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**Adenosquamous carcinoma of the pancreas: Molecular characterization of 23 patients along with a literature review**

Borazanci E *et al.* Molecular characterization/literature review of ASCP

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**Abstract**

Adenosquamous carcinoma of the pancreas (ASCP) is a rare entity. Like adenocarcinoma of the pancreas, overall survival is poor. Characteristics of ASCP include central tumor necrosis, along with osteoclasts and hypercalcemia. Various theories exist as to why this histological subtype exists, as normal pancreas tissue has no benign squamous epithelium. Due to the rarity of this disease, limited molecular analysis has been performed, and those reports indicate unique molecular features of ASCP.In this paper, we characterize 23 patients diagnosed with ASCP through molecular profiling using immunohistochemistry staining, fluorescent in-situ hybridization, chromogenic in situ hybridization, and gene sequencing, Additionally, we provide a comprehensive literature review of what is known to date of ASCP. Molecular characterization revealed overexpression in MRP1 (80%), MGMT (79%), TOP2A (75), RRM1 (42%), TOPO1 (42%), PTEN (45%), CMET (40%), and C-KIT (10%) among others. 100% of samples tested were positive for *KRAS* mutations. Thisanalysisshows heretofore unsuspected leads to be considered for treatments of this rare type of exocrine pancreas cancer. Molecular profiling may be appropriate to provide maximum information regarding the patient’s tumor. Further work should be pursued to better characterize this disease.

**Key words:** Adenosquamous carcinoma of the pancreas; Molecular profiling; Review

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**Core tip:** This analysis of 23 adenosquamous carcinoma of the pancreas in light of the reviewed literature highlights the potential to identify novel treatments when using a personalized medicine approach to patient tumor characterization.

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**INTRODUCTION**

Pancreas cancer remains a deadly disease. In 2014 it is estimated that 46420 new cases will occur, along with 39590 deaths, making it the fourth leading cause of cancer deaths in the United States[1]. The most commonly diagnosed pancreas cancer histology is adenocarcinoma, with an incidence of 85% of pancreas malignancies[2]. As shown in Table 1, other pancreas cancer histological subtypes include mucinous cyst adenocarcinoma, adenosquamous, undifferentiated/anaplastic, papillary mucinous, acinar cell, spindle cell, and pancreatoblastoma[2-4].

Adenosquamous carcinoma of the pancreas (ASCP) is a rare entity. Its estimated incidence in the literature is between 0.38% to 10% of all exocrine pancreatic tumors (Table 2)[2,5-19]. ASCP has also been referred to as adenoacanthoma, mixed squamous and adenocarcinoma, and mucoepidermoid carcinoma[20]. The entity was first described in 1907 by Gotthold Herheimer, who referred it as cancroide[20]. Adenosquamous histology is seen in cancers of other organ systems such as lung, esophagus, colon, stomach, salivary glands, and the female reproductive system[20]. Compared to pancreatic adenocarcinoma, which has a poor 5-year overall survival, survival is worse in patients with ASCP[12-15].

The etiology of ASCP is unknown. Most literature reports of this disease have come from Asia. The largest known case study showed that 79% of 415 patients with ASCP were Caucasian[12]. It is unknown if risk factors for the development of pancreatic adenocarcinoma such as chronic pancreatitis, ABO blood group, alcohol use, tobacco use, germline mutations such as BRCA2, PALB2, ATM, and p53 are also risk factors for the development of ASCP[12,21].

In this review, we will discuss the current understanding of ASCP. We have profiled 23 patients with ASCP and will present our findings, along with other molecular analyses reported in the literature. We will also discuss potential treatment strategies specifically targeting ASCP.

**PATHOLOGY**

Normal pancreas tissue has no benign squamous epithelial components[9,15,22]. Various hypotheses have been proposed regarding the histogenesis of ASCP. One theory hypothesizes that squamous metaplasia occurs as a result of ductal inflammation due to chronic pancreatitis or obstruction by an adenomatous tumor, and this process leads to ASCP[5,23]. Another theory, termed the collision theory, suggests that two histologically distinct tumors arise independently in the pancreas and are joined together leading to ASCP[20,23,24]. Finally, the third theory, the differentiation theory, suggests that a primitive pancreatic stem cell differentiates into either squamous or adenocarcinoma or becomes a combination of both[14,22,23]. Despite different hypotheses, there has been no study to elucidate the mechanism of origination of ASCP.

The pathology of ASCP includes the typical squamous carcinoma pattern that is characterized by epithelium with whorls, keratohyalin, or pearls[14,16], as seen in Figure 1. Compared to nuclei of benign squamous cells, the nuclei of malignant squamous cells are hyperchromic and pleomorphic[15,22]. The squamous carcinoma component of ASCP appears to be more focal in the tumor. An interesting histological feature is the finding in several case series of ASCP that the squamous cell carcinoma component is located in the periphery of the tumor, while the adenocarcinoma component is in the center[9,22]. There is a transitional zone where the glandular structure blends into the squamous component[22]. There is an entity descriptive of pure squamous cell carcinoma of the pancreas, but this classification has been debated and is considered to be more secondary to metastasis to the pancreas from a non-pancreas primary carcinoma[15,25,26].

Tumor cell necrosis is frequently seen in patients with ASCP, along with high tumor grade[25,27]. Other unusual reported pathology has included the presence of clear cell and rhabdoid components[27,28]. One pathology case report noted the presence of both osteoclast and giant tumor cells which were scattered individually within the stroma[3]. The presence of osteoclasts is not unique to ASCP, as osteoclasts have been seen in adenosquamous carcinoma of other organs, including the esophagus, gallbladder, and kidney[3,13]. Acantholysis has also been noted[3]. The squamous component of the cancer has been shown to be more likely to demonstrate vascular invasion, but less likely to metastasize to the lymph nodes[16]. One study found that pancreatic adenosquamous carcinoma grows at twice the rate of pancreatic adenocarcinoma[29].

The current guideline to diagnose adenosquamous pancreatic cancer requires the presence of at least 30% of squamous component in the pancreas tumor tissue[18,20,30]. However, this classification system is being debated, due to both the subjective nature of estimating percentage composition and the sampling method of a patient’s tumor at biopsy through fine needle aspiration (FNA) versus surgical resection. The clinical relevance of the degree of squamous cell differentiation in adenosquamous pancreas cancer is unknown[16,18]. The proportion of squamous differentiation in ASCP did not influence survival in one case series of 38 patients[22]. Some have proposed that presence of any squamous cell carcinoma component in a pancreatic tumor should classify the cancer as adenosquamous[16,26,31].

Prior immunohistochemistry (IHC) analysis on patients with adenosquamous carcinoma have shown the cancer to be positive for cytokeratin (CK) 5/6, CK 7, and p63 and negative for CK 20, p16, and p53[18,32]. IHC positivity for pancreatic adenocarcinoma includes CK7, CK20, mesothelin, cancer antigen 125 (CA-125), and lysozyme[18,33]. The KI-67 index for one patient with ASCP with approximately a 70%-80% squamous carcinoma component was 33%[32].

As in pancreatic adenocarcinoma, *KRAS* mutations have also been observed in ASCP[18,27,31,34]. A molecular study involving analysis for p53, Dpc4/SMAD4, p16, E-cadherin, EGFR protein expression levels, *KRAS* mutational analysis; *p16/CDKN2a* amplification, and HPV DNA detection was carried out on 8 patients with ASCP[27]. The *KRAS* mutations only screened for mutations in codons 12 and 13, which were present in 5/8 of the squamous component of the cancer samples. A homozygous deletion of the p16 gene was present in 3/8 squamous components. Regarding protein expression in the same patient samples, DPC4 was lost in 5/8 samples, p53 was positive in 5/8 samples, p16 was universally lost, E-cadherin was either reduced or lost in 7/8 samples, and P63 and EGFR were positive in all 8 samples[27]. The lack of protein expression of p16 was particularly interesting since the gene was present in 5/8 patient samples, suggesting other causes of loss of protein expression, such as gene silencing like DNA methylation. There was no HPV DNA detectable in the eight patients tested[27]. HPV status was looked at another analysis of 7 patients, and none of these patients were positive[13]. The lack of positivity of HPV is noteworthy due to its influence in the development of other squamous histology cancers such as the cervix, head and neck, and anus[13].

We have conducted a molecular characterization using a commercially available assay[35]. 23 patients with ASCP were identified and the results of the profiling are presented (Tables 3-5). The median age was 60 years old (range 41 to 86 years old), and 17/23 patients were male. Evaluation of protein expression by IHC analysis revealed the following: DNA topoisomerase2 (TOPO2A) overexpression was prevalent in 78% of the samples, which in some studies of other histologic types indicates sensitivity to agents such as doxorubicin or etoposide. Low expression of ribonucleotide reductase M1 (RRM1), which can indicate sensitivity to gemcitabine, was low in 57% of the patient samples. Low thymidylate synthase (TS) expression, found in 62% of patient samples, correlates to sensitivity in some tumor types to flouropyrimidines such as 5-FU, capecitabine, and pemetrexed. Low expression of excision repair cross-complementation group 1, or ERCC1, is associated with sensitivity to platinum-based therapies in some tumor types and was found to be low in 69% of patient samples. Other positive findings included 10% (1 in 10) positivity of c-KIT, and TOPO1 overexpression in 38% of patient samples. These biomarkers are correlated to sensitivities to imatinib and topotecan/irinotecan, respectively, in some tumor types. The high expression of both MRP1 and BCRP1 at 80% highlights the difficulty of treating ASCP, as these proteins are involved in drug resistance to chemotherapy. FISH/CISH analysis revealed an 11% overexpression of c-MET, an oncoprotein that is increasingly targeted in new drug development. Also, one sample had a mutation in c-MET. Of note, mutation analysis revealed *KRAS* mutations in all sixteen patient samples tested, but none had *EGFR* mutations.

Very little genomic sequencing data is available in the literature on adenosquamous pancreatic cancers. However, a study published examining whole genomic sequencing in eleven patients with advanced cancer included one patient with ASCP[36]. This patient’s sequencing included single nucleotide variations (SNV), whole genome sequencing (WGS), and whole transcriptome sequencing (WTS). Some of the variations found included the upregulation of two ligands, transforming growth factor (TGF)-beta 1 and TGF-beta 2 along with their accompanying receptor, TGF-beta receptor type II. These growth factors are involved in the epithelial to mesenchymal transition (EMT). Other members of a shared pathway, Lef-1, TCF8, and E2A, were also found to be upregulated. E-cadherin was found to be down-regulated, which is a hallmark of the EMT phenotype[33]. The EMT phenotype has been shown to play a crucial role in cancer cell metastasis along with resistance to chemotherapy and contributing to the formation of cancer stem cells[36]. This patient’s tumor did have a mutation in KRAS at codon 12 along with a mutation in *PI3KCA*. The patient’s sequencing was done during therapy and upon progression on gemcitabine and cisplatin. The patient was then enrolled on a phase I trial involving a combination PI3K and MEK inhibitor, and experienced a clinical benefit in the form of a dramatic decrease in his pain, along with tumor response[36].

Another genetic analysis done recently looked at 23 patients with ASCP through genomic sequencing and showed a mutation of the *UPF1* gene, which encodes a RNA helicase essential for the highly conserved RNA degradation pathway, nonsense-mediated RNA decay (NMD)[37]. This mutation was not seen in the adjacent normal tissue of these patient samples. The pathways that UPF1 is implicated in are not all known but appear to be involved in the normal splicing of RNA, affecting such genes as p53[37].

**IMAGING**

While there is not a definitive characteristic appearance of ASCP on Computed tomography (CT) imaging, they are usually not well circumscribed[38]. CT imaging of ASCP lesions commonly show the presence of central necrosis within the tumor mass[31,38,39], which is rarely seen in pancreatic ductal adenocarcinoma or in endocrine tumors of the pancreas[40,41]. Another common imaging finding is the propensity for vascular and nerve encasement[38].

A large series looking at ASCP through CT and magnetic resonance imaging (MRI) showed the presence of frequent intra-tumor necrosis, increased enhancement, and exophytic growth[42]. It is theorized that this phenomenon may reflect the presence of the squamous component causing rapid proliferation, as these characteristics are not seen as often in adenocarcinoma of the pancreas[43]. Other unique features noted in imaging evaluation with ASCP are the lack of pancreatic atrophy and mild duct dilatation, which are more common features of pancreatic adenocarcinoma[42]. Like adenocarcinoma of the pancreas, adenosquamous cancers of the pancreas may exhibit the double duct sign[38], which consists of simultaneous dilatation of the common bile and pancreatic ducts[44].

Gallium-67 is an older radioactive tracer that is taken up by some malignancies and infections and has been replaced by PET scans in relation to tumor staging[45]. Intense Gallium-67 uptake, which rarely is detected in pancreatic adenocarcinoma, has been observed in ASCP[45,46]. PET-CT imaging has been reported in a limited number of case reports. One case report noted a patient with localized ASCP to have a standardized uptake value (SUV) of 15.8, which was over 3 times higher than the SUV average for patients with pancreatic adenocarcinoma at their institution[47].

Figure 2 highlights several key imaging findings from patients we have treated with ASCP, including necrosis and mixed morphology. Of note is that in looking at one of our recent ASCP patients, the hypermetabolism that has been previously reported in patients with ASCP was not seen[47].

**CLINICAL CHARACTERISTICS**

The characteristics of patients with ASCP tend to favor more aggressive features with more node positive disease, more poorly differentiated disease, and more perineural invasion present compared to patients with pancreatic adenocarcinoma[16]. Patients with ASCP present with symptoms similar in nature to patients with pancreatic adenocarcinoma, with abdominal pain, weight loss, back pain, nausea, vomiting, anorexia, and jaundice being the most common presenting symptoms[19,20,38]. As with pancreatic adenocarcinoma, there appears to be an increased risk of deep vein thrombosis (DVT)[25]. In larger case series, patients are typically male, white, present in their sixth decade of life, and the tumor is located in the head of the pancreas[12,13,20]. Serum lab abnormalities may include elevated bilirubin, elevated alkaline phosphatase, anemia, and elevated carbohydrate antigen 19-9 (CA 19-9)[19,20,25]. Occasionally, patients may also have elevated levels of carcinoembroyonic antigen (CEA) or CA-125[38].

Long term survival overall is poor for ASCP. Survival, despite surgical resection, is slightly poorer for patients with ASCP than those with pancreatic adenocarcinoma. Those with ASCP have a 3-year survival rate of 14% with surgery, as opposed to 19% 3-year survival of resected pancreatic adenocarcinoma patients[29,48]. Like patients with pancreatic adenocarcinoma, patients with ASCP tend to present more commonly in advanced stage, with one large analysis through the California Cancer Registry database (CCR) indicating over 50% of ASCP patients presenting in advanced stage[11]. The mean tumor diameter in one series of resected ASCP patients was 46.3 mm *vs* 33.5 mm of adenocarcinoma pancreas patients (*P* value 0.0001)[11]. Comparisons between patients at single institutions and matching for stage have yielded an overall median survival of 6.51 mo *vs* 11.0 mo for ASCP versus adenocarcinoma[15]. In another large single institution analysis of patients with ASCP, the median survival of patients with resection was 10.9 mo, which was worse than those with pancreatic adenocarcinoma who underwent resection, which was 17.9 mo[16].

In an analysis of Surveillance, Epidemiology, and End Results **(**SEER) patients that identified 415 patients with ASCP, the mean age was found to be 66 years old and the tumor more likely to be in the head of the pancreas. Compared to patients with adenocarcinoma of the pancreas, patients with ASCP were more likely to be poorly differentiated (71.4% *vs* 45%), larger (5.7 cm *vs* 4.3 cm), and more likely to have positive lymph nodes (52.8% *vs* 47.1%)[12]. In patients with ASCP, overall 1 and 2 year survival was 21.2% and 10.8% compared to 24.7% and 10.9% in patients with pancreatic adenocarcinoma[12]. Patients with ASCP were found to have a median survival of 4 mo compared to 5 mo in patients with pancreatic adenocarcinoma[12]. Patients with ASCP who underwent resection had worse survival rates than those with adenocarcinoma pancreas cancer who underwent resection. 1 year and 2 year survival rates of 50.7% and 29% and median survival was 12 mo in patients with ASCP as opposed to 60.1% and 35.8% and median survival of 16 mo in those with adenocarcinoma of the pancreas[12]. After primary resection, recurrence may occur in a number of sites. Common sites of metastases include the liver, lung, retroperitoneum, and development of malignant ascites[16,38].

Several studies have examined various clinical features of survival in patients with ASCP. Lymph node status, tumor size, or resection in patients with ASCP does not impact survival when compared with patients with adenocarcinoma of the pancreas[12,16]. Not surprisingly, risk factors for poorer survival of patients with ASCP are those with distant disease, advanced age, and patients unable to undergo surgical resection[12]. In one study, only 40% of patients with ASCP were resectable[12]. A single institution case series from the Mayo clinic showed that patients with an R1 resection still benefited in survival compared to those who did not undergo surgery[49]. Patients from that study that had either an R0 or R1 resection had a median survival of 14.4 and 8 mo respectively, compared to 4.8 mo who received no surgical treatment[49]. Location of the tumor matters, with poorer survival noted if the location was in the body or tail as opposed to the head of the pancreas[48]. This was based on a chart analysis of 39 patients with ASCP and may be accounted for by size of these tumors by location as the ones located in the head were smaller (4.7 ± 1.9 cm) as opposed to the body/tail lesions (7.3 ± 1.8 cm)[48]. The likely reason for poorer survival is that patients with head of pancreas lesions tend to present with obstructive symptoms, which are clinically evident when the lesion is smaller in comparison to body/tail lesions of the pancreas.

It is unclear why patients with ASCP have such a poor prognosis. Due to the small sample size, data to shed light on this disease has been limited. One case series from Voong *et al*[16] looking at patients diagnosed with ASCP and who had undergone surgery showed via univariate analysis that only patients who received adjunct chemoradiation had a clinical significant improvement in survival[16]. The patients who received adjunct chemoradiation had a median survival of 13.6 mo as opposed to 8.6 mo for those that did not[16]. Other factors such as age, tumor size, differentiation, margin, node status, type of surgery were not shown to affect survival in this case series[16].

Malignancy associated hypercalcemia, which is a rare phenomenon of exocrine pancreatic carcinoma, has been described in ASCP[50,51]. Of note is that malignancy associated hypercalcemia is more commonly associated with squamous cell carcinomas of the head, neck, lung, and esophagus. Case reports have also described patients with adenosquamous pancreatic cancer having elevated levels of calcium due to elevated levels of parathyroid hormone related protein (PTH-rP)[50,51]. In both reported cases, hypercalcemia persisted despite bisphosphonate treatment[50,51]. Curiously, hyperglycemia has not been reported with great frequency in ASCP despite being reported in up to 80% of patients with pancreatic adenocarcinoma[52].

**MANAGEMENT**

Diagnosis of patients with ASCP requires biopsy along with pathology review using criteria of ASCP with at least 30% of the tumor positive for squamous histology. Staging is done in a similar manner as pancreatic adenocarcinoma with guidelines set forth by the American Joint Committee on Cancer (AJCC)/International Union Against Cancer (UICC). Unresectable disease is designated as Stage III and metastatic disease is designated as stage IV. One issue with diagnosis includes the current standard approach of using endoscopic ultrasound (EUS) for diagnosing pancreatic cancer and using FNA. In a retrospective review of patients at John Hopkins University and Emory University it was noted that in patients who eventually had a diagnosis of ASCP after surgical resection, two thirds of them (67%) were initially diagnosed as being pancreatic adenocarcinoma only. It is possible for pathologists to misclassify or ignore the squamous cell compartment in pancreatic FNA specimens, which not only leads to underreporting of ASCP but may also miss the diagnosis of malignancy altogether[15].

There are currently no guidelines for treating patients with ASCP. Literature reports often cite treatment regimens similar to adenocarcinoma[48]. Due to its relative infrequency in incidence there have been no published randomized clinical trials specifically targeting patients with ASCP. Treatments in years past have focused on resection of local adenosquamous pancreatic carcinoma using the same guidelines for pancreatic adenocarcinoma. These include pancreaticoduodenectomy (PD), pylorus-preserving PD (PPPD), distal pancreatectomy (DP), and total pancreatectomy (TP)[48]. These techniques are not modified for ASCP and surgical resection remains the best opportunity to achieve long lasting survival[48].

The role of neoadjuvant and adjuvant chemotherapy is unclear, mimicking some questions that continue to be explored in pancreatic adenocarcinoma[48]. Most case reports in the literature have used 5-fluorouracil based therapies for treatment around surgical procedures and have not examined the role of gemcitabine or more robust regimens such as FOLFIRINOX or nab-paclitaxel/gemcitabine[49]. In a retrospective series of 62 patients identified with pancreatic adenosquamous carcinoma, 14 patients received platinum therapy in the adjuvant setting as opposed to 48 who did not[53]. The patients who received platinum therapy in the adjuvant setting had an overall median survival of 19.1 months as opposed to 10.7 mo for those who did not (*P* = 0.011, hazard ratio of survival 0.48)[53].

The role of radiation therapy as an adjunct to resection of ASCP is also unclear[48,54,55]. Two retrospective studies examined adjuvant radiation therapy, but did not show a benefit in overall survival for those that received adjuvant therapy versus those who did not. In a previously published literature review of 30 patients who received radiation therapy either intra and/or postoperatively, the 2-year survival rate was 20% and median survival 13 mo[48]. In the patients who did not receive radiation therapy their 2-year survival rate was 9% and median survival period was 6 mo. Despite the differences in survival between the 2 groups, they did not reach statistical significance[48].

**CONCLUSION**

ASCP is an aggressive variation of carcinoma of the pancreas. Overall it carries a poor prognosis. A study to assess the percentage component of squamous carcinoma in ASCP and associating this with differences in clinical outcome is certainly warranted, but may be difficult to carry out due to the scarcity of this disease and the subjective evaluation needed by pathologists to determine percent squamous in a pancreas carcinoma specimen. Obtaining the proper amount of tissue makes diagnosis difficult and is akin to diagnosing patients with lymphoma by way of FNA: there may be diagnostic inaccuracies depending upon where the sample is biopsied. This role of subjective evaluation also makes interpreting retrospective analysis difficult, such as examining databases like SEER.

There is a need to better characterize the disease beyond traditional pathology analysis. Doing further work characterizing this disease on a molecular level may further elucidate the requirements for classifying pancreatic carcinomas as adenosquamous or adeno. Our work in molecular characterization, while small in sample size, points to the use of novel therapeutic combinations in patients with ASCP, such as epirubicin/cisplatin/5-FU, which may be tested in small clinical trials. Targeting novel pathways such as those affecting the epithelial to mesenchymal change pathway, using agents that target APC, WNT, B-catenin, along with those targeting chromatin remodeling may be worth trying against this disease. Using cell lines derived from ASCP patients and studying them in growth assays and xenograft models may yield clues regarding their response to newer anti-cancer agents in development[54,55]. Understanding the key genetic drivers for this disease may lead to better treatment outcomes since it is clear traditional treatments for pancreatic adenocarcinoma do not translate well to ASCP.

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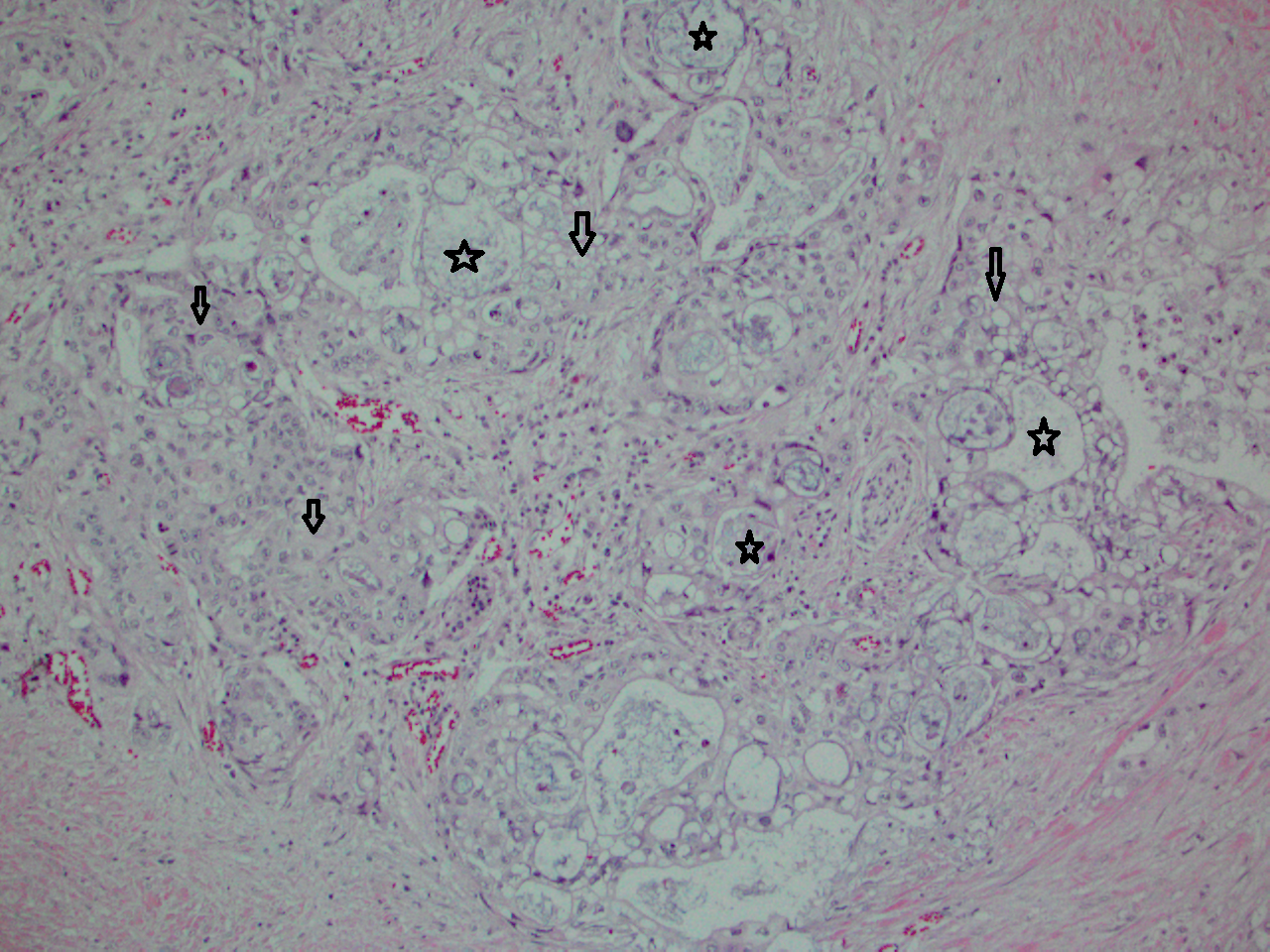
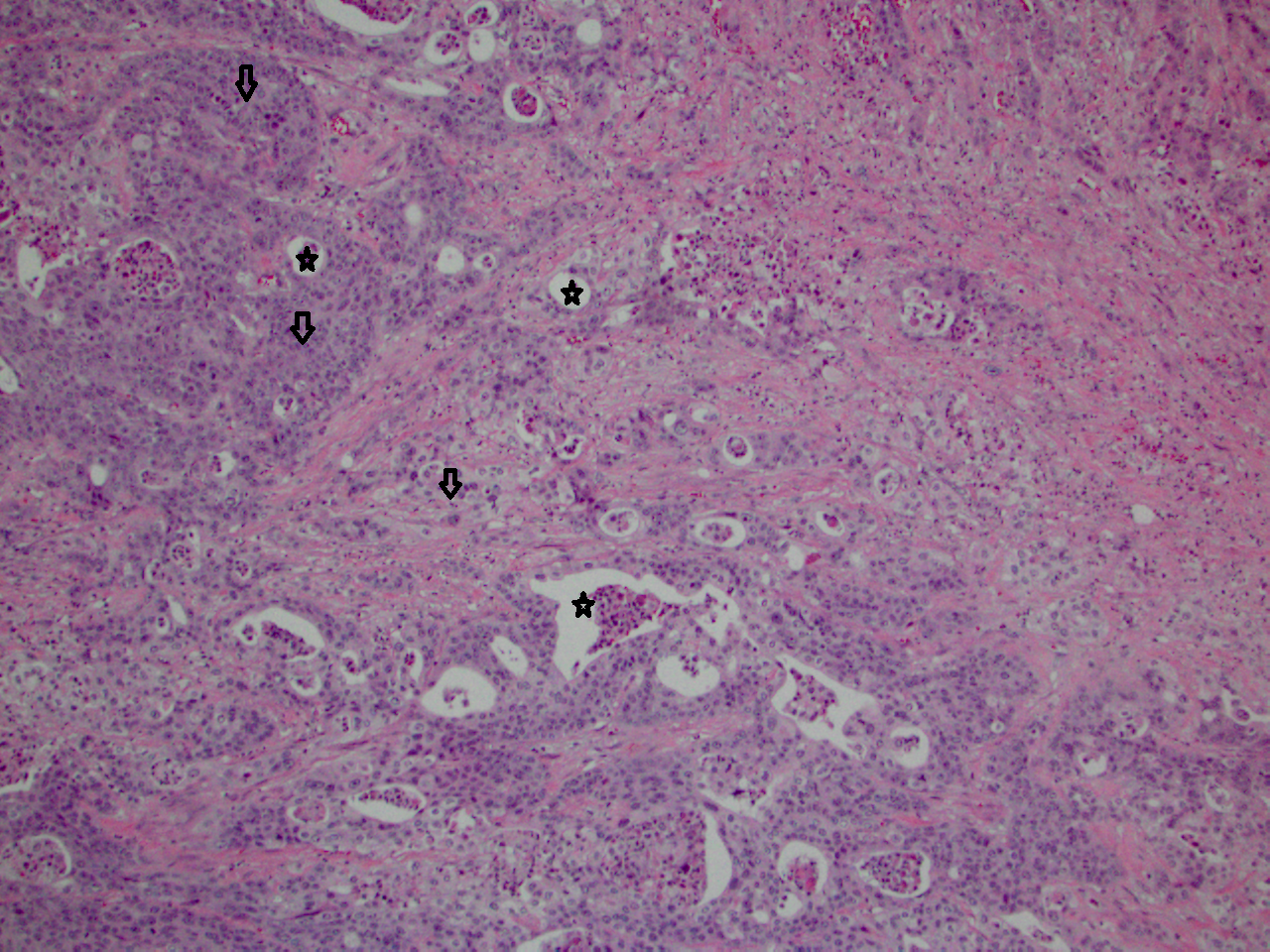
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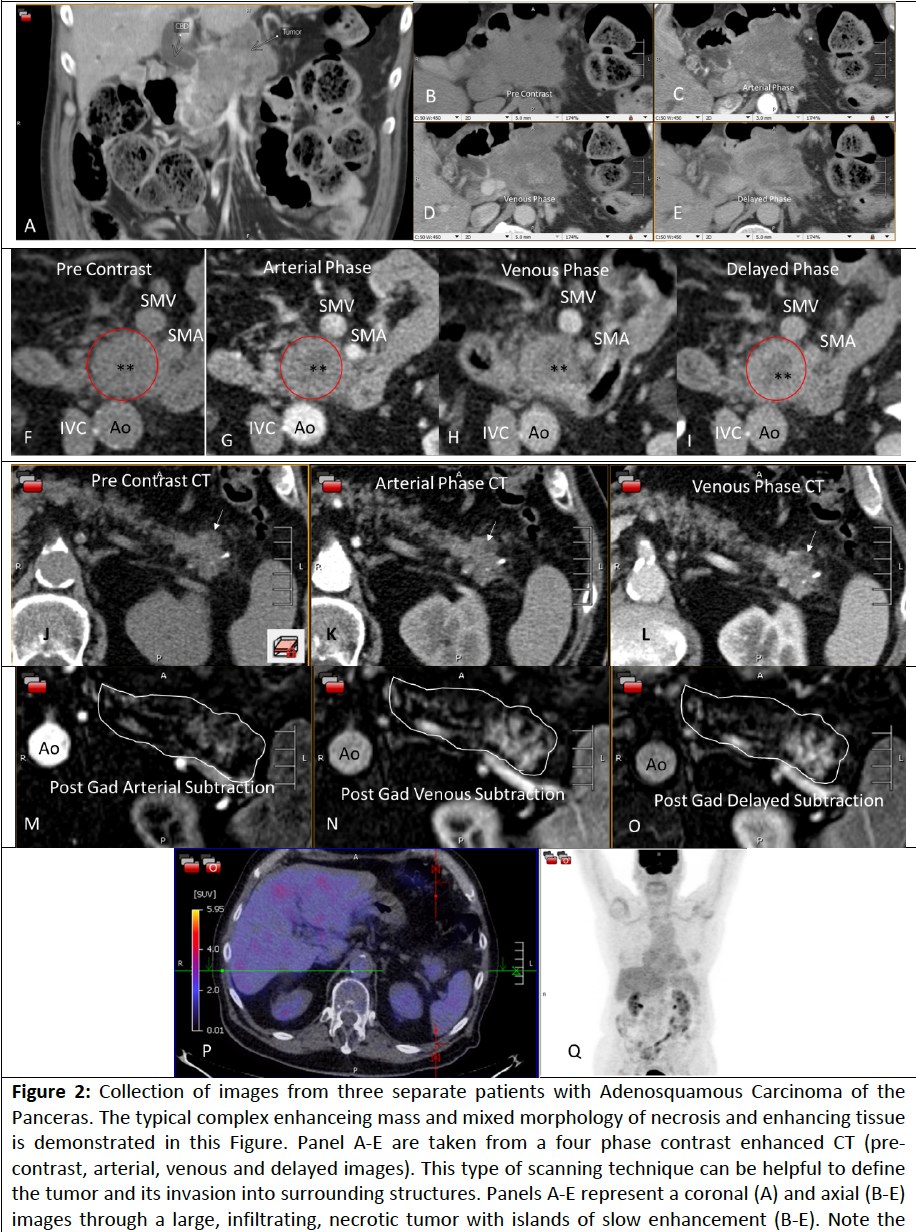
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**Figure 1 Typical pathology of Adenosquamous carcinoma of the pancreas.** H and E slides of two patient’s tissues, showing the adeno versus squamous component (Stars = adeno; arrows = squamous component). A: Tissue from head of pancreas; B: Tissue from tail of pancreas; both are G2, moderately differentiated cancers.



**Figure 2 Collection of images from three separate patients with adenosquamous carcinoma of the pancreas.** The typical complex enhancing mass and mixed morphology of necrosis and enhancing tissue is demonstrated in this figure. Panel A-E are taken from a four phase contrast enhanced CT (pre-contrast, arterial, venous and delayed images). This type of scanning technique can be helpful to define the tumor and its invasion into surrounding structrures. Panels A-E represent a coronal (A) and axial (B-E) images through a large, infiltrating, necrotic tumor with islands of slow enhancement (B-E). Note the islands of soft tissue enhancement increasing from arterial to delayed phase contrast enhanced CTs. These features are usually signs of very aggressive tumors. In another subject (Panels F-I) there is again a central area of necrosis (\*) surrounded by a ring of slowly enhancing tumor (red circle). Note the relative lack of surrounding soft tissue infiltration compared to the tumor on Panels A-E. Panels J-O are taken from a third subject and are an example of an atypical adenosquamous carcinoma involving the pancreas tail with a slowly enhancing, non-infiltrating lesion both on CT (Panel J-L) and post gadolinium subtraction MRI (Panel M-O). The white outline in Panels M-O outlines the contour of the pancreas with the enhancing lesion seen towards the tail of the pancreas. There is a small focus of necrosis present (arrow), a feature typical of adenosquamous carcinoma of the pancreas. The corresponding FDG PET/CT (Panel P and Q) is unusual in that it shows that this mass is not hypermetabolic unlike most Adenosquamous Pancreas Carcinomas. Ao: Aorta; IVC: Inferior vena caval; SMA: Superior Mesenteric Artery; SMV: Superior Mesenteric Vein.

**Table 1 Frequency of malignant exocrine pancreatic neoplasms**

|  |  |
| --- | --- |
| **Frequency of malignant exocrine pancreatic neoplasms** | |
| **Histological Subtype** | **Frequency** |
| Adenocarcinoma | 85% |
| Mucinous cyst adenocarcinoma | 2% |
| Adenosquamous | 0.38%-10% |
| Undifferentiated/anaplastic carcinoma | < 1% |
| Intraductal papillary mucinous carcinoma | 3% |
| Acinar cell carcinoma | < 1% |
| Rare subtypes | 4% |

Rare subtypes include signet ring cell carcinoma, giant cell tumor, cystadenocarcinoma, pancreatoblastoma, spindle cell carcinoma.

**Table 2 Incidence of adenosquamous carcinoma of the pancreas**

|  |  |  |
| --- | --- | --- |
| **Pancreatic cancer specimens** | **No. (%) of ASCP** | **Ref.** |
| 15185 | 81 (0.05) | [2] |
| 5075 | 46 (0.9) | [6] |
| 264 | 10 (3.8) | [8] |
| 391 | 13 (3.4) | [9] |
| 80 | 8(10) | [10] |
| 202 | 6 (3) | [11] |
| 3651 | 45 (1.2) | [12] |
| 45693 | 415 (0.9) | [13] |
| 237 | 7 (2.9) | [14] |
| 406 | 14 (4) | [15] |
| 1025 | 46 (4.5) | [16] |
| 24604 | 95 (0.38) | [17] |
| 635 | 20 (3.1) | [18] |
| 8372 | 25 (0.3) | [19] |
| 234 | 7 (2.9) | [20] |

ASCP: Adenosquamous carcinoma of the pancreas.

**Table 3 Molecular profiling of patients with adenosquamous carcinoma of the pancreas: Immunohistochemistry analysis**

|  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- |
| **IHC analysis percent positive expression (positive/number examined)** | | | | | | | | | | | | | | | | | | |
| **MRP1** | **BCRP** | **MGMT** | **TOP2A** | **TUBB3** | **PTEN** | **SPARC** | **TOPO1** | **RRM1** | **cMET** | **TLE3** | **TS** | **ERCC1** | **PGP** | **C-kit** | **PR** | **AR** | **ER** | **Her2** |
| 80 (8/10) | 80 (4/5) | 76 (16/21) | 78 (14/18) | 38 (3/8) | 41 (9/22) | 39 (9/23) | 38 (8/21) | 43 (9/21) | 33 (4/12) | 42 (5/12) | 38 (8/21) | 31 (4/13) | 11 (2/18) | 10 (1/10) | 5 (1/21) | 0 (0/21) | 5 (1/21) | 0 (0/22) |

IHC: Immunohistochemistry.

**Table 4 Molecular profiling of patients with adenosquamous carcinoma of the pancreas: fluorescence *in situ* hybridization/chromogenic *in situ h*ybridization analysis**

|  |  |  |  |  |
| --- | --- | --- | --- | --- |
| **FISH/CISH percent positive expression** **(Positive/number examined)** | | | | |
| **cMET** | **EGFR** | **Her2** | **TOP2A** | **ALK** |
| 9  (1/11) | 0  (0/6) | 0  (0/12) | 0  (0/2) | 0  (0/1) |

FISH: Fluorescence *in situ* hybridization; CISH: Chromogenic *in situ h*ybridization.

**Table 5 Molecular profiling of patients with adenosquamous carcinoma of the pancreas: Mutated gene analysis (either sanger or next generation sequencing)**

|  |  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- | --- |
| **Mutated genes percent positive** **(Number found/Examined)** | | | | | | | | |
| **cMET** | **KRAS** | **TP53** | **BRAF** | **NRAS** | **SMAD4** | **cKIT** | **PIK3CA** | **EGFR** |
| 13  (1/13) | 100  (16/16) | 50  (4/8) | 0  (0/9) | 0  (0/9) | 25  (2/8) | 0  (0/9) | 0  (0/11) | 0  (0/10) |