

## Species differences in regulation of renal proximal tubule transport by certain molecules

George Seki, Motonobu Nakamura, Masashi Suzuki, Nobuhiko Satoh, Shoko Horita

George Seki, Motonobu Nakamura, Masashi Suzuki, Nobuhiko Satoh, Shoko Horita, Department of Internal Medicine, Faculty of Medicine, University of Tokyo, Tokyo 113-0033, Japan

**Author contributions:** All the authors contributed to writing of this manuscript.

**Conflict-of-interest:** None.

**Open-Access:** This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

**Correspondence to:** George Seki, MD, Department of Internal Medicine, Faculty of Medicine, University of Tokyo, 7-3-1 Hongo, Bunkyo-ku, Tokyo 113-0033,

Japan. [georgeseki-tky@umin.ac.jp](mailto:georgeseki-tky@umin.ac.jp)

Telephone: +81-3-38155411

Fax: +81-3-58008806

Received: November 26, 2014

Peer-review started: November 27, 2014

First decision: December 18, 2014

Revised: December 24, 2014

Accepted: January 18, 2015

Article in press: January 20, 2015

Published online: May 6, 2015

### Abstract

Renal proximal tubules (PTs) play important roles in the regulation of acid/base, plasma volume and blood pressure. Recent studies suggest that there are substantial species differences in the regulation of PT transport. For example, thiazolidinediones (TZDs) are widely used for the treatment of type 2 diabetes mellitus, but the use of TZDs is associated with fluid overload. In addition to the transcriptional enhancement of sodium transport in distal nephrons, TZDs rapidly stimulate PT sodium transport *via* a non-genomic mechanism depending on

peroxisome proliferator activated receptor  $\gamma$ /Src/epidermal growth factor receptor (EGFR)/MEK/ERK. In mouse PTs, however, TZDs fail to stimulate PT transport probably due to constitutive activation of Src/EGFR/ERK pathway. This unique activation of Src/ERK may also affect the effect of high concentrations of insulin on mouse PT transport. On the other hand, the effect of angiotensin II (Ang II) on PT transport is known to be biphasic in rabbits, rats, and mice. However, Ang II induces a concentration-dependent, monophasic transport stimulation in human PTs. The contrasting responses to nitric oxide/guanosine 3',5'-cyclic monophosphate pathway may largely explain these different effects of Ang II on PT transport. In this review, we focus on the recent findings on the species differences in the regulation of PT transport, which may help understand the species-specific mechanisms underlying edema formation and/or hypertension occurrence.

**Key words:** Renal proximal tubule; Thiazolidinediones; Peroxisome proliferator activated receptor  $\gamma$ ; Insulin; Angiotensin II; Nitric oxide

© **The Author(s) 2015.** Published by Baishideng Publishing Group Inc. All rights reserved.

**Core tip:** Renal proximal tubule (PT) transport is essential for the regulation of plasma volume and blood pressure. Several species differences are found as to the stimulatory effects of thiazolidinediones, insulin, and angiotensin II on PT sodium transport. This review focuses on this topic, which may be relevant to species-specific mechanisms underlying edema formation and/or hypertension occurrence.

Seki G, Nakamura M, Suzuki M, Satoh N, Horita S. Species differences in regulation of renal proximal tubule transport by certain molecules. *World J Nephrol* 2015; 4(2): 307-312 Available from: URL: <http://www.wjgnet.com/2220-6124/full/v4/i2/307.htm> DOI: <http://dx.doi.org/10.5527/wjn.v4.i2.307>

## INTRODUCTION

The kidney plays an essential role in the homeostatic regulation of electrolytes, acid-base and plasma volume in the body. Some species differences are known to exist in the distribution patterns of solute transporters as well as the hormonal actions in distal nephron<sup>[1,2]</sup>. In proximal tubules (PTs), on the other hand, there are no major species differences in the distribution patterns and functions of main sodium transporters such as the apical Na<sup>+</sup>/H<sup>+</sup> exchanger type 3 NHE3 and the basolateral electrogenic Na<sup>+</sup>-HCO<sub>3</sub><sup>-</sup> cotransporter NBCe1<sup>[3,4]</sup>. While the transport stoichiometry of NBCe1 was reported to be 1Na<sup>+</sup> to 3HCO<sub>3</sub><sup>-</sup> in rat PTs *in vivo*<sup>[5]</sup>, it was found to be 1Na<sup>+</sup> to 2HCO<sub>3</sub><sup>-</sup> in rabbit PTs *in vitro*<sup>[6]</sup>. However, this difference was turned out to be due to the differences in experimental conditions<sup>[7,8]</sup>.

Nevertheless, recent studies identified substantial species differences in the regulation of PT transport, which might be important for uncovering the species-specific causes for edema and/or hypertension. Species differences in renal physiologic and/or metabolic responses may be also potentially important for understanding mechanisms for the different effects of several agents against diabetic nephropathy between animal models and human patients. For example, inhibition of advanced glycosylation end products by aminoguanidine or pyridoxamine was reported to be protective in rodent models of diabetic nephropathy<sup>[9-12]</sup>. However, these agents were not found effective in initial human trials in diabetic patients<sup>[13,14]</sup>. Effectiveness of a selective PKC-inhibitor Ruboxistaurin found in rodent models of diabetic nephropathy<sup>[15,16]</sup> was also not confirmed in human type 2 diabetic patients<sup>[17]</sup>. Indeed, rodent models of diabetic nephropathy exhibit proteinuria and pathological glomerular changes, but do not exhibit progressive renal failure<sup>[18]</sup>. Dogs, pigs and other non-human primates have been used for toxicology testing. However, ideal animal models have not been established for revealing drug side effects<sup>[19]</sup>. We focus on species differences in the regulation of PT transport in this review.

## EFFECTS OF THIAZOLIDINEDIONES

Thiazolidinediones (TZDs) activate a nuclear receptor peroxisome proliferator activated receptor  $\gamma$  (PPAR $\gamma$ ), thereby improving insulin resistance through the transcriptional modulation of the relevant genes<sup>[20]</sup>. TZDs have been widely used for the treatment of type 2 diabetes. However, fluid retention is a serious clinical problem, which may exert an adverse effect on the cardiovascular system<sup>[21]</sup>.

Initially, PPAR $\gamma$ -mediated transcriptional enhancement of the epithelial Na channel ENaC $\gamma$  subunit in collecting ducts was proposed to be a main mechanism for TZDs-induced volume expansion<sup>[22,23]</sup>. Subsequent analysis in renal principal cell culture models, however, did not support the central role of ENaC in TZDs-induced volume

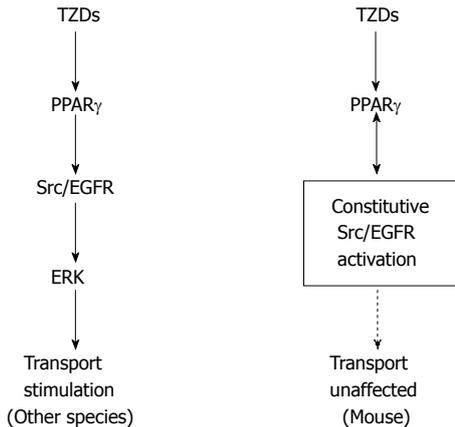
expansion<sup>[24]</sup>. Moreover, TZDs-induced volume expansion was preserved in collecting ducts specific ENaC deficient mice<sup>[25]</sup>. These results indicate that mechanism(s) other than the activation of ENaC in collecting ducts may be also involved in TZDs-induced volume expansion. Interestingly, Muto and colleagues reported the rapid stimulation of NBCe1 activity by troglitazone in isolated rabbit PTs<sup>[26]</sup>. Because TZDs were reported to stimulate PT transport also in humans<sup>[27]</sup>, we performed the detailed analysis on the mechanism of TZDs-induced stimulation of PT sodium transport.

We found that TZDs rapidly induced transport stimulation within minutes in isolated PTs from rabbits, rats, and humans. Our subsequent analysis revealed that TZDs-induced PT transport stimulation was dependent on the non-genomic signaling cascade consisting of PPAR $\gamma$ /Src/epidermal growth factor receptor (EGFR)/MEK/ERK<sup>[28]</sup>. Notably, however, TZDs failed to induce transport stimulation in isolated mouse PTs, despite the definite expression of PPAR $\gamma$  in these tubules. We speculated that some factor(s) specific for mouse PTs might interfere with TZDs-induced rapid signaling. Consistent with this view, Kiley *et al.*<sup>[29]</sup> reported the constitutive activation of Src/EGFR pathway that was found only in mouse PTs. This unique constitutive activation of Src/EGFR, which could potentially explain the different effects of exogenous EGF on unilateral ureteral obstruction in rats and mice<sup>[30]</sup>, was confined to mouse PTs and not found in other mouse tissues<sup>[29]</sup>. Because TZDs rapidly activated the Src/ERK pathway in kidney cortex of rabbits and rats but not mice, we concluded that the constitutive activation of Src/EGFR prevented TZDs-induced transport stimulation in mouse PTs<sup>[28]</sup>. Consistent with this conclusion, the constitutive activation of Src was also reported to interfere with the non-genomic signaling of another nuclear receptor for estrogen<sup>[31]</sup>.

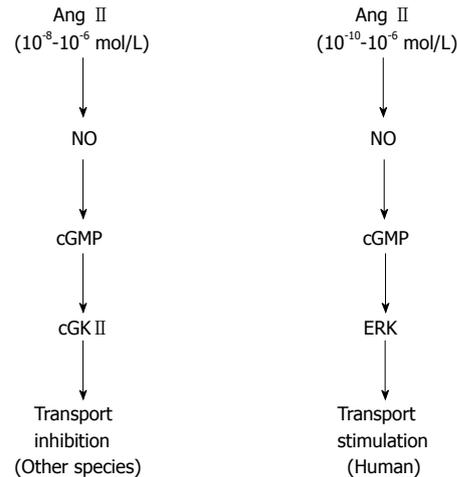
Most likely, TZDs-induced volume expansion is multifactorial, depending on both genomic and non-genomic actions of PPAR $\gamma$  on tubular sodium transport. One of potential targets of non-genomic actions of PPAR $\gamma$  may be distal nephron, where WNK kinases regulate the balance between renal NaCl absorption and K<sup>+</sup> secretion<sup>[32,33]</sup>. WNK4 is also known to affect Cl<sup>-</sup> transport in extrarenal epithelia<sup>[34]</sup>. It remains to be determined whether PPAR $\gamma$  regulates the WNK kinase system.

## EFFECTS OF INSULIN

Insulin is thought to enhance renal sodium retention by activating sodium transport in several nephron segments<sup>[35]</sup>. In PTs, insulin stimulates the major sodium transporters NHE3, NBCe1, and the Na<sup>+</sup>/K<sup>+</sup>-ATPase<sup>[36-38]</sup>. In isolated rabbit PTs, Baum found that insulin, at the concentrations between 10<sup>-10</sup> mol/L and 10<sup>-8</sup> mol/L, dose-dependently stimulates volume and bicarbonate absorption<sup>[39]</sup>. Recently, we also found the similar dose-dependent stimulation of NBCe1 by up to 10<sup>-8</sup> mol/L insulin in isolated PTs from rats



**Figure 1 Species differences in thiazolidinediones effects on proximal tubule transport.** Non-genomic activation of ERK by TZDs via PPAR $\gamma$ /Src/EGFR pathway is suppressed in mouse proximal tubule probably due to constitutive activation of Src/EGFR. TZDs: Thiazolidinediones; PPAR: Peroxisome proliferator activated receptor; Src: Sarcoma; EGFR: Epidermal growth factor receptor; ERK: Extracellular signal-regulated kinase.



**Figure 2 Species differences in angiotensin II effects on proximal tubule transport.** In human, NO/cGMP pathway mediates the concentration-dependent stimulatory effect of Ang II. In other species, NO/cGMP pathway mediates the inhibitory effect of high concentrations of Ang II. Ang II: Angiotensin II; NO: Nitrox oxide; cGMP: Guanosine 3',5'-cyclic monophosphate; cGK II: cGMP-dependent kinase II; ERK: Extracellular signal-regulated kinase.

and humans<sup>[40]</sup>. This stimulatory effect of insulin on PT sodium transport, which is completely preserved in insulin resistant rats and humans, may play an important role in the pathogenesis of hypertension associated with insulin resistance<sup>[40]</sup>.

Insulin actions are initiated by the activation of tyrosine kinase in the cell membrane receptor, which induces a series of phosphorylation in multiple insulin receptor substrates (IRSs). The two major IRS proteins, IRS1 and IRS2 often mediate distinct insulin actions, and defects at the levels of IRS1 or IRS2 may be responsible for the occurrence of selective insulin resistance in several tissues<sup>[41]</sup>. We have shown that IRS2 but not IRS1 mediates the stimulatory effect of insulin on PT transport in both mice and rats. Moreover, we have confirmed that insulin activates Akt in kidney cortex of both mice and rats<sup>[40,42]</sup>. We found that up to  $10^{-9}$  mol/L insulin stimulated mouse PT sodium transport. Unlike in rats and humans, however, the higher concentrations of insulin (*i.e.*, more than  $10^{-8}$  mol/L) failed to stimulate PT transport in mice<sup>[42]</sup>. In isolated mouse collecting ducts, by contrast, insulin at the high concentrations up to  $10^{-7}$  mol/L was reported to activate ENaC<sup>[43]</sup>. Importantly, insulin is known to activate both Akt and ERK pathways, and these two pathways are interconnected with each other<sup>[44,45]</sup>. It is therefore possible that the constitutive activation of Src/ERK may somehow interfere with the effects of high concentrations of insulin in mouse PTs.

## EFFECTS OF ANGIOTENSIN II

The stimulation of PT sodium transport by Angiotensin II (Ang II) may be essential for Ang II-induced hypertension<sup>[46,47]</sup>. Actually, however, Ang II is known to regulate PT transport in a biphasic way: transport is stimulated by low (picomolar to nanomolar) concen-

trations of Ang II, but inhibited by high (nanomolar to micromolar) concentrations of Ang II. This biphasic regulation of PT transport by Ang II has been confirmed in rats, mice, and rabbits<sup>[48-51]</sup>. Regarding the receptor subtype(s) responsible for the biphasic effects of Ang II, controversial results had been reported<sup>[52-54]</sup>. However, the analyses on isolated PTs from type 1A Ang II receptor (AT<sub>1A</sub>) deficient mice have clearly shown that AT<sub>1A</sub> mediates both the stimulatory and inhibitory effects of Ang II<sup>[49,51]</sup>.

The stimulatory effect of Ang II is dependent on the activation of protein kinase C and/or the decrease in the intracellular cAMP concentration, which may result in the activation of ERK<sup>[55,56]</sup>. On the other hand, the inhibitory effect of Ang II is dependent on the activation of phospholipase A<sub>2</sub>/arachidonic acid/5,6-epoxyeicosatrienoic acid (EET) pathway and/or nitric oxide (NO)/guanosine 3',5'-cyclic monophosphate (cGMP) pathway<sup>[53,55,57]</sup>. Because little had been known about the direct effects of Ang II on human PT transport, we tried to clarify this issue using isolated, intact human PTs obtained from nephrectomy surgery.

Surprisingly, we found that the inhibitory effect of Ang II is lost in human PTs. Actually, up to  $10^{-5}$  mol/L Ang II dose-dependently stimulated human PT transport. In view of high intrarenal concentrations of Ang II, these data suggest that Ang II may play an even more important role in the regulation of plasma volume and blood pressure in humans than in other species<sup>[58]</sup>.

The detailed analysis revealed that the contrasting responses to NO/cGMP could explain the different modes of PT transport regulation in humans and other species. While the NO/cGMP/ERK pathway mediates the dose-dependent stimulatory effect of Ang II in

humans, the NO/cGMP/cGMP-dependent kinase II (cGKII) pathway mediates the inhibitory effect of high concentrations of Ang II in mice<sup>[58]</sup>. In cGKII-deficient mice, the inhibitory effect of Ang II was lost, but the NO/cGMP pathway failed to stimulate PT transport. These results indicate that the loss of cGKII alone in mice cannot reproduce the NO/cGMP/ERK-dependent stimulatory effect of Ang II in humans.

Currently, the reason why the NO/cGMP pathway exerts different effects on PT transport in humans and other species remains unknown. Nevertheless, several previous reports supported the existence of such species differences. For example, renal NO production was enhanced by salt loading in rodents, which might facilitate sodium diuresis and prevent blood pressure elevation<sup>[59,60]</sup>. Thus, renal NO is thought to induce a natriuretic response to salt loading in rodents<sup>[61]</sup>. However, renal NO production was not significantly enhanced by salt loading in humans<sup>[62,63]</sup>, and an adaptive role of renal NO to salt loading is less clear in humans<sup>[64,65]</sup>. In any case, the stimulatory effect of NO/cGMP pathway on PT transport may represent a human-specific target for hypertension.

The NO/cGMP pathway is thought to be inhibitory on sodium transport in thick ascending limb and collecting ducts<sup>[61]</sup>. Therefore, it will be interesting to examine whether the similar species differences may also exist in the effects of NO/cGMP pathway on these nephron segments. Such knowledge may help understand the mechanism of fluid overload by selective endothelin receptor antagonism<sup>[66]</sup>.

## CONCLUSION

In summary, non-genomic stimulation of PT transport by TZDs is uniquely absent in mice probably because of the constitutive activation of Src/EGFR as shown in Figure 1. As shown in Figure 2, on the other hand, the inhibitory effect of Ang II is lost in human PTs, where the NO/cGMP pathway is stimulatory unlike in other species. These species differences may be at least partially responsible for the species-specific mechanisms underlying edema formation and/or hypertension occurrence.

## REFERENCES

- 1 **Biner HL**, Arpin-Bott MP, Loffing J, Wang X, Knepper M, Hebert SC, Kaissling B. Human cortical distal nephron: distribution of electrolyte and water transport pathways. *J Am Soc Nephrol* 2002; **13**: 836-847 [PMID: 11912242]
- 2 **Morel F**. Sites of hormone action in the mammalian nephron. *Am J Physiol* 1981; **240**: F159-F164 [PMID: 6259951]
- 3 **Alpern RJ**. Cell mechanisms of proximal tubule acidification. *Physiol Rev* 1990; **70**: 79-114 [PMID: 2404291]
- 4 **Boron WF**. Acid-base transport by the renal proximal tubule. *J Am Soc Nephrol* 2006; **17**: 2368-2382 [PMID: 16914536 DOI: 10.1681/ASN.2006060620]
- 5 **Yoshitomi K**, Burckhardt BC, Frömter E. Rheogenic sodium-bicarbonate cotransport in the peritubular cell membrane of rat renal proximal tubule. *Pflugers Arch* 1985; **405**: 360-366 [PMID: 3936018]
- 6 **Seki G**, Coppola S, Frömter E. The Na(+)-HCO<sub>3</sub>- cotransporter operates with a coupling ratio of 2 HCO<sub>3</sub>- to 1 Na+ in isolated rabbit renal proximal tubule. *Pflugers Arch* 1993; **425**: 409-416 [PMID: 8134258 DOI: 10.1007/BF00374866]
- 7 **Müller-Berger S**, Nesterov VV, Frömter E. Partial recovery of in vivo function by improved incubation conditions of isolated renal proximal tubule. II. Change of Na-HCO<sub>3</sub> cotransport stoichiometry and of response to acetazolamide. *Pflugers Arch* 1997; **434**: 383-391 [PMID: 9211803 DOI: 10.1007/s004240050411]
- 8 **Seki G**, Coppola S, Yoshitomi K, Burckhardt BC, Samarzija I, Müller-Berger S, Frömter E. On the mechanism of bicarbonate exit from renal proximal tubular cells. *Kidney Int* 1996; **49**: 1671-1677 [PMID: 8743474 DOI: 10.1038/ki.1996.244]
- 9 **Alderson NL**, Chachich ME, Youssef NN, Beattie RJ, Nachtigal M, Thorpe SR, Baynes JW. The AGE inhibitor pyridoxamine inhibits lipemia and development of renal and vascular disease in Zucker obese rats. *Kidney Int* 2003; **63**: 2123-2133 [PMID: 12753299 DOI: 10.1046/j.1523-1755.2003.00027.x]
- 10 **Bucala R**, Vlassara H. Advanced glycosylation end products in diabetic renal and vascular disease. *Am J Kidney Dis* 1995; **26**: 875-888 [PMID: 7503061]
- 11 **Degenhardt TP**, Alderson NL, Arrington DD, Beattie RJ, Basgen JM, Steffes MW, Thorpe SR, Baynes JW. Pyridoxamine inhibits early renal disease and dyslipidemia in the streptozotocin-diabetic rat. *Kidney Int* 2002; **61**: 939-950 [PMID: 11849448 DOI: 10.1046/j.1523-1755.2002.00207.x]
- 12 **Friedman EA**, Distant DA, Fleishhacker JF, Boyd TA, Cartwright K. Aminoguanidine prolongs survival in azotemic-induced diabetic rats. *Am J Kidney Dis* 1997; **30**: 253-259 [PMID: 9261037]
- 13 **Bolton WK**, Cattran DC, Williams ME, Adler SG, Appel GB, Cartwright K, Foiles PG, Freedman BI, Raskin P, Ratner RE, Spinowitz BS, Whittier FC, Wuerth JP. Randomized trial of an inhibitor of formation of advanced glycation end products in diabetic nephropathy. *Am J Nephrol* 2004; **24**: 32-40 [PMID: 14685005 DOI: 10.1159/000075627]
- 14 **Lewis EJ**, Greene T, Spitaler S, Blumenthal S, Berl T, Hunsicker LG, Pohl MA, Rohde RD, Raz I, Yerushalmy Y, Yagil Y, Hershkovits T, Atkins RC, Reutens AT, Packham DK, Lewis JB. Pyridoxamine in type 2 diabetic nephropathy. *J Am Soc Nephrol* 2012; **23**: 131-136 [PMID: 22034637 DOI: 10.1681/ASN.2011030272]
- 15 **Ishii H**, Jirousek MR, Koya D, Takagi C, Xia P, Clermont A, Bursell SE, Kern TS, Ballas LM, Heath WF, Stramm LE, Feener EP, King GL. Amelioration of vascular dysfunctions in diabetic rats by an oral PKC beta inhibitor. *Science* 1996; **272**: 728-731 [PMID: 8614835]
- 16 **Koya D**, Haneda M, Nakagawa H, Isshiki K, Sato H, Maeda S, Sugimoto T, Yasuda H, Kashiwagi A, Ways DK, King GL, Kikawa R. Amelioration of accelerated diabetic mesangial expansion by treatment with a PKC beta inhibitor in diabetic db/db mice, a rodent model for type 2 diabetes. *FASEB J* 2000; **14**: 439-447 [PMID: 10698958]
- 17 **Tuttle KR**, Bakris GL, Toto RD, McGill JB, Hu K, Anderson PW. The effect of ruboxistaurin on nephropathy in type 2 diabetes. *Diabetes Care* 2005; **28**: 2686-2690 [PMID: 16249540 DOI: 10.2337/diacare.28.11.2686]
- 18 **Breyer MD**, Böttinger E, Brosius FC, Coffman TM, Fogo A, Harris RC, Heilig CW, Sharma K. Diabetic nephropathy: of mice and men. *Adv Chronic Kidney Dis* 2005; **12**: 128-145 [PMID: 15822049 DOI: 10.1053/j.ackd.2005.01.004]
- 19 **Helke KL**, Swindle MM. Animal models of toxicology testing: the role of pigs. *Expert Opin Drug Metab Toxicol* 2013; **9**: 127-139 [PMID: 23216131 DOI: 10.1517/17425255.2013.739607]
- 20 **Yki-Järvinen H**. Thiazolidinediones. *N Engl J Med* 2004; **351**: 1106-1118 [PMID: 15356308 DOI: 10.1056/NEJMra041001]
- 21 **Mudaliar S**, Chang AR, Henry RR. Thiazolidinediones, peripheral edema, and type 2 diabetes: incidence, pathophysiology, and clinical implications. *Endocr Pract* 2003; **9**: 406-416 [PMID: 14583425 DOI: 10.4158/EP.9.5.406]
- 22 **Guan Y**, Hao C, Cha DR, Rao R, Lu W, Kohan DE, Magnuson

- MA, Redha R, Zhang Y, Breyer MD. Thiazolidinediones expand body fluid volume through PPAR $\gamma$  stimulation of ENaC-mediated renal salt absorption. *Nat Med* 2005; **11**: 861-866 [PMID: 16007095 DOI: 10.1038/nm1278]
- 23 **Zhang H**, Zhang A, Kohan DE, Nelson RD, Gonzalez FJ, Yang T. Collecting duct-specific deletion of peroxisome proliferator-activated receptor gamma blocks thiazolidinedione-induced fluid retention. *Proc Natl Acad Sci USA* 2005; **102**: 9406-9411 [PMID: 15956187 DOI: 10.1073/pnas.0501744102]
- 24 **Nofziger C**, Chen L, Shane MA, Smith CD, Brown KK, Blazer-Yost BL. PPAR $\gamma$  agonists do not directly enhance basal or insulin-stimulated Na(+) transport via the epithelial Na(+) channel. *Pflugers Arch* 2005; **451**: 445-453 [PMID: 16170524 DOI: 10.1007/s00424-005-1477-4]
- 25 **Vallon V**, Hummler E, Rieg T, Pochynyuk O, Bugaj V, Schroth J, Dechenes G, Rossier B, Cunard R, Stockand J. Thiazolidinedione-induced fluid retention is independent of collecting duct alphaENaC activity. *J Am Soc Nephrol* 2009; **20**: 721-729 [PMID: 19158355 DOI: 10.1681/ASN.2008040415]
- 26 **Muto S**, Miyata Y, Imai M, Asano Y. Troglitazone stimulates basolateral rheogenic Na<sup>+</sup>/HCO<sub>3</sub><sup>-</sup> cotransport activity in rabbit proximal straight tubules. *Exp Nephrol* 2001; **9**: 191-197 [PMID: 11340303 DOI: 10.1159/000052611]
- 27 **Zanchi A**, Chiolero A, Maillard M, Nussberger J, Brunner HR, Burnier M. Effects of the peroxisomal proliferator-activated receptor-gamma agonist pioglitazone on renal and hormonal responses to salt in healthy men. *J Clin Endocrinol Metab* 2004; **89**: 1140-1145 [PMID: 15001599 DOI: 10.1210/jc.2003-031526]
- 28 **Endo Y**, Suzuki M, Yamada H, Horita S, Kunimi M, Yamazaki O, Shirai A, Nakamura M, Iso-O N, Li Y, Hara M, Tsukamoto K, Moriyama N, Kudo A, Kawakami H, Yamauchi T, Kubota N, Kadowaki T, Kume H, Enomoto Y, Homma Y, Seki G, Fujita T. Thiazolidinediones enhance sodium-coupled bicarbonate absorption from renal proximal tubules via PPAR $\gamma$ -dependent nongenomic signaling. *Cell Metab* 2011; **13**: 550-561 [PMID: 21531337 DOI: 10.1016/j.cmet.2011.02.015]
- 29 **Kiley SC**, Chevalier RL. Species differences in renal Src activity direct EGF receptor regulation in life or death response to EGF. *Am J Physiol Renal Physiol* 2007; **293**: F895-F903 [PMID: 17626154 DOI: 10.1152/ajprenal.00227.2007]
- 30 **Kiley SC**, Thornhill BA, Belyea BC, Neale K, Forbes MS, Luetkeke NC, Lee DC, Chevalier RL. Epidermal growth factor potentiates renal cell death in hydronephrotic neonatal mice, but cell survival in rats. *Kidney Int* 2005; **68**: 504-514 [PMID: 16014027 DOI: 10.1111/j.1523-1755.2005.00428.x]
- 31 **Li L**, Hisamoto K, Kim KH, Haynes MP, Bauer PM, Sanjay A, Collinge M, Baron R, Sessa WC, Bender JR. Variant estrogen receptor-c-Src molecular interdependence and c-Src structural requirements for endothelial NO synthase activation. *Proc Natl Acad Sci USA* 2007; **104**: 16468-16473 [PMID: 17921256 DOI: 10.1073/pnas.0704315104]
- 32 **Kahle KT**, Wilson FH, Leng Q, Lalioti MD, O'Connell AD, Dong K, Rapson AK, MacGregor GG, Giebisch G, Hebert SC, Lifton RP. WNK4 regulates the balance between renal NaCl reabsorption and K<sup>+</sup> secretion. *Nat Genet* 2003; **35**: 372-376 [PMID: 14608358 DOI: 10.1038/ng1271]
- 33 **Yang CL**, Angell J, Mitchell R, Ellison DH. WNK kinases regulate thiazide-sensitive Na-Cl cotransport. *J Clin Invest* 2003; **111**: 1039-1045 [PMID: 12671053 DOI: 10.1172/JCI17443]
- 34 **Kahle KT**, Gimenez I, Hassan H, Wilson FH, Wong RD, Forbush B, Aronson PS, Lifton RP. WNK4 regulates apical and basolateral Cl<sup>-</sup> flux in extrarenal epithelia. *Proc Natl Acad Sci USA* 2004; **101**: 2064-2069 [PMID: 14769928 DOI: 10.1073/pnas.0308434100]
- 35 **Horita S**, Seki G, Yamada H, Suzuki M, Koike K, Fujita T. Insulin resistance, obesity, hypertension, and renal sodium transport. *Int J Hypertens* 2011; **2011**: 391762 [PMID: 21629870]
- 36 **Férraille E**, Carranza ML, Rousselot M, Favre H. Insulin enhances sodium sensitivity of Na-K-ATPase in isolated rat proximal convoluted tubule. *Am J Physiol* 1994; **267**: F55-F62 [PMID: 8048565]
- 37 **Gesek FA**, Schoolwerth AC. Insulin increases Na(+)–H<sup>+</sup> exchange activity in proximal tubules from normotensive and hypertensive rats. *Am J Physiol* 1991; **260**: F695-F703 [PMID: 1674643]
- 38 **Ruiz OS**, Qiu YY, Cardoso LR, Arruda JA. Regulation of the renal Na-HCO<sub>3</sub> cotransporter: IX. Modulation by insulin, epidermal growth factor and carbachol. *Regul Pept* 1998; **77**: 155-161 [PMID: 9809810 DOI: 10.1016/S0167-0115(98)00115-3]
- 39 **Baum M**. Insulin stimulates volume absorption in the rabbit proximal convoluted tubule. *J Clin Invest* 1987; **79**: 1104-1109 [PMID: 3549779]
- 40 **Nakamura M**, Yamazaki O, Shirai A, Horita S, Satoh N, Suzuki M, Hamasaki Y, Noiri E, Kume H, Enomoto Y, Homma Y, Seki G. Preserved Na/HCO<sub>3</sub> cotransporter sensitivity to insulin may promote hypertension in metabolic syndrome. *Kidney Int* 2014; **87**: 535-542 [PMID: 25354240 DOI: 10.1038/ki.2014.351]
- 41 **Thirone AC**, Huang C, Klip A. Tissue-specific roles of IRS proteins in insulin signaling and glucose transport. *Trends Endocrinol Metab* 2006; **17**: 72-78 [PMID: 16458527 DOI: 10.1016/j.tem.2006.01.005]
- 42 **Zheng Y**, Yamada H, Sakamoto K, Horita S, Kunimi M, Endo Y, Li Y, Tobe K, Terauchi Y, Kadowaki T, Seki G, Fujita T. Roles of insulin receptor substrates in insulin-induced stimulation of renal proximal bicarbonate absorption. *J Am Soc Nephrol* 2005; **16**: 2288-2295 [PMID: 15975995 DOI: 10.1681/ASN.2005020193]
- 43 **Pavlov TS**, Ilatovskaya DV, Levchenko V, Li L, Ecelbarger CM, Staruschenko A. Regulation of ENaC in mice lacking renal insulin receptors in the collecting duct. *FASEB J* 2013; **27**: 2723-2732 [PMID: 23558339]
- 44 **Taniguchi CM**, Emanuelli B, Kahn CR. Critical nodes in signalling pathways: insights into insulin action. *Nat Rev Mol Cell Biol* 2006; **7**: 85-96 [PMID: 16493415 DOI: 10.1038/nrm1837]
- 45 **Vigneri R**, Squatrito S, Sciacca L. Insulin and its analogs: actions via insulin and IGF receptors. *Acta Diabetol* 2010; **47**: 271-278 [PMID: 20730455 DOI: 10.1007/s00592-010-0215-3]
- 46 **Crowley SD**, Gurley SB, Herrera MJ, Ruiz P, Griffiths R, Kumar AP, Kim HS, Smithies O, Le TH, Coffman TM. Angiotensin II causes hypertension and cardiac hypertrophy through its receptors in the kidney. *Proc Natl Acad Sci USA* 2006; **103**: 17985-17990 [PMID: 17090678 DOI: 10.1073/pnas.0605545103]
- 47 **Gurley SB**, Riquier-Brison AD, Schnermann J, Sparks MA, Allen AM, Haase VH, Snouwaert JN, Le TH, McDonough AA, Koller BH, Coffman TM. AT1A angiotensin receptors in the renal proximal tubule regulate blood pressure. *Cell Metab* 2011; **13**: 469-475 [PMID: 21459331 DOI: 10.1016/j.cmet.2011.03.001]
- 48 **Harris PJ**, Young JA. Dose-dependent stimulation and inhibition of proximal tubular sodium reabsorption by angiotensin II in the rat kidney. *Pflugers Arch* 1977; **367**: 295-297 [PMID: 556854 DOI: 10.1007/BF00581370]
- 49 **Horita S**, Zheng Y, Hara C, Yamada H, Kunimi M, Taniguchi S, Uwatoko S, Sugaya T, Goto A, Fujita T, Seki G. Biphasic regulation of Na-HCO<sub>3</sub> cotransporter by angiotensin II type 1A receptor. *Hypertension* 2002; **40**: 707-712 [PMID: 12411466 DOI: 10.1161/01.HYP.0000036449.70110.DE]
- 50 **Schuster VL**, Kokko JP, Jacobson HR. Angiotensin II directly stimulates sodium transport in rabbit proximal convoluted tubules. *J Clin Invest* 1984; **73**: 507-515 [PMID: 6699174 DOI: 10.1172/JCI11237]
- 51 **Zheng Y**, Horita S, Hara C, Kunimi M, Yamada H, Sugaya T, Goto A, Fujita T, Seki G. Biphasic regulation of renal proximal bicarbonate absorption by luminal AT(1A) receptor. *J Am Soc Nephrol* 2003; **14**: 1116-1122 [PMID: 12707382]
- 52 **Haithecock D**, Jiao H, Cui XL, Hopfer U, Douglas JG. Renal proximal tubular AT2 receptor: signaling and transport. *J Am Soc Nephrol* 1999; **10** Suppl 11: S69-S74 [PMID: 9892143]
- 53 **Houillier P**, Chambrey R, Achard JM, Froissart M, Poggioli J, Paillard M. Signaling pathways in the biphasic effect of angiotensin II on apical Na/H antiporter activity in proximal tubule. *Kidney Int* 1996; **50**: 1496-1505 [PMID: 8914015 DOI: 10.1038/ki.1996.464]
- 54 **Poggioli J**, Lazar G, Houillier P, Gardin JP, Achard JM, Paillard M. Effects of angiotensin II and nonpeptide receptor antagonists on transduction pathways in rat proximal tubule. *Am J Physiol* 1992;

- 263: C750-C758 [PMID: 1329542]
- 55 **Li Y**, Yamada H, Kita Y, Kunimi M, Horita S, Suzuki M, Endo Y, Shimizu T, Seki G, Fujita T. Roles of ERK and cPLA2 in the angiotensin II-mediated biphasic regulation of Na<sup>+</sup>-HCO<sub>3</sub><sup>-</sup> transport. *J Am Soc Nephrol* 2008; **19**: 252-259 [PMID: 18094367 DOI: 10.1681/ASN.2007030289]
- 56 **Liu FY**, Cogan MG. Angiotensin II stimulates early proximal bicarbonate absorption in the rat by decreasing cyclic adenosine monophosphate. *J Clin Invest* 1989; **84**: 83-91 [PMID: 2544631 DOI: 10.1172/JCI114174]
- 57 **Zhang C**, Mayeux PR. NO/cGMP signaling modulates regulation of Na<sup>+</sup>-K<sup>+</sup>-ATPase activity by angiotensin II in rat proximal tubules. *Am J Physiol Renal Physiol* 2001; **280**: F474-F479 [PMID: 11181409]
- 58 **Shirai A**, Yamazaki O, Horita S, Nakamura M, Satoh N, Yamada H, Suzuki M, Kudo A, Kawakami H, Hofmann F, Nishiyama A, Kume H, Enomoto Y, Homma Y, Seki G. Angiotensin II dose-dependently stimulates human renal proximal tubule transport by the nitric oxide/guanosine 3',5'-cyclic monophosphate pathway. *J Am Soc Nephrol* 2014; **25**: 1523-1532 [PMID: 24511122 DOI: 10.1681/ASN.2013060596]
- 59 **Shultz PJ**, Tolins JP. Adaptation to increased dietary salt intake in the rat. Role of endogenous nitric oxide. *J Clin Invest* 1993; **91**: 642-650 [PMID: 7679414 DOI: 10.1172/JCI116244]
- 60 **Tolins JP**, Shultz PJ. Endogenous nitric oxide synthesis determines sensitivity to the pressor effect of salt. *Kidney Int* 1994; **46**: 230-236 [PMID: 7523754 DOI: 10.1038/ki.1994.264]
- 61 **Ortiz PA**, Garvin JL. Role of nitric oxide in the regulation of nephron transport. *Am J Physiol Renal Physiol* 2002; **282**: F777-F784 [PMID: 11934686]
- 62 **Facchini FS**, DoNascimento C, Reaven GM, Yip JW, Ni XP, Humphreys MH. Blood pressure, sodium intake, insulin resistance, and urinary nitrate excretion. *Hypertension* 1999; **33**: 1008-1012 [PMID: 10205239 DOI: 10.1161/01.HYP.33.4.1008]
- 63 **Schmidt RJ**, Beierwaltes WH, Baylis C. Effects of aging and alterations in dietary sodium intake on total nitric oxide production. *Am J Kidney Dis* 2001; **37**: 900-908 [PMID: 11325670]
- 64 **Parker JD**, Farrell B, Fenton T, Cohan M, Parker JO. Counter-regulatory responses to continuous and intermittent therapy with nitroglycerin. *Circulation* 1991; **84**: 2336-2345 [PMID: 1835676 DOI: 10.1161/01.CIR.84.6.2336]
- 65 **White WB**, Halley SE. Comparative renal effects of intravenous administration of fenoldopam mesylate and sodium nitroprusside in patients with severe hypertension. *Arch Intern Med* 1989; **149**: 870-874 [PMID: 2565105 DOI: 10.1001/archinte.1989.00390040080016]
- 66 **Sarafidis PA**, Lasaridis AN. Diabetic nephropathy: Endothelin antagonism for diabetic nephropathy. *Nat Rev Nephrol* 2010; **6**: 447-449 [PMID: 20664627 DOI: 10.1038/nrneph.2010.86]

**P- Reviewer:** Duan SB, Trimarchi H **S- Editor:** Tian YL  
**L- Editor:** A **E- Editor:** Lu YJ





Published by **Baishideng Publishing Group Inc**

8226 Regency Drive, Pleasanton, CA 94588, USA

Telephone: +1-925-223-8242

Fax: +1-925-223-8243

E-mail: [bpgoffice@wjgnet.com](mailto:bpgoffice@wjgnet.com)

Help Desk: <http://www.wjgnet.com/esps/helpdesk.aspx>

<http://www.wjgnet.com>

