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**Clinical significance of exosomes as potential biomarkers in cancer**

Wong CH *et al*. Clinical significance of exosomes

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**Abstract**

***BACKGROUND***

Exosomes are microvesicles with 30-100 nm in diameter. They are widely distributed in body fluids, including blood, bile, urine and saliva. Cancer-derived exosomes carry a wide variety of DNA, RNA, proteins and lipids, and may serve as novel biomarkers in cancer.

***AIM***

To summarize the performance of exosomal biomarkers in cancer diagnosis and prognosis.

***Methods***

Relevant literatures were identified using “PubMed” database until September 11, 2018. The quality of the included studies was assessed by QUADAS-2 and REMARK. For diagnostic biomarkers, 47 biomarkers and 2240 patients from 30 studies were included

***Results***

Our results suggested that these exosomal biomarkers had excellent diagnostic ability in various types of cancer with good sensitivity and specificity. For prognostic markers, 50 biomarkers and 4797 patients from 42 studies were included. We observed that exosomal biomarkers had prognostic values in overall survival, disease-free survival and recurrence-free survival.

***Conclusion***

Exosomes can function as potential biomarkers in cancer diagnosis and prognosis.

**Key words:** Exosome; Biomarker; Cancer; Diagnosis; Prognosis

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**Core tip:** Cancer-derived exosomes carry a wide variety of DNA, RNA, proteins and lipids, which may serve as novel biomarkers in cancer. The current systematic review and meta-analysis summarized the performance of exosomal biomarkers in cancer diagnosis and prognosis. We analyzed 47 diagnostic markers and 50 prognostic markers from 56 studies with various type of cancer. We found that exosomal biomarkers had both diagnostic and prognostic power in many cancers.

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**INTRODUCTION**

Cancer is the uncontrolled growth of cells and eventually leads to death. Cancer is the second cause of death which contributes to more than 8.8 million deaths every year[1,2]. Among various types of cancer, lung cancer, gastrointestinal cancers (GI cancer), including liver cancer, pancreatic cancer and colorectal cancer, and breast cancer are the most common cause of cancer-related death[2-4]. Although chemotherapy, targeted therapy, surgical recession and radiotherapy can effectively prolong survival of patients, the survival rate of cancer is still very low, especially in GI cancer with less than 20%[2]. One of the major reasons is the late diagnosis of cancer, in which patients are already with advanced and metastatic tumors. As a result, no therapies can effectively kill the cancer cells. The situation is even worse in pancreatic cancers at distant stage with 5-year survival rate of only 3%[2].

Since more than half of the patients present with locally advanced or metastatic stage, early diagnosis and early treatment are fundamentally important for better prognosis. Therefore, many tumor makers have been developed, aiming at accurately detecting various types of cancer and monitoring the disease progression. Blood test of tumor antigens carcinoembryonic antigen (CEA), carbohydrate antigen 19-9 (CA19-9), carbohydrate antigen 125 (CA125) are commonly used for detection of many cancers, such as GI cancers, ovarian cancer and breast cancer[5-8]. However, the sensitivity of these cancer biomarkers is unsatisfactory[9-12]. Also, the fecal occult blood test of colorectal cancer and the invasion endoscopic detection of gastric and colon cancer give great inconvenience to the patients. Therefore, highly sensitive and non-invasive diagnostic markers are urgently needed for early detection of cancer.

Exosomes are microvesicles with 30-100 nm diameter which are secreted by both normal cells and cancer cells. They are distributed in many body fluids such as blood, saliva and urine, and carry various types of biomolecules including RNA, proteins and lipids for inter-cellular communication[13-15]. During cancer development, cancer cells secrete more exosomes with significant changes in composition[16-18]. These facilitate the communication within tumor environment, acquisition of drug resistance and metastasis to distant organs[19-21]. Although many potential non-invasive biomarkers are developed using liquid biopsy such as serum and urine, studies found that these biomarkers are commonly located in the exosomes[22,23]. Enriching these exosomal biomarkers could achieve a higher diagnostic and prognostic efficiency[24-26]. This demonstrated that exosomal biomarkers can be novel targets in cancer diagnosis and prognosis.

The objective of this systemic review and meta-analysis is to evaluate the diagnostic and prognostic potential of exosomes in patients with various types of cancer, based on current available data. This helps the development of novel non-invasion biomarkers for sensitive and specific diagnosis and prognosis of cancer.

**MATERIALS AND METHODS**

***Search strategy***

Electronic literature search was performed using PubMed database without any language restriction. Articles related to exosomes in cancer from 2010 to September 11, 2018 were identified using the following keywords: “exosome” and “cancer” and ““diagnosis” or “prognosis””.

***Inclusion and exclusion criteria***

Articles were reviewed by their titles, keywords, abstracts and full text to identify eligible studies. Eligible studies were included based on the following inclusion criteria: (1) The original article was related to exosomal diagnostic or prognostic markers in cancer; (2) At least 10 patients and 10 matched controls were enrolled in the study; (3) For diagnostic markers, enough information such as specificity and sensitivity was provided to construct 2x2 table [true positive (TP), true negative (TN), false positive (FP), false negative (FN)]; (4) For prognostic markers, enough information was provided to estimate the hazard ratios and its confidence interval (CI); The exclusion criteria were as follows: (1) duplicate articles; (2) review articles, abstracts, comments, letters, case-report; (3) fundamental research or animal study; (4) diagnostic or prognostic marker was not specific to exosome; (5) size sample was less than 10; (6) the performance of the biomarker was not statistically significant; and(7) incomplete information to estimate its diagnostic or prognostic accuracy.

***Data extraction***

Two reviewers (Chi-Hin Wong and Yang-Chao Chen) independently reviewed and extracted the data from the eligible studies according to the listed criteria. Any disagreement was resolved by consensus among the authors. The following data from included studies were extracted: first author’s name, year of publication, sample size, cancer type, and country of origin, source of exosome, isolation method of exosome, and detection method of biomarkers. For diagnostic studies, cut-off value of tested targets, sensitivity, specificity, and area under the Receiver operating characteristics curve (AUC) were also extracted. For prognostic studies, survival analysis, cut-off value, multivariable hazard ratio (HR) and its 95% CI were extracted. If odds ratio (OR) was reported, OR was converted to relative risk using the formula introduced by Zhang[27]. If either OR or HR was not reported, method introduced by Tierney *et al*[28] was used to estimate the HR and its 95% CI from Kaplan-Meier plot.

***Quality assessment***

For diagnostic studies, the Quality Assessment of Diagnostic Accuracy Studies (QUADAS-2) was used to assess the quality of studies for the meta-analysis[29]. Briefly, 14 questions covering the patient selection, patient flow, index test and reference standard test were applied to each study and an answer of “Yes”, “No” or “Unclear” was given to each study. Only answer with “Yes” was given a score.

For prognostic study, the quality of studies was assessed according to reporting recommendations for tumor marker prognostic studies (REMARK)[30]. Briefly, a checklist of 20 items covering patients’ characteristics, samples’ source and storage, assay methods, statistical analysis and data interpretation. A score was given when the study fulfills the requirement of each item.

***Statistical analysis***

The statistical analysis of the diagnostic performance of biomarkers was performed using Meta-DiSc 1.4[31]. The 2x2 table of each study was used to assess the pooled sensitivity, specificity, positive likelihood ratio (PLR) and negative likelihood ratio (NLR). Also, the summary receiver operating characteristic curve (SROC) curve was plotted; and the area under the curve (AUC) was calculated and Q\*index was estimated to assess the overall performance in cancer diagnosis. An AUC of 0.5 suggests no diagnostic ability; 0.7-0.8 suggests acceptable diagnostic performance; 0.8-0.9 is considered excellent, and 0.9-1.0 suggests outstanding performance[32]. Q\* was defined at a point in which sensitivity and specificity are equal. For statistical analysis of the prognostic performance of biomarkers, forest plots were constructed using the HR and its 95%CI of each biomarkers to assess the overall prognostic performance of biomarkers on OS, DFS and RFS. Graphpad Prism 6 was used in constructing the forest plots. To elevate the heterogeneity between studies, Cochran-Q test and Inconsistency index (*I*2) statistics were calculated[33,34]. *P*-value of < 0.05 for Cochran-*Q* test or *I*2 >50% suggested the presence of heterogeneity.

**RESULTS**

***Literature search***

Initially, 1233 articles were identified based on the search strategies. Based on title and abstract screening, 705 were not related to exosome biomarkers in cancer diagnosis or prognosis, 287 were reviews articles. Upon further full-text review, 56 studies were basic studies; 42 studies with sample size less than 10 in either group (test group or control group); 12 studies analyzed the performance of combined markers; 70 studies did not provide enough information for analysis; 5 studies were without statistical significance. Finally, 56 eligible studies were included for systematic review (Figure 1). Of these, 22 candidate studies were related to diagnosis; 34 candidate studies were related to prognosis; and 8 studies were related to both diagnosis and prognosis.

***Assessment of study quality***

For diagnostic studies, QUADAS-2 system was used to assess the quality of study (Figure 2A). Most of the studies on diagnosis were with moderate-to-high quality, revealed by low risk of publication bias. However, there may be risk of bias in “patient selection” and “flow and timing”. This may due to control-based design in most of the studies. Also, time between the index test and the reference test is poorly reported. Importantly, many studies did not provide enough information on how the patients were selected and classified. Patients excluded from 2 x 2 table were often observed in some studies.

REMARK system was used to assess the quality of prognostic studies (Figure 2B). Most of the studies (> 90%) clearly stated the objective, biomarkers examined, the source of exosomes and the methodology of isolation and detection. Also, most of the studies clearly defined the clinical endpoints and the period of the follow-up time. However, details in patient’s characteristics during the follow-up period, such as the use of post-operative adjuvant therapy which significantly affects the overall survival and disease-free survival, were lacking in most of the studies. Importantly, some studies did not clearly report the clinicpathological characteristics of the patients enrolled. Also, some studies did not show the relationship of the tested biomarkers to prognostic variables, including tumor stages and tumor differentiation. 12 prognostic marker studies did not perform univariable or multivariable analysis. 28 of the enrolled studies reported multivariable analysis in prognostic markers, but only 5 studies clearly stated the adjustment factors.

**RESULTS**

**Diagnostic markers:** Diagnostic markers from 30 studies were recruited for meta-analysis (Table1). More than a half of enrolled studies were related to GI cancers (4 studies were about colon cancer; 5 studies were related to liver cancer; 4 studies were about pancreatic or pancreatobiliary tract cancer and 4 studies were related to gastric cancer). 2240 patients were enrolled in the meta-analysis. 12 studies enrolled < 50 patients; 16 studies enrolled 50-100 patients; and 6 studies enrolled > 100 patients. There were 47 diagnostic biomarkers analyzed in the meta-analysis. 42.6% of the biomarkers were miRNAs, followed by lncRNAs (36.2%) and proteins (19.1%). Notably, 6 studies analyzed the diagnostic performance of exosomal miR-21 in various types of cancer. Also, 61.3%, 16.1%, 12.9%, 3.2% and 3.2% of the biomarkers were detected in serum, plasma, urine, saliva and bile respectively.

Since a wide range of cancers was studied by different groups, we separated the diagnostic biomarkers according to cancer types and meta-analyzed cancer types with more than three biomarkers studied. Therefore, we focused on colorectal cancer (4 studies with 11 biomarkers), gastric cancer (4 studies with 5 biomarkers), pancreatic cancer (4 studies with 8 biomarkers), liver cancer (4 studies with 7 biomarkers) and prostate cancer (4 studies with 7 biomarkers (Figures 3-7). We observed that the pooled biomarkers had a good specificity of 0.87 but poor sensitivity of 0.57 and in colorectal cancer diagnosis (Figure 3A and B). The PLR and NLR were 2.02 and 0.21 respectively (Figure 3C and D). The diagnostic odds ratio was 20.35 (Figure 3E). Importantly, the AUC of the SROC curve was 0.89 and the Q\* was 0.82 (Figure 3F). In diagnosis of gastric cancer, we observed the pooled biomarkers had a good sensitivity of 0.77 and specificity of 0.73 with PLR, NLR, AUC of the SROC curve and Q\* of 2.94, 0.32, 9.88, 0.84 and 0.77 respectively (Figure 4). For diagnosis of pancreatic cancer, we also observed the pooled biomarkers had an excellent sensitivity of 0.91 and specificity of 0.90 with PLR, NLR, AUC of the SROC curve and Q\* of 6.35, 0.19, 40.71, 0.94 and 0.88 respectively (Figure 5). In liver cancer, the pooled biomarkers had a good diagnostic sensitivity of 0.76 and specificity of 0.80 with PLR, NLR, AUC of the SROC curve and Q\* of 3.51, 0.32, 12.45, 0.85 and 0.78 respectively (Figure 6). The pooled biomarkers also had a good sensitivity of 0.77 and specificity of 0.79 in detecting prostate cancer with PLR, NLR, AUC of the SROC curve and Q\* of 3.84, 0.28, 17.88, 0.88 and 0.80 respectively (Figure 7). The high sensitivity, specificity and Q\* demonstrated that the pooled biomarkers could effectively discriminating cancer patients from healthy people or non-cancer patients.

***Prognostic markers***

Prognostic biomarkers from 42 studies were recruited for systematic review (Table 2). 4797 patients were enrolled in this systemic review. 7 studies enrolled < 50 patients; 15 studies enrolled 50-100 patients; and 20 studies enrolled >100 patients. There were 50 prognostic biomarkers analyzed in the systematic review. 60% of the biomarkers were miRNAs, followed by lncRNAs (18%) and proteins (16%). Also, 50%, 43%, 2.4%, 2.4% and 2.4% of the biomarkers were detected in serum, plasma, bile, ascetic fluid and cell-free effusion supernatant respectively. 92.9%, 26.2% and 9.5% of the enrolled studies used OS, DFS and RFS respectively as the primary endpoints. In addition, a wide range of cancers was studied by different groups. More than a half of enrolled studies were related to GI cancers (11 studies were about colorectal or colon cancer; 5 studies were related to liver cancer; 5 studies were about pancreatic cancer and 4 studies were related to gastric cancer). In this meta-analysis, we separated studies based on clinical endpoints and focused on cancer types with more than three biomarkers studied.

For 13 biomarkers with OS reported in colon cancer, the pooled HR was 1.833 with *I2* of 62.14% and *P* = 0.002 (Figure 8A). Also, for 5 biomarkers with DFS reported in colon cancer, the pooled HR was 3.035 with *I2* of 0.00% and *P* = 0.536 (Figure 8B). Furthermore, for 4 biomarkers with RFS reported in colon cancer, the pooled HR was 1.645 with *I2* of 89.61% and *P* = 0.000 (Figure 8C). Apart from colon cancer, 4 biomarkers with OS reported in gastric cancer. The pooled HR was 1.836 with *I2* of 96.71 and *P* = 0.000 (Figure 9). In addition, 4 biomarkers with OS reported in pancreatic cancer. The pooled HR was 1.537 with *I2* of 81.50 and *P* = 0.001 (Figure 10). 5 biomarkers with the pooled HR was 1.828, *I2* of 84.48% and *P* = 0.000 were reported to prognosing OS in liver cancer (Figure 11). Also, 9 biomarkers with the pooled HR of 0.895, I2 of 89.50% and *P* = 0.000 were reported to function as prognostic biomarkers of OS in lung cancer (Figure 12). These results demonstrated that exosomes were associated with OS, DFS and RFS in various types of cancer.

**DISCUSSION**

Exosomes play important roles in cancer development via inter-cellular communication, promoting cell metastasis and developing drug resistance[19-21]. Importantly, exosomes are frequently secreted by the cancers and are widely distributed in many body fluids. Therefore, they can be detected in blood, saliva and urines. Exosomal biomarkers have better performances in cancer diagnosis and prognosis than only using liquid biopsy[24-26]. However, the methods of isolating exosomes from liquid biopsy varies between studies. Ultracentrifugation or the use of commercial isolation kits are common methods in extracting exosomes. Ultracentrifugation gives highly pure exosomes but the isolation efficiency is relatively low; whereas the use of commercial kits maximizes the efficiency with the loss of purity[95,96]. Therefore, a standardized protocol of detecting exosomal biomarkers is greatly needed.

There are some limitations of our meta-analysis. We excluded studies that utilized combined biomarkers because this cannot tell the performance of individual biomarkers[97,98]. For examples, a six-microRNA panel was development for diagnosis of lung cancer but miR-409-3p, miR-425-5p and miR-584-5p were not significantly dysregulated in patients’ exosomes[98]. This may reduce the diagnostic performance of other biomarkers in the same panel. Since many of the individual biomarkers in the panel were significantly differentially expressed in cancer exosomes, further studies may be needed to explore the correlation of these potential biomarkers with patients’ characteristics and their performances in cancer diagnosis and prognosis.

A further limitation is that we focused on exosomal markers only in cancer diagnosis and prognosis and excluded tissue-based biomarkers from this meta-analysis. In fact, many studies reported that expression levels in exosomes and in tissues are highly associated[35,66]. This suggested that many exosomal markers can reflect the situation in cancer cells and has been developed as potential biomarkers in various cancers. Importantly, this strong association may also suggest that many tissue-based biomarkers can be developed into non-invasive exosomal biomarkers in cancer diagnosis.

Notably, most of the included studies are retrospective that is performed on stored samples. However, the main disadvantage of retrospective study is the lack of complete clinicpathological information[30], which lowers the quality of study. Despite the above limitations, our meta-analysis indicates that exosomes can be potential biomarkers in cancer diagnosis and prognosis. Further large prospective studies are greatly needed to clarity the performance of exosomal biomarkers in cancer diagnosis and prognosis.

**ARTICLE HIGHLIGHTS**

***Research background***

Exosomes, which are widely distributed in body fluids, including blood, bile, urine and saliva, are microvesicles with 30-100 nm in diameter. Cancer-derived exosomes carry a wide variety of DNA, RNA, proteins and lipids, and may serve as novel biomarkers in cancer.

***Research motivation***

Exosomes may function as exosomal biomarkers in cancer diagnosis and prognosis.

***Research objectives***

To summarize the performance of exosomal biomarkers in cancer diagnosis and prognosis.

***Research methods***

Relevant literatures were identified using PubMed database. QUADAS-2 and REMARK were used to assess the quality of the included studies. For diagnostic biomarkers, 47 biomarkers and 2240 patients from 30 studies were included.

***Research results***

These exosomal biomarkers had excellent diagnostic ability in various types of cancer with good sensitivity and specificity. 50 biomarkers and 4797 patients from 42 studies were included for prognostic markers. We observed that exosomal biomarkers had prognostic values in overall survival, disease-free survival and recurrence-free survival.

***Research conclusions***

Exosomes could be potential biomarkers in cancer diagnosis and prognosis.

***Research perspectives***

Further large prospective studies are needed to clarity the performance of exosomal biomarkers in cancer diagnosis and prognosis, through exosomes can be potential biomarkers in cancer diagnosis and prognosis.

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**P-Reviewer:** Balaban YH, Treeprasertsuk S **S-Editor:** Cui LJ

**L-Editor:** **E-Editor:**

**Specialty type:** Medicine, research and experimental

**Country of origin:** China

**Peer-review report classification**

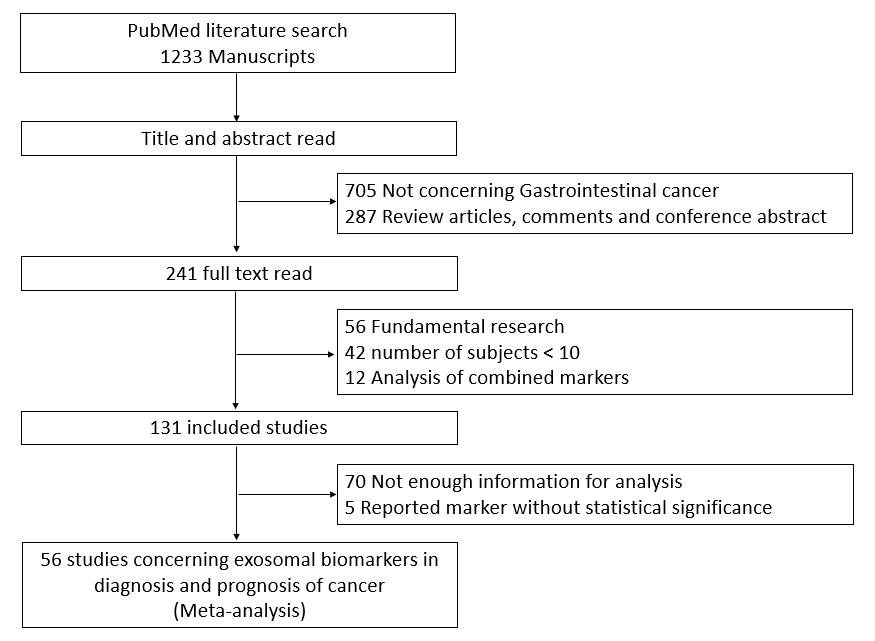
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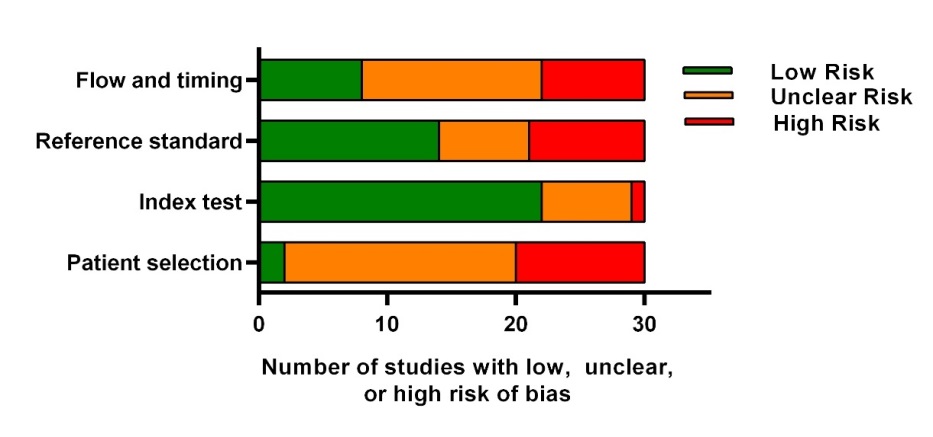
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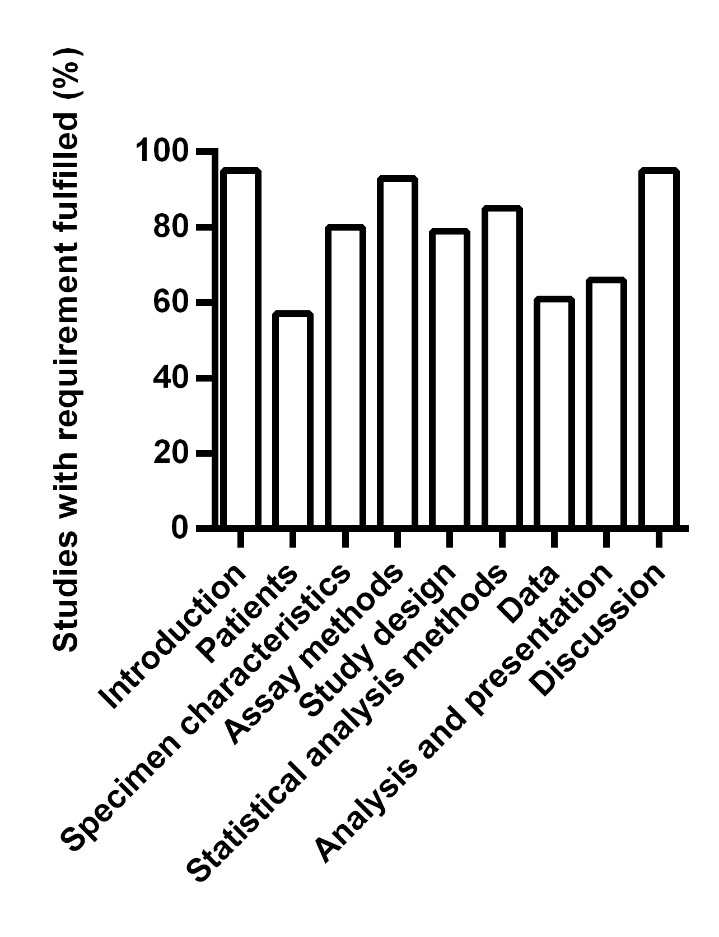


**Figure 1 Literature search process to select studies which elevating the diagnostic or prognostic performance of exosomal biomarkers in cancer.**

**A**



**B**



**Figure 2 Quality assessment of the studies in meta-analysis.** A: QUADAS-2 system was used to assess the quality of diagnostic studies; B: REMARK checklist was used to assess the quality of prognostic studies.

A

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自動產生的描述

B

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自動產生的描述

C

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自動產生的描述

D

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自動產生的描述

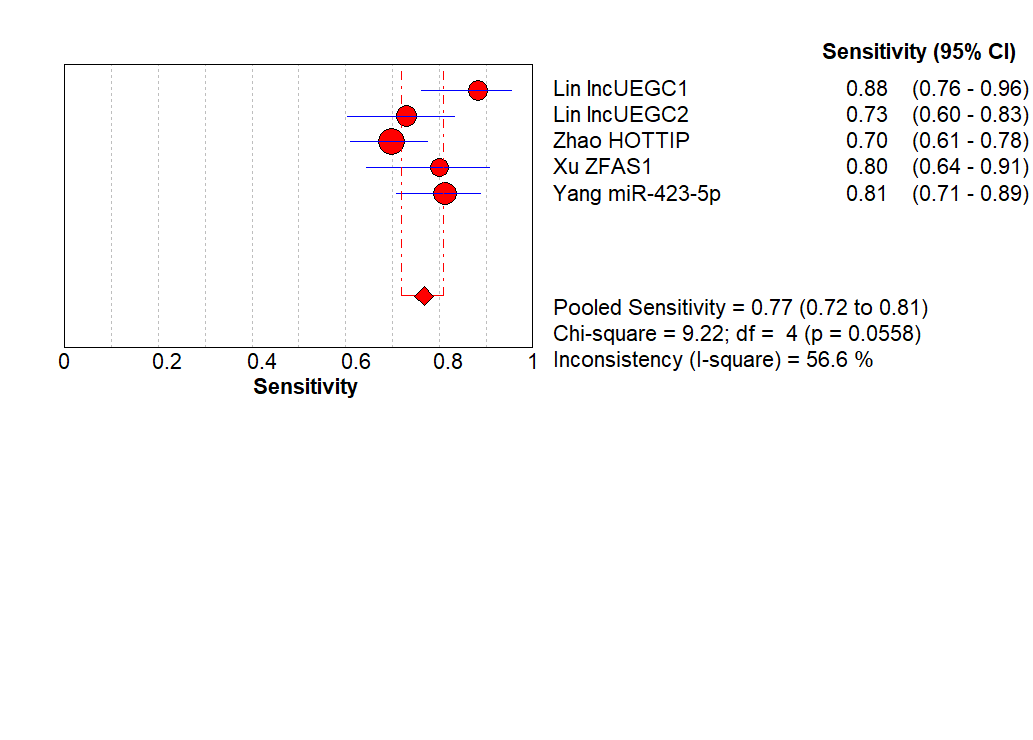
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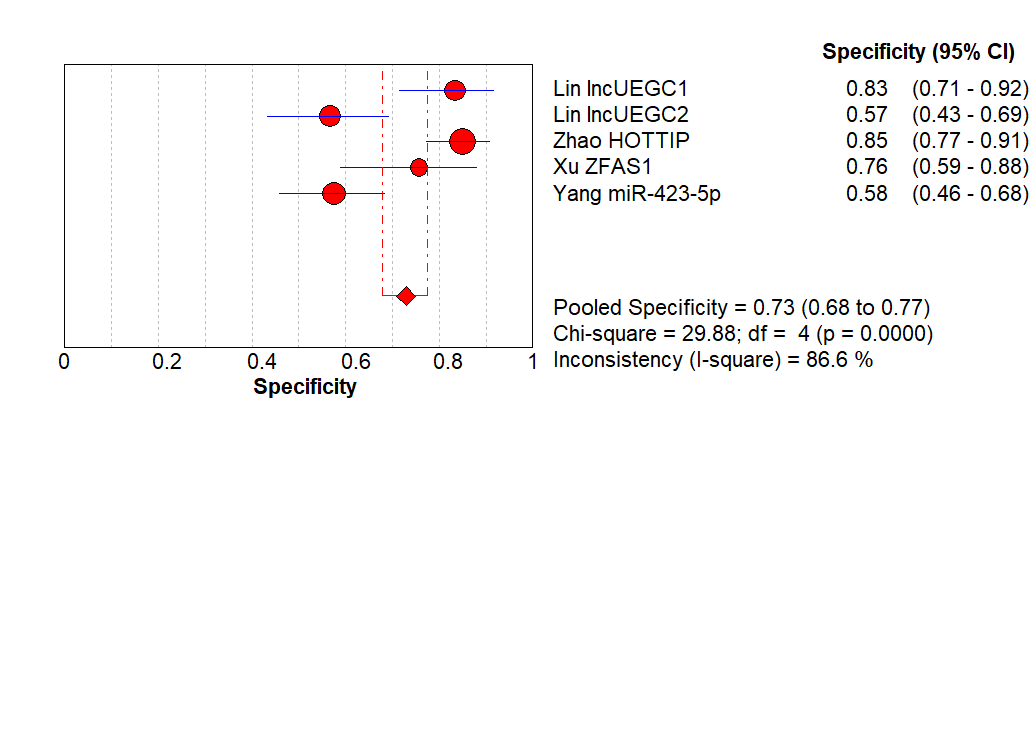
自動產生的描述

**Figure 3 Forest plot of pooled (A) sensitivity, (B) specificity, (C) positive likelihood ratio, (D) negative likelihood ratio, (E) diagnostic odds ratio and (F) SROC curve of exosomal biomarkers in diagnosis of colon cancer.**

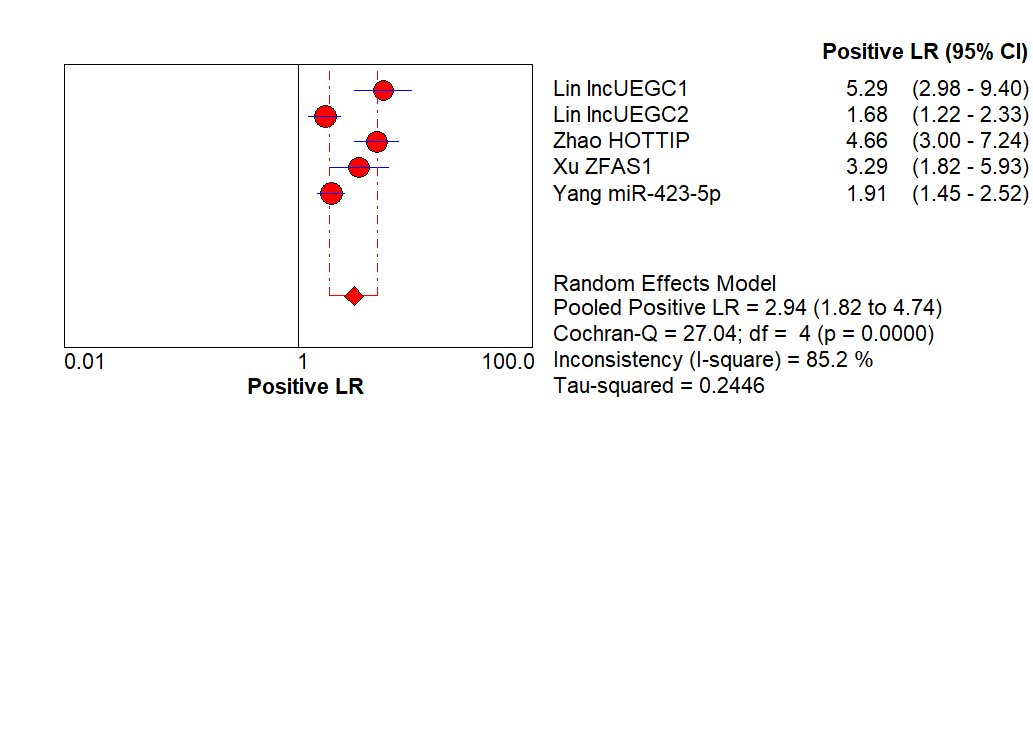
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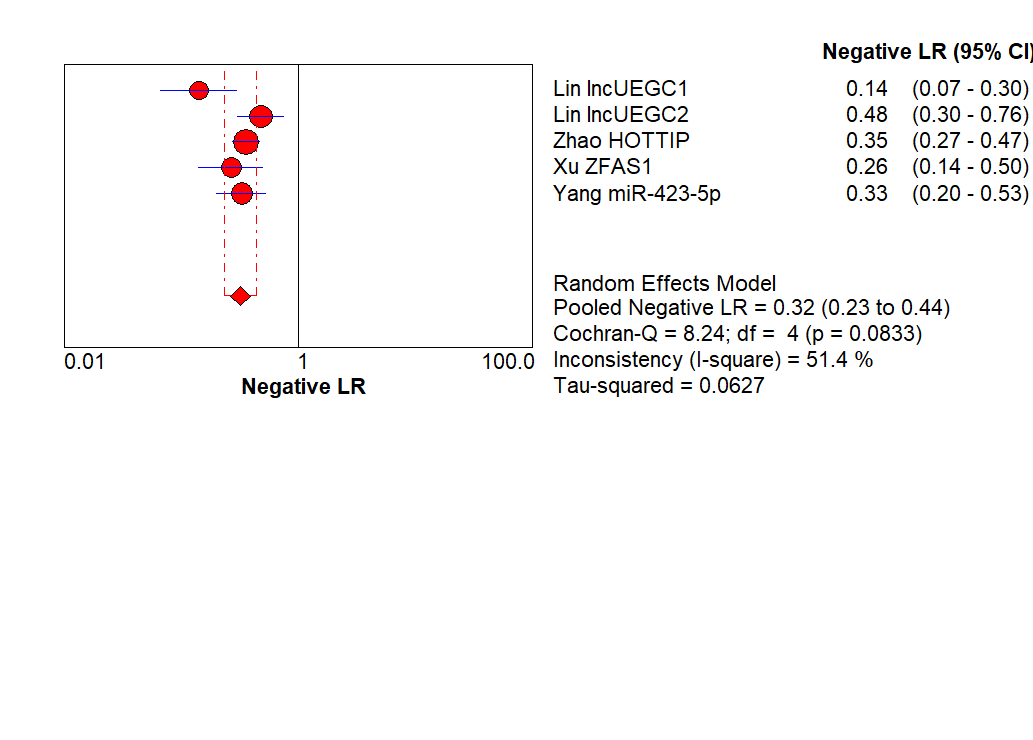
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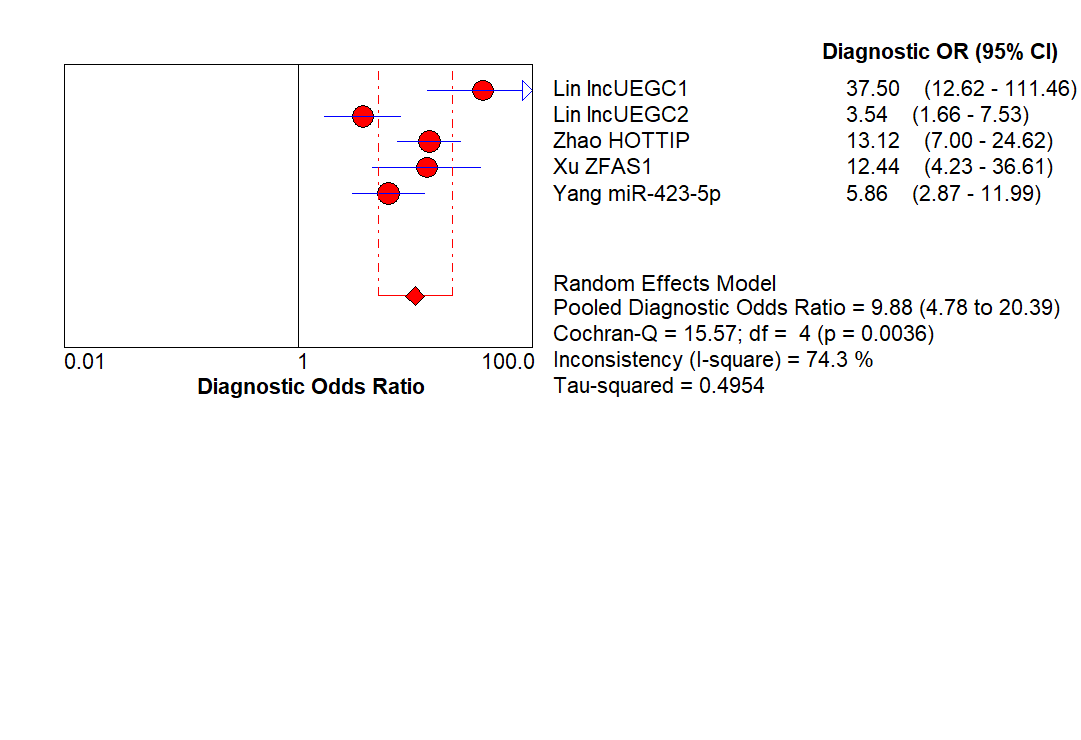
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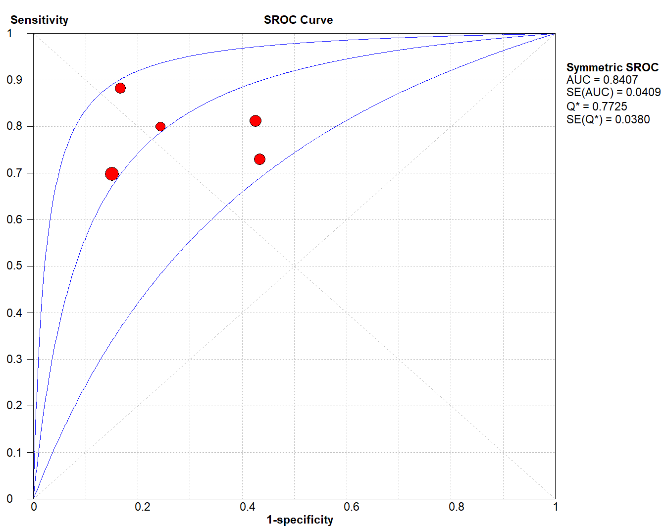
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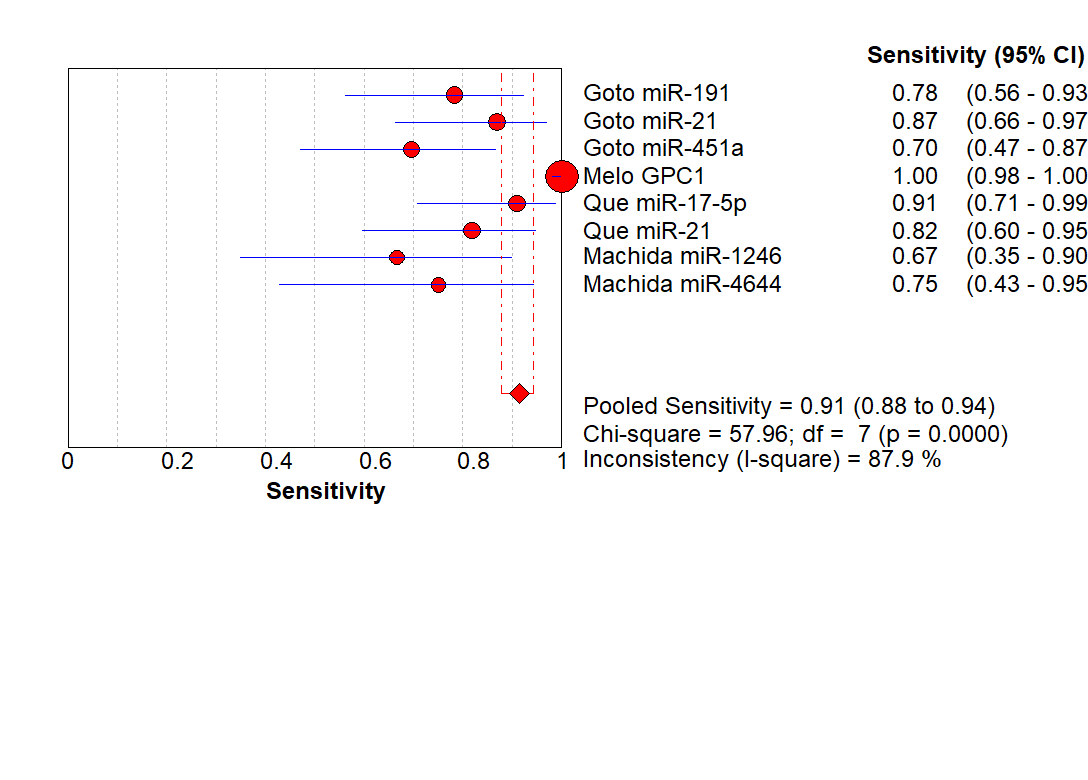


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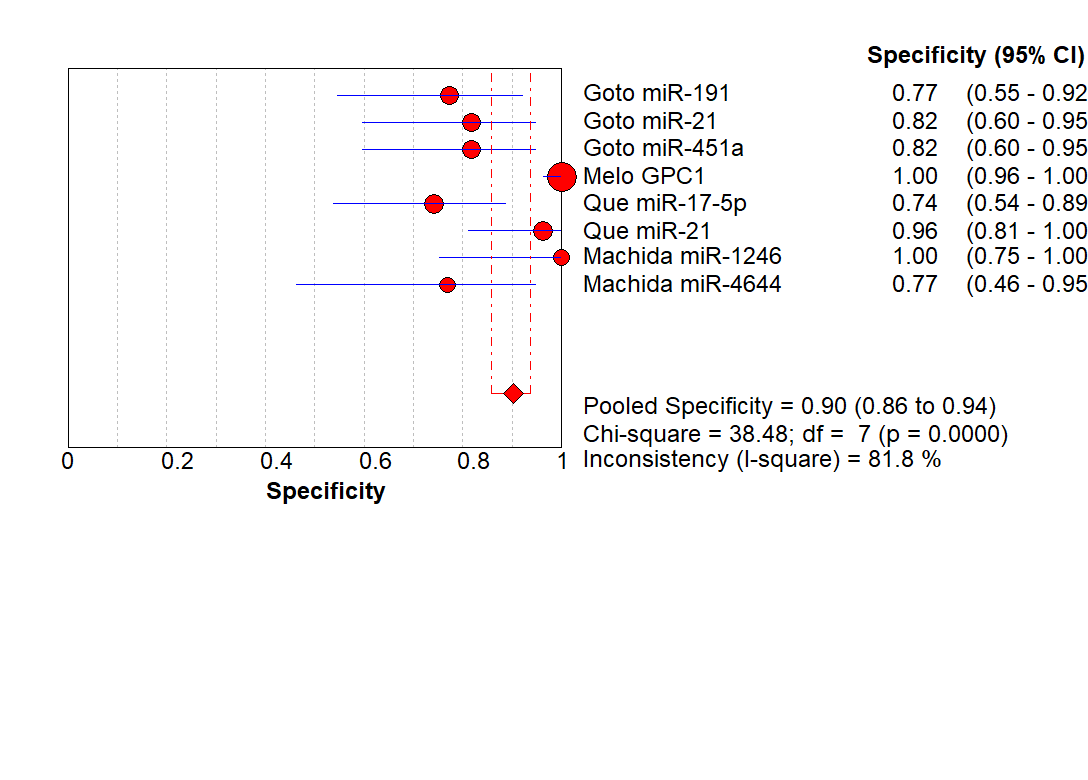


**Figure 4 Forest plot of pooled (A) sensitivity, (B) specificity, (C) positive likelihood ratio, (D) negative likelihood ratio, (E) diagnostic odds ratio, and (F) SROC curve of exosomal biomarkers in diagnosis of gastric cancer.**

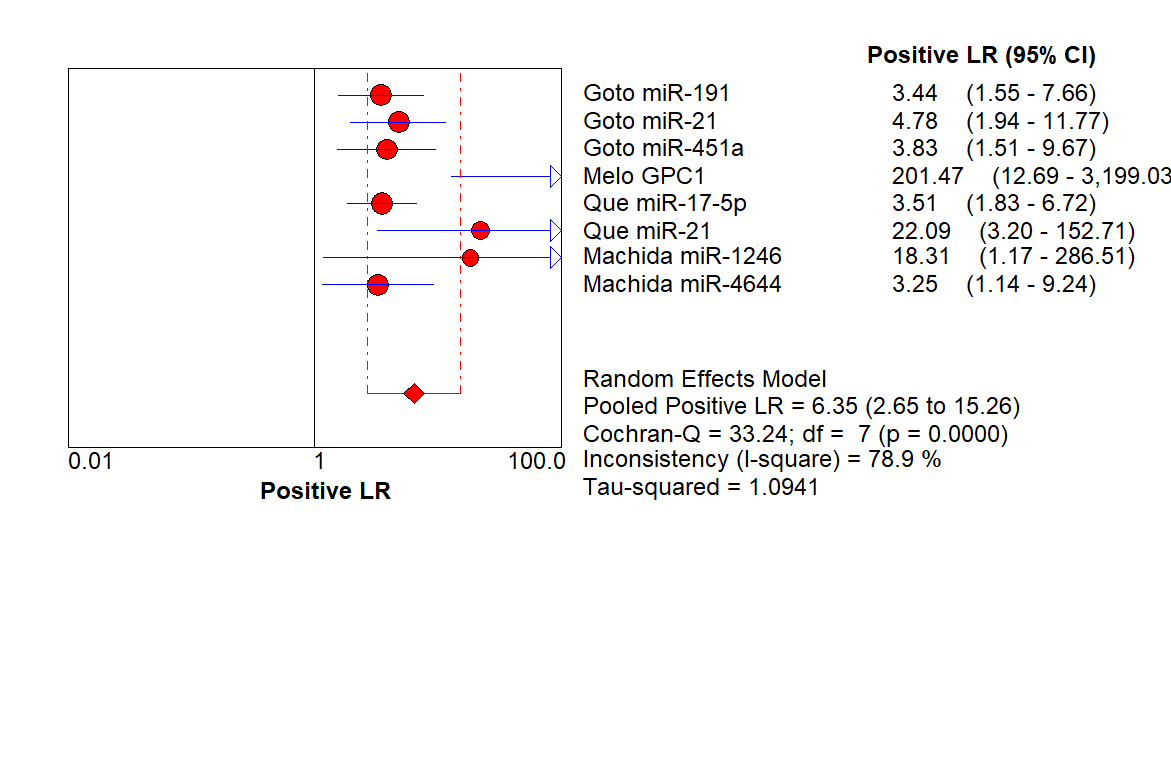
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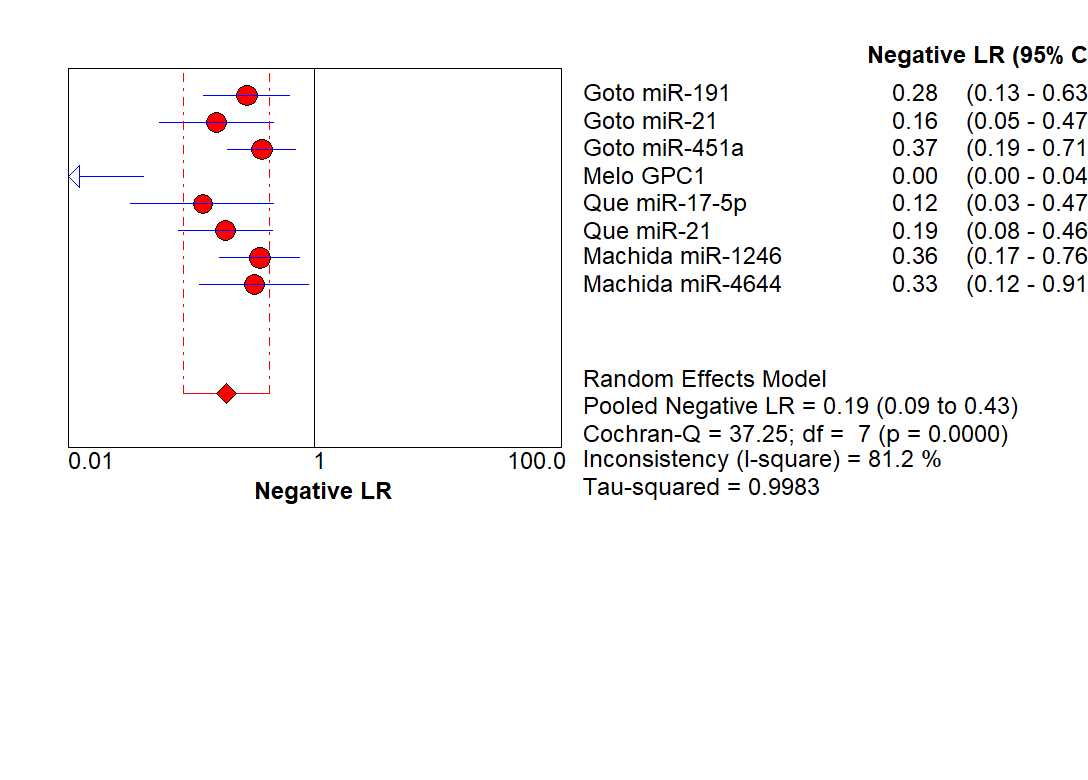
B



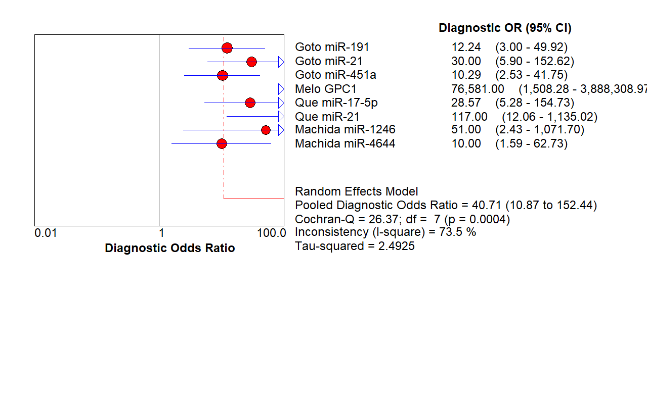
C



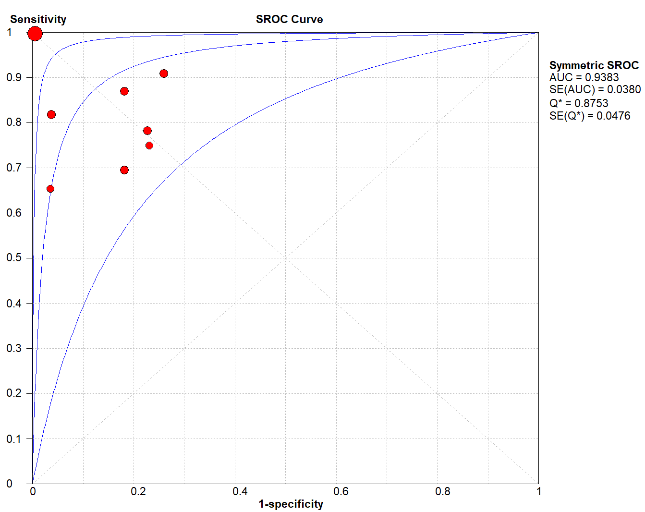
D



E

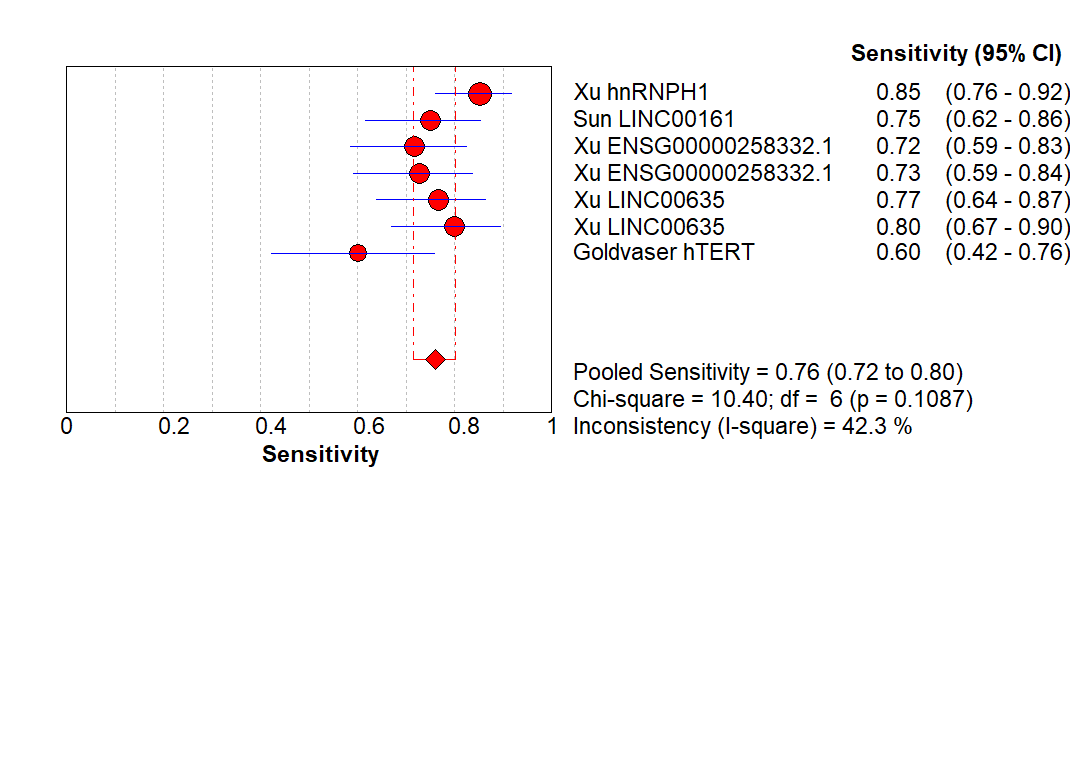


F

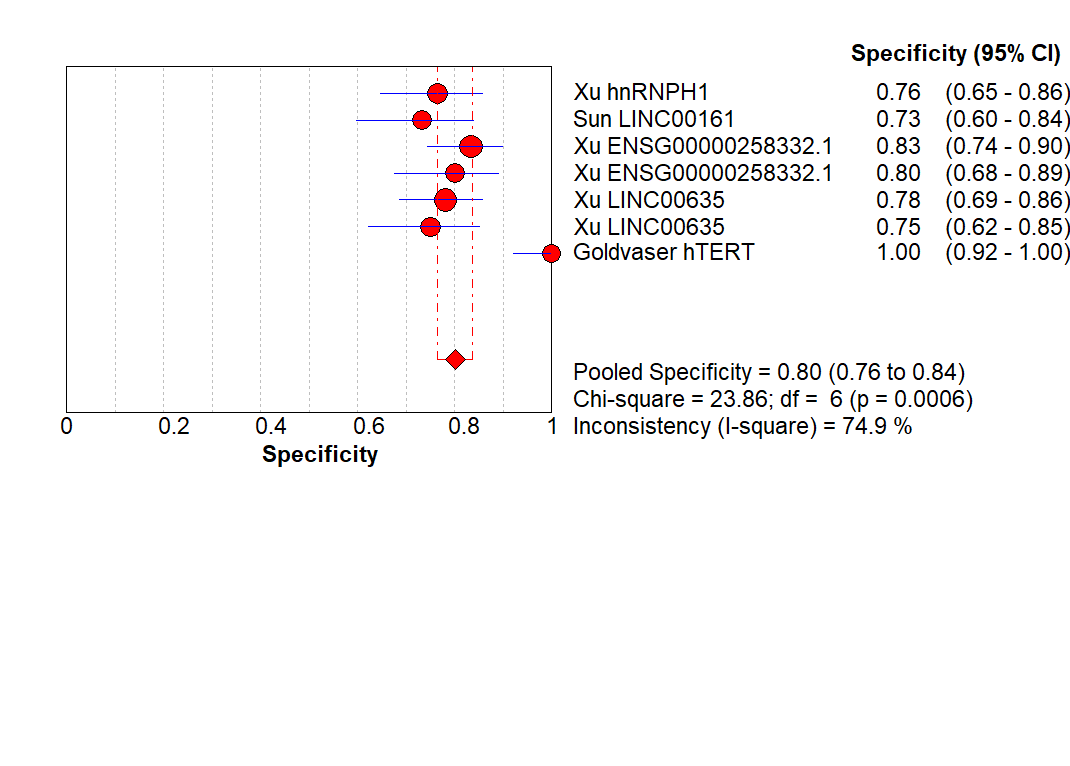


**Figure 5 Forest plot of pooled (A) sensitivity, (B) specificity, (C) positive likelihood ratio, (D) negative likelihood ratio, (E) diagnostic odds ratio and (F) SROC curve of exosomal biomarkers in diagnosis of pancreatic cancer.**

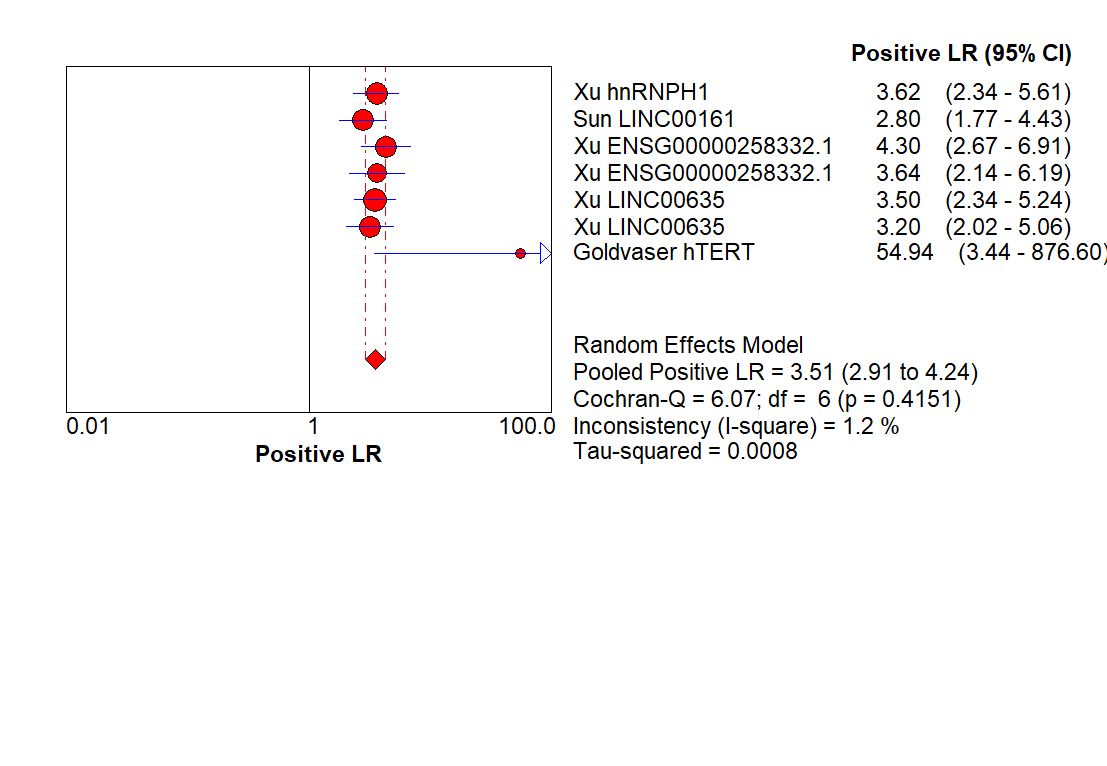
A



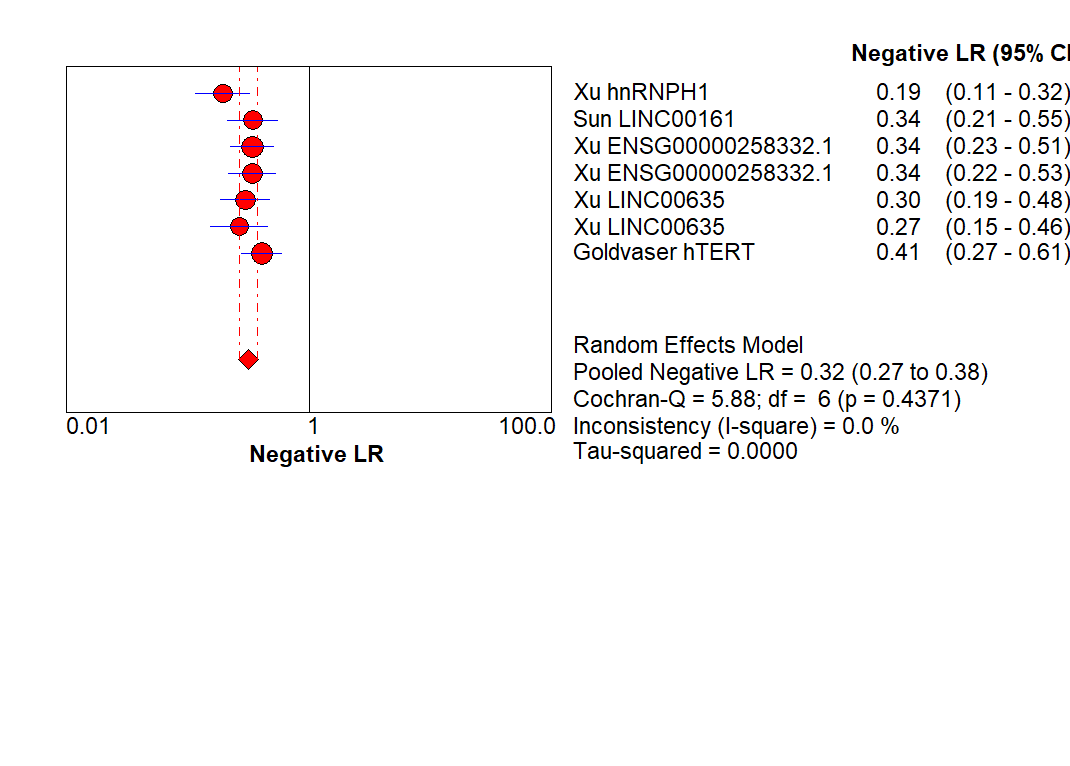
B



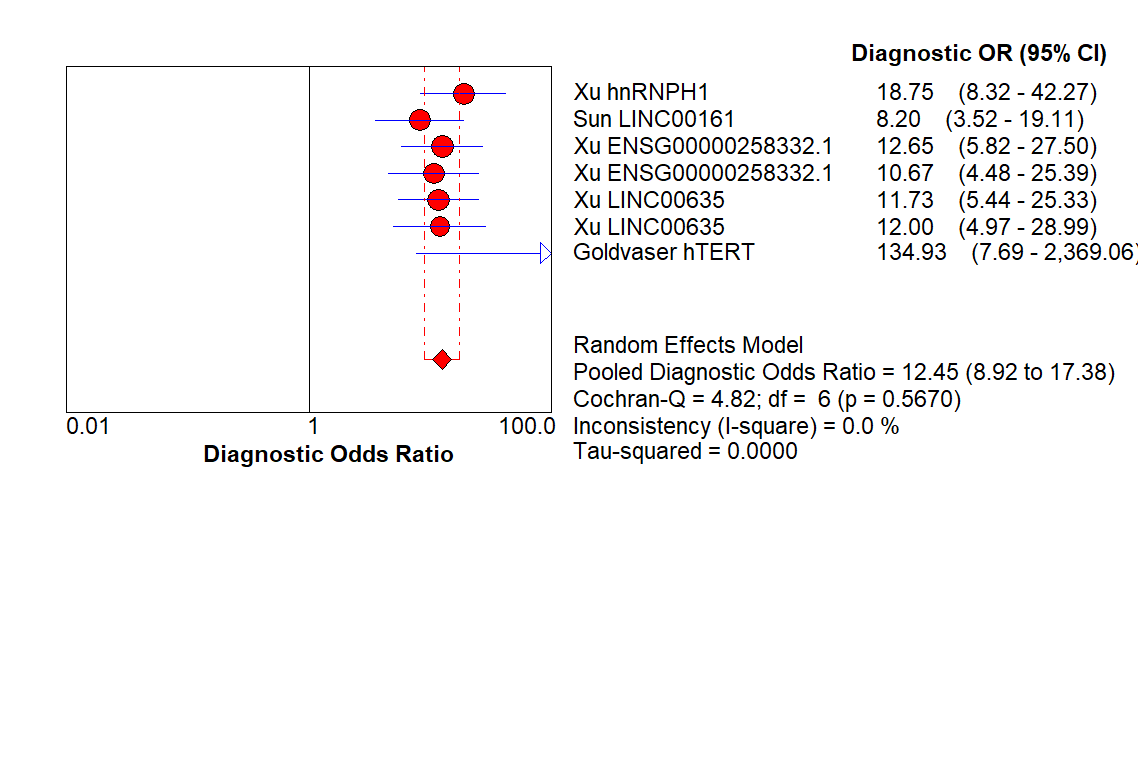
C



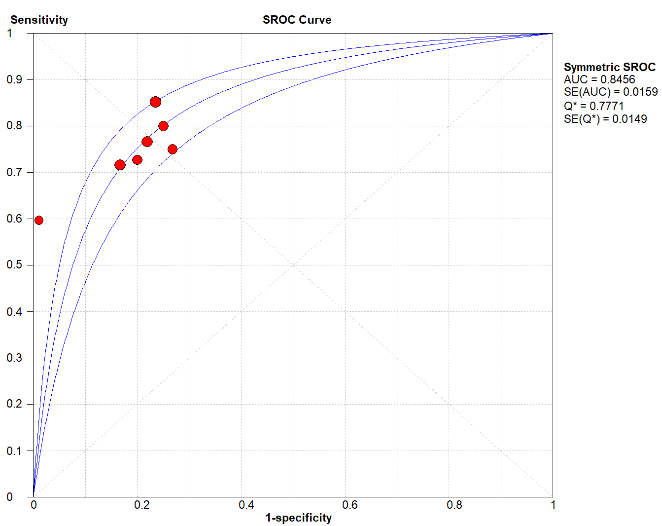
D



E

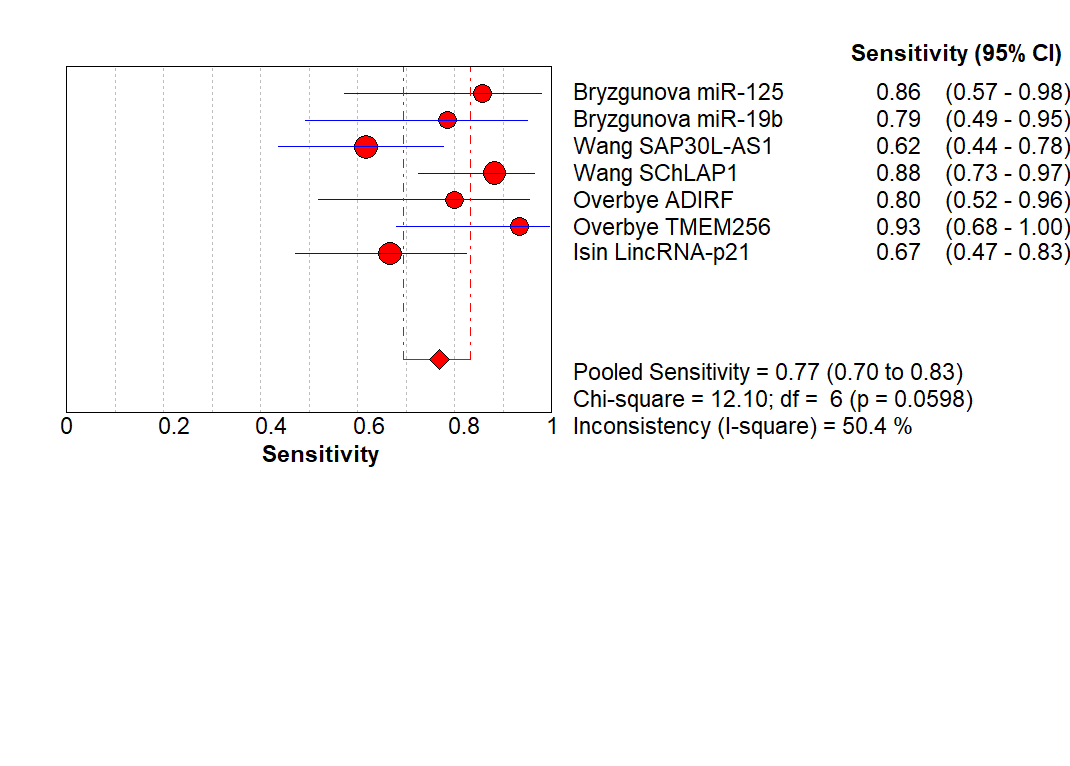


F

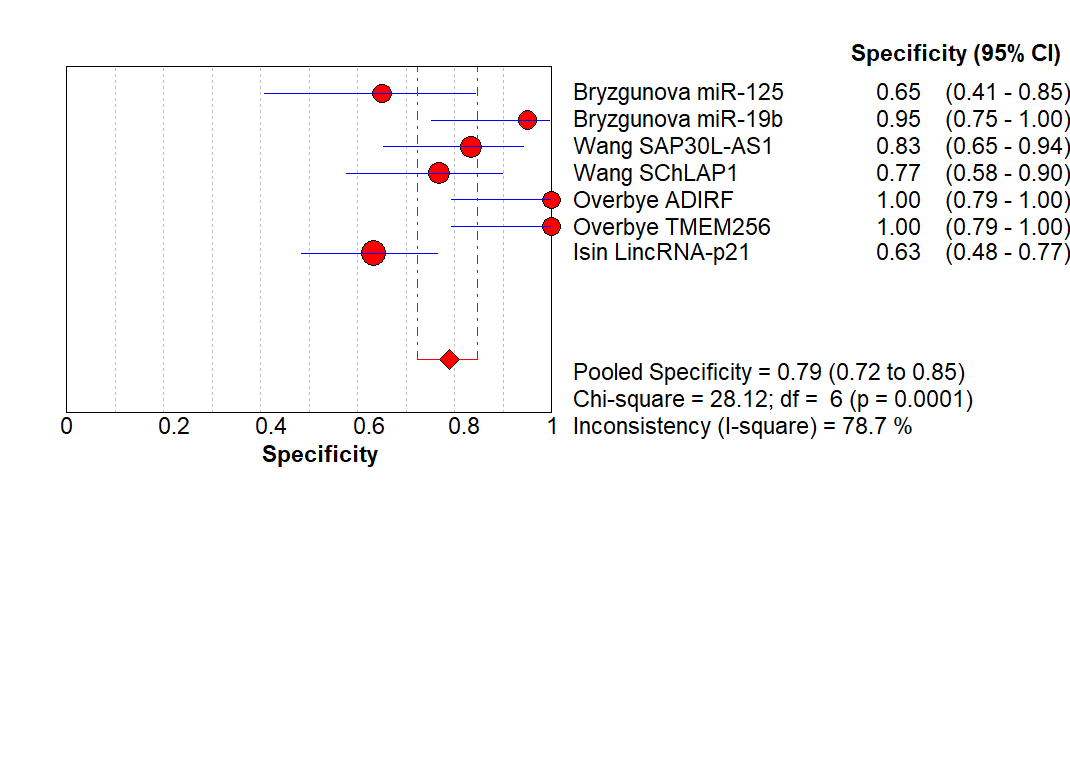


**Figure 6 Forest plot of pooled (A) sensitivity, (B) specificity, (C) positive likelihood ratio, (D) negative likelihood ratio, (E) diagnostic odds ratio, and (F) SROC curve of exosomal biomarkers in diagnosis of liver cancers.**

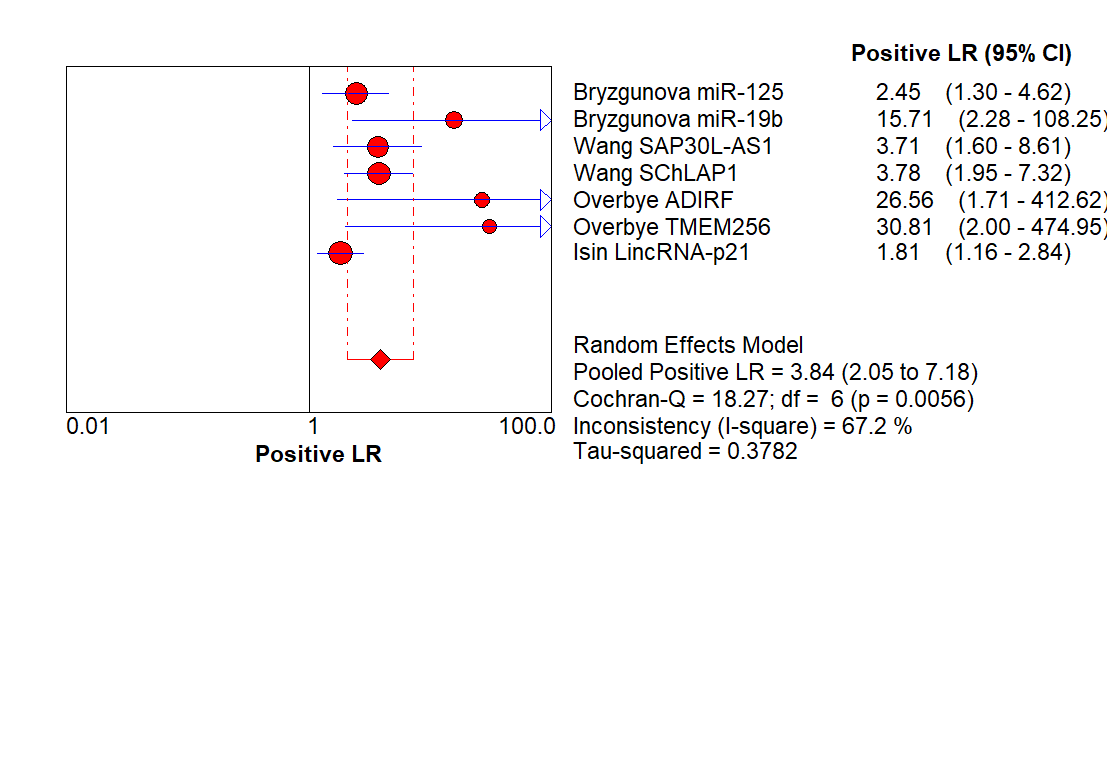
A



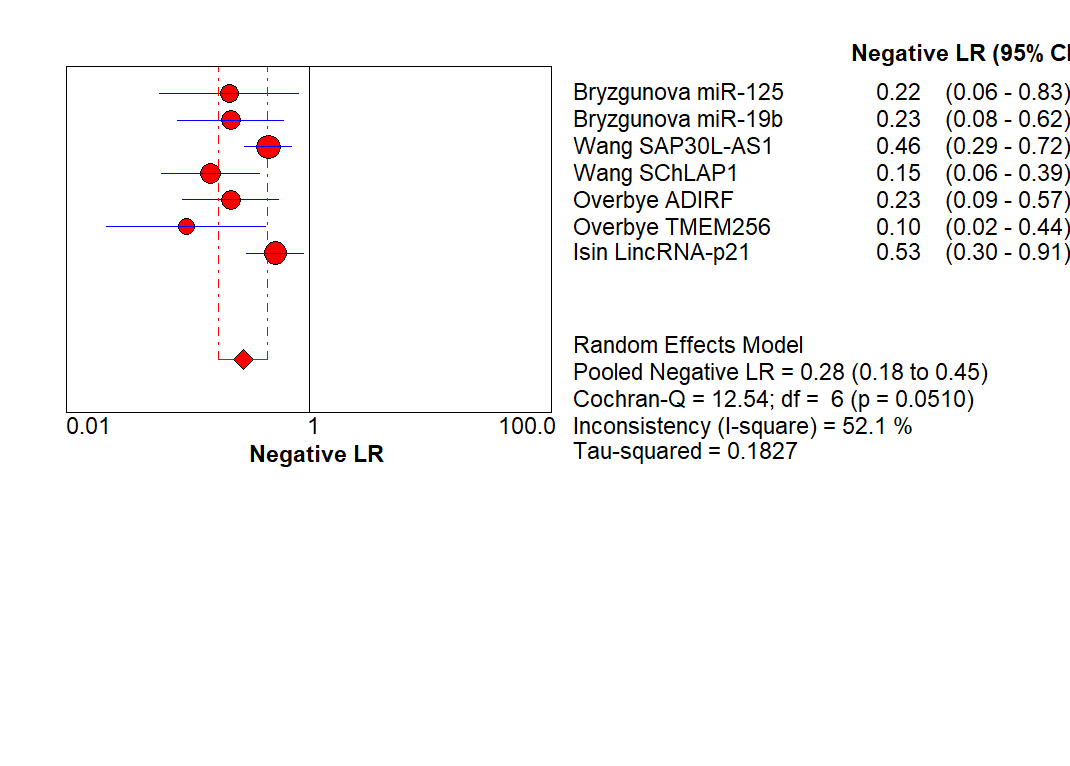
B



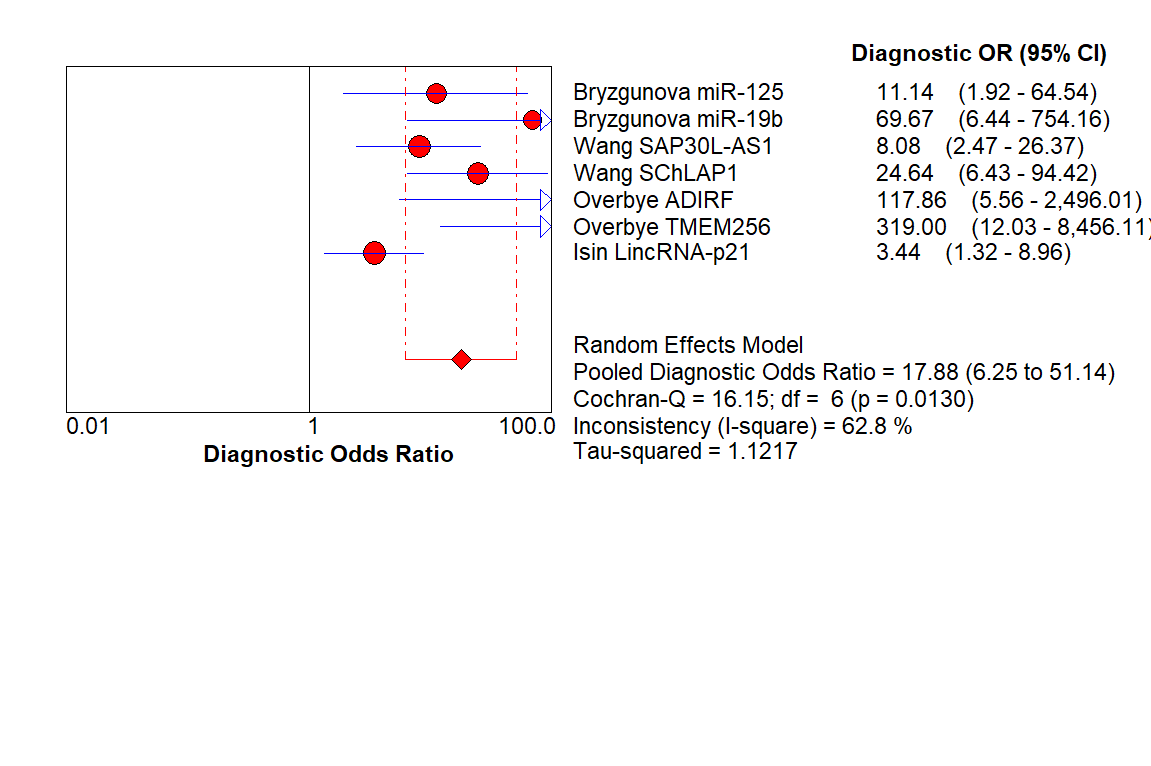
C



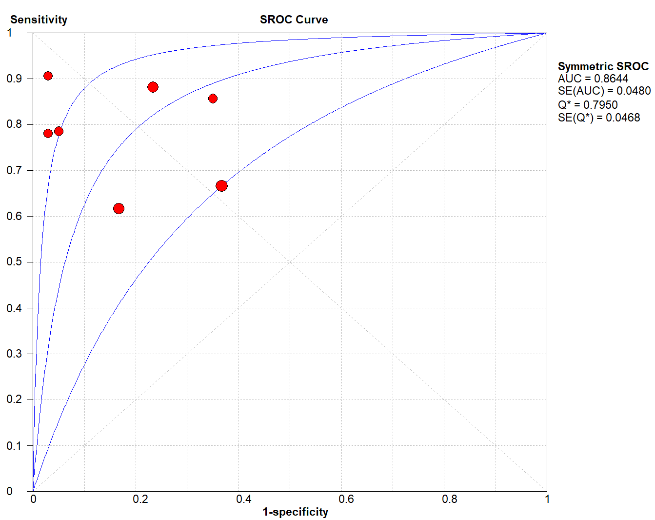
D



E

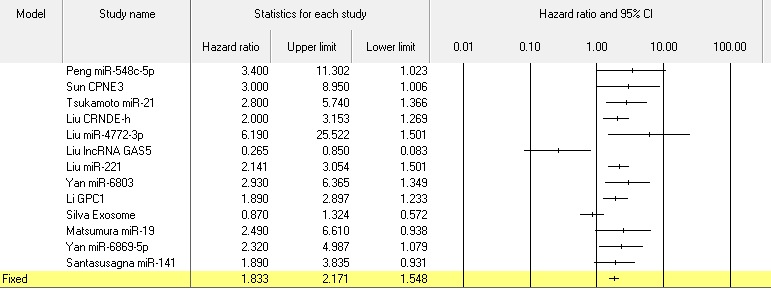


F

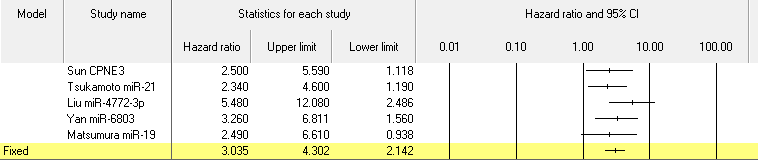


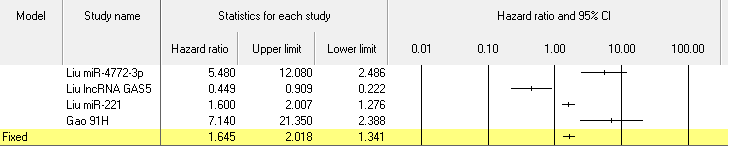
**Figure 7 Forest plot of pooled (A) sensitivity, (B) specificity, (C) positive likelihood ratio, (D) negative likelihood ratio, (E) diagnostic odds ratio, and (F) SROC curve of exosomal biomarkers in diagnosis of prostate cancers.**

A

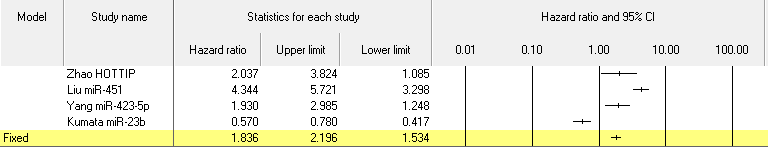


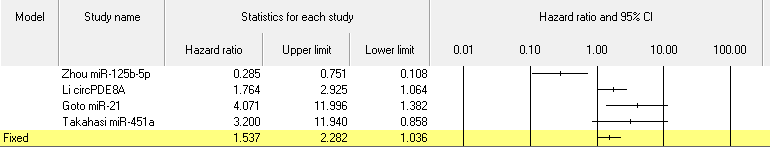
B



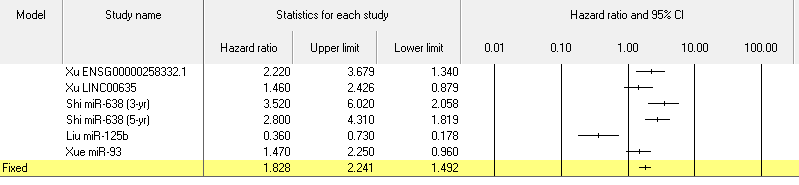
C

**Figure 8 Forest plot elevating the effect of exosomal markers on overall survival (A), disease-free survival (B), and (C) recurrence-free survival of patients with colon cancer.**

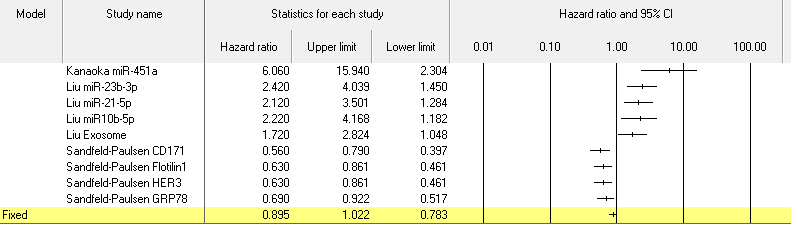
**Figure 9 Forest plot elevating the effect of exosomal markers on overall survival of patients with gastric cancer.**



**Figure 10 Forest plot elevating the effect of exosomal markers on overall survival of patients with pancreatic cancer.**



**Figure 11 Forest plot elevating the effect of exosomal markers on overall survival of patients with liver cancer.**



**Figure 12 Forest plot elevating the effect of exosomal markers on overall survival of patients with lung cancer.**

**Table 1 Studies included for meta-analysis of exosomal biomarkers in cancer diagnosis**

|  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- |
| **First author** | **Country** | **Cancer type** | **Stage** | **Control** | **Number of Control** | **Number of patients** | **Sample** | **Isolation method of exosome** | **Marker** | **Detection method** | **Cut-off** | **TP** | **TN** | **FP** | **FN** |
| Sun[35] | China | Colorectal | All | Healthy | 32 | 92 | Plasma | UC | CPNE3 | ELISA | 0.143pg/ug exosome | 62 | 27 | 5 | 30 |
| Ogata-Kawata[36] | Japan | Colorectal | All | Healthy | 11 | 88 | Serum | UC | miR-1246 | qRT-PCR | 1.45 | 84 | 10 | 4 | 1 |
| miR-23a | 0.3100 | 81 | 11 | 7 | 0 |
| miR-21 | 1.08 | 54 | 10 | 34 | 1 |
| miR-150 | 0.08 | 49 | 11 | 39 | 0 |
| let-7a | 0.9 | 44 | 10 | 44 | 1 |
| miR-223 | 1.72 | 41 | 10 | 47 | 1 |
| miR-1224-5p | 0.5 | 28 | 11 | 60 | 0 |
| miR-1229 | 0.06 | 20 | 11 | 68 | 0 |
| Liu[37] | China | Colorectal | All | Healthy and benign | 320 | 148 | Serum | ExoQuick | CRNDE-h | qRT-PCR | 0.02 | 104 | 302 | 18 | 44 |
| Uratani[38] | Japan | Colorectal | NR | Healthy | 47 | 26 | Serum | ExoQuick | miR-21 | qRT-PCR | Youden index | 18 | 38 | 9 | 8 |
| Lin[39] | China | Gastric | All | Healthy | 60 | 51 | Plasma | UC | lncUEGC1 | qRT-PCR | NR | 45 | 50 | 10 | 6 |
|  |  |  |  | lncUEGC2 | NR | 46 | 34 | 26 | 17 |
| Zhao[40] | China | Gastric | All | Healthy | 120 | 126 | Serum | NR | HOTTIP | qRT-PCR | 1.72 | 88 | 102 | 18 | 38 |
| Pang[41] | China | Gastric | All | Healthy | 37 | 40 | Serum | ExoQuick | ZFAS1 | qRT-PCR | NR | 32 | 28 | 9 | 8 |
| Yang[42] | China | Gastric | All | Healthy | 80 | 80 | Serum | ExoQuick | miR-423-5p | qRT-PCR | NR | 65 | 46 | 34 | 15 |
| Goto[43] | Japan | Pancreatic | All | Healthy and advanced pancreatic cancer | 22 | 23 | Serum | ExoQuick | miR-191 | qRT-PCR | Distance=  (1-sensitivity)2 + (1-specificity)2 in ROC curve | 18 | 17 | 5 | 5 |
| miR-21 | 20 | 18 | 4 | 3 |
| miR-451a | 16 | 18 | 4 | 7 |
| Melo[44] | Germany | Pancreatic | All | Healthy | 100 | 190 | Serum | UC | GPC1 | Flow cytometry | Youden index | 190 | 100 | 0 | 0 |
| Que[45] | China | Pancreatic | All | Non-PDAC | 27 | 22 | Serum | UC | miR-17-5p | qRT-PCR | 6.826 | 20 | 20 | 7 | 2 |
| miR-21 | 7.693 | 18 | 26 | 1 | 4 |
| Machida[46] | Japan | Pancreatobiliary tract | II-IV | Healthy | 13 | 12 | Saliva | Total exosome isolation kit | miR-1246 | qRT-PCR | 13.77 | 8 | 13 | 0 | 4 |
| miR-4644 | -5.205 | 9 | 10 | 3 | 3 |
| Xu[47] | China | Liver | All | Chronic hepatitis B | 68 | 88 | Serum | Total exosome isolation kit | hnRNPH1 | qRT-PCR | 0.67 | 75 | 52 | 16 | 13 |
| Sun[48] | China | Liver | All | Healthy | 56 | 56 | Serum | Total exosome isolation kit | LINC00161 | qRT-PCR | NR | 42 | 41 | 15 | 14 |
| Xu[49] | China | Liver | All | Chronic hepatitis B | 96 | 60 | Serum | Total exosome isolation kit | ENSG00000258332.1 | qRT-PCR | 1.345 | 43 | 80 | 16 | 17 |
| 60 | 55 | ENSG00000258332.1 | 1.366 | 40 | 48 | 12 | 15 |
| 96 | 60 | LINC00635 | 1.69 | 46 | 75 | 21 | 14 |
| 60 | 55 | LINC00635 | 1.532 | 44 | 45 | 15 | 11 |
| Goldvaser[50] | Israel | Pan-cancer  (not include liver) |  | Healthy | 45 | 98 | Serum | Total exosome isolation kit | hTERT | qRT-PCR | NR | 61 | 45 | 0 | 37 |
| Liver | NR | Healthy | 45 | 35 | NR | 21 | 45 | 0 | 14 |
| Zhang[51] | China | Lung | All | Healthy | 30 | 77 | Serum | ExoQuick | MALAT-1 | qRT-PCR | NR | 62 | 21 | 9 | 15 |
| Sun[52] | China | Lung | All | Healthy | 15 | 15 | Plasma | UC | 14-3-3ζ | ELISA |  | 9 | 12 | 3 | 6 |
| Li[53] | NR | Ovarian |  | Benign | 21 | 50 | Serum | UC | ephrinA2 | ELISA | 20.4ng/l | 44 | 17 | 4 | 6 |
| Meng[54] | NR | Ovarian | All | Benign | 20 | 163 | Serum | Total exosome isolation kit | miR-200a | PCR+  qRT-PCR | Youden index | 135 | 18 | 2 | 28 |
| miR-200b | 86 | 20 | 0 | 77 |
| miR-200c | 51 | 20 | 0 | 112 |
| Pan[55] | Germany | Ovarian | All | Healthy | 29 | 106 | Plasma | ExoQuick | miR-21 | PCR+  qRT-PCR | Youden index | 65 | 24 | 5 | 41 |
| miR-100 | 66 | 21 | 8 | 40 |
| miR-200b | 68 | 25 | 4 | 38 |
| miR-320 | 59 | 20 | 9 | 47 |
| Bryzgunova[56] | Russia | Prostate | All | Healthy | 20 | 14 | Urine | UC | miR-125 | qRT-PCR | NR | 12 | 13 | 7 | 2 |
| miR-19b | NR | 11 | 19 | 1 | 3 |
| Wang[57] | China | Prostate | II-IV | Healthy | 30 | 34 | Plasma | Total exosome isolation kit | SAP30L-AS1 | qRT-PCR | NR | 21 | 25 | 5 | 13 |
| SChLAP1 | NR | 30 | 23 | 7 | 4 |
| Øverbye[58] | NR | Prostate | All | Healthy | 15 | 16 | Urine | UC | ADIRF | Mass spectrometry | Youden index | 12 | 16 | 0 | 3 |
| TMEM256 | 14 | 16 | 0 | 1 |
| Işın[59] | NR | Prostate | All | BPH | 49 | 30 | Urine | Urine Exosome RNA Isolation Kit | LincRNA-p21 | qRT-PCR | 0.181 | 20 | 31 | 18 | 10 |
| Wang[60] | China | Laryngeal | All | Vocal cord polyps | 49 | 52 | Serum | ExoQuick | miR-21 | qRT-PCR | 0.043 | 36 | 40 | 9 | 16 |
| HOTAIR |  | 0.032 | 48 | 28 | 21 | 4 |
| Alegre[61] | NR | Melanoma | NR | Healthy | 25 | 53 | Serum | ExoQuick | exo-MIA | ELISA | 1.4ug/l | 42 | 20 | 5 | 11 |
| exo-S100B | ELISA | 0.015ug/l | 42 | 20 | 5 | 11 |
| Manterola[62] | France | GBM | NR | Healthy | 30 | 50 | Serum | ExoQuick | RNU6 | qRT-PCR | 0.372 | 33 | 20 | 10 | 17 |
| Chen[63] | Taiwan | Bladder | All | hernia | 81 | 140 | Urine | UC | TACSTD2 | ELISA | 2.47 ng/ml | 103 | 62 | 19 | 37 |
| Ge[64] | China | Cholangiocarcinoma | All | Biliary obstruction | 56 | 35 | Bile | UC | ENST00000588480.1 | qRT-PCR | NR | 22 | 41 | 15 | 13 |

UC: Ultracentrifugation; NR: Not reported.

**Table 2 Studies included for meta-analysis of exosomal biomarkers in cancer prognosis**

|  |  |  |  |  |  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- |
| **First author** | **Period** | **Country** | **Sample**  **Size** | **Cancer**  **Type** | **Stage** | **Sample** | **Isolation method of exosome** | **Marker** | **Detection method** | **Cut-off value** | **Survival analysis** | **HR (95%CI)** |
| Peng[65] | 2008-2014 | China | 108 | Colorectal | All | Serum | Total Exosome Isolation kit | miR-548c-5p | qRT-PCR | NR | OS | 3.40 (1.02‐11.27) |
| Sun[35] | 2012-2017 | China | 92 | Colorectal | All | Plasma | UC | CPNE3 | ELISA | ≥0.143pg/ug exosome | OS | 3.0 (1.0-8.9) |
| ≥0.143pg/ug exosome | DFS | 2.5 (1.1-5.5) |
| Tsukamoto[66] | 2002-2012 | Japan | 326 | Colorectal | II-IV | Plasma | UC | miR-21 | qRT-PCR | >median | OS | 2.28 (1.81-5.74) |
| DFS | 2.34 (1.87- 4.60) |
| Liu[37] | 2007-2010 | China | 148 | Colorectal | All | Serum | ExoQuick | CRNDE-h | qRT-PCR | >0.02 | OS | 2.000 (1.269-3.154) |
| Liu[67] | 2006-2011 | USA | 84 | Colorectal | II-III | Serum | ExoQuick | miR-4772-3p | qRT-PCR | ≥27.88 | OS | 6.19 (1.50-25.5) |
| ≥27.88 | RFS | 5.48 (2.49-12.1) |
| Liu[24] | 2013-2014 | China | 158 | Colorectal | All | Plasma | UC | lncRNA GAS5 | qRT-PCR | NR | OS | 0.265 (0.082 -0.844) |
|  |  |  | RFS | 0.449 (0.194- 0.909) |
| miR-221 | qRT-PCR | NR | OS | 2.141 (1.368-3.054) |
|  |  |  | RFS | 1.600 (1.162-2.007) |
| Gao[68] | 2011-2014 | China | 108 | Colorectal | All | Serum | ExoQuick | 91H | qRT-PCR | ≥ 0.85 | RFS | 7.14 (1.23-21.35) |
| Yan[69] | NR | NR | 168 | Colorectal | All | Serum | Total Exosome Isolation kit | miR-6803 | qRT-PCR | NR | OS | 2.93 (1.35-6.37) |
| DFS | 3.26 (1.56-6.81) |
| Li[70] | 2013-2015 | China | 85 | Colorectal | III | Plasma | ExoCapTM | GPC1 | Flow cytometry | >mean | OS | 1.89 (1.23-2.89) |
| Silva[71] | 2003-2009 | Spain | 91 | Colorectal | All | Plasma | UC | Exosome | Flow cytometry of EpCAM | High | OS | 0.87 (0.57-1.32) |
| Matsumura[72] | 1992-2007 | Japan | 209 | Colorectal | All | Serum | UC | miR-19 | qRT-PCR | >mean | O | 2.49 (1.12-6.61) |
| DFS | 2.49 (1.12-6.61) |
| Yan[73] | 2012-2015 | China | 142 | Colorectal | All | Serum | Total Exosome Isolation kit | miR-6869-5p | qRT-PCR | <mean | OS | 2.32 (1.08-4.99) |
| Santasusagna[25] | 2009-2013 | Spain | 32 | Colon | I-III | Plasma | UC | miR-141 | qRT-PCR | High | OS | 1.89 (0.93-3.83) |
| Zhao[40] | 2011-2012 | China | 126 | Gastric | All | Serum | NR | HOTTIP | qRT-PCR | >1.72 | OS | 2.037 (1.085-3.823) |
| Liu[74] | 2012-2017 | China | 76 | Gastric | All | Serum | Total Exosome Isolation kit | miR-451 | qRT-PCR | >median | 5yr-OS | 4.344 (2.853‐5.721) |
| Yang[42] | NR | China | 80 | Gastric | All | Serum | ExoQuick | miR-423-5p | qRT-PCR | >median | DFS | 1.93 (1.25-2.99) |
| OS | 1.42 (0.92-2.20) |
| Kumata[75] | 2006-2013 | Japan | 232 | Gastric | All | Plasma | UC | miR‑23b | qRT-PCR | >0.78 | OS | 0.57 (0.37‑0.78) |
| DFS | 0.64 (0.41‑0.91) |
| Zhou[76] | 2010-2014 | China | 152 | Pancreatic | All | Plasma | ExoQuick | miR-125b-5p | qRT-PCR | <median | OS | 0.285 (0.108-0.75) |
| Li[77] | 2012-2016 | China | 87 | Pancreatic | All | Plasma | NR | circPDE8A | qRT-PCR | >median | OS | 1.764 (1.064-2.925) |
| Goto[43] | 2013-2015 | Japan | 32 | Pancreatic | All | Serum | ExoQuick | miR-21 | qRT-PCR | >median | OS | 4.071 (1.832-11.996) |
| Takahasi[78] | 2013-2017 | Japan | 50 | Pancreatic | I-II | Plasma | UC | miR-451a | qRT-PCR | >1.75 | OS | 3.20 (1.07-11.94) |
| DFS | 2.87 (1.23-7.23) |
| Xu[49] | 2012-2016 | China | 60 | Liver | All | Serum | Total Exosome Isolation kit | ENSG00000258332.1 | qRT-PCR | >1.845 | OS | 2.22 (1.34-3.68) |
| LINC00635 | qRT-PCR | >2.100 | OS | 1.46 (0.88-2.43) |
| Shi[79] | 2008-2011 | China | 126 | Liver | All | Serum | Total Exosome Isolation kit | miR-638 | qRT-PCR | NR | 3yr-OS | 3.52 (1.37-6.02) |
| 5yr-OS | 2.80 (1.24-4.31) |
| Liu[26] | 2012 | China | 128 | Liver | All | Serum | ExoQuick | miR-125b | qRT-PCR | <median | RFS | 0.14 (0.07-0.29) |
| OS | 0.36 (0.18-0.74) |
| Xue [80] | 2015-2017 | China | 85 | Liver | All | Serum | Total Exosome Isolation kit | miR-93 | qRT-PCR | NR | OS | 1.47 (0.96-2.25) |
| Liu[81] | 2008-2013 | China | 32 | Hepatoblastoma (children) | All | Serum | ExoQuick | miR-21 | qRT-PCR | NR | EFS | 1.434 (1.257-2.766) |
| Matsumoto[82] | 2011-2012 | Japan | 66 | Esophageal | All | Plasma | Total Exosome Isolation kit | exosome | AChE activity | <600x108/ml | OS | 2.177 (1.085-3.605) |
| Lu[83] | 2007-2015 | China | 110 | Nasopharyngeal | All | Plasma | UC | miR-9 | qRT-PCR | NR | OS | 1.5 (1.03-2.18) |
| Ye[84] | 2011-2013 | China | 83 | Nasopharyngeal | II-IV | Serum | UC | protein concentration | BCA assay | >11 μg/ml | DFS | 214.22 (139.27-329.49) |
| Huang[85] | NR | NR | 23 | Prostate | All | Plasma | ExoQuick | miR-1290 | qRT-PCR | >mean | OS | 1.79(1.30-2.48) |
| miR-375 | qRT-PCR | >mean | OS | 2.69(1.52-4.77) |
| Tang[86] | NR | NR | 35 | Ovarian | All | Ascitic fluid | UC | E-cadherin | NR | >10 μg/ml | OS | 1.82 (0.53-3.58) |
| Vaksman[87] | 1998-2003 |  | 86 | Ovarian | III-IV | Effusion  supernatant | ExoQuick | miR-21 | qRT-PCR | >median | OS | 1.70 (1.1-2.59) |
| Kanaoka[88] | 2012-2017 | Japan | 285 | Lung | I-III | Plasma | UC | miR-451a | qRT-PCR | >1.45 | OS | 6.06 (2.61-15.94) |
| DFS | 2.55 (1.44-4.65) |
| Liu[89] | 2012-2014 | China | 196 | Lung | All | Plasma | ExoQuick | miR-23b-3p | qRT-PCR | High | OS | 2.42 (1.45-4.04) |
| miR-21-5p | qRT-PCR | OS | 2.12(1.28-3.49) |
| miR-10b-5p | qRT-PCR | OS | 2.22 (1.18-4.16) |
| Liu[90] | 2012-2014 | China | 208 | Lung | All | Plasma | ExoQuick | Exosome | AChE activity |  | OS | 1.72 (1.05-2.83) |
| Sandfeld-Paulsen[91] | 2011-2014 | Denmark | 276 | Lung | All | Plasma | / | CD171 | ELISA | NR | OS | 0.56 (0.41-0.79) |
| Flotilin1 | ELISA | NR | OS | 0.63 (0.46-0.86) |
| HER3 | ELISA | NR | OS | 0.63 (0.46-0.86) |
| GRP78 | ELISA | NR | OS | 0.69 (0.51-0.91) |
| Manier[92] | 2006-2008 | France | 156 | Multiple myeloma | All | Plasma | ExoQuick | let-7b | qRT-PCR | <median | OS | 2.83 (1.07-7.50) |
| let-7b | qRT-PCR | <median | DFS | 1.90 (1.22-2.94) |
| let-7e | qRT-PCR | <median | DFS | 2.01 (1.30-3.11) |
| miR-106a | qRT-PCR | <median | DFS | 2.34 (1.52-3.61) |
| miR-106b | qRT-PCR | <median | DFS | 3.54 (2.21-5.68) |
| miR-155 | qRT-PCR | <median | OS | 2.41 (0.96-6.05) |
| miR-155 | qRT-PCR | <median | DFS | 1.76 (1.15-2.69) |
| miR-16 | qRT-PCR | <median | DFS | 2.21 (1.41-3.47) |
| miR-17 | qRT-PCR | <median | DFS | 2.29 (1.48-3.55) |
| miR-18a | qRT-PCR | <median | DFS | 4.52 (1.57-12.98) |
| miR-18a | qRT-PCR | <median | OS | 2.76 (1.79-4.26) |
| miR-20a | qRT-PCR | <median | DFS | 2.31 (1.52-3.53) |
| Alegre[61] | NR | NR | 53 | Melanoma | NR | Serum | ExoQuick | MIA | ELISA | 2.5 μg/l | OS | 1.28 (0.65-2.51) |
| Lan[93] | 2011-2012 | China | 60 | Glioma | All | Serum | ExoQuick | miR-301a | qRT-PCR | >median | OS | 4.4 (3.1-9.6) |
| Ge[64] | NR | China | 35 | Cholangiocarcinoma | All | Bile | UC | ENST00000588480.1 | qRT-PCR | >median | OS | 2.40 (1.24-4.66) |
| ENST00000517758.1 | qRT-PCR | OS | 1.55 (0.80-3.01) |
| Fujii[94] | 2005-2014 | Japan | 108 | Renal cell | I-III | Serum | Total Exosome Isolation kit | miR-224 | qRT-PCR | >median | OS | 9.1 (1.8-166.1) |

UC: Ultracentrifugation; OS: Overall survival; DFS: Disease-free survival; RFS: Recurrence free survival; EFC: Even-free survival; NR: Not reported.