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***Basic Study***

**Prostaglandin E1 protects hepatocytes against endoplasmic reticulum stress-induced apoptosis *via* protein kinase A-dependent induction of** **glucose-regulated protein 78 expression**

Yang FW *et al*. PGE1 protects hepatocytes against ER stress-induced apoptosis

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**Abstract**

***AIM***

To investigate the protective effect of prostaglandin E1 (PGE1) against endoplasmic reticulum (ER) stress-induced hepatocyte apoptosis, and to explore its’ underlying mechanisms.

***METHODS***

Thapsigargin (TG) was used to induce ER stress in human hepatic cell line L02 and hepatocarcinoma derived cell-line HepG2. To evaluate the effects of PGE1 on TG-induced apoptosis, PGE1 was used an hour prior to TG treatment. Activation of unfolded protein response (UPR) signaling pathways were detected by western blotting and quantitative real-time RT-PCR. Apoptotic index and cell viability of L02 cells and HepG2 cells were determined with flow cytometry and MTS [3-(4,5-dimethylthiazol-2-yl)-5-(3-carboxymethoxyphenyl)-2-(4-sulfophenyl)-2H-tetrazolium] assay.

***RESULTS***

Pretreatment with 1 μmol/L PGE1 protected against TG-induced apoptosis in both L02 cells and HepG2 cells. PGE1 enhanced the TG-induced expression of C/EBP homologous protein (CHOP), glucose-regulated protein (GRP) 78 and spliced X box-binding protein 1 at 6 h. However, it attenuated their expressions after 24 h. PGE1 alone induced protein and mRNA expressions of GRP78; PGE1 also induced protein expression of DNA damage-inducible gene 34 and inhibited the expressions of phospho-PKR-like ER kinase, phospho-eukaryotic initiation factor 2α and CHOP. Treatment with protein kinase A (PKA)-inhibitor H89 or KT5720 blocked PGE1-induced upregulation of GRP78. Further, the cytoprotective effect of PGE1 on hepatocytes was not observed after blockade of GRP78 expression by H89 or siRNA specifically targeted against human GRP78.

***CONCLUSION***

Our study demonstrates that PGE1 protects against ER stress-induced hepatocyte apoptosis *via* PKA pathway-dependent induction of GRP78 expression.

**Key words**: Hepatocytes; Endoplasmic reticulum stress; Thapsigargin; Glucose-regulated protein 78; Protein kinase A; Apoptosis

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**Core tip**: The mechanism underlying the hepatoprotective effect of prostaglandin E1 (PGE1) remains unclear. In this study, we found that pretreatment with PGE1 protected hepatocytes against thapsigargin-induced apoptosis. PGE1 alone induced protein and mRNA expressions of glucose-regulated protein (GRP) 78. Treatment with protein kinase A (PKA)-inhibitor H89, KT5720 or siRNA specifically targeted against human GRP78 blocked PGE1-induced upregulation of GRP78. The hepatoprotective effect of PGE1 was lost by blocking GRP78 expression with either H89 or siRNA. Our study demonstrates for the first time that PGE1 protects against ER stress-induced hepatocyte apoptosis *via* PKA pathway-dependent induction of GRP78 expression.

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**INTRODUCTION**

Hepatocyte apoptosis can be triggered by intra- or extra-cellular signals[1]. The intracellular signals for hepatocyte apoptosis are induced by DNA damage, oxidative stress, growth factor deprivation, mitochondrial dysfunction, ATP depletion and endoplasmic reticulum (ER) stress[2,3]. A complex interaction occurs among these intracellular apoptotic signaling pathways. ER stress is known to induce hepatocyte apoptosis under various pathological conditions[4]. ER stress is implicated in the pathogenesis of various liver diseases such as obesity-associated fatty liver disease[5,6], viral hepatitis[7], alcohol-induced liver injury[8], drug-induced liver injury[9] and in ischemia/reperfusion injury of the liver[10,11]. Devising a treatment strategy to protect hepatocytes from ER stress-induced apoptosis will benefit most patients with liver diseases.

ER is a multifunctional intracellular organelle responsible for the synthesis, processing, and trafficking of proteins that are essential for cell growth and survival. ER also serves as a storage organelle for calcium[12]. When the homeostasis of ER is disturbed under various pathophysiological conditions, ER stress is induced and the unfolded protein response (UPR) is activated. The UPR activates three ER-transmembrane transducers, inositol-requiring enzyme (IRE) 1a, PKR-like ER kinase (PERK), and activating transcription factor (ATF) 6α[12-14]. Activation of these three UPR pathways enhances the ER’s protein folding *via* upregulation of the synthesis of glucose-regulated protein (GRP) 78. UPR signals also accelerate the degradation of misfolded proteins and reduce the synthesis of new proteins. Therefore, UPR during ER stress facilitates restoration of homeostasis. However, sustained or unresolved ER stress can activate a cascade of apoptotic signals that eventually result in cell death[15,16].

In several experimental models of liver injury, prostaglandin E1 (PGE1) has been shown to protect against hepatocyte apoptosis[17-19]. PGE1 is also effective in the treatment of patients with fulminant hepatitis and those with primary graft nonfunction after liver transplantation[20,21]. Thus, PGE1 appears to protect hepatocytes against apoptosis through various mechanisms[22-24]. However, the underlying mechanism of its hepatoprotective effect is not well understood.

In two recent studies, PGE1 was shown to induce expressions of heat-shock protein (HSP) and GRP78 in animal models of liver injury caused by ischemia reperfusion and hepatectomy[25,26]. These findings suggest that modulation of UPR may mediate the hepatoprotective effects of PGE1. However, the role of PGE1 in ER stress-induced apoptosis of hepatocytes is largely unknown.

In this study, we evaluated the protective effect of PGE1 against ER stress-induced hepatocyte apoptosis in both normal human hepatocyte cell line L02 and hepatocarcinoma derived cell-line HepG2 [27].

**MATERIALS AND METHODS**

***Chemical reagents***

RPMI 1640 was obtained from Thermo-Fisher Biochemical Products Co. Ltd (Beijing); fetal bovine serum (FBS), TG, protein kinase A (PKA) inhibitor H89 and KT5720, 4-phenylbutyric acid (PBA) and PGE1 were purchased from Sigma (St. Louis, MO, United States). Antibodies against GRP78, PERK, eukaryotic translation initiation factor-2α (eIF-2α), phospho-PERK (p-PERK) and phospho-eIF2α (p-eIF2α), C/EBP homologous protein (CHOP), spliced X box-binding protein1 (sXBP1), growth arrest and DNA damage-inducible gene34 (GADD34) and β-actin were purchased from Santa Cruz Biotechnology (Santa Cruz, CA, United States). Annexin V-FITC/PI apoptosis detection kit was purchased from Dojindo Laboratories (Kumamoto, Japan). siRNA scramble control and validated human GRP78-siRNA were purchased from Santa Cruz Biotechnology (Santa Cruz, CA, United States). All other chemicals and reagents were obtained from Sigma (St. Louis, MO, United States), unless stated otherwise.

***Cell culture***

Human hepatocyte cell line L02 and hepatocarcinoma derived cell line HepG2 were obtained from cell bank of the Type Culture Collection at the Chinese Academy of Sciences (Shanghai, China). L02 and HepG2 cells were propagated at 37 °C in 5% CO2 in RPMI 1640 medium containing 10% (v/v) FBS, 100 units/mL penicillin and were passaged every 5-7 d. L02 and HepG2 cells were cultured until they acquired 80%-100% confluence. Thereafter, the cells were rinsed three times with 10 mL of PBS and cultured in a medium lacking FBS for 24-36 h. To evaluate the effects of PGE1 or PBA on TG-induced apoptosis, PGE1 or PBA was used an hour prior to TG treatment and, thereafter, the medium was not changed. TG and H89 were dissolved in dimethyl sulfoxide (DMSO) at concentrations of 40 µmol/L and 20 µmol/L, respectively, as stock solution. PGE1 was dissolved in ethanol at a concentration of 2.82 mmol/L as stock solution. PBA was dissolved in RPMI 1640 medium. All control conditions included corresponding vehicles at the appropriate concentrations (ethanol for PGE1 and DMSO for TG and H89).

***Cell apoptosis analysis***

Apoptosis was determined using annexin V-FITC/PI apoptosis detection kit, according to the manufacturer’s instructions. Briefly, 2 × 106 cells were harvested using 0.05% trypsin with 0.5% mmol/L EDTA. To analyze the whole apoptotic cell population, nonadherent cells present in the culture medium were added to the harvested cells. The cells were then washed twice with pre-cold PBS and resuspended in 500 μL annexin binding buffer. 5 μL of annexin V-FITC and 5 μL of propidium iodide (PI) were added to each sample and incubated in dark at room temperature for 10 min. Flow cytometry (Beckman Coulter Gallios, United States) was performed according to the manufacturer’s specifications. The apoptotic index was calculated as the percentage of annexin V+ cells divided by the total number of cells in the gated region.

***Cell viability assay***

Cell viability was assessed with MTS [3-(4,5-dimethylthiazol-2-yl)-5-(3-carboxymethoxyphenyl)-2-(4-sulfophenyl)-2H-tetrazolium] method using the CellTiter 96® AQueous One Solution Cell Proliferation Assay kit (Promega Corporation, Madison, WI, United States) according to the manufacturer's instructions. In brief, early passage L02 or HepG2 cells were plated in triplicate on 96-well plates (10000 cells/well) and cultured in modified RPMI 1640 for 24 h. Thereafter, the cells were rinsed three times in 200 μL of PBS and cultured in a medium lacking FCS. Cell viability was determined by replacing the medium with 20 μL of MTS. After incubation of the cells at 37 °C for 3 h, the absorbance was measured at 490 nm using a microplate reader (Bio-Rad model 680; Bio-Rad, Hercules, CA, United States). Cell viability was normalized as a percentage of control. This experiment was performed five times.

***Western blotting***

Cell lysates containing 40 µg of protein were resolved by SDS-PAGE using 4%–20% polyacrylamide gradient gel, and the fractioned proteins were subsequently transferred to nitrocellulose membranes. After blocking with tris-based saline buffer containing 5% dry milk and 0.1% Tween 20 for 1 h, the membranes were blotted with the corresponding antibodies. The primary and secondary antibodies used were: rabbit anti-human GRP78 (1:500 dilution), PERK (1:1000 dilution), p-PERK (1:500 dilution), eIF-2α (1:500 dilution), p-eIF-2α (1:250 dilution), sXBP1 (1:500 dilution), CHOP (1:1000 dilution), GADD34 (1:1000 dilution), mouse anti-human β-actin, goat anti-rabbit IgG conjugated with horseradish peroxidase (HRP), and goat anti-mouse IgG conjugated with HRP. The membranes were developed using a chemiluminescence detection system and thereafter exposed to Kodak BioMax Light Film (Rochester, NY, United States). The band intensity for each protein was measured using Image-Pro Plus analysis software (MediaCybernetics, Silver Spring, MD, United States) and the expression normalized to that of β-actin.

***Quantitative real-time RT-PCR***

Total RNA was isolated from L02 cells using TriZol reagent (Invitrogen, CA, United States) according to the manufacturer’s instructions. RNase-free DNase I (TaKaRa, Dalian, China) was used before cDNA synthesis to remove any genomic contamination. A total of 1 μg RNA from each sample was used for cDNA synthesis with a reverse transcription kit (TaKaRa, Dalian, China). Reverse transcription (final volume of 20 μL) was performed at 42 °C for 10 min, followed by 75 °C for 5 min. Real-time PCR reaction (25 μL) containing 12.5 μL of 2 × SYBR Premix Ex Taq II (Tli RNaseH Plus) (TaKaRa, Dalian, China), 1 μL of each 10 μmol/L primers, 2 μL of cDNA template, and 8.5 μL RNAase-DNAse free water was performed on a CFX96 PCR system (Bio-Rad, Hercules, CA, United States). The reaction process was as follows: denaturation at 95 °C for 3 min, followed by 40 cycles of amplification (95 °C for 10 s and 60 °C for 30 s), ending with a melt curve ranging from 60 °C to 95 °C with a heating rate of 0.3 °C/15 s. All samples were run in triplicate. Relative expression of GRP78 was calculated using the delta-delta-Ct method with β-actin as the reference control.

Primers used in the PCR were: GRP78 forward, 5’-AAATAAGCCTCAGCGGTTTCTT-3’, and reverse, 5’-TCAAGTTCTTGCCGTTCAAGG-3’; β-actin forward, 5’- CGGGAAATCGTGCGTGAC-3’; and reverse, 5’- CAGGAAGGAAGGCTGGAAG-3’ (Takara, Dalian, China). Quantitative real time PCR was performed according to the MIQE guidelines[28].

***GRP78-siRNA transfection***

Briefly, cell culture plates containing 6-wells were seeded with 2 × 105 cells/well and cultured in RPMI 1640 medium containing 10% (v/v) FBS, 100 units/mL penicillin. siRNA scramble control and validated human GRP78-siRNA were used. GRP78 inhibition was performed using commercially available siRNA kit (Santa Cruz, CA, United States). Knockdown of the target molecule, GRP78, was monitored by western blotting and real time PCR.

***Statistical analysis***

Results of cell apoptosis and cell viability are expressed as mean ± SD. One-way analysis of variance (ANOVA) with Bonferroni's post hoc analysis was performed to compare multiple groups. The Student's *t*-test was used to assess between-group differences. The level of significance was set at *P* < 0.05.

**RESULTS**

***TG-induced ER stress and apoptosis in L02 cells and PGE1 protected L02 and HepG2 cells against ER stress-induced apoptosis***

We confirmed TG-induced ER stress in L02 cells (Figure 1). At 48 h, TG (1 μmol/L) caused significant enhancement in phosphorylation of PERK and GADD34 proteins, as compared to that observed at 6 h. Further, phosphorylation of eIF2α at 24 h was also upregulated as compared to that at 6 h. TG also induced a significant increase in the expressions of GRP78, CHOP and sXBP1 proteins.

We also confirmed TG-induced apoptosis in both L02 and HepG2 cells by means of flow cytometry and MTS assay. As shown in Figure 2A, the apoptotic index of L02 cells after TG treatment was significantly higher than that of control from 6 h to 48 h (*P* < 0.01). At 48 h, the apoptotic index reached 32.66%. TG also showed a dose-dependent increase in apoptotic index with increase in the concentration of TG (1 µmol/L, 2 µmol/L and 3 µmol/L; data not shown). Cell viability of both L02 and HepG2 cells significantly decreased from 24 h to 48 h after TG treatment (Figure 2B and C).

We assessed the effect of PGE1 on ER stress-induced apoptosis in L02 cells (Figure 2A). From 6 h to 48 h, 1 µmol/L PGE1 pretreatment significantly decreased TG-induced apoptotic index. At 48 h, 0.5 µmol/L and 1 µmol/L PGE1 showed dose-dependent protection against TG-induced apoptosis (data not shown). PBA is a low molecular weight chemical chaperone and an ER stress inhibitor. 10 nmol/L PBA significantly inhibited TG induced apoptosis (Figure 2A). MTS assay revealed that PGE1 significantly increased the viability of TG treated L02 cells and HepG2 cells (Figure 2B and C). These results demonstrate the protective effect of PGE1 on ER stress-induced apoptosis in L02 cells and in HepG2 cells.

***PGE1 promoted rapid recovery of ER stress***

After demonstration of the cytoprotective effect of PGE1, we investigated the effects of PGE1 on UPR signals induced by TG. We selected three indicators from UPR, GRP78, CHOP, and sXBP1, as GRP78 is a common downstream target of the three UPR signal pathways. CHOP is the major protein involved in apoptotic signals induced by ER stress, and sXBP1 is activated through the oligomerization of IRE1a[29]. As shown in Figure 3, 1 µmol/L PGE1 appeared to increase TG-induced GRP78, CHOP, and sXBP1 expressions till 6 h post-treatment. However, from 24 h to 48 h, PGE1 suppressed TG-induced GRP78 and CHOP expressions. These results indicate that although PGE1 increased the early expression of UPR signals, it promoted the rapid recovery of ER stress.

***PGE1 induced GRP78 expression via PKA pathway***

The observation that PGE1 attenuated the ER stress-induced apoptosis and promoted the rapid recovery from ER stress, prompted us to investigate the effect of PGE1 on GRP78, CHOP and GADD34 expressions. GRP78 is a critical chaperone which determines the outcome of ER stress. GADD34 is involved in both recovery and resumption of protein synthesis as well as in the ER stress-induced apoptosis[14,30].We also investigated the effects of PGE1 on phosphorylation of eIF2α and PERK, since the activation of PERK and eIF2α is known to induce CHOP and GADD34. From 3 h to 24 h after treatment with 1 µmol/L PGE1, a significant increase in GRP78 expression was observed in both L02 cells and HepG2 cells (Figure 4A and B). PGE1 also decreased CHOP expression in L02 cells and induced the expression of GADD34; however, it suppressed the expression of p-PERK and p-eIF2α proteins (Figure 4A). Results of quantitative real time RT-PCR also demonstrated that PGE1 induced the mRNA expression of GRP78 from 3 h to 24 h (Figure 4C).

To explore the signal pathways that may mediate the effect of PGE1 on GRP78 expression, we inhibited the PKA pathway by H89 or KT5720. Treatment with 10 µmol/L of H89 or 1 µmol/L KT5720 appeared to counteract the effect of PGE1 on GRP78 expression (Figure 5A and B). These results indicate that PGE1 increased GRP78 expression *via* PKA dependent pathway.

***PGE1 protected L02 and HepG2 cells against ER stress-induced apoptosis via PKA dependent induction of GRP78 expression***

10 µmol/L of H89 appeared to inhibit the protective effect of PGE1 on hepatocyte apoptosis, which suggests that the cytoprotective effect of PGE1 was mediated *via* PKA signaling pathway (Figure 5C). To further test whether the protective effect of PGE1 was dependant on the induction of GRP78 expression, we inhibited GRP78 expression by transfection with siRNA specifically targeted against human GRP78. On blocking the PGE1-induced upregulation of GRP78 expression by siRNA (Figure 6A-C), PGE1 appeared to lose its cytoprotective effect in both L02 cells and HepG2 cells (Figure 6D-G). These findings imply that PGE1 protected against hepatocyte apoptosis *via* PKA-dependent induction of GRP78 expression.

**DISCUSSION**

The key findings of this study are that PGE1 protected hepatocyte from ER stress-induced apoptosis; that PGE1 enhanced the expression of GRP78 *via* PKA pathway, and that the cytoprotective effect of PGE1 on hepatocytes was mediated *via* PKA dependent GRP78 induction.

TG are known to induce ER stress by blocking ER Ca2+ uptake, which leads to depletion of ER Ca2+ stores[31]. PGE1 can also increase intracellular Ca2+ level by promoting the influx of Ca2+ from the external medium as well as by mobilization of Ca2+ from intracellular stores[32]. As intracellular Ca2+ level is an important factor in hepatocyte apoptosis and necrosis, cotreatment with TG and PGE1 may have modulated apoptotic signals of L02 cells which eventually led to cell death by necrosis. Therefore, we assessed cell apoptosis and viability on flow cytometry and MTS assay. Our results strongly suggest that PGE1 protected hepatocytes from ER stress-induced apoptosis.

PGE1 is known to have a direct vasodilator as well as an anti-platelet effect[22,33]. Several *in vivo* studies have indicated that PGE1 protects against hepatocyte apoptosis and promotes hepatocyte proliferation *via* inducing downregulation of proinflammatory cytokine levels[24,26], suppression of tumor necrosis factor-α (TNF-α) receptor and adhesion molecule expression[23,24], and by upregulating cyclin C and cyclin D1 expressions[34]. PGE1 also inhibits oxidative stress and nitrosative stress-induced hepatocyte death by inhibiting production of superoxide anion, by enhancing nitric oxide synthase expression and by inhibiting NF-κB activation *in vitro*[19,35,36]. In this study, we demonstrated that PGE1 protected hepatocytes from ER stress-induced apoptosis. Our findings suggest that modulation of UPR may mediate the cytoprotective effect of PGE1 in various pathological conditions.

The mechanisms involved in ER stress-induced apoptosis are yet to be elucidated. The adaptive capacity of cell to ER stress is an important determinant of the outcomes of ER stress. Apoptosis results from sustained or strong ER stress when UPR fails to restore ER homeostasis[37,38]. In the present study, PGE1 treatment enhanced TG-induced CHOP, GRP78 and sXBP1 expressions at 6 h, and significantly attenuated TG-induced CHOP, GRP78 and sXBP1 expressions as assessed at 48 h. These results suggest that PGE1 boosted the initial expression of UPR and promoted restoration of ER homeostasis during ER stress.

During ER stress, UPR may have been activated to restore ER homeostasis. UPR is [precisely](http://dict.youdao.com/search?q=precisely&keyfrom=E2Ctranslation) regulated by three ER stress transducers and their downstream signals. GRP78 is considered as a master regulator of response to ER stress[39,40]. GRP78 binds with all three major UPR sensors (PERK, IRE1a, ATF6) and keeps them inactivated while the homeostasis of ER is maintained. During ER stress GRP78 dissociates from PERK, ATF6 and IRE1a and binds to unfolded or misfolded proteins, promotes their proper folding or directs them to degradation. The dissociation of GRP78 from PERK, ATF6 and IRE1a, triggers the UPR response and further enhances the expression of GRP78. Increased GRP78 augments the folding capacity of the ER, inactivates the three ER sensors and promotes restoration of ER homeostasis[29].

The protective role of GRP78 against apoptosis during ER stress has been demonstrated both *in vivo* and *in vitro*[41-44]. Insulin signaling pathway has been found to promote cell proliferation and improve cell survival *via* upregulation of GRP78 expression[45]. In this study, PGE1 significantly inhibited TG-induced apoptosis. PGE1 induced protein and mRNA expressions of GRP78. Further, inhibition of GRP78 expression *via* either H89 or siRNA hindered the protective role of PGE1 on TG induced apoptosis. These results demonstrate that the cytoprotective effect of PGE1 on hepatocytes was mediated *via* induction of GRP78 during ER stress. Figure 4A and B show that the protein expression of GRP78 induced by PGE1 peaked at 6 h; however, mRNA expression of GRP78 induced by PGE1 peaked at 12 h (Figure 4C). It is difficult to explain the difference in the time frame for attainment of peak levels of mRNA and protein expressions of GRP78. One explanation is that PGE1 regulated the GRP78 expression not only at the transcriptional level but also at the translational or posttranslational level. Whether PGE1 regulates the expression of GRP78 at the translational or posttranslational level warrants further studies.

The other important signal pathways in ER stress-induced apoptosis are mediated *via* induction of CHOP and GADD34. During UPR, phosphorylation of eIF2α *via* PERK pathway results in inhibition of mRNA translation and general protein synthesis. However, mRNA for activating transcription factor 4 (ATF4) is selectively upregulated. Activation of ATF4 results in induction of CHOP and GADD34 expressions. GADD34 and protein phosphatase 1 were shown to promote dephosphorylation of eIF2α and allow protein synthesis[46]. However, recent studies have shown that induction of GADD34 exacerbates the disturbance of ER homeostasis and leads to cell apoptosis by increasing oxidative stress[47]. CHOP has been identified as one of the most important mediators of ER stress-induced apoptosis; it induces apoptosis through various signal pathways[48].

In the current study, we had anticipated inhibition of GADD34 by PGE1. However, we observed an enhanced expression of GADD34 and significant inhibition of p-PERK, CHOP and p-eIF2α expressions in L02 cells. Our results indicate that the cytoprotective effect of PGE1 against hepatocyte apoptosis does not depend on the inhibition of GADD34. Since PGE1 lost its protective effects after inhibition of GRP78 expression in this study, it is possible that the increased expression of GADD34 in our study was the result of induction of GRP78 by PGE1 and that this represented restoration of ER homeostasis. Further study is needed to clarify the role of GADD34 in ER stress-induced apoptosis.

The biological effects of PGE result from its binding to its receptors, EP1 to EP4, and previous studies have shown that L02 cells express EP1 receptors[49]. Binding of PGE1 with its receptors stimulates the production of the second messenger cyclic 3, 5 adenosine monophosphate (cAMP). cAMP may act *via* distinct intracellular signaling effectors such as PKA and the exchange proteins activated by cAMP[50]. A previous study has shown that cAMP has PKA-independent interaction with Ca2+ stored in lymphocytes[51]. To test whether hepatoprotective effect of PGE1 is mediated *via* PKA-independent interaction with Ca2+ stores in L02 cells, we used H89 to block PKA pathway. The results showed that the protective effects of PGE1 were largely dependent on the PKA pathway. However, as showed in Figure 5A, H89 inhibited GRP78 expression most effectively at 6 h, and that at 12 h the inhibitory effect was alleviated; the increased apoptotic index lasted from 6 h to 48 h (Figure 5C). It is difficult to explain this result. One explanation is that H89 may also have inhibited other kinases in addition to PKA. H89 has been shown to inhibit at least 8 kinases beside PKA[52]. The role of PKA and other kinases in ER stress-induced apoptosis remains to be studied in future.

It is known that GRP78 expression is regulated at the transcriptional level by ER stress. Previous studies have shown that preconditioning to ER stress protects against cell death *via* induction of GRP78 or autophagy[53]. In this study, although PGE1 pretreatment significantly induced GRP78 expression and enhanced TG-induced early CHOP and sXBP1 expressions, PGE1 alone inhibited the expressions of p-PERK and p-eIF2α and CHOP. Therefore, whether PGE1 induced GRP78 expression *via* induction of ER stress is not known. In previous studies, leptin was shown to induce GRP78 expression in neuronal cells through the PI3K-mTOR pathway[54]; further, oncostatin M was also shown to induce GRP78 expression without triggering ER stress[55]. The mechanisms involved in the induction of GRP78 expression by PGE1 warrant further investigation.

In conclusion,this is the first study to demonstrate that the cytoprotective effect of PGE1 against ER stress-induced apoptosis is mediated *via* PKA-dependent induction of GRP78 expression in hepatocytes. Further studies are required for devising treatment strategies to protect hepatocyte against ER stress-induced apoptosis, which will be of much clinical relevance in the context of liver diseases.

**COMMENTS**

***Background***

Prostaglandin E1 (PGE1) has been shown to protect against hepatocyte apoptosis, however, the role of PGE1 in endoplasmic reticulum (ER) stress-induced apoptosis of hepatocytes is largely unknown.

***Research frontiers***

ER stress has been implicated in the pathogenesis of various liver diseases. Understanding the mechanisms underlying ER stress-induced apoptosis and devising a treatment strategy to protect hepatocytes from ER stress-induced apoptosis will benefit most patients with liver diseases.

***Innovations and breakthrough***

Pretreatment with PGE1 protected hepatocytes against thapsigargin-induced apoptosis. PGE1 alone induced protein and mRNA expressions of glucose-regulated protein (GRP) 78. Treatment with protein kinase A (PKA)-inhibitor H89, KT5720 or siRNA specifically targeted against human GRP78 blocked PGE1-induced upregulation of GRP78. The hepatoprotective effect of PGE1 was lost by blocking GRP78 expression by either H89 or siRNA. Our study demonstrates for the first time that PGE1 protects against ER stress -induced hepatocyte apoptosis *via* PKA pathway dependent induction of GRP78 expression.

***Applications***

PKA and GRP78 may be a new target for pharmacological treatment in patients with liver diseases.

***Peer-review***

The strengths of this study lie in the experimental design, methodology and that the authors describe the caveats and potential pitfalls in great detail. Overall, the study was well-designed and the paper is interesting and sound.

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**Specialty type:** Gastroenterology and hepatology

**Country of origin:** China

**Peer-review report classification**

Grade A (Excellent): 0

Grade B (Very good): B

Grade C (Good): C

Grade D (Fair): 0

Grade E (Poor): 0

****

125kDa

p-PERK

fold change 1.0 1.3 1.6a 1.8b 1.7b 1.6b

****

PERK

125kDa

****

p-eIF2α

36kDa

fold change1.0 1.3 1.5a 2.1b 1.5a 0.6a

****

36kDa

eIF2α

****

GADD34

73kDa

fold change1.0 1.3 2.8b 4.6b 4.0b 3.8b

****

42kDa

β-actin

(h)

TG

0 6 12 24 36 48



GRP78

78kDa

fold change 1.0 1.6a 3.8b 4.6b 6.0b 5.8b



55kDa

sXBP1

fold change 1.0 4.2b  3.2b 3.0b 2.2b 2.1b



29kDa

CHOP

fold change 1.0 2.4a 2.5b 4.3b 4.0b 2.5a



42kDa

β-actin

**Figure 1 Thapsigargin induced endoplasmic reticulum stress in L02 cells.** L02 cells were treated with 1 μmol/L TG for 6, 12, 24, 36, and 48 h; Expressions of p-PERK, p-eIF2α, GADD34, GRP78, sXBP1 and CHOP were assessed by western blotting. Representative blots from three independent experiments are presented. The results of densitometric analysis are presented as a fold change compared to that at 0 h (a*P* < 0.05; b*P* < 0.01). TG: Thapsigargin; ER: Endoplasmic reticulum; p-PERK: Phospho-PKR-like ER kinase; p-eIF2α: Phospho-eukaryotic translation initiation factor-2α; GADD34: Growth arrest and DNA damage-inducible gene34; GRP78: Glucose-regulated protein 78; sXBP1: Spliced X box-binding protein1; CHOP: C/EBP homologous protein.

A

B

C

**Figure 2 PGE1 protected against thapsigargin-induced apoptosis in both L02 cells and HepG2 cells.** L02 cells and HepG2 cells were pretreated with PGE1 or PBA for 1 h and treated with a final concentration of 1 μmol/L TG for 6, 24, and 48 h. A: Apoptotic index of L02 cells was determined on flow cytometry. Histograms represent mean ± SD of five separate experiments, each of which was performed in triplicate. *bP* <0.01 *vs* control at the same time point; *dP* <0.01 *vs* TG at the same time point. B and C: Cell viability of L02 cells and HepG2 cells was determined by MTS [3-(4,5-dimethylthiazol-2-yl)-5-(3-carboxymethoxyphenyl)-2-(4-sulfophenyl)-2H-tetrazolium ] assay. The absorbance was measured at 490 nm and cell viability was normalized as a percentage of control. Histograms represent mean ± SD of five separate experiments, each of which was performed in triplicate. *dP* < 0.01 *vs* TG at the same time point. TG: Thapsigargin; PGE1: Prostaglandin E1; PBA: 4-phenylbutyric acid.

|  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- |
|  | 6 | | 24 | | 48 (h) | |
| TG | **+** | **+** | **+** | **+** | **+** | **+** |
| PGE1 | **-** | **+** | **-** | **+** | **-** | **+** |

C:\Users\lenovo\Desktop\1.jpg

GRP78

78kDa

fold change 1.0 2.0b 1.0 0.9a 1.0 0.7a

C:\Users\lenovo\Desktop\2.jpg

sXBP1

55kDa

fold change 1.0 2.5b 1.0 1.2 1.0 0.4b

C:\Users\lenovo\Desktop\4.jpg

CHOP

29kDa

fold change 1.0 3.5b 1.0 0.8a 1.0 0.6a

C:\Users\lenovo\Desktop\3.jpg

β-actin

42kDa

**Figure 3 PGE1 promoted rapid recovery of endoplasmic reticulum stress.** L02 cells were pretreated with 1 μmol/L PGE1 for 1 h and at a final concentration of 1 μmol/L TG for 6, 24, and 48 h. Expressions of glucose-regulated protein 78 (GRP78), C/EBP homologous protein (CHOP) and spliced X box-binding protein1 (sXBP1) were assessed on western blotting. One representative blot each of three individual experiments is presented. The results of densitometric analysis are presented as a fold change compared to those of TG at the same time point (a*P* < 0.05, b*P* < 0.01). PGE1: Prostaglandin E1; TG: Thapsigargin.

**A**

(h)

PGE1

0 3 6 12 24



78kDa

GRP78

fold change 1.0 4.1b 17.5b 12b 11b



42kDa

β-actin

(h)

0 3 6 12 24

PGE1

****

29kDa

CHOP

fold change 1.4 1.1 0.9 0.6b 0.2b

****

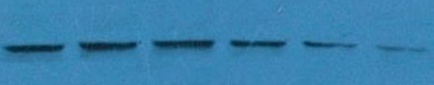
(h)

0 6 12 24 36 48

42kDa

β-actin

PGE1

****

125kDa

p-PERK

fold change 1.0 1.0 0.8 0.5b 0.2b 0.1b

****

125kDa

PERK

****

36kDa

p-eIF2α

fold change 1.0 0.9 0.9 0.8a 0.7a 0.5b

****

36kDa

eIF2α

****

73kDa

GADD34

fold change 1.0 1.1 1.2a 1.5b 1.6b 1.9b



42kDa

β-actin

**B**

(h)

PGE1

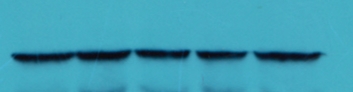
0 3 6 12 24



78kDa

GRP78

fold change 1.0 1.5 3.0b 3.2b 2.6b



42kDa

β-actin

**C**

**Figure 4 PGE1 induced glucose-regulated protein 78 protein and mRNA expressions**. To determine GRP78 and C/EBP homologous protein (CHOP) protein or mRNA expressions, L02 cells and HepG2 cells were treated with 1 μmol/L PGE1 for 3, 6, 12 and 24 h; for detecting growth arrest and DNA damage-inducible gene34 (GADD34) and the phospho-PKR-like ER kinase (p-PERK) and phospho-eukaryotic translation initiation factor-2α (p-eIF2α), L02 cells were treated with 1 μmol/L PGE1 for 6, 12, 24, 36 and 48 h. A: Expressions of GRP78, CHOP, GADD34, p-PERK, and p-eIF2α in L02 cells were assessed on western blotting. One representative blot each from the three individual experiments is presented. The results of densitometric analysis are presented as a fold change compared to those at 0 h (a*P* < 0.05, b*P* < 0.01). B: Expression of GRP78 in HepG2 cells was assessed on western blotting. One representative blot each from the three individual experiments is presented. The results of densitometric analysis are presented as a fold change compared to those at 0 h (b*P* < 0.01). C: mRNA expression of GRP78 in L02 cells was assessed by quantitative real time PCR (E). Histograms represent mean ± SD of three experiments (b*P* < 0.01 *vs* those at 0 h). GRP78：Glucose-regulated protein 78; PGE1: Prostaglandin E1; TG: Thapsigargin; ER: Endoplasmic reticulum.

**A**

|  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- |
|  | 3 | | 6 | | 12 | |
| PGE1 | **+** | **+** | **+** | **+** | **+** | **+** |
| H89 | **-** | **+** | **-** | **+** | **-** | **+** |

(h)



78kDa

GRP78

fold change 1.0 0.7b 1.0 0.5b 1.0 0.9b



42kDa

β-actin

**B**

|  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- |
|  | 3 | | 6 | | 12 | |
| PGE1 | **+** | **+** | **+** | **+** | **+** | **+** |
| KT5720 | **-** | **+** | **-** | **+** | **-** | **+** |

(h)

**2_看图王.jpg**

78kDa

GRP78

fold change 1.0 0.4b 1.0 0.5b 1.0 1.0b

**11_看图王.jpg**

42kDa

β-actin

**C**

**D**

**Figure 5 PGE1 induced glucose-regulated protein 78 expression and protected L02 cells against endoplasmic reticulum stress-induced apoptosis *via* protein kinase A dependent pathway.** A and B: L02 cells were pretreated with or without PKA inhibitor H89 (10 μmol/L) or KT5720 (1 μmol/L), and then 1 μmol/L PGE1 for 3 h, 6 h, and 12 h. The expressions of GRP78 were detected on western blotting. One representative blot each from three independent experiments is presented. The results of densitometric analysis are presented as a fold change compared to those of PGE1 at the same time points (b*P* < 0.01). C: L02 cells were pretreated with or without 10 μmol/L H89, and then 1 μmol/L PGE1 and a final concentration of 1 μmol/L TG for 6 h, 24 h and 48 h. The apoptotic index was determined on flow cytometry. Histograms represent mean ± SD of five separate experiments, each of which was performed in triplicate. (d*P* < 0.01 *vs* those of TG+PGE1 at the same time point; f*P* < 0.01 *vs* those of TG at the same time point). D: L02 cells were pretreated with or without 10 μmol/L H89, and then 1 μmol/L PGE1 and a final concentration of 1 μmol/L TG for 6 h, 24 h and 48 h. Cell viability of L02 cells was determined on MTS assays. Histograms represent mean ± SD of five separate experiments, each of which was performed in triplicate (d*P* < 0.01 *vs* those of TG+PGE1 at the same time point; f*P* < 0.01 *vs* those of TG at the same time point). GRP78：Glucose-regulated protein 78; PKA:Protein kinase A; PGE1: Prostaglandin E1; TG:Thapsigargin.

**A**

ConsiRNA

PGE1

GRP78siRNA

PGE1

0 12 24 0 12 24 0 12 24 (h)

PGE1



GRP78

78kDa

fold change 1.0 5.9 10.2b 1.0 0.7 0.2b 1.0 4.8 5.4b



β-actin

42kDa

**B**

ConsiRNA

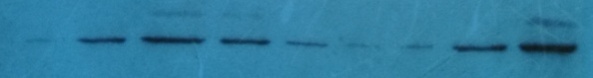
PGE1

GRP78siRNA

PGE1

PGE1

0 12 24 0 12 24 0 12 24 (h)



GRP78

78kDa

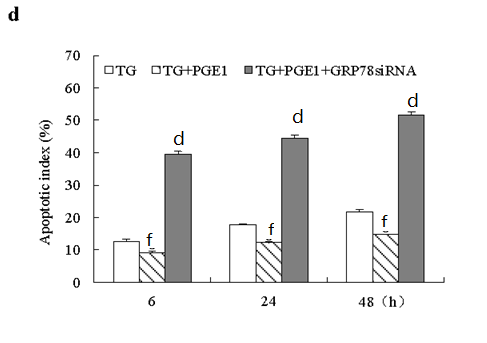
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β-actin

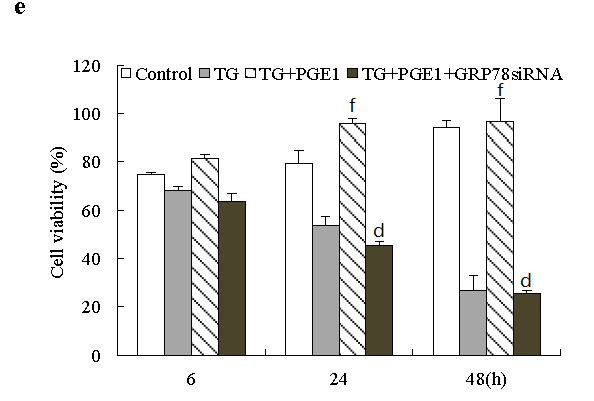
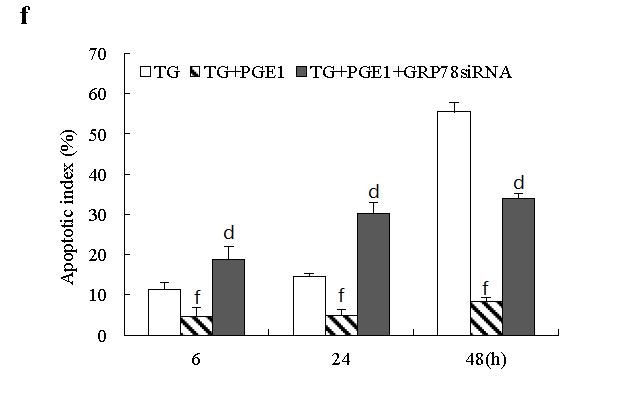
42kDa

**C**

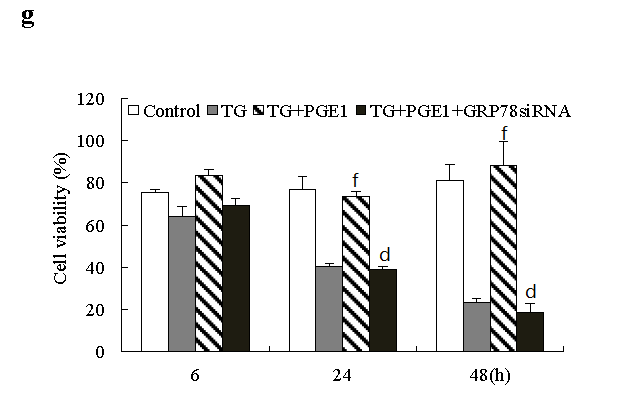


**D**

**E**



**F**



**G**

**Figure 6 PGE1 protected against endoplasmic reticulum stress-induced apoptosis *via* induction of glucose-regulated protein 78 expression in both L02 and HepG2 cells.** L02 and HepG2 cells were transfected with either siRNA scramble control (ConsiRNA) or siRNA against human GRP78 (GRP78 siRNA) for 48 h. The cells were treated with 1 μmol/L PGE1 for 12 h and 24 h. A and B: Expressions of GRP78 in L02 cells and in HepG2 cells were detected on western blotting. One representative blot each from the three individual experiments is presented. The results of densitometric analysis are presented as a fold change compared to those at 0 h (b*P* < 0.01). C: mRNA expression of GRP78 in L02 cells was detected by real time PCR. Histograms represent mean ± SD of three experiments (a*P* < 0.05; b*P* < 0.01 *vs* those at 0 h). D and F: Apoptotic indices of L02 cells and HepG2 cells were determined on flow cytometry. Histograms represent mean ± SD of five independent experiments, each of which was performed in triplicate (d*P* < 0.01 *vs* those of TG + PGE1 at the same time point; f*P* < 0.01 *vs* those of TG at the same time point). E and G: Cell viability of L02 cells and HepG2 cells was determined on MTS assay. Histograms represent mean (± SD) of five independent experiments, each of which was performed in triplicate (d*P* < 0.01 *vs* those of TG + PGE1 and f*P* < 0.01 *vs* those of TG at the same time point). GRP78: Glucose-regulated protein 78; PGE1: Prostaglandin E1; TG: Thapsigargin.