

World Journal of *Gastroenterology*

World J Gastroenterol 2019 January 21; 25(3): 282-397



Contents

Weekly Volume 25 Number 3 January 21, 2019

EDITORIAL

- 282** Chronic hepatitis B and metabolic risk factors: A call for rigorous longitudinal studies
Seto WK

REVIEW

- 287** Roles of Na⁺/Ca²⁺ exchanger 1 in digestive system physiology and pathophysiology
Liao QS, Du Q, Lou J, Xu JY, Xie R

MINIREVIEWS

- 300** Endoscopic resection techniques for colorectal neoplasia: Current developments
Dumoulin FL, Hildenbrand R
- 308** Elastography-based screening for esophageal varices in patients with advanced chronic liver disease
Paternostro R, Reiberger T, Bucsics T

ORIGINAL ARTICLE

Basic Study

- 330** NKX6.3 protects against gastric mucosal atrophy by downregulating β -amyloid production
Yoon JH, Lee YS, Kim O, Ashktorab H, Smoot DT, Nam SW, Park WS
- 346** Effects of positive acceleration (+Gz stress) on liver enzymes, energy metabolism, and liver histology in rats
Shi B, Wang XQ, Duan WD, Tan GD, Gao HJ, Pan YW, Guo QJ, Zhang HY

Retrospective Study

- 356** Incidence and treatment of mediastinal leakage after esophagectomy: Insights from the multicenter study on mediastinal leaks
Fumagalli U, Baiocchi GL, Celotti A, Parise P, Cossu A, Bonavina L, Bernardi D, de Manzoni G, Weindelmayer J, Verlato G, Santi S, Pallabazzer G, Portolani N, Degiuli M, Reddavid R, de Pascale S
- 367** Predicting gastroesophageal varices through spleen magnetic resonance elastography in pediatric liver fibrosis
Yoon H, Shin HJ, Kim MJ, Han SJ, Koh H, Kim S, Lee MJ

Observational Study

- 378** Differential hepatic features presenting in Wilson disease-associated cirrhosis and hepatitis B-associated cirrhosis
Zhong HJ, Sun HH, Xue LF, McGowan EM, Chen Y

Prospective Study

- 388** Do patients with gastroesophageal reflux disease and somatoform tendencies benefit from antireflux surgery?

Fuchs HF, Babic B, Fuchs KH, Breithaupt W, Varga G, Musial F

ABOUT COVER

Editorial board member of *World Journal of Gastroenterology*, Maria Gazouli, PhD, Associate Professor, Department of Basic Medical Sciences, Laboratory of Biology, Medical School, National and Kapodistrian University of Athens, Athens 11527, Greece

AIMS AND SCOPE

World Journal of Gastroenterology (*World J Gastroenterol*, *WJG*, print ISSN 1007-9327, online ISSN 2219-2840, DOI: 10.3748) is a peer-reviewed open access journal. The *WJG* Editorial Board consists of 642 experts in gastroenterology and hepatology from 59 countries.

The primary task of *WJG* is to rapidly publish high-quality original articles, reviews, and commentaries in the fields of gastroenterology, hepatology, gastrointestinal endoscopy, gastrointestinal surgery, hepatobiliary surgery, gastrointestinal oncology, gastrointestinal radiation oncology, etc. *WJG* is dedicated to become an influential and prestigious journal in gastroenterology and hepatology, to promote the development of above disciplines, and to improve the diagnostic and therapeutic skill and expertise of clinicians.

INDEXING/ABSTRACTING

World Journal of Gastroenterology (*WJG*) is now indexed in Current Contents®/Clinical Medicine, Science Citation Index Expanded (also known as SciSearch®), Journal Citation Reports®, Index Medicus, MEDLINE, PubMed, PubMed Central and Directory of Open Access Journals. The 2018 edition of Journal Citation Report® cites the 2017 impact factor for *WJG* as 3.300 (5-year impact factor: 3.387), ranking *WJG* as 35th among 80 journals in gastroenterology and hepatology (quartile in category Q2).

RESPONSIBLE EDITORS FOR THIS ISSUE

Responsible Electronic Editor: Yan Huang

Proofing Editorial Office Director: Ze-Mao Gong

NAME OF JOURNAL*World Journal of Gastroenterology***ISSN**

ISSN 1007-9327 (print) ISSN 2219-2840 (online)

LAUNCH DATE

October 1, 1995

FREQUENCY

Weekly

EDITORS-IN-CHIEF

Subrata Ghosh, Andrzej S Tarnawski

EDITORIAL BOARD MEMBERS<http://www.wjgnet.com/1007-9327/editorialboard.htm>**EDITORIAL OFFICE**

Ze-Mao Gong, Director

PUBLICATION DATE

January 21, 2019

COPYRIGHT

© 2019 Baishideng Publishing Group Inc

INSTRUCTIONS TO AUTHORS<https://www.wjgnet.com/bpg/gerinfo/204>**GUIDELINES FOR ETHICS DOCUMENTS**<https://www.wjgnet.com/bpg/GerInfo/287>**GUIDELINES FOR NON-NATIVE SPEAKERS OF ENGLISH**<https://www.wjgnet.com/bpg/gerinfo/240>**PUBLICATION MISCONDUCT**<https://www.wjgnet.com/bpg/gerinfo/208>**ARTICLE PROCESSING CHARGE**<https://www.wjgnet.com/bpg/gerinfo/242>**STEPS FOR SUBMITTING MANUSCRIPTS**<https://www.wjgnet.com/bpg/GerInfo/239>**ONLINE SUBMISSION**<https://www.f6publishing.com>



Roles of Na⁺/Ca²⁺ exchanger 1 in digestive system physiology and pathophysiology

Qiu-Shi Liao, Qian Du, Jun Lou, Jing-Yu Xu, Rui Xie

ORCID number: Qiu-Shi Liao (0000-0002-5555-2203); Qian Du (0000-0001-7056-5208); Jun Lou (0000-0001-9133-3879); Jing-Yu Xu (0000-0002-0545-0444); Rui Xie (0000-0003-3643-3388).

Author contributions: Liao QS and Du Q equally contributed to this study, and wrote the manuscript; Liao QS, Du Q, and Lou J participated in information collection, analysis, and organization; Xu JY primarily revised and finalized the manuscript; Xie R revised the manuscript for clarity and style; Xu JY and Xie R are the co-corresponding authors.

Supported by the National Natural Science Foundation of China, No. 816660412 to Xie R and No. 81160265 to Xu JY.

Conflict-of-interest statement: Authors declare no conflict of interests for this article.

Open-Access: This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

Manuscript source: Unsolicited

Qiu-Shi Liao, Qian Du, Jun Lou, Jing-Yu Xu, Rui Xie, Department of Gastroenterology, Affiliated Hospital to Zunyi Medical College, Zunyi 563000, Guizhou Province, China

Corresponding author: Rui Xie, MD, PhD, Professor, Department of Gastroenterology, Affiliated Hospital to Zunyi Medical College, No. 149, Dalian Road, Huichuan District, Zunyi 563000, Guizhou Province, China. xr19841029@aliyun.com

Telephone: +86-15120390646

Fax: +86-851-28609205

Abstract

The Na⁺/Ca²⁺ exchanger (NCX) protein family is a part of the cation/Ca²⁺ exchanger superfamily and participates in the regulation of cellular Ca²⁺ homeostasis. NCX1, the most important subtype in the NCX family, is expressed widely in various organs and tissues in mammals and plays an especially important role in the physiological and pathological processes of nerves and the cardiovascular system. In the past few years, the function of NCX1 in the digestive system has received increasing attention; NCX1 not only participates in the healing process of gastric ulcer and gastric mucosal injury but also mediates the development of digestive cancer, acute pancreatitis, and intestinal absorption. This review aims to explore the roles of NCX1 in digestive system physiology and pathophysiology in order to guide clinical treatments.

Key words: Na⁺/Ca²⁺ exchanger; Digestive system diseases; Ion channel; Sodium; Calcium

©The Author(s) 2019. Published by Baishideng Publishing Group Inc. All rights reserved.

Core tip: The Na⁺/Ca²⁺ exchange 1 protein (NCX1) is a membrane transporter and participates in the regulation of cellular Ca²⁺ homeostasis. As we known, NCX1 is expressed widely in various organs and tissues and plays an especially important role in the physiological and pathological processes of nerves and the cardiovascular system. This review aims to explore the roles of NCX1 in digestive system physiology and pathophysiology in order to guide clinical treatments.

Citation: Liao QS, Du Q, Lou J, Xu JY, Xie R. Roles of Na⁺/Ca²⁺ exchanger 1 in digestive system physiology and pathophysiology. *World J Gastroenterol* 2019; 25(3): 287-299

URL: <https://www.wjgnet.com/1007-9327/full/v25/i3/287.htm>

DOI: <https://dx.doi.org/10.3748/wjg.v25.i3.287>

manuscript

Received: November 21, 2018**Peer-review started:** November 22, 2018**First decision:** December 12, 2018**Revised:** December 20, 2018**Accepted:** December 27, 2018**Article in press:** December 27, 2018**Published online:** January 21, 2019

INTRODUCTION

Ca^{2+} is an important cellular signal. Changes in intracellular Ca^{2+} control various cellular processes that are relevant to the regulation of normal function and to the development of diseases. These processes include muscle contraction, blood coagulation, nerve excitation, angiogenesis, cell apoptosis^[1-3], and the development of cancer^[4,5]. The homeostasis of intracellular calcium is controlled by a variety of proteins and ion channels, including the plasma membrane $\text{Na}^+/\text{Ca}^{2+}$ exchanger (NCX). Members of the NCX family can exchange Na^+ and Ca^{2+} in either direction depending on the transmembrane electrochemical gradients and membrane potential^[6], and these exchangers have a two-way transport mode such that under physiological conditions, one Ca^{2+} ion exits and three Na^+ ions enter the cell but the reverse transport occurs under special conditions (such as cancer or inflammation), that is, three Na^+ ions exit and one Ca^{2+} ion enters^[7]. The NCX family contains three separate gene products exhibiting differential expression: NCX1, NCX2, and NCX3. NCX1 is widely expressed in mammalian organs and tissues^[8], and NCX2 and NCX3 are expressed mainly in nerves and skeletal muscle^[9]. Numerous studies have shown that NCX1 is involved in a variety of physiological and pathophysiological processes. For example, in the cardiovascular system, NCX1 can control the contraction and relaxation of vascular smooth muscle^[10], while NCX1 can regulate heart rhythm, which is related to arrhythmia^[11,12], and participate in the regulation of myocardial ischemia-reperfusion injury^[13]. In the nervous system, NCX1 regulates neurotransmitter release^[14] and microglia-related functions^[15], which is associated with cerebral ischemia-reperfusion and Alzheimer's disease^[16,17]. In the urinary system, NCX1 is involved in renal Ca^{2+} reabsorption and associated with renal ischemia-reperfusion^[18,19]. In the endocrine system, NCX1 can regulate insulin secretion^[20]. In the immune system, NCX1 is associated with the development of systemic lupus erythematosus^[21]. In recent years, NCX1 has been found to be expressed in all of the organs of the digestive system and play important roles in the physiological processes and digestive diseases (such as pancreatitis, gastric ulcer, and gastrointestinal cancers)^[22-24]. However, the mechanism and function of NCX1 in the gastrointestinal tract have not yet been completely elucidated, particularly relating to certain digestive diseases and tumors. This review intends to explore the roles of NCX1 in digestive system physiology and pathophysiology as well as current treatments utilizing NCX1-based therapeutics.

STRUCTURAL FEATURES OF NCX1

NCX1 is a transmembrane bidirectional transporter with a molecular weight of 110 kDa and consists of 970 amino acids. NCX1 has 9 transmembrane segments, forming a large central cytoplasmic loop between the 5th and 6th transmembrane segments^[25,26]. In addition, the NCX1 transmembrane segment has two internal repeat regions, the $\alpha 1$ and $\alpha 2$ repeat regions^[27]. The first half of the transmembrane segment, including the α -repeat region, may be involved in ion transport^[28-30]. In contrast, the second half, which contains the central cytoplasmic ring, has an inhibitory effect on the entire sodium-calcium exchanger^[31,32]. In addition, there are two binding sites that can regulate Na^+ and Ca^{2+} ^[33,34], and there is a secondary Ca^{2+} adapter site^[35] (Figure 1).

NCX1 AND THE ESOPHAGUS

The esophagus is a muscular portion of the digestive tract that transports food from the pharynx to the stomach depending on the contraction of muscle. In normal conditions, the tension of the lower esophageal smooth muscle (LES) depends mainly on the intracellular Ca^{2+} concentration. A high concentration of intracellular Ca^{2+} can cause smooth muscle contraction, and a low concentration of Ca^{2+} causes smooth muscle relaxation^[36]. However, the contraction of the esophageal body is mainly dependent on the gradient of extracellular calcium. The occurrence of esophagitis is also closely related to the regulation of Ca^{2+} in pathological conditions. Calcium channel blockers (CCBs) have been used in the treatment of esophageal-related diseases, such as achalasia^[36,37]. Achalasia is a kind of neuropathy where the smooth muscle fiber is not relaxed or cannot relax completely, a partial loss of esophageal body peristalsis occurs, and the motility is not coordinated. Gelfond *et al*^[38] first reported that use of the L-type CCB nifedipine can relax the esophageal smooth muscle to reduce the pressure of the lower esophageal sphincter by blocking the flow of calcium ions into the cells and intracellular calcium release, thereby achieving the

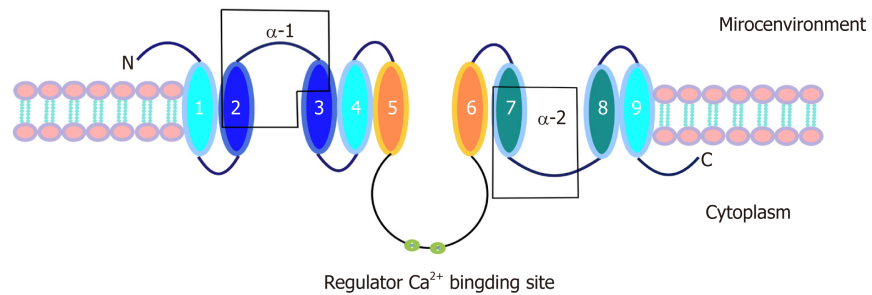


Figure 1 Structural features of NCX1.

purpose of treating achalasia. However, taking CCBs for a long time will cause the LES to become too relaxed and will lead to reflux esophagitis (RE)^[39]. There is no definitive evidence as to whether NCX1 plays a regulatory role in RE. However, Kim *et al.*^[40] found that NCX1 is widely expressed in the esophageal muscle layer. Furthermore, the estrogen E2-induced inhibition of smooth muscle contraction in the esophagus and the mucus secretion in the esophageal mucosa are mainly achieved by downregulating the expression of calcium-related genes such as NCX1, CaBP-9k, and PMCA1 and decreasing the intracellular calcium level. It is suggested that NCX1 may play an important role in the regulation of contraction and relaxation in the LES. In addition, at present, our research group also confirmed that NCX1 expression was significantly increased along with the expression of TRPC6, TRPV4, and other acid-sensitive calcium channels that have a regulatory role in Barrett's esophagus or reflux esophagitis caused by acid reflux or bile reflux. Interfering with NCX1 can significantly inhibit the release of inflammatory mediators and the expression of intestinal metaplasia genes caused by the aforementioned pathogenic factors. Therefore, NCX1 is likely to be an important treatment target for esophageal functional diseases.

NCX1 also plays an important role in the correlation between smoking and the pathogenesis of esophageal squamous cell carcinoma (ESCC)^[41]. Clinical evidence showed that the expression of NCX1 in ESCC tissue was significantly higher than that in esophageal noncancerous tissue and demonstrated a positive correlation between the NCX1 expression level and the smoking status of ESCC patients. The tobacco-derived carcinogen 4-(methylnitrosamino)-1-(3-pyridyl)-1-butanone (NNK) can significantly enhance NCX1 expression in normal esophageal cells and human ESCC cells. NNK mediates an increase in the intracellular Ca^{2+} concentration through NCX1 activation and promotes the proliferation and migration of human ESCC cells^[41]. These findings indicate that tobacco smoking could cause Ca^{2+} entry through enhanced expression and function of NCX1, finally resulting in the pathogenesis of ESCC. Furthermore, NCX1 is also involved in the proliferation and migration of ESCC cells. To elucidate the mechanism of NCX1 in ESCC, it is necessary to study the function of NCX1 in ESCC in the future.

EXPRESSION AND FUNCTION OF NCX1 IN THE STOMACH

The stomach, the main digestive organ of the human body, is connected to the esophagus and the duodenum. It is known that the plasma membrane Ca^{2+} -ATPase (PMCA), NCX, and the endoplasmic reticulum (ER) Ca^{2+} -ATPase are the main mechanisms for the transport of intracellular Ca^{2+} to the extracellular space of gastric smooth muscle cells^[42,43]. Studies have shown that NCX1 is widely expressed in the antrum of guinea pigs, while NCX2 has higher expression in the fundus^[44]. Researchers believe that different NCX subtypes, which have different physiological functions, are expressed in different parts of the stomach and regulate each other. NCX1 is mainly involved in gastric antral motility, while NCX2 mainly changes the intracellular Ca^{2+} homeostasis in fundus smooth muscle cells to control the contraction and relaxation of the fundus smooth muscle^[44]. This information suggests that the NCX family may control the movement of the whole stomach by controlling the movement of the gastric antrum and fundus smooth muscle.

In a study of gastrointestinal motility diseases, Hagi *et al.*^[45] found that NO and PACAP act as important mediators of the transient and sustained relaxation in the mouse gastric fundus. A change in functional coupling and/or collaborative functions between NO signaling and PACAP signaling may cause intracellular Ca^{2+} concentration changes, thereby controlling gastric fundus relaxation in mice. The

overexpression of NCX1 in smooth muscle may result in increased functional coupling and/or collaborative functions between NO signaling and PACAP signaling, resulting in the occurrence of functional gastrointestinal disorders^[46]. Interestingly, studies on experimental gastric ulcer (GU) indicate that nitric oxide synthase (NOS) activity may be an important marker of neutrophil infiltration^[47,48]. NO also contributes to ethanol-induced gastric ulceration and inflammatory bowel diseases (IBD) due to its role in the stimulation of cell proliferation in the gastric mucosa^[49-51]. However, the overexpression of NCX1 in gastrointestinal smooth muscle may affect the function of the NO signaling pathway, and whether this change will promote GU and IBD process through the NO signaling pathway needs further study. It has also been reported that electric field stimulation (EFS) can induce sustained relaxation of the stomach fundus, but not other intestinal regions^[52-54]. The study suggested that this sustained status may be closely related to the regulatory functions of NCX1 and NCX2. In the past few years, studies have shown that NCX1 and NCX2 are expressed in smooth muscles and neurons to regulate the relaxation and motility of the gastric fundus. Upon NCX1 or NCX2 heterozygosity deletion, the fundus relaxation and the gastric peristalsis can be enhanced by EFS^[55]. Therefore, the NCX family may be an important treatment target for functional gastrointestinal diseases. In addition, Lajos V also found that NCX family members (NCX1, NCX2, and NCX3) are expressed extensively in human gastric myofibroblasts and participate in the regulation of intracellular calcium oscillations. Knockdown of NCX1 significantly inhibited the migration and proliferation of gastric myofibroblasts induced by insulin-like growth factor II (IGF-II)^[22]. Gastric myofibroblasts are a kind of contractile, nonexcitatory cell induced by inflammatory factors such as transforming growth factor (TGF- β), and these cells are localized to the subepithelium throughout the whole gastrointestinal tract^[56,57]. It is known that gastric myofibroblasts can not only regulate the secretion of extracellular matrix proteins and the formation of new blood vessels, promoting the healing process of ulcers^[58,59], but also participate in the development of chronic gastritis and gastric cancer cell invasion and metastasis^[60,61]. Further study on the role of NCX in gastrointestinal smooth muscle may elucidate the regulatory mechanisms of ulcer healing and tumor invasion and metastasis.

NCX1 AND THE INTESTINE

NCX1 and the small intestine

The expression of NCX1 protein has been detected in the small intestine, colon, and rectum^[62,63], and it participates in the physiological regulation mechanism of intestinal calcium absorption, bicarbonate secretion, ileal smooth muscle movement and so on. The small intestine absorbs 90% of calcium^[3,64,65]. Additionally, NCX1 mainly participates in the process of extracellular discharge of calcium ions from the basal membrane of intestinal epithelial cells. NCX1 can transport three Na⁺ ions into the cell and transport one Ca²⁺ ion out of the cell; this transport is regulated by vitamin D and 1,25-(OH)₂D₃^[66], which can enhance the expression and activity of NCX on the basal membrane and promote the transport of Ca²⁺ from the cell to the outside^[66]. Moreover, Wongdee *et al* found that vitamin D can upregulate the expression of the Ca²⁺ transporter gene NCX1, thus enhancing the Ca²⁺ transmembrane transport^[67].

It is well known that the gastric acid defense barrier involves bicarbonate, and the hyposecretion of bicarbonate is one of the key mechanisms for the pathogenesis of duodenal ulcers. It has been reported that intracellular calcium signals can promote HCO₃⁻ secretion depending on the activation of the HCO₃⁻ secreting channel cystic fibrosis transmembrane conductance regulator (CFTR) or on the activation of the intermediate-conductance Ca²⁺ activated K⁺ channel (IKCa²⁺), which provide a driving force for HCO₃⁻ secretion^[68]. However, NCX1 may be the key to the regulation of intracellular calcium changes. Dong *et al* found that the NCX1 protein is functionally expressed in the mouse duodenal mucosal epithelium, and a dynamic calcium ion experiment determined that the reverse regulation mode of NCX1 occurs, causing Na⁺ efflux and Ca²⁺ entry to regulate HCO₃⁻ secretion^[69]. Subsequent research confirmed that the muscarinic receptor agonists carbachol and 5-hydroxytryptamine (5-HT) can increase the intracellular calcium concentration and promote duodenal bicarbonate secretion after stimulating mouse duodenal mucosal or epithelial cells and confirmed that disturbing or inhibiting the function of NCX1 can obviously block the intracellular calcium change and promote bicarbonate secretion^[70]. These results suggest that NCX1 and its mediated Ca²⁺ influx play a critical and extensive role in regulating the secretion of HCO₃⁻ in the duodenal mucosa. Other studies have shown that NCX1 and NCX2 are also involved in ileal smooth muscle contraction and ileal motility regulation^[71]. Nishiyama *et al* found that NCX2 regulates ileal motility

primarily by controlling the sensitivity of acetylcholine (AChE) and substance P (SP) in smooth muscle^[71]. Compared with that in the wild-type model, the contraction amplitude induced by AChE and SP after NCX2 knockout was significantly reduced; although NCX1 also plays a role in the regulation of ileal contraction, this decline was not evident in the NCX1 knockout model^[71]. This finding suggests that NCX2 plays a more important role than NCX1 in ileal movement.

NCX1 and the colon

NCXs (NCX1 and NCX2) are also widely expressed in colonic smooth muscle and the myenteric plexus layers^[72]. Nishiyama *et al* showed that NCX1 overexpression in the mouse distal colon enhanced the relaxation amplitude induced by EFS, suggesting that NCX1 can affect the distal colonic smooth muscle movement in mice^[71]. Furthermore, it was found that the secretion of the mucin MUC5AC induced by ATP depends on the influx of Ca^{2+} into colonic goblet cells and that ATP requires the activation of TRPM5 channels to increase intracellular Na^+ , which activates the NCX reverse transport mode and increases intracellular Ca^{2+} uptake; thus, inhibiting NCX can significantly reduce the MUC5AC secretion in goblet cells^[73]. There is also evidence that the NCX family may also be involved in the pathogenesis of diarrhea. Although NCX1 and NCX2 were found to be expressed in the myenteric nerve plexus of the proximal colon and the colon transversum as well as longitudinal and annular muscular layers, the function of NCX1 and NCX2 in intermuscular neurons may be different from that in smooth muscle^[74]. Kazuhiro *et al* have found that in a diarrhea model induced with magnesium sulfate or 5-HT, the diarrhea in NCX2 heterozygous knockout mice (NCX2 HET) was more serious than that in wild-type mice (WT), but the diarrhea in NCX1 heterozygous knockout mice (NCX1 HET) showed no significant changes from that of WT^[75]. Magnesium sulfate-induced diarrhea was exacerbated in NCX2 HET by decreasing normal and soft fecal materials and increasing watery fecal materials, however, PGE2-induced diarrhea in NCX1 HET and NCX2 HET was similar to that in the WT^[75]. The researchers believe that this finding may be due to the mechanism of 5-HT-induced diarrhea involving stimulation of the 5-HT₃ receptor in myenteric plexus neurons and its downstream cholinergic and tachykinin excitatory pathways^[76,77]. However, PGE2 acts directly on smooth muscle and stimulates fluid accumulation to induce diarrhea^[78,79]. Therefore, NCX2 rather than NCX1 in the myenteric plexus may play a critical role in the occurrence and development of diarrhea. Further study of NCX may provide new targets for the diarrhea caused by gastrointestinal dysfunction.

NCX1 AND THE PANCREAS

Expression and distribution of NCX in the pancreas

NCX1 is functionally expressed in the β -cells, acinar cells, and ductal cells of the rat pancreas and has a distinct pattern of distribution in pancreatic ducts depending on their size and proximity to acini^[80]. Two splicing variants of NCX1, NCX1.3 and NCX1.7, are mainly expressed in rat pancreatic cells^[81-84]; however, three other variants, NCX1.2, NCX1.9, and NCX1.13, were also found in guinea pigs, hamsters and mice. In the past few years, studies have proved that different types of NCX1 have different expression levels between different species.

Role of NCX1 in pancreatic physiological processes

The physiological functions of the pancreas include secreting various digestive enzymes and insulin. Normally, the intracellular ATP/ADP ratio increases after pancreatic β -cells uptake glucose, *via* the closure of K^+ -ATP channels, causing β -cell depolarization and inducing extracellular calcium influx through calcium channels in the membrane; the intracellular calcium increase causes fusion of the vesicular membrane containing insulin with the cytoplasmic membrane and the subsequent secretion of insulin from cells *via* vesicular exocytosis^[85]. It has been found that both the voltage-dependent calcium channel (CaV) and the intracellular IP₃-sensitive calcium pool are important in insulin secretion regulation in the past few years^[86], but the role of the NCX family in this process is just beginning to be evaluated.

In normal pancreatic islet β -cells, NCX1 is mainly responsible for Ca^{2+} efflux from cells. The aim is to control the Ca^{2+} concentration within the normal physiological range in order to accurately control the insulin release level^[83,87]. In native pancreatic ducts, the NCX1 expression level is downregulated by acetylcholine and secretin but upregulated by insulin^[80]; as the main physiological stimulant of insulin release, glucose has the reverse regulatory effect on the transcription, expression, and activity of NCX^[88].

NCX1 and pancreatic diseases

Pathologically, NCX also has a regulatory mechanism affecting the insulin secretion from the β -cells of diabetic patients. In the past few years, research has shown that NCX overexpression can lead to ER stress and Ca^{2+} release from the ER, thus promoting β -cell apoptosis, reducing β -cell proliferation, and decreasing insulin secretion^[88]. Herchuelz *et al*^[89] found that heterozygous inactivation of NCX1 (Ncx1+/-) leads to an increase in β -cell function and a 5-fold increase in both β -cell mass and proliferation. The mutation also increases the β -cell resistance to hypoxia, and Ncx1+/- islets show a 2-4 times higher rate of curing diabetes than Ncx1+/+ islets when transplanted into diabetic animals. However, in some cases, NCX may change into the reverse regulation mode to promote Ca^{2+} entry, prolonging the duration of the peak electrical activity associated with glucose and increasing insulin release^[90]. In summary, the different NCX1 expression and transport modes can regulate insulin secretion, so selective inhibition of NCX1 may improve insulin secretion, which provides more theoretical evidence for novel glucose-sensitive insulinotropic drugs for type 2 diabetes that target NCX1.

In addition to regulating physiological insulin secretion, the Ca^{2+} homeostasis is also a key factor leading to pancreatitis, hypercalcemia, pancreatic cancer, and other diseases. Pancreatitis is one of the most common acute abdomen problems, and the pathogenesis is the abnormal accumulation of intracellular Ca^{2+} (calcium overload) to promote excessive activation of trypsinogen, resulting in pancreatic autodigestive injury^[91]. Previous studies have reported that the calcium overload in acute pancreatitis may be related to calcium channels such as CRAC/TRPV1/TRPV3^[92,93]; however, it has recently been found that the NCX1 reverse regulation mode may also be involved in this overload. Yu *et al* confirmed that the mRNA and protein expression of NCX1 in tissues of acute pancreatitis induced by cerulein was significantly increased in cell experiments and animal experiments^[94]. The authors found that the expression of inflammatory mediators such as TNF- α and interleukin-6 (IL-6) caused by cerulein was decreased significantly after treatment with KB-R7943 (a specific inhibitor of NCX1)^[94]. This finding suggests that NCX1 may play a critical role in the occurrence and development of acute pancreatitis. In addition, pancreatic cancer is a kind of cancer with high malignancy and poor prognosis, and duct cell carcinoma is the main pathological type of pancreatic cancer^[95,96]. However, it is generally accepted that alterations in TGF- β signaling and its downstream SMAD pathway play an important role in pancreatic cancer development^[97]. The study by Chow *et al* found that TRPC1 and NCX1 are expressed and functional in pancreatic cancer cells. TGF- β activates TRPC1 and NCX1 channels to mediate a cytoplasmic Ca^{2+} concentration increase in pancreatic cancer cells, which activates the downstream PKC/SMAD4 pathway to regulate pancreatic cancer cell motility^[98]. These results suggest that NCX1 may be involved in the malignant biological behavior regulation of pancreatic cancer (Figure 2).

NCX1 AND THE LIVER

NCX1 and hepatic ischemia-reperfusion

Although NCX1 is expressed in normal livers, liver fibrosis, and liver cancer, the transcription levels and regulation modes are different under these three conditions, suggesting that NCX may have different functions and effects in the development of hepatitis, liver fibrosis, and liver cancer^[99]. In the study of liver ischemia-reperfusion injury, intracellular calcium accumulation is a critical mechanism of cell apoptosis and injury^[100]. NCX mainly adopts the forward control mode in ischemic-reperfusion injury. Trisulfated disaccharide (TD) can transport excess intracellular calcium out of the cell by activating NCX1, thus reducing the serum levels of inflammation markers (TNF- α , IL-6, and IL-10) and the lipid peroxidation after liver injury^[101].

NCX1 and liver fibrosis

Hepatic fibrosis is a necessary process in the progression from chronic hepatitis to cirrhosis. The activation and proliferation of hepatic stellate cells (HSCs) are the central link of hepatic fibrosis. Nakamura *et al* found that the NCX mRNA and protein expression levels were significantly upregulated in response to the activation of rat HSCs induced by CCl₄^[102]. It was also reported that NCX expression is upregulated in cirrhotic tissue, although the specific mechanism is not clear; NCX may be a new target in liver fibrosis or cirrhosis research^[93].

NCX1 and liver cancer

Finally, in hepatocellular carcinoma (HCC) research, our research group has

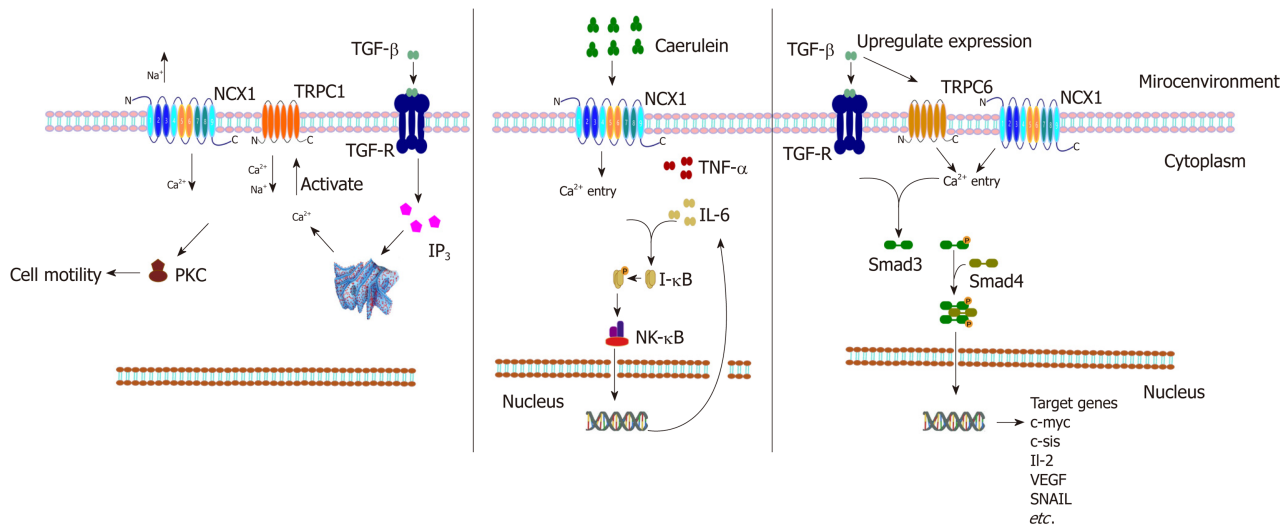


Figure 2 The regulated signal pathways and transcription factors of NCX1 in digestive diseases. Transforming growth factor- β (TGF- β) stimulates the activation of PLC-IP3 and Ca^{2+} release from the endoplasmic reticulum, which activates TRPC1 and the reverse mode of NCX1 resulting in Ca^{2+} influx, and the increase of Ca^{2+} mediates cell motility directly or indirectly via activation of Ca^{2+} -dependent PKC in pancreatic cancer. Cerulein activates NCX1 and induces activation of inflammatory factors TNF- α and IL-6 and the downstream NF- κ B pathway in pancreatic cells. TGF- β can upregulate the expression of NCX1 and TRPC6 and activate the downstream SMAD pathway to regulate the migration and invasion of hepatocellular carcinoma cells.

published articles confirming that the expression of NCX1 is obviously upregulated in hepatoma cells and tissues and that NCX1 can affect the intracellular calcium level to affect the cytokines TGF- β or IL-6 in the malignant behavior of hepatoma cells. A study found that TGF- β can upregulate the expression of NCX1 and the transient receptor potential channel TRPC6 in hepatoma cells and further induce intracellular calcium activation in HCC to promote the formation of a complex between NCX1 and TRPC6, thus activating the downstream SMAD pathway, which can regulate HCC malignant biological behaviors such as migration and invasion^[103]. Not only was the phosphorylation of Smad proteins dependent on TRPC6 and NCX1, but also the Smad signaling, especially the phosphorylation of Smad2, augmented the expression of TRPC6 and NCX1. At the same time the upregulated expression of TRPC6 and NCX1 can be strongly correlated with the stage and pathologic grade of HCC, which may become useful biomarkers for monitoring disease progression of liver cancer patients^[103,104]. In a related study of IL-6 and liver cancer, it was also confirmed that an intracellular pH regulator (NHE1), NCX1, and calmodulin (CaM) coexisted in the same lipid-crossing structure of the cell membrane and that their expression levels were upregulated in liver cancer tissues^[105] (Figure 2). Moreover, IL-6 activated NHE1 to promote H^+ excretion and an NCX1-induced external Ca^{2+} influx, and NHE1 pumped H^+ in exchange for Na^+ influx to promote NCX1 activation, which enhanced the interaction between NCX1 and CaM, thus promoting the occurrence and development of liver cancer^[106]. The above findings provide the basis for the important role of NCX in hepatic carcinogenesis and also provide a new possibility for drug development for early intervention in inflammation-associated tumors.

CONCLUSION

In summary, the NCX1 channel protein regulates the Ca^{2+} signaling pathway *via* its forward/reverse modes in the digestive system and regulates the cell function, thus participating in the occurrence and development of digestive system diseases (Figures 3 and 4). The functions of NCX1 in inflammation-associated digestive diseases (such as inflammatory bowel disease and hepatitis) will become a new research hotspot. NCX1 could be a new molecular marker for the digestive system disease diagnosis and treatment, and drug development targeting NCX1 will represent a new direction of the treatment of digestive system diseases.

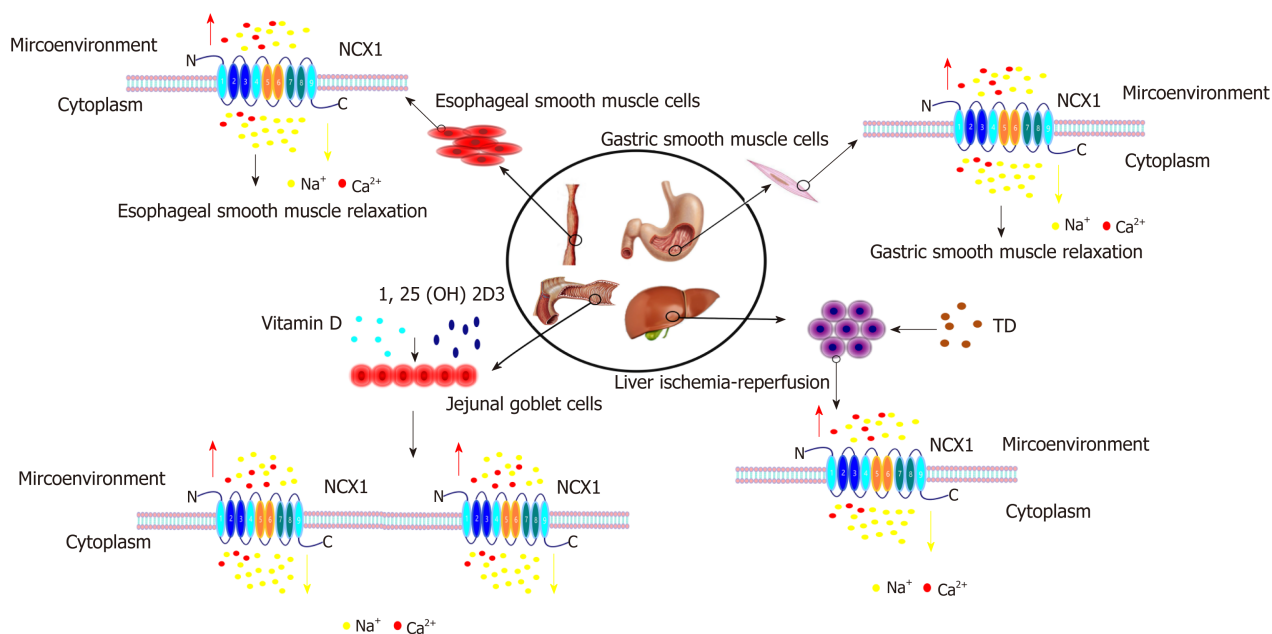


Figure 3 The effects of NCX1 positive mode in the digestive system. Under normal circumstances, NCX1 adopts the positive mode in esophageal smooth muscle and gastric smooth muscle, excreting Ca^{2+} from the cells, reducing intracellular concentration and inducing smooth muscle relaxation. In the jejunum, vitamin D and $1,25\text{-(OH)}_2\text{D}_3$ can enhance the expression and activity of NCX1 to increase the excretion of Ca^{2+} . NCX1 mainly adopts the forward control mode in ischemic-reperfusion injury. Trisulfated disaccharide (TD) can transport excess intracellular Ca^{2+} out of the cell by activating NCX1.

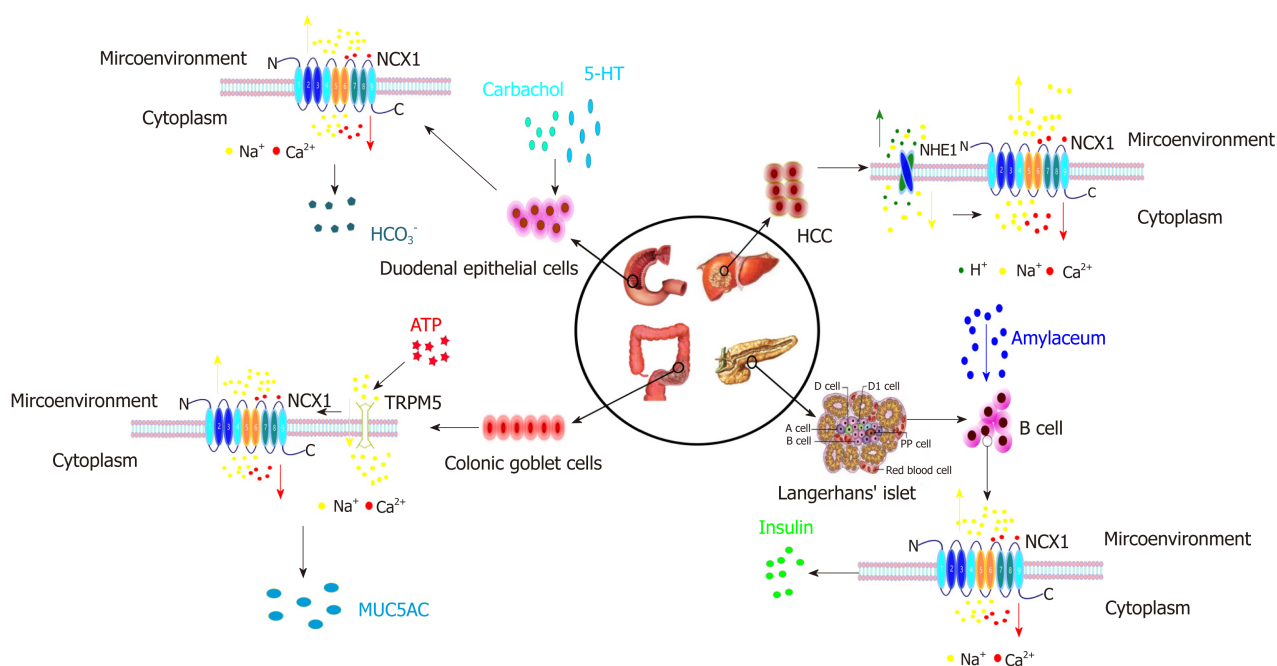


Figure 4 The roles of NCX1 reverse mode in the digestive system. In duodenal epithelial cells, carbachol and 5-HT can activate the reverse mode of NCX1, enhancing Ca^{2+} influx to release HCO_3^- . In hepatoma cells, NHE1 can promote H^+ excretion and Na^+ influx and activate the reverse mode of NCX1 to induce Ca^{2+} influx. In colon goblet cells, ATP activates the TRPM5 channel to induce Na^+ influx, and an increase of Na^+ concentration starts the NCX1 reverse mode and increases Ca^{2+} influx and MUC5AC expression. In pancreatic islet β cells, under glucose stimulation, NCX1 can be converted to a reverse mode to promote Ca^{2+} influx to increase insulin secretion.

ACKNOWLEDGEMENTS

We thank Professor Biguang Tuo (Department of Gastroenterology, Affiliated Hospital to Zunyi Medical College) for highly professional services.

REFERENCES

- 1 **Berridge MJ**, Lipp P, Bootman MD. The versatility and universality of calcium signalling. *Nat Rev Mol Cell Biol* 2000; **1**: 11-21 [PMID: [11413485](#) DOI: [10.1038/35036035](#)]
- 2 **Clapham DE**. Calcium signaling. *Cell* 2007; **131**: 1047-1058 [PMID: [18083096](#) DOI: [10.1016/j.cell.2007.11.028](#)]
- 3 **Hoenderop JG**, Nilius B, Bindels RJ. Calcium absorption across epithelia. *Physiol Rev* 2005; **85**: 373-422 [PMID: [15618484](#) DOI: [10.1152/physrev.00003.2004](#)]
- 4 **Rachow S**, Zorn-Kruppa M, Ohnemus U, Kirschner N, Vidal-y-Sy S, von den Driesch P, Börnchen C, Eberle J, Mildner M, Vettorazzi E, Rosenthal R, Moll I, Brandner JM. Occludin is involved in adhesion, apoptosis, differentiation and Ca²⁺-homeostasis of human keratinocytes: implications for tumorigenesis. *PLoS One* 2013; **8**: e55116 [PMID: [23390516](#) DOI: [10.1371/journal.pone.0055116](#)]
- 5 **Zeng F**, Chen X, Cui W, Wen W, Lu F, Sun X, Ma D, Yuan Y, Li Z, Hou N, Zhao H, Bi X, Zhao J, Zhou J, Zhang Y, Xiao RP, Cai J, Zhang X. RIPK1 Binds MCU to Mediate Induction of Mitochondrial Ca²⁺ Uptake and Promotes Colorectal Oncogenesis. *Cancer Res* 2018; **78**: 2876-2885 [PMID: [29531160](#) DOI: [10.1158/0008-5472.can-17-3082](#)]
- 6 **Khananshvil D**. Sodium-calcium exchangers (NCX): molecular hallmarks underlying the tissue-specific and systemic functions. *Pflugers Arch* 2014; **466**: 43-60 [PMID: [24281864](#) DOI: [10.1007/s00424-013-1405-y](#)]
- 7 **Khananshvil D**. The SLC8 gene family of sodium-calcium exchangers (NCX) - structure, function, and regulation in health and disease. *Mol Aspects Med* 2013; **34**: 220-235 [PMID: [23506867](#) DOI: [10.1016/j.mam.2012.07.003](#)]
- 8 **Liao J**, Li H, Zeng W, Sauer DB, Belmares R, Jiang Y. Structural insight into the ion-exchange mechanism of the sodium/calcium exchanger. *Science* 2012; **335**: 686-690 [PMID: [22323814](#) DOI: [10.1126/science.1215759](#)]
- 9 **Papa M**, Canitano A, Boscia F, Castaldo P, Sellitti S, Porzig H, Tagliatela M, Annunziato L. Differential expression of the Na⁺-Ca²⁺ exchanger transcripts and proteins in rat brain regions. *J Comp Neurol* 2003; **461**: 31-48 [PMID: [12722103](#) DOI: [10.1002/cne.10665](#)]
- 10 **Blaustein MP**, Leenen FH, Chen L, Golovina VA, Hamlyn JM, Pallone TL, Van Huysse JW, Zhang J, Wier WG. How NaCl raises blood pressure: a new paradigm for the pathogenesis of salt-dependent hypertension. *Am J Physiol Heart Circ Physiol* 2012; **302**: H1031-H1049 [PMID: [22058154](#) DOI: [10.1152/ajpheart.00899.2011](#)]
- 11 **Antoons G**, Willems R, Sipido KR. Alternative strategies in arrhythmia therapy: evaluation of Na/Ca exchange as an anti-arrhythmic target. *Pharmacol Ther* 2012; **134**: 26-42 [PMID: [22197992](#) DOI: [10.1016/j.pharmthera.2011.12.001](#)]
- 12 **Herrmann S**, Lipp P, Wiesen K, Stieber J, Nguyen H, Kaiser E, Ludwig A. The cardiac sodium-calcium exchanger NCX1 is a key player in the initiation and maintenance of a stable heart rhythm. *Cardiovasc Res* 2013; **99**: 780-788 [PMID: [23761399](#) DOI: [10.1093/cvr/cvt154](#)]
- 13 **Ohtsuka M**, Takano H, Suzuki M, Zou Y, Akazawa H, Tamagawa M, Wakimoto K, Nakaya H, Komuro I. Role of Na⁺-Ca²⁺ exchanger in myocardial ischemia/reperfusion injury: evaluation using a heterozygous Na⁺-Ca²⁺ exchanger knockout mouse model. *Biochem Biophys Res Commun* 2004; **314**: 849-853 [PMID: [14741714](#)]
- 14 **Roome CJ**, Power EM, Empson RM. Transient reversal of the sodium/calcium exchanger boosts presynaptic calcium and synaptic transmission at a cerebellar synapse. *J Neurophysiol* 2013; **109**: 1669-1680 [PMID: [23255722](#) DOI: [10.1152/jn.00854.2012](#)]
- 15 **Noda M**, Ifuku M, Mori Y, Verkhratsky A. Calcium influx through reversed NCX controls migration of microglia. *Adv Exp Med Biol* 2013; **961**: 289-294 [PMID: [23224888](#) DOI: [10.1007/978-1-4614-4756-6_24](#)]
- 16 **Morimoto N**, Kita S, Shimazawa M, Namimatsu H, Tsuruma K, Hayakawa K, Mishima K, Egashira N, Iyoda T, Horie I, Gotoh Y, Iwasaki K, Fujiwara M, Matsuda T, Baba A, Komuro I, Horie K, Takeda J, Iwamoto T, Hara H. Preferential involvement of Na⁺/Ca²⁺ exchanger type-1 in the brain damage caused by transient focal cerebral ischemia in mice. *Biochem Biophys Res Commun* 2012; **429**: 186-190 [PMID: [23137542](#) DOI: [10.1016/j.bbrc.2012.10.114](#)]
- 17 **Annunziato L**, Pignataro G, Di Renzo GF. Pharmacology of brain Na⁺/Ca²⁺ exchanger: from molecular biology to therapeutic perspectives. *Pharmacol Rev* 2004; **56**: 633-654 [PMID: [15602012](#) DOI: [10.1124/pr.56.4.5](#)]
- 18 **Moor MB**, Haenzi B, Legrand F, Koesters R, Hynes NE, Bonny O. Renal Memo1 Differentially Regulates the Expression of Vitamin D-Dependent Distal Renal Tubular Calcium Transporters. *Front Physiol* 2018; **9**: 874 [PMID: [30038585](#) DOI: [10.3389/fphys.2018.00874](#)]
- 19 **Yamashita J**, Kita S, Iwamoto T, Ogata M, Takaoka M, Tazawa N, Nishikawa M, Wakimoto K, Shigekawa M, Komuro I, Matsumura Y. Attenuation of ischemia/reperfusion-induced renal injury in mice deficient in Na⁺/Ca²⁺ exchanger. *J Pharmacol Exp Ther* 2003; **304**: 284-293 [PMID: [12490603](#) DOI: [10.1124/jpet.102.039024](#)]
- 20 **Hamming KS**, Soliman D, Webster NJ, Searle GJ, Matemisz LC, Liknes DA, Dai XQ, Pulinilkunnit T, Riedel MJ, Dyck JR, Macdonald PE, Light PE. Inhibition of beta-cell sodium-calcium exchange enhances glucose-dependent elevations in cytoplasmic calcium and insulin secretion. *Diabetes* 2010; **59**: 1686-1693 [PMID: [20413506](#) DOI: [10.2337/db09-0630](#)]
- 21 **Vasques ER**, Cunha JEM, Kubrusly MS, Coelho AM, Sanpietri SN, Nader HB, Tersariol ILS, Lima MA, Chaib E, D'Albuquerque LAC. The M-RNA, expression of SERCA2 and NCX1 in the process of pharmacological cell protection in experimental acute pancreatitis induced by taurocholate. *Arq Bras Cir Dig* 2018; **31**: e1352 [PMID: [29947686](#) DOI: [10.1590/0102-672020180001e1352](#)]
- 22 **Kemény LV**, Schnúr A, Czepán M, Rakonczay Z Jr, Gál E, Lonovics J, Lázár G, Simonka Z, Venglovecz V, Maléth J, Judák L, Németh IB, Szabó K, Almássy J, Virág L, Geisz A, Tiszlavicz L, Yule DI, Wittmann T, Varró A, Hegyi P. Na⁺/Ca²⁺ exchangers regulate the migration and proliferation of human gastric myofibroblasts. *Am J Physiol Gastrointest Liver Physiol* 2013; **305**: G552-G563 [PMID: [23907822](#) DOI: [10.1152/ajpgi.00394.2012](#)]
- 23 **Xu J**, Jiang Y, Xie R, Wen G, Dong H, Tuo B. Su1578 Expression and Functional Role of Ncx1 in Human Hepatocellular Carcinoma. *Gastroenterology* 2012; **142**: S-970-S-970
- 24 **Tang B**, Chow JY, Dong TX, Yang SM, Lu DS, Carethers JM, Dong H. Calcium sensing receptor

- suppresses human pancreatic tumorigenesis through a novel NCX1/Ca(2+)/ β -catenin signaling pathway. *Cancer Lett* 2016; **377**: 44-54 [PMID: 27108064 DOI: 10.1016/j.canlet.2016.04.027]
- 25 **Plain F**, Turnbull D, Fraser NJ, Fuller W. Understanding the rules governing NCX1 palmitoylation. *Channels (Austin)* 2017; **11**: 377-379 [PMID: 28617626 DOI: 10.1080/19336950.2017.1342501]
 - 26 **Reilly L**, Howie J, Wypijewski K, Ashford ML, Hilgemann DW, Fuller W. Palmitoylation of the Na/Ca exchanger cytoplasmic loop controls its inactivation and internalization during stress signaling. *FASEB J* 2015; **29**: 4532-4543 [PMID: 26174834 DOI: 10.1096/fj.15-276493]
 - 27 **Nicoll DA**, Ren X, Ottolia M, Phillips M, Paredes AR, Abramson J, Philipson KD. What we know about the structure of NCX1 and how it relates to its function. *Ann N Y Acad Sci* 2007; **1099**: 1-6 [PMID: 17303833 DOI: 10.1196/annals.1387.014]
 - 28 **Nicoll DA**, Hryshko LV, Matsuoka S, Frank JS, Philipson KD. Mutation of amino acid residues in the putative transmembrane segments of the cardiac sarcolemmal Na⁺-Ca²⁺ exchanger. *J Biol Chem* 1996; **271**: 13385-13391 [PMID: 8662775]
 - 29 **Doering AE**, Nicoll DA, Lu Y, Lu L, Weiss JN, Philipson KD. Topology of a functionally important region of the cardiac Na⁺/Ca²⁺ exchanger. *J Biol Chem* 1998; **273**: 778-783 [PMID: 9422731]
 - 30 **Iwamoto T**, Uehara A, Imanaga I, Shigekawa M. The Na⁺/Ca²⁺ exchanger NCX1 has oppositely oriented reentrant loop domains that contain conserved aspartic acids whose mutation alters its apparent Ca²⁺ affinity. *J Biol Chem* 2000; **275**: 38571-38580 [PMID: 10967097 DOI: 10.1074/jbc.M003788200]
 - 31 **Li Z**, Nicoll DA, Collins A, Hilgemann DW, Filoteo AG, Penniston JT, Weiss JN, Tomich JM, Philipson KD. Identification of a peptide inhibitor of the cardiac sarcolemmal Na(+)–Ca²⁺ exchanger. *J Biol Chem* 1991; **266**: 1014-1020 [PMID: 1985930]
 - 32 **Matsuoka S**, Nicoll DA, He Z, Philipson KD. Regulation of cardiac Na(+)–Ca²⁺ exchanger by the endogenous XIP region. *J Gen Physiol* 1997; **109**: 273-286 [PMID: 9041455]
 - 33 **Levitsky DO**, Nicoll DA, Philipson KD. Identification of the high affinity Ca(2+)-binding domain of the cardiac Na(+)–Ca²⁺ exchanger. *J Biol Chem* 1994; **269**: 22847-22852 [PMID: 8077237]
 - 34 **Matsuoka S**, Nicoll DA, Hryshko LV, Levitsky DO, Weiss JN, Philipson KD. Regulation of the cardiac Na(+)–Ca²⁺ exchanger by Ca²⁺. Mutational analysis of the Ca(2+)-binding domain. *J Gen Physiol* 1995; **105**: 403-420 [PMID: 7769381]
 - 35 **Reyes RC**, Verkhatsky A, Parpura V. Plasmalemmal Na⁺/Ca²⁺ exchanger modulates Ca²⁺-dependent exocytotic release of glutamate from rat cortical astrocytes. *ASN Neuro* 2012; **4**: pii: e00075 [PMID: 22268447 DOI: 10.1042/an20110059]
 - 36 **Liu JF**, Lu HL, Wen SW, Wu RF. Effects of acetylcholine on sling and clasp fibers of the human lower esophageal sphincter. *J Gastroenterol Hepatol* 2011; **26**: 1309-1317 [PMID: 21443668 DOI: 10.1111/j.1440-1746.2011.06731.x]
 - 37 **Hoogerwerf WA**, Pasricha PJ. Pharmacologic therapy in treating achalasia. *Gastrointest Endosc Clin N Am* 2001; **11**: 311-324, vii [PMID: 11319064]
 - 38 **Gelfond M**, Rozen P, Keren S, Gilat T. Effect of nitrates on LOS pressure in achalasia: a potential therapeutic aid. *Gut* 1981; **22**: 312-318 [PMID: 7239323]
 - 39 **Kellerman R**, Kintanar T. Gastroesophageal Reflux Disease. *Prim Care* 2017; **44**: 561-573 [PMID: 29132520 DOI: 10.1016/j.pop.2017.07.001]
 - 40 **Kim K**, Lee D, Ahn C, Kang HY, An BS, Seong YH, Jeung EB. Effects of estrogen on esophageal function through regulation of Ca²⁺-related proteins. *J Gastroenterol* 2017; **52**: 929-939 [PMID: 28078471 DOI: 10.1007/s00535-016-1305-y]
 - 41 **Wen J**, Pang Y, Zhou T, Qi X, Zhao M, Xuan B, Meng X, Guo Y, Liu Q, Liang H, Li Y, Dong H, Wang Y. Essential role of Na⁺/Ca²⁺ exchanger 1 in smoking-induced growth and migration of esophageal squamous cell carcinoma. *Oncotarget* 2016; **7**: 63816-63828 [PMID: 27588478 DOI: 10.18632/oncotarget.11695]
 - 42 **Wray S**, Burdya T. Sarcoplasmic reticulum function in smooth muscle. *Physiol Rev* 2010; **90**: 113-178 [PMID: 20086075 DOI: 10.1152/physrev.00018.2008]
 - 43 **Webb RC**. Smooth muscle contraction and relaxation. *Adv Physiol Educ* 2003; **27**: 201-206 [PMID: 14627618 DOI: 10.1152/advan.00025.2003]
 - 44 **Sakai Y**, Kinoshita H, Saitou K, Homma I, Nobe K, Iwamoto T. Functional differences of Na⁺/Ca²⁺ exchanger expression in Ca²⁺ transport system of smooth muscle of guinea pig stomach. *Can J Physiol Pharmacol* 2005; **83**: 791-797 [PMID: 16333381 DOI: 10.1139/y05-079]
 - 45 **Hagi K**, Azuma YT, Nakajima H, Shintani N, Hashimoto H, Baba A, Takeuchi T. Involvements of PHI-nitric oxide and PACAP-BK channel in the sustained relaxation of mouse gastric fundus. *Eur J Pharmacol* 2008; **590**: 80-86 [PMID: 18602629 DOI: 10.1016/j.ejphar.2008.05.045]
 - 46 **Fujimoto Y**, Hayashi S, Azuma YT, Mukai K, Nishiyama K, Kita S, Morioka A, Nakajima H, Iwamoto T, Takeuchi T. Overexpression of Na⁺/Ca²⁺ exchanger 1 display enhanced relaxation in the gastric fundus. *J Pharmacol Sci* 2016; **132**: 181-186 [PMID: 27816547 DOI: 10.1016/j.jphs.2016.10.003]
 - 47 **Coskun T**, Yeğen BC, Alican I, Peker O, Kurtel H. Cold restraint stress-induced gastric mucosal dysfunction. Role of nitric oxide. *Dig Dis Sci* 1996; **41**: 956-963 [PMID: 8625769]
 - 48 **Neurath MF**. Cytokines in inflammatory bowel disease. *Nat Rev Immunol* 2014; **14**: 329-342 [PMID: 24751956 DOI: 10.1038/nri3661]
 - 49 **Li Y**, Wang WP, Wang HY, Cho CH. Intragastric administration of heparin enhances gastric ulcer healing through a nitric oxide-dependent mechanism in rats. *Eur J Pharmacol* 2000; **399**: 205-214
 - 50 **Soufli I**, Toumi R, Rafa H, Touil-Boukoffa C. Overview of cytokines and nitric oxide involvement in immuno-pathogenesis of inflammatory bowel diseases. *World J Gastrointest Pharmacol Ther* 2016; **7**: 353-360 [PMID: 27602236 DOI: 10.4292/wjgpt.v7.i3.353]
 - 51 **Boutemine IM**, Amri M, Amir ZC, Fitting C, Mecherara-Idjeri S, Layaida K, Sennoun N, Berkane S, Cavaillon JM, Touil-Boukoffa C. Gastro-protective, therapeutic and anti-inflammatory activities of Pistacia lentiscus L. fatty oil against ethanol-induced gastric ulcers in rats. *J Ethnopharmacol* 2018; **224**: 273-282 [PMID: 29859303 DOI: 10.1016/j.jep.2018.05.040]
 - 52 **Mukai K**, Takeuchi T, Toyoshima M, Satoh Y, Fujita A, Shintani N, Hashimoto H, Baba A, Hata F. PACAP- and PHI-mediated sustained relaxation in circular muscle of gastric fundus: findings obtained in PACAP knockout mice. *Regul Pept* 2006; **133**: 54-61 [PMID: 16229904 DOI: 10.1016/j.reg.2006.04.001]

- 10.1016/j.regpep.2005.09.019]
- 53 **Mulè F**, Serio R. NANC inhibitory neurotransmission in mouse isolated stomach: involvement of nitric oxide, ATP and vasoactive intestinal polypeptide. *Br J Pharmacol* 2003; **140**: 431-437 [PMID: 12970100 DOI: 10.1038/sj.bjp.0705431]
 - 54 **Baccari MC**, Calamai F. Modulation of nitrergic relaxant responses by peptides in the mouse gastric fundus. *Regul Pept* 2001; **98**: 27-32 [PMID: 11179775]
 - 55 **Azuma YT**, Hayashi S, Nishiyama K, Kita S, Mukai K, Nakajima H, Iwamoto T, Takeuchi T. Na(+) /Ca(2+) exchanger-heterozygote knockout mice display increased relaxation in gastric fundus and accelerated gastric transit in vivo. *Neurogastroenterol Motil* 2016; **28**: 827-836 [PMID: 26787195 DOI: 10.1111/nmo.12779]
 - 56 **Hinz B**, Phan SH, Thannickal VJ, Galli A, Bochaton-Piallat ML, Gabbiani G. The myofibroblast: one function, multiple origins. *Am J Pathol* 2007; **170**: 1807-1816 [PMID: 17525249 DOI: 10.2353/ajpath.2007.070112]
 - 57 **Desmoulière A**, Chaponnier C, Gabbiani G. Tissue repair, contraction, and the myofibroblast. *Wound Repair Regen* 2005; **13**: 7-12 [PMID: 15659031 DOI: 10.1111/j.1067-1927.2005.130102.x]
 - 58 **Nishida T**, Tsuji S, Kimura A, Tsujii M, Ishii S, Yoshio T, Shinzaki S, Egawa S, Irie T, Yasumaru M, Iijima H, Murata H, Kawano S, Hayashi N. Endothelin-1, an ulcer inducer, promotes gastric ulcer healing via mobilizing gastric myofibroblasts and stimulates production of stroma-derived factors. *Am J Physiol Gastrointest Liver Physiol* 2006; **290**: G1041-G1050 [PMID: 16384872 DOI: 10.1152/ajpgi.00462.2005]
 - 59 **Chai J**, Norng M, Tarnawski AS, Chow J. A critical role of serum response factor in myofibroblast differentiation during experimental oesophageal ulcer healing in rats. *Gut* 2007; **56**: 621-630 [PMID: 17068115 DOI: 10.1136/gut.2006.106674]
 - 60 **Guo X**, Oshima H, Kitamura T, Taketo MM, Oshima M. Stromal fibroblasts activated by tumor cells promote angiogenesis in mouse gastric cancer. *J Biol Chem* 2008; **283**: 19864-19871 [PMID: 18495668 DOI: 10.1074/jbc.M800798200]
 - 61 **McCaig C**, Duval C, Hemers E, Steele I, Pritchard DM, Przemeck S, Dimaline R, Ahmed S, Bodger K, Kerrigan DD, Wang TC, Dockray GJ, Varro A. The role of matrix metalloproteinase-7 in redefining the gastric microenvironment in response to *Helicobacter pylori*. *Gastroenterology* 2006; **130**: 1754-1763 [PMID: 16697739 DOI: 10.1053/j.gastro.2006.02.031]
 - 62 **Kim JA**, Yang H, Hwang I, Jung EM, Choi KC, Jeung EB. Expression patterns and potential action of the calcium transport genes Trpv5, Trpv6, Ncx1 and Pmca1b in the canine duodenum, kidney and uterus. *In Vivo* 2011; **25**: 773-780 [PMID: 21753133]
 - 63 **Hwang I**, Jung EM, Yang H, Choi KC, Jeung EB. Tissue-specific expression of the calcium transporter genes TRPV5, TRPV6, NCX1, and PMCA1b in the duodenum, kidney and heart of *Equus caballus*. *J Vet Med Sci* 2011; **73**: 1437-1444 [PMID: 21737966]
 - 64 **Wasserman RH**. Vitamin D and the dual processes of intestinal calcium absorption. *J Nutr* 2004; **134**: 3137-3139 [PMID: 15514288 DOI: 10.1093/jn/134.11.3137]
 - 65 **Bronner F**. Recent developments in intestinal calcium absorption. *Nutr Rev* 2009; **67**: 109-113 [PMID: 19178653 DOI: 10.1111/j.1753-4887.2008.00147.x]
 - 66 **Khuituan P**, Wongdee K, Jantarajit W, Suntornsaratoon P, Krishnamra N, Charoenphandhu N. Fibroblast growth factor-23 negates 1,25(OH)2D3-induced intestinal calcium transport by reducing the transcellular and paracellular calcium fluxes. *Arch Biochem Biophys* 2013; **536**: 46-52 [PMID: 23747333 DOI: 10.1016/j.abb.2013.05.009]
 - 67 **Wongdee K**, Charoenphandhu N. Vitamin D-enhanced duodenal calcium transport. *Vitam Horm* 2015; **98**: 407-440 [PMID: 25817876 DOI: 10.1016/bs.vh.2014.12.010]
 - 68 **Xie R**, Dong X, Wong C, Vallon V, Tang B, Sun J, Yang S, Dong H. Molecular mechanisms of calcium-sensing receptor-mediated calcium signaling in the modulation of epithelial ion transport and bicarbonate secretion. *J Biol Chem* 2014; **289**: 34642-34653 [PMID: 25331955 DOI: 10.1074/jbc.M114.592774]
 - 69 **Dong H**, Sellers ZM, Smith A, Chow JY, Barrett KE. Na(+)/Ca(2+) exchange regulates Ca(2+)-dependent duodenal mucosal ion transport and HCO(3)(-) secretion in mice. *Am J Physiol Gastrointest Liver Physiol* 2005; **288**: G457-G465 [PMID: 15499079 DOI: 10.1152/ajpgi.00381.2004]
 - 70 **Smith AJ**, Chappell AE, Buret AG, Barrett KE, Dong H. 5-Hydroxytryptamine contributes significantly to a reflex pathway by which the duodenal mucosa protects itself from gastric acid injury. *FASEB J* 2006; **20**: 2486-2495 [PMID: 17142798 DOI: 10.1096/fj.06-6391.com]
 - 71 **Nishiyama K**, Azuma YT, Morioka A, Yoshida N, Teramoto M, Tanioka K, Kita S, Hayashi S, Nakajima H, Iwamoto T, Takeuchi T. Roles of Na(+)/Ca(2+) exchanger isoforms NCX1 and NCX2 in motility in mouse ileum. *Naunyn Schmiedeberg's Arch Pharmacol* 2016; **389**: 1081-1090 [PMID: 27411318 DOI: 10.1007/s00210-016-1271-1]
 - 72 **Nishiyama K**, Morioka A, Kita S, Nakajima H, Iwamoto T, Azuma YT, Takeuchi T. Na/Ca(2+) exchanger 1 transgenic mice display increased relaxation in the distal colon. *Pharmacology* 2014; **94**: 230-238 [PMID: 25427675 DOI: 10.1159/000363246]
 - 73 **Mitrovic S**, Nogueira C, Cantero-Recasens G, Kiefer K, Fernández-Fernández JM, Popoff JF, Casano L, Bard FA, Gomez R, Valverde MA, Malhotra V. TRPM5-mediated calcium uptake regulates mucin secretion from human colon goblet cells. *Elife* 2013; **2**: e00658 [PMID: 23741618 DOI: 10.7554/eLife.00658]
 - 74 **Nishiyama K**, Azuma YT, Kita S, Azuma N, Hayashi S, Nakajima H, Iwamoto T, Takeuchi T. Na/Ca²⁺ exchanger 1/2 double-heterozygote knockout mice display increased nitric oxide component and altered colonic motility. *J Pharmacol Sci* 2013; **123**: 235-245 [PMID: 24162024]
 - 75 **Nishiyama K**, Tanioka K, Azuma YT, Hayashi S, Fujimoto Y, Yoshida N, Kita S, Suzuki S, Nakajima H, Iwamoto T, Takeuchi T. Na⁺/Ca²⁺ exchanger contributes to stool transport in mice with experimental diarrhea. *J Vet Med Sci* 2017; **79**: 403-411 [PMID: 27928109 DOI: 10.1292/jvms.16-0475]
 - 76 **Briejer MR**, Schuurkes JA. 5-HT3 and 5-HT4 receptors and cholinergic and tachykinergic neurotransmission in the guinea-pig proximal colon. *Eur J Pharmacol* 1996; **308**: 173-180 [PMID: 8840129]
 - 77 **Tuladhar BR**, Costall B, Naylor RJ. 5-HT3 and 5-HT4 receptor-mediated facilitation of the emptying phase of the peristaltic reflex in the marmoset isolated ileum. *Br J Pharmacol* 1996; **117**: 1679-1684 [PMID: 8732276]
 - 78 **Rivière PJ**, Farmer SC, Burks TF, Porreca F. Prostaglandin E2-induced diarrhea in mice:

- importance of colonic secretion. *J Pharmacol Exp Ther* 1991; **256**: 547-552 [PMID: [1993994](#)]
- 79 **Wardle TD**, Hall L, Turnberg LA. Inter-relationships between inflammatory mediators released from colonic mucosa in ulcerative colitis and their effects on colonic secretion. *Gut* 1993; **34**: 503-508 [PMID: [8491398](#)]
- 80 **Ankorina-Stark I**, Amstrup J, Novak I. Regulation of the Na⁺/Ca²⁺ exchanger in rat pancreatic ducts. *J Membr Biol* 2002; **186**: 43-53 [PMID: [11891588](#) DOI: [10.1007/s00232-001-0134-x](#)]
- 81 **Quednau BD**, Nicoll DA, Philipson KD. Tissue specificity and alternative splicing of the Na⁺/Ca²⁺ exchanger isoforms NCX1, NCX2, and NCX3 in rat. *Am J Physiol* 1997; **272**: C1250-C1261 [PMID: [9142850](#) DOI: [10.1152/ajpcell.1997.272.4.C1250](#)]
- 82 **Hamming KS**, Riedel MJ, Soliman D, Maternisz LC, Webster NJ, Searle GJ, MacDonald PE, Light PE. Splice variant-dependent regulation of beta-cell sodium-calcium exchange by acyl-coenzyme A. *Mol Endocrinol* 2008; **22**: 2293-2306 [PMID: [18635667](#) DOI: [10.1210/me.2008-0053](#)]
- 83 **Herchuelz A**, Diaz-Horta O, van Eylen F. Na⁺/Ca exchange and Ca²⁺ homeostasis in the pancreatic beta-cell. *Diabetes Metab* 2002; **28**: 3554-60; discussion 35108-12 [PMID: [12688634](#)]
- 84 **Van Eylen F**, Bollen A, Herchuelz A. NCX1 Na/Ca exchanger splice variants in pancreatic islet cells. *J Endocrinol* 2001; **168**: 517-526 [PMID: [11241183](#)]
- 85 **Rorsman P**, Renström E. Insulin granule dynamics in pancreatic beta cells. *Diabetologia* 2003; **46**: 1029-1045 [PMID: [12879249](#) DOI: [10.1007/s00125-003-1153-1](#)]
- 86 **Bruce JL**, Yang X, Ferguson CJ, Elliott AC, Steward MC, Case RM, Riccardi D. Molecular and functional identification of a Ca²⁺ (polyvalent cation)-sensing receptor in rat pancreas. *J Biol Chem* 1999; **274**: 20561-20568 [PMID: [10400686](#)]
- 87 **Herchuelz A**, Diaz-Horta O, Van Eylen F. Na/Ca exchange in function, growth, and demise of beta-cells. *Ann N Y Acad Sci* 2002; **976**: 315-324 [PMID: [12502574](#)]
- 88 **Herchuelz A**, Kamagate A, Ximenes H, Van Eylen F. Role of Na/Ca exchange and the plasma membrane Ca²⁺-ATPase in beta cell function and death. *Ann N Y Acad Sci* 2007; **1099**: 456-467 [PMID: [17446486](#) DOI: [10.1196/annals.1387.048](#)]
- 89 **Herchuelz A**, Pachera N. The Na⁺/Ca²⁺ exchanger and the Plasma Membrane Ca²⁺-ATPase in β -cell function and diabetes. *Neurosci Lett* 2018; **663**: 72-78 [PMID: [28780165](#) DOI: [10.1016/j.neulet.2017.08.009](#)]
- 90 **Van Eylen F**, Horta OD, Barez A, Kamagate A, Flatt PR, Macianskiene R, Mubagwa K, Herchuelz A. Overexpression of the Na/Ca exchanger shapes stimulus-induced cytosolic Ca(2+) oscillations in insulin-producing BRIN-BD11 cells. *Diabetes* 2002; **51**: 366-375 [PMID: [11812743](#)]
- 91 **Petersen OH**, Tepikin AV, Gerasimenko JV, Gerasimenko OV, Sutton R, Criddle DN. Fatty acids, alcohol and fatty acid ethyl esters: toxic Ca²⁺ signal generation and pancreatitis. *Cell Calcium* 2009; **45**: 634-642 [PMID: [19327825](#) DOI: [10.1016/j.ceca.2009.02.005](#)]
- 92 **Gerasimenko JV**, Gryshchenko O, Ferdek PE, Stapleton E, Hébert TO, Bychkova S, Peng S, Begg M, Gerasimenko OV, Petersen OH. Ca²⁺ release-activated Ca²⁺ channel blockade as a potential tool in antipancreatitis therapy. *Proc Natl Acad Sci U S A* 2013; **110**: 13186-13191 [PMID: [23878235](#) DOI: [10.1073/pnas.1300910110](#)]
- 93 **Vigna SR**, Shahid RA, Liddle RA. Ethanol contributes to neurogenic pancreatitis by activation of TRPV1. *FASEB J* 2014; **28**: 891-896 [PMID: [24221085](#) DOI: [10.1096/fj.13-236208](#)]
- 94 **Yu SY**, Xie R, Chen YY, Yang SM, Zhu B. Role of sodium-calcium exchanger-1 in development of rat acute pancreatitis. *Di-San Junyi Daxue Xuebao* 2015; **37**: 1325-1330 [DOI: [10.16016/j.1000-5404.201501236](#)]
- 95 **Jemal A**, Siegel R, Ward E, Hao Y, Xu J, Murray T, Thun MJ. Cancer statistics, 2008. *CA Cancer J Clin* 2008; **58**: 71-96 [PMID: [18287387](#) DOI: [10.3322/ca.2007.0010](#)]
- 96 **Schmidt CM**, Powell ES, Yiannoutsos CT, Howard TJ, Wiebke EA, Wiesenauer CA, Baumgardner JA, Cummings OW, Jacobson LE, Broadie TA, Canal DF, Goulet RJ Jr, Curie EA, Cardenes H, Watkins JM, Loehrer PJ, Lillemoe KD, Madura JA. Pancreaticoduodenectomy: a 20-year experience in 516 patients. *Arch Surg* 2004; **139**: 718-25; discussion 725-7 [PMID: [15249403](#) DOI: [10.1001/archsurg.139.7.718](#)]
- 97 **Truty MJ**, Urrutia R. Basics of TGF-beta and pancreatic cancer. *Pancreatolgy* 2007; **7**: 423-435 [PMID: [17898532](#) DOI: [10.1159/000108959](#)]
- 98 **Dong H**, Shim KN, Li JM, Estrema C, Ornelas TA, Nguyen F, Liu S, Ramamoorthy SL, Ho S, Carethers JM, Chow JY. Molecular mechanisms underlying Ca²⁺-mediated motility of human pancreatic duct cells. *Am J Physiol Cell Physiol* 2010; **299**: C1493-C1503 [PMID: [20861471](#) DOI: [10.1152/ajpcell.00242.2010](#)]
- 99 **Xu JY**, Jiang YX, Xie R, Jin H, Wen GR, Tuo BG. The expression of NCX1 and its effect on proliferation and migration of hepatocellular carcinoma cells through regulation of intracellular Ca²⁺. *Zhongguo Aizheng Zazhi* 2016; **26**: 735-742 [DOI: [10.19401/j.cnki.1007-3639.2016.09.003](#)]
- 100 **Guan LY**, Fu PY, Li PD, Li ZN, Liu HY, Xin MG, Li W. Mechanisms of hepatic ischemia-reperfusion injury and protective effects of nitric oxide. *World J Gastrointest Surg* 2014; **6**: 122-128 [PMID: [25068009](#) DOI: [10.4240/wjgs.v6.i7.122](#)]
- 101 **Vasques ER**, Cunha JE, Coelho AM, Sampietre SN, Patzina RA, Abdo EE, Nader HB, Tersariol IL, Lima MA, Godoy CM, Rodrigues T, Chaib E, D'Albuquerque LA. Trisulfate Disaccharide Decreases Calcium Overload and Protects Liver Injury Secondary to Liver Ischemia/Reperfusion. *PLoS One* 2016; **11**: e0149630 [PMID: [26901764](#) DOI: [10.1371/journal.pone.0149630](#)]
- 102 **Nakamura T**, Arii S, Monden K, Furutani M, Takeda Y, Imamura M, Tominaga M, Okada Y. Expression of the Na⁺/Ca²⁺ exchanger emerges in hepatic stellate cells after activation in association with liver fibrosis. *Proc Natl Acad Sci U S A* 1998; **95**: 5389-5394 [PMID: [9560286](#)]
- 103 **Xu J**, Yang Y, Xie R, Liu J, Nie X, An J, Wen G, Liu X, Jin H, Tuo B. The NCX1/TRPC6 Complex Mediates TGF β -Driven Migration and Invasion of Human Hepatocellular Carcinoma Cells. *Cancer Res* 2018; **78**: 2564-2576 [PMID: [29500176](#) DOI: [10.1158/0008-5472.can-17-2061](#)]
- 104 **Tian Y**, Zhu MX. A novel TRPC6-dependent mechanism of TGF- β -induced migration and invasion of human hepatocellular carcinoma cells. *Sci China Life Sci* 2018; **61**: 1120-1122 [PMID: [30136057](#) DOI: [10.1007/s11427-018-9365-7](#)]
- 105 **Rusolo F**, Pucci B, Colonna G, Capone F, Guerriero E, Milone MR, Nazzaro M, Volpe MG, Di Bernardo G, Castello G, Costantini S. Evaluation of selenite effects on selenoproteins and cytokine in human hepatoma cell lines. *Molecules* 2013; **18**: 2549-2562 [PMID: [23442931](#) DOI: [10.3390/molecules18032549](#)]

- 106 **Xu J**, Ji B, Wen G, Yang Y, Jin H, Liu X, Xie R, Song W, Song P, Dong H, Tuo B. Na⁺/H⁺ exchanger 1, Na⁺/Ca²⁺ exchanger 1 and calmodulin complex regulates interleukin 6-mediated cellular behavior of human hepatocellular carcinoma. *Carcinogenesis* 2016; **37**: 290-300 [PMID: 26775040 DOI: 10.1093/carcin/bgw004]

P- Reviewer: Dong H, Sandow SL, Touil-Boukoffa C

S- Editor: Gong ZM **L- Editor:** Wang TQ **E- Editor:** Huang Y





Published By Baishideng Publishing Group Inc
7901 Stoneridge Drive, Suite 501, Pleasanton, CA 94588, USA
Telephone: +1-925-2238242
Fax: +1-925-2238243
E-mail: bpgoffice@wjgnet.com
Help Desk: <http://www.f6publishing.com/helpdesk>
<http://www.wjgnet.com>

