**Name of Journal: *World Journal of Stem Cells***

**Manuscript NO: 34815**

**Manuscript Type:** **Therapeutics Advances**

**Amyotrophic lateral sclerosis as a protein level, non-genomic disease: Therapy with S2RM exosome released molecules**

Maguire G. ALS therapy with stem cell molecules

**Greg Maguire**

**Greg Maguire,** BioRegenerative Sciences, Inc., South La Jolla, CA 92037, United States

**ORCID number:** Greg Maguire ([0000-0001-5484-6157](http://orcid.org/0000-0001-5484-6157" \t "_blank)).

**Author contributions:** Maguire G wrote the artcle.

**Conflict-of-interest statement:** Maguire G is owner of BioRegenerative Sciences.

**Open-Access:** This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: http://creativecommons.org/licenses/by-nc/4.0/

**Manuscript source:** Invited manuscript

**Correspondence to:** **Dr. Greg Maguire,** BioRegenerative Sciences, Inc., 505 Coast Blvd. South La Jolla, CA 92037, United States. gmaguire@bioregenerativesciences.com

**Telephone:** +1-858-5219203

**Received:** May 29, 2017

**Peer-review started:** June 1, 2016

**First decision:** July 17, 2017

**Revised:** August 23, 2017

**Accepted:** September 3, 2017

**Article in press:**

**Published online:**

**Abstract**

Amyotrophic lateral sclerosis (ALS) is a rapidly progressing neurodegenerative disease that leads to death. No effective treatments are currently available. Based on data from epidemiological, etiological, laboratory, and clinical studies, I offer a new way of thinking about ALS and its treatment. This paper describes a host of extrinsic factors, including the exposome, that disrupt the extracellular matrix and protein function such that a spreading, prion-like disease leads to neurodegeneration in the motor tracts. A treatment regimen is described using the stem cell released molecules from a number of types of adult stem cells to provide tissue dependent molecules that restore homeostasis, including proteostasis, in the ALS patient. Because stem cells themselves as a therapeutic are cumbersome and expensive, and when implanted in a host cause aging of the host tissue and often fail to engraft or remain viable, only the S2RM molecules are used. Rebuilding of the extracellular matrix and repair of the dysfunctional proteins in the ALS patient ensues.

**Key words:** Amyotrophic lateral sclerosis; Proteins; Stem cells; Extracellular matrix; Exposome; Heat shock protein; Stem cell molecules

**© The Author(s) 2017.** Published by Baishideng Publishing Group Inc. All rights reserved.

**Core tip**: The author propose that amyotrophic lateral sclerosis is a disease of damaged and misfolded proteins induced by environmental regulators, occurring at the level of translation and post-translation, and not at the level of the genome. The damaged proteins disrupt the extracellular matrix surrounding neurons, precluding normal communication between neurons and the neural stem cells that surround the neurons. As a consequence, the neural stem cells no longer properly shuttle heat shock proteins to the neurons, and many proteins within the neuron misfold. The misfolded proteins are prion-like, leading to a spreading, destructive sequelae within and between neurons and other cells in the nervous system.

Maguire G. Amyotrophic lateral sclerosis as a protein level, non-genomic disease: Therapy with S2RM exosome released molecules. *World J Stem Cell* 2017; In press

**INTRODUCTION**

Epidemiological studies of amyotrophic lateral sclerosis (ALS) are in progress, but so far we know the incidence is related to aging, primarily those over 60, two in 100000 new cases per year, and men are afflicted 1.5 times more than women. Despite suspected clusters being studied in the United States, including around bodies of water where contaminated seafood was ingested[1] the only cluster of ALS patients was confirmed in the Western Pacific (Guam) in the 1940’s where locals were ingesting neurotoxins[2]. Some physicians report that they are “seeing increasing numbers of younger ALS patients”. Agreeing with what the epidemiology suggests, the etiology of ALS is multifactorial, including exposure to various chemicals[3].Most ALS experts think these brain afflictions have multiple causes, including genetics, epigenetics[4,5], poor diet, lack of exercise, stress, and a variety of environmental agents, including pathogens[6], pesticides, for example, pentachlorobenzene, and other chemicals, such as PCBs and formaldehyde[7]. A recent New England study found pesticides, solvents, or heavy metals, increased the risk of ALS[8], and Seneff *et al*[9] propose that glyphosate (Roundup) causes a number of mechanistic abnormalities associated with ALS, including protein misfolding, and elevated levels of glutamate[10].Surprisingly, even silver nanoparticles 100 nm in diameter, commonly used in personal, commercial, and industrial products can, when in aerosol form, be absorbed by the nasal mucosa and travel to the brain where they elicit a microglial, potentially inflammatory, response[11].

Neurological diseases are very complex, involving many underlying causative pathway, and, as such, using poly pharmacological approaches may increase efficacy when compared with single molecule therapies[12]. Therefore, neurodegenerative disorders such as ALS might require system therapeutic approaches[13] such as S2RM technology to obtain long-lasting, efficacious results[12]. In addition to the motor system dysfunction, deficits in mentation are reported for 50% of the ALS patients[14] demonstrating the complexity of this disease as measured by clinical endpoints.

The mechanisms underlying the destruction of motor neurons (MN) and other cell types in amyotrophic lateral sclerosis (ALS) are beginning to be understood, with many recent studies demonstrating key irregularities at the molecular and tissue levels in animal models and human ALS patients. Laboratory evidence suggests that stem cell therapy can positively affect ALS[15]. I will explain these distinct irregularities, show how the distinct components are interrelated, and then, based on the evidence, provide a rationale for using the stem cell-based S2RM technology to treat ALS.

In the clinic, the practitioner usually has no way to determine whether the stem cells used for therapy are viable, whether they perfuse to the site of injury following administration, and whether the cells engraft into the tissue and remain functional over time. Limited clinical research suggests that the stem cells often don’t engraft and therefore don’t remain functional in the patient. Laboratory models with detailed analysis show that stem cell injections into host tissue often fail for various reasons, including programmed cell death (apoptosis), a process that may be activated when stem cell numbers (shown for mesenchymal stem cells) become too high in the tissue[16]. As a therapeutic strategy, given that 80% or more of the therapeutic properties of stem cells is derived from the release of SRM, the better choice is to use the molecules derived from stem cells grown in optimal conditions in the laboratory. Those molecules can then be more easily, less expensively, and with better efficacy, directly administered to the patient[17].

Further, recent work in humans shows that stem cell therapy may induce advanced aging because forced cellular regeneration that accompanies re-engraftment induces intrinsic stem cell aging as measured using a *p16* biomarker[18]. In mouse models, serial transplant readily “exhausts” stem cells evidenced by a diminished capacity to produce new somatic cells[19]. Using only the molecules released from stem cells means that the S2RM technology will not induced intrinsic aging caused by engraftment of the stem cells. Considering the use of iPSCs for therapeutic use, these cells suffer from genetic and epigenetic reprogramming errors, and the older the patient, the more mutations found in mitochondrial DNA along with increased metabolic problems[20].

I will now describe the mechanisms underlying ALS, and then propose a treatment regimen based on the use of S2RM technology.

**MECHANISMS UNDERLYING ALS**

The multifactorial disease ALS is a result of some of the mechanisms listed below. Evidence suggest that these many factors, such as exposure to harmful chemicals during conception, development, and throughout life, coupled with aging, infection, and poor lifestyle can lead to the breakdown of the all-important extracellular matrix (ECM) and peri-neuronal nets (PNN) that support the motor tracts. The breakdown of matrix is likely a generalized phenomenon in ALS patients given that in animal models a breakdown of the matrix in the gut leads to “leaky gut,” followed by the induction of inflammation[21]. At the root of the breakdown of the ECM and PNN is a loss of homeostasis, and particularly a loss of proteostasis, meaning a loss of the building blocks for the ECM and PNN. As a major consequence of the ECM breakdown, heat shock proteins (HSP) and chaperone proteins from neighboring cells cannot supply the motor neurons with their necessary proteins that enable autophagy, and proper protein folding. These results are deadly for motor neurons.

**ALS IS NOT GENETIC, AND MUTATIONS ARE NOT SPECIFIC TO ONE GENE**

First, because of overlapping genes, a mutation at one nucleotide can have effects in multiple genes[22]. The very concept of a gene is in dispute[23], and the nebulous concept of a gene as a cause for disease is in question[24], especially given the “buffering” capacity of a redundant system to preclude simple mutations as disease causative agents[25] so well espoused in a new book by Professor Nobel of Oxford University[26].

“Genetic Factors Are Not the Major Causes of Chronic Diseases” is the title of highly regarded PLoSOne article by Professor Stephen Rappaport at UC Berkeley[27]. Indeed because of the failure of genetic studies to predict disease, where over 2000 genome-wide association studies have infrequently found risks greater than 1.2[24],geneticists are now using whole-genome sequencing (WGS) to search for “missing heritability”. The precision with which we can analyze the genome is very seductive, but as is the case with many seductresses, the outcome may not be positive. Rather than genetics, the empirical evidence suggests that exposures, extrinsic factors, are necessary determinants of disease that may or may not be operating in a background of genetic diversity, intrinsic factors. Thus, the extrinsic factors may be operating “above” the genome, for example, at the level of translation and post-translation. That the symptoms of ALS are largely a result of protein dysfunction is indisputable. Over sixty years ago the Millers[28] first discovered covalent binding of tissue proteins and carcinogen. Today we understand that exogenous (*i.e.,* xenobiotics) and endogenous chemicals (*i.e.,* oxidative stress factors), such as electrophiles, are a primary cause of protein dysfunction through, for example, covalent modifications[29]. Clearly genetics alone will not predict disease, whereas the exposome in general, and protein function specifically will be important in understanding many diseases and indications.

Although reports continue to claim genetic causes for ALs, such as mutations in C9orf72[30], studies of the loss of function of C9orf72 is insufficient to cause motor neuron disease by itself[31]. Likewise in SOD1 mutant mice, a predominant model for ALS, transcription and splicing are normal, suggesting that the abnormality occurs at the level of translation or post-translation[32]. Further, proteins, such as SOD1 can become pathogenic through non-heritable modifications such as oxidation, with characteristics similar to the SOD1 mutant in familial ALS, suggesting that conformational abnormalities due to WT-SOD1 protein translation or post-translational events can underlie sporadic ALS pathogenesis[33]. As MIT’s Evelyn Fox Keller[23] and Oxford’s Denis Noble[26] explain so well, mutations in the DNA often don’t express at the level of protein function, and simple sequence analysis of DNA alone will tell us little or nothing about disease. Let us now look at some of the mechanisms known to underlie ALS.

**EXTRACELLULAR MATRIX**

First, the extracellular matrix (ECM), particularly the perineuronal nets (PNN), surrounding motor neurons are dysregulated in ALS animal models. Reactive astrocytes and the abnormal accumulation of chondroitin sulfate proteoglycans (CSPGs) in the ECM[34] may create a microenvironment that is non-permissive for neural regeneration in ALS. For example, proteoglycans, specifically Decorin and Perlecan, control cell autophagy, meaning that these ECM components are important in clearing the motor neuron of misfolded proteins[35]. Further, breakdown of the ECM will lead to fragments that upregulate kainate receptors and exacerbate excitatory neurotoxicity[36]. PNNs are specialized for neurons and are condensed versions of ECM, the material that surrounds our individual cells. If these PNNs are restored through mesenchymal stem cell therapy by intrathecal injection (IT), then the motor function of the treated animals can be significantly improved. The mode of action in restoring the PNN is thought to be the release of a multitude of trophic factors, including neurotrophic factors, from the implanted stem cells[15]. In contrast to the success of this study, and like studies where cells or multiple factors are used, administration of only one neurotrophic factor failed to provide any benefit[37,38]. Recent safety studies show that administration (IT or IM) of stem cells to ALS patients is safe, and early indications are that some benefit was provided[39]. The aforementioned data provide excellent evidence that the S2RM methodology will work well in treating ALS.

Evidence suggests that the breakdown of ECM may be generalized phenomenon throughout the body in ALS patients given that the basement membrane breakdowns[40], as does the matrix in the gut, which leads to “Leaky gut” and an ensuing inflammatory response[21].

**MUSCLE AND NEUROMUSCULAR JUNCTION**

Evidence that pathological mechanisms begin at the neuromuscular junction (NMJ) rather than within the motor neurons[41-43] suggests that ALS may be a distal axonopathy. Cultures of satellite cells from biopsies of ALS patient biopsies have been shown to proliferate similarly to cells obtained from healthy muscle. However, the morphology of the ALS derived cells resembles that of senescent cells[44]. Myoblasts obtained from ALS patients are unable to normally differentiate into myotubes[44,45].

Human ALS samples are rare, but autopsy of an ALS patient demonstrated skeletal muscle changes with clear signs of denervation and reinnervation. However, the patient had normal appearing motor neurons; thus pathological changes in skeletal muscle appear to be present before the motor neurons are affected[41], thus providing human evidence for the “dying-back” phenomenon. *In vitro* cultures of SCs from ALS patients demonstrate a senescent-like morphology, disturbed differentiation, and an apparent inability to proceed through the myogenic program resulting in a decreased ability to regenerate and mature to functional myofibres[46]. Proteostasis is critical for maintaining aged satellite cells (muscle stem cell) function, preventing the stem cells from becoming senescent[47]. Proteostasis is dependent on a normal ECM and cytoskeleton. Exercise has been shown to increase the number of MSCs in muscle, where the MSCs act to release growth factors, inducing muscle regeneration[48]. At the NMJ, agrin, an ECM protein, is critical to maintenance of the synapse, and can rescue the NMJ in animal models of neuromuscular degeneration[49]. Agrin is expressed by MSCs, including bone marrow stem cells[50]. Again, agrin constitutes but one part of maintaining homeostasis at the NMJ, and if used alone to treat ALS, would represent a minimally effective, reductionist treatment regimen.

**SUPEROXIDE DISMUTASE**

The superoxide dismutase(SOD) protein is misfolded in the motor neurons of ALS patients, which means that SOD will not clear out oxygen radicals developed in the highly active motor neurons – thus, the motor neuron will die. Although some correlations of SOD misfolding with DNA mutations has been reported, Horwich’s lab at Yale has shown that the transcripts in SOD1 mice are largely normal[32], meaning that the RNA for SOD is normal and that the protein dysfunction is a result of translation, or post-translational mechanisms. Misfolded SOD1 will have many negative consequences, including binding the Voltage Dependent Anion Channel isoform 1 (VDAC1) in mitochondria, creating organelle dysfunction[51].

**TDP43**

TDP-43 is a protein that helps to regulates gene expression, including RNA transcription, splicing, transport, and translation. CNS cells, in particular motor neurons, are highly vulnerable to TDP-43 dysfunction. In TDP-43 knockdown mice, the effects are predominantly observed in astrocytes within the spinal cord, suggesting that TDP-43 dysfunction in astrocytes may play an important role in motor neuron degeneration. TDP-43 is misfolded in ALS, and spreads in a prion-like fashion[52].

TDP-43 can also impair mitochondrial dynamics and function in motor neurons. Previous studies described how ALS-associated mutations transform the cytosolic SOD1 into a membrane-interacting protein, and therefore are then capable of associating with organelles such as mitochondria. Recently the peptides derived from the TDP-43 prion-like domain were shown to have membrane-damaging capacity[53].

**DJ-1 PROTEIN IS MISFOLDED**

DJ-1 responds to oxidative stress to protect cells, *i.e.,* motor neurons, from radical oxygen species[54,55]. Proper function of this protein is also controlled by Heat shock proteins[56], that must be donated to the motor neurons from adjacent cells. DJ-1 is also responsible for DNA repair, acting as guanine glycation repair protein[57]. Thus, if DJ-1 is misfolded, guanine glycation occurs and with resultant mutation frequency, DNA strand breaks, and cytotoxicity.

**VGF**

Fragment depletion of VGF is likely involved in the onset and progression of ALS. In fibroblasts and plasma samples from ALS patients in an advanced stage, VGF C-terminus peptides were reduced in both fibroblast and plasma. In the G93A-SOD1 mice, the same VGF peptides were decreased in plasma in the late-symptomatic stage, while in the spinal cord down-regulation occurred earlier. Immunohistochemistry studies suggests that a large amount of gray matter is VGF C-terminus positive in control mice (including nerve terminals, axons and some somata of motoneurons), while a significant reduction of VGF peptides has already occurred in the pre-symptomatic stage[58].

**FUS**

In a small number of patients, mutations in fused in sarcoma/translocated in liposarcoma (FUS/TLS or FUS) are associated with ALS. Animal models provide evidence that mutant FUS exerts a gain-of-toxic function in the cytoplasm of motor neurons resulting in cellular degeneration. Recombinant FUS proteins perfused into squid axoplasm inhibits anterograde and retrograde transport. Regardless of whether the mutation affects the nuclear localization signal (*e.g.,* R521G or R495X) or the glycine-rich domain (*e.g.,* G230C) of FUS, each of the disease variants tested inhibited FAT. When mutant FUS is mized with the chaperone Hsp110, the impairment is reduced suggesting that a misfolded FUS is responsible for the gain-of-toxic function[59].

**DIPEPTIDE REPEAT PROTEINS**

One of the most common genetic changes associated with ALS and frontotemporal dementia (FTD) are aberrant hexanucleotide repeat expansions in C9orf72. Transcripts containing these expansions undergo repeat-associated non-ATG translation (RAN-T) to form five dipeptide repeat proteins (DPRs). In C9orf72-ALS/FTD patients, DPR aggregates are found throughout the CNS. DPRs can also cause degeneration when expressed *in vitro* in neuronal cultures and in vivo in animal models. The prion-like spread of the misfolded proteins is thought to lead to the spreading progression of pathology in the patients of many neurodegenerative diseases. Whether DPRs spread has yet to be determined. Using a number of experimental cell culture techniques, including spinal motor neurons derived from iPScs of C9orf72-ALS patients, data suggests cell-to-cell spreading of DPRs by exosome-dependent and exosome-independent pathways[60].

**MITOCHONDRIAL DYSFUNCTION**

Mitochondria are responsible for efficiently creating energy in our cells, including motor neurons, through a process called oxidative phosphorylation. In ALS, mitochondria function is compromised, and cells go into an inefficient energy producing state called glycolysis[61] that is 16 times less efficient than oxidative phosphorylation.

Inflammation is an essential component of immunity, but excessive response has been shown to damage tissue and lead to autoimmune disease. The excessive response is avoided with parallel, integrated regulatory responses that begin the healing. One of the keys to this regulatory response is interleukin-10 (IL-10). IL-10 is particularly important because it limits macrophage proinflammatory functions. IL-10 modifies macrophage function by enhancing the clearance of damaged mitochondria and modulating cellular metabolism to limit inflammation[62]. IL-10 is part of the molecular mix in the S2RM.

**HEAT SHOCK PROTEINS**

Neurons, once differentiated from their stem cell precursors, stop producing HSP. Without endogenous HSP, the motor neurons are dependent on surrounding cells to shuttle the HSP from the surrounding cell to the motor neuron (Figure 1). Mesenchymal stem cells release HSP, chaperone proteins[63], and HSP is transferred by exosomes from one cell to another to maintain protein homeostasis[64]. A compromised ECM impedes HSP transfer from one cell to another (Figure 2).

**EXOSOMES**

Secretion of exosomes and their intercellular transmission of chaperone protein are key mechanisms for non–cell-autonomous proteostasis. Hsp40, Hsp70, and Hsp90, are secreted from cells *via* exosomes under physiological conditions[64]. Thus exosomes from healthy cells, including healthy stem cells, are necessary for the transfer of HSP to motor neurons. Exosome transfer from one cell to another is facilitated by the ECM. Exosomes from MSCs have been shown to alleviate some of the abnormal SOD1 aggregation in neural cells in an *in vitro* mouse model of ALS[65]. Exosomes that are loaded with the normal set of molecules, and/or the ability of the exosome to travel from healthy stem cells to cells in need of rescue may be compromised in ALS.

**GLUTAMATE IS ELEVATED**

Astrocytes associated with an ALS mouse model, called AbA cells, do not express detectable levels of the specific glial glutamate transporter GLT-1. Lack of a glutamate transporter in the glial cells of the ALS model may explain, at least partially, excitotoxic damage to motor neurons. Furthermore, the induction of neurotoxicity using the conditioned medium from AbA cells is specific to motor neurons[66]. As part of the support system, the astrocytes and the ECM, including the PNN, are critical to maintaining glutamate levels surrounding the motor neurons and preventing the glutamate induced excitatory neurotoxicity. Breakdown of the ECM also upregulates kainate receptors that are responsive to glutamate and mediate excitotoxicity[36].

**ASTROCYTES**

Astrocytes derived from the spinal cord of persons with sporadic and familial ALS also can kill motoneurons when co-cultured[67].AbA cells in culture demonstrate increased proliferation and lack of replicative senescence, thus suggesting aberrant contact inhibition, and therefore a dysregulated ECM that fails to regulate the cells to a normal phenotype[66].

**OLIGODENDROCYTES**

Oligodendrocytes are severely affected in ALS and their degeneration has been shown to precede motor neuron death in the mutant SOD1 (mSOD1) mouse model[68]. Also, oligodendrocyte progenitors rapidly proliferate in the spinal cord of mSOD1G93A mice, but fail to replace degenerating oligodendrocytes, thus leaving motor neuron axons demyelinated. Recent studies suggest that SOD1 in ALS models is directly or indirectly contributing to oligodendrocyte pathology, and suggest that in this cell type some of the damage is irreversible[69].

**PRION-LIKE PROTEINS**

Misfolded proteins in the ALS model have similarities to prions. Key to the SOD1 and TDP43 proteins in ALS is that they self-template themselves, meaning that the proteins replicate themselves without the involvement of DNA and RNA, and then spread to other parts of the nervous system. These prion-like proteins will spread not only throughout the cell, but traveling in exosomes spread from cell to cell[70]. However, unlike true prions, the ALS prion-like proteins are able to be refolded or cleared from the body, correcting the destructive properties of the protein[71].

Protein-RNA assemblies serve as the foci of protein misfolding and their maturation into insoluble structures in the ALS state. The assemblies then recruit native proteins, turning them into misfolded forms. This self-perpetuation of misfolding proteins is a “twisted version of classical prion replication” that leads to amplification of pathological protein complexes that spread throughout the neuraxis, offering a pathogenic principle that underlies the rapid disease progression that characterizes ALS, and other neurodegenerative diseases[72].

**CYTOSKELETON**

The cytoskeleton in general[73] and actin[74], microtubules and neurofilaments[75] in particular, are believed to be dysregulated in ALS. Understanding that the rescue of neurons through the delivery of HSP and mitochondria is dependent on the ECM and actin-based tunnelling nanotubes (TNT) emanating from the cytoskeleton, and conveyed though the ECM of surrounding healthy stem cells, we can well believe that motor neurons in ALS are put at extreme risk by the interruption of the ECM/cytoskeleton.

**BASEMENT MEMBRANE – BLOOD BRAIN BARRIER DYSFUNCTION**

Basement membrane dysfunction leading to irregularities in blood/nutrient flow to neurons is a hallmark of macular degeneration, and has recently become evident in ALS[76]. Here again, breakdown of ECM can lead to vascular irregularities in the motor tracts and induce an inflammatory state. In the healthy adult with a stable ECM and basement membrane on which to attach, microglia are ramified and relatively quiescent. With breakdown of the basement membrane, microglia become mobile and reactive, and can further degrade the ECM[77]. A chronic para-inflammatory state in the motor tracts ensues. Blood-derived neurotoxic hemoglobin and iron accumulate in the spinal cord, leading to early motor-neuron degeneration in SOD1G93A mice. Chelation of blood-derived iron and antioxidant treatment early in the ALS sequalae mitigated early motor-neuronal injury[78]. Chronic inflammation with MMP-9 presence, as an example, can lead to laminin-III breakdown and a resulting breakdown of the blood brain barrier. Transplant of bone marrow stem cells into a mouse model of ALS repaired some of the vascular pathology[76].

**ACHE – LOSS OF FUNCTION AT THE SYNAPSE BECAUSE ACHE BECOMES UNBOUND FROM THE ECM AT NMJ**

Interestingly, muscle biopsies revealed a reduction in the AChE level of ALS patients[79], and analysis of the patient’s plasma revealed a large increase in circulating AChE[80]. The increase in circulating AChE may reflect a disruption of extracellularly bound AChE at the NMJ[81], leading to functional deficits in the muscle. Although in a study where nerve-muscle integrity was altered, and a similar reduction of AChE at the animals NMJ was measured, with an associated increase in plasma levels[82,83], the exact source of AChE increase in ALS plasma has not been demonstrated. Again, the data are suggestive of an early disruption of ECM, including that of the NMJ.

**STRESS**

Traditionally we considered inflammation as a defense in response to infection or injury. Inflammation, however, is often induced by tissue stress and malfunction in the absence of infection or overt tissue damage. Stress as a chronic phenomenon, will elicit a chronic, self-perpetuating para-inflammatory state with an active immune response.

Interestingly, activation of autoreactive T-cell receptors by non-commensal microbes might be a common trigger of autoimmune diseases in the immunoprivileged CNS[84]. This means that stress, and other factors that degrade the microbiota of the gut can have devastating long term effects through chronic inflammation and or elicit a full-blown autoimmune response in the nervous system.

Although the data are poorly collected in third world nations, the incidence of ALS in underdeveloped countries is reportedly lower. This could be due to many factors, including less stress and fewer dangerous chemicals in the environment[85].

**EXPOSOME**

Chemical exposure is thought to contribute about 70% of our health state[86]. Exposure to various chemicals, including PCBs and pesticides has been correlated to human ALS, but not to their matched controls[3]. The specificity with which pesticides and other environmental chemicals can act to destroy components of the nervous system is exemplified by the chronic exposure of the brain to the lipophilic pesticide rotenone causing a specific loss of dopaminergic neurons[87].

**PGC-1α**

Mesenchymal stem cells better engraft into host tissue when they overexpress PGC-1α[88]. Exosomes from adipose-derived stem cells ameliorate Huntington's disease phenotypes in an *in vitro* model through the activation of PGC-1α[89]. This has yet to be explored in ALS.

**SPATACSIN**

In one study of Spatacsin, variants of the spatascin gene were associated with amyotrophic lateral sclerosis neuropathology in one member of one of the 10 families studied[90]. Given the limited tissue samples available for genetic analysis, and the lack of RNA or protein anlaysis, while the data warrant further study, no conclusions from this one study can be made.

**LEAKY GUT AND DYSBIOSIS**

Intestinal homeostasis and the microbiome have recently been shown to play essential roles in neurological diseases, such as Parkinson's disease and ALS. Animal model studies suggests a potential role of the intestinal epithelium and microbiome in the very early progression of ALS[21] The ALS mouse model, G93A, which expresses mutant superoxide dismutase (SOD1G93A), showed a number of gut abnormalities. Damaged tight junction structure was present leading to increased permeability. Also, significant reductions in the protein expression levels of tight junction ZO‐1 and the adherens junction E‐cadherin were seen. Increased numbers of abnormal Paneth cells were reported. Because Paneth cells are specialized epithelial cells that can sense microbes and secrete antimicrobial peptides, they are key to host innate immune responses and helping to shape the gut microbiome. A decreased level of defensin 5 alpha, an antimicrobial peptide, was also found in the ALS intestine. A shift in the profile of the intestinal microbiome was also observed, including reduced levels of *Butyrivibrio Fibrisolvens,* *Escherichia coli*, and *Fermicus*, in the G93A mice. Increased expression of the inflammatory cytokine IL-17, and reduced levels of autophagic lysozyme 1, which leads to a reduced ability to clear misfolded proteins) were additional attributes of a disrupted microbiome. Importantly, the dysbiotic properties of the gut were present when the mice were 2 months old, before any other symptoms of ALS had yet developed. A comparison of healthy subjects *vs* ALS patients also has shown dysbiosis in the ALS patients with a decrease in healthy microbes and decrease in unhealthy microbes of the intestine[91].

Dysbiosis of the gut may also be involved in the misfolded alpha-synuclein (AS) and other proteins involved in neurodegenerative disorders that display prion-like transmission of protein aggregation. The role of amyloid proteins made by gut microbiota in the development of ALS was studied using aged rats and transgenic C. elegans. Both animals were exposed to E. coli producing the extracellular bacterial amyloid protein curli. Rats exposed to curli-producing bacteria displayed increased neuronal AS deposition in both gut and brain. Also observed was enhanced microgliosis and astrogliosis compared to rats exposed to either mutant bacteria unable to synthesize curli, or to vehicle alone[92]. The study showed that bacterial amyloid functions as a trigger to initiate AS aggregation through cross-seeding, while also priming responses of the innate immune system.

Relevant to ALS, and neurodegeneration in general, a probiotic mixture of 8 different strains of bacteria, namely Streptococcus thermophilus DSM24731, Bifidobacterium breve DSM24732, Bifidobacterium longum DSM24736, Bifidobacterium infantis DSM24737, Lactobacillus acidophilus DSM24735, Lactobacillus plantarum DSM24730, Lactobacillus paracasei DSM24733, Lactobacillus delbrueckii subspecies Bulgaricus DSM24734, has shown reparation of leaky gut and a concomitant improvement in a number of neural functions in aged animal models of neurodegenerative disease[93].

**AGE/RAGE**

Fibroblasts are abundant in most tissues and in particular the nervous system. They are crucial for homeostatic maintenance and for building the ECM, as well as pathological ECM alterations observed in tissue[94].Proteins in the ECM with a long half-life, such as collagen, can nonenzymatically react with high levels of glucose to form advanced glycation end products (AGEs). AGE-modified collagen increases stiffness of the matrix making the matrix resistant to hydrolytic turnover, resulting in ECM proteins accumulating in the microenvironment of cells. In addition, AGE will activate the receptor called RAGE (The receptor for advanced glycation endproducts). RAGE activation in turn can negatively impact normal ECM formation[95].

Many lines of evidence suggest that inflammatory responses play a critical role in the pathogenesis of motor neuron degeneration in ALS. RAGE is a key component in regulation of both innate and adaptive immunity in different pathologies associated with inflammation. RAGE mediates inflammatory responses and microglia stimulation in the brain[96], leading to neuronal damage, neurodegeneration, and resulting in symptomatic brain disorders. Immunohistochemical analysis of RAGE in control and ALS patients showed increased expression in reactive glial cells in both gray (ventral horn) and white matter in the spinal cord[97]. Expression of RAGE is also higher in the SOD1 transgenic mouse model of ALS *vs* wild-type mouse spinal cord. Further, treatment of SOD1 transgenic mice with soluble RAGE (sRAGE), a natural competitor of RAGE that sequesters RAGE ligands and blocks their interaction with cell surface RAGE, significantly delays the progression of ALS and prolongs life span as compared to vehicle treatment[98].

Part of the therapeutic strategy of S2RM is to return proteostasis to a normal state using the S2RM technology, thus rebuilding the ECM to a normal state, and reducing the deleterious build-up of AGE and the subsequent activation of RAGE.

**WHAT LEADS TO BREAKDOWN OF THE ECM**

Environmental toxins, xenobiotic electrophiles for example, can disrupt DNA and protein function and degrade ECM[99]. Damage to ECM triggers inflammation and further degradation of the extracellular matrix (ECM). The ECM is an intricately arranged biochemical and mechanical scaffold composed of secreted proteins and complex sugars that collectively support cell function and survival. Some ECM molecules become aberrantly expressed following damage, whereas other proteins are cleaved into bioactive fragments known as damage-associated molecular patterns (DAMPs) or “alarmins”. The DAMPs are able to bind different types of pattern recognition receptors (PRRs), and can influence the phenotype and magnitude of inflammation. Further, the enzymes and inflammatory mediators released by immune cells resulting from the damage further degrade or alter the composition of the ECM[100].

Knowing that the breakdown of the ECM/cytoskeleton is a key determinant in ALS, we can then ask, what causes the breakdown of the ECM/cytoskeleton? The exposome, *i.e.,* chemical exposure, is thought to contribute about 70% of health state[86]. This coupled with lifestyle, stress, and the aging process itself are all risk factors. To summarize, the following factors are known to cause ECM breakdown: (1) chemical exposure – numerous mechanisms, including breakdown of matrix; (2) stress - signals originating in the sympathetic nervous system can suppress osteoblast function and control the attraction of stem cells to their niche[101]. Stress, acting through cortisol directly breaks down ECM; (3) aging – degradation of ECM, loss of stem cell function; (4) glycation – binding of sugars with ECM causing ECM dysfunction[98]; and (5) inflammation, MMPs – chronic para-inflammation or inflammation as a result of ECM breakdown leads to further inflammation and an activation of macrophages (microglia) and a release of many molecules, including MMPs that will exacerbate the inflammatory state thus damaging neurons and muscle.

Collectively these studies suggest that a key determinant of ALS is a dysfunctional ECM and cytoskeleton. I now present the rationale for our therapeutic approach based on the evidence.

**S2RM METHODOLOGY FOR TREATING ALS**

Simply stated, I believe the predominant cause of ALS is a breakdown of the ECM/microenvironment in the motor neuron tracts of the nervous system; all other mechanism underlying ALS derive from this breakdown. As a major consequence of the ECM breakdown, HSP and chaperone proteins from neighboring cells cannot supply the motor neurons with their necessary proteins that enable autophagy, and proper protein folding. These results are deadly for motor neurons.

My strategy uses a combination of stem cell types to treat ALS. And instead of injecting cells into the patient, we inject the molecules from the multiple cell types into the patient. Here’s why. The molecules released from the stem cells are doing all of the work in treating ALS, and the cells are difficult and expensive to work with outside of the laboratory setting, making clinical procedures with the stem cells onerous. Further, once the cells are injected into the patient, the functionality of the stem cells is not measurable, and we know that most stem cells injected into patients are not functional shortly after implantation, failing to make a live graft[102]. Therefore, our strategy is to grow and stimulate the stem cells in the laboratory to optimize the set of SRM, collect the therapeutic molecules that the stem cells release, and then use those molecules to directly treat the patient. In this manner, we know the “healing molecules” are being delivered to the patient in a defined manner, both in space and in time. The molecules we use are derived from several types of stem cells known to be active in the brain, to stimulate neuron and glial cell growth[103], build the PNNs, set the ECM to a permissive state for neuro-regeneration, and to correct some of the mechanistic failures and alleviate the symptoms of ALS (Figure 3).

**S2RM DELIVERED IN EXOSOMES**

As stated above, exosomes are key to this technology given their protective, penetrating, and delivery qualities. They are also immunoprivileged, and have been shown to be used successfully for patient to patient injection without negative immune reaction. Recently, exosomes from human cancer patients have been demonstrated to protect their protein contents from dephosphorylation from endogenous phosphatases. The exosomes were successfully used for protein-based diagnostics of cancer patients for up to 5 years following sampling from the patient[104]. This is strong evidence for the ability of our exosome-based therapeutic to be protected in the body as it courses to the target site in the nervous system of the ALS patient.

**CELLS USED TO CREATE S2RM-EXOSOME (1-4 ADMINSTERED INTRANASAL, 1-5 ADMINSTERED INTRATHECAL)**

First, the use of allogeneic MSCs or their exosomes have been used to treat a number of patients to treat graft-*vs*-host disease with success, meaning that stem cells themselves, and especially their exosomes, will not elicit a negative immune reaction[105-107]. Because of their small size, unlike the stem cells themselves, exosomes can easily be sterilized by passing them through a 0.22-µm sterile filter. The cell types used to develop the therapeutic are as follows: (1) bone marrow mesenchymal stem cells (BMSC) – conditioned media induces angiogenesis and rescue mitochondrial function – these cells are stimulated and cultured in a special way, explained in methods section below (Figures 4 and 5); (2) neural progenitor cells – neural growth factors; (3) fetal fibroblasts – less than 24 wk post-gestation (secreting growth factors and building blocks of ECM and PNN, allowing a permissive environment for cellular reprogramming of the afflicted motor neurons and astrocytes); (4) fetal astrocytes – homeostasis of motor neuron support cells; (5) satellite cells (muscle stem cells) – known to express HSP. Administered only at spine through Intrathecal Injection. The SRM from the cells listed as 1-4 are collected and can be dosed intra-nasally to the patient. The SRM from the cells listed in 1-5 are collected and can dosed by intrathecal injection to the patient.

**CAN THE DISEASE BE CURED? CONSIDERATIONS OF ALS AS A CHRONIC PARA-INFLAMMATORY STATE AND HOMEOSTATIC RENORMALIZATION**

Tissue damage, independent of infection, such as breakdown of the ECM, Basal Lamina, and PeriNeuronal Nets, can induce a tissue-repair response that involves the activation of both innate and adaptive immunity, such as activation of regulatory T cells, group 2 innate lymphoid cells (ILC2 cells), eosinophils, and M2 macrophages[108]. This can lead to a state of chronic parainflammation that is self-perpetuating. Let me explain. Between basal homeostatic conditions and true inflammation a “para-inflammation” state exists[109]. Para-inflammation is an adaptive response of the immune system to low levels of tissue damage, such as in aging whereby oxidative stress or cortisol release for example, accumulates for many decades. The physiological role of para-inflammation is to maintain homeostasis (or re-set the homeostatic threshold of the tissue) and restore tissue functionality. Unfortunately, because of aging, and living in the industrial world with all of the newly created stressors and an exposome that lacks a corresponding evolved homeostatic renormalization function in our bodies, homeostasis is constantly disrupted. This para-inflammation theory helps to explain many phenomena observed in various chronic disease conditions, an example of which is “inflammaging”[110].Why the ECM and Peri-Neuronal Nets will be differentially affected and cause ALS will be hard to figure, but is likely to involve the exposome, what you’ve been exposed to during your lifetime, at conception, during development, and throughout your adult life, along with other lifestyle factors such as stress, exercise, and diet. Further, epigenetic regulation of your parents, and their parents, by what they did and what they encountered in their environments will also have major consequences to your health[111-114]. This is termed transgenerational epigenetic inheritance. For example, even dietary effects at the epigenetic level can be passed for three generations through an RNA-based mechanism[115,116], and the drugs consumed by the father[117]. The types, quantities, and timing of these factors are likely to be critical in determining whether one will develop ALS, or any other disease of inflammation and aging.

**HOMEOSTATIC RENORMALIZATION USING S2RM**

The chronic parainflammatory state underlying ALS can best be treated with S2RM technology to renormalize homeostasis. The renormalization process, which includes proteostatic renormalization to rebuild the ECM, TNT, and Peri Neuronal Nets will allow for key HSP and chaperone proteins, along with protein conglomerates called proteasomes, to fold, refold, repair, or expunge the misfolded and damaged proteins, SOD1, DG-1, TDP-43, that lead to ALS. An imbalance in proteostasis leads to many dysfunctions including, for example, inhibition of FGFBP1 expression in muscle cells by increased accumulation of the transforming growth factor-β1 (TGF-β1) in skeletal muscles and at their NMJs. In the absence of FGFBP1, NMJs exhibit structural abnormalities[118].

Further, mitochondrial function will be repaired in the neurons and muscles, and TNTs to transfer healthy mitochondria from healthy cells to compromised cells reestablished – this too is critical in ALS therapy because mitochondria are damaged[119] by phylogenetically bacterial symbionts of early eukaryotic cells, that when damaged, release mitochondrial damage-associated molecular patterns (formyl peptides and mitochondrial DNA) with evolutionarily conserved similarities to bacterial pathogen-associated molecular patterns. These are released into the circulation and are powerful activators of innate immunity and Nlrp3 inflammasome[120,121]. Fitting with our hypothesis and treatment strategy, formyl peptides (chemotactic peptides), for example, rearrange the cytoskeleton and change migration patterns of cells, likely interrupting the cell to cell connections that convey HSP, chaperones, exosomes, and mitochondria from healthy stem cells to damaged motor neurons and muscles. These protein factors, along with key microRNA[122] to renormalize dysregulated microRNA in the ALS patient[123], contained in the exosomes of S2RM, are the major factors in S2RM therapeutics for ALS.

**SPECIAL CULTURE TECHNOLOGY**

Our strategy is to use only the molecules that are fully formed and secreted by the stem cells. Crushing the cells and extracting the molecules by harsh techniques is inadvisable as incomplete and misfolded proteins and other molecules may be extracted in this manner. Incomplete and misfolded proteins as we have seen in the aforementioned studies are exactly what we want to avoid in ALS. Stem cells secrete many molecules into the media, and also a rich self-assembly of some of those molecules forms an ECM on the bottom of the culture dish.

Figure 4 diagrams how HSP is induced in MSCs, and how TNTs are formed between the stressed cells and the normal cells. The culture procedure has two steps: (1) MSCs are cultured under normal conditions, but pulsed with higher temperatures at 39 °C for 24 h in order to slightly stress the cells so the HSPs are formed. On completing of the 24 h stress period at 39 °C, the cells are then removed from the culture vessel using standard techniques so that they can be co-cultured with normal, non-stressed MSCs; and (2) the stressed MSCs are then co-cultured with normal MSCs at a ratio of 1 normal cell to 4 stressed cells. Under these conditions TNTs will form between the normal and stressed cells using actin and microtubules emanating from the stressed cells and guided through the developing ECM at the bottom of the dish that serves as a matrix for the growing cells. In this manner, HSPs, mitochondria, and exosomes can transfer from one cell to another.

Our strategy is to: (1) capture the conditioned media with all of its soluble molecules, as well as the molecules in the exosomes; and (2) capture all of the molecules that have formed the ECM at the bottom of the dish by the process shown below in Figure 5. Although our therapeutic is protected by issued and pending patents and trade secrets, we can disclose here that our therapeutic is developed using the SRM from five different types of adult stem cells, including lineages from mesenchyme and nervous system.

**CONCLUSION**

ALS is a disease of protein dysfunction without an RNA or DNA component. Extrinsic factors, including the exposome, cause disruption of proteins that build and maintain the ECM. Once the ECM is disrupted, the support function of stem cells, including neural stem cells, which maintain the proper folding of proteins in the nervous system is degraded. Once the proteins in the nervous system misfold, the misfolded proteins self-template themselves and act in a prion-like manner to spread and destroy neurons. The best way to treat this disease of degraded ECM and misfolded proteins is to restore homeostasis, and in particular, proteostasis, so that the ECM and proper folding of proteins is restored. Proteostasis will allow, for example, tunneling nanotube formation and the transfer of mitochondria from healthy cells to those in need of recue[124]. Homeostatic renormalization is accomplished by administering S2RM stem cell molecule technology to the nervous system.

**REFERENCES**

1. **Banack SA**, Metcalf JS, Bradley WG, Cox PA. Detection of cyanobacterial neurotoxin β-N-methylamino-l-alanine within shellfish in the diet of an ALS patient in Florida. *Toxicon* 2014; **90**: 167-173 [PMID: 25123936 DOI: 10.1016/j.toxicon.2014.07.018]
2. **Delzor A**, Couratier P, Boumédiène F, Nicol M, Druet-Cabanac M, Paraf F, Méjean A, Ploux O, Leleu JP, Brient L, Lengronne M, Pichon V, Combès A, El Abdellaoui S, Bonneterre V, Lagrange E, Besson G, Bicout DJ, Boutonnat J, Camu W, Pageot N, Juntas-Morales R, Rigau V, Masseret E, Abadie E, Preux PM, Marin B. Searching for a link between the L-BMAA neurotoxin and amyotrophic lateral sclerosis: a study protocol of the French BMAALS programme. *BMJ Open* 2014; **4**: e005528 [PMID: 25180055 DOI: 10.1136/bmjopen-2014-005528]
3. **Su FC**, Goutman SA, Chernyak S, Mukherjee B, Callaghan BC, Batterman S, Feldman EL. Association of Environmental Toxins With Amyotrophic Lateral Sclerosis. *JAMA Neurol* 2016; **73**: 803-811 [PMID: 27159543 DOI: 10.1001/jamaneurol.2016.0594]
4. **Xi Z**, Zinman L, Moreno D, Schymick J, Liang Y, Sato C, Zheng Y, Ghani M, Dib S, Keith J, Robertson J, Rogaeva E. Hypermethylation of the CpG island near the G4C2 repeat in ALS with a C9orf72 expansion. *Am J Hum Genet* 2013; **92**: 981-989 [PMID: 23731538 DOI: 10.1016/j.ajhg.2013.04.017]
5. **Paez-Colasante X**, Figueroa-Romero C, Sakowski SA, Goutman SA, Feldman EL. Amyotrophic lateral sclerosis: mechanisms and therapeutics in the epigenomic era. *Nat Rev Neurol* 2015; **11**: 266-279 [PMID: 25896087 DOI: 10.1038/nrneurol.2015.57]
6. **Li W**, Lee MH, Henderson L, Tyagi R, Bachani M, Steiner J, Campanac E, Hoffman DA, von Geldern G, Johnson K, Maric D, Morris HD, Lentz M, Pak K, Mammen A, Ostrow L, Rothstein J, Nath A. Human endogenous retrovirus-K contributes to motor neuron disease. *Sci Transl Med* 2015; **7**: 307ra153 [PMID: 26424568 DOI: 10.1126/scitranslmed.aac8201]
7. **Su FC**, Goutman SA, Chernyak S, Mukherjee B, Callaghan BC, Batterman S, Feldman EL. Association of Environmental Toxins With Amyotrophic Lateral Sclerosis. *JAMA Neurol* 2016; **73**: 803-811 [PMID: 27159543 DOI: 10.1001/jamaneurol.2016.0594]
8. **Andrew AS**, Caller TA, Tandan R, Duell EJ, Henegan PL, Field NC, Bradley WG, Stommel EW. Environmental and Occupational Exposures and Amyotrophic Lateral Sclerosis in New England. *Neurodegener Dis* 2017; **17**: 110-116 [PMID: 28122372 DOI: 10.1159/000453359]
9. **Seneff S,** Morley WA, Hadden MJ, Michener MC. Does glyphosate acting as a glycine analogue contribute to ALS? *J Bioinfo Proteomics Rev* 2017; **3:** 1-21 [DOI: 10.15436/2381-0793.16.1173]
10. **Cattani D**, de Liz Oliveira Cavalli VL, Heinz Rieg CE, Domingues JT, Dal-Cim T, Tasca CI, Mena Barreto Silva FR, Zamoner A. Mechanisms underlying the neurotoxicity induced by glyphosate-based herbicide in immature rat hippocampus: involvement of glutamate excitotoxicity. *Toxicology* 2014; **320**: 34-45 [PMID: 24636977 DOI: 10.1016/j.tox.2014.03.001]
11. **Patchin ES**, Anderson DS, Silva RM, Uyeminami DL, Scott GM, Guo T, Van Winkle LS, Pinkerton KE. Size-Dependent Deposition, Translocation, and Microglial Activation of Inhaled Silver Nanoparticles in the Rodent Nose and Brain. *Environ Health Perspect* 2016; **124**: 1870-1875 [PMID: 27152509 DOI: 10.1289/EHP234]
12. **Valera E**, Masliah E. Combination therapies: The next logical Step for the treatment of synucleinopathies? *Mov Disord* 2016; **31**: 225-234 [PMID: 26388203 DOI: 10.1002/mds.26428]
13. **Maguire G**. Systems biology approach to developing "systems therapeutics". *ACS Med Chem Lett* 2014; **5**: 453-455 [PMID: 24900858 DOI: 10.1021/ml5000614]
14. **Ringholz GM**, Appel SH, Bradshaw M, Cooke NA, Mosnik DM, Schulz PE. Prevalence and patterns of cognitive impairment in sporadic ALS. *Neurology* 2005; **65**: 586-590 [PMID: 16116120 DOI: 10.1212/01.wnl.0000172911.39167.b6]
15. **Forostyak S**, Homola A, Turnovcova K, Svitil P, Jendelova P, Sykova E. Intrathecal delivery of mesenchymal stromal cells protects the structure of altered perineuronal nets in SOD1 rats and amends the course of ALS. *Stem Cells* 2014; **32**: 3163-3172 [PMID: 25113670 DOI: 10.1002/stem.1812]
16. **Chen TL**, Zhu GL, Wang JA, Wang Y, He XL, Jiang J. Apoptosis of bone marrow mesenchymal stem cells caused by hypoxia/reoxygenation via multiple pathways. *Int J Clin Exp Med* 2014; **7**: 4686-4697 [PMID: 25663966]
17. **Maguire G**. Therapeutics from Adult Stem Cells and the Hype Curve. *ACS Med Chem Lett* 2016; **7**: 441-443 [PMID: 27190588 DOI: 10.1021/acsmedchemlett.6b00125]
18. **Wood WA**, Krishnamurthy J, Mitin N, Torrice C, Parker JS, Snavely AC, Shea TC, Serody JS, Sharpless NE. Chemotherapy and Stem Cell Transplantation Increase p16<sup>INK4a</sup> Expression, a Biomarker of T-cell Aging. *EBioMedicine* 2016; **11**: 227-238 [PMID: 27591832 DOI: 10.1016/j.ebiom.2016.08.029]
19. **Harrison DE**. Proliferative capacity of erythropoietic stem cell lines and aging: an overview. *Mech Ageing Dev* 1979; **9**: 409-426 [PMID: 37376]
20. **Kang E**, Wang X, Tippner-Hedges R, Ma H, Folmes CD, Gutierrez NM, Lee Y, Van Dyken C, Ahmed R, Li Y, Koski A, Hayama T, Luo S, Harding CO, Amato P, Jensen J, Battaglia D, Lee D, Wu D, Terzic A, Wolf DP, Huang T, Mitalipov S. Age-Related Accumulation of Somatic Mitochondrial DNA Mutations in Adult-Derived Human iPSCs. *Cell Stem Cell* 2016; **18**: 625-636 [PMID: 27151456 DOI: 10.1016/j.stem.2016.02.005]
21. **Wu S**, Yi J, Zhang YG, Zhou J, Sun J. Leaky intestine and impaired microbiome in an amyotrophic lateral sclerosis mouse model. *Physiol Rep* 2015; **3**: pii: e12356 [PMID: 25847918 DOI: 10.14814/phy2.12356]
22. **Veeramachaneni V**, Makałowski W, Galdzicki M, Sood R, Makałowska I. Mammalian overlapping genes: the comparative perspective. *Genome Res* 2004; **14**: 280-286 [PMID: 14762064 DOI: 10.1101/gr.1590904]
23. **Keller EF**. The postgenomic genome. In: Altman R, Ankeny RA, Bliss C, Dupré J, Fortun M, Keller EF, Leonelli S, Mackenzie A, Moinester M, Panofsky A, Shostak S. Postgenomics: Perspectives on Biology after the Genome. Duke University Press; 2015 [DOI: 10.1215/9780822375449-002]
24. **Chang CQ**, Yesupriya A, Rowell JL, Pimentel CB, Clyne M, Gwinn M, Khoury MJ, Wulf A, Schully SD. A systematic review of cancer GWAS and candidate gene meta-analyses reveals limited overlap but similar effect sizes. *Eur J Hum Genet* 2014; **22**: 402-408 [PMID: 23881057 DOI: 10.1038/ejhg.2013.161]
25. **Nobel D**. The Music of Life. Oxford University Press; 2006
26. **Nobel D**.Dance to the Tune of Life: Biological Relativity. Cambridge University Press, Cambridge UK; 2017
27. **Rappaport SM**. Genetic Factors Are Not the Major Causes of Chronic Diseases. *PLoS One* 2016; **11**: e0154387 [PMID: 27105432 DOI: 10.1371/journal.pone.0154387]
28. **Miller EC**, Miller JA. The presence and significance of bound aminoazo dyes in the livers of rats fed p-dimethylaminoazobenzene. *Cancer Res* 1947; **7**: 468-480
29. **Liebler DC**. Protein damage by reactive electrophiles: targets and consequences. *Chem Res Toxicol* 2008; **21**: 117-128 [PMID: 18052106 DOI: 10.1021/tx700235t]
30. **Renton AE**, Majounie E, Waite A, Simón-Sánchez J, Rollinson S, Gibbs JR, Schymick JC, Laaksovirta H, van Swieten JC, Myllykangas L, Kalimo H, Paetau A, Abramzon Y, Remes AM, Kaganovich A, Scholz SW, Duckworth J, Ding J, Harmer DW, Hernandez DG, Johnson JO, Mok K, Ryten M, Trabzuni D, Guerreiro RJ, Orrell RW, Neal J, Murray A, Pearson J, Jansen IE, Sondervan D, Seelaar H, Blake D, Young K, Halliwell N, Callister JB, Toulson G, Richardson A, Gerhard A, Snowden J, Mann D, Neary D, Nalls MA, Peuralinna T, Jansson L, Isoviita VM, Kaivorinne AL, Hölttä-Vuori M, Ikonen E, Sulkava R, Benatar M, Wuu J, Chiò A, Restagno G, Borghero G, Sabatelli M; ITALSGEN Consortium, Heckerman D, Rogaeva E, Zinman L, Rothstein JD, Sendtner M, Drepper C, Eichler EE, Alkan C, Abdullaev Z, Pack SD, Dutra A, Pak E, Hardy J, Singleton A, Williams NM, Heutink P, Pickering-Brown S, Morris HR, Tienari PJ, Traynor BJ. A hexanucleotide repeat expansion in C9ORF72 is the cause of chromosome 9p21-linked ALS-FTD. *Neuron* 2011; **72**: 257-268 [PMID: 21944779 DOI: 10.1016/j.neuron.2011.09.010]
31. **Koppers M**, Blokhuis AM, Westeneng HJ, Terpstra ML, Zundel CA, Vieira de Sá R, Schellevis RD, Waite AJ, Blake DJ, Veldink JH, van den Berg LH, Pasterkamp RJ. C9orf72 ablation in mice does not cause motor neuron degeneration or motor deficits. *Ann Neurol* 2015; **78**: 426-438 [PMID: 26044557 DOI: 10.1002/ana.24453]
32. **Bandyopadhyay U**, Cotney J, Nagy M, Oh S, Leng J, Mahajan M, Mane S, Fenton WA, Noonan JP, Horwich AL. RNA-Seq profiling of spinal cord motor neurons from a presymptomatic SOD1 ALS mouse. *PLoS One* 2013; **8**: e53575 [PMID: 23301088 DOI: 10.1371/journal.pone.0053575]
33. **Bosco DA**, Morfini G, Karabacak NM, Song Y, Gros-Louis F, Pasinelli P, Goolsby H, Fontaine BA, Lemay N, McKenna-Yasek D, Frosch MP, Agar JN, Julien JP, Brady ST, Brown RH Jr. Wild-type and mutant SOD1 share an aberrant conformation and a common pathogenic pathway in ALS. *Nat Neurosci* 2010; **13**: 1396-1403 [PMID: 20953194 DOI: 10.1038/nn.2660]
34. **Mizuno H**, Warita H, Aoki M, Itoyama Y. Accumulation of chondroitin sulfate proteoglycans in the microenvironment of spinal motor neurons in amyotrophic lateral sclerosis transgenic rats. *J Neurosci Res* 2008; **86**: 2512-2523 [PMID: 18438943 DOI: 10.1002/jnr.21702]
35. **Gubbiotti MA**, Iozzo RV. Proteoglycans regulate autophagy via outside-in signaling: an emerging new concept. *Matrix Biol* 2015; **48**: 6-13 [PMID: 26462577 DOI: 10.1016/j.matbio.2015.10.002]
36. **Chen ZL**, Yu H, Yu WM, Pawlak R, Strickland S. Proteolytic fragments of laminin promote excitotoxic neurodegeneration by up-regulation of the KA1 subunit of the kainate receptor. *J Cell Biol* 2008; **183**: 1299-1313 [PMID: 19114596 DOI: 10.1083/jcb.200803107]
37. **Miller RG**, Petajan JH, Bryan WW, Armon C, Barohn RJ, Goodpasture JC, Hoagland RJ, Parry GJ, Ross MA, Stromatt SC. A placebo-controlled trial of recombinant human ciliary neurotrophic (rhCNTF) factor in amyotrophic lateral sclerosis. rhCNTF ALS Study Group. *Ann Neurol* 1996; **39**: 256-260 [PMID: 8967757 DOI: 10.1002/ana.410390215]
38. A controlled trial of recombinant methionyl human BDNF in ALS: The BDNF Study Group (Phase III). *Neurology* 1999; **52**: 1427-1433 [PMID: 10227630]
39. **Petrou P**, Gothelf Y, Argov Z, Gotkine M, Levy YS, Kassis I, Vaknin-Dembinsky A, Ben-Hur T, Offen D, Abramsky O, Melamed E, Karussis D. Safety and Clinical Effects of Mesenchymal Stem Cells Secreting Neurotrophic Factor Transplantation in Patients With Amyotrophic Lateral Sclerosis: Results of Phase 1/2 and 2a Clinical Trials. *JAMA Neurol* 2016; **73**: 337-344 [PMID: 26751635 DOI: 10.1001/jamaneurol.2015.4321]
40. **Garbuzova-Davis S**, Sanberg PR. Blood-CNS Barrier Impairment in ALS patients versus an animal model. *Front Cell Neurosci* 2014; **8**: 21 [PMID: 24550780 DOI: 10.3389/fncel.2014.00021]
41. **Fischer LR**, Culver DG, Tennant P, Davis AA, Wang M, Castellano-Sanchez A, Khan J, Polak MA, Glass JD. Amyotrophic lateral sclerosis is a distal axonopathy: evidence in mice and man. *Exp Neurol* 2004; **185**: 232-240 [PMID: 14736504]
42. **Jokic N**, Gonzalez de Aguilar JL, Dimou L, Lin S, Fergani A, Ruegg MA, Schwab ME, Dupuis L, Loeffler JP. The neurite outgrowth inhibitor Nogo-A promotes denervation in an amyotrophic lateral sclerosis model. *EMBO Rep* 2006; **7**: 1162-1167 [PMID: 17039253 DOI: 10.1038/sj.embor.7400826]
43. **Dupuis L**, di Scala F, Rene F, de Tapia M, Oudart H, Pradat PF, Meininger V, Loeffler JP. Up-regulation of mitochondrial uncoupling protein 3 reveals an early muscular metabolic defect in amyotrophic lateral sclerosis. *FASEB J* 2003; **17**: 2091-2093 [PMID: 14500553 DOI: 10.1096/fj.02-1182fje]
44. **Pradat PF**, Barani A, Wanschitz J, Dubourg O, Lombès A, Bigot A, Mouly V, Bruneteau G, Salachas F, Lenglet T, Meininger V, Butler-Browne G. Abnormalities of satellite cells function in amyotrophic lateral sclerosis. *Amyotroph Lateral Scler* 2011; **12**: 264-271 [PMID: 21473708 DOI: 10.3109/17482968.2011.566618]
45. **Scaramozza A**, Marchese V, Papa V, Salaroli R, Sorarù G, Angelini C, Cenacchi G. Skeletal muscle satellite cells in amyotrophic lateral sclerosis. *Ultrastruct Pathol* 2014; **38**: 295-302 [PMID: 25079897 DOI: 10.3109/01913123.2014.937842]
46. **Jensen L**, Jørgensen LH, Bech RD, Frandsen U, Schrøder HD. Skeletal Muscle Remodelling as a Function of Disease Progression in Amyotrophic Lateral Sclerosis. *Biomed Res Int* 2016; **2016**: 5930621 [PMID: 27195289 DOI: 10.1155/2016/5930621]
47. **García-Prat L**, Martínez-Vicente M, Perdiguero E, Ortet L, Rodríguez-Ubreva J, Rebollo E, Ruiz-Bonilla V, Gutarra S, Ballestar E, Serrano AL, Sandri M, Muñoz-Cánoves P. Autophagy maintains stemness by preventing senescence. *Nature* 2016; **529**: 37-42 [PMID: 26738589 DOI: 10.1038/nature16187]
48. **Valero MC**, Huntsman HD, Liu J, Zou K, Boppart MD. Eccentric exercise facilitates mesenchymal stem cell appearance in skeletal muscle. *PLoS One* 2012; **7**: e29760 [PMID: 22253772 DOI: 10.1371/journal.pone.0029760]
49. **Hettwer S**, Lin S, Kucsera S, Haubitz M, Oliveri F, Fariello RG, Ruegg MA, Vrijbloed JW. Injection of a soluble fragment of neural agrin (NT-1654) considerably improves the muscle pathology caused by the disassembly of the neuromuscular junction. *PLoS One* 2014; **9**: e88739 [PMID: 24520420 DOI: 10.1371/journal.pone.0088739]
50. **Mazzon C**, Anselmo A, Cibella J, Soldani C, Destro A, Kim N, Roncalli M, Burden SJ, Dustin ML, Sarukhan A, Viola A. The critical role of agrin in the hematopoietic stem cell niche. *Blood* 2011; **118**: 2733-2742 [PMID: 21653324 DOI: 10.1182/blood-2011-01-331272]
51. **Magrì A**, Belfiore R, Reina S, Tomasello MF, Di Rosa MC, Guarino F, Leggio L, De Pinto V, Messina A. Hexokinase I N-terminal based peptide prevents the VDAC1-SOD1 G93A interaction and re-establishes ALS cell viability. *Sci Rep* 2016; **6**: 34802 [PMID: 27721436 DOI: 10.1038/srep34802]
52. **Brettschneider J**, Del Tredici K, Toledo JB, Robinson JL, Irwin DJ, Grossman M, Suh E, Van Deerlin VM, Wood EM, Baek Y, Kwong L, Lee EB, Elman L, McCluskey L, Fang L, Feldengut S, Ludolph AC, Lee VM, Braak H, Trojanowski JQ. Stages of pTDP-43 pathology in amyotrophic lateral sclerosis. *Ann Neurol* 2013; **74**: 20-38 [PMID: 23686809 DOI: 10.1002/ana.23937]
53. **Lim L**, Wei Y, Lu Y, Song J. ALS-Causing Mutations Significantly Perturb the Self-Assembly and Interaction with Nucleic Acid of the Intrinsically Disordered Prion-Like Domain of TDP-43. *PLoS Biol* 2016; **14**: e1002338 [PMID: 26735904 DOI: 10.1371/journal.pbio.1002338]
54. **Mitsumoto A**, Nakagawa Y. DJ-1 is an indicator for endogenous reactive oxygen species elicited by endotoxin. *Free Radic Res* 2001; **35**: 885-893 [PMID: 11811539]
55. **Canet-Avilés RM**, Wilson MA, Miller DW, Ahmad R, McLendon C, Bandyopadhyay S, Baptista MJ, Ringe D, Petsko GA, Cookson MR. The Parkinson's disease protein DJ-1 is neuroprotective due to cysteine-sulfinic acid-driven mitochondrial localization. *Proc Natl Acad Sci U S A* 2004; **101**: 9103-9108 [PMID: 15181200 DOI: 10.1073/pnas.0402959101]
56. **Mathiassen SG**, Larsen IB, Poulsen EG, Madsen CT, Papaleo E, Lindorff-Larsen K, Kragelund BB, Nielsen ML, Kriegenburg F, Hartmann-Petersen R. A Two-step Protein Quality Control Pathway for a Misfolded DJ-1 Variant in Fission Yeast. *J Biol Chem* 2015; **290**: 21141-21153 [PMID: 26152728 DOI: 10.1074/jbc.M115.662312]
57. **Richarme G**, Liu C, Mihoub M, Abdallah J, Leger T, Joly N, Liebart JC, Jurkunas UV, Nadal M, Bouloc P, Dairou J, Lamouri A. Guanine glycation repair by DJ-1/Park7 and its bacterial homologs. *Science* 2017; **357**: 208-211 [PMID: 28596309 DOI: 10.1126/science.aag1095]
58. **Brancia C**, Noli B, Boido M, Boi A, Puddu R, Borghero G, Marrosu F, Bongioanni P, Orrù S, Manconi B, D'Amato F, Messana I, Vincenzoni F, Vercelli A, Ferri GL, Cocco C. VGF Protein and Its C-Terminal Derived Peptides in Amyotrophic Lateral Sclerosis: Human and Animal Model Studies. *PLoS One* 2016; **11**: e0164689 [PMID: 27737014 DOI: 10.1371/journal.pone.0164689]
59. **Sama RR**, Fallini C, Gatto R, McKeon JE, Song Y, Rotunno MS, Penaranda S, Abdurakhmanov I, Landers JE, Morfini G, Brady ST, Bosco DA. ALS-linked FUS exerts a gain of toxic function involving aberrant p38 MAPK activation. *Sci Rep* 2017; **7**: 115 [PMID: 28273913 DOI: 10.1038/s41598-017-00091-1]
60. **Westergard T**, Jensen BK, Wen X, Cai J, Kropf E, Iacovitti L, Pasinelli P, Trotti D. Cell-to-Cell Transmission of Dipeptide Repeat Proteins Linked to C9orf72-ALS/FTD. *Cell Rep* 2016; **17**: 645-652 [PMID: 27732842 DOI: 10.1016/j.celrep.2016.09.032]
61. **Sabatelli M**, Zollino M, Conte A, Del Grande A, Marangi G, Lucchini M, Mirabella M, Romano A, Piacentini R, Bisogni G, Lattante S, Luigetti M, Rossini PM, Moncada A. Primary fibroblasts cultures reveal TDP-43 abnormalities in amyotrophic lateral sclerosis patients with and without SOD1 mutations. *Neurobiol Aging* 2015; **36**: 2005.e5-2005.e13 [PMID: 25792239 DOI: 10.1016/j.neurobiolaging.2015.02.009]
62. **Ip WKE**, Hoshi N, Shouval DS, Snapper S, Medzhitov R. Anti-inflammatory effect of IL-10 mediated by metabolic reprogramming of macrophages. *Science* 2017; **356**: 513-519 [PMID: 28473584 DOI: 10.1126/science.aal3535]
63. **Chiellini C**, Cochet O, Negroni L, Samson M, Poggi M, Ailhaud G, Alessi MC, Dani C, Amri EZ. Characterization of human mesenchymal stem cell secretome at early steps of adipocyte and osteoblast differentiation. *BMC Mol Biol* 2008; **9**: 26 [PMID: 18302751 DOI: 10.1186/1471-2199-9-26]
64. **Takeuchi T**, Suzuki M, Fujikake N, Popiel HA, Kikuchi H, Futaki S, Wada K, Nagai Y. Intercellular chaperone transmission via exosomes contributes to maintenance of protein homeostasis at the organismal level. *Proc Natl Acad Sci U S A* 2015; **112**: E2497-E2506 [PMID: 25918398 DOI: 10.1073/pnas.1412651112]
65. **Lee M**, Liu T, Im W, Kim M. Exosomes from adipose-derived stem cells ameliorate phenotype of Huntington's disease in vitro model. *Eur J Neurosci* 2016; **44**: 2114-2119 [PMID: 27177616 DOI: 10.1111/ejn.13275]
66. **Díaz-Amarilla P**, Olivera-Bravo S, Trias E, Cragnolini A, Martínez-Palma L, Cassina P, Beckman J, Barbeito L. Phenotypically aberrant astrocytes that promote motoneuron damage in a model of inherited amyotrophic lateral sclerosis. *Proc Natl Acad Sci U S A* 2011; **108**: 18126-18131 [PMID: 22010221 DOI: 10.1073/pnas.1110689108]
67. **Haidet-Phillips AM**, Hester ME, Miranda CJ, Meyer K, Braun L, Frakes A, Song S, Likhite S, Murtha MJ, Foust KD, Rao M, Eagle A, Kammesheidt A, Christensen A, Mendell JR, Burghes AH, Kaspar BK. Astrocytes from familial and sporadic ALS patients are toxic to motor neurons. *Nat Biotechnol* 2011; **29**: 824-828 [PMID: 21832997 DOI: 10.1038/nbt.1957]
68. **Philips T**, Bento-Abreu A, Nonneman A, Haeck W, Staats K, Geelen V, Hersmus N, Küsters B, Van Den Bosch L, Van Damme P, Richardson WD, Robberecht W. Oligodendrocyte dysfunction in the pathogenesis of amyotrophic lateral sclerosis. *Brain* 2013; **136**: 471-482 [PMID: 23378219 DOI: 10.1093/brain/aws339]
69. **Ferraiuolo L**, Meyer K, Sherwood TW, Vick J, Likhite S, Frakes A, Miranda CJ, Braun L, Heath PR, Pineda R, Beattie CE, Shaw PJ, Askwith CC, McTigue D, Kaspar BK. Oligodendrocytes contribute to motor neuron death in ALS via SOD1-dependent mechanism. *Proc Natl Acad Sci U S A* 2016; **113**: E6496-E6505 [PMID: 27688759 DOI: 10.1073/pnas.1607496113]
70. **Grad LI**, Pokrishevsky E, Silverman JM, Cashman NR. Exosome-dependent and independent mechanisms are involved in prion-like transmission of propagated Cu/Zn superoxide dismutase misfolding. *Prion* 2014; **8**: 331-335 [PMID: 25551548 DOI: 10.4161/19336896.2014.983398]
71. **Lee S**, Kim HJ. Prion-like Mechanism in Amyotrophic Lateral Sclerosis: are Protein Aggregates the Key? *Exp Neurobiol* 2015; **24**: 1-7 [PMID: 25792864 DOI: 10.5607/en.2015.24.1.1]
72. **Polymenidou M**, Cleveland DW. Biological Spectrum of Amyotrophic Lateral Sclerosis Prions. *Cold Spring Harb Perspect Med* 2017; Epub ahead of print [PMID: 28062558 DOI: 10.1101/cshperspect.a024133]
73. **Maximino JR**, de Oliveira GP, Alves CJ, Chadi G. Deregulated expression of cytoskeleton related genes in the spinal cord and sciatic nerve of presymptomatic SOD1(G93A) Amyotrophic Lateral Sclerosis mouse model. *Front Cell Neurosci* 2014; **8**: 148 [PMID: 24904291 DOI: 10.3389/fncel.2014.00148]
74. **Wu CH**, Fallini C, Ticozzi N, Keagle PJ, Sapp PC, Piotrowska K, Lowe P, Koppers M, McKenna-Yasek D, Baron DM, Kost JE, Gonzalez-Perez P, Fox AD, Adams J, Taroni F, Tiloca C, Leclerc AL, Chafe SC, Mangroo D, Moore MJ, Zitzewitz JA, Xu ZS, van den Berg LH, Glass JD, Siciliano G, Cirulli ET, Goldstein DB, Salachas F, Meininger V, Rossoll W, Ratti A, Gellera C, Bosco DA, Bassell GJ, Silani V, Drory VE, Brown RH Jr, Landers JE. Mutations in the profilin 1 gene cause familial amyotrophic lateral sclerosis. *Nature* 2012; **488**: 499-503 [PMID: 22801503 DOI: 10.1038/nature11280]
75. **Kaus A**, Sareen D. ALS Patient Stem Cells for Unveiling Disease Signatures of Motoneuron Susceptibility: Perspectives on the Deadly Mitochondria, ER Stress and Calcium Triad. *Front Cell Neurosci* 2015; **9**: 448 [PMID: 26635528 DOI: 10.3389/fncel.2015.00448]
76. **Garbuzova-Davis S**, Kurien C, Thomson A, Falco D, Ahmad S, Staffetti J, Steiner G, Abraham S, James G, Mahendrasah A, Sanberg PR, Borlongan CV. Endothelial and Astrocytic Support by Human Bone Marrow Stem Cell Grafts into Symptomatic ALS Mice towards Blood-Spinal Cord Barrier Repair. *Sci Rep* 2017; **7**: 884 [PMID: 28408761 DOI: 10.1038/s41598-017-00993-0]
77. **Vincent C**, Siddiqui TA, Schlichter LC. Podosomes in migrating microglia: components and matrix degradation. *J Neuroinflammation* 2012; **9**: 190 [PMID: 22873355 DOI: 10.1186/1742-2094-9-190]
78. **Winkler EA**, Sengillo JD, Sagare AP, Zhao Z, Ma Q, Zuniga E, Wang Y, Zhong Z, Sullivan JS, Griffin JH, Cleveland DW, Zlokovic BV. Blood-spinal cord barrier disruption contributes to early motor-neuron degeneration in ALS-model mice. *Proc Natl Acad Sci U S A* 2014; **111**: E1035-E1042 [PMID: 24591593 DOI: 10.1073/pnas.1401595111]
79. **Rasool CG**, Bradley WG, Connolly B, Baruah JK. Acetylcholinesterase and ATPases in motor neuron degenerative diseases. *Muscle Nerve* 1983; **6**: 430-435 [PMID: 6137769 DOI: 10.1002/mus.88006060]
80. **Festoff BW**, Fernandez HL. Plasma and red blood cell acetylcholinesterase in amyotrophic lateral sclerosis. *Muscle Nerve* 1981; **4**: 41-47 [PMID: 7231444 DOI: 10.1002/mus.880040108]
81. **Campanari ML**, García-Ayllón MS, Ciura S, Sáez-Valero J, Kabashi E. Neuromuscular Junction Impairment in Amyotrophic Lateral Sclerosis: Reassessing the Role of Acetylcholinesterase. *Front Mol Neurosci* 2016; **9**: 160 [PMID: 28082868 DOI: 10.3389/fnmol.2016.00160]
82. **Wilson BW**, Linkhart TA, Walker CR, Nieberg PS. Tissue acetylcholinesterase in plasma of chick embryos and dystrophic chickens. *J Neurol Sci* 1973; **18**: 333-350 [PMID: 4698314]
83. **Ferdandez HL**, Inestrosa NC. Role of axoplasmic transport in neurotrophic regulation of muscle end plate acetylcholinesterase. *Nature* 1976; **262**: 55-56 [PMID: 59316]
84. **Horai R**, Zárate-Bladés CR, Dillenburg-Pilla P, Chen J, Kielczewski JL, Silver PB, Jittayasothorn Y, Chan CC, Yamane H, Honda K, Caspi RR. Microbiota-Dependent Activation of an Autoreactive T Cell Receptor Provokes Autoimmunity in an Immunologically Privileged Site. *Immunity* 2015; **43**: 343-353 [PMID: 26287682 DOI: 10.1016/j.immuni.2015.07.014]
85. **Oskarsson B**, Horton DK, Mitsumoto H. Potential Environmental Factors in Amyotrophic Lateral Sclerosis. *Neurol Clin* 2015; **33**: 877-888 [PMID: 26515627 DOI: 10.1016/j.ncl.2015.07.009]
86. **Rappaport SM**, Smith MT. Epidemiology. Environment and disease risks. *Science* 2010; **330**: 460-461 [PMID: 20966241 DOI: 10.1126/science.1192603]
87. **Betarbet R,** Sherer TB, Kim JH, Baptista M, Cookson MR, Greenamyre JT. Rotenone models of Parkinson's disease: altered alpha-synuclein expression following chronic inhibition of complex I. Soc Neurosci 2002; 316: Abstr 386
88. **Lu D**, Zhang L, Wang H, Zhang Y, Liu J, Xu J, Liang Z, Deng W, Jiang Y, Wu Q, Li S, Ai Z, Zhong Y, Ying Y, Liu H, Gao F, Zhang Z, Chen B. Peroxisome proliferator-activated receptor-γ coactivator-1α (PGC-1α) enhances engraftment and angiogenesis of mesenchymal stem cells in diabetic hindlimb ischemia. *Diabetes* 2012; **61**: 1153-1159 [PMID: 22266669 DOI: 10.2337/db11-1271]
89. **Lee CY**, Kang JY, Lim S, Ham O, Chang W, Jang DH. Hypoxic conditioned medium from mesenchymal stem cells promotes lymphangiogenesis by regulation of mitochondrial-related proteins. *Stem Cell Res Ther* 2016; **7**: 38 [PMID: 26968383 DOI: 10.1186/s13287-016-0296-1]
90. **Orlacchio A**, Babalini C, Borreca A, Patrono C, Massa R, Basaran S, Munhoz RP, Rogaeva EA, St George-Hyslop PH, Bernardi G, Kawarai T. SPATACSIN mutations cause autosomal recessive juvenile amyotrophic lateral sclerosis. *Brain* 2010; **133**: 591-598 [PMID: 20110243 DOI: 10.1093/brain/awp325]
91. **Fang X**, Wang X, Yang S, Meng F, Wang X, Wei H, Chen T. Evaluation of the Microbial Diversity in Amyotrophic Lateral Sclerosis Using High-Throughput Sequencing. *Front Microbiol* 2016; **7**: 1479 [PMID: 27703453 DOI: 10.3389/fmicb.2016.01479]
92. **Chen SG**, Stribinskis V, Rane MJ, Demuth DR, Gozal E, Roberts AM, Jagadapillai R, Liu R, Choe K, Shivakumar B, Son F, Jin S, Kerber R, Adame A, Masliah E, Friedland RP. Exposure to the Functional Bacterial Amyloid Protein Curli Enhances Alpha-Synuclein Aggregation in Aged Fischer 344 Rats and Caenorhabditis elegans. *Sci Rep* 2016; **6**: 34477 [PMID: 27708338 DOI: 10.1038/srep34477]
93. **Distrutti E**, O'Reilly JA, McDonald C, Cipriani S, Renga B, Lynch MA, Fiorucci S. Modulation of intestinal microbiota by the probiotic VSL#3 resets brain gene expression and ameliorates the age-related deficit in LTP. *PLoS One* 2014; **9**: e106503 [PMID: 25202975 DOI: 10.1371/journal.pone.0106503]
94. **Stewart JA Jr**, Cashatt DO, Borck AC, Brown JE, Carver WE. 17beta-estradiol modulation of angiotensin II-stimulated response in cardiac fibroblasts. *J Mol Cell Cardiol* 2006; **41**: 97-107 [PMID: 16765374 DOI: 10.1016/j.yjmcc.2006.04.019]
95. **Serban AI**, Stanca L, Geicu OI, Munteanu MC, Dinischiotu A. RAGE and TGF-β1 Cross-Talk Regulate Extracellular Matrix Turnover and Cytokine Synthesis in AGEs Exposed Fibroblast Cells. *PLoS One* 2016; **11**: e0152376 [PMID: 27015414 DOI: 10.1371/journal.pone.0152376]
96. **Bianchi R**, Kastrisianaki E, Giambanco I, Donato R. S100B protein stimulates microglia migration via RAGE-dependent up-regulation of chemokine expression and release. *J Biol Chem* 2011; **286**: 7214-7226 [PMID: 21209080 DOI: 10.1074/jbc.M110.169342]
97. **Casula M**, Iyer AM, Spliet WG, Anink JJ, Steentjes K, Sta M, Troost D, Aronica E. Toll-like receptor signaling in amyotrophic lateral sclerosis spinal cord tissue. *Neuroscience* 2011; **179**: 233-243 [PMID: 21303685 DOI: 10.1016/j.neuroscience.2011.02.001]
98. **Juranek JK**, Daffu GK, Wojtkiewicz J, Lacomis D, Kofler J, Schmidt AM. Receptor for Advanced Glycation End Products and its Inflammatory Ligands are Upregulated in Amyotrophic Lateral Sclerosis. *Front Cell Neurosci* 2015; **9**: 485 [PMID: 26733811 DOI: 10.3389/fncel.2015.00485]
99. **Rappaport SM**, Li H, Grigoryan H, Funk WE, Williams ER. Adductomics: characterizing exposures to reactive electrophiles. *Toxicol Lett* 2012; **213**: 83-90 [PMID: 21501670 DOI: 10.1016/j.toxlet.2011.04.002]
100. **Gaudet AD**, Popovich PG. Extracellular matrix regulation of inflammation in the healthy and injured spinal cord. *Exp Neurol* 2014; **258**: 24-34 [PMID: 25017885 DOI: 10.1016/j.expneurol.2013.11.020]
101. **Katayama Y**, Battista M, Kao WM, Hidalgo A, Peired AJ, Thomas SA, Frenette PS. Signals from the sympathetic nervous system regulate hematopoietic stem cell egress from bone marrow. *Cell* 2006; **124**: 407-421 [PMID: 16439213 DOI: 10.1016/j.cell.2005.10.041]
102. **Olsson R**, Remberger M, Schaffer M, Berggren DM, Svahn BM, Mattsson J, Ringden O. Graft failure in the modern era of allogeneic hematopoietic SCT. *Bone Marrow Transplant* 2013; **48**: 537-543 [PMID: 23222384 DOI: 10.1038/bmt.2012.239]
103. **Estin C**, Vernadakis A. Primary glial cells and brain fibroblasts: interactions in culture. *Brain Res Bull* 1986; **16**: 723-731 [PMID: 2874876]
104. **Chen IH**, Xue L, Hsu CC, Paez JS, Pan L, Andaluz H, Wendt MK, Iliuk AB, Zhu JK, Tao WA. Phosphoproteins in extracellular vesicles as candidate markers for breast cancer. *Proc Natl Acad Sci U S A* 2017; **114**: 3175-3180 [PMID: 28270605 DOI: 10.1073/pnas.1618088114]
105. **Le Blanc K**, Rasmusson I, Sundberg B, Götherström C, Hassan M, Uzunel M, Ringdén O. Treatment of severe acute graft-versus-host disease with third party haploidentical mesenchymal stem cells. *Lancet* 2004; **363**: 1439-1441 [PMID: 15121408 DOI: 10.1016/S0140-6736(04)16104-7]
106. **Baron F**, Storb R. Mesenchymal stromal cells: a new tool against graft-versus-host disease? *Biol Blood Marrow Transplant* 2012; **18**: 822-840 [PMID: 21963621 DOI: 10.1016/j.bbmt.2011.09.003]
107. **Kordelas L**, Rebmann V, Ludwig AK, Radtke S, Ruesing J, Doeppner TR, Epple M, Horn PA, Beelen DW, Giebel B. MSC-derived exosomes: a novel tool to treat therapy-refractory graft-versus-host disease. *Leukemia* 2014; **28**: 970-973 [PMID: 24445866 DOI: 10.1038/leu.2014.41]
108. **Burzyn D**, Kuswanto W, Kolodin D, Shadrach JL, Cerletti M, Jang Y, Sefik E, Tan TG, Wagers AJ, Benoist C, Mathis D. A special population of regulatory T cells potentiates muscle repair. *Cell* 2013; **155**: 1282-1295 [PMID: 24315098 DOI: 10.1016/j.cell.2013.10.054]
109. **Medzhitov R**. Origin and physiological roles of inflammation. *Nature* 2008; **454**: 428-435 [PMID: 18650913 DOI: 10.1038/nature07201]
110. **Franceschi C**, Campisi J. Chronic inflammation (inflammaging) and its potential contribution to age-associated diseases. *J Gerontol A Biol Sci Med Sci* 2014; **69** Suppl 1: S4-S9 [PMID: 24833586 DOI: 10.1093/gerona/glu057]
111. **Rassoulzadegan M**, Grandjean V, Gounon P, Vincent S, Gillot I, Cuzin F. RNA-mediated non-mendelian inheritance of an epigenetic change in the mouse. *Nature* 2006; **441**: 469-474 [PMID: 16724059 DOI: 10.1038/nature04674]
112. **Carone BR**, Fauquier L, Habib N, Shea JM, Hart CE, Li R, Bock C, Li C, Gu H, Zamore PD, Meissner A, Weng Z, Hofmann HA, Friedman N, Rando OJ. Paternally induced transgenerational environmental reprogramming of metabolic gene expression in mammals. *Cell* 2010; **143**: 1084-1096 [PMID: 21183072 DOI: 10.1016/j.cell.2010.12.008]
113. **Rando OJ**. Intergenerational Transfer of Epigenetic Information in Sperm. *Cold Spring Harb Perspect Med* 2016; **6**: pii: a022988 [PMID: 26801897 DOI: 10.1101/cshperspect.a022988]
114. **Sharma U**, Conine CC, Shea JM, Boskovic A, Derr AG, Bing XY, Belleannee C, Kucukural A, Serra RW, Sun F, Song L, Carone BR, Ricci EP, Li XZ, Fauquier L, Moore MJ, Sullivan R, Mello CC, Garber M, Rando OJ. Biogenesis and function of tRNA fragments during sperm maturation and fertilization in mammals. *Science* 2016; **351**: 391-396 [PMID: 26721685 DOI: 10.1126/science.aad6780]
115. **Rechavi O**, Houri-Ze'evi L, Anava S, Goh WS, Kerk SY, Hannon GJ, Hobert O. Starvation-induced transgenerational inheritance of small RNAs in C. elegans. *Cell* 2014; **158**: 277-287 [PMID: 25018105 DOI: 10.1016/j.cell.2014.06.020]
116. **Houri-Ze'evi L**, Korem Y, Sheftel H, Faigenbloom L, Toker IA, Dagan Y, Awad L, Degani L, Alon U, Rechavi O. A Tunable Mechanism Determines the Duration of the Transgenerational Small RNA Inheritance in C. elegans. *Cell* 2016; **165**: 88-99 [PMID: 27015309 DOI: 10.1016/j.cell.2016.02.057]
117. **Jia H**, Morris CD, Williams RM, Loring JF, Thomas EA. HDAC inhibition imparts beneficial transgenerational effects in Huntington's disease mice via altered DNA and histone methylation. *Proc Natl Acad Sci U S A* 2015; **112**: E56-E64 [PMID: 25535382 DOI: 10.1073/pnas.1415195112]
118. **Taetzsch T**, Tenga MJ, Valdez G. Muscle Fibers Secrete FGFBP1 to Slow Degeneration of Neuromuscular Synapses during Aging and Progression of ALS. *J Neurosci* 2017; **37**: 70-82 [PMID: 28053031 DOI: 10.1523/JNEUROSCI.2992-16.2016]
119. **Cozzolino M**, Rossi S, Mirra A, Carrì MT. Mitochondrial dynamism and the pathogenesis of Amyotrophic Lateral Sclerosis. *Front Cell Neurosci* 2015; **9**: 31 [PMID: 25713513 DOI: 10.3389/fncel.2015.00031]
120. **Zhang Q**, Raoof M, Chen Y, Sumi Y, Sursal T, Junger W, Brohi K, Itagaki K, Hauser CJ. Circulating mitochondrial DAMPs cause inflammatory responses to injury. *Nature* 2010; **464**: 104-107 [PMID: 20203610 DOI: 10.1038/nature08780]
121. **Iyer SS**, He Q, Janczy JR, Elliott EI, Zhong Z, Olivier AK, Sadler JJ, Knepper-Adrian V, Han R, Qiao L, Eisenbarth SC, Nauseef WM, Cassel SL, Sutterwala FS. Mitochondrial cardiolipin is required for Nlrp3 inflammasome activation. *Immunity* 2013; **39**: 311-323 [PMID: 23954133 DOI: 10.1016/j.immuni.2013.08.001]
122. **Williams AH**, Valdez G, Moresi V, Qi X, McAnally J, Elliott JL, Bassel-Duby R, Sanes JR, Olson EN. MicroRNA-206 delays ALS progression and promotes regeneration of neuromuscular synapses in mice. *Science* 2009; **326**: 1549-1554 [PMID: 20007902 DOI: 10.1126/science.1181046]
123. **Kye MJ**, Gonçalves Ido C. The role of miRNA in motor neuron disease. *Front Cell Neurosci* 2014; **8**: 15 [PMID: 24523674 DOI: 10.3389/fncel.2014.00015]
124. **Wang X**, Gerdes HH. Transfer of mitochondria via tunneling nanotubes rescues apoptotic PC12 cells. *Cell Death Differ* 2015; **22**: 1181-1191 [PMID: 25571977 DOI: 10.1038/cdd.2014.211]

**P-Reviewer:** Orlacchio A, Yao Dengfu **S-Editor:** Kong JX **L-Editor: E-Editor:**

**Specialty type:** Cell and tissue engineering

**Country of origin:** United States

**Peer-review report classification**

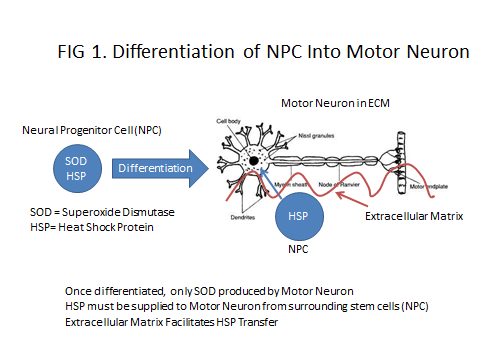
Grade A (Excellent): 0

Grade B (Very good): B

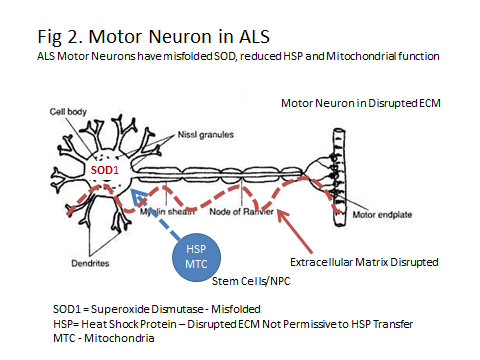
Grade C (Good): C

Grade D (Fair): 0

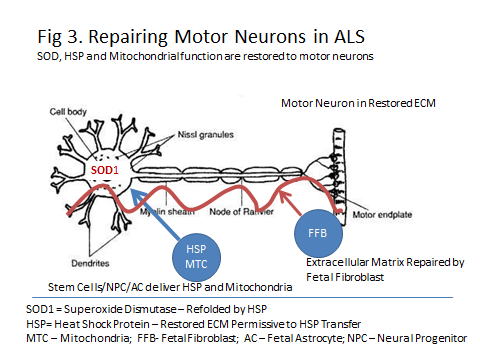
Grade E (Poor): 0



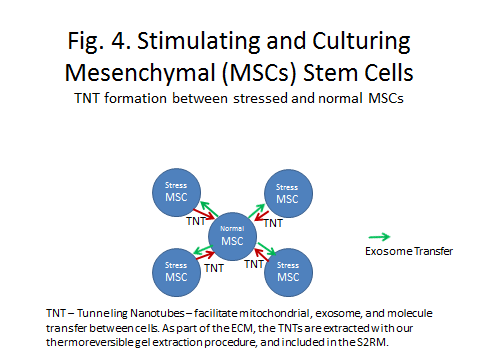
**Figure 1 Differentiation of neural progenitor cells into motor neuron.** Once differenciated, only SOD produced by motor neuron. HSP must be supplied to motor neuron from surrounding stem cells (NPC). ECM facilitates HSP transfer. SOD: Superoxide dismutase; NPC: Neural progenitor cells; ECM: Extracellular matrix; HSP: Heat shock proteins.

****

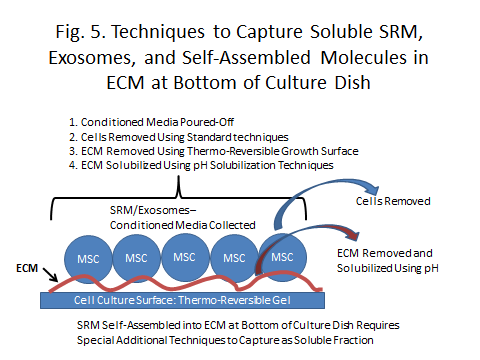
**Figure 2 Motor neuron in amyotrophic lateral sclerosis.** ALS Motor neurons have misfolded SOD, reduced HSP and mitochondrial function. ALS: Amyotrophic lateral sclerosis; SOD: Superoxide dismutase; SOD1: Superoxide dismutase – misfolded; HSP: Heat shock proteins – disrupted ECM not permissive to HSP transfer; MTC: Mitochondria; ECM: Extracellular matrix.

****

**Figure 3 Repairing motor neurons in amyotrophic lateral sclerosis.** SOD, HSP and mitochondrial function are restores to motor neurons. SOD: Superoxide dismutase; SOD1: Superoxide dismutase – misfolded; HSP: Heat shock proteins – disrupted ECM not permissive to HSP transfer; MTC: Mitochondria; NPC: Neural progenitor cells; ECM: Extracellular matrix; FFB: Fetal fibroblast; AC: Fetal astrocyte.

****

**Figure 4 Stimulating and culturing mesenchymal stem cells.** TNT formation between stressed and normal MSCs. TNTs facilitate mitochondrial, exosome and molecule transfer between cells. As part of the ECM, the TNTs are extracted with out thermoreversible gel sxtraction procedure, and included in the S2RM. TNT:Tunnelling nanotubes; ECM: Extracellular matrix.

****

**Figure 5 Techniques to capture soluble SRM, exosomes, and self-assembled molecules in extracellular matrix at bottom of culture dish.**