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**Regression of cardiovascular remodeling in hypertension: Novel relevant mechanisms**

Jalil JE *et al*. Cardiovascular remodeling regression in hypertension

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**Abstract**

Asymptomatic organ damage due to progressive kidney damage, cardiac hypertrophy and remodeling put hypertensive patients at high risk for developing heart and renal failure, myocardial infarction and stroke. Current antihypertensive treatment normalizes high blood pressure, partially reverses organ damage, and reduces the incidence of heart and renal failure. Activation of the renin–angiotensin system (RAS) is a primary mechanism of progressive organ damage and, specifically, a major cause of both renal and cardiovascular fibrosis. Currently, inhibition of the RAS system (mainly with (ACE) inhibitors or angiotensin II (Ang II) receptor antagonists) is the most effective antihypertensive strategy for normalizing blood pressure and preventing target organ damage. However, residual organ damage and consequently high risk for cardiovascular events and renal failure still persist. Accordingly, in hypertension, it is relevant to develop new therapeutic perspectives, beyond reducing blood pressure to further prevent/reduce target organ damage by acting on pathways that trigger and maintain cardiovascular and renal remodeling. We review here relevant novel mechanisms of target organ damage in hypertension, their role and evidence in prevention/regression of cardiovascular remodeling and their possible clinical impact as well. Specifically, we focus on the signaling pathway RhoA/Rho kinase, on the impact of the vasodilatory peptides from the RAS and some insights on the role of estrogens and myocardial chymase in cardiovascular hypertensive remodeling.

**Key words:** Remodeling; Hypertrophy; Rho kinase; MYPT-1; Angiotensin; Angiotensin1-9; Chymase; Angiotensin1-7

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**Core tip**: Antihypertensive treatment normalizes high blood pressure, partially reverses organ damage, and reduces the incidence of heart and renal failure. However, residual organ damage and high risk for cardiovascular events still persist. We review here novel relevant mechanisms of cardiovascular damage in hypertension, their role and evidence in prevention/regression of cardiovascular remodeling and their possible clinical impact. We focus on the signaling pathway RhoA/Rho kinase, on the impact of the vasodilatory peptides from the renin angiotensin system and some insights on the role of estrogens and myocardial chymase in cardiovascular remodeling due to hypertension.

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**INTRODUCTION**

Target organ damage in hypertension causing asymptomatic renal dysfunction, atrial size enlargement along with cardiac hypertrophy and remodeling place hypertensive subjects at a very high risk condition to develop cardiac failure, progressive kidney disease, myocardial infarction and stroke as well. Up-to-date antihypertensive drug therapy reduces to normality elevated arterial blood pressure, doe revert organ damage to some extent, and diminishes the occurrence of cardiac and renal disease. However, the clinical impact of all antihypertensive drug classes is not substantially different among most clinical outcomes when the blood pressure effect is equivalent[1]. Besides, permanent stimulation of the renin–angiotensin axis is a fundamental process of continuing damage to the target organs and a main cause of fibrosis both in the kidney an also in the myocardium. Current pharmacological blockade of the renin–angiotensin axis (primarily with inhibitors of ACE or by blocking the angiotensin II receptor) is a most effective antihypertensive strategy for normalizing high blood pressure and for preventing continuing end organ damage[2]. However, both residual damage in the target organs and consequently, a condition of high hazard for experiencing major clinical events still persist.

Thus, in hypertension, it is most crucial the development of new therapeutic viewpoints, further than only reducing blood pressure to better prevent/decrease target organ damage by aiming to paths triggering and maintaining cardiovascular remodeling and also in the kidney[2]. Our purpose is to review here three novel mechanisms of target organ damage in hypertension, their role and evidence on regression of cardiovascular remodeling and their possible clinical impact as well. Specifically, we will concentrate on the signaling pathway RhoA/Rho kinase, on the impact of the vasodilatory peptides from the renin angiotensin system and on the role of estrogens and the myocardial chymase-angiotensin II pathway in cardiovascular hypertensive remodeling. Interestingly, the 3 aforementioned mechanisms interact strongly with the renin angiotensin system at the cardiovascular level.

***Rho kinase, hypertension and cardiovascular remodeling***

The small protein Rho (a guanosine triphosphatase) and its target Rho kinase (ROCK), have important functions in blood pressure modulation, by regulating smooth muscle contraction and additionally in cardiovascular remodeling. Agonists of receptors coupled to the G protein in the cell membrane (such as endothelin, angiotensin II, or noradrenalin), growth factors and cytokines activate Rho[4-6] (Figure 1). Some actomyosin-associated proteins, such as myosin light chain (MLC) phosphatase, myosin light chain 2, LIM-kinase, ezrin radixin-moesin and adducin are considered physiological ROCK substrates[7-9] (Table 1 and Figure 1). In non-hypertensive people, activation of the classical renin angiotensin system induced by low-salt diet does increase RhoA-ROCK signaling and does stimulate activation of the RhoA guanine exchange factor Arhgef1, which is implicated in vascular tone regulation and in hypertension induced by angiotensin II) in circulating mononuclear cells[10]. Immediately after Rho activation, this small protein is translocated to the cell membrane where it phosphorylates and activates ROCK (Figure 1), controlling in this way several cellular functions (Table 2), the majority of them related to remodeling. Activated ROCK does phosphorylate MLC phosphatase, which becomes inhibited. This cascade does stimulate tonic contraction of the smooth muscle within the vessels, development of stress fibers, and also cell migration. Thus, activation of both Rho and ROCK has significant effects on numerous cardiovascular diseases[4-6,11,12], especially in hypertension[13].

Administration of ROCK inhibitors reduces blood pressure effectively in the rat with spontaneous hypertension (SHR)[14-17], in the DOCA salt hypertensive model in rats[14,15,18], renal hypertensive rodent[14], L-NAME hypertensive rats[19,20] and in also in normotensive rats[14,15,17,20] which indicates that blood pressure fall by inhibitors of Rho kinase does not depend on the mechanism of hypertension[21]. Furthermore, the ROCK intracellular signaling cascade is activated in human hypertension[22,23] where elevated ROCK activity appears to be a consequence derived from up-regulation of the renin angiotensin system and also from higher levels of reactive oxygen species (ROS)[8,9]. ROCK inhibition decreases smooth muscle contractility by reducing MLC phosphorylation in the smooth muscle cell and by enhancing endothelial function through reestablishing eNOS activity and NO production[8,14,18,24-28].

Cardiovascular inflammation and remodeling are also reduced by ROCK inhibition[8] through: (1) suppressing the levels of cytokines and adhesion molecules such as PAI-1, MCP-1 and the transforming growth factor 1 in endothelial and in smooth muscle cells[18,24,29,30]; (2) by inhibiting in endothelial cells ROS production through down-regulation of NADPH oxidases[24,31,32]; (3) by reducing in smooth muscle cells secretion of cyclophilin A[33] and also (4) by augmenting the levels of angiotensin 1-9[18]. Moreover, ROCK inhibitors delivered in the brainstem reduce blood pressure and sympathetic nerve in hypertensive rodents[34,35].

In hypertension, there are experimental data available on the significant role of ROCK activation on developing myocardiac hypertrophy, remodeling and ventricular dysfunction. In the rodent with salt-sensitive hypertension, cardiac hypertrophy was importantly reduced by using Y-27632, a specific ROCK inhibitor[36]. In this experimental model, upregulated RhoA, ROCK gene expression and phosphorylated MLC in the stage with hypertrophy were also inhibited by ROCK inhibition[36]. Besides, fasudil attenuated cardiac fibrosis possibly throughout inhibition of inflammatory cells myocardial infiltration in hypertensive rats[37]. Additionally, activated ROCK in the aorta observed in rats with genetically determined elevated levels of angiotensin converting enzyme and the peptide angiotensin II, is reduced by Fasudil administration causing reduced gene expression that stimulate vascular remodeling (like transforming growth factor 1, plasminogen activator inhibitor 1 and MCP-1) and also enhances oxidative species in the vasculature[24].

Long-term inhibition of ROCK using fasudil ameliorated diastolic cardiac failure in the Dahl hypertensive rat[38]. Besides, in rats with LVH due to pressure overload, inhibition of ROCK with GSK-576371 recovered LV chamber geometry, improved diastolic function and reduced myocardial fibrosis[39] and recently, long term treatment of DOCA-salt and LNAME hypertensive rats with the more potent ROCK inhibitor SAR407899 reduced hypertension and cardiac and renal remodeling in a dose-dependent way in both models[19]. Interestingly, in DOCA hypertensive rats, blood pressure reduction and protective effects on hypertensive organ damage of SAR407899 were superior compared to amlodipine and also to ramipril[19] and hearts of hypertensive DOCA or LNAME animals treated with SAR407899 had significantly better systolic LV function (measured as heart power *in vitro*). Additionally, endothelial-dependent relaxation was significantly and dose-dependently improved after long-term treatment with SAR407899[19]. An important amelioration of myocardial interstitial fibrosis and expression of collagen genes and of CD3 and CD68 (markers of infiltrating macrophages and leukocytes) in both models was observed, possibly explained by the relevant Rho kinase function in cellular migration and cytokinesis through cytoskeleton modulation[19].

In more advanced heart disease secondary to hypertension, and evident impairment in cardiac function (both diastolic and systolic), it is very likely that ROCK activity levels be rather similar to Rho kinase activation levels observed in cardiac failure secondary to different mechanisms and its inhibition could produce in this situation similar benefits. In the mouse overexpressing Gαq,deletion of ROCK1 gene did prevent LV chamber dilatation and improved cardiac contractility[40]. Furthermore, in cardiomyocytes in culture, ROCK activation up-regulates Bax *via* p53 to induce apoptosis[41]. In the transgenic mouse that overexpress the isoform MYPT2, activation of myosin phosphatase induced LV function decline and remodeling, probably by reducing calcium sensitivity, along with impairing the myofibrillar organization, which is the original report about the functions of both MYPT2 and myosin phosphatase, and the consequences of *in-vivo* cardiac MLC phosphorylation[42].

**Assessing Rho kinase activation in human circulating leukocytes, a possible marker of cardiovascular remodeling and risk:** In people diagnosed with metabolic syndrome (MetS), Liu *et al*[43] reported for the first time significantly increased ROCK activity by 31% through the measurement of ROCK phosphorylationin circulating leukocytes using the approach of determining myosin binding subunit phosphorylation (MBS). At the same time they observed that plasma concentrations of high-sensitivity C-reactive protein were substantially higher and that circulating levels of adiponectin were significantly lower in MetS subjects as compared with control subjects. Additionally, in this population they found that increased ROCK activation was significantly related with body mass, waist circumference, fasting glucose, high-sensitivity C reactive protein, and additionally with triglyceride levels[43]. In this clinical study the probability of increased ROCK activity was considerably increased with the amount of MetS components. Experimental findings indicate that insulin resistance promoted by Rho kinase is implicated in myocardial damage in rats with MetS and that this action of Rho kinase is probably through the IRS-1-PI3-kinase-AKT signaling cascade[44]. In humans, ROCK activation inleukocytes is also enhanced by smoking and does predict endothelial dysfunction[45].

In circulating leukocytes, Hata *et al*[46] measured the activity of Rho kinase by assessing the relation amid phosphorylated myosin-binding subunit (p-MBS) on myosin light chain phosphatase to the total MBS and also the change on the blood flow in the forearm (FBF) as a pharmacological action of the distinctive Rho kinase inhibitor fasudil using strain-gauge plethysmography in control subjects and also in subjects diagnosed with a cardiovascular illness. Compared to healthy subjects, they found that leukocyte p-MBS/total-MBS ratio was substantially higher (by 90%) in the diagnosed patients[46]. Besides, they found that the characteristic inhibitor of ROCK fasudil increased FBF by 300% only in their patients with cardiovascular disease, but this was not the case not in the healthy control group[46]. Moreover, they found an important relationship between leukocyte p-MBS/total-MBS and maximal FBF induced by fasudil in the group with cardiovascular disease (r=0.59), not in the healthy subjects.

Lately, we have evaluated the level of ROCK activation in leukocytes obtained from venous blood, by quantifying the relationship of phospho to total MLC phosphatase 1 (known as MYPT1-P/T) as a potential remodeling marker in untreated hypertensive patients (HT), in HT patients with LVH or with type II diabetes mellitus receiving specific treatment and also in patients with congestive cardiac failure and LV systolic reduced function[47-49].

In a recent clinical follow up trial with the aim to determine the correlation amid the observed activity of ROCK and a first main cardiovascular event along with hospitalization rates for congestive cardiac failure, the levels of ROCK activity were determined in leukocytes by the technique of Western blot in more than 600 subjects who undertook a health-screening examination[50]. After a median period of 42 mo of follow-up, 29 deaths were registered (10 of them because of cardiovascular causes), 2 of them were diagnosed with a myocardial infarction, in 20 of them at least one revascularization procedure was performed, 15 developed a stroke, and 17 required hospitalization for congestive cardiac failure[50]. In the above-mentioned study, once the adjustment for several confounding variables (such as age, gender, known risk factors and other relevant predictors of cardiovascular illness) was performed, ROCK activity remained as a robust independent indicator of a first main cardiovascular event (the hazard ratio was 2.19), of death as a consequence of cardiovascular disease (hazard ratio, 2.57), cerebrovascular accident (hazard ratio, 2.14), and the clinical necessity for revascularization (reported hazard ratio was 2.68)[50]. The authors concluded that ROCK activity levels determined in circulating leukocytes may be a new marker of cardiovascular events and propose that its inhibition may be a novel therapeutical approach to achieve effective prevention of cardiovascular disease[50].

**Rho kinase activation in subjects with essential hypertension:** In a case-control study with the goal of comparing ROCK activation levels in subjects diagnosed with hypertension against a healthy normotensive control group in regard to the existence of LVH assessed by 2D-echocardiography we measured LV mass and dimensions in addition to LV performance and ROCK activation levels in leukocytes from venous blood (by MYPT1-p/t levels through Western blot)[47].Compared to non-hypertensive controls, MYPT1-p/t was considerably higher by 450% in the group lacking LVH and by 900% in the group with confirmed LVH by 2D-echocardiography (Figure 2). In contrast with the hypertensive subjects without LVH, MYPT1-p/t was considerable higher (by 200%) in the subjects with hypertension along with echocardiographic confirmed LVH[47]. Additionally, in the hypertensive subjects with evidence of eccentric LVH had an MYPT1-p/t relationship remarkably higher (by 400%) compared to hypertensive subjects and no eccentric LVH. Patients having an E/e´ ratio measured in the transmitral diastolic flow ≥15 showed a substantially higher MYPT1-p/t relationship (26%) as related to the levels in those subjects with a smaller E/e´ ratio. This study concluded that ROCK activation levels determined in leukocytes from venous blood are significantly raised up in hypertensive subjects with definite cardiac hypertrophy compared with HT patients without LVH. ROCK activation is additionally increased when eccentric hypertrophy is present. Therefore, in subjects with essential hypertension, ROCK phosphorylation/activation determined in leukocytes from venous blood is correlated to pathological myocardial remodeling and could contribute as one indicator of LVH[47].

Similarly, Hata *et al*[51] recently observed significantly higher Rho kinase activation levels in circulating leukocytes in subjects with essential hypertension when contrasted to healthy individuals by 37%. Besides, in these hypertensive patients under antihypertensive treatment, ROCK activity levels were substantially lowered in patients using calcium channel antagonists compared to patients receiving as antihypertensive treatment inhibitors of the renin-angiotensin-aldosterone axis, thiazides, or β-blockers[51]. These observations suggest that increased Rho kinase activity associated to hypertension may cause activation of leukocytes along with leukocyte infiltration into the vessel wall, which favors atherosclerosis progression[51] (as well as remodeling) and suggest the possible clinical relevance of determining in this context the degrees of Rho kinase activation. In a randomized clinical study with the aim of assessing the impact of the specific aldosterone receptor blocker eplerenone, on the endothelial function determined by flow mediated dilatation (FMD) and on Rho kinase activation as well, determined in leukocytes obtained from venous blood in subjects with essential hypertesion, 60 patients were received eplerenone, the antagonist of calcium channels nifedipine, or losartan for 48 wk[52]. They observed that FMD was increased and leukocyte Rho kinase activity was reduced with eplerenone, wheras nifedipine reduced Rho kinase activity but did not modify FMD[52]. In the aforementioned clinical trial losartan augmented FMD but did not modify Rho kinase activity. In this clinical study, both the blood pressure reducing effect and also the vasodilation levels induced by nitroglycerin were similar with the different three antihypertensive drugs throughout the follow-up[52].

Rho kinase activity is importantly increased in subjects with very high cardiovascular risk and remodeling hazard, as is frequently observed in subjects with the combined diagnosis of both type 2 diabetes mellitus along with essential hypertension. In a cross sectional clinical study carried through comparing three groups of subjects[48]: Essential hypertensive patients under no medical treatment, patients with both hypertension and type II diabetes under treatment (with similar degrees of left cardiac mass) and normotensive control subjects, in the patients having the two aforementioned medical conditions, increased ROCK activation (determined in venous blood leukocytes) was found as compared to hypertensive subjects not getting pharmacological antihypertensive treatment[48] (Figure 3). In this clinical study, in the diabetic hypertensive patients compared to both non-diabetic hypertensives and to normotensive controls, increased levels of oxidative stress were found[48]. These findings were correlated with reduced arterial compliance and could help to explain the unfavorable vascular remodeling that is commonly detected in hypertensive plus diabetic patients receiving treatment[48].

In a very recent prospective clinical trial aimed to evaluate the effect of the specific ROCK inhibitor fasudil on diastolic LV function parameters observed in a group of individuals diagnosed with type 2 diabetes presenting with preserved systolic performance, 250 patients with the established clinical diagnosis (62% of them hypertensives), were allocated to receive the ROCK inhibitor (fourteen days, 30 mg *iv* twice per day) or to placebo[53]. As planned, echocardiographic parameters were determined before and after 1 mo receiving this treatment. In relationship with the group that was randomized to placebo, in the subjects that were randomized to fasudil, an important reduction in both blood pressure (diastolic) and in echocardiographic late diastolic transmitral flow was observed[53]. In the aforementioned clinical study, deceleration time, relaxation time (isovolumic), peak early annular diastolic velocity (e'), peak of late diastolic annular velocity, as well as the E to e' ratio also showed an important recovery by fasudil administration for one month[53]. Moreover, the Em to Am ratio, both the relaxation (isovolumic) and deceleration times, along with E to e' ratio values observed after receiving treatment with fasudil in the subjects with baseline diminished left ventricular chamber relaxation diverged significantly from what was noticed in the patients presenting with normal left ventricular chamber relaxation. Accordingly, clinical ROCK inhibition by using fasudil improved for the first time left ventricular chamber diastolic function parameters in diabetic patients (the majority of them hypertensives) and normal systolic performance[53].

**Rho kinase increased signaling in subjects with progressive cardiac remodeling: the case of congestive cardiac failure with systolic function decline:** Rho kinase activity is markedly augmented in those subjects with established cardiac failure (CF) due to systolic dysfunction. In a cross sectional study comparing control healthy subjects with patients with chronic and clinically stable CF due to systolic dysfunction under optimal medical treatment, we observed that Rho kinase activation (determined in circulating leukocytes as the MYPT1-P/T ratio) was increased by 100-fold and that it was inversely related with ejection fraction[49]. Interestingly, in those patients with CF with LV diameter ≥ 60 mm MYPT1-P/T was significantly more elevated than in the CF subjects with LV diameter < 60 mm. Thus, ROCK activity is markedly augmented in patients with stable chronic CF receiving optimal medical treatment, and Rho kinase signaling is robustly related to pathologic left ventricular chamber remodeling and to systolic function decline as well[49]. Another clinical study examined whether ROCK activation (determined in venous blood leucocytes) is increased in congestive CF and how it is related with the clinical prognosis in 170 patients admitted with this clinical condition[54]. Patients were prospectively followed up for 14.4 ± 7.2 mo or up to the event of cardiac death. Observed Rho kinase signaling in the patients with congestive CF was significantly higher than that of two control groups. The protein concentrations of both Rho kinase isoforms (ROCK1 and ROCK2) as well as the measured activation of the up-river Rho kinase cascade GTPase RhoA in the congestive CF patients were significantly higher than what was observed in both control groups[54]. Dyspnea at rest, reduced left ventricular systolic function and impaired renal function were all independent factors predicting Rho kinase activity levels at baseline in the subjects with congestive CF[54]. By combining Rho kinase activity with N terminal pro brain natriuretic peptide (NT-proBNP) an incremental value in the estimate of long-term mortality (when compared with only the NT-proBNP measurement) was observed. Thus, Rho kinase activity is elevated in these patients with extreme myocardial remodeling, it is also associated with higher mortality and it might be an additional biomarker to congestive CF risk assessment[54]. In a clinical study to evaluate whether Rho kinase activity in venous blood leukocytes is elevated in subjects presenting with an established acute coronary syndrome and if Rho kinase activation does predict long‐term cardiovascular events, 188 patients with ACS and 61 control subjects were evaluated[55]. The authors found significantly increased ROCK activity in the two clinical groups (myocardial infarction and unstable angina) when it was compared to control subjects. Besides, patients with both elevated NT-proBNP and Rho kinase activity on admission had a five-fold hazard of a major cardiovascular outcome in relation to the observed hazard in those subjects with low NT-proBNP and low ROCK activity[55]. Their main conclusion was that by combining Rho kinase activity and NT-proBNP levels a subset of acute coronary syndrome patients at particularly high risk might be identified[55].

Altogether, these findings strongly suggest that ROCK activation in circulating human leukocytes is directly related to pathological cardiovascular remodeling, from early target organ damage in hypertension to extreme cardiovascular disease. Besides, as this measurement possibly mirrors remodeling it has prognosis value for disease progression, clinical events and conceivably for target organ damage/disease prevention and regression.

***Impact of the vasodilatory peptides from the renin angiotensin system on mechanisms in hypertension and cardiovascular remodeling***

Remodeling of the cardiovascular (CV) structures does occur as a response, not only to modifications in blood arterial pressure or ﬂow but also to variations in the neural and hormonal milieus, where the renin-angiotensin-aldosterone axis exerts a major inﬂuence[56]. The aforementioned neurohormonal system, one of the oldest phylogenetically hormonal systems, is most recognized because of its fundamental role in regulating hydromineral and cardiovascular homeostasis[56,57]. The renin-angiotensin-aldosterone axis is a fundamental element of CV physiology having a main pathophysiological role by regulating vascular tone, blood pressure, sodium and potassium balance and vascular responses to both injury and inflammation[58]. Long lasting activation of the renin angiotensin system, throughout both the octapeptide angiotensin II and the mineralocorticoid hormone aldosterone, causes hypertension and at the same time does stimulate prohypertrophy, proinflammation, prothrombosis, and atherogenesis pathways strongly linked with hypertensive organ damage.

During a long period, a main research focus has been production and signaling of Angiotensin II, highlighting both the angiotensin I converting enzyme (ACE) and renin regarding its production. With the precise description of both main angiotensin II receptors 1 (AT1R ) and 2 (AT2R), most of the issues related to the biochemistry, pharmacology, and physiology of the renin-angiotensin-aldosterone system seemed to be resolved[57,59]. Extensive studies of the RAAS for a long time, were mainly focused on four classes of drugs targeting the renin-angiotensin-aldosterone system at different levels: angiotensin-converting enzyme inhibitors (known as -prils), Angs receptor blockers (-sartans), renin inhibitors (-kirens) and mineralocorticoid receptor antagonists (Figure 4). All of them are cornerstones in the treatment of HT. However, there are more recent discoveries extending our understanding of important properties of the renin-angiotensin-aldosterone system. In the most recent decades, increasing evidence has been accumulated indicating an exceeding complexity of the RAAS. Among some novel discoveries is the finding and location of the physiologically important aions of Ang-(1–7)[60], Ang-(1-9)[61] and alamandine[62] acting through identifiable tissue receptors[60,62,63] (Figure 4).

**New vasodilatory peptides in the RAAS and hypertensive remodeling:** The therapeutic effectiveness of “classic” blocking the RAAS for treating HT and also related cardiovascular illness has been extensively well established. However, in the most recent decades growing facts and data indicate that the roles and biological functions of the RAAS go beyond the effects initially described. Currently, the amount of biologically relevant end-products of the RAAS is even now increasing, which raises new possibilities to attack through this axis cardiovascular disease, and specifically hypertensive CV remodeling. These facts are particularly accurate for the peptides Ang (1–7) along with Ang (1-9) and lately, for alamandine (Figure 4), and in most of the situations these peptides display biological effects opposing to Angiotensin II.

**Ang-(1-7)**: Production of the active molecule Ang-(1–7) mostly depends on the cleavage of the octapeptide Ang II which is performed by ACE2 (Figure 4). Moreover, Ang-(1-7) can be also formed through hydrolysis of the decapeptide Ang I performed by other peptidases such as prolyl-endopeptidase, neutral-endo- peptidase (NEP), and timeth-oligopeptidase which cleave the Pro7-Phe8 bond to remove the final three amino acids. The function of the prolyl endopeptidase (PEP)[64],oligopeptidase (TOP) and of NEP[65] in the enzymatic hydrolysis from Ang I onto Ang-(1–7) depends on tissue distribution and substrate availability of the enzymes. Neprilysin behaves as an especially active enzyme which has been primarily located in the vessel endothelial cells while the thimet oligopeptidase enzymatic activity is relevant in the cleavage toward Ang-(1–7) occurring within the smooth muscle vascular cells[66,67]. Ang-(1-7) is also formed from Ang II by ACE2[68]. The level of Ang‑(1-7) is regulated by the action of ACE which hydrolyzes Ang-(1-7) to Ang-(1-5)[65] (Figure 4).

Ang-(1-7) produces its biological effects and acts throughout its own receptor, the Mas receptor (MasR), one of the membrane receptors coupled to G protein[8]. This receptor mediates the current known actions of this heptapeptide, as most of them can be prevented by the specific blocker, D-Ala7-Ang-(1-7) (A779)[8]. The Ang-(1-7) observed effects acting throughout the MasR in the CV system consistently include vasodilation, antihypertrophy, antiarrhythmogenesis, antifibrogenesis and antithrombogenesis[69-71].

**Angiotensin (1-9)**: The first observations about Ang-(1-9) (Figure 4) showed a rapid appearance of leucine after the injection of radiolabeled Ang I into dog renal and pulmonary arteries[72,73], by the enzymatic action of a carbopeptidase that was breaking down Ang I forming des-Leu10 Ang I. It is possible to generate Ang-(1-9) from Ang I through the effect produced by several enzymes (carboxypeptidase-type), including cathepsin A and ACE2[74-76], although at a relatively slow rate compared to the making of Ang (1–7) starting from Ang II[77]. Moreover, it was observed that an inhibitor of ACE2 doesn’t have an effect on Ang-(1–9) formation, but benzylsuccinate, a CxA inhibitor, does stop the formation of Ang-(1–9) and rises the levels of Ang I in heart membranes[78]. Alternatively, it is possible to cleave Ang-(1–9) to Ang-(1–7) by the ACE carboxypeptidase or by the effects of other enzymes including prolyl endopeptidase (POP), neutral endopeptidase (NEP) and thimetoligopeptidase 1 (TOD)[79,80]. Recently, it was found that Ang-(1–9) has the capability to be hydrolyzed into the peptide Ang-(2–9) by aminopeptidase A[81] (Figure 4).

Initially Ang-(1-9) was considered as a biologically non active peptide, operating indirectly by competing with Ang I for the ACE active site, and therefore reducing Ang II levels while increasing Ang-(1-7)’s[76,77,82]. However, increasing evidence has confirmed that Ang-(1-9) does work as a molecule with relevant cardiovascular effects both *in vitro* in addition to *in vivo*, through the AT2R[61,63,83,84].

**Alamandine:** Recently a new member of RAAS has been discovered, the heptapeptide Ala-Arg-Val-Tyr-Ile-His-Pro, known as alamandine[62] (Figure 4). By using mass spectrometry, alamandine was identified as a chemical product of a catalytic hydrolysis of Ang A, an octapeptide, by ACE2[62]. Alamandine is composed by a sequence of amino acids which is extremely similar to Ang-(1–7). Both peptides diverge only in one amino acid residue (alanine in place of an aspartate residue) located at the amine-terminus. Alamandine may be also synthesized by decarboxylation of the N-terminal aspartate amino acid residue from Ang-(1–7). The enzyme which is in charge of the ultimate reaction remains unknown[62]. However, Alamandine degradation is not yet completely understood. Nevertheless, it is possible that aminopeptidases have a most relevant role, since the subtraction of Ala1 could conduct to form Ang-(2-7), deemed as a non-active molecule, although it shows inhibitory activity of ACE[85]. Other Ang-(1-7)-degrading enzymes, such as NEP or neprilysin, may also participate, given alamandine’s similarity to Ang-(1-7)[86] (Figure 4).

Although Alamandine is rather similar to the peptide Ang-(1-7) and the biochemical effects of both molecules look to be close, alamandine acts through a different receptor, the Mas-related G protein–coupled receptor (MrgD)[62]. Alamandine produces endothelial-dependent vasodilation in rat and mice aortic rings[62]. It has recently been observed that oral delivery of alamandine (by including it within HP-β cyclodextrin), produced similar effects to those already observed for Ang-(1-7) such as a long-term antihypertensive effect in SHR and a main reduction of cardiac deposition levels of collagens I and III as well as fibronectin in isoproterenol-treated rats[62].

***Target receptors for the second RAAS arm***

**Mas receptor and AT2 receptor:** the two receptors, the AT2R and the Mas receptor, are GPCRs (Figure 4) with their conventional seven transmembrane domains[60,68]. Interestingly, even though their signaling mechanisms are quite unusual for GPCRs and not completely understood, again, major resemblances have been observed. For both receptors, signaling by activating phosphatases, particularly the Src homology 2 domain-containing protein tyrosine phosphatase (SHP)-1 and SHP-2, seems to be crucial[87–90]. In both situations, it has been found that phosphatase activation does interfere, in an inhibitory way, with kinase driven signaling cascades, producing inflammation or hypertrophy involving molecules like the mitogen-activated protein kinases or the nuclear factor *κ*B[91,92]. Another shared signaling mechanism of importance is the augmented nitric oxide (NO) synthesis and consequent increase of cGMP levels, which does mediate the vasodilation effects of both receptors[93–95]. Furthermore, these two receptors are able to develop dimers with the AT1R which results in a functional inhibition of this latter one[96,97].

**The Mas-related gene D (MrgD) receptor:** the Mas-related gene D (MrgD) belongs to the G protein coupled receptors (GPCRs) family and it is associated to the MasR[98]. The MrgD is located in the myocardium in addition to the vessels wall[62]. Shinohara *et al*[99] found that a small amino acid, β-alanine could internalize the MrgD, and thus be able to induce intracellular influx of calcium and to inhibit production of cAMP in Chinese hamster ovary cultured cells (CHO) that expressed rat, mouse, or human MrgD[99]. The effect in calcium influx can be understood by the connection of the MrgD with the G-protein α subunit (Gq), and the cAMP suppression does suggest an interaction concerning the MrgD with the inhibitory G protein (Gi)[99]. Furthermore, MrgD activation through β-alanine also suppressed KCNQ/M-type potassium channels, and in this way also increased neuron excitability by means of the Gq and phospholipase C (PLC) pathways[100]. In addition, the incubation of alamandine with CHO cells that are transfected with MrgD induces significant NO release[62].

It has been proposed that this MrgD receptor is related to pain sensation[101], sensitiveness to thermic and mechanical stimulation[102], and tumorigenic activity[103]. It was also observed that MRGD is able to transduce intracellular signaling of Ang-(1–7)[104].

***Clinical approach to the Ang-(1-7)-MasR, Ang-(1-9)-AT2R and Alamandine-MrgD axis in hypertension and cardiovascular remodeling***

**Ang-(1-7)-MasR Axis:** ACE inhibitors (ACEI) and Ang II receptor blockers (ARB) can affect in part the ACE2-Ang-(1-7) system. Experimental studies in myocardial infarcted rats showed that chronic administration of enalapril prevented myocardial hypertrophy and contractile dysfunction in addition to increased ACE2 activity in plasma and in the ventricular wall[105].

ACEI increase the Ang I levels, which are hydrolyzed to produce Ang-(1-7), through the actions of both ACE2 and NEP. The arterial pressure reduction effects due to ACEI are also associated to increased excretion of Ang-(1-7), an observation reported in urine collected from subjects with essential hypertension receiving the ACEI captopril during 6 mo[106]. It´s well recognized that ACEI are able to diminish excretion of urinary protein in subjects with established type 2 diabetes mellitus[107], and interestingly, in the ACE2 knock-out mice the proteinuric blocking effect of ACEI disappears[108]. At this respect, ARBs may be here markedly effective because elevated Ang II levels as a consequence of them will promote Ang-(1-7) production[109]. Additionally, the low affinity binding of Ang-(1-7) to the AT1 receptor may allow this peptide to work as an antagonist in the presence of Ang II[110]. In this regard, normotensive rodents with high ACE and Ang II along with low NEP activity[111] and Ang-(1-7) concentration[112] (genetically determined) showed a higher hypertensive response (chronic) after renovascular hypertension induction[113]. Besides, the inverse correlation observed among the amounts of both Ang II in addition to Ang-(1-7) in the aforementioned rodents, determined increased cardiac fibrous tissue deposition after isoproterenol administration[114] and also ROCK activation in the aortic wall as well as stimulation of genes that promote vascular remodeling (such as the monocyte chemoattractant protein 1 gene, the transforming growth factor 1 gene, and plasminogen activator inhibitor gene)[24] and also higher oxidative stress levels in the vessels wall in normotensive rodents[115].

In humans, similar relationships have been observed[116,117]. Particularly, in hypertensive patients having the DD-ACE genotype (with increased ACE levels), the Ang-(1-7) blood levels were reduced by 4 fold as compared to those observed in patients having the II-ACE genotype (and consequently lower ACE levels)[116]. In these subjects we reported an important effect of the I/D ACE genotype on circulating NEP enzymatic activity in addition to an interactive effect amid the I/D ACE genotype status and the hypertensive condition[117].

By blocking the classic ACE-Ang II- AT1R axis a well-recognized and effective anti-hypertensive and antiproteinuric treatment is obtained. More recently, a few patients have received activators of the ACE2-Ang-(1-7)-Mas receptor pathway, which can be separated in two main types: (1) those compounds that augment the enzymatic activity of ACE2 and will impact the system by increasing Ang II inactivation of[118] and (2) those molecules that increase Ang-(1-7) production and and are particularly oriented to stimulate the MasR[119]. At this time, in the case of ACE2, little molecules have been developed which activate ACE2[120]. In rats with spontaneous hypertension rats (SHR), a leading ACE2 activator compound (XNT) diminishes BP and does recover ventricular function[121]. The recombinant human ACE2 has been also developed as a different attempt to use the possible therapeutic capabilities of ACE2. At this respect, it has recently been observed that rhACE2 administration attenuates diabetic kidney damage through a mechanism involving both Ang II reduction and Ang-(1-7) increasing signaling[122]. AVE 0991 is the first synthetic compound (non-peptide) developed in order to stimulate the MasR[123]. This molecule is an orally active MasR agonist that imitates the consequences of administering Ang-(1-7) on the kidney, the vessels, and on the heart as well[124,125]. AVE0991 does considerably prevent organ damage in SHR and also in rats with hypertension induced by L-NAME by preserving cardiac contractility, avoiding hypertension, and by reducing urinary protein excretion[123]. Two new designed peptides, CGEN-856 as well as CGEN-857, target the other activator of G protein coupled receptor, and also show high specificity for the Mas receptor[126].

**The Ang-(1-9)-AT2R axis:** The first observations regarding the biological actions of ACE2 and Ang-(1-9) counter-regulating the ACE/Ang II axis were made by Ocaranza *et al*[105] (Figure 4). In MI rats, down regulation of circulating and cardiac ACE2 enzymatic activity is observed in the chronic phase of LV dysfunction and this effect is precluded by enalapril[105]. When rats with MI or with the sham procedure received the ACEI enalapril for 2 mo, Ang-(1-9) levels in plasma were increased significantly but Ang-(1-7) levels were not modified[105]. Thus, by taking into account these observations, it was proposed that Ang-(1-9) rather than Ang-(1-7) acts as a counter-regulator of Ang II in this model of heart failure[105].

Ang (1–9) does regulate cardiac hypertrophy both *in vivo* in addition to *in vitro*[61,63]. In rats with myocardial infarction (MI) that received vehicle, enalapril, or candesartan during 8 wk, Ang (1–9) did prevent myocardial hypertrophy and increased plasma Ang‑(1‑9) circulating levels by several fold[61]. Besides, in those experiments, Ang-(1-9) plasma levels correlated inversely with several markers of cardiac hypertrophy, even by adjusting for reduction of blood pressure[61]. This observed action was very specific, since no correlation was found between cardiac hypertrophy with Ang-(1–7), nor with Ang II neither with the bradykinin levels. In other experiments, chronic treatment with Ang-(1-9) to rats with MI by using osmotic pumps diminished circulating levels of Ang II and enzymatic activity of the ACE and also to prevented myocardial hypertrophy[61]. Since there are available *in vitro* data showing that Ang-(1-9) incubation with ACE does generate Ang-(1-7)[76], and this peptide negatively regulates hypertrophy[127,128], the blocker of the Mas receptor A779 was utilized in order to assess whether Ang-(1-7) could intervene in the actions of Ang-(1-9). Even though A779 did augment blood levels of Ang-(1-7) by almost 3 fold, this specific blocker did not alter the Ang-(1-9) suppression effect on cardiomyocyte (CM) hypertrophy secondary to MI[61]. In experiments using cultured CM incubated with noradrenaline, IGF-1[61] or Ang II[63], Ang-(1-9) prevented hypertrophy of cardiac cells and this action was mediated through the AT2R[63]. In addition, by the same AT2R mediated mechanism, Ang-(1-9) treatment did alleviate streptosotozin (STZ) induced cardiomyopathy dose dependently and did attenuate cardiac dysfunction in rats with diabetes induced by STZ[129].

Recently, it has been described that long term treatment with Ang-(1-9) significantly reduced HT and hypertensive cardiovascular damage in two experimental models: the Ang II infusion model and the Goldblatt model (2K-1C) as well[84]. In these experiments, Ang-(1-9) also blunted the modifications in LV systolic function (ejection fraction) in both hypertensive models, without having an effect in the control rats[84]. Co-administration of Ang-(1-9) together with A779 did not modify the antihypertensive capability of Ang-(1–9) but PD123319, a specific AT2R antagonist, did entirely abolish the favorable effect of Ang-(1-9) on hypertension and on cardiovascular remodeling[84]. In cultured cardiac rat fibroblasts, we hace recently observed that Ang-(1-9) was able to reduce fibroblast proliferation promoted throughout Ang II and also collagen content with no effects on differentiation of fibroblasts onto myofibroblasts[84]. The biological effects of Ang-(1-9) on hypertensive CV remodeling were corroborated in the rat with spontaneous hypertension which is stroke-prone (SHRSP)[130]**.** Those facts demonstrate that activation of the AT2R produced by Ang-(1-9) has a significant myocardial antifibrotic effect that may be associated to a direct effect on cardiac fibroblasts. This preclinical findings suggest a possible clinical approach for cardiovascular complications from hypertension, by stimulating the AT2R using Ang-(1-9) and obtaining in this way antihypertrophic and antifibrotic protective effects.

The AT2R stimulation activates among other mechanisms the NO–cGMP dependent pathway[131]. This happens through direct or indirect effects *via* bradykinins or by augmented activity or expression of endothelial NOS[131]. Additionally, AT2R activation might be able to induce relaxation by inverse regulation of the Rho kinase pathway in the vascular wall[132].

The release of endothelial vasodilators in response to Ang-(1-9) may be a mechanism involved in the beneficial consequences of Ang-(1-9) observed in hypertensive rodents. In *ex vivo* resistance arteries from Ang II treated rats*,* it has been observed that Ang-(1-9) does preserve relaxation induced by Ach (which is dependent from endothelium)[84]. Ang-(1-9) did also augment the concentration of eNOS mRNA in the aortic wall, which is associated to higher plasma concentrations of nitrate. These observed effects of Ang-(1-9) were completely inhibited by using PD123319, which is coherent with the concept that Ang-(1-9) does increase bioavailability of NO through a mechanism mediated by the AT2R[84]. In keeping with these results by blocking Enos a significantly increased contractile response to Phe in aortic rings of SHRSP chronically treated with Ang-(1-9) infusion has been found[130]. Furthermore, Ang-(1-9) does stimulate secretion of ANP without modifying the atrial contractility[83] and this observed effect of Ang-(1-9) is attenuated by using an AT2R antagonist but not when using AT1R nor MasR pharmacologic antagonism. Furthermore, by using inhibitors of phosphatidylinositol 3-kinase (PI3K), nitric oxide synthase, protein kinase B (Akt), or soluble guanylyl cyclase, Ang-(1-9)-induced ANP secretion is blocked. The above-mentioned observations consistently suggest that the Ang-(1-9) peptide does stimulate secretion of ANP throughout the AT2R-PI3K-Akt-NO-cGMP cascade[83]. The release of arachidonic acid - another potent vasodilator - may be also actively implicated here, in addition to the NO[133].

Regulation of Ang-(1-9) by Rho kinase was assessed by Ocaranza and coworkers for first time in hypertensive DOCA-salt rats, by inhibiting Rho-kinase with fasudil[18]. In the above mentioned experimental model, it was noticed that over expression of genes promoting cardiovascular remodeling such as transforming growth factor 1, plasminogen activator inhibitor 1 and the MCP-1 molecule were lower by using the specific Rho kinase inhibitor, whereas both ACE2 enzymatic activity and blood levels of Ang-(1-9) were substantially increased. Remarkably, the changes observed in ACE, ACE2 and in the levels of Ang-(1–9) were clearly observed in both experimental groups receiving fasudil (the sham group and the DOCA group with hypertension)[18]. Thus, this new action of Rho kinase inhibition on ACE2 (gene expression/enzymatic activity) in addition to lowering Ang-(1-9) levels might also conduce to its salutary effects in HT, atherosclerosis disease, and CV remodeling.

AT2R agonists could represent a new class of drugs aimed to preclude and reverse hypertensive CV remodeling. In isolated conductance and resistance vessels, several investigators have shown that CGP42112A (a peptide agonist)[134,135] and more recently the agonist C21[136] induce vasorelaxation, consistent with the concept of AT2R opposing to the AT1R. A recent study assessed AT2R-stimulation with C21 on post-MI cardiac function[137]. Treatment using C21 began 24 hours after MI and it was administered during seven days. The aforementioned treatment with C21 diminished the scar size and this reduction was due to a favorable effect of C21 on myocardial remodeling port MI that also did improve systolic and diastolic cardiac dysfunction. Besides, C21 treatment diminished the content of inflammatory cytokines (IL-1b, IL-2, IL-6) and pro-apoptotic markers (caspase-3, Fas-ligand). Moreover, both the monocyte chemoattractant-protein 1 and levels of myeloperoxidase, a biomarker of oxidative stress, were significantly decreased by C21[137]. Therefore, AT2R stimulation (direct) did improve cardiac function following an experimental MI throughout both anti-inflammatory and anti-oxidant mechanisms and by a more favorable scar remodeling.

Amelioration of inflammation seems to be a fundamental mechanism of action of AT2R-agonism[138]. It has been recently observed that AT2R-stimulation with C21 inhibits NF-kB activity and subsequent synthesis of interleukin 6 and other cytokines that promote inflammation by activating tyrosine-phosphatases, serine/threonine-phosphatases and also CYP2C/2J enzymes leading in this way to increased 11, 12-EET synthesis[91]. 11,12-EET has been shown by Node *et al*[139] in 1999 to have anti-inflammatory characteristics by inhibiting NF-kB. The fact that the described action of C21 on IL-6 promoter activity was similar in strength to the effect of hydrocortisone administered at an equivalent dose indicates that the AT2R could be clinically useful as a beneficial target in cardiovascular disease, and also in inflammatory clinical conditions[140]; an hypothesis which needs to be tested in future experiments.

**Alamandine–MrgD axis:** Alamandine produces vasorelaxation in phenylephrine-contracted aortic rings and when microinjected into central areas critically involved in BP control, such as caudal ventrolateral and rostral ventrolateral medulla. Alamandine produces a decrease vs. increase in BP in the former vs. latter, respectively, revealing that it acts locally and centrally in a rather similar manner with respect to Ang-(1-7) (Figure 4). Additionally, oral alamandine administration produces a longstanding antihypertensive effect in SHR rats[62], suggesting a therapeutic potential for the conditions with underlying cardiovascular remodeling. Furthermore, alamandine has direct effects on remodeling by diminishing cardiac deposition of collagen as well as fibronectin in the rat model of myocardial fibrosis induced by isoproterenol[62]. Since alamandine is a very new molecule within the RAS, currently there are scant data about its participation in disease, although it has been observed elevated plasma alamandine concentrations is in subjects with nephropathy, suggesting that alamandine may be involved in some pathophysiological conditions[62].

The identification of alamandine and its MrgDR contributes to provide novel insights to the knowledgement of the RAS pathophysiology and also opens new possibilities in order to develop therapeutic approaches oriented to prevent/treat CV remodeling, particularly in hypertension.

**POSSIBLE ROLE OF ESTROGENS IN PREVENTING HYPERTENSIVE CARDIAC HYPERTROPHY THROUGH MYOCARDIAL CHYMASE AND ANGIOTENSIN II**

Since cardiovascular disease in females is consistently increasing, it becomes relevant to better know the connections among age, gender, and cardiovascular health status in a more precise way. With the assumption that estrogens do prevent pathological cardiac remodeling secondary to pressure overload throughout inhibition of mast cell chymase release, Li *et al*[141] recently performed thoracic aortic constriction in intact and in ovariectomized rats (an experimental model resembling postmenopausal hypertension). Three days previously to the aortic constriction surgery, ovariectomized rats began to receive 17ß-Estradiol, an inhibitor of the chymase (or a mast cell stabilizer)[141]. Their main findings were that both density and degranulation of the mast cells, circulating chymase levels - able to hydrolyze angiotensin I onto the octapeptide angiotensin II - and cardiac transforming growth factor-1 were augmented due to the constriction procedure in ovariectomized rats and that replacement therapy with estrogens significantly diminished the cardiac levels of increased chymase, both degranulation and density of the mast cells, circulating chymase levels and active transforming growth factor-1 in the myocardium as well[141]. They also observed in this experimental model that estrogens did prevent myocardial hypertrophy and fibrosis[141] (which is rather similar to cardiac failure with normal systolic performance commonly associated to hypertensive diastolic dysfunction)[142]. By using the above mentioned experimental model they concluded that the mast cell derived chymase release, which is able to be inhibited by estrogens, is responsible for myocardial protection in the case of adverse cardiac remodeling resulting from transverse aortic constriction[141].

Novel observations derived from experimental models that emulate the cardiovascular phenotype of women having reduced estrogens levels (such in early ovarian failure or in postmenopause), including the ovariectomized congenic mRen2. Lewis rat and transverse thoracic aortic constriction in ovariectomized rats models provide substantial data in the sense that estrogens actively modulate the tissue renin–angiotensin–aldosterone system, and signaling intracellular pathways related to nitric oxide synthase, in some measure through the G protein–coupled receptor 30 (in the membrane), additionally identified as the G protein–coupled estrogen receptor 1[143] which might also be connected to the pro remodeling Rho A/Rho kinase signaling pathway[142].

Current limitations and challenges: more potent ROCK inhibition is a main challenge at this time even though recent preclinical evidence of newer inhibitors in this regard is available[15,19]. As a consequence, clinical studies with ROCK inhibitors in hypertension will follow. In the field of the vasodilatory peptides from the RAS, Ang 1-9 is effective as antihypertensive and anti-remodeling. However, human data are necessary as well as pharmacodynamic information and means for appropriate delivery. With relationship to the implications of the estrogens-myocardial chymase interaction, from our point of view, more preclinical data are required since the number of studies is small.

**CONCLUSION**

The discussed evidences in this review about the three aforementioned novel mechanisms of hypertensive myocardial remodeling: the Rho kinase intracellular signaling pathway, the vasodilatory peptides from the RAS and the estrogens-myocardial chymase interaction, open new therapeutic opportunities to effectively get better quality of life, reduce/avoid hypertensive cardiovascular remodeling and residual hypertensive risk.

**REFERENCES**

1 **Thomopoulos C**, Parati G, Zanchetti A. Effects of blood pressure-lowering on outcome incidence in hypertension: 5. Head-to-head comparisons of various classes of antihypertensive drugs - overview and meta-analyses. *J Hypertens* 2015; **33**: 1321-1341 [PMID: 26039526 DOI: 10.1097/HJH.0000000000000614]

2 **Jalil JE**, Jalil R, Ocaranza MP. Combined rho kinase and renin-angiotensin system inhibition: a new therapeutic perspective for renal and cardiovascular remodeling. *Hypertens Res* 2010; **33**: 883-885 [PMID: 20664549 DOI: 10.1038/hr.2010.129]

3 **Nakagawa O**, Fujisawa K, Ishizaki T, Saito Y, Nakao K, Narumiya S. ROCK-I and ROCK-II, two isoforms of Rho-associated coiled-coil forming protein serine/threonine kinase in mice. *FEBS Lett* 1996; **392**: 189-193 [PMID: 8772201 DOI: 10.1016/0014-5793(96)00811-3]

4 **Shimokawa H**, Takeshita A. Rho-kinase is an important therapeutic target in cardiovascular medicine. *Arterioscler Thromb Vasc Biol* 2005; **25**: 1767-1775 [PMID: 16002741 DOI: 10.1161/01.ATV.0000176193.83629.c8]

5 **Jalil J**, Lavandero S, Chiong M, Ocaranza MP. [Rho/Rho kinase signal transduction pathway in cardiovascular disease and cardiovascular remodeling]. *Rev Esp Cardiol* 2005; **58**: 951-961 [PMID: 16053829 DOI: 10.1157/13078132]

6 **Shimokawa H**, Rashid M. Development of Rho-kinase inhibitors for cardiovascular medicine. *Trends Pharmacol Sci* 2007; **28**: 296-302 [PMID: 17482681 DOI: 10.1016/j.tips.2007.04.006]

7 **Watanabe H**, Iino K, Ito H. Rho-kinase in leukocytes. An emerging biomarker for heart failure. *Circ J* 2013; **77**: 2471-2472 [PMID: 23912838 DOI: 10.1253/circj.CJ-13-0935]

8 **Shi J**, Wei L. Rho kinases in cardiovascular physiology and pathophysiology: the effect of fasudil. *J Cardiovasc Pharmacol* 2013; **62**: 341-354 [PMID: 23921309 DOI: 10.1097/FJC.0b013e3182a3718f]

9 **Julian L**, Olson MF. Rho-associated coiled-coil containing kinases (ROCK): structure, regulation, and functions. *Small GTPases* 2014; **5**: e29846 [PMID: 25010901 DOI: 10.4161/sgtp.29846]

10 **Carbone ML**, Brégeon J, Devos N, Chadeuf G, Blanchard A, Azizi M, Pacaud P, Jeunemaître X, Loirand G. Angiotensin II activates the RhoA exchange factor Arhgef1 in humans. *Hypertension* 2015; **65**: 1273-1278 [PMID: 25870189 DOI: 10.1161/HYPERTENSIONAHA.114.05065]

11 **Dong M**, Yan BP, Liao JK, Lam YY, Yip GW, Yu CM. Rho-kinase inhibition: a novel therapeutic target for the treatment of cardiovascular diseases. *Drug Discov Today* 2010; **15**: 622-629 [PMID: 20601092 DOI: 10.1016/j.drudis.2010.06.011]

12 **Zhou Q**, Gensch C, Liao JK. Rho-associated coiled-coil-forming kinases (ROCKs): potential targets for the treatment of atherosclerosis and vascular disease. *Trends Pharmacol Sci* 2011; **32**: 167-173 [PMID: 21242007 DOI: 10.1016/j.tips.2010.12.006]

13 **Loirand G**, Pacaud P. Involvement of Rho GTPases and their regulators in the pathogenesis of hypertension. *Small GTPases* 2014; **5**: 1-10 [PMID: 25496262 DOI: 10.4161/sgtp.28846]

14 **Uehata M**, Ishizaki T, Satoh H, Ono T, Kawahara T, Morishita T, Tamakawa H, Yamagami K, Inui J, Maekawa M, Narumiya S. Calcium sensitization of smooth muscle mediated by a Rho-associated protein kinase in hypertension. *Nature* 1997; **389**: 990-994 [PMID: 9353125]

15 **Löhn M**, Plettenburg O, Ivashchenko Y, Kannt A, Hofmeister A, Kadereit D, Schaefer M, Linz W, Kohlmann M, Herbert JM, Janiak P, O'Connor SE, Ruetten H. Pharmacological characterization of SAR407899, a novel rho-kinase inhibitor. *Hypertension* 2009; **54**: 676-683 [PMID: 19597037 DOI: 10.1161/HYPERTENSIONAHA.109.134353]

16 **Sehon CA**, Wang GZ, Viet AQ, Goodman KB, Dowdell SE, Elkins PA, Semus SF, Evans C, Jolivette LJ, Kirkpatrick RB, Dul E, Khandekar SS, Yi T, Wright LL, Smith GK, Behm DJ, Bentley R, Doe CP, Hu E, Lee D. Potent, selective and orally bioavailable dihydropyrimidine inhibitors of Rho kinase (ROCK1) as potential therapeutic agents for cardiovascular diseases. *J Med Chem* 2008; **51**: 6631-6634 [PMID: 18842034 DOI: 10.1021/jm8005096]

17 **Kast R**, Schirok H, Figueroa-Pérez S, Mittendorf J, Gnoth MJ, Apeler H, Lenz J, Franz JK, Knorr A, Hütter J, Lobell M, Zimmermann K, Münter K, Augstein KH, Ehmke H, Stasch JP. Cardiovascular effects of a novel potent and highly selective azaindole-based inhibitor of Rho-kinase. *Br J Pharmacol* 2007; **152**: 1070-1080 [PMID: 17934515]

18 **Ocaranza MP**, Rivera P, Novoa U, Pinto M, González L, Chiong M, Lavandero S, Jalil JE. Rho kinase inhibition activates the homologous angiotensin-converting enzyme-angiotensin-(1-9) axis in experimental hypertension. *J Hypertens* 2011; **29**: 706-715 [PMID: 21330937 DOI: 10.1097/HJH.0b013e3283440665]

19 **Löhn M**, Plettenburg O, Kannt A, Kohlmann M, Hofmeister A, Kadereit D, Monecke P, Schiffer A, Schulte A, Ruetten H, Ivashchenko Y. End-organ protection in hypertension by the novel and selective Rho-kinase inhibitor, SAR407899. *World J Cardiol* 2015; **7**: 31-42 [PMID: 25632317 DOI: 10.4330/wjc.v7.i1.31]

20 **Dhaliwal JS**, Casey DB, Greco AJ, Badejo AM Jr, Gallen TB, Murthy SN, Nossaman BD, Hyman AL, Kadowitz PJ. Rho kinase and Ca2 entry mediate increased pulmonary and systemic vascular resistance in L-NAME-treated rats. *Am J Physiol Lung Cell Mol Physiol* 2007; **293**: L1306-1313 [PMID: 17766587]

21 **Grisk O**. Potential benefits of rho-kinase inhibition in arterial hypertension. *Curr Hypertens Rep* 2013; **15**: 506-513 [PMID: 23852615 DOI: 10.1007/s11906-013-0373-0]

22 **Masumoto A**, Hirooka Y, Shimokawa H, Hironaga K, Setoguchi S, Takeshita A. Possible involvement of Rho-kinase in the pathogenesis of hypertension in humans. *Hypertension* 2001; **38**: 1307-1310 [PMID: 11751708]

23 **Smith CJ**, Santhanam L, Alexander LM. Rho-Kinase activity and cutaneous vasoconstriction is upregulated in essential hypertensive humans. *Microvasc Res* 2013; **87**: 58-64 [PMID: 23481864 DOI: 10.1016/j.mvr.2013.02.005]

24 **Rivera P**, Ocaranza MP, Lavandero S, Jalil JE. Rho kinase activation and gene expression related to vascular remodeling in normotensive rats with high angiotensin I converting enzyme levels. *Hypertension* 2007; **50**: 792-798 [PMID: 17785632 DOI: 10.1161/HYPERTENSIONAHA.107.095117]

25 **Seko T**, Ito M, Kureishi Y, Okamoto R, Moriki N, Onishi K, Isaka N, Hartshorne DJ, Nakano T. Activation of RhoA and inhibition of myosin phosphatase as important components in hypertension in vascular smooth muscle. *Circ Res* 2003; **92**: 411-418 [PMID: 12600888 DOI: 10.1161/01.RES.0000059987.90200.44]

26 **Tsounapi P**, Saito M, Kitatani K, Dimitriadis F, Ohmasa F, Shimizu S, Kinoshita Y, Takenaka A, Satoh K. Fasudil improves the endothelial dysfunction in the aorta of spontaneously hypertensive rats. *Eur J Pharmacol* 2012; **691**: 182-189 [PMID: 22819709 DOI: 10.1016/j.ejphar.2012.07.016]

27 **Hassona MD**, Abouelnaga ZA, Elnakish MT, Awad MM, Alhaj M, Goldschmidt-Clermont PJ, Hassanain H. Vascular hypertrophy-associated hypertension of profilin1 transgenic mouse model leads to functional remodeling of peripheral arteries. *Am J Physiol Heart Circ Physiol* 2010; **298**: H2112-H2120 [PMID: 20400688 DOI: 10.1152/ajpheart.00016.2010]

28 **Chan CK**, Mak JC, Man RY, Vanhoutte PM. Rho kinase inhibitors prevent endothelium-dependent contractions in the rat aorta. *J Pharmacol Exp Ther* 2009; **329**: 820-826 [PMID: 19193928 DOI: 10.1124/jpet.108.148247 DOI: 10.1124/jpet.108.148247]

29 **Takeda K**, Ichiki T, Tokunou T, Iino N, Fujii S, Kitabatake A, Shimokawa H, Takeshita A. Critical role of Rho-kinase and MEK/ERK pathways for angiotensin II-induced plasminogen activator inhibitor type-1 gene expression. *Arterioscler Thromb Vasc Biol* 2001; **21**: 868-873 [PMID: 11348889 DOI: 10.1161/01.ATV.21.5.868]

30 **Rikitake Y**, Liao JK. Rho-kinase mediates hyperglycemia-induced plasminogen activator inhibitor-1 expression in vascular endothelial cells. *Circulation* 2005; **111**: 3261-3268 [PMID: 15956119 DOI: 10.1161/CIRCULATIONAHA.105.534024]

31 **Sun Q**, Yue P, Ying Z, Cardounel AJ, Brook RD, Devlin R, Hwang JS, Zweier JL, Chen LC, Rajagopalan S. Air pollution exposure potentiates hypertension through reactive oxygen species-mediated activation of Rho/ROCK. *Arterioscler Thromb Vasc Biol* 2008; **28**: 1760-1766 [PMID: 18599801 DOI: 10.1161/ATVBAHA.108.166967]

32 **Higashi M**, Shimokawa H, Hattori T, Hiroki J, Mukai Y, Morikawa K, Ichiki T, Takahashi S, Takeshita A. Long-term inhibition of Rho-kinase suppresses angiotensin II-induced cardiovascular hypertrophy in rats in vivo: effect on endothelial NAD(P)H oxidase system. *Circ Res* 2003; **93**: 767-775 [PMID: 14500337]

33 **Satoh K**, Fukumoto Y, Shimokawa H. Rho-kinase: important new therapeutic target in cardiovascular diseases. *Am J Physiol Heart Circ Physiol* 2011; **301**: H287-H296 [PMID: 21622831 DOI: 10.1152/ajpheart.00327.2011]

34 **Ito K**, Hirooka Y, Sakai K, Kishi T, Kaibuchi K, Shimokawa H, Takeshita A. Rho/Rho-kinase pathway in brain stem contributes to blood pressure regulation via sympathetic nervous system: possible involvement in neural mechanisms of hypertension. *Circ Res* 2003; **92**: 1337-1343 [PMID: 12791705]

35 **Ito K**, Hirooka Y, Kishi T, Kimura Y, Kaibuchi K, Shimokawa H, Takeshita A. Rho/Rho-kinase pathway in the brainstem contributes to hypertension caused by chronic nitric oxide synthase inhibition. *Hypertension* 2004; **43**: 156-162 [PMID: 14732730]

36 **Mita S**, Kobayashi N, Yoshida K, Nakano S, Matsuoka H. Cardioprotective mechanisms of Rho-kinase inhibition associated with eNOS and oxidative stress-LOX-1 pathway in Dahl salt-sensitive hypertensive rats. *J Hypertens* 2005; **23**: 87-96 [PMID: 15643129]

37 **Ishimaru K**, Ueno H, Kagitani S, Takabayashi D, Takata M, Inoue H. Fasudil attenuates myocardial fibrosis in association with inhibition of monocyte/macrophage infiltration in the heart of DOCA/salt hypertensive rats. *J Cardiovasc Pharmacol* 2007; **50**: 187-194 [PMID: 17703135]

38 **Fukui S**, Fukumoto Y, Suzuki J, Saji K, Nawata J, Tawara S, Shinozaki T, Kagaya Y, Shimokawa H. Long-term inhibition of Rho-kinase ameliorates diastolic heart failure in hypertensive rats. *J Cardiovasc Pharmacol* 2008; **51**: 317-326 [PMID: 18356698 DOI: 10.1097/FJC.0b013e31816533b7]

39 **Phrommintikul A**, Tran L, Kompa A, Wang B, Adrahtas A, Cantwell D, Kelly DJ, Krum H. Effects of a Rho kinase inhibitor on pressure overload induced cardiac hypertrophy and associated diastolic dysfunction. *Am J Physiol Heart Circ Physiol* 2008; **294**: H1804-H1814 [PMID: 18245565 DOI: 10.1152/ajpheart.01078.2007]

40 **Shi J**, Zhang YW, Summers LJ, Dorn GW, Wei L. Disruption of ROCK1 gene attenuates cardiac dilation and improves contractile function in pathological cardiac hypertrophy. *J Mol Cell Cardiol* 2008; **44**: 551-560 [PMID: 18178218 DOI: 10.1016/j.yjmcc.2007.11.018]

41 **Del Re DP**, Miyamoto S, Brown JH. RhoA/Rho kinase up-regulate Bax to activate a mitochondrial death pathway and induce cardiomyocyte apoptosis. *J Biol Chem* 2007; **282**: 8069-8078 [PMID: 17234627]

42 **Mizutani H**, Okamoto R, Moriki N, Konishi K, Taniguchi M, Fujita S, Dohi K, Onishi K, Suzuki N, Satoh S, Makino N, Itoh T, Hartshorne DJ, Ito M. Overexpression of myosin phosphatase reduces Ca(2+) sensitivity of contraction and impairs cardiac function. *Circ J* 2010; **74**: 120-128 [PMID: 19966500]

43 **Liu PY**, Chen JH, Lin LJ, Liao JK. Increased Rho kinase activity in a Taiwanese population with metabolic syndrome. *J Am Coll Cardiol* 2007; **49**: 1619-1624 [PMID: 17433952]

44 **Li CB**, Li XX, Chen YG, Gao HQ, Bao CM, Liu XQ, Bu PL, Zhang J, Zhang Y, Ji XP. Myocardial remodeling in rats with metabolic syndrome: role of Rho-kinase mediated insulin resistance. *Acta Biochim Pol* 2012; **59**: 249-254 [PMID: 22693684]

45 **Hidaka T**, Hata T, Soga J, Fujii Y, Idei N, Fujimura N, Kihara Y, Noma K, Liao JK, Higashi Y. Increased leukocyte rho kinase (ROCK) activity and endothelial dysfunction in cigarette smokers. *Hypertens Res* 2010; **33**: 354-359 [PMID: 20139919 DOI: 10.1038/hr.2010.3]

46 **Hata T**, Goto C, Soga J, Hidaka T, Fujii Y, Idei N, Fujimura N, Maruhashi T, Mikami S, Kihara Y, Chayama K, Noma K, Liao JK, Higashi Y. Measurement of Rho-associated kinase (ROCK) activity in humans: validity of leukocyte p-MBS/t-MBS in comparison with vascular response to fasudil. *Atherosclerosis* 2011; **214**: 117-121 [PMID: 21035804 DOI: 10.1016/j.atherosclerosis.2010.10.005]

47 **Gabrielli L**, Winter JL, Godoy I, McNab P, Padilla I, Cordova S, Rigotti P, Novoa U, Mora I, García L, Ocaranza MP, Jalil JE. Increased rho-kinase activity in hypertensive patients with left ventricular hypertrophy. *Am J Hypertens* 2014; **27**: 838-845 [PMID: 24363278 DOI: 10.1093/ajh/hpt234]

48 **Gabrielli L**, Berkovitz A, Mora I, Novoa U, Godoy I, MacNab P, Córdova S, Padilla I, Rigotti P, García L, Lavandero S, Ocaranza MP, Jalil J. Increased Rho-kinase activity in peripheral leukocytes is associated to oxidative stress and decreased compliance of the arterial wall in diabetic hypertensive patients. *Rev Chil Cardiol* 2010; **30**: 34-41

49 **Ocaranza MP**, Gabrielli L, Mora I, Garcia L, McNab P, Godoy I, Braun S, Córdova S, Castro P, Novoa U, Chiong M, Lavandero S, Jalil JE. Markedly increased Rho-kinase activity in circulating leukocytes in patients with chronic heart failure. *Am Heart J* 2011; **161**: 931-937 [PMID: 21570525]

50 **Kajikawa M**, Noma K, Maruhashi T, Mikami S, Iwamoto Y, Iwamoto A, Matsumoto T, Hidaka T, Kihara Y, Chayama K, Nakashima A, Goto C, Liao JK, Higashi Y. Rho-associated kinase activity is a predictor of cardiovascular outcomes. *Hypertension* 2014; **63**: 856-864 [PMID: 24379190 DOI: 0.1161/HYPERTENSIONAHA.113.02296]

51 **Hata T**, Soga J, Hidaka T, Idei N, Fujii Y, Fujimura N, Mikami S, Maruhashi T, Kihara Y, Chayama K, Kato H, Noma K, Liao JK, Higashi Y. Calcium channel blocker and Rho-associated kinase activity in patients with hypertension. *J Hypertens* 2011; **29**: 373-379 [PMID: 21063203 DOI: 10.1097/HJH.0b013e328340902d]

52 **Fujimura N**, Noma K, Hata T, Soga J, Hidaka T, Idei N, Fujii Y, Mikami S, Maruhashi T, Iwamoto Y, Kihara Y, Chayama K, Kato H, Liao JK, Higashi Y. Mineralocorticoid receptor blocker eplerenone improves endothelial function and inhibits Rho-associated kinase activity in patients with hypertension. *Clin Pharmacol Ther* 2012; **91**: 289-297 [PMID: 22205191 DOI: 10.1038/clpt.2011.227]

53 **Guo R**, Su Y, Yan J, Sun H, Wu J, Liu W, Xu Y. Fasudil improves short-term echocardiographic parameters of diastolic function in patients with type 2 diabetes with preserved left ventricular ejection fraction: a pilot study. *Heart Vessels* 2015; **30**: 89-97 [PMID: 24390764 DOI: 10.1007/s00380-013-0458-3]

54 **Dong M**, Liao JK, Fang F, Lee AP, Yan BP, Liu M, Yu CM. Increased Rho kinase activity in congestive heart failure. *Eur J Heart Fail* 2012; **14**: 965-973 [PMID: 22588320 DOI: 10.1093/eurjhf/hfs068]

55 **Dong M**, Liao JK, Yan B, Li R, Zhang M, Yu CM. A combination of increased Rho kinase activity and N-terminal pro-B-type natriuretic peptide predicts worse cardiovascular outcome in patients with acute coronary syndrome. *Int J Cardiol* 2013; **167**: 2813-2819 [PMID: 22921817 DOI: 10.1016/j.ijcard.2012.07.007]

56 **Intengan HD**, Schiffrin EL. Structure and mechanical properties of resistance arteries in hypertension: role of adhesion molecules and extracellular matrix determinants. *Hypertension* 2000; **36**: 312-318 [PMID: 10988257 DOI: 10.1161/01.HYP.36.3.312]

57 **Unger T**. The angiotensin type 2 receptor: variations on an enigmatic theme. *J Hypertens* 1999; **17**: 1775-1786 [PMID: 10703869 DOI: 10.1097/00004872-199917121-00001]

58 **Ferrario CM**. Role of angiotensin II in cardiovascular disease therapeutic implications of more than a century of research. *J Renin Angiotensin Aldosterone Syst* 2006; **7**: 3-14 [PMID: 17083068 DOI: 10.3317/jraas.2006.003]

59 **de Gasparo M**, Catt KJ, Inagami T, Wright JW, Unger T. International union of pharmacology. XXIII. The angiotensin II receptors. *Pharmacol Rev* 2000; **52**: 415-472 [PMID: 10977869]

60 **Santos RA**, Simoes e Silva AC, Maric C, Silva DM, Machado RP, de Buhr I, Heringer-Walther S, Pinheiro SV, Lopes MT, Bader M, Mendes EP, Lemos VS, Campagnole-Santos MJ, Schultheiss HP, Speth R, Walther T. Angiotensin-(1-7) is an endogenous ligand for the G protein-coupled receptor Mas. *Proc Natl Acad Sci USA* 2003; **100**: 8258-8263 [PMID: 12829792 DOI: 10.1073/pnas.1432869100]

61 **Ocaranza MP**, Lavandero S, Jalil JE, Moya J, Pinto M, Novoa U, Apablaza F, Gonzalez L, Hernandez C, Varas M, Lopez R, Godoy I, Verdejo H, Chiong M. Angiotensin-(1-9) regulates cardiac hypertrophy in vivo and in vitro. *J Hypertens* 2010; **28**: 1054-1064 [PMID: 20411619 DOI: 10.1097/HJH.0b013e328335d291]

62 **Lautner RQ**, Villela DC, Fraga-Silva RA, Silva N, Verano-Braga T, Costa-Fraga F, Jankowski J, Jankowski V, Sousa F, Alzamora A, Soares E, Barbosa C, Kjeldsen F, Oliveira A, Braga J, Savergnini S, Maia G, Peluso AB, Passos-Silva D, Ferreira A, Alves F, Martins A, Raizada M, Paula R, Motta-Santos D, Klempin F, Pimenta A, Alenina N, Sinisterra R, Bader M, Campagnole-Santos MJ, Santos RA. Discovery and characterization of alamandine: a novel component of the renin-angiotensin system. *Circ Res* 2013; **112**: 1104-1111 [PMID: 23446738 DOI: 10.1161/CIRCRESAHA.113.301077]

63 **Flores-Muñoz M**, Smith NJ, Haggerty C, Milligan G, Nicklin SA. Angiotensin1-9 antagonises pro-hypertrophic signalling in cardiomyocytes via the angiotensin type 2 receptor. *J Physiol* 2011; **589**: 939-951 [PMID: 21173078 DOI: 10.1113/jphysiol.2010.203075]

64 **Welches WR**, Santos RA, Chappell MC, Brosnihan KB, Greene LJ, Ferrario CM. Evidence that prolyl endopeptidase participates in the processing of brain angiotensin. *J Hypertens* 1991; **9**: 631-638 [PMID: 1653799 DOI: 10.1097/00004872-199107000-00008]

65 **Chappell MC**, Pirro NT, Sykes A, Ferrario CM. Metabolism of angiotensin-(1-7) by angiotensin-converting enzyme. *Hypertension* 1998; **31**: 362-367 [PMID: 9453329 DOI: 10.1161/01.HYP.31.1.362]

66 **Welches WR**, Brosnihan KB, Ferrario CM. A comparison of the properties and enzymatic activities of three angiotensin processing enzymes: angiotensin converting enzyme, prolyl endopeptidase and neutral endopeptidase 24.11. *Life Sci* 1993; **52**: 1461-1480 [PMID: 8387132 DOI: 10.1016/0024-3205(93)90108-F]

67 **Ferrario CM**, Chappell MC. A new myocardial conversion of angiotensin I. *Curr Opin Cardiol* 1994; **9**: 520-526 [PMID: 7987030 DOI: 10.1097/00001573-199409000-00004]

68 **Tipnis SR**, Hooper NM, Hyde R, Karran E, Christie G, Turner AJ. A human homolog of angiotensin-converting enzyme. Cloning and functional expression as a captopril-insensitive carboxypeptidase. *J Biol Chem* 2000; **275**: 33238-33243 [PMID: 10924499 DOI: 10.1074/jbc.M002615200]

69 **Ferreira AJ**, Santos RA. Cardiovascular actions of angiotensin-(1-7). *Braz J Med Biol Res* 2005; **38**: 499-507 [PMID: 15962175 DOI: 10.1590/S0100-879X2005000400003]

70 **Santos RA**, Ferreira AJ, Simões E Silva AC. Recent advances in the angiotensin-converting enzyme 2-angiotensin(1-7)-Mas axis. *Exp Physiol* 2008; **93**: 519-527 [PMID: 18310257 DOI: 10.1113/expphysiol.2008.042002]

71 **Giani JF**, Mayer MA, Muñoz MC, Silberman EA, Höcht C, Taira CA, Gironacci MM, Turyn D, Dominici FP. Chronic infusion of angiotensin-(1-7) improves insulin resistance and hypertension induced by a high-fructose diet in rats. *Am J Physiol Endocrinol Metab* 2009; **296**: E262-E271 [PMID: 19001546 DOI: 10.1152/ajpendo.90678.2008]

72 **Oparil S**, Sanders CA, Haber E. In-vivo and in-vitro conversion of angiotensin I to angiotensin II in dog blood. *Circ Res* 1970; **26**: 591-599 [PMID: 4315298 DOI: 10.1161/01.RES.26.5.591]

73 **Oparil S**, Tregear GW, Koerner T, Barnes BA, Haber E. Mechanism of pulmonary conversion of angiotensin I to angiotensin II in the dog. *Circ Res* 1971; **29**: 682-690 [PMID: 4331484]

74 **Jackman HL**, Massad MG, Sekosan M, Tan F, Brovkovych V, Marcic BM, Erdös EG. Angiotensin 1-9 and 1-7 release in human heart: role of cathepsin A. *Hypertension* 2002; **39**: 976-981 [PMID: 12019279]

75 **Kokkonen JO**, Saarinen J, Kovanen PT. Regulation of local angiotensin II formation in the human heart in the presence of interstitial fluid. Inhibition of chymase by protease inhibitors of interstitial fluid and of angiotensin-converting enzyme by Ang-(1-9) formed by heart carboxypeptidase A-like activity. *Circulation* 1997; **95**: 1455-1463 [PMID: 9118513]

76 **Donoghue M**, Hsieh F, Baronas E, Godbout K, Gosselin M, Stagliano N, Donovan M, Woolf B, Robison K, Jeyaseelan R, Breitbart RE, Acton S. A novel angiotensin-converting enzyme-related carboxypeptidase (ACE2) converts angiotensin I to angiotensin 1-9. *Circ Res* 2000; **87**: E1-E9 [PMID: 10969042]

77 **Rice GI**, Thomas DA, Grant PJ, Turner AJ, Hooper NM. Evaluation of angiotensin-converting enzyme (ACE), its homologue ACE2 and neprilysin in angiotensin peptide metabolism. *Biochem J* 2004; **383**: 45-51 [PMID: 15283675]

78 **Garabelli PJ**, Modrall JG, Penninger JM, Ferrario CM, Chappell MC. Distinct roles for angiotensin-converting enzyme 2 and carboxypeptidase A in the processing of angiotensins within the murine heart. *Exp Physiol* 2008; **93**: 613-621 [PMID: 18356559]

79 **Keidar S**, Kaplan M, Gamliel-Lazarovich A. ACE2 of the heart: From angiotensin I to angiotensin (1-7). *Cardiovasc Res* 2007; **73**: 463-469 [PMID: 17049503]

80 **Trask AJ**, Averill DB, Ganten D, Chappell MC, Ferrario CM. Primary role of angiotensin-converting enzyme-2 in cardiac production of angiotensin-(1-7) in transgenic Ren-2 hypertensive rats. *Am J Physiol Heart Circ Physiol* 2007; **292**: H3019-H3024 [PMID: 17308000]

81 **Schwacke JH**, Spainhour JC, Ierardi JL, Chaves JM, Arthur JM, Janech MG, Velez JC. Network modeling reveals steps in angiotensin peptide processing. *Hypertension* 2013; **61**: 690-700 [PMID: 23283355]

82 **Drummer OH**, Kourtis S, Johnson H. Effect of chronic enalapril treatment on enzymes responsible for the catabolism of angiotensin I and formation of angiotensin II. *Biochem Pharmacol* 1990; **39**: 513-518 [PMID: 2407244]

83 **Cha SA**, Park BM, Gao S, Kim SH. Stimulation of ANP by angiotensin-(1-9) via the angiotensin type 2 receptor. *Life Sci* 2013; **93**: 934-940 [PMID: 24177599]

84 **Ocaranza MP**, Moya J, Barrientos V, Alzamora R, Hevia D, Morales C, Pinto M, Escudero N, García L, Novoa U, Ayala P, Díaz-Araya G, Godoy I, Chiong M, Lavandero S, Jalil JE, Michea L. Angiotensin-(1-9) reverses experimental hypertension and cardiovascular damage by inhibition of the angiotensin converting enzyme/Ang II axis. *J Hypertens* 2014; **32**: 771-783 [PMID: 24463937]

85 **Paula RD**, Lima CV, Britto RR, Campagnole-Santos MJ, Khosla MC, Santos RA. Potentiation of the hypotensive effect of bradykinin by angiotensin-(1-7)-related peptides. *Peptides* 1999; **20**: 493-500 [PMID: 10458520]

86 **Etelvino GM**, Peluso AA, Santos RA. New components of the renin-angiotensin system: alamandine and the MAS-related G protein-coupled receptor D. *Curr Hypertens Rep* 2014; **16**: 433 [PMID: 24760442]

87 **Bedecs K**, Elbaz N, Sutren M, Masson M, Susini C, Strosberg AD, Nahmias C. Angiotensin II type 2 receptors mediate inhibition of mitogen-activated protein kinase cascade and functional activation of SHP-1 tyrosine phosphatase. *Biochem J* 1997; **325 ( Pt 2)**: 449-454 [PMID: 9230127]

88 **Gava E**, Samad-Zadeh A, Zimpelmann J, Bahramifarid N, Kitten GT, Santos RA, Touyz RM, Burns KD. Angiotensin-(1-7) activates a tyrosine phosphatase and inhibits glucose-induced signalling in proximal tubular cells. *Nephrol Dial Transplant* 2009; **24**: 1766-1773 [PMID: 19144997]

89 **Nouet S**, Nahmias C. Signal transduction from the angiotensin II AT2 receptor. *Trends Endocrinol Metab* 2007; **11**: 1-6 [PMID: 10652498]

90 **Sampaio WO**, Henrique de Castro C, Santos RA, Schiffrin EL, Touyz RM. Angiotensin-(1-7) counterregulates angiotensin II signaling in human endothelial cells. *Hypertension* 2007; **50**: 1093-1098 [PMID: 17984366]

91 **Rompe F**, Artuc M, Hallberg A, Alterman M, Ströder K, Thöne-Reineke C, Reichenbach A, Schacherl J, Dahlöf B, Bader M, Alenina N, Schwaninger M, Zuberbier T, Funke-Kaiser H, Schmidt C, Schunck WH, Unger T, Steckelings UM. Direct angiotensin II type 2 receptor stimulation acts anti-inflammatory through epoxyeicosatrienoic acid and inhibition of nuclear factor kappaB. *Hypertension* 2010; **55**: 924-931 [PMID: 20157051]

92 **El-Hashim AZ**, Renno WM, Raghupathy R, Abduo HT, Akhtar S, Benter IF. Angiotensin-(1-7) inhibits allergic inflammation, via the MAS1 receptor, through suppression of ERK1/2- and NF-κB-dependent pathways. *Br J Pharmacol* 2012; **166**: 1964-1976 [PMID: 22339213]

93 **Gohlke P**, Pees C, Unger T. AT2 receptor stimulation increases aortic cyclic GMP in SHRSP by a kinin-dependent mechanism. *Hypertension* 1998; **31**: 349-355 [PMID: 9453327]

94 **Heitsch H**, Brovkovych S, Malinski T, Wiemer G. Angiotensin-(1-7)-Stimulated Nitric Oxide and Superoxide Release From Endothelial Cells. *Hypertension* 2001; **37**: 72-76 [PMID: 11208759]

95 **Stegbauer J**, Potthoff SA, Quack I, Mergia E, Clasen T, Friedrich S, Vonend O, Woznowski M, Königshausen E, Sellin L, Rump LC. Chronic treatment with angiotensin-(1-7) improves renal endothelial dysfunction in apolipoproteinE-deficient mice. *Br J Pharmacol* 2011; **163**: 974-983 [PMID: 21371005]

96 **AbdAlla S**, Lother H, Abdel-tawab AM, Quitterer U. The angiotensin II AT2 receptor is an AT1 receptor antagonist. *J Biol Chem* 2001; **276**: 39721-39726 [PMID: 11507095]

97 **Kostenis E**, Milligan G, Christopoulos A, Sanchez-Ferrer CF, Heringer-Walther S, Sexton PM, Gembardt F, Kellett E, Martini L, Vanderheyden P, Schultheiss HP, Walther T. G-protein-coupled receptor Mas is a physiological antagonist of the angiotensin II type 1 receptor. *Circulation* 2005; **111**: 1806-1813 [PMID: 15809376]

98 **Milasta S**, Pediani J, Appelbe S, Trim S, Wyatt M, Cox P, Fidock M, Milligan G. Interactions between the Mas-related receptors MrgD and MrgE alter signalling and trafficking of MrgD. *Mol Pharmacol* 2006; **69**: 479-491 [PMID: 16282220]

99 **Shinohara T**, Harada M, Ogi K, Maruyama M, Fujii R, Tanaka H, Fukusumi S, Komatsu H, Hosoya M, Noguchi Y, Watanabe T, Moriya T, Itoh Y, Hinuma S. Identification of a G protein-coupled receptor specifically responsive to beta-alanine. *J Biol Chem* 2004; **279**: 23559-23564 [PMID: 15037633]

100 **Crozier RA**, Ajit SK, Kaftan EJ, Pausch MH. MrgD activation inhibits KCNQ/M-currents and contributes to enhanced neuronal excitability. *J Neurosci* 2007; **27**: 4492-4496 [PMID: 17442834]

101 **Dong X**, Han S, Zylka MJ, Simon MI, Anderson DJ. A diverse family of GPCRs expressed in specific subsets of nociceptive sensory neurons. *Cell* 2001; **106**: 619-632 [PMID: 11551509]

102 **Rau KK**, McIlwrath SL, Wang H, Lawson JJ, Jankowski MP, Zylka MJ, Anderson DJ, Koerber HR. Mrgprd enhances excitability in specific populations of cutaneous murine polymodal nociceptors. *J Neurosci* 2009; **29**: 8612-8619 [PMID: 19571152]

103 **Nishimura S**, Uno M, Kaneta Y, Fukuchi K, Nishigohri H, Hasegawa J, Komori H, Takeda S, Enomoto K, Nara F, Agatsuma T. MRGD, a MAS-related G-protein coupled receptor, promotes tumorigenisis and is highly expressed in lung cancer. *PLoS One* 2012; **7**: e38618 [PMID: 22715397]

104 **Gembardt F**, Grajewski S, Vahl M, Schultheiss HP, Walther T. Angiotensin metabolites can stimulate receptors of the Mas-related genes family. *Mol Cell Biochem* 2008; **319**: 115-123 [PMID: 18636314]

105 **Ocaranza MP**, Godoy I, Jalil JE, Varas M, Collantes P, Pinto M, Roman M, Ramirez C, Copaja M, Diaz-Araya G, Castro P, Lavandero S. Enalapril attenuates downregulation of Angiotensin-converting enzyme 2 in the late phase of ventricular dysfunction in myocardial infarcted rat. *Hypertension* 2006; **48**: 572-578 [PMID: 16908757]

106 **Luque M**, Martin P, Martell N, Fernandez C, Brosnihan KB, Ferrario CM. Effects of captopril related to increased levels of prostacyclin and angiotensin-(1-7) in essential hypertension. *J Hypertens* 1996; **14**: 799-805 [PMID: 8793704]

107 **Jennings DL**, Kalus JS, Coleman CI, Manierski C, Yee J. Combination therapy with an ACE inhibitor and an angiotensin receptor blocker for diabetic nephropathy: a meta-analysis. *Diabet Med* 2007; **24**: 486-493 [PMID: 17367311]

108 **Tikellis C**, Bialkowski K, Pete J, Sheehy K, Su Q, Johnston C, Cooper ME, Thomas MC. ACE2 deficiency modifies renoprotection afforded by ACE inhibition in experimental diabetes. *Diabetes* 2008; **57**: 1018-1025 [PMID: 18235039]

109 **Igase M**, Kohara K, Nagai T, Miki T, Ferrario CM. Increased expression of angiotensin converting enzyme 2 in conjunction with reduction of neointima by angiotensin II type 1 receptor blockade. *Hypertens Res* 2008; **31**: 553-559 [PMID: 18497476]

110 **Bosnyak S**, Jones ES, Christopoulos A, Aguilar MI, Thomas WG, Widdop RE. Relative affinity of angiotensin peptides and novel ligands at AT1 and AT2 receptors. *Clin Sci (Lond)* 2011; **121**: 297-303 [PMID: 21542804]

111 **Oliveri C**, Ocaranza MP, Campos X, Lavandero S, Jalil JE. Angiotensin I-converting enzyme modulates neutral endopeptidase activity in the rat. *Hypertension* 2001; **38**: 650-654 [PMID: 11566949]

112 **Ocaranza MP**, Palomera C, Román M, Bargetto J, Lavandero S, Jalil JE. Effect of hypertension on angiotensin-(1-7) levels in rats with different angiotensin-I converting enzyme polymorphism. *Life Sci* 2006; **78**: 1535-1542 [PMID: 16229862]

113 **Ocaranza MP**, Piddo AM, Faúndez P, Lavandero S, Jalil JE. Angiotensin I-converting enzyme gene polymorphism influences chronic hypertensive response in the rat Goldblatt model. *J Hypertens* 2002; **20**: 413-420 [PMID: 11875308]

114 **Ocaranza MP**, Díaz-Araya G, Carreño JE, Muñoz D, Riveros JP, Jalil JE, Lavandero S. Polymorphism in gene coding for ACE determines different development of myocardial fibrosis in rats. *Am J Physiol Heart Circ Physiol* 2004; **286**: H498-H506 [PMID: 14527934]

115 **Jalil JE**, Pérez A, Ocaranza MP, Bargetto J, Galaz A, Lavandero S. Increased aortic NADPH oxidase activity in rats with genetically high angiotensin-converting enzyme levels. *Hypertension* 2005; **46**: 1362-1367 [PMID: 16230508]

116 **Jalil JE**, Palomera C, Ocaranza MP, Godoy I, Román M, Chiong M, Lavandero S. Levels of plasma angiotensin-(1-7) in patients with hypertension who have the angiotensin-I-converting enzyme deletion/deletion genotype. *Am J Cardiol* 2003; **92**: 749-751 [PMID: 12972127]

117 **Jalil JE**, Ocaranza MP, Oliveri C, Córdova S, Godoy I, Chamorro G, Braun S, Fardella C, Michel JB, Lavandero S. Neutral endopeptidase and angiotensin I converting enzyme insertion/deletion gene polymorphism in humans. *J Hum Hypertens* 2004; **18**: 119-125 [PMID: 14730327]

118 **Ferrario CM**. ACE2: more of Ang-(1-7) or less Ang II? *Curr Opin Nephrol Hypertens* 2011; **20**: 1-6 [PMID: 21045683]

119 **Grace JA**, Klein S, Herath CB, Granzow M, Schierwagen R, Masing N, Walther T, Sauerbruch T, Burrell LM, Angus PW, Trebicka J. Activation of the MAS receptor by angiotensin-(1-7) in the renin-angiotensin system mediates mesenteric vasodilatation in cirrhosis. *Gastroenterology* 2013; **145**: 874-884.e5 [PMID: 23796456]

120 **Hernández Prada JA**, Ferreira AJ, Katovich MJ, Shenoy V, Qi Y, Santos RA, Castellano RK, Lampkins AJ, Gubala V, Ostrov DA, Raizada MK. Structure-based identification of small-molecule angiotensin-converting enzyme 2 activators as novel antihypertensive agents. *Hypertension* 2008; **51**: 1312-1317 [PMID: 18391097]

121 **Ferreira AJ**, Shenoy V, Qi Y, Fraga-Silva RA, Santos RA, Katovich MJ, Raizada MK. Angiotensin-converting enzyme 2 activation protects against hypertension-induced cardiac fibrosis involving extracellular signal-regulated kinases. *Exp Physiol* 2011; **96**: 287-294 [PMID: 21148624]

122 **Oudit GY**, Liu GC, Zhong J, Basu R, Chow FL, Zhou J, Loibner H, Janzek E, Schuster M, Penninger JM, Herzenberg AM, Kassiri Z, Scholey JW. Human recombinant ACE2 reduces the progression of diabetic nephropathy. *Diabetes* 2010; **59**: 529-538 [PMID: 19934006]

123 **Wiemer G**, Dobrucki LW, Louka FR, Malinski T, Heitsch H. AVE 0991, a nonpeptide mimic of the effects of angiotensin-(1-7) on the endothelium. *Hypertension* 2002; **40**: 847-852 [PMID: 12468568]

124 **Santos RA**, Frézard F, Ferreira AJ. Angiotensin-(1-7): blood, heart, and blood vessels. *Curr Med Chem Cardiovasc Hematol Agents* 2005; **3**: 383-391 [PMID: 16250869]

125 **Pinheiro SV**, Simões e Silva AC, Sampaio WO, de Paula RD, Mendes EP, Bontempo ED, Pesquero JB, Walther T, Alenina N, Bader M, Bleich M, Santos RA. Nonpeptide AVE 0991 is an angiotensin-(1-7) receptor Mas agonist in the mouse kidney. *Hypertension* 2004; **44**: 490-496 [PMID: 15326087]

126 **Shemesh R**, Toporik A, Levine Z, Hecht I, Rotman G, Wool A, Dahary D, Gofer E, Kliger Y, Soffer MA, Rosenberg A, Eshel D, Cohen Y. Discovery and validation of novel peptide agonists for G-protein-coupled receptors. *J Biol Chem* 2008; **283**: 34643-34649 [PMID: 18854305]

127 **Tallant EA**, Ferrario CM, Gallagher PE. Angiotensin-(1-7) inhibits growth of cardiac myocytes through activation of the mas receptor. *Am J Physiol Heart Circ Physiol* 2005; **289**: H1560-H1566 [PMID: 15951342]

128 **Grobe JL**, Mecca AP, Lingis M, Shenoy V, Bolton TA, Machado JM, Speth RC, Raizada MK, Katovich MJ. Prevention of angiotensin II-induced cardiac remodeling by angiotensin-(1-7). *Am J Physiol Heart Circ Physiol* 2007; **292**: H736-H742 [PMID: 17098828]

129 **Zheng H**, Pu SY, Fan XF, Li XS, Zhang Y, Yuan J, Zhang YF, Yang JL. Treatment with angiotensin-(1-9) alleviates the cardiomyopathy in streptozotocin-induced diabetic rats. *Biochem Pharmacol* 2015; **95**: 38-45 [PMID: 25801006]

130 **Flores-Munoz M**, Work LM, Douglas K, Denby L, Dominiczak AF, Graham D, Nicklin SA. Angiotensin-(1-9) attenuates cardiac fibrosis in the stroke-prone spontaneously hypertensive rat via the angiotensin type 2 receptor. *Hypertension* 2012; **59**: 300-307 [PMID: 22184331]

131 **Abadir PM**, Carey RM, Siragy HM. Angiotensin AT2 receptors directly stimulate renal nitric oxide in bradykinin B2-receptor-null mice. *Hypertension* 2003; **42**: 600-604 [PMID: 12953015]

132 **Savoia C**, Tabet F, Yao G, Schiffrin EL, Touyz RM. Negative regulation of RhoA/Rho kinase by angiotensin II type 2 receptor in vascular smooth muscle cells: role in angiotensin II-induced vasodilation in stroke-prone spontaneously hypertensive rats. *J Hypertens* 2005; **23**: 1037-1045 [PMID: 15834290]

133 **Chen Z**, Tan F, Erdös EG, Deddish PA. Hydrolysis of angiotensin peptides by human angiotensin I-converting enzyme and the resensitization of B2 kinin receptors. *Hypertension* 2005; **46**: 1368-1373 [PMID: 16246972]

134 **Henrion D**, Kubis N, Lévy BI. Physiological and pathophysiological functions of the AT(2) subtype receptor of angiotensin II: from large arteries to the microcirculation. *Hypertension* 2001; **38**: 1150-1157 [PMID: 11711513]

135 **Widdop RE**, Matrougui K, Levy BI, Henrion D. AT2 receptor-mediated relaxation is preserved after long-term AT1 receptor blockade. *Hypertension* 2002; **40**: 516-520 [PMID: 12364356]

136 **Bosnyak S**, Welungoda IK, Hallberg A, Alterman M, Widdop RE, Jones ES. Stimulation of angiotensin AT2 receptors by the non-peptide agonist, Compound 21, evokes vasodepressor effects in conscious spontaneously hypertensive rats. *Br J Pharmacol* 2010; **159**: 709-716 [PMID: 20128808]

137 **Kaschina E**, Grzesiak A, Li J, Foryst-Ludwig A, Timm M, Rompe F, Sommerfeld M, Kemnitz UR, Curato C, Namsolleck P, Tschöpe C, Hallberg A, Alterman M, Hucko T, Paetsch I, Dietrich T, Schnackenburg B, Graf K, Dahlöf B, Kintscher U, Unger T, Steckelings UM. Angiotensin II type 2 receptor stimulation: a novel option of therapeutic interference with the renin-angiotensin system in myocardial infarction? *Circulation* 2008; **118**: 2523-2532 [PMID: 19029468]

138 **Rompe F**, Unger T, Steckelings UM. The angiotensin AT2 receptor in inflammation. *Drug News Perspect* 2010; **23**: 104-111 [PMID: 20369075]

139 **Node K**, Huo Y, Ruan X, Yang B, Spiecker M, Ley K, Zeldin DC, Liao JK. Anti-inflammatory properties of cytochrome P450 epoxygenase-derived eicosanoids. *Science* 1999; **285**: 1276-1279 [PMID: 10455056]

140 **Walters PE**, Gaspari TA, Widdop RE. Angiotensin-(1-7) acts as a vasodepressor agent via angiotensin II type 2 receptors in conscious rats. *Hypertension* 2005; **45**: 960-966 [PMID: 15767466]

141 **Li J**, Jubair S, Janicki JS. Estrogen inhibits mast cell chymase release to prevent pressure overload-induced adverse cardiac remodeling. *Hypertension* 2015; **65**: 328-334 [PMID: 25403608 DOI: 10.1161/HYPERTENSIONAHA.114.04238]

142 **Jalil JE**, Ocaranza MP. Estrogens and myocardial chymase: new insights into pathological hypertrophy and remodeling. *Hypertension* 2015; **65**: 271-272 [PMID: 25403603 DOI: 10.1161/HYPERTENSIONAHA.114.04375]

143 **Zhao Z**, Wang H, Jessup JA, Lindsey SH, Chappell MC, Groban L. Role of estrogen in diastolic dysfunction. *Am J Physiol Heart Circ Physiol* 2014; **306**: H628-H640 [PMID: 24414072 DOI: 10.1152/ajpheart.00859.2013]

144 **Guan R**, Xu X, Chen M, Hu H, Ge H, Wen S, Zhou S, Pi R. Advances in the studies of roles of Rho/Rho-kinase in diseases and the development of its inhibitors. *Eur J Med Chem* 2013; **70**: 613-622 [PMID: 24211637 DOI: 10.1016/j.ejmech.2013.10.048]

145 **Surma M**, Wei L, Shi J. Rho kinase as a therapeutic target in cardiovascular disease. *Future Cardiol* 2011; **7**: 657-671 [PMID: 21929346 DOI: 10.2217/fca.11.51]

146 **Mendoza-Torres E**, Oyarzún A, Mondaca-Ruff D, Azocar A, Castro PF, Jalil JE, Chiong M, Lavandero S, Ocaranza MP. ACE2 and vasoactive peptides: novel players in cardiovascular/renal remodeling and hypertension. *Ther Adv Cardiovasc Dis* 2015; **9**: 217-237 [PMID: 26275770]

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**Table 1 Rho kinase downstream target proteins, some signaling pathways and cellular functions**[8]

|  |  |  |
| --- | --- | --- |
| **ROCK downstream target protein** | **Signaling pathway** | **Function** |
| Myosin binding subunit of MLC/MYPT1 | MYPT1/MLC | Stress fiber formation |
| MLC2 | MYPT1/MLC2 | Mediates calcium sensitization and thereby enhances and sustains contraction in the vascular bed |
| LIM kinase/cofilin | LIM kinase/cofilin | Stress fiber formation |
| Ezrin/radixin/moesin |  |  |
| Adducin |  |  |

ROCK: Rho kinase; MLC: Myosin light chain; MYPT1: Phosphatase;

**Table 2 Cellular functions controlled by the RhoA/Rho kinase pathway[7-9]**

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| --- |
|  |

**Cytoskeletal dynamics**

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| --- |
|  |

Actin organization (formation of stress fibers and focal adhesion complexes)

Cell contraction

Adhesion

Morphology

Motility

Transcriptional regulation (pro remodeling genes)

Cell proliferation and cytokinesis

Differentiation

Apoptosis

Insulin-stimulated insulin receptor substrate (IRS)-1 phosphorylation

Development

|  |
| --- |
|  |



**Figure 1** **Rho kinase activation and downstream effects on cardiovascular remodeling in hypertension (as well as in cardiovascular disease)**. GEFs: Guanine nucleotide exchange factors; GAPs: GTPase activating proteins; GTP: Guanosine-triphosphate; GDP: Guanosine-diphosphate; MYPT1: Myosin binding subunit of myosin light chain phosphatase 1; MLC2: Myosin light chain 2; ERM: Ezrin/radixin/moesin; SMC: Smoorth muscle cell. Adapted from references[2,47,144,145].

**Figure 2** **Comparative levels of Rho kinase activity in circulating leukocytes (determined as phosphorylated/total MYPT-1 ratio) in healthy normotensive controls, in untreated hypertensive patients without LVH and in untreated hypertensive patients with left ventricular hypertrophy** **(Data shown as Mean + SEM).** 1: *P* < 0.01 *vs* Controls; 2: *P* < 0.01 *vs* untreated hypertensive patients without left ventricular hypertrophy (after significant ANOVA, respectively). Adapted, with permission from reference[47].LVH: Left ventricular hypertrophy.



**Figure 3** **Rho kinase activity in circulating leukocytes (determined as phosphorylated/total MYPT-1 ratio) in untreated hypertensive patients (white bar, mean age 48 years, mean BP 121 mmHg) and in hypertensive diabetic patients under antihypertensive and anti-diabetic pharmacological treatment** **(black bar, mean age 51, mean BP 111 mmHg).** Data shown as Mean + SEM, adapted with permission from reference[48].

**Figure 4 Schematic view of the current renin angiotensin system and sites of possible therapeutic interventions in hypertension and cardiovascular remodeling**. Letters in blue: Receptor agonists or enzyme activators; surrounded by interrupted lines: Enzyme/receptor inhibitors. AT1R: Angiotensin II receptor type 1; AT2R: Angiotensin II receptor type 2; ACE: Angiotensin converting enzyme I; ACE2: Angiotensin converting enzyme 2; MrgDR: Mas-related G protein–coupled receptor; NEP: Neutral endopeptidase; TOP: Thimet oligopeptidase; POP: Prolyl oligopeptidase; PCP: Prolyl carboxypeptidase. Adapted and modified from reference[146].