations between FPP and Qp/Qa (*r* = -0.707, *P* < 0.001), and between FPP and Ip/Ia (*r* = -0.759, *P* < 0.001) in the canine fibrosis model. Prediction of elevated FPP based on Qp/Qa and Ip/Ia was highly sensitive as assessed by the area under the receiver operating curve (0.866 and 0.895, respectively).

**CONCLUSION:** CEUS is a potential method to accurately but noninvasively estimate the portal venous pressure through measurement of Qp/Qa and Ip/Ia parameters.

**Key words:** Animal model; Contrast-enhanced ultrasound; Liver fibrosis; Noninvasive technique; Portal venous pressure

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**Core tip:** The measurement of portal pressure plays an important role in the evaluation of the progression of liver disease. Contrast-enhanced ultrasound techniques can provide hemodynamic parameters of blood circulation and tissue perfusion. The introduction of the area under the time-intensity curve and intensity at portal/arterial phases parameters has obvious advantages over absolute values, such as peak intensity, as the approach reduces other sources of interference. These parameters were negatively correlated with free portal pressure, supporting the use of contrast-enhanced ultrasound as a method to noninvasively estimate portal pressure.

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**INTRODUCTION**

Portal hypertension is a severe complication of chronic liver disease and cirrhosis, which causes many clinical abnormalities due to hemodynamic changes in portal pressure. The measurement of the portal venous pressure thus plays an important role in the evaluation of the progression of liver disease[1].

There are two reliable methods for accurately evaluating portal pressure: hepatic venous pressure gradient (HVPG) and free portal pressure (FPP) measurements[2]. However, both methods are performed under invasive procedures and cannot be routinely used for assessing and monitoring the progression of chronic liver disease. Therefore, a noninvasive and more reliable method to quantitatively evaluate portal pressure would present distinct advantages for patient evaluation over time.

Conventional grey scale ultrasound is the first-line imaging modality in the screening of portal hypertension[3–5]. The enlargement of the portal vein is a simple indicator for a clinical diagnosis of portal hypertension. Doppler ultrasound can provide valuable parameters for the evaluation of portal hypertension, such as velocity of the portal blood flow, direction of the portal flow, the hepatic vein waveforms, the pulsatility index, and the resistance index of hepatic and splenic arteries. However, all of these parameters have a poor linear relationship with portal venous pressure. Thus, the lack of reliability and reproducibility of the ultrasound technique limits its clinical utility for the assessment of the status of portal hypertension.

Newly developed ultrasound contrast agents and contrast-enhanced ultrasound (CEUS) techniques now provide hemodynamic parameters regarding blood circulation and tissue perfusion. As microbubble-based ultrasound contrast agent is neither discharged from the capillaries nor diffused into the interstitium, the signal intensity of CEUS is related to the concentration of the agent within the blood vessel. Investigators have found that several CEUS features, such as the arrival time of the contrast agent to the hepatic vein, and the quantitative analysis of the enhancement level of the liver parenchyma, may be useful for indirect assessment of liver cirrhosis and portal hypertension[6–9].

To validate a new imaging technique for the measurement of portal pressure, a large animal model with chronic liver disease is necessary. Currently, the canine animal model where liver fibrosis and cirrhosis is induced by carbon tetrachloride (CCl4) is often used to study hemodynamic changes of portal hypertension. Furthermore, the canine model is similar to human liver cirrhosis in serologic and pathologic physiology. Studies have demonstrated that canines develop liver fibrosis with elevation of portal pressure within 3–6 mo after injection of CCl4[10].

In this study, portal pressure was quantitatively estimated with CEUS in the hepatic fibrosis canine model and compared to catheter-based portal pressure measurement as the gold standard.

**MATERIALS AND METHODS**

***Ethics statement***

The study was reviewed and approved by the Beijing Friendship Hospital Institutional Review Board (Beijing, China). All procedures involving animals were reviewed and approved by the Institutional Animal Care and Use Committee of Beijing Friendship Hospital (IACUC protocol number: 12-5001). Animals were housed in the animal facility at the Beijing Friendship Hospital and were maintained under constant conditions at room temperature (20 ± 2 °C) and a relative humidity of 65% ± 10%. Animals were fed commercial canine food with access to tap water, and were quarantined for a two-week period before the study began.

***Animal model***

Healthy adult Beagles (*n* = 14; 7 males and 7 females weighing 10.5–14.5 kg) were obtained from the Beijing Rixin Technology Company (Beijing, China). To establish the liver fibrosis model, the animals received a subcutaneous injection of 60% CCl4 **olive oil** emulsion on the dorsal area of the body, at a dose of 1.0–1.5 mL/kg every 7 d. Penicillin (40 U/g, im, qd.) was administered on the first three consecutive days to prevent infection. After the injection, all animals were fed with granule feedstuff mixed with 10% lard. The amount of granule feedstuff was 15 g/kg per day for the first three days, and not controlled from the fourth to seventh day. The animal experiment lasted six months.

***CEUS technique and measurement***

CEUS was performed with an IU22 system (Philips Healthcare, Amsterdam, Netherlands) and a 2–5 MHz broadband convex array transducer. Real-time harmonic contrast imaging with a mechanical index of 0.06 was utilized. The depth of the image was maintained at 8–10 cm, and all parameters were consistent for all animals. The second-generation contrast agent SonoVue (Bracco Diagnostic Inc., Milan, Italy), which contains sulfur hexafluoride–filled microbubbles, was used for CEUS imaging of the liver and renal parenchyma.

The animals were anesthetized with pentobarbital sodium (30 mg/kg) through a 20-gauge catheter in an auricular vein after fasting for 12 h. All imaging was performed in the supine position. After conventional B-mode and color Doppler imaging was performed, the probe was placed in the subcostal position to display the portal vessels in the liver-kidney section. A bolus of SonoVue (0.5 mL) was then administered and immediately followed by an injection of normal saline (5 mL). The time for the agent to arrive at the kidney and liver was recorded with the timer on the ultrasound system. The clips of the real-time contrast imaging were stored in the built-in hard drive for later measurement and analysis.

In the liver-kidney section, the regions of interest (ROIs) were set on the hepatic parenchyma and the renal cortex (Figure 1). The ROIs were carefully placed on each section of the liver in order to avoid intrahepatic large vessels. Time-intensity curves (TICs) of CEUS for both ROIs were generated simultaneously with QLAB software on the IU22 system. The quality of the fit for the TICs was over 80%.

Because the liver has two independent blood supply systems, the hepatic artery and the portal vein, the arterial and portal phases cannot be clearly differentiated on the TIC of hepatic parenchyma. Thus, the computed tomography (CT) method for the evaluation of portal hypertension was adapted, *i.e.* the time to the maximum adjacent organ (splenic or renal cortical) enhancement as the demarcation point for the hepatic artery and portal vein phases[11]. Here, the time point when the peak renal intensity was reached was used as the demarcation between the hepatic arterial and portal venous phases on the TIC. The TIC parameters for the liver-kidney sectional imaging were defined as follows: (1) time of the hepatic arterial phase (Ta): the time when the renal intensity had reached the peak on the TIC; (2) intensity of the hepatic arterial phase (Ia): intensity at the time of Ta on the TIC of the hepatic parenchyma; (3) intensity of the hepatic parenchyma (Ipeak): maximum intensity from the TIC of the hepatic parenchyma; (4) peak time of the hepatic parenchyma (Tpeak): time when the TIC arrived at Ipeak; (5) intensity of the portal venous phase (Ip): Ipeak-Ia, representing the continuously increasing intensity during the portal venous phase; (6) Qp/Qa: area under the curve of the portal venous phase/hepatic arterial phase=/, representing the proportion of portal vein and hepatic arterial perfusion; and (7) Ip/Ia: intensity of the portal venous phase/hepatic arterial phase.

***Free portal pressure measurement***

All animals underwent portal pressure measurements immediately following CEUS. The free portal venous pressure (FPP) was measured with a multi-channel physiologic signal acquisition system equipped with a baroreceptor (RM6240; Chengdu Instrument Factory, Chengdu, China). Measurements were made with a percutaneous approach at the baseline and 8, 16, and 24 wk after the induction of liver fibrosis in each canine. Under general anesthesia, a 21-gauge needle over a guide wire was inserted into the right branch of the portal vein. The needle core was removed and connected to the baroreceptor. The pressure tracing was recorded for 45–60 s in order to obtain a stable measurement and to demonstrate fluctuation of respiration. The FPP was defined as the mean of five consecutive waveforms.

***Liver biopsy***

After the FPP measurement, all animals underwent percutaneous biopsy of the right lobe of the liver under ultrasound guidance in the same experiment. Two core specimens > 1.5 cm in length were obtained from each animal with an 18-gauge needle biopsy device (BARD Magnum; BARD Inc., Tempe, AZ, USA). Biopsies were pathologically evaluated, and hepatic fibrosis was classified according to the Metavir scoring system as follows: F0, no fibrosis; F1, portal fibrosis without septa; F2, portal fibrosis and few septa; F3, numerous septa without cirrhosis; and F4, cirrhosis[12].

***Statistical analysis***

Quantitative variables are expressed as the mean ± standard deviation. The repeated measures analysis of variance was utilized to compare FPP and CEUS parameters at multiple time points in all canines. A Pearson correlation test was used to compare Ip/Ia and Qp/Qa with FPP for all canines. *P* < 0.05 was considered statistically significant.

**RESULTS**

Pathologic examination of liver biopsies (*n* = 42) obtained from animals at 8, 16, and 24 wk revealed that different stages of liver fibrosis were successfully created in 14 canines. Fibrosis in the canine liver biopsies was staged according to the Metavir scoring system as follows: F0 (*n* = 3), F1 (*n* = 12), F2 (*n* = 14), F3 (*n* = 11), and F4 (*n* = 2) (Table 1). Although all stages of fibrosis were represented, cirrhosis was evident in only two cases.

The values of FPP increased gradually in the 14 canines over the 24-wk period. There were significant differences for FPP at 8, 16, and 24 wk compared to baseline FPP values from the canines (*P* < 0.001) (Table 2).

The TICs obtained from baseline and at 24 wk (F4) of induced fibrosis are presented in Figures 2 and 3. The duration of the hepatic arterial phase was shorter in liver fibrosis than the duration of the portal venous phase of baseline liver (Figure 2). In the liver fibrosis model, the duration of the hepatic arterial phase increased while the duration of portal venous phase decreased (Figure 3). Qp/Qa and Ip/Ia declined in the canine model, and there were significant differences at 8, 16, and 24 wk compared to baseline (Table 2).

There was a statistically significant negative correlation between FPP and Qp/Qa (*r* = -0.707; *P* < 0.001), and between FPP and Ip/Ia (*r* = -0.759; *P* < 0.001) in the canine model. Linear regression equations were y = -4.556x + 71.14 and y = -32.828x + 55.82, respectively (Figures 4 and 5).

The utility of the CEUS measurements in the prediction of elevated FPP was evaluated by the area under the receiver operating characteristic curves: Qp/Qa was 0.866 for elevated portal pressure (FPP ≥ 18 cmH2O), and 0.895 for Ip/Ia (Figures 6 and 7). The sensitivity and the specificity of Qp/Qa were 76 and 86%, and 85 and 87%, respectively, for Ip/Ia. These results indicated that Qp/Qa and Ip/Ia were favorable predictors for elevated portal pressure in liver fibrosis.

**DISCUSSION**

There are two reliable methods for accurately evaluating portal pressure: HVPG and FPP measurements. The measurement of the HVPG has been accepted as the gold standard for assessing the degree of portal hypertension[13–16]. The HVPG displays the difference in pressure between a free position and a wedged position in the hepatic vein. HVPG actually reflects the sinusoidal pressure, but it is not routinely performed due to its invasive nature and the special equipment and technical expertise required. While FPP is a measurement that directly reflects the portal pressure, it is also obtained intraoperatively, and therefore, not feasible for repeated assessment in a clinical setting. In addition, FPP cannot be used to determine the portal pressure in the early stages of liver disease and thus, the technique is inappropriate for monitoring disease progression. In this study, ultrasound-guided transportal puncture was used to measure FPP. This method is simple, easy to perform, and can be conducted repeatedly over the course of disease progression. Furthermore, the baroreceptor used in this study is more sensitive than the traditional catheter. FPP in the canine model increased gradually with the progress of the experiment, and FPP was successfully measured in all canines.

CEUS provides hemodynamic information of blood circulation and tissue perfusion. Several previous studieshave suggested that the severity of portal hypertension in patients with chronic liver disease is strongly correlated with a variety of CEUS features, such as arrival time and transit time of the hepatic vein[17,18]. However, other investigators question the approach, and suggest that CEUS has no clear diagnostic value in particular for mild and moderately elevated portal pressure[19]. Recently, the CEUS methodology has expanded the potential of hemodynamic studies performed in the United States[20]. The Ip/Ia and Qp/Qa proposed in this study are new CEUS features for evaluating the blood flow ratio of the portal vein/hepatic artery. Quantitative analysis of CEUS is affected by many factors, including probe frequency, mechanical index, analysis software, and concentration of the agent, and the area, shape, depth, and position of the ROI. In order to reduce the influence of these other factors, the introduction of Ip/Ia and Qp/Qa parameters, which are obtained from the same area, has obvious advantages over absolute values, such as the peak intensity. The ratio eliminates the influences mentioned above, and reflects the theoretic significance of hepatic parenchymal perfusion parameter ratio changes.

Both the portal vein and the hepatic artery feed the hepatic blood supply. Scholars believe that there is a significant correlation between intrahepatic blood flow and the portal pressure[21]. Current research indicates that the initiating factor in portal hypertension is increased resistance in the portal vein. Over the course of chronic liver disease, various factors lead to increased intrahepatic vascular resistance, such as the deposition of collagen fibers, the elastic decline of liver sinusoidal endothelial cells, the transformation of stellate cells into fibroblasts, and microthrombosis in the distal vein[22,23]. Moreover, intrahepatic hemodynamic changes, such as arteriovenous or portovenous shunting and the arterialization of capillary beds in the liver, also contribute to high portal pressure[24–26]. All of the above factors lead to a reduction in the amount of portal blood flow. Meanwhile, a compensatory increase in the blood flow of the hepatic artery, due to the buffer response, increases the proportion of the blood flow in the hepatic artery[27,28].

The histopathologic changes of the canine model induced by CCl4 are similar to human liver fibrosis and cirrhosis. Previous studies in canine models revealed that, in the early stages of liver fibrosis, the possible cause of elevated portal pressure was increased resistance of the blood stream due to sinusoid capillarization and activation of hepatic stellate cells[10]. Our data, that Qp/Qa and Ip/Ia declined, support these findings; the blood flow ratio of the portal vein/hepatic artery decreased, while the portal pressure increased, which is consistent with the histopathologic and hemodynamic changes of increasing portal pressure as mentioned previously. As liver fibrosis progressed and portal pressure increased, the TIC pattern from the liver parenchyma later in the experiment was almost the same as that of the renal cortex, as under these conditions, liver parenchyma infusion is mainly from the hepatic artery.

In the present study, established CT methods were adapted for evaluation of portal hypertension. Some investigators have previously used CT perfusion imaging and concluded that hepatic arterial blood flow increases while the portal venous blood flow decreases, as the portal pressure is increased[11,29]. The correlation between the parameters of spiral CT perfusion imaging and portal vein pressure in normal canines has been evaluated, and the results demonstrated that portal pressure was negatively correlated with portal venous perfusion[30]. CT has certain advantages in the evaluation of portal hypertension. However, the technique also has limitations. First, the frame rate is too low, making boundaries for the hepatic artery and portal venous perfusion phase inaccurate. Second, the iodinated contrast agent will diffuse from the capillary walls into the tissue space and impact the accuracy of portal venous perfusion phase detection. In contrast, CEUS has high temporal resolution, with a frame rate of 10 frames per second or more, and can more accurately distinguish enhanced phase, speed, and intensity between the hepatic artery and the portal vein.

The present study has some limitations. First, the number of experimental animals was limited. Second, measurements were conducted under anesthesia, so that respiratory motion artifacts of CEUS images may have affected the accuracy of the TICs. This particular issue was overcome in part by maintaining a low breathing amplitude in order to limit the range of movement. However, the quality of the fit of the TICs was over 80%. Finally, few biopsies were staged as F4. In the future, more biopsies representing each stage will be obtained in order to rigorously assess the utility of the CEUS measurements.

In conclusion, CEUS is a potential method to quantitatively and noninvasively estimate the portal venous pressure by determination of Qp/Qa and Ip/Ia parameters. Animal experiments demonstrated that Qp/Qa and Ip/Ia were favorable predictors for the elevated portal pressure in liver fibrosis, which can provide the basis for effectively measuring the portal pressure level during the progression of chronic liver disease. More measurements in larger cohorts are needed to confirm these preliminary results as well as to further demonstrate clinical utility.

**COMMENTS**

***Background***

Portal hypertension is a severe complication of chronic liver disease and cirrhosis, which directly leads to many clinical complications due to hemodynamic changes of portal pressure. Hepatic venous pressure gradient and free portal pressure (FPP) measurements are the two currently reliable methods for accurately evaluating portal pressure, but the methods are only performed under invasive procedures. Therefore, a noninvasive method to evaluate portal pressure over time would present distinct advantages clinically. The hepatic blood supply is fed by both the portal vein and hepatic artery. Current research indicates that the initiating factor in portal hypertension is increased resistance of the portal vein, which leads to a reduction in the amount of portal blood flow. Contrast-enhanced ultrasound (CEUS) techniques can provide the hemodynamic parameters of blood circulation and tissue perfusion. Thus, the aim of this study was to verify whether there is a significant correlation between CEUS parameters and portal pressure.

***Research frontiers***

The author aimed to establish a canine liver fibrosis model and to investigate the feasibility of a noninvasive quantitative estimation of portal pressure with CEUS during the development of liver fibrosis. The intensity and area under the curve of the portal venous phase/hepatic arterial phase (Ip/Ia and Qp/Qa, respectively) parameters used in this study are new CEUS features for evaluating the blood flow ratio of the portal vein/hepatic artery.

***Innovations and breakthroughs***

The introduction of Ip/Ia and Qp/Qa parameters has obvious advantages over the absolute value, such as peak intensity, as this approach reduces the influence of numerous factors that affect CEUS. This study demonstrates for the first time that there was a statistically significant negative correlation of FPP with Ip/Ia and Qp/Qa in the progression of liver fibrosis in a canine model.

***Applications***

Based on the results of the correlation of FPP with Ip/Ia and Qp/Qa, this study supports CEUS as a potential method to noninvasively estimate the portal pressure by measurement of Qp/Qa and Ip/Ia parameters.

***Terminology***

CEUS is the use of intravenous microbubble contrast agents in ultrasonography. The use of such contrast agents has been shown to improve the characterization of the vasculature inside the organ of interest and provide the hemodynamic parameters of blood circulation and tissue perfusion. The Ip/Ia and Qp/Qa proposed in this study are CEUS parameters for evaluating the blood flow ratio of portal vein/hepatic artery.

***Peer-review***

This manuscript demonstrates that new methods for CEUS detect fibrosis *in vivo*. This is a noninvasive method combined with CT and known parameters, such as FPP, Qp/Qa, and Ip/Ia. Previous publications have demonstrated that portal pressure was negatively correlated with portal hypertension. However, accuracy could be improved. In this study, the authors introduce CEUS with a higher temporal resolution to resolve the problem. The paper is well written, and they include crucial information concerning fibrosis.

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**Figure 1 Contrast-enhanced ultrasonography of the right lobe of the liver and the upper pole of the kidney in a sagittal view.** The region defined in yellow is the region of interest (ROI) of the hepatic parenchyma and in red, the ROI of the renal cortex.



**Figure 2 Time-intensity curve of normal canine hepatic parenchyma and renal cortex in the liver-kidney section.** The red line represents the time-intensity curve (TIC) of the renal cortex, and the yellow represents the TIC of hepatic parenchyma. ‘Ta’ represents the time at which renal intensity reaches the peak of the TIC. ‘Ia’ represents the intensity at the time of Ta in the TIC of the hepatic parenchyma. Ipeak and Tpeak represent the maximum intensity of the TIC of the hepatic parenchyma and the time at which the TIC arrives at Ipeak, respectively. The area under the curve of portal venous time and the hepatic arterial time are expressed as Qp and Qa, respectively.



**Figure 3 Time-intensity curves of the hepatic parenchyma and the renal cortex in the liver-kidney section of the canine model at 24 wk.** The red and yellow lines represent the time-intensity curves (TICs) of the hepatic parenchyma and the renal cortex, respectively, in a treated animal. Points on the curve are highlighted where the data for the parameters indicated were derived. ‘Ta’ represents the time at which renal intensity reaches the peak of the TIC. ‘Ia’ represents the intensity at the time of Ta in the TIC of the hepatic parenchyma. Ipeak and Tpeak represent the maximum intensity of the TIC of the hepatic parenchyma and the time at which the TIC arrives at Ipeak, respectively. The area under the curve of portal venous time and the hepatic arterial time are expressed as Qp and Qa, respectively. The proportion of hepatic arterial infusion increased, and Qp/Qa and Ip/Ia were reduced.



**Figure 4 Relationship between the area under the curve of the portal venous phase/hepatic arterial phase (Qp/Qa) and free portal pressure in the canine model.** Free portal pressure measurements from model animals at all time points were plotted as a function of Qp/Qa in order to assess the relationship between the two parameters.



**Figure 5 Relationship between the intensity of the portal venous phase/hepatic arterial phase (Ip/Ia) and free portal pressure in the canine model.** Free portal pressure measurements from model animals at all time points were plotted as a function of Ip/Ia in order to assess the relationship between the two parameters.



**Figure 6 Receiver operating characteristic curve showing that the prediction of elevated portal pressure (free portal pressure ≥ 18 cmH2O) based on the area under the curve of the portal venous phase/hepatic arterial phase was 0.866.**



**Figure 7 Receiver operating characteristic curve showing that the prediction of elevated portal pressure (free portal pressure ≥ 18 cmH2O) based on the intensity of the portal venous phase/hepatic arterial phase was 0.895.**

**Table 1 Pathologic staging of liver biopsies from the canine model**

|  |  |  |  |  |
| --- | --- | --- | --- | --- |
| **Stage** | **8th wk** | **16th wk** | **24th wk** | **Total** |
| F0 | 3 | 0 | 0 | 3 |
| F1 | 9 | 3 | 0 | 12 |
| F2 | 2 | 8 | 4 | 14 |
| F3 | 0 | 3 | 8 | 11 |
| F4 | 0 | 0 | 2 | 2 |
| Total | 14 | 14 | 14 | 42 |

**Table 2** **Free portal pressure and contrast-enhanced ultrasound parameters in normal and treated canines (*n* = 14)**

|  |  |  |  |
| --- | --- | --- | --- |
| **Time in weeks** | **FPP in cmH2O** | **Qp/Qa** | **Ip/Ia** |
| Normal | 10.26 ± 2.12 | 19.85 ± 3.30 | 1.77 ± 0.37 |
| 8  | 18.31 ± 3.17 | 11.32 ± 0.92 | 1.04 ± 0.12 |
| 16  | 28.20 ± 2.57 | 9.43 ± 0.85 | 0.85 ± 0.07 |
| 24  | 36.30 ± 2.21 | 7.27 ± 0.81 | 0.66 ± 0.13 |
| *P*  | < 0.001 | < 0.001 | < 0.001 |

Ip/Ia: Intensity of the portal venous phase/hepatic arterial phase; Qp/Qa: Area under the curve of the portal venous phase/hepatic arterial phase. FPP: Free portal pressure.