

## Epigenetic targets of rheumatoid arthritis

Ghazi Chabchoub

Ghazi Chabchoub, Laboratoire de Génétique Moléculaire Humaine, Faculté de Médecine de Sfax, 3000 Sfax, Tunisia

Ghazi Chabchoub, Caisse National d'assurance Maladie, 3021 Sakiet Ezzit, Tunisia

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Correspondence to: Ghazi Chabchoub, MD, PhD, Médecin Inspecteur, Caisse National d'assurance Maladie, Rue Mongi Slim Sakiet Ezzit, 3021 Sakiet Ezzit, Tunisia. [ghazi.chabchoub@laposte.net](mailto:ghazi.chabchoub@laposte.net)  
Telephone: +216-98-291120

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### Abstract

Rheumatoid arthritis (RA) is a systemic, inflammatory and autoimmune disorder, characterized by chronic arthritis with progressive joint destruction. It has a multifactorial aetiology involving both genetic and environmental factors. Epigenetics can be defined as modifications of DNA that result in altered gene expression. The two main epigenetic mechanisms are post translational modifications to

histone tails and DNA methylation. Recent evidence has suggested that epigenetic mechanisms may be an important contributor to RA susceptibility. The aim of this editorial is to present evidence for the role of epigenetic mechanisms in the pathogenesis of RA and the potential to therapeutic target. Several studies targeting histone modification and DNA methylation in animal models of inflammatory arthritis will be reviewed and alterations in the epigenetic signature of genes of key RA related pathways such as pro-inflammatory cytokines, proteases and regulators of cellular proliferation.

**Key words:** Rheumatoid arthritis; Epigenetic; DNA methylation; Histone modification

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**Core tip:** This paper has highlighted the numerous processes involved in the pathogenesis of rheumatoid arthritis (RA) that are modulated by epigenetic mechanisms. This is important hypotheses to explore a novel therapeutic target in RA.

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### INTRODUCTION

Rheumatoid arthritis (RA) is a systemic, inflammatory and autoimmune disorder, characterized by chronic arthritis with progressive joint destruction<sup>[1]</sup>. A typical feature is the over-production of pro-inflammatory cytokines including tumor necrosis factor (TNF) and interleukin-1 (IL-1), which lead to up-regulation of other pro-inflammatory molecules and proteases<sup>[2]</sup>.

RA is a complex disease, and his etiology involves an interaction of both genetic and environmental factors. Genome scans have identified multiple regions

linked to disease<sup>[3-5]</sup>. Although interesting associations have been reported<sup>[6,7]</sup>, only alleles at the HLA-DRB1 locus have consistently demonstrated both linkage and association<sup>[8]</sup>.

Treatment of RA for most patients involves the administration of disease-modifying antirheumatic drugs (DMARDs) such as methotrexate, leflunomide and sulphasalazine. If DMARDs treatment fails a new drug targets need to be identified to provide therapy for these patients who do not respond to either conventional treatment. Epigenetic mechanisms, which have been implicated in RA, systemic lupus erythematosus and systemic sclerosis<sup>[9,10]</sup>, offers both new targets for therapy but also may help predict the successful outcome of drug treatments in patients.

## EPIGENETICS MECHANISMS

Epigenetics can be defined as modifications of DNA or associated proteins without a change in the DNA sequence itself that result in altered gene expression. The two main epigenetic mechanisms are post translational modifications to histone tails and methylation of DNA, which determine the chromatin state and there by access of transcription factors to gene promoter regions. In cells, DNA is packaged by being wound around nucleosome octomers containing two each of histone H2A, H2B, H3 and H4. These histone proteins have N-terminal tails that are susceptible to a number of post-translational alterations including acetylation, methylation, phosphorylation, sumolation, isomerisation of proline and ubiquitination<sup>[11,12]</sup>.

The most widely studied histone modification is the addition of acetyl groups to the lysine residues of the N-terminal tails of histones H3 and H4. Histone acetyltransferases add the acetyl groups to lysine residues within the histone tails from the donor acetyl coenzyme A. In turn the acetylation can be reversed by histone deacetylases (HDAC).

DNA methylation results from the addition of a methyl group to the 5-carbon position ring of cytosine. The methylation blocks binding of transcription factors and other co-activators to the DNA and recruits transcriptional repressors to the promoter<sup>[13]</sup>. Three active DNA methyltransferases (DNMT) have been identified. De novo methylation is performed by DNMT3A and DNMT3B, whereas methylation is maintained following cell division by DNMT1<sup>[14]</sup>.

## EPIGENETIC REGULATION OF IMMUNE CELLS AND PRO-INFLAMMATORY MOLECULES

Both DNA methylation and histone modifications contribute to the expression of a number of T-cell associated cytokines, including interferon gamma (IFN- $\gamma$ ), IL-2 and IL-7R $\alpha$ <sup>[15-21]</sup>. At the IFN- $\gamma$  promoter differential methylation is seen in naïve, effector and memory T-cells

being highest in naïve cells. There is rapid loss of methylation upon re-stimulation in memory T-cells that is associated with these cells rapid ability to produce IFN- $\gamma$  in response to reinfection by a virus<sup>[17]</sup>. Interestingly, STAT4, which is involved in the stimulation of IFN- $\gamma$  expression, is also regulated by DNA methylation suggesting that if methylation levels change at both these promoters together there could be a multiplicative effect on the quantity of IFN- $\gamma$  produced<sup>[18]</sup>. Protein kinase C activators and phosphatase inhibitors attenuate DNA methylation levels, possibly by altering the activity of DNMT1, correlating with the induction of IFN- $\gamma$  providing one possible mechanism for changing methylation levels<sup>[22]</sup>. IL-7 is involved in chronic inflammation in RA, not only does it increase osteoclast formation and thereby enhances bone loss but it is also important in maintaining T-cell homeostasis<sup>[23]</sup>. Expression of the IL-7 receptor, IL-7R $\alpha$ , on T-cells is regulated by DNA methylation with T-cells showing only low surface IL-7R $\alpha$  having higher methylation at the gene promoter<sup>[21]</sup>.

TNF is a major player in the pathology of RA confirmed by the efficacy of anti-TNF treatment in many patients. There is increasing evidence that epigenetic mechanisms play a role in regulating TNF expression. Phosphorylation of Serine (S)<sup>10</sup> of histone H3 and demethylation of Lysine (K)<sup>7</sup> has been observed at the TNF promoter in THP-1 cells, following LPS stimulation<sup>[24]</sup>. These changes to the histone code occur in conjunction with loss of heterochromatin binding protein 1 $\alpha$  (HP1 $\alpha$ ) from the TNF promoter<sup>[12]</sup>. The role of histone acetylation in the regulation of TNF production remains unclear. In animal models of arthritis it appears that HDAC inhibitors attenuate the expression of TNF whereas treatment of monocyte or macrophages with various inhibitors including sodium butyrate, Chlamydocin and HC-toxin generally increased TNF production with sodium butyrate increasing expression levels by 22 fold in THP-1 cells<sup>[24]</sup>.

Epigenetic regulation has been implicated in the modulation of both IL-1 $\alpha$  and IL-1 $\beta$ . Methylation of the proximal promoter regulates the expression of IL-1 $\alpha$  and this methylation is responsible for the allele specific expression of this cytokine in CD4<sup>+</sup> T cells<sup>[25,26]</sup>. Histone modifications appear the major epigenetic mechanism regulating IL-1 $\beta$ . Inhibiting HDAC activity with fibroblasts trichostatin A (TSA) caused an increase in LPS-induced IL-1 $\beta$  expression in choriodecidual explants<sup>[27]</sup>. Interestingly DNA methylation appears to have a role in regulating IL-6 gene expression<sup>[2]</sup>.

IL-6 is another important pro-inflammatory cytokine involved in the hepatic acute phase of RA and with its role confirmed by the success of the IL-6 receptor antibody, tocilizumab, in reducing disease activity and bone erosions<sup>[2]</sup>. Interestingly DNA methylation appears to have a role in regulating IL-6 gene expression. In RA patients reduced DNA methylation at a single CpG site within the promoter (-1099) in comparison to healthy for controls has been identified<sup>[28]</sup>. Reduced

methylation at this site was associated with increased IL-6 mRNA production in response to LPS. In addition to the role of epigenetics in regulating expression of pro-inflammatory cytokines, these molecules once produced may also have a role in epigenetic regulation. IL-6 has been shown to modulate the expression and activity of DNMTs leading to increased DNA methylation at the tumour suppressor gene p53 and Foxp3<sup>[29,30]</sup>. Methylation induced by IL-6 at the p53 promoter could be playing a role in silencing pro-apoptotic genes in the rheumatoid synovium<sup>[29]</sup>.

Numerous HDAC inhibitors have been developed mainly to target histone acetylation in RA models. Various studies using topical application of suberoylanilide hydroxamic acid (SAHA), MS-275 and FK228 established their potential to decrease serum IL-6, IL-1 $\beta$  and TNF levels suggesting an important role for HDAC in regulating production of pro-inflammatory cytokines<sup>[31,32]</sup>.

## EPIGENETIC MECHANISMS AND REGULATION OF THE HOMEOSTASIS IN THE SYNOVIAL JOINT

Increasing evidence suggests that histone tail modifications have an important role in regulating synovial hyperplasia in the RA joint. HDAC inhibitors have been tested in several models of RA in both rats and mice. These studies demonstrated the efficacy of TSA, phenylbutyrate FK228, SAHA and MS275 in ameliorating joint swelling and inflammation associated with inflammatory arthritis<sup>[31-34]</sup>. Treatment of RA synovial fibroblasts (RASf) with phenylbutyrate, TSA and FK228 causes histone hyperacetylation at p16<sup>INK4</sup> and p21<sup>CIP1</sup> promoters associated with expression of these two proteins which involved in the reduction of RASf numbers<sup>[35]</sup>. Interestingly, treatment with TSA and phenyl butyrate established the potential of HDAC inhibitors to reduce paw swelling in an adjuvant arthritis model in rats. It was reported that treatment had to start early for the inhibitors to prevent pannus formation and associated joint damage. In addition, TSA was found to have a greater ability to suppress synovial hyperplasia than phenylbutyrate<sup>[31]</sup>.

Methylation of DNA may also play a role in regulating cartilage integrity. Demethylation of specific loci within the MMP-3, MMP-9, MMP-13 and ADAMTS-4 promoters is present in cartilage from patients with osteoarthritis compared to controls which, is seen in conjunction with other expression of these enzymes in osteoarthritis cartilage<sup>[36]</sup>. Osteogenic protein-1 (OP-1) is a potent anabolic growth factor for articular chondrocytes, an aging-related increase in OP-1 promoter methylation that leads to decreased expression may contribute to cartilage loss seen with aging and in particular with the progression of osteoarthritis in older adults<sup>[37]</sup>. In addition, when comparing DNA methylation patterns in chondrocytes and mesenchymal stem cells undergoing

chondrogenesis, loss of methylation at two CpG sites within the promoter of type X collagen is associated with the production of this collagen in the latter cell type<sup>[38]</sup>. These findings demonstrate the importance of DNA methylation in regulating the homeostasis cartilage.

## CONCLUSION

This editorial has highlighted the numerous processes involved in the pathogenesis of RA that are modulated by epigenetic mechanisms. Key aspects of the production of pro-inflammatory molecules, inappropriate immune cell responses, and abnormalities in the synovium and cartilage degradation have all been shown to be modulated by histone modifications and DNA methylation. However, the complete picture of how epigenetic mechanisms modulate cellular differentiation and response to activation remains unclear. The success of HDAC inhibitors in ameliorating the symptoms of inflammatory arthritis in animal models is an exciting new development for the treatment of RA. Further development in HDAC inhibitors especially as more complete clinical trials, will lead to further knowledge generation on their mechanisms, targets and ability to treat RA.

## REFERENCES

- 1 **McInnes IB**, Schett G. The pathogenesis of rheumatoid arthritis. *N Engl J Med* 2011; **365**: 2205-2219 [PMID: 22150039 DOI: 10.1056/NEJMra1004965]
- 2 **Brennan FM**, McInnes IB. Evidence that cytokines play a role in rheumatoid arthritis. *J Clin Invest* 2008; **118**: 3537-3545 [PMID: 18982160 DOI: 10.1172/JCI36389]
- 3 **Okada Y**, Wu D, Trynka G, Raj T, Terao C, Ikari K, Kochi Y, Ohmura K, Suzuki A, Yoshida S, Graham RR, Manoharan A, Ortmann W, Bhangale T, Denny JC, Carroll RJ, Eyler AE, Greenberg JD, Kremer JM, Pappas DA, Jiang L, Yin J, Ye L, Su DF, Yang J, Xie G, Keystone E, Westra HJ, Esko T, Metspalu A, Zhou X, Gupta N, Mirel D, Stahl EA, Diogo D, Cui J, Liao K, Guo MH, Myouzen K, Kawaguchi T, Coenen MJ, van Riel PL, van de Laar MA, Guchelaar HJ, Huizinga TW, Dieudé P, Mariette X, Bridges SL, Zernakova A, Toes RE, Tak PP, Miceli-Richard C, Bang SY, Lee HS, Martin J, Gonzalez-Gay MA, Rodriguez-Rodriguez L, Rantapää-Dahlqvist S, Arlestig L, Choi HK, Kamatani Y, Galan P, Lathrop M, Eyre S, Bowes J, Barton A, de Vries N, Moreland LW, Criswell LA, Karlson EW, Taniguchi A, Yamada R, Kubo M, Liu JS, Bae SC, Worthington J, Padyukov L, Klareskog L, Gregersen PK, Raychaudhuri S, Stranger BE, De Jager PL, Franke L, Visscher PM, Brown MA, Yamanaka H, Mimori T, Takahashi A, Xu H, Behrens TW, Siminovitch KA, Momohara S, Matsuda F, Yamamoto K, Plenge RM. Genetics of rheumatoid arthritis contributes to biology and drug discovery. *Nature* 2014; **506**: 376-381 [PMID: 24390342]
- 4 **Zhang MM**, Jiang YS, Lv HC, Mu HB, Li J, Shang ZW, Zhang RJ. Pathway-based association analysis of two genome-wide screening data identifies rheumatoid arthritis-related pathways. *Genes Immun* 2014; **15**: 487-494 [PMID: 25101796]
- 5 **Neidhart M**, Karouzakis E. Genetics: a new interpretation of genetic studies in RA. *Nat Rev Rheumatol* 2014; **10**: 199-200 [PMID: 24567061]
- 6 **Zhang C**, Chen L, Gu Y. Polymorphisms of MMP-1 and MMP-3 and susceptibility to rheumatoid arthritis. A meta-analysis. *Z Rheumatol* 2015; **74**: 258-262 [PMID: 25854159 DOI: 10.1007/s00393-014-1537-2]
- 7 **Abd-Allah SH**, el-Shal AS, Shalaby SM, Pasha HF, Abou el-Saoud

- AM, el-Najjar AR, el-Shahawy EE. PADI4 polymorphisms and related haplotype in rheumatoid arthritis patients. *Joint Bone Spine* 2012; **79**: 124-128 [PMID: 21981985]
- 8 **Reynolds RJ**, Ahmed AF, Danila MI, Hughes LB, Gregersen PK, Raychaudhuri S, Plenge RM, Bridges SL. HLA-DRB1-associated rheumatoid arthritis risk at multiple levels in African Americans: hierarchical classification systems, amino acid positions, and residues. *Arthritis Rheumatol* 2014; **66**: 3274-3282 [PMID: 25524867]
- 9 **Glant TT**, Mikecz K, Rauch TA. Epigenetics in the pathogenesis of rheumatoid arthritis. *BMC Med* 2014; **12**: 35 [PMID: 24568138]
- 10 **Sánchez-Pernaute O**, Ospelt C, Neidhart M, Gay S. Epigenetic clues to rheumatoid arthritis. *J Autoimmun* 2014; **30**: 12-20 [PMID: 18155418 DOI: 10.1016/j.jaut.2007.11.006]
- 11 **An W**. Histone acetylation and methylation: combinatorial players for transcriptional regulation. *Subcell Biochem* 2007; **41**: 351-369 [PMID: 17484136 DOI: 10.1007/1-4020-5466-1\_16]
- 12 **Berger SL**. The complex language of chromatin regulation during transcription. *Nature* 2007; **447**: 407-412 [PMID: 17522673 DOI: 10.1038/nature05915]
- 13 **Meng H**, Cao Y, Qin J, Song X, Zhang Q, Shi Y, Cao L. DNA methylation, its mediators and genome integrity. *Int J Biol Sci* 2015; **11**: 604-617 [PMID: 25892967 DOI: 10.7150/ijbs.11218]
- 14 **Baubec T**, Colombo DF, Wirbelauer C, Schmidt J, Burger L, Krebs AR, Akalin A, Schübeler D. Genomic profiling of DNA methyltransferases reveals a role for DNMT3B in genic methylation. *Nature* 2015; **520**: 243-247 [PMID: 25607372]
- 15 **Berkley AM**, Hendricks DW, Simmons KB, Fink PJ. Recent thymic emigrants and mature naive T cells exhibit differential DNA methylation at key cytokine loci. *J Immunol* 2013; **190**: 6180-6186 [PMID: 23686491]
- 16 **Komori HK**, Hart T, LaMere SA, Chew PV, Salomon DR. Defining CD4 T cell memory by the epigenetic landscape of CpG DNA methylation. *J Immunol* 2015; **194**: 1565-1579 [PMID: 25576597]
- 17 **Gray SM**, Kaech SM, Staron MM. The interface between transcriptional and epigenetic control of effector and memory CD8+ T-cell differentiation. *Immunol Rev* 2014; **261**: 157-168 [PMID: 25123283]
- 18 **Williams CL**, Schilling MM, Cho SH, Lee K, Wei M, Aditi M. STAT4 and T-bet are required for the plasticity of IFN- $\gamma$  expression across Th2 ontogeny and influence changes in Ifng promoter DNA methylation. *J Immunol* 2013; **191**: 678-687 [PMID: 23761633]
- 19 **Dong J**, Chang HD, Ivascu C, Qian Y, Rezai S, Okhrimenko A, Cosmi L, Maggi L, Eckhardt F, Wu P, Sieper J, Alexander T, Annunziato F, Gossen M, Li J, Radbruch A, Thiel A. Loss of methylation at the IFNG promoter and CNS-1 is associated with the development of functional IFN- $\gamma$  memory in human CD4(+) T lymphocytes. *Eur J Immunol* 2013; **43**: 793-804 [PMID: 23255246 DOI: 10.1002/eji.201242858]
- 20 **Northrop JK**, Thomas RM, Wells AD, Shen H. Epigenetic remodeling of the IL-2 and IFN-gamma loci in memory CD8 T cells is influenced by CD4 T cells. *J Immunol* 2006; **177**: 1062-1069 [PMID: 16818762 DOI: 10.4049/jimmunol.177.2.1062]
- 21 **Kim HR**, Hwang KA, Kim KC, Kang I. Down-regulation of IL-7Ralpha expression in human T cells via DNA methylation. *J Immunol* 2007; **178**: 5473-5479 [PMID: 17442928 DOI: 10.4049/jimmunol.178.9.5473]
- 22 **Bonilla-Henao V**, Martínez R, Sobrino F, Pintado E. Different signaling pathways inhibit DNA methylation activity and up-regulate IFN-gamma in human lymphocytes. *J Leukoc Biol* 2005; **78**: 1339-1346 [PMID: 16204617 DOI: 10.1189/jlb.1004604]
- 23 **Churchman SM**, El-Jawhari JJ, Burska AN, Parmar R, Goëb V, Conaghan PG, Emery P, Ponchel F. Modulation of peripheral T-cell function by interleukin-7 in rheumatoid arthritis. *Arthritis Res Ther* 2014; **16**: 511 [PMID: 25533722]
- 24 **Falvo JV**, Jasenosky LD, Kruidenier L, Goldfeld AE. Epigenetic control of cytokine gene expression: regulation of the TNF/LT locus and T helper cell differentiation. *Adv Immunol* 2013; **118**: 37-128 [PMID: 23683942]
- 25 **van Rietschoten JG**, Gal-Yam EN, Jeong S, Cortez CC, Verweij CL, Jones PA. Epigenetic regulation and nucleosome positioning in the human TATA-less IL-1 alpha promoter. *Genes Immun* 2008; **9**: 582-590 [PMID: 18615092]
- 26 **van Rietschoten JG**, Verzijlbergen KF, Gringhuis SI, van der Pouw Kraan TC, Bayley JP, Wierenga EA, Jones PA, Kooter JM, Verweij CL. Differentially methylated alleles in a distinct region of the human interleukin-1alpha promoter are associated with allele-specific expression of IL-1alpha in CD4+ T cells. *Blood* 2006; **108**: 2143-2149 [PMID: 16788102 DOI: 10.1182/blood-2006-01-021147]
- 27 **Sato TA**, Mitchell MD. Molecular inhibition of histone deacetylation results in major enhancement of the production of IL-1beta in response to LPS. *Am J Physiol Endocrinol Metab* 2006; **290**: E490-E493 [PMID: 16234266 DOI: 10.1152/ajpendo.00406.2005]
- 28 **Ishida K**, Kobayashi T, Ito S, Komatsu Y, Yokoyama T, Okada M, Abe A, Murasawa A, Yoshie H. Interleukin-6 gene promoter methylation in rheumatoid arthritis and chronic periodontitis. *J Periodontol* 2012; **83**: 917-925 [PMID: 22122521]
- 29 **Hodge DR**, Peng B, Cherry JC, Hurt EM, Fox SD, Kelley JA, Munroe DJ, Farrar WL. Interleukin 6 supports the maintenance of p53 tumor suppressor gene promoter methylation. *Cancer Res* 2005; **65**: 4673-4682 [PMID: 15930285 DOI: 10.1158/0008-5472.CAN-04-3589]
- 30 **Lal G**, Zhang N, van der Touw W, Ding Y, Ju W, Bottinger EP, Reid SP, Levy DE, Bromberg JS. Epigenetic regulation of Foxp3 expression in regulatory T cells by DNA methylation. *J Immunol* 2009; **182**: 259-273 [PMID: 19109157 DOI: 10.4049/jimmunol.182.1.259]
- 31 **Nishida K**, Komiyama T, Miyazawa S, Shen ZN, Furumatsu T, Doi H, Yoshida A, Yamana J, Yamamura M, Ninomiya Y, Inoue H, Asahara H. Histone deacetylase inhibitor suppression of autoantibody-mediated arthritis in mice via regulation of p16INK4a and p21(WAF1/Cip1) expression. *Arthritis Rheum* 2004; **50**: 3365-3376 [PMID: 15476220 DOI: 10.1002/art.20709]
- 32 **Lin HS**, Hu CY, Chan HY, Liew YY, Huang HP, Lepescheux L, Bastianelli E, Baron R, Rawadi G, Clément-Lacroix P. Anti-rheumatic activities of histone deacetylase (HDAC) inhibitors in vivo in collagen-induced arthritis in rodents. *Br J Pharmacol* 2007; **150**: 862-872 [PMID: 17325656 DOI: 10.1038/sj.bjpp.0707165]
- 33 **Ocker M**, Schneider-Stock R. Histone deacetylase inhibitors: signalling towards p21cip1/waf1. *Int J Biochem Cell Biol* 2007; **39**: 1367-1374 [PMID: 17412634]
- 34 **Zhou X**, Hua X, Ding X, Bian Y, Wang X. Trichostatin differentially regulates Th1 and Th2 responses and alleviates rheumatoid arthritis in mice. *J Clin Immunol* 2011; **31**: 395-405 [PMID: 21305388]
- 35 **Morinobu A**, Wang B, Liu J, Yoshiya S, Kurosaka M, Kumagai S. Trichostatin A cooperates with Fas-mediated signal to induce apoptosis in rheumatoid arthritis synovial fibroblasts. *J Rheumatol* 2006; **33**: 1052-1060 [PMID: 16755652]
- 36 **Cheung KS**, Hashimoto K, Yamada N, Roach HI. Expression of ADAMTS-4 by chondrocytes in the surface zone of human osteoarthritic cartilage is regulated by epigenetic DNA de-methylation. *Rheumatol Int* 2009; **29**: 525-534 [PMID: 18941754]
- 37 **Loeser RF**, Im HJ, Richardson B, Lu Q, Chubinskaya S. Methylation of the OP-1 promoter: potential role in the age-related decline in OP-1 expression in cartilage. *Osteoarthritis Cartilage* 2009; **17**: 513-517 [PMID: 18829350]
- 38 **Zimmermann P**, Boeuf S, Dickhut A, Boehmer S, Olek S, Richter W. Correlation of COL10A1 induction during chondrogenesis of mesenchymal stem cells with demethylation of two CpG sites in the COL10A1 promoter. *Arthritis Rheum* 2008; **58**: 2743-2753 [PMID: 18759285 DOI: 10.1002/art.23736]

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