World Journal of Hepatology

World J Hepatol 2018 January 27; 10(1): 1-171





Contents

Monthly Volume 10 Number 1 January 27, 2018

MINIREVIEWS

1 Role of inflammatory response in liver diseases: Therapeutic strategies

Del Campo JA, Gallego P, Grande L

ORIGINAL ARTICLE

Basic Study

- Preserved liver regeneration capacity after partial hepatectomy in rats with non-alcoholic steatohepatitis Haldrup D, Heebøll S, Thomsen KL, Andersen KJ, Meier M, Mortensen FV, Nyengaard JR, Hamilton-Dutoit S, Grønbæk H
- Bioengineered humanized livers as better three-dimensional drug testing model system Vishwakarma SK, Bardia A, Lakkireddy C, Nagarapu R, Habeeb MA, Khan AA

Retrospective Cohort Study

- Risk factors for hepatic steatosis in adults with cystic fibrosis: Similarities to non-alcoholic fatty liver disease Ayoub F, Trillo-Alvarez C, Morelli G, Lascano J
- Fatty liver disease, an emerging etiology of hepatocellular carcinoma in Argentina

 Piñero F, Pages J, Marciano S, Fernández N, Silva J, Anders M, Zerega A, Ridruejo E, Ameigeiras B, D'Amico C, Gaite L,

 Bermúdez C, Cobos M, Rosales C, Romero G, McCormack L, Reggiardo V, Colombato L, Gadano A, Silva M
- Current state and clinical outcome in Turkish patients with hepatocellular carcinoma

 Ekinci O, Baran B, Ormeci AC, Soyer OM, Gokturk S, Evirgen S, Poyanli A, Gulluoglu M, Akyuz F, Karaca C, Demir K,

 Besisik F, Kaymakoglu S

Retrospective Study

- Predicting early outcomes of liver transplantation in young children: The EARLY study Alobaidi R, Anton N, Cave D, Moez EK, Joffe AR
- Collagen proportionate area correlates to hepatic venous pressure gradient in non-abstinent cirrhotic patients with alcoholic liver disease
 - Restellini S, Goossens N, Clément S, Lanthier N, Negro F, Rubbia-Brandt L, Spahr L
- Ratio of mean platelet volume to platelet count is a potential surrogate marker predicting liver cirrhosis *Iida H, Kaibori M, Matsui K, Ishizaki M, Kon M*
- 88 Efficacy of direct-acting antiviral treatment for chronic hepatitis C: A single hospital experience Kaneko R, Nakazaki N, Omori R, Yano Y, Ogawa M, Sato Y



Contents

95 Efficacy of intra-arterial contrast-enhanced ultrasonography during transarterial chemoembolization with drug-eluting beads for hepatocellular carcinoma

Shiozawa K, Watanabe M, Ikehara T, Yamamoto S, Matsui T, Saigusa Y, Igarashi Y, Maetani I

Clinical Practice Study

105 Proton nuclear magnetic resonance-based metabonomic models for non-invasive diagnosis of liver fibrosis in chronic hepatitis C: Optimizing the classification of intermediate fibrosis

Batista AD, Barros CJP, Costa TBBC, Godoy MMG, Silva RD, Santos JC, de Melo Lira MM, Jucá NT, Lopes EPA, Silva RO

Observational Study

High burden of hepatocellular carcinoma and viral hepatitis in Southern and Central Vietnam: Experience of a large tertiary referral center, 2010 to 2016

Nguyen-Dinh SH, Do A, Pham TND, Dao DY, Nguy TN, Chen Jr MS

Toll-like receptor 4 polymorphisms and bacterial infections in patients with cirrhosis and ascites

Alvarado-Tapias E, Guarner-Argente C, Oblitas E, Sánchez E, Vidal S, Román E, Concepción M, Poca M, Gely C, Pavel O,

Nieto JC, Juárez C, Guarner C, Soriano G

Prospective Study

134 Effect of transplant center volume on post-transplant survival in patients listed for simultaneous liver and kidney transplantation

Modi RM, Tumin D, Kruger AJ, Beal EW, Hayes Jr D, Hanje J, Michaels AJ, Washburn K, Conteh LF, Black SM, Mumtaz K

META-ANALYSIS

- 142 Vitamin D levels do not predict the stage of hepatic fibrosis in patients with non-alcoholic fatty liver disease: A PRISMA compliant systematic review and meta-analysis of pooled data Saberi B, Dadabhai AS, Nanavati J, Wang L, Shinohara RT, Mullin GE
- Epigenetic basis of hepatocellular carcinoma: A network-based integrative meta-analysis Bhat V, Srinathan S, Pasini E, Angeli M, Chen E, Baciu C, Bhat M

CASE REPORT

Contrast uptake in primary hepatic angiosarcoma on gadolinium-ethoxybenzyl-diethylenetriamine pentaacetic acid-enhanced magnetic resonance imaging in the hepatobiliary phase

Hayashi M, Kawana S, Sekino H, Abe K, Matsuoka N, Kashiwagi M, Okai K, Kanno Y, Takahashi A, Ito H, Hashimoto Y,

Ohira H



Contents

World Journal of Hepatology Volume 10 Number 1 January 27, 2018

ABOUT COVER

Editorial Board Member of *World Journal of Hepatology*, Konstantinos Tziomalos, MD, MSc, PhD, Assistant Professor, First Propedeutic Department of Internal Medicine, Medical School, Aristotle University of Thessaloniki, Thessaloniki 54636, Greece

AIM AND SCOPE

World Journal of Hepatology (World J Hepatol, WJH, online ISSN 1948-5182, DOI: 10.4254), is a peer-reviewed open access academic journal that aims to guide clinical practice and improve diagnostic and therapeutic skills of clinicians.

WJH covers topics concerning liver biology/pathology, cirrhosis and its complications, liver fibrosis, liver failure, portal hypertension, hepatitis B and C and inflammatory disorders, steatohepatitis and metabolic liver disease, hepatocellular carcinoma, biliary tract disease, autoimmune disease, cholestatic and biliary disease, transplantation, genetics, epidemiology, microbiology, molecular and cell biology, nutrition, geriatric and pediatric hepatology, diagnosis and screening, endoscopy, imaging, and advanced technology. Priority publication will be given to articles concerning diagnosis and treatment of hepatology diseases. The following aspects are covered: Clinical diagnosis, laboratory diagnosis, differential diagnosis, imaging tests, pathological diagnosis, molecular biological diagnosis, immunological diagnosis, genetic diagnosis, functional diagnostics, and physical diagnosis; and comprehensive therapy, drug therapy, surgical therapy, interventional treatment, minimally invasive therapy, and robot-assisted therapy.

We encourage authors to submit their manuscripts to WJH. We will give priority to manuscripts that are supported by major national and international foundations and those that are of great basic and clinical significance.

INDEXING/ABSTRACTING

World Journal of Hepatology is now indexed in Emerging Sources Citation Index (Web of Science), PubMed, PubMed Central, and Scopus.

EDITORS FOR THIS ISSUE

Responsible Assistant Editor: Xiang Li Responsible Electronic Editor: Rui-Fang Li Proofing Editor-in-Chief: Lian-Sheng Ma Responsible Science Editor: Li-Jun Cui
Proofing Editorial Office Director: Xiu-Xia Song

NAME OF TOURNAL

World Journal of Hepatology

ISSN

ISSN 1948-5182 (online)

LAUNCH DATE October 31, 2009

FREQUENCY

Monthly

EDITOR-IN-CHIEF

Wan-Long Chuang, MD, PhD, Doctor, Professor, Hepatobiliary Division, Department of Internal Medicine, Kaohsiung Medical University Hospital, Kaohsiung Medical University, Kaohsiung 807, Taiwan

EDITORIAL BOARD MEMBERS

All editorial board members resources online at http://www.wjgnet.com/1948-5182/editorialboard.htm

EDITORIAL OFFICE

Xiu-Xia Song, Director

World Journal of Hepatology
Baishideng Publishing Group Inc
7901 Stoneridge Drive, Suite 501,
Pleasanton, CA 94588, USA
Telephone: +1-925-2238242
Fax: +1-925-2238243
E-mail: editorialoffice@wjgnet.com
Help Desk: http://www.fopublishing.com/helpdesk
http://www.wjgnet.com

PUBLISHER

Baishideng Publishing Group Inc
7901 Stoneridge Drive, Suite 501,
Pleasanton, CA 94588, USA
Telephone: +1-925-2238242
Fax: +1-925-2238243
E-mail: bpgoffice@wjgnet.com
Help Desk: http://www.fpublishing.com/helpdesk
http://www.wignet.com

PUBLICATION DATE

January 27, 2018

COPYRIGHT

© 2018 Baishideng Publishing Group Inc. Articles published by this Open Access journal are distributed under the terms of the Creative Commons Attribution Noncommercial License, which permits use, distribution, and reproduction in any medium, provided the original work is properly cited, the use is non commercial and is otherwise in compliance with the license.

SPECIAL STATEMENT

All articles published in journals owned by the Baishideng Publishing Group (BPG) represent the views and opinions of their authors, and not the views, opinions or policies of the BPG, except where otherwise explicitly indicated.

INSTRUCTIONS TO AUTHORS

http://www.wjgnet.com/bpg/gerinfo/204

ONLINE SUBMISSION

http://www.f6publishing.com



Submit a Manuscript: http://www.f6publishing.com

World J Hepatol 2018 January 27; 10(1): 1-7

DOI: 10.4254/wjh.v10.i1.1 ISSN 1948-5182 (online)

MINIREVIEWS

Role of inflammatory response in liver diseases: Therapeutic strategies

1

José A Del Campo, Paloma Gallego, Lourdes Grande

José A Del Campo, Paloma Gallego, Lourdes Grande, Department of Digestive Diseases, Valme University Hospital and CIBERehd, Sevilla 41014, Spain

ORCID number: José A Del Campo (0000-0002-6037-1028); Paloma Gallego (0000-0002-7233-0127); Lourdes Grande (0000-0001-7685-0464).

Author contributions: Del Campo JA designed the study and drafted the manuscript; Gallego P and Grande L drafted the manuscript and designed the figure.

Supported by Andalusian Government, No. PI0892-2012; Instituto de Salud Carlos III, PI14/01349 co-financed by the European Regional Development Fund (ERDF); JA Del Campo supported by Nicolás Monardes Program from Servicio Andaluz de Salud (SAS).

Conflict-of-interest statement: Authors declare no conflicts of interest.

Open-Access: This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: http://creativecommons.org/licenses/by-nc/4.0/

Manuscript source: Invited manuscript

Correspondence to: José A Del Campo, PhD, Department of Digestive Diseases, Valme University Hospital and CIBERehd,

Avda. Bellavista s/n, Sevilla 41014, Spain. jantonio.delcampo@ciberehd.org

Telephone: +34-955-015485 Fax: +34-955-015485

Received: November 8, 2017

Peer-review started: November 9, 2017 First decision: December 4, 2017 Revised: December 21, 2017 Accepted: January 15, 2018 Article in press: January 15, 2018 Published online: January 27, 2018

Abstract

Inflammation and tumorigenesis are tightly linked pathways impacting cancer development. Inflammasomes are key signalling platforms that detect pathogenic microorganisms, including hepatitis C virus (HCV) infection, and sterile stressors (oxidative stress, insulin resistance, lipotoxicity) able to activate pro-inflammatory cytokines interleukin-1ß and IL-18. Most of the inflammasome complexes that have been described to date contain a NOD-like receptor sensor molecule. Redox state and autophagy can regulate inflammasome complex and, depending on the conditions, can be either pro- or antiapoptotic. Acute and chronic liver diseases are cytokinedriven diseases as several proinflammatory cytokines (IL- 1α , IL- 1β , tumor necrosis factor-alpha, and IL-6) are critically involved in inflammation, steatosis, fibrosis, and cancer development. NLRP3 inflammasome gain of function aggravates liver disease, resulting in severe liver fibrosis and highlighting this pathway in the pathogenesis of non-alcoholic fatty liver disease. On the other hand, HCV infection is the primary catalyst for progressive liver disease and development of liver cancer. It is well established that HCV-induced IL-1B production by hepatic macrophages plays a critical and central process that promotes liver inflammation and disease. In this review, we aim to clarify the role of the inflammasome in the aggravation of liver disease, and how selective blockade of this main pathway may be a useful strategy to delay fibrosis progression in liver diseases.

Key words: Caspase-1; Fibrosis; Hepatitis C virus; Inflammasome; Interleukin- 1α ; Interleukin- 1β ; Liver disease; Non-alcoholic fatty liver disease; NLRP3; Tumor necrosis factor-alpha

© The Author(s) 2018. Published by Baishideng Publishing



Group Inc. All rights reserved.

Core tip: Inflammasomes are newly recognized vital players in innate immunity. Several factors have been identified able to activate the NLRP3 inflammasome. Inappropriate activation of NLRP3 can contribute to the onset and progression of various diseases, particularly age-related diseases. It is well established that hepatitis C virus infection plays a critical role in the promotion of liver inflammation and disease, inducing the production of IL-1 β and the activation of NLRP3. NLRP3 inflammasome gain of function aggravates liver disease, resulting in severe liver fibrosis and lately, hepatocellular carcinoma. In non-alcoholic fatty liver disease, the regulation of inflammation processes may prevent the progression of non-alcoholic steatohepatitis to fibrosis.

Del Campo JA, Gallego P, Grande L. Role of inflammatory response in liver diseases: Therapeutic strategies. *World J Hepatol* 2018; 10(1): 1-7 Available from: URL: http://www.wjgnet.com/1948-5182/full/v10/i1/1.htm DOI: http://dx.doi.org/10.4254/wjh.v10.i1.1

INTRODUCTION

Hepatic inflammation is a common trigger of liver disease, and is considered the main driver of hepatic tissue damage, triggering the progression from non-alcoholic fatty liver disease (NAFLD) to severe fibrogenesis and, finally, hepatocellular carcinoma (HCC).

Liver diseases, whose etiology can be diverse, are becoming one of the most serious public health problems. The diseases usually occur in response to chronic hepatocellular injury caused mainly by the abuse of alcoholic intake, chronic infections such as those caused by the hepatitis C virus (HCV), bile duct damage, NAFLD or non-alcoholic steatohepatitis (NASH)[1]. NAFLD was defined for the first time in 1980^[2] as an accumulation of fat (> 5%) in liver cells in the absence of excessive alcohol intake^[3]. The disease affects more than 30% of the population of the western world, especially patients suffering from metabolic syndrome, obesity (76%), and type II diabetes (50%)^[4]. The histological spectrum of NAFLD begins in a simple benign steatosis, evolving to a NASH. From this point, the consequent scarring and tissue replacement begins with type I collagen, developing fibrosis, cirrhosis and finally, in many cases complicating a HCC[3]. This pathogenesis, which is complex, is explained by two main impacts or damages to the liver tissue: on the one hand, a lipid accumulation in the hepatocytes, to which a second oxidative stress damage is added^[5]. It is this second damage, which produces a lipotoxicit that triggers the inflammatory response by the release of danger-activated molecular patterns (DAMPS) and pathogenic-activated molecular patterns (PAMPs), such

as lipopolysaccharide (LPS). Finally, they activate the innate immunity that causes the hepatic inflammation, being able to aggravate the process of fibrosis, cirrhosis, causing HCC.

That is why the knowledge of the relationship between liver disease and uncontrolled activation of the immune response, resulting in aggravating liver inflammation of disease progression, can be crucial for the design of therapeutic strategies blocking the immune response uncontrolled.

PATHOGENESIS OF NAFLD

The pathogenesis of this disease is complex, and is explained by two major impacts or damage to the liver tissue: on the one hand, a lipid accumulation in the hepatocytes, to which a second oxidative stress damage is added^[5].

Lipid accumulation in hepatocytes

The first of these is the abnormal accumulation of triglycerides in the hepatocyte, either due to a high intake of saturated fats and obesity, genetic deficiencies or insulin resistance (IR) due to hyperglycemia and hyperinsulinemia^[6], what is known as metabolic syndrome. Hyperinsulinemia and increased hepatic glucose production induce the expression of the sterol regulatory element binding protein (SREBP-1c) and the carbohydrate response element binding protein (ChREBP), respectively, which activate in turn the transcription of most of the genes involved in the enzymatic machinery necessary for free fatty acids (FFA) synthesis, decreasing the beta oxidation thereof^[7].

In addition, there are many mediating molecules such as peroxisome proliferator activating ligand receptors (PPAR), among which PPAR-7, whose expression in conditions of liver damage is usually high, contribute to the accumulation of FFA^[7]. The liver X receptor (LXR), another important mediator, activates genes involved in the synthesis of FFA, such as SREBP-1c and ChREBP, contributing to steatosis^[8]. Finally, the AMP-activated protein kinase (AMPK) functions as a sensor of the energy levels of the cell, stimulating catabolic pathways such as mitochondrial beta oxidation, and inhibiting ATP-consuming processes, such as lipogenesis. All this is done by phosphorylating different proteins involved in these pathways, such as acetyl-CoA carboxylase (ACC), ChREBP and SREBP-1c, which end up being inhibited.

In the process of lipid accumulation, several molecules act as DAMPs and pathogen-activated molecular patterns (PAMPs), that is, molecules that trigger inflammation. These may include the fatty acids, adenosine triphosphate (ATP), uric acid or proteins derived from the extracellular matrix, among others. In addition, recent research suggests that in the liver tissue of humans and mice with steatohepatitis, cholesterol crystals are present within hepatocytes, and they can act as DAMPs.



Oxidative stress and liver inflammation

A second impact that explains the characteristic histological lesions of NAFLD is oxidative stress and lipid peroxidation [5,9]. As a result of liver damage and liver inflammation, the inflammatory cells and the hepatocyte itself release cytokines, such as TNF- α and reactive oxygen species (ROS). These mediators can cause peroxidation of plasma and mitochondrial membranes, which leads to cell death due to necrosis or apoptosis [5,9]. All this activates endothelial cells of the liver, which increase the expression of cytokines, and finally activates the hepatic stellate cells (HSCs), producing a phenotypic change, associated with the acquisition of pro-fibrogenic and pro-inflammatory functions [10].

Significant accumulation of triglycerides and cholesterol in VLDL and LDL particles, added to the oxidative stress developed in the hepatocytes, produces the oxidation of the cholesterol linked to LDL, generating particles of LDL oxidase (oxLDL) that constitute an important risk factor of the disease^[11]. Recent studies show that oxLDL also contribute to the process of cellular inflammation and apoptosis.

ACTIVATION OF THE INNATE IMMUNE RESPONSE DURING NAFLD

Excessive lipids accumulation leads to hepatocyte damage, activating an inflammatory response that aggravates the progress of the liver disease, and which, in turn, feeds back the activation of the inflammation^[12].

The immune response of the liver is produced by the action of immune cells such as Kupffer cells, monocytes, neutrophils, dendritic cells (DCs), natural killer cells (NK), and NK T cells (NKT), which initiate and maintain the hepatic inflammation through the production of cytokines and chemokines, especially TNF and interleukin (IL) -1 β , as well as reactive oxygen species^[13].

The triggering of hepatic inflammation, as noted above, is caused by the accumulation of infectious and non-infectious material, which is released during cell damage and is recognized by pattern recognition receptors (PRRs). These PRRs include Toll-like receptors (TLRs), NOD-like receptors (NLRs), C-type lectin receptors (CLRs), and several other receptors [14].

The endogenous molecules produced by cellular damage or stress result in an inflammatory response (DAMPs). These molecules are very diverse and do not have a common structure between them. In the pathology of the liver disease and liver inflammation, induced-inflammation particles are included, such as cholesterol crystals. These cholesterol crystals produce the development, not only of the atheroma plaques, but of the inflammation, being present in humans and mice with NASH^[15] candidates are free fatty acids (FFA) that are released during liver damage. A specific mechanism by which they act is to activate TLR4, while fetuin-A, a 64-kDa protein specific for hepatocytes,

is required. It has been proven that the reduction of fetuin-A in mice with high fat diet results in a decrease of inflammatory signalling mediated by TLR4 in adipose tissue. Normally, in patients with NAFLD, the expression of fetuin-A is high^[16]. Palmitic acid (PA) is another molecule that causes liver inflammation, and it has recently been linked to the TLR2 receptor. The palmitic acid ligand of TLR2 induces the activation of caspase- Other 1 and the release of IL-1 α and IL-1 β , having an evident role in the liver inflammatory process^[17].

Many of these DAMPs activate PRR present in immune cells, of which the TLRs are the best characterized. In liver disease, saturation of fatty acids produces inflammation in the hepatocytes, which increases the induction of caspase-1 activation and the release of IL- 1β . This results in a release of more DAMPs from the hepatocytes, generating a feedback that amplifies the inflammatory response.

Among the various NLR receptors, the NLRP3 inflammasome is the best characterized and associated with a wider range of diseases including infections, autoinflammatory diseases and other autoimmune diseases. The NLRP3 inflammasome has the characteristic of forming a complex with apoptosis-associated specklike protein to CARD (ASC), to activate caspase-1 and induce the maturation and secretion of important proinflammatory cytokines such as IL-1 β and IL-18. Cytokines act directly against the damage and infection of the liver tissue $^{[18]}$.

Role of NLRP3 inflammasome in liver inflammation

Activation of the NLRP3 inflammasome is important in the inflammatory process, and occurs in two steps. The first includes the activation of TLRs by different molecules DAMPs and PAMPs (Figure 1). TLRs belong to the family of PRRs and their function is to maintain tissue homeostasis through the regulation of inflammatory responses. In the liver, TLRs are expressed in Kupffer cells, endothelial cells, DCs, epithelial biliary cells, HSCs and hepatocytes. Activated TLRs activate the cells and contribute to the release of cytokines that facilitate the progression of liver disease. There are at least 13 known TLRs, whose structure is characterized by having a leucine-rich repeat structure (LRR) in the extracellular domain and a Toll/IL-1 receptor (TIR) in its intracellular domain^[19].

TLR4 has an interesting role in liver inflammation and fibrogenesis [20]. The Kupffer cells of the liver are the first to be attacked by the damage produced, and express TLR4 to which lipopolysaccharides (LPS) bind. This produces the activation of NF-kB, mitogenactivated protein kinase (MAPK)[21] extracellular signal-regulated kinase-1 (ERK1), p38, c-jun N-terminal kinase (JNK) and interferon regulatory factor 3(IRF3)[22], which finally triggers the production of proinflammatory cytokines and enhances IFN- β and STAT1 expression[23]. The proinflammatory stimulus facilitates hepatocyte damage, contributing to the secretion of profibrogenic

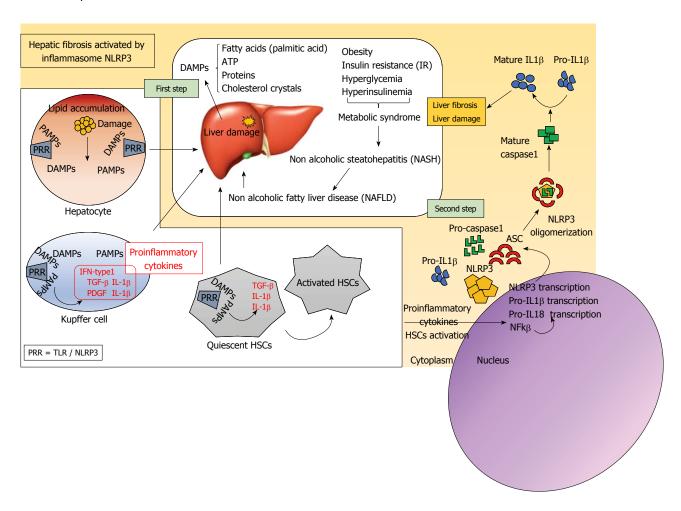


Figure 1 Hepatic fibrosis caused by the activation of the inflammasome-NLRP3 complex. The accumulation of lipids derived from the metabolic syndrome produces a non-alcoholic steatohepatitis (NASH) that progresses to a non-alcoholic fatty liver disease (NAFLD). The accumulated lipids, fatty acids, cholesterol crystals, among others, produce an alteration in the homeostasis of the liver cells, producing liver damage. Danger-activated molecular patterns (DAMPs) and pathogenic-activated molecular patterns (PAMPs) bind to and activate PRR receptors (TLRs, NLRPs, etc.) in hepatocytes, immune cells, hepatic stellate cells (HSCs), endothelial cells and DCs. Activated TLRs activate the cells and contribute to the release of cytokines that facilitate progression of liver disease. The proinflammatory stimulus facilitates hepatocyte damage, contributing to the secretion of profibrogenic cytokines such as transforming growth factor beta (TGF-β), promoting the activation of HSCs. This activation, in turn, up-regulates transcription of inflammasome-related components, including inactive NLRP3, pro-IL-1β and proIL-18. The second step of inflammation activation is the oligomerization of NLRP3 and subsequent assembly of NLRP3, ASC, and procaspase-1 into a complex. This triggers the transformation of procaspase-1 to caspase-1, as well as the production and secretion of mature IL-1β and IL-18, which produces liver damage and facilitates the progression of hepatic fibrosis.

cytokines such as transforming growth factor beta (TGF- β) and the platelet-derived growth factor (PDGF), promoting the activation of HSCs. This activation, in turn, up-regulates transcription of inflammasome-related components, including inactive NLRP3, pro-IL-1 β and proIL-18^[24,25].

The second step of inflammasome activation is the oligomerization of NLRP3 and subsequent assembly of NLRP3, ASC, and procaspase-1 into a complex (Figure 1). This triggers the transformation of procaspase-1 to caspase-1, as well as the production and secretion of mature IL-1 β and IL-18^[18].

NLRP3 signaling during liver inflammation

After NLRs overexpression, members of this family play an important role in the formation of intracellular multiprotein complexes called inflammasomes. The union of microbial components or activators of the infla-

mmasome (DAMPs and PAMPs) that enter the cytoplasm are detected by these cytosolic NLRs, activating the inflammasome formation. The inflammasome consists of an NLR protein, the adapter molecule ASC and procaspase-1, which is an effector molecule^[26]. The formation of this complex serves to activate cysteine protease caspase-1, which, in turn, produces the maturation of proinflammatory cytokines, including IL-1b and IL-18, and the proteolytic inactivation of IL-33^[27].

In short, and specifically in NAFLD, the saturated fatty acids from the excess lipid in the liver represent harmful endogenous molecules that finally induce the activation of the inflammasome. In addition, hepatocytes exposed to these saturated fatty acids release signals that trigger the activation of immune cells. Activation of the inflammasome activates caspase-1 in Kupffer cells, inducing proinflammatory signaling and activation of HSCs. In this way, collagen

deposition occurs that triggers liver fibrosis^[28] have shown that cholesterol crystals could be one pathway to activate the inflammasome in NASH. To test whether inflammasome blockade alters inflammatory recruitment, they used a drug called MCC950, which has already been shown to block NLRP3 activation, in an attempt to reduce liver injury in NASH. This drug partly reversed liver inflammation, particularly in obese diabetic mice.

Release of cytokines by immune cells

Inflammatory signaling of the liver is regulated by cytokines capable of activating effector functions in immune cells. Kupffer cells are the first to detect the presence of PAMPs and DAMPs through TLRs, activating the release of cytokines such as TNF- α , IL-1 and IL-6, as well as chemokines chemokine (C-X-C motif) ligand 13 (CXCL13), chemokine (C-X-C motif) ligand 8 CXCL-8 and Chemokine (C-C motif) ligand 24 (CCL24), which initiate an acute phase inflammatory response. These cytokines can produce apoptosis of hepatocytes, steatosis and inflammation, including the onset of a fibrosis process after interaction with HSCs via TGF-β. Cytokines can also activate hepatic sinusoidal endothelial cells that are ultimately involved in the recruitment of neutrophils, monocytes and NKT cells, being a feature of acute liver damage. Neutrophils can be activated, and their phenotype changed, releasing ROS, defensins and other chemokines that attract more neutrophils and monocytes. Monocytes can be differentiated into TNF- α , IL-1 β , granulocyte colony stimulating factor (G-CSF) and granulocytemacrophage colony stimulating factor (GM-CSF) macrophages, increasing the life expectancy of these neutrophils^[29,30]. Finally, the inflammation resolves with the apoptosis of the neutrophils, being these apoptotic neutrophils signals involved in the onset of phagocytosis and in the increase of IL-10 and TGF-β, cytokines related to the end of the inflammatory response and the repair of the tissue^[31].

Mesenchymal stem cells (MSCs) are a heterogeneous subset of stromal stem cells with immunonodulatory characteristics. MSCs are considered to act through multiple mechanisms to coordinate a dynamic, integrated response to liver inflammation and fibrosis, which prevents the progressive distortion of hepatic architecture. Management of inflammatory patterns is crucial also in case of potential treatment of liver diseases with stem cells^[32]. Adult stem cells have gained in attractiveness over embryonic stem cells for liver cell therapy due to their origin, multipotentiality, and the possibility of autologous transplantation.

ACTIVATION OF HSCS AND HEPATIC FIBROSIS

Any of these necroinflammatory mechanisms described above can activate HSCs, the main ones involved in

the process of hepatic fibrogenesis^[33]. In normal liver, stellate cells are described as being in a quiescent state. Quiescent stellate cells represent 5%-8% of the total number of liver cells^[34]. When the liver is damaged, stellate cells can change into an activated state. The activated stellate cell is characterized by proliferation, contractility, and chemotaxis. This state of the stellate cell is the main source of extracellular matrix production in liver injury^[35].

The hepatic stellate cells are characterized by having in their cytoplasm small droplets of fat 1-2 microns in diameter that allow the storage of vitamin A, one of its main functions. Besides, they control intercellular communication through the release of mediators, and participate in the homeostasis of the MEC of the liver by the production of collagens and noncollagens, synthesis of metalloproteinases (MMP) that catabolize the components of the MEC, and synthesis of inhibitors of MPPs, also called tissue inhibitors of metalloproteinases (TIMP) that control the catalytic activity of MMPs to maintain a homeostasis of the MEC^[36].

Therefore, after the chronic injury of the liver tissue, both the HSC and other cells producing the ECM undergo activation, a pathological process characterized by the loss of fat droplets, an increase in the number and size of the cells, and phenotypic transdifferentiation to proliferating, fibrogenic and contractile cells, which will be very similar to myofibroblasts. All this is mediated by different factors that induce cell proliferation, fibrogenic mediators such as TGF- $\beta1$ and IL-6, inducers of HSC contraction, such as endothelin-1, thrombin or angiotensin II and finally mediators of anti-inflammatory and anti-fibrogenic activity, such as IL-10 and interferon- γ (IFN- γ)[36].

Because of the activation of HSCs, phenotypic changes occur that affect the development of hepatic fibrosis, such as the production of collagen and noncollagen proteins of the MEC (collagen type I, type III, type IV, laminin, elastin, fibronectin and various proteoglycans) by the CEH^[37].

CONCLUSION

Many evidences reported in the literature suggest that the activation of the NLRP3 inflammasome complex and the consequent generation of the acute inflammatory response in the liver, facilitates the progress of steatohepatitis to liver fibrosis, cirrhosis and finally, HCC development. However, recent studies show that the KO mice of NRLP6 and NLRP3 inflammasomes have a worse progression in the NAFLD/NASH disease. The absence of the inflammasome is associated with changes in the homeostasis of the intestinal microbiota, resulting in a strong hepatic steatosis and inflammation through TLR receptor agonists, such as TLR4, which allows the release of TNF- α which leads to the progression of NASH^[38]. This demonstrates the complexity of the effects of the activation



of the inflammasome, which can generate an acute inflammation or, conversely, be protective. However and in general terms, this activation is usually proinflammatory in the liver, and its inactivation and absence in relation to the microbiota should be carefully studied. Recently, Pierantonelli *et al*^[39] have shown that the progression of liver fibrosis is associated with the downregulation of NLRP3 in the gut which, together with the current evidence of a strong correlation between intestinal changes (including modification of microbiota composition) and liver disease, makes the role of NLRP3 in the intestine extremely attractive as a protective factor. Finally, MCC950 has been proven as an effective NLRP3 inhibitor, being able to reduce liver injury and inflammation.

REFERENCES

- Younossi ZM, Loomba R, Anstee QM, Rinella ME, Bugianesi E, Marchesini G, Neuschwander-Tetri BA, Serfaty L, Negro F, Caldwell SH, Ratziu V, Corey KE, Friedman SL, Abdelmalek MF, Harrison SA, Sanyal AJ, Lavine JE, Mathurin P, Charlton MR, Goodman ZD, Chalasani NP, Kowdley KV, George J, Lindor K. Diagnostic Modalities for Non-alcoholic Fatty Liver Disease (NAFLD), Non-alcoholic Steatohepatitis (NASH) and Associated Fibrosis. *Hepatology* 2017; Epub ahead of print [PMID: 29222917 DOI: 10.1002/hep.29721]
- 2 Ludwig J, Viggiano TR, McGill DB, Oh BJ. Nonalcoholic steatohepatitis: Mayo Clinic experiences with a hitherto unnamed disease. Mayo Clin Proc 1980; 55: 434-438 [PMID: 7382552]
- 3 Chalasani N, Younossi Z, Lavine JE, Diehl AM, Brunt EM, Cusi K, Charlton M, Sanyal AJ. The diagnosis and management of non-alcoholic fatty liver disease: practice Guideline by the American Association for the Study of Liver Diseases, American College of Gastroenterology, and the American Gastroenterological Association. *Hepatology* 2012; 55: 2005-2023 [PMID: 22488764 DOI: 10.1002/hep.25762]
- 4 Loomba R, Sanyal AJ. The global NAFLD epidemic. Nat Rev Gastroenterol Hepatol 2013; 10: 686-690 [PMID: 24042449 DOI: 10.1038/nrgastro.2013.171]
- 5 Buzzetti E, Pinzani M, Tsochatzis EA. The multiple-hit pathogenesis of non-alcoholic fatty liver disease (NAFLD). Metabolism 2016; 65: 1038-1048 [PMID: 26823198 DOI: 10.1016/ j.metabol.2015.12.012]
- 6 Brunt EM, Wong VW, Nobili V, Day CP, Sookoian S, Maher JJ, Bugianesi E, Sirlin CB, Neuschwander-Tetri BA, Rinella ME. Nonalcoholic fatty liver disease. *Nat Rev Dis Primers* 2015; 1: 15080 [PMID: 27188459 DOI: 10.1038/nrdp.2015.80]
- 7 Hassan K, Bhalla V, El Regal ME, A-Kader HH. Nonalcoholic fatty liver disease: a comprehensive review of a growing epidemic. World J Gastroenterol 2014; 20: 12082-12101 [PMID: 25232245 DOI: 10.3748/wjg.v20.i34.12082]
- 8 **Chisholm JW**, Hong J, Mills SA, Lawn RM. The LXR ligand T0901317 induces severe lipogenesis in the db/db diabetic mouse. *J Lipid Res* 2003; **44**: 2039-2048 [PMID: 12923232]
- 9 Begriche K, Massart J, Robin MA, Bonnet F, Fromenty B. Mitochondrial adaptations and dysfunctions in nonalcoholic fatty liver disease. *Hepatology* 2013; 58: 1497-1507 [PMID: 23299992 DOI: 10.1002/hep.26226]
- Diehl AM, Li ZP, Lin HZ, Yang SQ. Cytokines and the pathogenesis of non-alcoholic steatohepatitis. *Gut* 2005; 54: 303-306 [PMID: 15647199 DOI: 10.1136/gut.2003.024935]
- Walenbergh SM, Koek GH, Bieghs V, Shiri-Sverdlov R. Nonalcoholic steatohepatitis: the role of oxidized low-density lipoproteins. *J Hepatol* 2013; 58: 801-810 [PMID: 23183522 DOI: 10.1016/j.jhep.2012.11.014]
- 12 Wree A, McGeough MD, Peña CA, Schlattjan M, Li H, Inzaugarat

- ME, Messer K, Canbay A, Hoffman HM, Feldstein AE. NLRP3 inflammasome activation is required for fibrosis development in NAFLD. *J Mol Med* (Berl) 2014; **92**: 1069-1082 [PMID: 24861026 DOI: 10.1007/s00109-014-1170-1]
- Tilg H, Diehl AM. Cytokines in alcoholic and nonalcoholic steatohepatitis. N Engl J Med 2000; 343: 1467-1476 [PMID: 11078773 DOI: 10.1056/NEJM200011163432007]
- Martinon F, Burns K, Tschopp J. The inflammasome: a molecular platform triggering activation of inflammatory caspases and processing of proIL-beta. *Mol Cell* 2002; 10: 417-426 [PMID: 12191486 DOI: 10.1016/S1097-2765(02)00599-3]
- 15 Ioannou GN, Haigh WG, Thorning D, Savard C. Hepatic cholesterol crystals and crown-like structures distinguish NASH from simple steatosis. *J Lipid Res* 2013; 54: 1326-1334 [PMID: 23417738 DOI: 10.1194/jlr.M034876]
- 16 Ix JH, Sharma K. Mechanisms linking obesity, chronic kidney disease, and fatty liver disease: the roles of fetuin-A, adiponectin, and AMPK. J Am Soc Nephrol 2010; 21: 406-412 [PMID: 20150538 DOI: 10.1681/ASN.2009080820]
- Miura K, Yang L, van Rooijen N, Brenner DA, Ohnishi H, Seki E. Toll-like receptor 2 and palmitic acid cooperatively contribute to the development of nonalcoholic steatohepatitis through inflammasome activation in mice. *Hepatology* 2013; 57: 577-589 [PMID: 22987396 DOI: 10.1002/hep.26081]
- Ozaki E, Campbell M, Doyle SL. Targeting the NLRP3 inflammasome in chronic inflammatory diseases: current perspectives. *J Inflamm Res* 2015; 8: 15-27 [PMID: 25653548 DOI: 10.2147/JIR.S51250]
- 19 Takeuchi O, Akira S. Pattern recognition receptors and inflammation. *Cell* 2010; 140: 805-820 [PMID: 20303872 DOI: 10.1016/j.cell.2010.01.022]
- 20 Iracheta-Vellve A, Petrasek J, Gyongyosi B, Satishchandran A, Lowe P, Kodys K, Catalano D, Calenda CD, Kurt-Jones EA, Fitzgerald KA, Szabo G. Endoplasmic Reticulum Stress-induced Hepatocellular Death Pathways Mediate Liver Injury and Fibrosis via Stimulator of Interferon Genes. *J Biol Chem* 2016; 291: 26794-26805 [PMID: 27810900 DOI: 10.1074/jbc.M116.736991]
- 21 Bian H, Li F, Wang W, Zhao Q, Gao S, Ma J, Li X, Ren W, Qin C, Qi J. MAPK/p38 regulation of cytoskeleton rearrangement accelerates induction of macrophage activation by TLR4, but not TLR3. *Int J Mol Med* 2017; 40: 1495-1503 [PMID: 28949380 DOI: 10.3892/ijmm.2017.3143]
- 22 Hu W, Jain A, Gao Y, Dozmorov IM, Mandraju R, Wakeland EK, Pasare C. Differential outcome of TRIF-mediated signaling in TLR4 and TLR3 induced DC maturation. *Proc Natl Acad Sci USA* 2015; 112: 13994-13999 [PMID: 26508631 DOI: 10.1073/pnas.1510760112]
- Kanda T, Steele R, Ray R, Ray RB. Hepatitis C virus infection induces the beta interferon signaling pathway in immortalized human hepatocytes. *J Virol* 2007; 81: 12375-12381 [PMID: 17804510 DOI: 10.1128/JVI.01695-07]
- 24 Seki E, Brenner DA. Toll-like receptors and adaptor molecules in liver disease: update. *Hepatology* 2008; 48: 322-335 [PMID: 18506843 DOI: 10.1002/hep.22306]
- 25 Zhang B, Xu D, She L, Wang Z, Yang N, Sun R, Zhang Y, Yan C, Wei Q, Aa J, Liu B, Wang G, Xie Y. Silybin inhibits NLRP3 inflammasome assembly through the NAD + /SIRT2 pathway in mice with nonalcoholic fatty liver disease. FASEB J 2017 [PMID: 28970254 DOI: 10.1096/fj.201700602R]
- 26 Dolunay A, Senol SP, Temiz-Resitoglu M, Guden DS, Sari AN, Sahan-Firat S, Tunctan B. Inhibition of NLRP3 Inflammasome Prevents LPS-Induced Inflammatory Hyperalgesia in Mice: Contribution of NF-κB, Caspase-1/11, ASC, NOX, and NOS Isoforms. *Inflammation* 2017; 40: 366-386 [PMID: 27924425 DOI: 10.1007/s10753-016-0483-3]
- 27 Csak T, Velayudham A, Hritz I, Petrasek J, Levin I, Lippai D, Catalano D, Mandrekar P, Dolganiuc A, Kurt-Jones E, Szabo G. Deficiency in myeloid differentiation factor-2 and toll-like receptor 4 expression attenuates nonalcoholic steatohepatitis and fibrosis in mice. Am J Physiol Gastrointest Liver Physiol 2011; 300:



- G433-G441 [PMID: 21233280 DOI: 10.1152/ajpgi.00163.2009]
- Mridha AR, Wree A, Robertson AAB, Yeh MM, Johnson CD, Van Rooyen DM, Haczeyni F, Teoh NC, Savard C, Ioannou GN, Masters SL, Schroder K, Cooper MA, Feldstein AE, Farrell GC. NLRP3 inflammasome blockade reduces liver inflammation and fibrosis in experimental NASH in mice. *J Hepatol* 2017; 66: 1037-1046 [PMID: 28167322 DOI: 10.1016/j.jhep.2017.01.022]
- 29 Bhargava P, Lee CH. Role and function of macrophages in the metabolic syndrome. *Biochem J* 2012; 442: 253-262 [PMID: 22329799 DOI: 10.1042/BJ20111708]
- 30 Nallagangula KS, Nagaraj SK, Venkataswamy L, Chandrappa M. Liver fibrosis: a compilation on the biomarkers status and their significance during disease progression. *Future Sci OA* 2017; 4: FSO250 [PMID: 29255622 DOI: 10.4155/fsoa-2017-0083]
- 31 Soehnlein O, Lindbom L. Phagocyte partnership during the onset and resolution of inflammation. *Nat Rev Immunol* 2010; 10: 427-439 [PMID: 20498669 DOI: 10.1038/nri2779]
- 32 Fagoonee S, Famulari ES, Silengo L, Camussi G, Altruda F. Prospects for Adult Stem Cells in the Treatment of Liver Diseases. Stem Cells Dev 2016; Epub ahead of print [PMID: 27503633 DOI: 10.1089/scd.2016.0144]
- 33 Wu X, Wu X, Ma Y, Shao F, Tan Y, Tan T, Gu L, Zhou Y, Sun B, Sun Y, Wu X, Xu Q. CUG-binding protein 1 regulates HSC activation and liver fibrogenesis. *Nat Commun* 2016; 7: 13498 [PMID: 27853137 DOI: 10.1038/ncomms13498]
- 34 Geerts A. History, heterogeneity, developmental biology, and

- functions of quiescent hepatic stellate cells. *Semin Liver Dis* 2001; **21**: 311-335 [PMID: 11586463 DOI: 10.1055/s-2001-17550]
- 35 Eng FJ, Friedman SL. Fibrogenesis I. New insights into hepatic stellate cell activation: the simple becomes complex. Am J Physiol Gastrointest Liver Physiol 2000; 279: G7-G11 [PMID: 10898741 DOI: 10.1152/ajpgi.2000.279.1.G7]
- 36 Seki E, Schwabe RF. Hepatic inflammation and fibrosis: functional links and key pathways. *Hepatology* 2015; 61: 1066-1079 [PMID: 25066777 DOI: 10.1002/hep.27332]
- 37 Duarte S, Baber J, Fujii T, Coito AJ. Matrix metalloproteinases in liver injury, repair and fibrosis. *Matrix Biol* 2015; 44-46: 147-156 [PMID: 25599939 DOI: 10.1016/j.matbio.2015.01.004]
- 38 Henao-Mejia J, Elinav E, Jin C, Hao L, Mehal WZ, Strowig T, Thaiss CA, Kau AL, Eisenbarth SC, Jurczak MJ, Camporez JP, Shulman GI, Gordon JI, Hoffman HM, Flavell RA. Inflammasome-mediated dysbiosis regulates progression of NAFLD and obesity. *Nature* 2012; 482: 179-185 [PMID: 22297845 DOI: 10.1038/nature10809]
- Pierantonelli I, Rychlicki C, Agostinelli L, Giordano DM, Gaggini M, Fraumene C, Saponaro C, Manghina V, Sartini L, Mingarelli E, Pinto C, Buzzigoli E, Trozzi L, Giordano A, Marzioni M, De Minicis S, Uzzau S, Cinti S, Gastalderi A, Svegliati-Baroni G. Lack of NLRP3-inflammasome leads to gut-liver axis derangement, gut dysbiosis and a worsened phenotype in a mouse model of NAFLD. Sci Rep 2017; 7: 12200 [PMID: 28939830 DOI: 10.1038/s41598-017-11744-6]

P- Reviewer: Kanda T, Ozenirler S, Pellicano R, Sazci A S- Editor: Cui LJ L- Editor: A E- Editor: Li RF





Published by Baishideng Publishing Group Inc

7901 Stoneridge Drive, Suite 501, Pleasanton, CA 94588, USA

Telephone: +1-925-223-8242

Fax: +1-925-223-8243

E-mail: bpgoffice@wjgnet.com

Help Desk: http://www.f6publishing.com/helpdesk

http://www.wjgnet.com

