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## *Retrospective Study*

## Ketogenic diet imposes significant effect on imbalanced gut microbiota in infants with refractory epilepsy

Gan X *et al.* Gut microbiota altered by ketogenic diet on epilepsy

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***AIM***

To investigate if patients with refractory epilepsy and healthy infants differed in gut microbiota (GM), and how ketogenic diet (KD) alter GM.

***METHODS***

A total of 14 epileptic and 30 healthy infants were recruited and seizure frequencies were recorded. Stool samples were collected for 16S rDNA sequencing using Illumina Miseq platform. The composition of GM in each sample was analyzed by MOTHUR, and inter-group comparison was conducted by R software.

***RESULTS***

After being on KD treatment for a week, 64% of epileptic infants showed an obvious improvement, with a 50% decrease in seizure frequency. GM structure in epileptic infants (P1 group) differed dramatically from that in healthy infants (Health group). Proteobacteria, which had accumulated significantly in P1 group, decreased dramatically after KD treatment (P2 group). *Cronobacter* was dominated in P1 group and remained at a low level both in Health and P2 group. *Bacteroides* increased significantly in P2 group, in whom *Prevotella* and *Bifidobacterium* also grew in numbers and kept continued to increase.

***CONCLUSION***

GM pattern in healthy infants differed dramatically from that of epileptic group. KD could significantly modify symptoms of epilepsy and reshape the GM of epileptic infants.

**Key words:** Ketogenic diet; Epilepsy; Seizures; Gut microbiota; *Cronobacter*

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**Core tip:** Many infants with epilepsy are refractory to current antiepileptic drugs, and ketogenic diet (KD) could help to moderate seizures frequency as an alternative treatment. A large number of reports have demonstrated that gut microbiota (GM) can affect children’s neurodevelopment. Concurrently, GM could be dramatically affected by diet. KD could rapidly alter GM and alleviate seizure frequency in infants with refractory epilepsy. The GM structure of epileptic infants—comprising large numbers of pathogens, such as *Streptococcus*-differs from that of healthy controls. After KD therapy, GM of epileptic patients changed significantly, with fewer pathogens and more beneficial bacteria.

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**INTRODUCTION**

Pediatric epilepsy is widespread, with complications including cognitive impairment, delayed neurodevelopment and loss of bodily control[1,2]. Disequilibrium between excitation and depression of the central nervous system is acknowledged as the main factor in epilepsy incidence[3]. Prior reports have identified increased inflammatory reactions and pro-inflammatory cytokines, such as interleukin-6 (IL-6), interleukin-17 (IL-17) and interferon, in the cerebrospinal fluid (CSF)[4]. AEDs and surgery are the main conventional treatments for infants with epilepsy[5]. However, there are still 30% of epileptic infants who suffered from therapeutic futility and recurrent attacks.

A growing number of reports indicated that KD was a promising therapeutic alternative for infants with refractory epilepsy, as it has been shown to ameliorate their clinical symptoms, including the frequency of seizures[6-10]. It remains unclear exactly how this occurs. Several reports implicated changed neurotransmitters after KD therapy, including γ-aminobutyric acid (GABA), monoamines and glutamate[7,11]. Dahlin *et al*[12,13] also identified increased ketone bodies (KBs) and decreased dopamine as well as serotonin[12,13]. However, Sariego-Jamardo *et al*[14]found little change of neurotransmitters, pterins and amino acids in the CSF of KD responders as opposed to non-responders. These discrepant findings suggested a need for the further elucidation of the mechanisms of KD therapy.

A variety of findings shown that diet posed significant effect on GM[8,15]. A high-fat diet induced selective enrichment of bile-metabolizing microbiota, such as *Bacteroides*[16]. Whilst, high-fiber foods promoted the accumulation of plant-polysaccharide fermenting microbial organisms, including *Prevotella* and *Clostridium*[16]. A number of reports implicated involvement of GM in enteric nervous system (ENS), blood-brain barrier (BBB) and glial cell development, all of which were pivotal to behavioral control and cognitive progression[17,18]. GM could produce neurotransmitters and gut hormones directly[19] or indirectly by producing signaling molecules to regulate host cells[20]. GM-derived short-chain fatty acids (SCFAs) could stimulate enterochromaffin cells to produce serotonin[21]. Wikoff *et al*[22] also documented decreased serotonin in peripheral serum in the absence of GM. Moreover, *Clostridium sporogenes* and *Ruminococcus gnavus* promoted decarboxylation of tryptophan to tryptamine, which modulated mood and appetite through amine-associated receptors[23]. Based on the involvement of GM in the gut-brain axis, increasing reports demonstrated imbalanced GM in neurogenic diseases (NDs), including autism-spectrum disorder (ASD), Parkinson’s disease, and depression[24]. However, GM dysbiosis in childhood epilepsy remain unexplored.

Previous studies declared that short-term dietary intake could rapidly alter human GM[8,15]. In this study, we performed comparison between diseased infants (before and after KD treatment) and healthy controls, to explore if and how GM of infants with refractory epilepsy differed with age-matched healthy subjects. We also evaluated the therapeutic effect of KD on refractory epilepsy and the changes in GM after treatment. It is hoped that his research will help to bridge some gaps in the current understanding of refractory childhood epilepsy.

**MATERIALS AND METHODS**

***Sample collection***

We enrolled 14 pediatric patients with refractory epilepsy (aged 1.95 ± 3.10 years, 11 male and 3 female) in Shenzhen Children’s Hospital, according to the following inclusion criteria: convulsed more than four times per week after ≥ 3 AEDs treatment; no antibiotic exposure for at least 1 mo; no known genetic metabolic disorders or severe systemic illnesses; successive KD therapy for at least 1 wk. KD was provided by Zeneca, Shenzhen, China, including Qitong ketogenic liquid milk (3.4 g protein, 8.0 g lipid and 0.6 g carbohydrate per 100 g milk), Qitong ketogenic cookies and Qitong ketogenic set-meal packages[25].

Healthy subjects (aged up to 3 years, 15 male and 15 female) were also recruited based on the following criteria: no antibiotic exposure for at least 1 month before this study, no disease symptoms for at least 1 month following recruitment, and no history of seizures (Supplementary Table 1). Fisher’s test was used to evaluate the effect of gender and age on GM composition.

***DNA extraction, library construction and sequencing***

The genomic DNA of microbiota was extracted from stool samples using the Power Soil DNA Isolation Kit (Mo Bio Laboratories, Carlsbad) following the manufacture’s protocol. The hyper variable V3-V4 region of the 16S rRNA gene was amplified using PCR kit (TransGenAP221-02, Peking), and DNA products were quantified by gel electrophoresis and Qubit (Thermo Fisher, Singapore). After library construction, the qualified libraries were sequenced by Illumina MiSeq Sequencing platform (Illumina, San Diego).

***Taxonomy classification and diversity detection***

After filtration, overlapped paired reads were assembled as tags by FLASH (v1.2.11), and clustered to operational taxonomic units (OTUs) through USEARCH (v7.0.1090)[26]. Representative OTUs were mapped against the Greengenes database (v201305)[27] and classified by RDP classifier (v2.2)[28]. The diversity of microbiota was calculated by MOTHUR (v1.31.2)[29].

***Principal component analysis and statistical analysis***

### Principal component analysis (PCA) was performed by R software (v3.2.5). Wilcoxon rank-sum test was used to compare GM in diseased infants and healthy controls. Comparative analysis between the P1 and P2 group was conducted by Wilcoxon signed-rank test. Linear discriminant analysis Effect Size (LEfSe) analysis was used to identify microbial species, which were apparently enriched in specific group.

**RESULTS**

***Data output and patients’ characteristics***

High-quality 117196 sequencing reads were produced for each sample on average, ranging from 31900 to 305190. The number of assembled tags averaged 22800, with a range from 12655 to 27337. Both gender and age had no statistical significance for affecting GM, with *P*-value 0.069 and 0.234 respectively.

***GM of healthy*** ***individuals differed dramatically with that of diseased infants***

Shannon index analysis indicated higher GM diversity in healthy infants, by comparison with infants with refractory epilepsy (Figure 1, Supplementary Table 2). PCA analysis of GM profile also identified that healthy infants could be clearly distinguished from patients (Figure 2, Supplementary Table 3). Phylum Firmicutes was dominated in patients (45.82%) and was unchanged after KD therapy (47.00%) (Supplementary Table 4). Bacteroidetes accounted for 53.01% of GM in healthy infants, followed by Firmicutes (34.38%). After KD treatment, Bacteroidetes increased from 26.75% to 38.71%. Actinobacteria was enriched in healthy infants (8.49%) and occupied fewer percent in patients (2.38% when pre-treatment and 2.92% after treatment). Proteobacteria was highly accumulated in infants with refractory epilepsy (24.34%) and decreased dramatically after KD therapy (10.77%). At genus level, *Cronobacter* was dominant in the patients (23.30% *vs* 0.00% in healthy group). By contrast, healthy subjects harbored more than twice *Bacteroides* (42.68%) than that in infants with refractory epilepsy (17.93%). *Prevotella* and *Bifidobacterium* were also accumulated in healthy group (7.25%, 7.84% respectively) (Supplementary Table 5).

***KD therapy ameliorated epilepsy and GM of patients started to improve***

After a week of KD therapy, 3 patients (21%) were seizure-free and 6 (43%) patients had a 50% to 90% decrease of seizure frequency (Supplementary Table 1). The remaining 5 infants experienced no significant improvement in seizure control (36%) (Supplementary Table 1). GM of P2 group was more similar to that of Health group, by comparison with P1 group (Figure 3 and 4). After KD treatment, *Bacteroides* increased significantly, by 24.42%. *Prevotella* also increased in P1 group from 0.37% to 1.85% after KD treatment (Figure 3 and Supplementary Table 5). *Cronobacter* decreased sharply in after-treatment patients, from 23.3% to 10.44 % (Figure 3, Figure 4 and Supplementary Table 5). KD exposure also induced a decrease in *Erysipelatoclostridium* (by 8.67% in P1 and 4.89% in P2 group); it represented just 0.64% in healthy infants (Figure 3, Figure 4 and Supplementary Table 5). *Streptococcus*, *Alistipes*, *Ruminiclostridium*, *Barnesiella* and *Enterococcus* also decreased after KD therapy (Figure 3, Figure 4 and Supplementary Table 5).

**DISCUSSION**

KD is increasingly used for the treatment of refractory epilepsy in childhood, but the mechanism remains unclear. Previous reports indicated that GM played an important role in gut-brain-axis[24], and was affected significantly by intake of high-fat food[16]. This study focused on differed GM structure of healthy with epileptic infants, as well as alteration in the GM of patients after one week of KD treatment. The results pointed to an imbalanced GM in patients and a significant improvement after KD therapy.

Proteobacteria comprise a variety of notorious pathogens, such as *Escherichia*, *Salmonella* and *Vibrio*. It accounted for 24.34% in pediatric patients and decreased dramatically after KD treatment. Bacteroidetes was dominant in healthy infants and increased largely for after-treatment patients.

We identified accumulated *Bac**teroides* in healthy subjects as well as in patients after treatment. *Bacteroides* was reported to digest and metabolize high-fat food and to regulate the secretion of IL-6 and IL-17 in dendritic cells (DCs), a process strongly associated with seizure severity of epileptic patients[4,16]. However, patients-enriched *Cronobacter* decreased dramatically after KD therapy. Prior reports demonstrated that there were multiple virulence determinants of *Cronobacter*, including C*ronobacter*plasminogen activator and ferric ion transporter protein, which paly a detrimental role in human health[30-32]. *Prevotella* was robust producer of SCFAs[33], which could protect intestinal mucosa and function as neurotransmitters. Previous reports also indicated that SCFAs mediatednervous impulse and mitigated Parkinson’s disease[33,34]. Similarly, we identified increased *Prevotella* in Health and P2 group, when compared with P1 group. Some other genera also offer clues to epilepsy recovery, such as *Erysipelatoclostri**dium*, *Blautia, Bifidobacterium* and *Streptococcus*. *Bifidobacterium* was well known to be beneficial to health[35], and *Streptococcus,* a common pathogen, played a role especially in respiratory diseases[36]. Although GM imbalance in diseased infants was identified and GM improved after KD treatment, more exploration was needed to elucidate the contribution of a healthy GM to epilepsy onset/recovery.

This study revealed that KD can mitigate the symptoms of epilepsy and correct an imbalanced GM in epileptic infants. However, further analysis is needed to unravel how GM may be involved in epilepsy onset/recovery.

There are some limitations that need to be clarified. Firstly, 16S rDNA analysis identified microbes at genus level, which make it difficult to unravel different microbes at species or function level. Secondly, it would be more useful to evaluate the efficacy of KD treatment and its effect on the GM if this could be done with a longer period of follow-up. Thirdly, an animal model might be applied to demonstrate whether GM imbalance could induce epilepsy associated symptoms. Considering these limitations, we are planning to perform metagenomic analysis on GM of healthy and epileptic infants. This will provide more insights into distinct metabolic networks in imbalanced GM.

In this study, we found that GM of infants with refractory epilepsy differed dramatically from that of healthy infants. Concurrently, epileptic patients harbored significantly enriched pathogens and decreased beneficial bacteria. Although this study provides new insight into the involvement of GM in pediatric refractory epilepsy, the gap between KD and epilepsy recovery is still huge. To uncover the mechanism and pathogens involved refractory infantile epilepsy, further research should underscore functional gene networks in GM. In summary, this work lays a foundation for GM alteration of GM in epilepsy in order to promote the amelioration of this condition.

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**COMMENTS**

***Background***

Infants with refractory epilepsy couldn’t be cured by several AEDs and ketogenic diet (KD) was increasingly used as an alternative therapy to refractory epilepsy. High-fat diet was reported to pose significant impact on gut microbiota (GM), which could regulate neurological system.

***Research frontiers***

Previous reports demonstrated that GM could affect neural systems by secreting metabolites as neurotransmitters. In parallel, the gut-brain axis is a research hot spot in biomedicine, including the study of autism, Parkinson’s disease, and depression.

***Innovations and breakthroughs***

This study showed that the GM pattern of diseased infants differs significantly from that of the healthy. The decreased number of dominant pathogens and significantly increased number of beneficial bacteria after KD treatment offers new insight to KD therapy for epilepsy.

***Applications***

This study found several bacteria representing alterations in the GM; suggesting that these bacteria could be monitored as biomarkers, providing an important reference for epilepsy treatment.

***Terminology***

The GM-which consist of many kinds of bacteria including pathogens, commensals, and probiotics—play an important role in the human body**.**

***Peer-review***

The authors have performed important research in pediatric epilepsy. They discovered that the composition of the GM in health and disease was significantly different, specifically in healthy infants as opposed to those with refractory epilepsy. Bacterial patterns were dramatically changed after KD therapy, and this was associated with a reduction in the frequency of seizures. These findings should enhance our knowledge of the relationship between epilepsy and GM and provide new insight into the clinical treatment of epilepsy. However, environmental factors and clinical parameters should be studied more closely in further research.

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**Figure 1 Gut microbial diversity of three groups.** Distribution of Shannon index (evenness). Red, blue, and green represent the Health, P1, and P2 groups, respectively. The gut microbiota (GM) of the healthy infants was more stable than that of the other two groups.



**Figure 2 Principal components analysis.** Each plot in the Principal components analysis (PCA) graph stands for a sample. Red, blue and green colors represent Health, P1 and P2 group, respectively.



**Figure 3 Gut microbiota structure in the Health, P1, and P2 groups at the genus level.** SVG package (version 1.1) was used to produce the paragraph. The size of the circle representing each genus was determined by the relative abundance of the three groups, and the width of line linking the P1, P2, and Health groups indicates the relative abundance of each group.

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**Figure 4 Significantly enriched Gut microbiota components in the Health, P1, and P2 groups.** LEfSe analysis was applied to detect the gut microbiota (GM) components in the three groups. Red, green, and blue represent the Health, P1, and P2 groups, respectively. The LDA score was set as ≥ 2. The enrichment degree is proportional to the LDA score.