**Status of autoimmune diabetes 20-years after generation of**

**BDC2.5-TCR transgenic NOD mouse**

Lourdes Ramirez and Abdel Rahim A.R. Hamad

Department of Pathology, Johns Hopkins University School of Medicine, Ross 664G, 720 Rutland Ave, Baltimore, MD 21205

Address correspondence to Abdel Rahim A. Hamad at [ahamad@jhmi.edu](mailto:ahamad@jhmi.edu), phone 410-614-3021; Fax 410-614-3548

Key words: autoimmune diabetes; immunotherapy; T cells; BDC2.5 T cells, anti-CD3, immunosuppression.

Running title: Lessons from BDC2.5 TCR-Transgenic NOD mouse

**Core tip**

Our understanding of type 1 diabetes pathogenesis has significantly improved over the last three decades. We went from not knowing very little to acquisition of significant details about the role of the immune system and different T cell subsets in the disease process. The NOD mouse model contributed and continues to contribute to our understanding of the disease process. This article pays tributes to the major role BDC2.5 TCR transgenic mouse played in shaping of our understanding of the disease process. We also divulge to briefly discuss current challenges facing development of a safe immunotherapy for the disease.

**Abstract**

Type 1 diabetes (T1D) is an autoimmune disease that results from the destruction of insulin-producing β cells by autoreactive T cells, leading to lifelong dependency on insulin therapy and increased risk of long-term cardiovascular complications. Here we take the opportunity of the 20th anniversary of the generation of the BDC2.5 TCR transgenic NOD (non-obese diabetic) mouse model, to provide a brief overview of the significant progress that has been made during the last 20 years in our understanding of the role of T cells in the disease pathogenesis that included development of hundreds of reagents that block or even reverse new-onset disease by directly or indirectly controlling T cells. In the last part, we reflect on the sobering fact that none of these strategies has shown significant efficacy in clinical trials and discuss potential reasons hindering translation of the preclinical findings into successful therapeutic strategies and potential ways forward.

Diabetes is a heterogeneous metabolic disease caused by glucose intolerance and manifested clinically as hyperglycemia. Based on the underlying cause of the hyperglycemia, diabetes is divided into type 1 (T1D) and 2 (T2D). Type 1 diabetes is autoimmune in nature and results from the destruction of insulin-producing β cells by autoreactive T cells, leading to insulin deficiency and dependency on exogenous insulin to maintain glucose homeostasis. In contrast, T2D is a complex metabolic disorder associated with insulin resistance in peripheral tissues. Currently, there is no cure for either type of diabetes. In the interim, T1D is managed by multiple daily injections of insulin, whereas T2D is controlled by medications that improve insulin sensitivity and/or reduce glucose production by the liver. Maintenance of glucose homeostasis, however, is challenging and most patients eventually develop fatal cardiovascular complications. Intensive efforts are therefore being directed toward development of cure or prevention strategies. Small animal models play profoundly important roles in these efforts, particularly in T1D research.

Small animal research in T1D began in earnest with the development and use of spontaneous and induced disease models in 1970s and 80s. Among several T1D models, the NOD (non-obese diabetic) mouse became the most commonly used and favorite model soon after its development about 33 years ago [1](#_ENREF_1). The value of the NOD mouse in understanding the disease mechanism increased exponentially in the late 80s and early 90s following development of technologies that allowed engineering of the genome to generate mice bearing particular transgenes or lacking specific molecules to interrogate their roles in the disease process [2](#_ENREF_2). Consequently, more than 250 different genetically modified NOD mice were produced and characterized (http://jaxmice.jax.org/findmice/index.html). Results of these efforts uncovered a wealth of information about the roles of various cell types and molecules in modulating T cells and established key cellular and molecular events in the disease process.

Of interest is that uncovering the role of T cells in autoimmune diabetes traversed several key steps that culminated in the generation of the NOD mouse bearing TCR transgenic T cells [reviewed in detail in by Haskins et. al [3](#_ENREF_3)]. Considerable evidence accumulated in the early 1990s indicating a central role for T cells in mediating T1D in mice. These included demonstration that the disease development can be prevented by immunosuppressive agents that target T cells [4](#_ENREF_4), and by anti-CD4 and anti-CD8 antibody treatments [5](#_ENREF_5), [6](#_ENREF_6). Furthermore, the disease was shown to be transferrable to neonatal NOD mice and immunodeficient NOD.SCID (severe combined immunodeficiency mice) by adoptive transfer of T cells from spontaneously diabetic NOD donors [7](#_ENREF_7). A clearer picture of the role of T cells began to emerge with the generation of islet antigen-specific T cell clones. Several groups independently generated islet antigen-specific T cell clones capable of transferring the disease to susceptible recipients [4](#_ENREF_4). It was found that different T cell clones expressed different TCRs, suggesting for the first time that islet-specific T cells recognize several different islet antigens and pointing to the complexity of the disease. Among the well-characterized clones is the BDC2.5 clone, the T-cell receptor (TCR) that was later used to generate the BDC2.5 TCR tg mouse in 1993 [8](#_ENREF_8). Thus, generation of T cell clones was crucial in cementing the role of T cell in the disease pathogenesis and the existence of diabetogenic T cells in autoimmune-prone hosts. Yet clones have limited value in providing details regarding the nature and in vivo action mechanisms of diabetogenic T cells. Among the pressing questions (some of which are still incompletely understood) are how autoreactive T cells escape negative selection, where they reside in the periphery, what triggers them to become diabetogenic, and how they cause the disease. Diabetogenic T cells among the peripheral T cell repertoire are rare and the lack of appropriate reagents that permit their identification in vivo precluded addressing these questions directly in vivo in unmanipulated NOD mice. To overcome this problem, researchers generated TCR tg mice by using TCRs derived from generated clones. Among the widely used TCR transgenic mice in autoimmune diabetes is the BDC2.5 TCR tg mouse generated in 1993 by Katz et al. [8](#_ENREF_8)**,** in which all T cells express the TCRα (Vα1) and β (Vβ4) chain genes from the BDC2.5 TCR CD4 T cell clone [9](#_ENREF_9). Unlike in wild type NOD mice, which harbor a diverse repertoire where autoreactive T cells are very rare and are difficult to track in vivo, all T cells in BDC2.5 tg mice recognize and respond uniformly to an elusive islet autoantigen [It was recently reported by two groups [10](#_ENREF_10), [11](#_ENREF_11) that BDC2.5 T cells recognize peptides from chromogranin A (ChgA)].Therefore, by studying T cells in BDC2.5 tg mice, the authors were able to track the behavior and fate of diabetogenic T cells in vivo and test hypotheses pertaining to roles of thymic selection, site of priming and peripheral activation of diabetogenic T cells, trafficking, and timing of response to islet autoantigens. Results showed that diabetogenic TCR can be produced in a large proportion of thymocytes in the TCR αβ tg mice, are positively selected without undergoing massive clonal deletion, and migrate to the periphery where they constitute the majority of the T cell repertoire. The model is still providing an important platform for in vivo dissecting of diabetogenic T cells, including roles of various molecules and cell types in modulating their pathogenicity. It has not only resulted in a wealth of information regarding pathogenesis of autoimmune diabetes, but also shed light on the immune system and autoimmunity.

Tracking disease development in BDC2.5 TCR tg mice showed that initiation of the disease is highly regulated with two important checkpoints controlling the diabetogenic process. These two checkpoints are especially evident and synchronous in BDC2.5 tg mice. The autoreactive T cells appear to ignore the β cells for the first 2 weeks of life. Soon after, BDC2.5 T cells abruptly invade the pancreatic islets resulting in insulitis that progresses rapidly, with almost all islets heavily infiltrated by the age of 3 to 4 weeks. Surprising at the time, however, was the observation that insulitis in most BDC2.5 tg mice never progresses to full-blown diabetes. But when the BDC2.5 transgene is introduced into NOD-Rag-1 knockout mice, they do develop aggressive disease at a very early age. Failure of BDC2.5 TCR tg mice to develop full-blown disease in Rag-1-sufficient background was due to incomplete allelic exclusion of endogenous TCRβ chains, resulting in developing thymocytes that differentiate into regulatory T cells that oppose the pathogenic effect of diabetogenic T cells leading to standstill insulitis. On the other hand, in the absence of the Rag-1 gene all developing T cells bear the BDC2.5 TCR transgene, resulting in a pathogenic repertoire devoid of regulatory cells, inducing a rapid onset of aggressive disease. The results provide critical hints of a major role for regulatory T cells in opposing the disease development. The synchronous development of the disease in BDC2.5 mice combined with other studies, including adoptive transfer of BDC2.5 T cells, led to the concept that immunoregulatory mechanisms exist at two check points, at the pancreatic draining lymph nodes (PLNs) and the islet itself, respectively. Breach of these checkpoints by diabetogenic T cells is clearly visualized in NOD mice by using adoptive transfer of BDC2.5 in appropriate hosts [12](#_ENREF_12), [13](#_ENREF_13). This paradigm is depicted in **Figure 1**. Subsequent studies revealed critical roles for regulatory T and B cells and various molecules involved in controlling the major checkpoints, and prevention and cure of the disease in the NOD mouse. Over the last two decades, vast numbers of molecules necessary for maintaining immunoregulatory mechanisms and others that facilitate their subversion have been identified. Targeting these molecules identified more than 250 interventions capable of preventing the disease in the NOD mouse. Some, like treatment with anti-CD3 [14](#_ENREF_14) and anti-CD20 [15](#_ENREF_15) reversed the disease in as many as 30 to 50% of new-onset cases,raising hope of developing strategies to reverse disease in newly diabetic patients.Consequently, in the last few years, clinical trials have been conducted to test efficacy of several molecules including anti-CD3 and anti-CD20.

**Sobering reality facing translation of preclinical data into effective immunotherapeutics and ways forward:**

Translating immunotherapies found effective in preclinical studies into human therapies is proving challenging [16](#_ENREF_16), at least for now. Several high profile clinical trials including phase III have failed to demonstrate significant efficacy for all those tested [17](#_ENREF_17), [18](#_ENREF_18). The disappointing results in the clinic are forcing a retreat to drawing boards and generating second thoughts about whether the NOD mouse has surpassed its life expectancy as a research model and even the value of NOD mice in predicting and evaluating immunotherapy for T1D. It is easy to lay the blame on biologic differences between humans and mice, accentuated by more than 60 million years since their divergence into two species that differ in size, lifespan**,** and lifestyle (habitat / environment). The immune system in humans and mice, however, are generally quite similar, and with few notable exceptions, most paradigms translate well between them. Thus, the intangible efficacy of modalities such as anti-CD3 in humans is not entirelyjustified by biologic differences between the two species.

We argue that environmental factors play a dominant, if not the dominant role, in subverting therapeutic efficacy of modulators acting alone or in synergy with genetic factors [19](#_ENREF_19), [20](#_ENREF_20). This is acutely evident in the NOD mouse itself. For instance, the variability of anti-CD3 efficacy in reversing new-onset hyperglycemia ranges from about 30 to 80% in newly diabetic NOD mice housed in the same facility [14](#_ENREF_14), [21](#_ENREF_21) and mostly likely mice in the same cage responded differently. The low efficacy in NOD mice given the extremely small variations in their genetic makeup and exogenous influence of the environment suggests that treating the same mice under virtually identical conditions, the treatment would be successful only once out of at least two attempts. Applying the comparison to patients with markedly different genetic backgrounds, types of food, environment, and microbiota, the odds of success would be extremely low. Therefore, there is still much to be learned in the NOD mouse to uncover causes of variability on rate of disease onset, timing and response to treatment. In addition, understanding why females are more susceptible to disease than males[22-24](#_ENREF_22) and why NOD mice housed in conventional facilities do not develop disease remains unclear [25](#_ENREF_25). It will also be important to understand why inactivation of molecules such as Fas death receptor or its ligand prevents disease in NOD mice [13](#_ENREF_13), [16](#_ENREF_16), [26-29](#_ENREF_26). Understanding mechanisms underlying these observations would provide important clues that could potentially facilitate the development of therapeutic strategies with high efficacy rates that are effective in both mice and men.

**Acknowledgement**

The authors acknowledge support of our diabetes research by grants from the NIH (1R56AI099027 and 1R01AI099027-01) and American Heart Association (10GRNT4200003). We apologize for not having the opportunity to cite all seminal work describing islet reactive clones and transgenic mice expressing islet antigens.

**Figures legend**

**Figure 1.** Briefly, islet autoantigens are picked up by antigen present cells (APC) from the pancreas, which then migrate to draining lymph nodes (PLN) and present the autoantigens to autoreactive T cells, leading to their priming. Activated autoreactive T cells undergo proliferation, differentiation, and acquire homing molecules that allow them to direct to the pancreas, and infiltrate the islets resulting in insulitis and β cell destruction.

**References**

1. **Makino S**, Kunimoto K, Muraoka Y, Mizushima Y, Katagiri K, Tochino Y: Breeding of a non-obese, diabetic strain of mice, Jikken dobutsu Experimental animals 1980, 29:1-13

2. **Anderson MS**, Bluestone JA: The NOD mouse: a model of immune dysregulation, Annu Rev Immunol 2005, 23:447-485

3. **Haskins K**: Pathogenic T-cell clones in autoimmune diabetes: more lessons from the NOD mouse, Adv Immunol 2005, 87:123-162

4. **Haskins K**, Wegmann D: Diabetogenic T-cell clones, Diabetes 1996, 45:1299-1305

5. **Miller BJ**, Appel MC, O'Neil JJ, Wicker LS: Both the Lyt-2+ and L3T4+ T cell subsets are required for the transfer of diabetes in nonobese diabetic mice, J Immunol 1988, 140:52-58

6. **Phillips JM**, Harach SZ, Parish NM, Fehervari Z, Haskins K, Cooke A: Nondepleting anti-CD4 has an immediate action on diabetogenic effector cells, halting their destruction of pancreatic beta cells, J Immunol 2000, 165:1949-1955

7. **Bendelac A**, Carnaud C, Boitard C, Bach JF: Syngeneic transfer of autoimmune diabetes from diabetic NOD mice to healthy neonates. Requirement for both L3T4+ and Lyt-2+ T cells, J Exp Med 1987, 166:823-832

8. **Katz JD**, Wang B, Haskins K, Benoist C, Mathis D: Following a diabetogenic T cell from genesis through pathogenesis, Cell 1993, 74:1089-1100

9. **Haskins K**, Portas M, Bradley B, Wegmann D, Lafferty K: T-lymphocyte clone specific for pancreatic islet antigen, Diabetes 1988, 37:1444-1448

10. **Nikoopour E**, Sandrock C, Huszarik K, Krougly O, Lee-Chan E, Masteller EL, Bluestone JA, Singh B: Cutting edge: vasostatin-1-derived peptide ChgA29-42 is an antigenic epitope of diabetogenic BDC2.5 T cells in nonobese diabetic mice, J Immunol 2011, 186:3831-3835

11. **Stadinski BD**, Delong T, Reisdorph N, Reisdorph R, Powell RL, Armstrong M, Piganelli JD, Barbour G, Bradley B, Crawford F, Marrack P, Mahata SK, Kappler JW, Haskins K: Chromogranin A is an autoantigen in type 1 diabetes, Nat Immunol 2010, 11:225-231

12. **Andre I**, Gonzalez A, Wang B, Katz J, Benoist C, Mathis D: Checkpoints in the progression of autoimmune disease: lessons from diabetes models, Proc Natl Acad Sci U S A 1996, 93:2260-2263

13. **Mohamood AS**, Guler ML, Xiao Z, Zheng D, Hess A, Wang Y, Yagita H, Schneck JP, Hamad AR: Protection from autoimmune diabetes and T-cell lymphoproliferation induced by FasL mutation are differentially regulated and can be uncoupled pharmacologically, Am J Pathol 2007, 171:97-106

14. **Chatenoud L**, Thervet E, Primo J, Bach JF: Anti-CD3 antibody induces long-term remission of overt autoimmunity in nonobese diabetic mice, Proc Natl Acad Sci U S A 1994, 91:123-127

15. **Hu CY**, Rodriguez-Pinto D, Du W, Ahuja A, Henegariu O, Wong FS, Shlomchik MJ, Wen L: Treatment with CD20-specific antibody prevents and reverses autoimmune diabetes in mice, J Clin Invest 2007, 117:3857-3867

16. **Gutfreund R**, Hamad AR: Immunotherapy for Type 1 Diabetes: Necessity, Challenges and Unconventional Opportunities. Edited by Wager D. Janeza Trdine 9, INTECH, 2011, p. pp. 409-424

17. **Staeva TP**, Chatenoud L, Insel R, Atkinson MA: Recent lessons learned from prevention and recent-onset type 1 diabetes immunotherapy trials, Diabetes 2013, 62:9-17

18. **Michels AW**, von Herrath M: 2011 Update: antigen-specific therapy in type 1 diabetes, Curr Opin Endocrinol Diabetes Obes 2011, 18:235-240

19. **Wicker LS**, Todd JA, Peterson LB: Genetic control of autoimmune diabetes in the NOD mouse, Annu Rev Immunol 1995, 13:179-200

20. **Chervonsky A**: Innate receptors and microbes in induction of autoimmunity, Curr Opin Immunol 2009, 21:641-647

21. **Bresson D**, Togher L, Rodrigo E, Chen Y, Bluestone JA, Herold KC, von Herrath M: Anti-CD3 and nasal proinsulin combination therapy enhances remission from recent-onset autoimmune diabetes by inducing Tregs, J Clin Invest 2006, 116:1371-1381

22. **Ivakine EA**, Fox CJ, Paterson AD, Mortin-Toth SM, Canty A, Walton DS, Aleksa K, Ito S, Danska JS: Sex-specific effect of insulin-dependent diabetes 4 on regulation of diabetes pathogenesis in the nonobese diabetic mouse, J Immunol 2005, 174:7129-7140

23. **Ivakine EA**, Mortin-Toth SM, Gulban OM, Valova A, Canty A, Scott C, Danska JS: The idd4 locus displays sex-specific epistatic effects on type 1 diabetes susceptibility in nonobese diabetic mice, Diabetes 2006, 55:3611-3619

24. **Markle JG**, Frank DN, Mortin-Toth S, Robertson CE, Feazel LM, Rolle-Kampczyk U, von Bergen M, McCoy KD, Macpherson AJ, Danska JS: Sex differences in the gut microbiome drive hormone-dependent regulation of autoimmunity, Science 2013, 339:1084-1088

25. **Wen L**, Ley RE, Volchkov PY, Stranges PB, Avanesyan L, Stonebraker AC, Hu C, Wong FS, Szot GL, Bluestone JA, Gordon JI, Chervonsky AV: Innate immunity and intestinal microbiota in the development of Type 1 diabetes, Nature 2008, 455:1109-1113

26. **Brunner T**, Mog R, LaFace D, Yoo N, Mahboubl A, Echeverri F, Martin S, Force W, Lynch D, Ware C, Green D: Cell autonomous Fas (CD95) / Fas ligand interaction mediates activation induced apoptosis in T cell hybridomas, Nature 1995, 373:441-444

27. **Desbarats J**, Duke RC, Newell MK: Newly discovered role for Fas ligand in the cell-cycle arrest of CD4+ T cells, Nat Med 1998, 4:1377-1382

28. **Atkinson MA**, Bluestone JA, Eisenbarth GS, Hebrok M, Herold KC, Accili D, Pietropaolo M, Arvan PR, Von Herrath M, Markel DS, Rhodes CJ: How does type 1 diabetes develop?: the notion of homicide or beta-cell suicide revisited, Diabetes 2011, 60:1370-1379

29. **von Herrath M**: Combination therapies for type 1 diabetes: why not now?, Immunotherapy 2010, 2:289-291