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| CORE TIP | A retrospective analysis of 226 patients from Auckland, New Zealand found BK polyomavirus (BKV) as an uncommon cause of graft loss. Renal units with­out a formal BKV surveillance programme showed a similar incidence and outcomes for BK polyomavirus nephropathy (BKVN) to centres with an active screening programme. When designing a cost effective screening programme for BKV infection, it should be centre specific in relation to the units immunosuppression and monitoring protocol, epidemiology and outcomes of BKVN. |
| KEY WORDS | BK virus; BK polyomavirus nephropathy; Kidney transplantation; Screening |
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**Retrospective Cohort Study**

Outcomes of renal transplant recipients with BK virus infection and BK virus surveillance in the Auckland region from 2006 to 2012

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**Abstract**

**AIM**

To evaluate incidence, risk factors and treatment outcome of BK polyomavirus nephropathy (BKVN) in a cohort of renal transplant recipients in the Auckland region without a formal BK polyomavirus (BKV) surveillance programme.

**METHODS**

A cohort of 226 patients who received their renal trans­plants from 2006 to 2012 was retrospectively reviewed.

**RESULTS**

Seventy-six recipients (33.6%) had a BK viral load (BKVL) test and 9 patients (3.9%) developed BKVN. Cold ischaemia time (HR = 1.18, 95%CI: 1.04-1.35) was found to be a risk factor for BKVN. Four recipients with BKVN had complete resolution of their BKV infection; 1 recipient had BKVL less than 625 copies/mL; 3 reci­pients had BKVL more than 1000 copies/mL and 1 had graft failure from BKVN. BKVN has a negative impact on graft function [median estimated glomerular filtration rate (eGFR) 22.5 (IQR 18.5-53.0) mL/min per 1.73 m2, *P* = 0.015), but no statistically significant difference (*P* = 0.374) in renal allograft function was found among negative BK viraemia group [median eGFR 60.0 (IQR 48.5-74.2) mL/min per 1.73 m2), positive BK viraemia without BKVN group [median eGFR 55.0 (IQR 47.0-76.0) mL/min per 1.73 m2] and unknown BKV status group [median eGFR 54.0 (IQR 43.8-71.0) mL/min per 1.73 m2]. The incidence and treatment outcomes of BKVN were similar to some centres with BKV surveillance programmes.

**CONCLUSION**

Recipients with BVKN have poorer graft function. Although active surveillance for BKV has been shown to be effective in reducing incidence of BKVN, it should be tailored specifically to that transplant centre based on its epidemiology and outcomes of BKVN, particularly in centres with limited resources.

**Key words:** BK virus; BK polyomavirus nephropathy; Kidney transplantation; Screening

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**Core tip:** A retrospective analysis of 226 patients from Auckland, New Zealand found BK polyomavirus (BKV) as an uncommon cause of graft loss. Renal units with­out a formal BKV surveillance programme showed a similar incidence and outcomes for BK polyomavirus nephropathy (BKVN) to centres with an active screening programme. When designing a cost effective screening programme for BKV infection, it should be centre specific in relation to the units immunosuppression and monitoring protocol, epidemiology and outcomes of BKVN.

INTRODUCTION

Since the first discovery of BK polyomavirus (BKV) isolated from the urine of a renal allograft recipient with ureteric obstruction in 1971[1], BKV infection has emerged as an important cause of renal allograft dysfunction. In the modern era, with the use of more potent immuno­suppressive agents, BKV nephropathy (BKVN) has resulted in a significant rate of graft loss[2].

Seroprevalence of BKV in immunocompetent adults ranges from 65% to 90% and BKV remains latent pre­dominately in the urinary tract[3]. Immunosuppression, inflammation and insufficient anti-viral immune responses play an integral role in the reactivation and replication of BKV and progression to BKVN in renal transplant recipients[4]. Dendritic cell deficiency, seronegativity for BKV and impaired BKV-specific T-cell response are found to be associated with BK viraemia[5-8]. Other possible risk factors for BK viraemia include a long cold ischaemia time (CIT)[9], acute rejection[10,11], placement of a ureteric stent[12,13], human leukocyte antigen (HLA) mismatch[11,14], lymphocyte-depleting agents[15-18], tacrolimus[19,20] and steroids[16,19,20]. Given the complexity in the pathogenesis of BKVN, the intensity of immunosuppression may not be solely responsible for the development of BKVN and it may not be appropriate to generalise these predisposing risk factors in all renal transplant recipients. However, as the immunosuppressive burden appears a significant risk factor for the development of BK polyomavirus, it is possible that the immunosuppression regimen used accounts at least in part for the substantial variation in the prevalence of BK viruria (30%-62%), BK viraemia (11.5%-20%) and BKVN (1%-10%) among transplant centres[10,12,21,22].

No effective pharmacological treatment has con­sistently emerged for the treatment of BKV infection apart from a reduction of immunosuppressants[23,24]. Many investigators have recommended screening BK viraemia and pre-emptive reduction in immunosuppressants as a strategy to preserve graft function and reduce the risk of BKVN occurring[25-28]. However, a screening programme may not be suitable in a transplant centre with a low rate of BVKN[29].

We conducted a retrospective review of renal tran­splant recipients from our region, where no lymphocyte depleting induction treatment is used and the pre­dominant calcineurin inhibitor utilised is Ciclosporin, to determine the incidence of BKVN and/or BK viraemia and to evaluate the characteristics that are potentially associated with the development of BKVN and related treatment outcome.

MATERIALS AND METHODS

Patients

We searched through the Auckland Renal Transplant Group’s database to identify patients who received renal transplants from January 2006 to December 2012. Patients resident outside the Auckland region were not included in this retrospective review. We excluded those who died or had primary graft failure within 1 mo of receiving a renal allograft. Patients’ clinical notes with clinical and demographic data were obtained from the clinical electronic portal system, Concerto® and 3M® Health Information Systems, with data censored for 31 December 2013.

Testing for serum BKV viral load is performed at a single laboratory, LabPlus™, in the Auckland region. The laboratory does not routinely perform BKV viral load in urine samples, due to the high prevalence of positive BK viruria in renal transplant recipients. Recipients are considered to have BK viraemia for any level of viral load. Testing for serum creatinine is performed at both hospital and community laboratories in the Auckland region. Estimated glomerular filtration rate (eGFR) was calculated using the CKD-EPI equation[30]. The recipients’ renal biopsies performed in the Auckland region were processed and interpreted at a single laboratory: LabPlus™.

BKV surveillance

There is no formal screening programme among the three renal units in the Auckland region. One unit rou­tinely screened for BKV from the year 2012. One of these hospitals trialled screening for BKV in 2009, but halted this strategy due to cost, and BK viral load (BKVL) test is performed only if there is a biopsy-proven BKVN. The third unit checks for BKV if there is a clinical indication or a biopsy-proven BKVN.

Immunosuppression protocol

Basiliximab was routinely used for induction from August 2010. All male recipients were given Ciclosporin as their maintenance calcineurin inhibitor, but female recipients were given a choice of either Tacrolimus or Ciclosporin. The tacrolimus trough concentration in each recipient was kept between 10 to 15 g/L for the first 2 mo after transplant, and then the target concentration was kept between 5 to 10 g/L. The Ciclosporin (2 h post dose) target concentration was 1700 g/L for the first month after transplant and 1500 g/L for the second month. The target concentration was then gradually reduced to equal to or less than 800 g/L after 12 mo of transplantation. Every recipient received Mycophenolate mofetil (MMF) 1 g twice daily if on Ciclosporin or 750 mg twice daily if on Tacrolimus. Concentration monitoring for MMF is not routinely performed. All recipients re­ceived Methylprednisolone 1 g at induction followed by prednisone as maintenance. The prednisone dose was 30 mg daily for all recipients and it was tapered down to 7.5 mg daily or lower at months 4 post-transplant.

Acute rejection treatment protocol

Recipients who have acute rejection Banff grade Ⅰ receive Methylprednisolone 500 mg daily for 3 d. For Banff grade Ⅱ, Ⅲ and steroid-resistant rejections, patients receive lymphocyte-depleting agents. Biopsy-proven rejection in the presence of therapeutic Ciclosporin concentrations necessitates a conversion to Tacrolimus unless con­traindicated. The target Tacrolimus concentration is maintained between 7 to 10 g/L.

Renal transplant biopsy protocol

A protocol biopsy at 3 mo after transplantation is per­formed in all renal transplant recipients. Patients with a > 20% decline in graft function without a clear cause and all reversible non-renal causes excluded undergo renal biopsy.

Statistical analysis

SPSS version 22 was used to perform statistical analysis. Results are expressed as numbers (percentages), median (interquartile range Q1-Q3) and mean (± SD) unless otherwise stated. The 95%CIs are based on exact confidence limits. Data were compared by 2 test, Fisher’s exact test, non-parametric Mann-Whitney *U* test or Kruskal-Wallis test as appropriate. Cox regression was used to identify possible risk factors for BKVN. Those who died, developed graft failure unrelated to BKVN and were transferred to outside of the Auckland region were censored in this model.

**RESULTS**

Description of the study population

Four hundred and twenty-eight patients underwent renal transplantation between January 2006 and December 2012. After excluding 194 patients from outside of the Auckland region and 8 patients who died or developed primary graft failure at one month after transplant, 226 were included in the study (Figure 1).

Seventy-six recipients were tested for BKV (33.6%) at any point in time over the study period. Twenty-eight patients of 76 tested patients had BK viraemia (36.8%). Twenty of these 28 patients were managed with a reduction of their immunosuppressants; by reducing or stopping MMF and maintaining Tacrolimus trough con­centration and Ciclosporin trough concentration between 4-6 g/L and 100-150 g/L, respectively, depending on BKVLs’ responses. Seven patients received Leflunomide concurrently; one BKVN recipient was given additional Ciprofloxacin as the BKVL failed to decline despite reducing immunosuppressants, and another BKVN recipient also received Ciprofloxacin and subsequently intravenous Immunoglobulin due to persistent high level of BK virae­mia, worsening graft dysfunction and a repeat graft biopsy showed features of acute rejection in addition to BKVN. The remaining 8 BK viraemic recipients did not have a reduction of immunosuppressants due to low viral loads (all less than 1250 copies/mL).

Of these 28 patients, 16 patients had transplant biopsies for renal allograft dysfunction. Nine patients had biopsy-proven BKV nephropathy equivalent to an incidence of 11.8% (95%CI: 5.6%-21.3%) of the cohort tested for BKV viral load (9/76) or 4.0% (95%CI: 1.8%-7.4%) of the entire cohort (9/226). Eight of these nine patients were tested for BKVLs after their transplant biopsies diagnosed BKVN. The other patient with BKVN had the transplant biopsy and serum BKVL requested concurrently as part of investigation of graft dysfunction. The remaining seven transplant recipients with transplant biopsies for graft dysfunction did not have BKVN. The other 12 recipients did not have transplant biopsies as their graft functions were stable, and 7 of these 12 recipients had their immunosuppression reduced.

Three (33.3%) of the recipients with BKVN did not have acute rejection prior to the diagnosis of BKVN. The other 6 recipients (66.7%) had at least one episode of biopsy proven acute rejection requiring a pulse methy­lprednisolone course prior to the development of BKVN. Ten of the 19 recipients (53.7%) with BK viraemia did not have acute rejection prior to the diagnosis of BK viraemia and the others (47.3%) had at least one acute rejection prior.

Associations with BKV nephropathy

When comparing features between recipients with BKVN and without BKVN, we found that BKVN was more common in Māori, Pacific Islanders and Asians than in Europeans (European 1.5%% *vs* Asian 9.3% *vs* Māori and Pacific Islanders 7.4%, *P =* 0.038). The renal allografts of the recipients that developed BKVN were all from deceased donors (*P* = 0.005). The BKVN group had a longer CIT (median 19 h *vs* 7 h, *P* = 0.001). The group was also more likely to have at least 1 episode of acute rejection at any time point, though it was not statistically significant (*P* = 0.069). There was no significant difference observed in age, gender, co-morbidity, dialysis vintage, HLA mismatch, Tacrolimus, Basiliximab induction, or use of lymphocyte-depleting agents for acute rejection between the positive (+) BKVN and negative (-) BKVN groups (Table 1).

Comparison of recipients with BK viraemia only and recipients with BKVN

There was no statistical difference in ethnicity, acute rejection at any time point and other demographics when comparing the BKVN group and the (+) BKV group (recipients with only BK viraemia and no biopsy-proven BKVN). The (+) BKV group had fewer deceased donors (47.4%, *P* = 0.01) and shorter CIT (median 5.4 h, *P* < 0.0001) than patients who developed BKV nephropathy. BK viraemic recipients (47.4%) and recipients with BKVN (66.7%) were more likely to have acute rejection prior to the diagnosis of BKVN and BK viraemia as opposed to recipients without BK viraemia (27.1%, *P* = 0.048).

Risk factors for BKVN

In a univariable Cox regression analysis, Asian recipients had a greater risk of developing BKVN compared with European recipients (unadjusted HR = 6.36, 95%CI: 1.96-38.16, *P =* 0.043), but it was not seen in Māori and Pacific islands recipients (unadjusted HR = 4.75, 95%CI: 0.87-25.94, *P =* 0.072) (Table 2). Because CIT is dependent on donor source, donor source was not used in the analysis. Recipients with longer CIT had a higher risk of developing BKVN (unadjusted HR = 1.18, 95%CI: 1.06-1.32, *P =* 0.003). While acute rejection appeared to be associated with BKVN, this did not reach statistical significance (unadjusted HR = 3.72, 95%CI: 0.93-14.91, *P =* 0.063).

We included only the variables that had *P* value< 0.1 in a multivariable model. Ethnicity was not found to be significant, and Longer CIT was the only risk factor for BKVN (HR = 1.18, 95%CI: 1.05-1.39, *P* = 0.009).

Effect of BKVN and BK viraemia on renal allograft function

Renal allograft function in the BKVN group was signi­ficantly lower comparing with those in the other BKV status groups (*P* = 0.015), and the median graft func­tion of the recipients who were never checked for BK viraemia was similar to those with and without BK viraemia using the non-parametric Kruskal-Wallist test (*P* = 0.374) (Figure 2). After controlling for those factors (age at tran­splant, comorbidity, donor source, HLA mismatch, acute rejection, Basiliximab induction, calcineurin inhibitor and eGFR at 1 mo after transplant) that could potentially affect graft function, the mean eGFR of the recipients with BKVN (taken just before the censored date or before recipients were transferred, but those who developed graft failure were not included) was still 17.0 mL/min per 1.73 m2 (95%CI: -32.5 to -1.5 mL/min per 1.73 m2, *P =* 0.032) lower than that of those without BKVN. However, no significant impact on graft function was found in reci­pients with only BK viraemia (-4.1 mL/min per 1.73 m2, *P* = 0.464) and unknown BKV status (-1.1 mL/min per 1.73 m2, *P* = 0.754) comparing with the negative BK viraemic recipients.

One of the 9 recipients (11.1%) with BKVN had graft failure compared to 8 (3.7%) of those without, but there was no statistical difference found using a log-rank test in the Kaplan-Meier survival analysis (*P*log-rank= 0.283) (Figure 3). Similarly, no difference was found between those with known and unknown BKV status (0.92 for positive BKV and BKVN, 0.90 for negative BKV, and 0.92 for unknown BKV status, *P*log-rank= 0.568) (Figure 4).

Outcome of management of BKVN and BK viraemia

For those diagnosed with BKVN, BKVLs were unde­tectable in 4 recipients and the BKVL was less than 625 copies/mL in one recipient at the time of censoring. The other 3 patients still had persistent BK viraemia but viral loads were declining at the end of study period (Table 3). One patient developed graft failure due to BKVN, but it is important to note that this recipient’s graft function was poor prior to the diagnosis of BKVN (eGFR 23-27 mL/min per 1.73 m2) with no clear explanation despite 4 graft biopsies that did not show any significant abnormality. BKVLs of the recipients with BK viraemia were either undetectable or declining during the study period. None of these 19 patients, including six of them with BKVLs more than 104 copies/mL, progressed to BKVN over the study period.

Of the patients who had a reduction in immuno­suppression none of the 11 patients with BK viraemia had acute rejection, while two of the 9 recipients with BKVN developed an acute T-cell mediated rejection grade 1B at 30 mo and 2 mo after their immunosuppressive therapy was reduced. The acute rejections were successfully treated with intravenous pulse methylprednisolone, and their calcineurin inhibitors were already switched to Tacrolimus from previous acute rejection prior to the diagnosis of BKVN.

DISCUSSION

This retrospective study has demonstrated that only one patient had graft failure due to BKVN in our cohort of patients. We identified 28 patients with BKVinfection with a rate of 36.8% in the cohort that was screened. However, this has likely overestimated the incidence as 150 patients (66.4%) were not tested for BK viraemia. When considering the entire cohort the rate of BKV was 12.4% (28 of 226). The incidence of biopsy proven BKVN was 11.8% (9/76). It is important to note that it is unlikely that any patient with clinically significant BKV did not undergo testing due to the regional protocol to perform renal biopsy. However, it is possible that early BKVN could be missed by renal biopsy due to its focal nature[25]. Using the overall cohort we noted an incidence of 4.0% (9/226) which is comparable to that of other transplant centres with an active BKV screening programmes (0.8% to 6.4%)[11,26-28,31-33].

A long CIT was identified as a potential risk factor for BKVN in our study, and is known to cause severe ischaemia-reperfusion injury resulting in intragraft inflammation, which in turn is found to stimulate BK polyomavirus DNA replication[34,35]. Acute rejection results in tubulointerstitial inflammation and typically leads to also an increased burden of immunosuppression. How­ever, interestingly in our cohort, acute rejection was not found to be a risk factor for progressing to BKVN. Although others have demonstrated an association be­tween Tacrolimus use and BKVN, Tacrolimus was not found to be associated with BKVN in our cohort. Despite lymphocyte-depleting agents were used for severe acute rejection episodes in this cohort, our analysis did not demonstrate more recipients with BK viraemia and BKVN. Other studies have also failed to show any correlation between lymphocyte-depleting agents given at induction and BKV infection[26,28].

Because of the difficulty in predicting BKVN in trans­plant recipients, a latent period from BK viraemia to the development of BKVN, and possible sampling errors in diagnosing early BKVN due to its focal nature[25], screening of the blood or urine for BKV infection and pre-emptive reduction of immunosuppressants when BKV is detected have been shown to reduce, the risk of the development of BKVN (0.8%-1.3%)[27,28,32,33]. Interestingly, some centres report that despite having a BKV screening programme in the literature, their incidence of BKVN is greater than that seen in our cohort (4.2%-6.4%)[11,26,31], This includes one centre that screened for decoys cells in urine fortnightly for the first 3 mo, then at 6 and 12 mo and yearly after transplant[31]. It appears that frequent monthly BKV monitoring is essential in early detection of BK viraemia and intervention resulting in the lower incidences of BKVN described in the abovementioned studies, with one centre that performed a cheaper urine cytology screening fortnightly from 0 to 3 mo after trans­plant, monthly from 3 to 6 mo and then every 2 mo from 6 to 12 mo[33].

Because all patients with a > 20% decline in renal function undergo routine biopsy and all patients undergo a protocol biopsy at 3 mo after transplantation in our study cohort, we postulate that a large proportion (66.4%) of our cohort with unknown BKV status had no episodes of graft dysfunction due to BKVN. Bohl and his co-inves­tigators looked at other studies and commented that BKVLs exceeding 104 copies/mL only have a positive predictive value of 50%-85% for diagnosing BKVN[36]. BKVLs that are less than 104 copies/mL do not require intervention[28,37]. This is reflected not only on the re­cipients who never had any BKVL tests, but also on the eight patients with low BKVLs who did not progress to BKVN even without a reduction of immunosuppressants and further BKV monitoring.

BKVN impacted on graft function; one patient lost their allograft and the other eight patients had a mean eGFR more than over 18 mL/min worse than patients without BKVN. This finding of poorer graft function from BKVN is also found in a prospective study that adopted a rigorous monthly screening programme - the mean eGFR of recipients with BKVN (39.0 ± 14.3 mL/min per 1.73 m2) was lower than that of those without BKVN (52.3 ± 19.9 mL/min per 1.73 m2), though there was no graft failure due to BKVN[28].

Frequent monthly BKV monitoring and pre-emptive reduction of immunosuppression have been shown to be beneficial in reducing the risk of BKVN occurring. This strategy is effective in improving overall graft outcomes in renal transplant recipients as there is no definitive medical therapy for BKVN with graft failure still occurring. However, this approach may be cost prohibitive for those resource restricted centres especially where a low incidence of BKVN is identified[29]. Screening for decoy cells in the urine first may be a cheaper option[33], but there is a high prevalence rate of BK viruria even in immunocompetent adults and not all centres have resources to perform this test. Though cost saving can be achieved by reductions of immunosuppression as described in the literature[38,39], they may not necessarily cover the cost of screening for BKV if immunosup­pressants are inexpensive. To reduce the financial burden by increasing the monitoring interval to every 3 mo or longer, it may not reduce the incidence of BKVN as seen in the other studies. Nevertheless, either screening approach may not necessarily preserve graft function for those with BKVN. It is also interesting to see that the group with unknown BKV status in this cohort had a similar median eGFR comparing to those without BK viraemia, thus questioning the necessity of screening in this context. Our study is likely under-powered and increasing differences among these groups would likely have been observed in a large sample size. Targeting only those recipients who have significant predisposing risk factors could potentially reduce screening cost, but there is no consistency in what these risk factors are in the literature, thereby failing to identify BKVN cases early. Another cost saving option would be to identify recipients with positive BK viraemia early by performing an intensive monthly BKV screening only in the first 3 mo of renal transplant when the degree of immunosuppression is the greatest followed by a 3 mo screening until 12 mo after transplant. Therefore, when designing a BKV surveillance programme, it should be centre specific by taking the epidemiology of BKV infection, immunosuppression and monitoring protocols and related costs in a transplant centre into consideration to make it viable and cost effective.

There are several limitations in this retrospective study. Due to the lack of a comprehensive screening program we may have underestimated the incidence of BKVN. With our current approach of undertaking renal biopsy in all patients with a significant decline in renal function, it is unlikely that there are many patients with clinically significant BKVN that are not recognised. Interestingly, three patients with BKVLs of more than 160000 copies/mL did not have transplant biopsies due to stable graft function. Because of the selected cohort, the sizes of the comparative groups were small and there were only 9 recipients with BKVN which allow only a small number of variables to be included in the multivariable Cox regression model. As a result, the study is likely under-powered. In addition, there might be other risk factors for BKVN that were not identified, and tacrolimus and ciclosporin levels were not included in this study. Due to a large number of the recipients not tested for BKV, we cannot make effective comparisons, evaluate risk factors for BK viraemia or perform a cost analysis in this cohort.

A comprehensive BK virus surveillance program and reduction of immunosuppressive therapy is the reco­gnised management strategy to reduce the risk of BKVN occurring, because BKVN significantly impairs graft function. This study highlights that in our cohort the incidence of BKVN and graft failure due to BKVN without a formal screening programme is low and comparable to some transplant centres that have a BKV surveillance programme. Long CIT is associated with BKVN. The risk factor for BKVN identified is not consistent with other studies suggesting intricacy in the pathogenesis of BKVN and different protocols adopted by various transplant centres. Though the outcomes of this study remain speculative particularly the incidence of BKVN due to the study’s limitations and it requires further validation in larger trials, it provides a similar perspective in BK virus screening to Kiberd’s and Smith’s studies[29,38]. Transplant centres should evaluate its immunosuppression and monitoring protocols, the epidemiology of BKV infection and related costs before designing a BKV surveillance programme to make it centre specific and cost effective.

COMMENTS

Background

Screening for BK viraemia is an important strategy in reducing the risk of BK polyomavirus nephropathy (BKVN) and requires monthly monitoring in the first year of transplant in order for a BK virus surveillance to be effective. Applying this surveillance strategy in any transplant centres may not necessarily produce the best possible outcomes due to resource constraints and a low incidence of BK virus infection. This retrospective study was performed to compare outcomes of BK virus infection in the Auckland region without a formal BK virus screening program with those in centres with a BK virus surveillance program reported in literature.

Research frontiers

There are many risk factors, such as immunosuppressive burden, human leukocyte antigen mismatch, implicated in BK virus infection and the development of BKVN in renal transplant recipients. Therefore, immunosuppression and monitoring protocols play a role in BK virus infection. When designing a surveillance program for BK virus, the authors feel that immunosuppression and monitoring protocols, the epidemiology of BK virus infection and related costs should also be evaluated.

Innovations and breakthroughs

Outcomes of BKV infection, particularly BKVN, in this studied cohort are similar to some of transplant centres with a formal BK screening program that has less frequent testing for BKV.

Applications

Screening for BKV infection is important. Some transplant centres with limited resources may not afford frequent BKV testing or have capacity to perform BKV test. The study results suggest that a BKV screening program should be centre specific to be cost effective and achieve best possible outcomes.

Peer-review

This retrospective study looked at the incidence of BK viremia, BK nephropathy and graft outcomes among kidney transplant recipients in Auckland region. Study is well conducted and written clearly.

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Figure Legends

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**Figure 1 Description of the study population from January 2006 to December 2012 with follow-up until December 2013.** BKV: BK polyomavirus; BKVN: BK polyomavirus nephropathy.

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**Figure 2 Comparison of unadjusted graft function (median estimated glomerular filtration rate mL/min per 1.73 m2) taken before the censored date among BK polyomavirus status groups, excluding those who developed graft failure.** BKV: BK polyomavirus; eGFR: Estimated glomerular filtration rate.

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**Figure 3 Kaplan-Meier plot showing graft failure rates comparing recipients with/without BK polyomavirus nephropathy censored for death without graft failure and recipients transferred.** BKVN: BK polyomavirus nephropathy.

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**Figure 4 Kaplan-Meier plot showing graft failure rates comparing renal transplant recipients with different BK polyomavirus status censored for death without graft failure and recipients transferred.** BKV: BK polyomavirus; BKVN: BK polyomavirus nephropathy.

Footnotes

Institutional review board statement: This retrospective review was approved by the Northern X Regional Ethics Com­mittee (NTX/EXP).

Informed consent statement:Patients were not required to give informed consent to the study because the analysis used anonymous, de-identified clinical data.

Conflict-of-interest statement: There are no conflicts of interest in the publication of this paper.

Data sharing statement: The original de-identified dataset is available on request from the corresponding author at chunyuan.hsiao@middlemore.co.nz.

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**Table 1 Demographics and clinical characteristics of renal transplant recipients with/without BK polyomavirus nephropathy and comparison between those with BK polyomavirus nephropathy and BK viraemia *n* (%)**

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
|  | (-) BKVN (*n* = 217) | (+) BKVN (*n* = 9) | *P* value | (+) BKV (*n* = 19) | *P* value |
| Age (yr) | 48 (35.3-56.8) | 55 (34.9-60.5) | NS | 50 (27.1-60) | NS |
| Gender (female) | 92 (42.3) | 3 (33.3) | NS | 6 (31.6) | NS |
| Ethnicity |  |  |  |  |  |
| European | 131 (98.5) | 2 (1.5) | 0.038 | 11 (57.9) | 0.191 |
| Asian | 32 (90.7) | 3 (9.3) |  | 4 (21.1) |  |
| Māori and Pacific Islander | 54 (92.6) | 4 (7.4) |  | 4 (21.1) |  |
| Cause of ESKF |  |  |  |  |  |
| Glomerulonephritis | 91 (41.9) | 5 (55.6) | NS | 8 (42.1) | NS |
| Diabetes mellitus | 26 (12.0) | 2 (22.2) |  | 2 (10.5) |  |
| Others | 55 (46.1) | 2 (22.2) |  | 9 (47.4) |  |
| Co-morbidity |  |  |  |  |  |
| Diabetes mellitus | 26 (11.9) | 2 (22.2) | NS | 2 (10.5) | NS |
| Hypertension | 130 (59.9) | 5 (55.5) | NS | 12 (63.2) | NS |
| Cardiac disease | 31 (14.2) | 2 (22.2) | NS | 3 (15.8) | NS |
| Dialysis vintage (yr) | 2 (0.2-5) | 5 (1-6) | NS | 2.5 (0-5) | NS |
| Donor source (deceased) | 118 (54.3) | 9 (100) | 0.005 | 9 (47.4) | 0.01 |
| HLA mismatch | 3 (2-4) | 4 (2.5-4) | NS | 3.5 (2-5) | NS |
| Basiliximab induction | 95 (43.7) | 3 (33.3) | NS | 13 (68.4) | NS |
| Tacrolimus | 92 (42.3) | 5 (55.6) | NS | 10 (52.6) | NS |
| Cold ischaemia time (h) | 7 (4-15) | 19 (14.2-20.4) | 0.001 | 5.4 (4-12.5) | < 0.0001 |
| AR at any time point |  |  |  |  |  |
| 0 episode | 144 (66.4) | 3 (33.3) | 0.069 | 9 (47.4) | NS |
| ≥ 1 episode(s) | 73 (33.6%) | 6 (66.7) |  | 10 (52.6) |  |
| AR before known BKV/BKVN status |  |  |  |  |  |
| 10 → 0 episode | - | 3 (33.3) |  | 10 (52.6) | NS |
| 11 → ≥ 1 episode(s) | - | 6 (66.7) |  | 9 (47.6) |  |
| Thymoglobulin for acute rejection | 33 (15.2) | 0 (0) | NS | 5 (26.3) | NS |

Values are expressed as numbers (percentages) and medians (interquartile range Q1-Q3). *P* value are calculated using non-parametric test and Fisher’s exact test, and actual values are shown if *P* < 0.1. BKVN: BK polyomavirus nephropathy; (+) BKV: Recipients with BK viraemia without biopsy-proven BKV nephropathy; NS: Not significant; AR: Acute rejection; ESKF: End-stage kidney failure; HLA: Human leukocyte antigen.

**Table 2 Univariable and multivariable Cox regression to assess potential risk factors for BK polyomavirus nephropathy**

|  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- |
|  | **Crude HR (+) BKVN** | **95%CI** | ***P* value** | **Adjusted HR (+) BKVN** | **95%CI** | ***P* value** |
| Gender (female) | 0.66 | 0.16-2.66 | NS |  |  |  |
| Ethnicity (reference: European) |  |  |  |  |  |  |
| Asian | 6.36 | 1.06-38.16 | 0.043 | 3.731 | 0.61-22.92 | 0.154 |
| Māori/Pacific Islander | 4.75 | 0.87-25.94 | 0.072 | 2.631 | 0.45-15.25 | 0.279 |
| Co-morbidity |  |  |  |  |  |  |
| Diabetes mellitus | 2.06 | 0.42-9.94 | NS |  |  |  |
| Basiliximab induction | 0.75 | 0.18-3.03 | NS |  |  |  |
| Tacrolimus | 1.64 | 0.44-6.11 | NS |  |  |  |
| Thymoglobulin for rejection | 0.03 | 0.00-88.65 | NS |  |  |  |
| Cold ischaemia time | 1.18 | 1.06-1.32 | 0.003 | 1.181 | 1.04-1.35 | 0.009 |
| Acute rejection (≥ 1 episode)2 | 3.72 | 0.93-14.91 | 0.063 | 4.051 | 0.99-16.53 | 0.051 |
| HLA mismatch | 1.15 | 0.75-1.77 | NS |  |  |  |

1Variables included in the multivariable Cox regression model; 2Acute rejection at any time point. HR: Hazard ratio; CI: Confidence interval; (+) BKVN: BK polyomavirus nephropathy nephropathy; HLA: Human leukocyte antigen; NS: No significance.

**Table 3 Clinical outcome of the renal transplant recipients with BK polyomavirus nephropathy after a reduction of immu­nosuppressive therapy**

|  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- |
| **Recipients with BKVN** | **BKVL at diagnosis** | **eGFR at 1 mo** | **eGFR at CD** | **BKVL outcome1** | **Time to outcome1** | **Acute rejection (post reduction of IS)** |
| 1 | 1100000 | 48.7 | 73 | Undetectable | 28 mo | No |
| 2 | 3250 | 17.8 | 22 | Undetectable | 44 mo | Yes (at 30 mo) |
| 3 | 364700 | 36.9 | 23 | Undetectable | 31 mo | Yes (at 2 mo) |
| 4 | 265650 | 21.5 | 59 | Undetectable | 17 mo | No |
| 5 | 47225 | 23 | 18 | < 625 | 7 mo | No |
| 6 | 1794502 | 57 | 35 | 3725 | 59 mo | No |
| 7 | 59736502,3 | 33.4 | 20 | 41175 | 12 mo | No |
| 8 | 295608752,3,4 | 50.2 | 16 | 30425 | 8 mo | No |
| 9 | 56125 | 27.9 | Graft failure | Graft failure | 21 mo | No |

1Follow-up censored at December 2013; 2Use of Leflunomide; 3Use of Ciprofloxacin; 4Use of intravenous Immunoglobulin. BKVL: BK viral load (copies/mL); BKVN: BKV Nephropathy; IS: Immunosuppressant; CD: Censored date; eGFR: Estimated glomerular filtration rate (mL/min per 1.73 m2).