

Influence of gut bacteria on development and progression of non-alcoholic fatty liver disease

Ali Abdul-Hai, Ali Abdallah, Stephen DH Malnick

Ali Abdul-Hai, Ali Abdallah, Stephen DH Malnick, Division of Internal Medicine, Kaplan Medical Center, Affiliated to the Hebrew University, Rehovot 76100, Israel

Author contributions: All the authors were involved in the writing of various sections of the manuscript; Malnick SDH conceived the idea of writing the review.

Conflict-of-interest: The authors have no conflict of interest to declare.

Open-Access: This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

Correspondence to: Dr. Stephen DH Malnick, Division of Internal Medicine, Kaplan Medical Center, Affiliated to the Hebrew University, 1 Pasternak, Rehovot 76100, Israel. stephen@malnick.net
 Telephone: +972-89-441371
 Fax: +972-89-441852

Received: December 17, 2014
 Peer-review started: December 18, 2014
 First decision: March 6, 2015
 Revised: April 8, 2015
 Accepted: April 16, 2015
 Article in press: April 20, 2015
 Published online: June 28, 2015

Abstract

The intestine of the human contains a dynamic population of microbes that have a symbiotic relationship with the host. In addition, there is an effect of the intestinal microbiota on metabolism and digestion. Non-alcoholic fatty liver disease (NAFLD) is a common cause worldwide

of hepatic pathology and is thought to be the hepatic manifestation of the metabolic syndrome. In this review we examine the effect of the human microbiome on the components and pathogenesis of the metabolic syndrome. We are now on the threshold of therapeutic interventions on the human microbiome in order to effect human disease including NAFLD.

Key words: Microbiome; Metabolic syndrome; Stool transplantation; Non-alcoholic fatty liver disease

© **The Author(s) 2015.** Published by Baishideng Publishing Group Inc. All rights reserved.

Core tip: The human intestine contains more bacterial cells than mammalian cells. These have a symbiotic relationship with the host. Non-alcoholic fatty liver disease is the hepatic manifestation of the metabolic syndrome and a major cause of hepatic morbidity as a consequence of the obesity epidemic. We examine the effect of the human microbiome on the components of the metabolic syndrome and fatty liver and mention the possibility of therapeutic interventions in humans.

Abdul-Hai A, Abdallah A, Malnick SDH. Influence of gut bacteria on development and progression of non-alcoholic fatty liver disease. *World J Hepatol* 2015; 7(12): 1679-1684 Available from: URL: <http://www.wjgnet.com/1948-5182/full/v7/i12/1679.htm> DOI: <http://dx.doi.org/10.4254/wjh.v7.i12.1679>

INTRODUCTION

Non-alcoholic fatty liver disease (NAFLD) is considered the hepatic manifestation of the metabolic syndrome. The metabolic syndrome is defined by clear clinical and laboratory criteria (Table 1). NAFLD encompasses a range of liver damage ranging from simple steatosis to non-alcoholic steatohepatitis (NASH), fibrosis, cirrhosis and its complications. NAFLD is present in approximately

Table 1 The definitions of the metabolic syndrome

	NCEP ATP III	IDF
Absolutely required	None	Central obesity (waist circumference) ≥ 94 cm in males or ≥ 80 cm in females European origin ≥ 90 cm in males or ≥ 80 cm in females Central obesity plus two of the four criteria below
Criteria	Any three of the five criteria below	
Obesity	(1) waist circumference > 40 inches in males, or > 35 inches in females	
Hyperglycemia	(2) fasting glucose ≥ 100 mg/dL or treated for DM	(1) fasting glucose ≥ 100 mg/dL
Dyslipidemia	(3) TG ≥ 150 mg/dL or treated for dyslipidemia Or (4) HDL cholesterol < 40 mg/dL in males, or < 50 mg/dL in females or under treatment	(2) TG ≥ 150 mg/dL or treated for dyslipidemia Or (3) HDL cholesterol < 40 mg/dL in males, or < 50 mg/dL in females or under treatment
Hypertension	(5) > 130 mmHg systolic or > 85 mmHg diastolic or treated for HTN	(4) > 130 mmHg systolic or > 85 mmHg diastolic or treated for HTN

NCEP ATP III: National Cholesterol and Education Program - Adult Treatment Panel III; IDF: International diabetes federation; DM: Diabetes mellitus; TG: Triglycerides; HTN: Hypertension.

1/3 of the United States population, who have isolated steatosis of the liver^[1]. Of the patients with NAFLD, approximately 30% have NASH^[2]. NASH refers to those patients who have developed liver inflammation and fibrosis. It is those patients with NASH who may develop stage 3 or 4 fibrosis (cirrhosis)^[3].

Many factors including diet, sedentary lifestyle and genetics have been shown to influence the progression from steatosis through NASH to cirrhosis. However, not all people who are obese develop NAFLD and neither are all patients with NAFLD obese.

GUT MICROBIOTA

The intestinal microbiome is attracting an increasing amount of attention^[4]. It is becoming apparent that there is a symbiotic relationship between the intestine and its microbiota and that disturbance in this relationship can be associated with the pathogenesis of many disorders. The most striking example of such an association is *Clostridium difficile* infection, for which fecal transplantation from healthy donors is now an accepted treatment^[5].

Distinct gut microbiota profiles are linked with specific metabolomes. Ninety-five percent of the gut microbiota of humans consists of the *Firmicutes*, *Bacteroidetes* and *Actinobacteria* phyla. The species level of the human microbiota, however, has higher diversity, with approximately 200 highly prevalent and up to 1000 less common bacterial species^[6]. In humans as in mice, each individual has a unique bacterial species profile^[7]. The gut bacteria may alter in response to a high fat diet (HFD), which could be responsible for some of the responses to an HFD.

Bacteria from human stools can be transferred to germ free (GF) mice and result in a similar microbiome in the host mice^[8]. This can result in the appearance of human gut enzymatic activities in GF rodents after human fecal transplantation^[9,10].

Recently, the transfer of human gut microbiome from obesity discordant twins to GF mice was shown to result

in the transfer of the adiposity phenotype of the donor twin^[11]. Thus, the transfer of human fecal microbiota to GF mice may result in the development of human diseases and provide an experimental study system.

INTESTINAL MICROBIOTA ARE RELATED TO OBESITY AND INSULIN RESISTANCE

The gut microbiota is now recognized as contributing to obesity and NAFLD^[12]. GF mice have been found to gain less weight than conventional mice after being fed a high sugar and fat diet in spite of a higher amount of food consumption^[13,14]. Furthermore, GF mice on an HFD develop an increase in insulin sensitivity^[15] and GF mice colonized with conventional mouse intestinal microbiota develop an increase in body fat content^[13]. There are, however, wide variations in the development of HFD-associated features^[16,17], but the responsible factors are still undefined.

The insulin resistance index can be transferred by gut microbiota transplantation^[18]. Gut microbiota affects both macrophage fat accumulation and systemic glucose metabolism by different mechanisms^[19]. In a diet-induced obesity mouse model, administration of antibiotics improved fasting glycemia and insulin resistance independently of both food intake or adiposity^[20]. Furthermore the improved insulin sensitivity correlated with less hepatic lipogenesis and steatosis in the antibiotic-treated mice^[21]. Taken together, these findings suggest that the gut microbiota influences both host glucose metabolism and liver function.

A study in humans showed that transfer of intestinal microbiota from lean donors to males with the metabolic syndrome resulted in increased insulin sensitivity^[21]. Dietary factors and changes in diet influence the composition of the microbiome. The intestinal microbiota of obese individuals has a different microbial diversity compared to lean persons. They have less *Bacteroides* and more *Firmicutes*^[22]. Furthermore, an HFD increases the proportion of Gram-negative to

Gram-positive microbes, resulting in the production of lipopolysaccharide (LPS) which is responsible for inflammation^[23]. Gram-positive microbes are increased following the administration of prebiotics^[24]. A prebiotic is a nondigestible food substrate which increases the growth of intestinal bacteria that can result in health benefits for the host.

The intestinal microbiome in obesity has an increased capacity to extract energy from the host diet. Bacterial enzymes extract calories from otherwise indigestible dietary polysaccharides^[25]. Enteric bacteria suppress the synthesis and secretion of small intestinal fasting-induced adipocyte factor, resulting in an increased activity of lipoprotein lipase and increased liver triglyceride^[13,14].

GF lean mice that were resistant to becoming obese on a fat-enriched diet had an increase of phosphorylated adenosine monophosphate-activated protein kinase (AMPK) in both the skeletal muscle and liver. AMPK phosphorylates acetyl coenzyme A (CoA) carboxylase, resulting in decreased malonyl CoA levels. Malonyl CoA controls the rate-limiting step of long-chain fatty acyl CoA entry to the mitochondria by blocking carnitine palmitoyltransferase which promotes the oxidation of fatty acid and results in a lower storage of fat^[14,26].

Thus, the intestinal microbiome has an effect on both obesity and insulin resistance, as well as hepatic fat content.

GUT MICROBIOTA AND NAFLD

In view of the intimate connection between the metabolic syndrome with its concomitant insulin resistance and NAFLD, it is expected that there is an effect of the intestinal microbiome on NAFLD.

The fecal microbiota in NAFLD and NASH patients has been examined using quantitative polymerase chain reaction (PCR) and deep sequencing of a conserved region in the bacterial 16S ribosomal RNA gene^[27-30]. A recent review provides a summary of the changes in the intestinal microbiota associated with NAFLD and NASH^[12]. Many of these studies have variable and often contradictory findings. This may be due to differences in patient mix, methodology and documentation of liver disease.

In addition to the mixture of bacteria in the colon, patients with obesity or NAFLD have more small intestinal bacterial overgrowth^[31,32]. Small intestinal bacterial overgrowth was found in 50% of patients with NASH, significantly more than that in a control population^[33]. The intestinal permeability and bacterial overgrowth were shown to be related to the degree of hepatic steatosis but not inflammation or fibrosis^[31].

However, it is not clear if the assessment of small bowel bacterial overgrowth by breath tests is accurate since an estimate of total fecal bacterial count by real-time PCR did not detect any difference between healthy controls and patients with NAFLD and NASH^[28].

Possible mediators of the link between the enteric microbiome and the host include alcohol, choline

and endotoxins. Obese animals have been shown to have higher levels of alcohol in breath tests than thin animals^[34]. Alcohol reaches the liver *via* the portal blood and can cause triglyceride accumulation in hepatocytes^[35]. In addition, alcohol may provide the "second hit" to the liver for making the transformation from steatosis to steatohepatitis^[36].

Choline may also be involved in the development of NAFLD and NASH. It is well known that choline deficiency may result in chronic liver disease^[37]. In animal models choline-deficient diets were utilized, but it is now known that choline deficiency can exist while there is a diet that is not deficient. HFDs produce intestinal microbiota that converts dietary choline into methylamines. This results in a reduction of serum level of phosphatidylcholine which can cause NASH^[26]. Phosphatidylcholine is important for the production of very low-density lipoprotein (VLDL)^[38] and thus choline deficiency secondary to the intestinal microbiome will result in lower hepatic secretion of VLDL and result in triglyceride accumulation in hepatocytes.

The products of the intestinal microbiota are also implicated in the development of NAFLD and NASH. Endotoxemia has been found in patients with NASH^[39]. Toll-like receptor 4, a receptor for LPS, in hematopoietic-derived cells is necessary for the development of hepatic steatosis but not for obesity in mice^[40]. Mice that are deficient in sensing pathogen-associated molecular patterns (PAMPs) or downstream signaling are resistant to NASH^[41,42].

The microbial products reach the liver *via* the portal vein and cause inflammation. Mice that are genetically obese are more sensitive to endotoxin-induced hepatotoxicity and develop steatohepatitis after being exposed to low doses of LPS^[43]. NAFLD patients have an increased intestinal permeability and changes in the intestinal tight junctions, as compared to healthy individuals^[31]. The increased permeability, in combination with bacterial overgrowth, increases the hepatic exposure to endotoxins.

Alteration of the fecal microbiome by administration of probiotics has been shown to decrease the amount of intrahepatic triglyceride content in addition to a decrease in *Firmicutes* and an increase in *Bacteroidetes*^[30]. A meta-analysis of the published trials of probiotics in patients with NAFLD, showed a reduction in serum transaminases, total cholesterol, tumor necrosis factor- α and an improvement in insulin resistance^[44].

Dysbiosis can induce intestinal inflammation. Indeed GF mice are protected from inflammation of the small intestine^[45]. Mice deficient in Nlrp3 and Nlrp6 are unable to form cytoplasmic multiprotein complexes composed of nucleotide-binding domain and leucine-rich repeat-containing proteins (NLR) family, inflammasomes. Inflammasomes are sensors of exogenous PAMPs that regulate cleavage of precursors of inflammatory cytokines including pro-interleukin 1 beta (pro-IL1 β) and pro-IL18. In mice, loss of Nlrp3 and Nlrp6 inflammasomes is associated with intestinal dysbiosis and colonic inflammation *via* CCL5. Dysbiosis is linked to an increase in *Prevotella*^[46]. The consequent translocation of bacteria

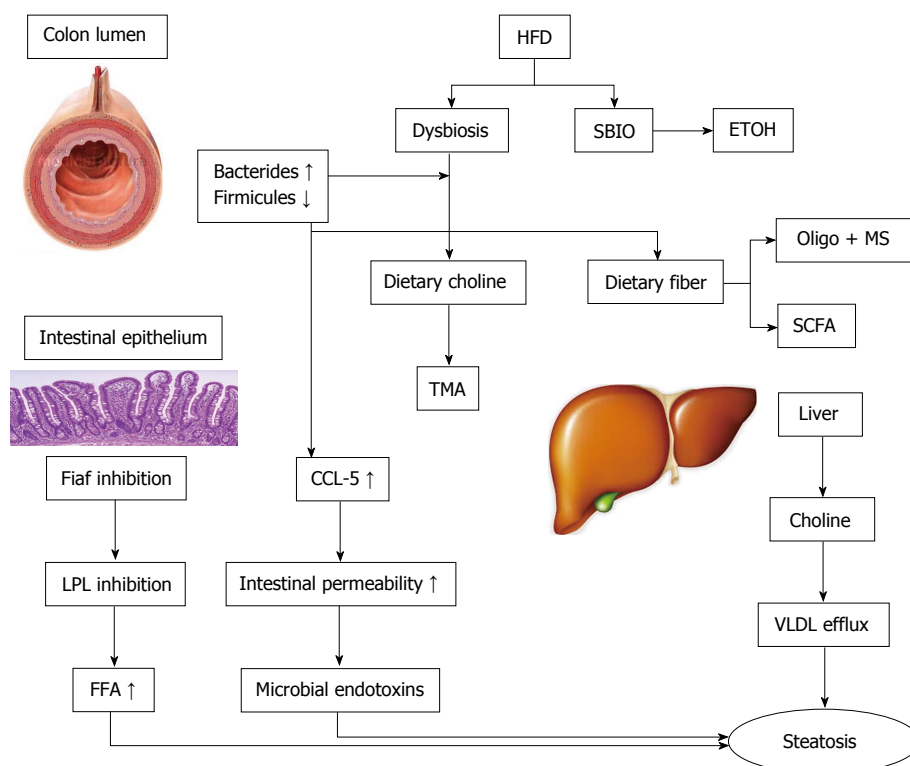


Figure 1 The effect of the intestinal microbiota on non-alcoholic fatty liver disease. High fat diets (HFD) produce dysbiosis and small bowel intestinal overgrowth (SBIO). There is an increase in energy extraction and fermentation of dietary fibers to oligo- and mono-saccharides and short chain fatty acids (SCFA). There is also an increase in ethanol (ETOH) production. The microbiota metabolize choline to trimethylamine (TMA). There is a choline deficiency which decreases very low-density lipoprotein (VLDL) efflux and hepatic steatosis. In addition the intestinal microbiota suppresses the production of fasting induced adipocyte factor (Fiaf) in intestinal epithelia, which increases the activity of lipoprotein lipase and the levels of free fatty acids (FFA). Dysbiosis results in a disruption of tight junctions in the enterocytes via chemokine (C-C motif) ligand 5 (CCL-5). The resulting increase in intestinal permeability results in the translocation of microbial products to the liver and inflammation; MS: Monosaccharides; LPL: Lipoprotein lipase.

leads to an increase in bacterial products including LPS and bacterial DNA in the portal vein. The ensuing hepatic inflammatory response promotes progression of NAFLD to NASH (Figure 1). This change in phenotype can be transmitted by co-housing wild-type and NASH-prone mice^[46].

Thus, intestinal dysbiosis can induce colonic inflammation and bacterial translocation which accelerates the progression of simple steatosis to NASH. As a result of these findings attention is beginning to be directed at fecal microbiota transplantation (FMT). FMT was first used in China more than 1500 years ago^[47]. In 1958, 4 cases of treatment of pseudomembranous colitis by fecal enemas were reported^[48]. This is now an established treatment^[49]. At present there is only one report of FMT for metabolic syndrome. Vrieze *et al.*^[21] reported 18 patients with the metabolic syndrome who underwent a stool transplant that was either autologous or from lean healthy volunteers. Six weeks following the FMT there was a significant increase in insulin sensitivity together with an increase in the levels of butyrate-producing intestinal microbiota.

In summary, there appears to be an effect of the fecal microbiome on the development of the metabolic syndrome and its hepatic manifestation NAFLD and NASH. Further investigation of this relationship will increase our understanding of this connection. There

is evidence that manipulation of the fecal microbiome may result in a change in the metabolic syndrome and an improvement in the features of NAFLD. This needs to be explored further in order to investigate if there will be an improvement in clinically significant end points.

REFERENCES

- 1 **Browning JD**, Szczepaniak LS, Dobbins R, Nuremberg P, Horton JD, Cohen JC, Grundy SM, Hobbs HH. Prevalence of hepatic steatosis in an urban population in the United States: impact of ethnicity. *Hepatology* 2004; **40**: 1387-1395 [PMID: 15565570 DOI: 10.1002/hep.20466]
- 2 **Williams CD**, Stengel J, Asike MI, Torres DM, Shaw J, Contreras M, Landt CL, Harrison SA. Prevalence of nonalcoholic fatty liver disease and nonalcoholic steatohepatitis among a largely middle-aged population utilizing ultrasound and liver biopsy: a prospective study. *Gastroenterology* 2011; **140**: 124-131 [PMID: 20858492 DOI: 10.1053/j.gastro.2010.09.038]
- 3 **Caldwell S**, Argo C. The natural history of non-alcoholic fatty liver disease. *Dig Dis* 2010; **28**: 162-168 [PMID: 20460906]
- 4 **Nicholson JK**, Holmes E, Kinross J, Burcelin R, Gibson G, Jia W, Pettersson S. Host-gut microbiota metabolic interactions. *Science* 2012; **336**: 1262-1267 [PMID: 22674330 DOI: 10.1126/science.1223813]
- 5 **Floch MH**. The power of poop: probiotics and fecal microbial transplant. *J Clin Gastroenterol* 2012; **46**: 625-626 [PMID: 22874805 DOI: 10.1097/MCG.0b013e3182667a93]
- 6 **Ley RE**, Lozupone CA, Hamady M, Knight R, Gordon JI. Worlds within worlds: evolution of the vertebrate gut microbiota. *Nat Rev Microbiol* 2008; **6**: 776-788 [PMID: 18794915 DOI: 10.1038/

- nrmicro1978]
- 7 **Zoetendal EG**, Akkermans AD, De Vos WM. Temperature gradient gel electrophoresis analysis of 16S rRNA from human fecal samples reveals stable and host-specific communities of active bacteria. *Appl Environ Microbiol* 1998; **64**: 3854-3859 [PMID: 9758810]
 - 8 **Turnbaugh PJ**, Ridaura VK, Faith JJ, Rey FE, Knight R, Gordon JI. The effect of diet on the human gut microbiome: a metagenomic analysis in humanized gnotobiotic mice. *Sci Transl Med* 2009; **1**: 6ra14 [PMID: 20368178 DOI: 10.1126/scitranslmed.3000322]
 - 9 **Bowey E**, Adlercreutz H, Rowland I. Metabolism of isoflavones and lignans by the gut microflora: a study in germ-free and human flora associated rats. *Food Chem Toxicol* 2003; **41**: 631-636 [PMID: 12659715]
 - 10 **Gérard P**, Béguet F, Lepercq P, Rigottier-Gois L, Rochet V, Andrieux C, Juste C. Gnotobiotic rats harboring human intestinal microbiota as a model for studying cholesterol-to-coprostanol conversion. *FEMS Microbiol Ecol* 2004; **47**: 337-343 [PMID: 19712322 DOI: 10.1016/S0168-6496(03)00285-X]
 - 11 **Ridaura VK**, Faith JJ, Rey FE, Cheng J, Duncan AE, Kau AL, Griffin NW, Lombard V, Henrissat B, Bain JR, Muehlbauer MJ, Ilkayeva O, Semenkovich CF, Funai K, Hayashi DK, Lyle BJ, Martini MC, Ursell LK, Clemente JC, Van Treuren W, Walters WA, Knight R, Newgard CB, Heath AC, Gordon JI. Gut microbiota from twins discordant for obesity modulate metabolism in mice. *Science* 2013; **341**: 1241214 [PMID: 24009397 DOI: 10.1126/science.1241214]
 - 12 **Schnabl B**, Brenner DA. Interactions between the intestinal microbiome and liver diseases. *Gastroenterology* 2014; **146**: 1513-1524 [PMID: 24440671 DOI: 10.1053/j.gastro.2014.01.020]
 - 13 **Bäckhed F**, Ding H, Wang T, Hooper LV, Koh GY, Nagy A, Semenkovich CF, Gordon JI. The gut microbiota as an environmental factor that regulates fat storage. *Proc Natl Acad Sci USA* 2004; **101**: 15718-15723 [PMID: 15505215 DOI: 10.1073/pnas.0407076101]
 - 14 **Bäckhed F**, Manchester JK, Semenkovich CF, Gordon JI. Mechanisms underlying the resistance to diet-induced obesity in germ-free mice. *Proc Natl Acad Sci USA* 2007; **104**: 979-984 [PMID: 17210919 DOI: 10.1073/pnas.0605374104]
 - 15 **Rabot S**, Membrez M, Bruneau A, Gérard P, Harach T, Moser M, Raymond F, Mansourian R, Chou CJ. Germ-free C57BL/6J mice are resistant to high-fat-diet-induced insulin resistance and have altered cholesterol metabolism. *FASEB J* 2010; **24**: 4948-4959 [PMID: 20724524 DOI: 10.1096/fj.10-164921]
 - 16 **Jang I**, Hwang D, Lee J, Chae K, Kim Y, Kang T, Kim C, Shin D, Hwang J, Huh Y, Cho J. Physiological difference between dietary obesity-susceptible and obesity-resistant Sprague Dawley rats in response to moderate high fat diet. *Exp Anim* 2003; **52**: 99-107 [PMID: 12806884]
 - 17 **Li H**, Xie Z, Lin J, Song H, Wang Q, Wang K, Su M, Qiu Y, Zhao T, Song K, Wang X, Zhou M, Liu P, Zhao G, Zhang Q, Jia W. Transcriptomic and metabonomic profiling of obesity-prone and obesity-resistant rats under high fat diet. *J Proteome Res* 2008; **7**: 4775-4783 [PMID: 18828625]
 - 18 **Le Roy T**, Llopis M, Lepage P, Bruneau A, Rabot S, Bevilacqua C, Martin P, Philippe C, Walker F, Bado A, Perlemuter G, Cassard-Doulcier AM, Gérard P. Intestinal microbiota determines development of non-alcoholic fatty liver disease in mice. *Gut* 2013; **62**: 1787-1794 [PMID: 23197411 DOI: 10.1136/gutjnl-2012-303816]
 - 19 **Caesar R**, Reigstad CS, Bäckhed HK, Reinhardt C, Ketonen M, Lundén GÖ, Cani PD, Bäckhed F. Gut-derived lipopolysaccharide augments adipose macrophage accumulation but is not essential for impaired glucose or insulin tolerance in mice. *Gut* 2012; **61**: 1701-1707 [PMID: 22535377 DOI: 10.1136/gutjnl-2011-301689]
 - 20 **Membrez M**, Blancher F, Jaquet M, Bibiloni R, Cani PD, Burcelin RG, Corthesy I, Macé K, Chou CJ. Gut microbiota modulation with norfloxacin and ampicillin enhances glucose tolerance in mice. *FASEB J* 2008; **22**: 2416-2426 [PMID: 18326786 DOI: 10.1096/fj.07-102723]
 - 21 **Vrieze A**, Van Nood E, Holleman F, Salojärvi J, Kootte RS, Bartelsman JF, Dallinga-Thie GM, Ackermans MT, Serlie MJ, Oozeer R, Derrien M, Druenes A, Van Hylckama Vlieg JE, Bloks VW, Groen AK, Heilig HG, Zoetendal EG, Stoes ES, de Vos WM, Hoekstra JB, Nieuwdorp M. Transfer of intestinal microbiota from lean donors increases insulin sensitivity in individuals with metabolic syndrome. *Gastroenterology* 2012; **143**: 913-6.e7 [PMID: 22728514 DOI: 10.1053/j.gastro.2012.06.031]
 - 22 **Turnbaugh PJ**, Bäckhed F, Fulton L, Gordon JI. Diet-induced obesity is linked to marked but reversible alterations in the mouse distal gut microbiome. *Cell Host Microbe* 2008; **3**: 213-223 [PMID: 18407065 DOI: 10.1016/j.chom.2008.02.015]
 - 23 **Cani PD**, Amar J, Iglesias MA, Poggi M, Knauf C, Bastelica D, Neyrinck AM, Fava F, Tuohy KM, Chabo C, Waget A, Delmée E, Cousin B, Sulpice T, Chamontin B, Ferrières J, Tanti JF, Gibson GR, Casteilla L, Delzenne NM, Alessi MC, Burcelin R. Metabolic endotoxemia initiates obesity and insulin resistance. *Diabetes* 2007; **56**: 1761-1772 [PMID: 17456850 DOI: 10.2337/db06-1491]
 - 24 **Cani PD**, Neyrinck AM, Fava F, Knauf C, Burcelin RG, Tuohy KM, Gibson GR, Delzenne NM. Selective increases of bifidobacteria in gut microflora improve high-fat-diet-induced diabetes in mice through a mechanism associated with endotoxaemia. *Diabetologia* 2007; **50**: 2374-2383 [PMID: 17823788 DOI: 10.1007/s00125-007-0791-0]
 - 25 **Turnbaugh PJ**, Ley RE, Mahowald MA, Magrini V, Mardis ER, Gordon JI. An obesity-associated gut microbiome with increased capacity for energy harvest. *Nature* 2006; **444**: 1027-1031 [PMID: 17183312 DOI: 10.1038/nature05414]
 - 26 **Dumas ME**, Barton RH, Toye A, Cloarec O, Blancher C, Rothwell A, Fearnside J, Tatoud R, Blanc V, Lindon JC, Mitchell SC, Holmes E, McCarthy MI, Scott J, Gauguier D, Nicholson JK. Metabolic profiling reveals a contribution of gut microbiota to fatty liver phenotype in insulin-resistant mice. *Proc Natl Acad Sci USA* 2006; **103**: 12511-12516 [PMID: 16895997 DOI: 10.1073/pnas.0601056103]
 - 27 **Raman M**, Ahmed I, Gillevet PM, Probert CS, Ratcliffe NM, Smith S, Greenwood R, Sikaroodi M, Lam V, Crotty P, Bailey J, Myers RP, Rioux KP. Fecal microbiome and volatile organic compound metabolome in obese humans with nonalcoholic fatty liver disease. *Clin Gastroenterol Hepatol* 2013; **11**: 868-875.e1-3 [PMID: 23454028]
 - 28 **Mouzaki M**, Comelli EM, Arendt BM, Bonengel J, Fung SK, Fischer SE, McGilvray ID, Allard JP. Intestinal microbiota in patients with nonalcoholic fatty liver disease. *Hepatology* 2013; **58**: 120-127 [PMID: 23401313]
 - 29 **Zhu L**, Baker SS, Gill C, Liu W, Alkhouri R, Baker RD, Gill SR. Characterization of gut microbiomes in nonalcoholic steatohepatitis (NASH) patients: a connection between endogenous alcohol and NASH. *Hepatology* 2013; **57**: 601-609 [PMID: 23055155 DOI: 10.1002/hep.26093]
 - 30 **Wong VW**, Tse CH, Lam TT, Wong GL, Chim AM, Chu WC, Yeung DK, Law PT, Kwan HS, Yu J, Sung JJ, Chan HL. Molecular characterization of the fecal microbiota in patients with nonalcoholic steatohepatitis—a longitudinal study. *PLoS One* 2013; **8**: e62885 [PMID: 23638162 DOI: 10.1371/journal.pone.0062885]
 - 31 **Miele L**, Valenza V, La Torre G, Montalto M, Cammarota G, Ricci R, Mascianà R, Forgione A, Gabrieli ML, Perotti G, Vecchio FM, Rapaccini G, Gasbarrini G, Day CP, Grieco A. Increased intestinal permeability and tight junction alterations in nonalcoholic fatty liver disease. *Hepatology* 2009; **49**: 1877-1887 [PMID: 19291785 DOI: 10.1002/hep.22848]
 - 32 **Sabaté JM**, Jouët P, Harnois F, Mechler C, Msika S, Grossin M, Coffin B. High prevalence of small intestinal bacterial overgrowth in patients with morbid obesity: a contributor to severe hepatic steatosis. *Obes Surg* 2008; **18**: 371-377 [PMID: 18286348 DOI: 10.1007/s11695-007-9398-2]
 - 33 **Wigg AJ**, Roberts-Thomson IC, Dymock RB, McCarthy PJ, Grose RH, Cummins AG. The role of small intestinal bacterial overgrowth, intestinal permeability, endotoxaemia, and tumour necrosis factor alpha in the pathogenesis of non-alcoholic steatohepatitis. *Gut* 2001; **48**: 206-211 [PMID: 11156641]
 - 34 **Cope K**, Risby T, Diehl AM. Increased gastrointestinal ethanol

- production in obese mice: implications for fatty liver disease pathogenesis. *Gastroenterology* 2000; **119**: 1340-1347 [PMID: 11054393]
- 35 **Hartmann P**, Chen WC, Schnabl B. The intestinal microbiome and the leaky gut as therapeutic targets in alcoholic liver disease. *Front Physiol* 2012; **3**: 402 [PMID: 23087650 DOI: 10.3389/fphys.2012.00402]
 - 36 **Day CP**. Pathogenesis of steatohepatitis. *Best Pract Res Clin Gastroenterol* 2002; **16**: 663-678 [PMID: 12406438]
 - 37 **Blumberg H**, McCollum EV. The prevention by choline of liver cirrhosis in rats on high fat, low protein diets. *Science* 1941; **93**: 598-599 [PMID: 17795968 DOI: 10.1126/science.93.2425.598]
 - 38 **Jiang XC**, Li Z, Liu R, Yang XP, Pan M, Lagrost L, Fisher EA, Williams KJ. Phospholipid transfer protein deficiency impairs apolipoprotein-B secretion from hepatocytes by stimulating a proteolytic pathway through a relative deficiency of vitamin E and an increase in intracellular oxidants. *J Biol Chem* 2005; **280**: 18336-18340 [PMID: 15734742 DOI: 10.1074/jbc.M500007200]
 - 39 **Farhadi A**, Gundlapalli S, Shaikh M, Frantzides C, Harrell L, Kwasny MM, Keshavarzian A. Susceptibility to gut leakiness: a possible mechanism for endotoxaemia in non-alcoholic steatohepatitis. *Liver Int* 2008; **28**: 1026-1033 [PMID: 18397235 DOI: 10.1111/j.1478-3231.2008.01723.x]
 - 40 **Saberi M**, Woods NB, de Luca C, Schenk S, Lu JC, Bandyopadhyay G, Verma IM, Olefsky JM. Hematopoietic cell-specific deletion of toll-like receptor 4 ameliorates hepatic and adipose tissue insulin resistance in high-fat-fed mice. *Cell Metab* 2009; **10**: 419-429 [PMID: 19883619 DOI: 10.1016/j.cmet.2009.09.006]
 - 41 **Miura K**, Kodama Y, Inokuchi S, Schnabl B, Aoyama T, Ohnishi H, Olefsky JM, Brenner DA, Seki E. Toll-like receptor 9 promotes steatohepatitis by induction of interleukin-1 β in mice. *Gastroenterology* 2010; **139**: 323-34.e7 [PMID: 20347818 DOI: 10.1053/j.gastro.2010.03.052]
 - 42 **Rivera CA**, Adegboyega P, van Rooijen N, Tagalicud A, Allman M, Wallace M. Toll-like receptor-4 signaling and Kupffer cells play pivotal roles in the pathogenesis of non-alcoholic steatohepatitis. *J Hepatol* 2007; **47**: 571-579 [PMID: 17644211 DOI: 10.1016/j.jhep.2007.04.019]
 - 43 **Yang SQ**, Lin HZ, Lane MD, Clemens M, Diehl AM. Obesity increases sensitivity to endotoxin liver injury: implications for the pathogenesis of steatohepatitis. *Proc Natl Acad Sci USA* 1997; **94**: 2557-2562 [PMID: 9122234]
 - 44 **Ma YY**, Li L, Yu CH, Shen Z, Chen LH, Li YM. Effects of probiotics on nonalcoholic fatty liver disease: a meta-analysis. *World J Gastroenterol* 2013; **19**: 6911-6918 [PMID: 24187469]
 - 45 **Ding S**, Chi MM, Scull BP, Rigby R, Schwerbrock NM, Magness S, Jobin C, Lund PK. High-fat diet: bacteria interactions promote intestinal inflammation which precedes and correlates with obesity and insulin resistance in mouse. *PLoS One* 2010; **5**: e12191 [PMID: 20808947 DOI: 10.3748/wjg.v19.i40.6911]
 - 46 **Hena-Mejia J**, Elinav E, Jin C, Hao L, Mehal WZ, Strowig T, Thaiss CA, Kau AL, Eisenbarth SC, Jurczak MJ, Camporez JP, Shulman GI, Gordon JI, Hoffman HM, Flavell RA. Inflammasome-mediated dysbiosis regulates progression of NAFLD and obesity. *Nature* 2012; **482**: 179-185 [PMID: 22297845 DOI: 10.1038/nature10809]
 - 47 **Zhang F**, Luo W, Shi Y, Fan Z, Ji G. Should we standardize the 1,700-year-old fecal microbiota transplantation? *Am J Gastroenterol* 2012; **107**: 1755; author reply p.1755-p.1756 [PMID: 23160295 DOI: 10.1038/ajg.2012.251]
 - 48 **Eiseman B**, Silen W, Bascom GS, Kauvar AJ. Fecal enema as an adjunct in the treatment of pseudomembranous enterocolitis. *Surgery* 1958; **44**: 854-859 [PMID: 13592638]
 - 49 **van Nood E**, Vrieze A, Nieuwdorp M, Fuentes S, Zoetendal EG, de Vos WM, Visser CE, Kuijper EJ, Bartelsman JF, Tijssen JG, Speelman P, Dijkgraaf MG, Keller JJ. Duodenal infusion of donor feces for recurrent *Clostridium difficile*. *N Engl J Med* 2013; **368**: 407-415 [PMID: 23323867 DOI: 10.1056/NEJMoa1205037]

P- Reviewer: Abdel-Salam OME, Balaban YH,

Hsieh SY, Julie NL, Rajeshwari K, Wong GLH

S- Editor: Ji FF **L- Editor:** Wang TQ **E- Editor:** Liu SQ





Published by **Baishideng Publishing Group Inc**

8226 Regency Drive, Pleasanton, CA 94588, USA

Telephone: +1-925-223-8242

Fax: +1-925-223-8243

E-mail: bpgoffice@wjgnet.com

Help Desk: <http://www.wjgnet.com/esps/helpdesk.aspx>

<http://www.wjgnet.com>

