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***Basic Study***

**Alterations in g****ut microbiota during remission and** **recurrence of diabetes after** **duodenal-jejunal bypass in rats**

Zhong MW *et al.* Gut microbiota, duodenal-jejunal bypass and T2DM

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**Abstract**

**AIM:** To observe the alterations in gut microbiota in high-fat diet (HFD)-induced diabetes recurrence after duodenal-jejunal bypass (DJB) in rats.

**METHODS:** We assigned HDF- and low-dose streptozotocin-induced diabetic rats into two major groups to receive DJB and sham operation respectively. When DJB completed, we used HFD to induce the diabetes recurrence. Then we grouped the DJB-operated rats by blood glucose level into the DJB-remission (DJB-RM) group and the DJB-recurrence (DJB-RC) group. At a sequence of time points after operations, we would compare calorie content in the food intake (calorie intake), oral glucose tolerance test, homeostasis model assessment of insulin resistance (HOMA-IR), concentrations of glucagon-like peptide 1 (GLP-1), serum insulin, total bile acids (TBAs) and lipopolysaccharide (LPS) and alterations in colonic microbiota.

**RESULTS:** The relative abundance of *Firmicutes* in the control (58.06% ± 11.12%; *P <* 0.05 *vs* sham; *P <* 0.05 *vs* DJB-RC)and DJB-RM (55.58% ± 6.16%;*P <* 0.05 *vs* sham; *P <* 0.05 *vs* DJB-RC) groups was higher than that in the sham (29.04% ± 1.36%) and DJB-RC (27.44% ± 2.17%) groups; but the relative abundance of *Bacteroidetes* was lower (control group: 33.46% ± 10.52%,*P <* 0.05 *vs* sham 46.88% ± 2.34%; *P <* 0.05 *vs* DJB-RC 47.41% ± 5.67%. DJB-RM group: 34.63% ± 3.37%, *P <* 0.05 *vs* sham; *P <* 0.05 *vs* DJB-RC). *Escherichia coli* was higher in the sham (15.72% ± 1.67%; *P <* 0.05 *vs* control; *P <* 0.05 *vs* DJB-RM) and DJB-RC (16.42% ± 3.00%; *P <* 0.05 *vs* control; *P <* 0.05 *vs* DJB-RM) groups than in the control (3.58% ± 3.67%) and DJB-RM (4.15% ± 2.76%) groups. Improved HOMA-IR (2.82 ± 0.73; *P <* 0.05 *vs* DJB-RC4.23 ± 0.72), increased TBAs (27803.17 ± 4673.42 ng/mL; *P <* 0.05 *vs* DJB-RC18744.00 ± 3047.26 ng/mL), and decreased LPS (0.12 ± 0.04 ng/mL; *P <* 0.05 *vs* DJB-RC 0.19 ± 0.03 ng/mL) were observed the in DJB-RM group; however, these improvements were reversed in the DJB-RC group, with the exception of GLP-1 (DJB-RM *vs* DJB-RC *P >* 0.05).

**CONCLUSION:**Alterations in gut microbiota may be responsible for the diabetes remission and recurrence after DJB, possibly by influencing serum LPS and TBAs.

**Key words:**Duodenal-jejunal bypass; Diabetes recurrence; Gut microbiota; Lipopolysaccharide; Total bile acids

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**Core tip:** To determine the alteration in gut microbiota during diabetes recurrence after the performance of duodenal-jejunal bypass (DJB), high-fat diet-fed (HFD) and low-dose streptozotocin-injected diabetic rats received DJB. We used postoperative HFD to cause the diabetes recurrence. Relative abundance of *Firmicutes* in diabetes-recurrence rats is lower than that in diabetes-remission rats, whereas higher relative abundance of *Bacteroidetes* and *Escherichia coli* is observed in diabetes-recurrence rats*.* Alterations in gut microbiota may cause diabetes to reappear postoperatively by influencing levels of serum lipopolysaccharide and total bile acids, which have links with low-grade inflammation and glycolipid metabolism in diabetes.

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**INTRODUCTION**

Bariatric surgery can provide the most significant effectiveness in treating type 2 diabetes mellitus (T2DM)[1,2], and the past decade saw an steady growth in the amount of bariatric procedures performance all over the word[3]. Although bariatric surgery offers a rapid resolution of T2DM, the long-term remission rate usually decreases over time, and several previous studies have observed that, a part of patients, achieving initial postoperative resolution, experienced T2DM again[4-7]. Several factors have been found to be closely related to diabetes recurrence, including preoperative body mass index(BMI), age, course and gravity of T2DM, percentage of excess body weight loss(%EBWL), weight regain, postoperative diet and lifestyle[8-12].

Approximately 100 trillion bacterial cells, 10 times of human cells, colonize in the human gut[13]. According to recent data, gut microbiota can regulate the host physiology and metabolism by harvesting more energy from diet, modulating lipid metabolism, regulating bile acid biosynthesis, and increasing inflammatory tone[14]. Gut microbial dysbiosis, a considered environmental factor, can regulate obesity and T2DM[15,16]. Gastric bypass, the most effective way in treating obesity and T2DM, can affect host’s gut microbiota. A study including with individuals undergoing gastric bypass surgery has reported the changes in the abundance of specific gut microbe[17]. By the transformation of gut microbiota from mice with gastric bypass to germ-free mice, a research has proved the causal link between gut microbiota and the effect of gastric bypass[18].

A previous study of us demonstrated an effect in deteriorating glucose tolerance of a high-fat diet (HFD) after initial improvement in diabetic rats[19]. HFD seems to be one of the primary factors in obesity and T2DM[20]. And the diet, served as the major force, contributes much to the composition formation of gut microbiota[21-23]. Alterations in gut microbiota, furthermore, exert considerable influence on diabetes’ pathogenesis and postoperative remission. The association between gut microbiota and postoperative diabetes recurrence has not been explored so far. We also speculated that the postoperative diabetes recurrence may be involved with alterations in gut microbiota.

In our study, we performed DJB on HFD- and STZ-induced diabetic rats. Postoperatively, the HFD is used to induce diabetes recurrence. The gut microbiota, glucose profiles and serum parameters including levels of glucagon-like peptide 1 (GLP-1), insulin, total bile acids (TBAs) and lipopolysaccharide (LPS) were determined and compared between the groups.

**MATERIALS AND METHODS**

***Animals and diets***

With the condition that constant temperature is 24 to 26 °C, humidity is 50% to 60%, and light dark alternates each 12 h, eight-week-old Wistar rat, provided by Laboratory Animal Center of Shandong University, were separately housed in independently ventilated cages. Then we picked 7 from 30 male rats as the control group, which were given free access to standard chow (14% calories from fat, Shandong University Laboratory Animal Center) till the study ended. In order to induce insulin resistance, we fed the remaining rats for 4 wk with a HFD (Huafukang Biotech Company, China), which contains 40% fat as calories. A twelve-hour-fasting period was succeeded by intraperitoneal injection with 2% STZ (Sigma, United States), at the dose of 35 mg/kg weight, so the rats reached a diabetic state. After two weeks, 21 rats met the diabetic criterion — a blood glucose level at least 16.7 mmol/L during oral glucose tolerance test (OGTT) — and were randomly divided to two groups receiving sham (*n =* 7) or DJB operation (*n =* 14), in which rats were fed HFD postoperatively. Twelve weeks after surgery, the DJB group was subdivided into the DJB-recurrence group (DJB-RC, 5 rats, defined as having blood glucose ≥ 16.7 mmol/L during OGTT) and the DJB-remission group (DJB-RM group, 6 rats).

Body weight and calorie content in the food intake (calorie intake) were measured at baseline, 4, 8, and 12 wk postoperatively. Finally, all rats were euthanized by chloral hydrate overdose (intraperitoneal injection, 15 mL/kg, 10% chloral hydrate) for tissue collection. All animal experimental procedure involving in our study has been approved by the Animal Care and Utilization Committee of Qilu Hospital of Shandong University, Jinan, China.

***Surgical techniques***

Fifteen days after induction of diabetes, a low-residue diet was administered to the rats in sham group and DJB group from 48 h before surgery to 72 h after surgery. We performed DJB or sham surgery on rats under anesthesia using 10% chloral hydrate at the dose of 3 mL/kg. All of the surgeries were completed within 3 days. Seventy-two postoperative hours, all rats in both groups were allowed access to the HFD and water ad libitum.

**DJB:** We transected the duodenum just at the site of connection with the pylorus, and 7–0 silk sutures (Ningbo Medical Needle, China) were applied in the closure of the duodenum distal end, followed by transecting jejunum 15 cm at the site of distal to the Treitz’s ligament. Then, we proceeded anastomose of the proximal end of the duodenum and distal jejunum (duodenojejunal anastomosis). Moreover, we anastomosed jejunum proximal end to the antimesenteric border of the alimentary limb 15 cm distal to the duodenojejunal anastomosis, to form an end-to-side anastomosis.

**Sham:** We transected the intestines at the same sites with those in enterotomies of DJB, and then carried out re-anastomosis *in situ*. Because of similar stress from surgery and anaesthesia, the duration of the sham operation was similar to DJB.

***Oral glucose tolerance test***

We conducted the oral glucose tolerance test (OGTT) at several time points of baseline, postoperative 4 and 12 wk. Eight-hour fasting completing, all rats would receive 1 g/kg glucose by oral gavage. Then we estimated the levels of blood glucose at six time points (baseline, 15, 30, 60, 90, 120 min administration) respectively.

***Insulin, GLP-1, TBAs and LPS***

During the OGTT at 4 and 12 weeks postoperatively, we respectively gathered blood samples by retrobulbar venous plexus approach at time points of baseline 15, 30, 60, and 120 min after gavage with glucose, to collect serum by centrifugation (1006 × *g,* 4 °C, 15 min) and stored at −80 °C for further measurement. Concentrations of insulin and GLP-1 in serum, LPS in fasting serum were tested by enzyme-linked immune sorbent assay (ELISA) kits of Millipore( MA, United States),kits of Uscn Life Science Inc. (Wuhan, China), and Rat LPS ELISA Kit (Bio-Swamp, Wuhan, China) respectively. Levels TBAs in Fasting serum were detected by Hitachi automatic biochemical analyzer (Japan).

***Homeostasis model assessment of insulin resistance***

The calculations of homeostasis model assessment of insulin resistance (HOMA-IR), in 4 and 12 wk postoperatively, aim to evaluate insulin resistance using this formula: HOMA−IR=fasting insulin (mIU/L) × fasting glucose (mmol/L)/22.5[24].

***16S rDNA-based study of gut microbiota***

Because host metabolism can be influenced by the biological activity of microbiota in the colon, we focused our study on the colonic microbiota. At 12 wk after surgery, all rats were narcotized with 10% chloral hydrate (3 mL/kg). A three- centimeter segment of colon was ligatured and removed from enterocoelia. The colon was incised longitudinally, then, we gathered the colonic contents and stored them at −80 °C. All operations were performed under aseptic conditions. Genomic DNA of colonic microbiota was isolated based on the protocol of an E.Z.N.A. Soil DNA kit (Omega, Norcross, GA, United States). The method to amplify the V4 region of microbial 16S rDNA g by PCR has been reported previously[25]. We use Illumina MiSeq platform (BGI Technology, China) to sequence the amplified V4 region. The raw data were filtered to eliminate the adapter pollution and low quality and to obtain clean reads. Then paired-end reads with overlap were merged to tags, which were clustered, at 97% sequence similarity, to operational taxonomic unit (OTU). By using Ribosomal Database Project (RDP) Na, e Bayesian Classifier v.2.2, we assigned taxonomic ranks to OTU representative sequence. At last, the different species screening tests were analyzed based on OTU and taxonomic ranks.

***Statistical analysis***

All quantitative data were presented as mean±SD. By the use of trapezoidal integration, we calculated the area under the curves for OGTT (AUCOGTT). Body weight, calorie intake, AUCOGTT, HOMA-IR, TBAs and LPS were evaluated with the use of one-way analysis of variance (ANOVA) followed by Bonferroni *post hoc* comparison. All the data of the insulin and total GLP-1 concentrations were analyzed by the mixed model ANOVA followed by Bonferroni *post hoc* comparison after glucose administered by gavage to rats. All statistical calculations were processed with the use of SPSS version 19.0 (IBM, United States), at an alpha level of 0.05.

The information of the 16S rDNA-based study involving gut microbiota was analyzed by BGI Technology. Package 'ade4' of software R (v3.0.3) was used in principal component analysis (PCA). Heatmap was generated using the package “gplots” of software R (v3.0.3). The relative abundance of specific microbes between groups was analyzed by the Wilcoxontest.

**RESULTS**

When the study ended 12 wk after surgery, 7, 7, 5, and 6 rats were alive in the control, sham, DJB -RC and DJB-RM groups, respectively. Three rats died of anastomotic leak after DJB.

***Body weight and calorie intake***

Compared withrats inthe other three groups at any time points, the rats in the control grouphadmarkedly lowerbody weight and calorie intake (*P value <* 0.05) (Figure 1a and 1b). The differences of other groups were not significant.

***OGTT***

As Figure 2c showed, compared with other three groups, preoperative AUCOGTT of the control group (797.04 ± 81.01)was statistically lower (*P <* 0.05 *vs* sham1913.46 ± 184.55; *P <* 0.05 *vs* DJB-RC1869.90 ± 216.37; *P <* 0.05 *vs* DJB-RM, 1935.88 ± 250.73), with no differences among the other three groups. The postoperative AUCOGTT of the control group remained unchanged. Compared with the sham group (4 wk after surgery: 1935.88 ± 250.73; 12 wk after surgery: 1985.57 ± 152.56), both the DJB-RM (4 wk after surgery: 920.88 ± 62.90, *P <* 0.05 *vs* sham; 12 wk after surgery: 1009.86 ± 119.90*, P <* 0.05 *vs* sham)and control (4 wk after surgery: 827.86 ± 63.83,*P <* 0.05 *vs* sham; 12 wk after surgery: 895.39 ± 44.80*, P <* 0.05 *vs* sham) groups showed the lower AUCOGTT at all postoperative time points, indicating a deterioration in diabetes in sham rats. Notably, the AUCOGTT in the DJB-RC group (890.85 ± 114.41) was comparable to that in the DJB-RM group (920.88 ± 62.90, *P >* 0.05*vs*DJB-RC) 4 wk postoperatively, which was higher when compared with DJB-RM group 12 wk postoperatively (*P <* 0.05DJB-RC *vs* DJB-RM, 1876.20 ± 178.48 *vs* 1009.86 ± 119.90), demonstrating that the improved glucose tolerance in the DJB-RC group was reversed.

***Serum insulin***

Figure 2a and 2b showed the postoperative curves of serum insulin levels during the OGTT. It can be seen that no statistically difference was observed in serum insulin between the groups 4 wk and 12 wk postoperatively.

***HOMA-IR***

Postoperative HOMA-IR is shown in Figure 2d. Four weeks after surgery, compared with other three groups, HOMA-IR in the sham group (5.18 ± 0.98) was higher (*P <* 0.05 *vs* control 2.08 ± 0.38; *P <* 0.05 *vs* DJB-RC 2.30 ± 0.46; *P <* 0.05 *vs* DJB-RM, 2.17 ± 0.60), and no difference was observed among the control group, DJB-RC group and DJB-RM group. Consistent with the results of OGTT, 12 wk after surgery, the DJB-RC group (4.23 ± 0.72) exhibited higher HOMA-IR values than the DJB-RM group (2.82 ± 0.73, *P <* 0.05 *vs* DJB-RC), indicating re-impaired insulin sensitivity in the DJB-RC group 12 wk after surgery.

***GLP-1***

At both 4 (Figure 3a) and 12 wk (Figure 3b) after surgery, total serum GLP-1 secretion was comparable among the control group, DJB-RM group and DJB-RC groups (*P >* 0.05), and rats in the sham group showed a lower total serum GLP-1 secretion(*P <* 0.05 *vs* control; *P <* 0.05 *vs* DJB-RC; *P <* 0.05 *vs* DJB-RM).

***Fasting serum TBAs***

As shown in Figure 4a, at both 4 and 12 wk after surgery, TBAs levels were higher in the DJB-RC (4 wk after surgery: 21161.60 ± 2550.10 ng/mL, *P <* 0.05 *vs* control, *P <* 0.05 *vs* sham; 12 wk after surgery: 18744.00 ± 3047.26 ng/mL, *P <* 0.05 *vs* control, *P <* 0.05 *vs* sham) and DJB-RM (4 wk after surgery: 19543.00 ± 2639.35 ng/mL, *P <* 0.05 *vs* control, *P <* 0.05 *vs* sham; 12 wk after surgery: 27803.17 ± 4673.42 ng/mL, *P <* 0.05 *vs* control, *P <* 0.05 *vs* sham) groups than in the control (4 wk after surgery: 11608.00 ± 1248.10 ng/mL;12 wk after surgery: 13001.86 ± 1613.55 ng/mL) and sham (4 wk after surgery: 13190.14 ± 1237.38 ng/mL;12 wk after surgery: 12064.86 ± 1809.36 ng/mL) groups. Notably, at 12 wk postoperatively, rats in DJB-RM group showed higher TBAs than rats in DJB-RC group (*P <* 0.05).

***Fasting serum LPS***

As shown in Figure 4b**,** 4 wk after surgery, levels of fasting serum LPS in the sham group (0.20 ± 0.03 ng/mL) were higher compared with other three groups(*P <* 0.05 *vs* control 0.10 ± 0.02 ng/mL; *P <* 0.05 *vs* DJB-RC 0.13 ± 0.02 ng/mL; *P <* 0.05 *vs* DJB-RM, 0.12 ± 0.04 ng/mL). Twelve weeks after surgery, the sham (0.22 ± 0.04 ng/mL) and DJB-RC (0.19 ± 0.03 ng/mL) groups exhibited similar levels of LPS, but higher levels than the control(0.11 ± 0.02 ng/mL*; P <* 0.05 *vs* sham; *P <* 0.05 *vs* DJB-RC) and DJB-RM (0.12 ± 0.04 ng/mL*; P <* 0.05 *vs* sham; *P <* 0.05 *vs* DJB-RC) groups.

***Gut microbiota***

As demonstrated by PCA (Figure 5a), the gut microbiota showed marked changes in the four groups, which indicated a relative-centralized tendency intra-group and a relative-dispersed distribution inter-group. These tendencies were also observed in the Heatmap Analysis at the phylum level (Figure 5b).

The taxonomic composition distribution histograms, at the phylum level, in each group of rats are shown in Figure 6a. *Bacteroidetes* were the predominant gut microbes in the sham (46.88% ± 2.34%) and DJB-RC (47.41% ± 5.67%) groups (Figure 6b), while *Firmicutes* were predominant in the control (58.06% ± 11.12%) and DJB-RM groups (55.58% ± 6.16%, Figure 6c). The relative abundance of *Proteobacteria* was higher in the sham (24.14% ± 2.89%) and DJB-RC (21.83% ± 5.09%) groups than in the control (6.86% ± 3.79%; *P <* 0.05 *vs* sham; *P <* 0.05 *vs* DJB-RC) and DJB-RM groups (8.31% ± 4.39%; *P <* 0.05*vs*sham; *P <* 0.05 *vs* DJB-RC; Figure 6d).

It is noteworthy that although resolution at the species level was low, the relative abundance of *Escherichia coli* (*E. coli*) in the sham (15.72% ± 1.67%) and DJB-RC (16.42% ± 3.00%) groups was remarkably higher than that in the control(3.58% ± 3.67%; *P <* 0.05*vs*sham; *P <* 0.05 *vs* DJB-RC) and DJB-RM groups (4.15% ± 2.76%; *P <* 0.05*vs*sham; *P <* 0.05 *vs* DJB-RC; Figure 6e).

**DISCUSSION**

With the understanding of bariatric surgery developing, more and more clinicians and patients consider it as their first choice of treating obesity and T2DM, during which procedure Roux-en-Y gastric bypass (RYGB) is performed most frequently as before[3]. Bariatric surgery achieves a higher diabetes remission rate than non-surgical way in treating diabetes[2,26]. However, the recurrence of diabetes after initial remission should not be ignored. Due to limitations in clinical research, we established an animal model to investigate the mechanisms of diabetes recurrence after surgery. Our research group previously reported that the re-impairment of insulin sensitivity was a major factor of diabetes postoperative recurrence in an animal study[19]. The HFD, a widely used insulin resistance inducer in rats, was adopted in the present and in our previous studies to reverse the improvement in diabetes[19,27]. In the model of diabetes remission and recurrence, we hoped to delineate the major mechanisms of diabetes postoperative recurrence.

In this study, statistically significant differences in body weight were not seen among the sham, DJB-RC and DJB-RM groups. Furthermore, the body weight of all DJB rats increased after surgery. Our study including the DJB model was devised to estimate that whether the bypass surgery has the weight-independent anti-diabetic effects, and then the effects were proved[28]. This indicated that the remission and recurrence of diabetes after surgery were independent of body weight, which was consistent with our previous studies and with the common view that the rapid anti-diabetic effect postoperatively has nothing to do with weight loss[29,30]. Some clinical trials have reported that the diabetes recurrence was linked to inadequate weight loss and weight regain[1,31], which contradicted the results obtained in our study.

T2DM is a metabolic disease featured with insulin resistance and function loss of pancreatic beta-cells[32]. The present study has observed that impaired insulin secretion during the OGTT did not increase after surgery, demonstrating that the remission of diabetes was the result of improved insulin sensitivity represented by a lower HOMA-IR, which was in accordance with our previous study[33]. Taking into account that insulin resistance and function loss of pancreatic beta-cells could lead to T2DM, and compared with DJB-RM rats, DJB-RC rats exhibited comparable secretion of insulin after gavage and re-impaired insulin sensitivity represented by a higher HOMA-IR at the end of this study, we concluded that the recurrence of diabetes after initial remission in DJB rats was due to the re-deterioration of improved insulin sensitivity. Interestingly both DJB-RC and DJB-RM rats experienced the same pre- and post-operative management and procedures, these rats, however, showed opposite glucose profiles and different insulin sensitivity. To analyze the intrinsic mechanisms involved in the re-deterioration of insulin sensitivity, factors related to insulin resistance including gut microbiota, serum TBAs, LPS and GLP-1 were measured.

The gut microbiota, known as an environmental factor, can affect obesity and diabetes, and it can take effect between the host genotype and diet, responsible for the metabolic process of host glucose and lipid being modulated[14]. [Larsen](http://www.ncbi.nlm.nih.gov/pubmed/?term=Larsen%20N%5BAuthor%5D&cauthor=true&cauthor_uid=20140211) *et al*[34] recently reported that gut microbial composition was related to T2DM, and observed that, compared with non-diabetic people, *Firmicutes* in T2DM patients significantly decreased. Moreover, the positive correlation existed between the *Bacteroidetes*-to-*Firmicutes* ratio and the plasma glucose level, rather than exist between the ratio and BMI. The present study has observed that, compared with control group, diabetic rats in sham groups offered fewer *Firmicutes* and more *Bacteroidetes* in terms of relative abundance.

After surgery, *Firmicutes*, compared with sham rats, increased remarkably in DJB-RM rats and whereas *Bacteroidetes* decreased, and these results were consistent with what reported by Ryan *et al*[35]. Compared with DJB-RM rats, DJB-RC rats showed the reverse microbial composition in terms of relative abundance of *Firmicutes* and *Bacteroidetes*, the trend was similar to that in sham rats. Based on the above findings, a conclusion could be made that there was a close connection between the altered *Firmicutes*-to-*Bacteroidetes* and diabetes postoperative remission and recurrence.

Except for *Firmicutes* and *Bacteroidetes*, compared with the control group and DJB-RM group, the sham group and DJB-RC group had greater relative abundance of *Proteobacteria*. Further species annotation indicated that the difference in *Proteobacteria* was due to alterations in *E. coli*. In addition, serum LPS levels showed the same tendency as the relative abundance of *E. coli*, where sham and DJB-RC rats had higher serum LPS than control and DJB-RM rats. LPS, a structure material in cell wall of *E. coli* is a low-grade inflammation and insulin resistance inducer. As a result of chronic infusion of LPS through 4 wk, the chow-fed mice demonstrated increased adiposity and infiltration of macrophage in adipose tissue, and the inflammation and insulin resistance liver of developed[36]. As evidence shown above, we surmised that the alterations in *E. coli* may be a major factor in triggering postoperative diabetes by influencing serum LPS levels. However, our study only observed a similar trend of LPS to the trend of the relative abundance of *E. coli*. The alterations in LPS may be the result of combined effects of gut microbiota, gut barrier function and host’s immunity. We could not demonstrate the exact causal relationship between LPS and *E. coli,* which need to be verified in further studies.

Gut microbiota exerts a role of regulation in synthetizing bile acids synthesis and producing[37-40]. Bile acids can convey signaling information and regulate metabolic process of lipid, glucose and energy by farnesoid X receptor (FXR) and G-protein-coupled receptor 5 (TGR5)[41]. Our study showed that, fasting serum TBAs levels increased 4 wk after surgery in all DJB rats, which had no difference between the control and sham groups. Similar changes were observed by other researchers and in our previous studies[42,43]. When the study ended 12 wk after surgery, the fasting serum TBAs levels in DJB-RC rats decreased and were lower than those in DJB-RM rats, which was consistent with changes in insulin sensitivity as assessed by HOMA-IR. These results suggested that the changes in TBAs may be other factors influencing the remission and recurrence of diabetes after DJB.

GLP-1 is the most significant jut hormones, the source of which is L cells mostly existing in the epithelium of distal ileum and colon[44]. GLP-1 functions as a regulator of glucose homeostasis through the improvement of insulin secretion, inhibition of glucagon secretion and apoptosis of beta cells, and promotion of proliferation of beta cells[45]. Mounting evidence has confirmed that postprandial GLP-1 secretion would be enhanced after DJB[46], which is reconfirmed by our study. No statistically significant difference, however, between the DJB-RC group and DJB-RM group, indicated that GLP-1 was not related to the recurrence of diabetes, and enhancement of GLP-1 secretion after surgery partially contributed to diabetes remission.

There are some limitations in this research. First, the gut microbiota is a dynamically changing process, which is distributed throughout the entire digestive tract. Based on measurements of gut microbiota composition in different segments and at different time points, the function of gut microbiota after surgery could be well explained. Second, this study mainly focused on the alterations in gut microbiota during the remission and recurrence of diabetes after surgery, and LPS and TBAs were examined to help understand the role of microbes in the changes of glucose tolerance. Further studies on the mechanisms of diabetes remission and recurrence with alterations in gut microbiota are anticipated. Third, calorie intake in our study didn’t show the difference among the sham, DJB-RC and DJB-RM groups. However, it is a pity that the calorie content in faeces was not measured. So, the calorie absorbed from food intake could not be calculated.

In conclusion, a postoperative HFD can re-exacerbate insulin resistance and induce recurrence of diabetes after initial remission in DJB-operated rats. Alterations in gut microbiota may be responsible for the recurrence of diabetes after DJB, possibly by influencing serum LPS and TBAs.

**COMMENTS**

***Background***

Bariatric surgery can contribute to remission of type 2 diabetes mellitus. Some patients, however, experienced diabetes again postoperatively. Several factors have been found to be closely related to diabetes recurrence, including percentage of excess body weight loss, body mass index before operation, age, duration and severity of diabetes, weight regain, postoperative diet and lifestyle. The intrinsic mechanism of diabetes postoperative recurrence, however, remains unclear.

***Research frontiers***

Not only can gut microbiota modulate host metabolism, but it also exerts an influence on diabetes postoperative remission. Alterations in gut microbiota after bariatric surgery may be responsible for diabetes remission possibly by influencing serum levels of lipopolysaccharide (LPS) and total bile acids (TBAs).

***Innovations and breakthroughs***

This study created the high-fat diet-induced rat models of diabetes recurrence after duodenal-jejunal bypass (DJB) and determined the alterations in gut microbiota during diabetes recurrence after DJB. The rats with diabetes recurrence presented a phenomena with a reduced relative abundance of *Firmicutes* and an increased *Bacteroidetes* and *Escherichia coli*. Alterations in gut microbiota may be responsible for the diabetes postoperative recurrence, possibly by influencing serum LPS and TBAs.

***Applications***

The findings in this study could enable investigators focus more on the link between gut microbiota and diabetes postoperative recurrence. A new design of therapeutic interventions aiming to target gut microbiota may prevent diabetes postoperative recurrence.

***Terminology***

DJB, served as an experimental procedure, was devised to estimate the anti-diabetic effects independent from weight of Roux-en-Y gastric, which is the most effective treatment for diabetes.

***Peer-review***

This is a very well designed, performed and written experimental study for investigation of the role of alterations in gut microbiota in the pathogenesis and remission of type 2 diabetes after bariatric surgery. For investigation of this aim the authors created and used a rat model of diabetes recurrence after DJB and compared the results obtained in diabetes rats induced by high-fat diet after bypass and in the group with sham operation.

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**Figure 1 Body weight and calorie content in the food intake (****calorie intake).** Shown are body weight (a) and calorie intake (b) of rats at baseline, 4, 8, and 12 wk after surgery. a*P* < 0.05 *vs* control group.



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**Figure 2** **Serum insulin after oral glucose administration.** Shown are levels of serum insulin after oral glucose gavage (1 g/kg) at 4 wk (**a**) and 12 wk (**b**) after surgery, between which there are no significant differences with the use of mixed model one-way analysis of variance followed by Bonferroni *post hoc* comparisons. area under the curves for oral glucose tolerance test and homeostasis model assessment of insulin resistance were demonstrated in Figure 2c and 2d. a*P* < 0.05 *vs* control group; c*P* < 0.05 *vs* sham group; e*P* < 0.05 *vs* DJB-RC group. DJB-RC: duodenal-jejunal bypass recurrence group; DJB-RM: duodenal-jejunal bypass-remission group.



**Figure 3 Level of glucagon-like peptide 1 after administration of oral glucose**. Shown are glucagon-like peptide 1 (GLP-1) level after oral glucose gavage (1 g/kg) at 4 wk (panel a) and 12 wk (panel b) after surgery. In terms of global GLP-1 concentration, the illustrations in the rectangles show that there are significant differences between the groups with the use of mixed model one-way analysis of variance followed by Bonferroni *post hoc* comparisons. DJB-RC: duodenal-jejunal bypass recurrence group; DJB-RM: duodenal-jejunal bypass-remission group.



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**Figure 4 Fasting serum total bile acids and lipopolysaccharide**. Fasting serum TBAs (a) and LPS (b) were measured at 4 and 12 wk after surgery. a*P* < 0.05 *vs* control group; c*P* < 0.05 *vs* sham group; e*P* < 0.05 *vs* DJB-RC group. DJB-RC: duodenal-jejunal bypass recurrence group; DJB-RM: duodenal-jejunal bypass-remission group; TBAs: total bile acids; LPS: lipopolysaccharide.



**Figure 5 principal component analysis and Heatmap analysis.** A: Principal component analysis was used to construct a 2-D graph to summarize the factors mainly responsible for this difference. Similarity was high when two samples were closely located. Number in brackets represents contributions of principal components to differences among samples; b: Heatmapanalysis: The longitudinal clustering indicates the similarity of all species among different samples, and horizontal clustering indicates the similarity of certain species among different samples, the closer the distance and the shorter the branch length, the more similar the species composition is between the samples.



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**Figure 6 Species annotation**. Species annotation (a) is the taxonomic composition distribution histogram of each sample at the phylum level. The ratios of each phylum in certain samples are displayed. The relative abundance of *Bacteroidetes* (b), *Firmicutes* (c), *Proteobacteria* (d) and *Escherichia coli* (e) between the groups was analyzed by the Wilcoxontest. a*P* < 0.05 *vs* control group; c*P* < 0.05 *vs* sham group; e*P* < 0.05 *vs* DJB-RC group. DJB-RC: duodenal-jejunal bypass recurrence group; DJB-RM: duodenal-jejunal bypass-remission group.