**Name of Journal:** *World Journal of Clinical Cases*

**Manuscript NO:** 47651

**Manuscript Type:** REVIEW

**Bone alterations in inflammatory bowel diseases**

Sgambato D *et al*. Bone and IBD

Dolores Sgambato, Francesca Gimigliano, Cristiana De Musis, Antimo Moretti, Giuseppe Toro, Emanuele Ferrante, Agnese Miranda, Domenico De Mauro, Lorenzo Romano, Giovanni Iolascon, Marco Romano

**Dolores Sgambato, Cristiana De Musis, Emanuele Ferrante, Agnese Miranda, Domenico De Mauro, Marco Romano,** Departments of Precision Medicine and Polyspecialistic Internal Medicine, University of Campania ‘‘Luigi Vanvitelli’’ and University Hospital, Naples 80131, Italy

**Francesca Gimigliano,** Department of Physical and Mental Health, University of Campania “Luigi Vanvitelli”, Naples 80131, Italy

**Antimo Moretti, Giuseppe Toro, Giovanni Iolascon,** Department of Medical and Surgical Specialties and Dentistry, University of Campania “Luigi Vanvitelli”, Naples 80131, Italy

**Lorenzo Romano,** Surgical Digestive Endoscopy, Department of Clinical Medicine and Surgery, Federico II University, Naples 80131, Italy

**ORCID number:** Dolores Sgambato (0000-0002-7501-3792); Francesca Gimigliano (0000-0002-1905-6405); Cristiana De Musis (0000-0001-7011-5047); Antimo Moretti (0000-0002-4598-2891); Giuseppe Toro (0000-0002-8560-721X); Emanuele Ferrante (0000-0001-5612-0560);, Agnese Miranda (0000-0003-4682-9087); Domenico De Mauro (0000-0002-4484-0963); Lorenzo Romano (0000-0002-6581-7930); Giovanni Iolascon (0000-0002-0976-925X); Marco Romano (0000-0002-3271-349X).

**Author contributions:** All authors contributed to this paper with conception and design of the study, literature review and analysis, drafting and critical revision and editing, and final approval of the final version. In particular DS and FG equally contributed to the manuscript.

**Conflict-of-interest statement:** No potential conflicts of interest. No financial support.

**Open-Access:** This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: http://creativecommons.org/licenses/by-nc/4.0/

**Manuscript source:** Invited manuscript

**Corresponding author:** Marco Romano, MD, Professor, Departments of Precision Medicine and Polyspecialistic Internal Medicine, University of Campania ‘‘Luigi Vanvitelli’’ and University Hospital. University of Campania “Luigi Vanvitelli”, Via Pansini 5, Naples 80131, Italy. marco.romano@unicampania.it

Telephone: +39-081-5665718

Fax: +39-081-5665714

**Received:** March 20, 2019

**Peer-review started:** March 20, 2019

**First decision:** May 9, 2019

**Revised:** June 14, 2019

**Accepted:** June 14, 2019

**Article in press:** June 26, 2019

**Published online:** August 6, 2019

Abstract

Inflammatory bowel diseases (IBDs) are characterized by a multifactorial partially unknown etiology that involves genetic, immunological and environmental factors. Up to 50% of IBD patients experience at least one extraintestinal manifestation; among them is the involvement of bone density which is referred to as metabolic bone disease (MBD), including osteopenia and osteoporosis. Bone alterations in IBDs population appear to have a multifactorial etiology: decreased physical activity, inflammation-related bone resorption, multiple intestinal resections, dietary malabsorption of minerals and vitamin D deficiency, genetic factors, gut-bone immune signaling interaction, steroid treatment, microbiota and pathogenic micro-organisms interaction, and dietary malabsorption of minerals, that, all together or individually, may contribute to the alteration of bone mineral density. This review aims to summarize the prevalence and pathophysiology of metabolic bone alterations in IBD subjects outlining the main risk factors of bone fragility. We, also, want to underline the role of the screening and prophylaxis of bone alterations in Crohn’s disease and ulcerative colitis patients and the importance of treating appropriately MBD.

**Key words:** Inflammatory bowel diseases; Bone alterations; Bone mineral density; Osteoporosis; Osteopenia; Ulcerative colitis; Crohn’s disease

**© The Author(s) 2019.** Published by Baishideng Publishing Group Inc. All rights reserved.

**Core tip:** Up to 50% of inflammatory bowel disease (IBD) patients experience at least one extraintestinal manifestation; among them is the involvement of bone density which is referred to as metabolic bone disease (MBD), including osteopenia and osteoporosis. Bone alterations in IBD population appear to have a multifactorial etiology. This review summarizes the prevalence and pathophysiology of metabolic bone alterations in IBD subjects outlining the main risk factors of bone fragility. We, also, want to underline the role of the screening and prophylaxis of bone alterations in Crohn’s disease and ulcerative colitis patients and the importance of treating appropriately MBD.

**Citation:** Sgambato D, Gimigliano F, De Musis C, Moretti A, Toro G, Ferrante E, Miranda A, De Mauro D, Romano L, Iolascon G, Romano M. Bone alterations in inflammatory bowel diseases. *World J Clin Cases* 2019; 7(15): 1908-1925

**URL:** https://www.wjgnet.com/2307-8960/full/v7/i15/1908.htm

**DOI:** https://dx.doi.org/10.12998/wjcc.v7.i15.1908

INTRODUCTION

Inflammatory bowel diseases (IBDs) are mainly represented by Crohn’s disease (CD) and ulcerative colitis (UC), both characterized by a multifactorial, partially unknown etiology that involves genetic, immunological and environmental factors including intestinal microbiota[1]. A dysregulated immune response to an unknown trigger leading to a sustained pro-inflammatory response within the gastrointestinal (GI) tract seems to play a major pathogenic role. Also, 10%-40% of IBD patients may suffer from at least one extraintestinal manifestation (EIM)[2], even before the occurrence of the intestinal disease[3].

Among the most frequent EIMs, there are those affecting the musculoskeletal system and in particular the bone tissue, such as osteoporosis (OP)[4].

OP is a systemic disease characterized by an increased risk of fractures even after a low energy trauma (fragility fracture). The reduction of bone strength is a consequence of a decrease in bone mineral density (BMD) and a deterioration in bone quality[5]. According to the World Health Organization the operational diagnosis of OP is based on BMD values equal or lower than 2.5 standard deviations (SD) from the average values for young healthy women (T-score < -2.5 SD) in post-menopausal women and men aged ≥ 50 years, while, osteopenia is defined by BMD values between -1 to -2.5 SD (T-score -1< and > -2.5).

BMD is measured trough dual-energy X-ray absorptiometry (DXA)[6,7]; T-score is a parameter comparing the BMD of a given patient with the average bone density of young healthy adults of the same sex, while, Z-score compares each BMD with the average BMD of a person with the same age and sex[8].

OP is classified into two main groups: primary (or idiopathic) and secondary. The first one is the most common and includes juvenile, postmenopausal and senile OP. Secondary OP might be caused by several conditions (*i.e.*, endocrine, hematological, GI, rheumatic or renal diseases) that negatively affect bone metabolism leading to poor bone strength. Other causes might be the chronic use of some medications, particularly glucocorticoids (GCs), anticoagulants, and anticonvulsants[9,10]. Therefore, the term “secondary OP” refers to all those clinical conditions in which the bone involvement is a consequence of the primary disease or results from the related treatments (*i.e.*, GCs). Secondary OP affects about 60% of males and more than 50% of premenopausal women[11,12].

In IBD population, there are several pathological mechanisms that might result in low BMD and poor bone strength, thus leading to OP.

This review aims to summarize the prevalence and pathophysiology of metabolic bone alterations in IBD subjects outlining their main risk factors. We also underline the role of the screening and prophylaxis of BMD in CD and UC patients and the importance of early treatment.

EPIDEMIOLOGY

OP is one of the most common noncommunicable diseases[13] and its incidence is increasing worldwide[14]. According to Svedbom *et al*[15], 22 million women and 5.5 million men were estimated to have OP in Europe, with a reported incidence of 3.5 million new osteoporotic fractures in 2010, (620000 hip fractures, 520000 vertebral fractures, 560000 forearm fractures and 1800000 fractures in other skeletal sites). OP and its consequences (*i.e.*, fractures) carry a considerable economic burden on the health care systems[15] and, in particular, the socioeconomic costs of an osteoporotic hip fracture are equivalent to those of myocardial infarction and stroke[16].

In IBD subjects, the prevalence of low BMD ranges from 22% to 77% and that of fragility fractures from 17% to 41%[17]. These wide ranges across different studies might be explained by the small number of samples and by the heterogeneity of the studies and of the populations.

Several studies have been performed to describe the relationship between IBD and bone alterations. Sheth *et al*[2] showed that both osteopenia and OP are frequently associated with IBD, ranging from 32% to 36% for osteopenia and from 7% to 15% for OP. The same study reported an increased relative risk of fragility fractures in CD patients and a prevalence of < 0.5% for osteonecrosis, a clinical condition characterized by the death of the bone tissue, commonly described as a complication of steroid therapy in IBD patients. A study of Boubaker *et al*[18] reported that in a Tunisian group of 67 patients, OP represented the most frequent EIM in CD patients with a prevalence of low BMD at hip and spine in 31.8% and 40.9% of cases, respectively. OP is strongly associated with CD in females, thus suggesting that female gender might be one of the risk factors for bone loss in IBD. A prospective study in Romania found osteopenia in 48.07% of UC patients and in 56.41% of CD patients, while OP was shown in 18.26% of UC patients and in 15.38% of CD patients[19]. A Swiss IBD cohort study performed on 877 patients showed a prevalence of bone density alteration in 20% of IBD patients and identified, by multivariate logistic regression analysis, corticosteroid usage, long disease duration and perianal disease as independent risk factors[4].A Japanese study reported that two-thirds of IBD patients showed a loss of BMD, with a prevalence of OP of about 13% in their cohort of patients with mean age of 43 years. The prevalence of OP is more frequent in Western IBD population than in the Asiatic one[20] and, therefore, the prevalence of bone metabolism alterations generally varies depending on study population, location and design of the study performed.

Notably, the risk of fragility fractures seems to be increased in IBD population[21] although the literature shows controversial results. Recently, Komaki *et al*[22] showed that there was no increase in the risk of overall fractures in IBD patients, but they reported more fractures at the spine, associated with steroids therapy.

Pediatric IBDs seem to show a similar association with osteopenia and OP as in adults. In fact, it is known that children with IBD have a higher risk of low BMD[23]. The overall prevalence of osteopenia and OP in pediatric and young IBDs patients seems to vary from 20% to 50%[24]. A recent study suggested a positive association between BMD and physical activity and between low BMD and fractures in the childhood[25]. Incidence of fragility fractures is higher in the young IBD population and is likely to be associated to the use of GC[26,27]. Pediatric CD patients appear to be more severely compromised than those with UC, probably because CD inhibits linear growth more frequently than UC[28].

CD and UC, while being both classified as IBDs, show considerable differences in the anatomic location and distribution of the intestinal lesions as well as in the underlying pathogenic mechanisms. This might have an influence on the incidence of bone alterations in each condition. Bjarnason *et al*[29] described no significant differences in T scores for spine or hip between patients with CD and those with UC. On the other side, in a study by Jahnsen *et al*[30], BMD resulted significantly reduced in CD subjects at all measured sites compared with UC patients, and healthy subjects. Interestingly, the authors did not describe significant differences in BMD between UC and healthy subjects. Ardizzone *et al*, in Italy, evaluated differences between CD and UC with respect to the pathogenic mechanisms underlying bone loss. Crohn’s Disease Activity Index (CDAI)[31] and Truelove and Witts’ Score[32] were used to grade disease activity of CD and UC, respectively. The distribution of normal, osteopenic and osteoporotic BMD values among CD and UC showed no significant differences also between patients with different disease activity. Also, in a study conducted in Sri-Lanka, there were no significant statistical differences in the frequency of OP between CD and UC, whereas the occurrence of OP among IBDs patients (13.5%) was higher than in healthy controls[33]. Recently, Vázquez *et al*[34] did not find any difference between patients with CD and those with UC regarding the prevalence of alterations of bone density.

The discrepancy between different studies may be due to variability in patient selection, differences in the methods used to evaluate bone density, and the body sites studied at DEXA (*i.e.*, radius *vs* lumbar spine or hip).

PATHOPHYSIOLOGY

Both bone quality and quantity (BMD) depend on physiological mechanisms, such as bone modeling and remodeling that in turn are regulated by biochemical and mechanical factors, including osteoprotegerin (OPG), receptor activator of nuclear factor kappa-B ligand (RANKL), receptor activator of nuclear factor kappa-B (RANK), and weight bearing activities. In particular, the RANKL secreted by osteoblasts binds to the RANK receptor, located on pre-osteoclasts and mature osteoclasts, inducing osteoclast proliferation, activity and survival. OPG, a molecule secreted by osteoblasts, modulates bone turnover by inhibiting the binding of RANKL to RANK. The balance between OPG and RANKL release regulates osteoclast activity that in turn can be influenced by several hormones and cytokines, including vitamin D, estrogens, testosterone, GCs, parathormone (PTH), as well as pro-inflammatory mediators, such as interleukin-1 (IL-1) and tumor necrosis factor-alfa (TNFα). In OP, an imbalance between the serum levels of OPG and RANKL occurs, with excessive bone resorption and impaired bone formation, with consequent overall reduction of bone mass[35].

Other key modulators of bone turnover are the Wnt/β-catenin signaling and sclerostin, which acts mainly on osteoblasts and osteocytes. Sclerostin is a glycoprotein produced almost exclusively by osteocytes and its expression is influenced by many factors, including serum PTH and mechanical loading[36]. Animal studies have shown that the mechanical load reduces serum sclerostin, whereas unloading increases the transcription of *SOST*, the gene encoding for sclerostin. Once secreted, sclerostin through osteocyte canalicular system reaches the bone lining cells (capable of activation into mature osteoblasts), where it binds to specific co-receptors (LRP-5 and -6) to inhibit the Wnt pathway, with consequent reduction of osteoblastogenesis and bone formation. On the contrary, a reduction in sclerostin levels is associated with an activation of the Wnt/β-catenin pathway with subsequent enhancement of osteoblast activity and survival[37]. Moreover, there is a correlation between hypersclerostinemia and bone loss in subjects forced to prolonged immobilization, thus supporting the key role of sclerostin in the development of OP following reduced mechanical load[38].

Among the most frequent secondary forms of OP, GCs use has been shown to decrease the number of osteoblast precursors and to increase the apoptosis of mature osteoblasts. The reduction in osteoblast differentiation is in partly mediated by the inhibition of the Wnt/β-catenin pathway along with increased expression of sclerostin, that antagonizes the Wnt signaling[39]. On the other hand, GCs decrease also osteoclast proliferation, although their activity tends to increase, through both the increase in RANKL and the reduction of OPG levels[40]. During GCs therapy, bone loss occurs rapidly with BMD reduction of 6%-12% within the first year, followed by a constant and gradual loss throughout the treatment period[41]. These two pathways are the basis of the most modern pharmacological approaches to OP: a human monoclonal antibody against the RANKL (denosumab) and a humanized monoclonal antibody that targets sclerostin (romosozumab).

Among the most frequent causes of secondary OP there are GI disorders, including IBDs. Chronic gut inflammation in IBD may contribute to OP through the activation of T lymphocytes resulting in enhanced release of inflammatory cytokines, such as TNFα, that modulates the OPG/RANKL/RANK pathway thus inducing bone loss[42] (Figure 1). Moreover, TNFα enhances sclerostin production that results in decreased bone formation. Interestingly, in patients affected by CD, bone loss occurs before GCs administration, supporting the detrimental role of systemic inflammation on bone health[43].

There is a close relationship between bone and GI system that allows calcium absorption and bone mineralization; the GI tract may communicate with bone tissue through different mechanisms such as blood, nerves and immune cells, defining a characteristic gut-to-bone signaling axis that involves also incretins, serotonin and GI microbiota[44] (Figure 2).

Although in the general population female sex, early menopause, hormonal imbalance, smoke and old age are the main risk factors correlated with the onset of metabolic bone disease (MBD), in IBD patients the main risk factor seems to be the prolonged use of GCs[45,46]. Activity and severity of gut inflammation, intestinal malabsorption and calcium and vitamin D deficiency are also directly involved in the loss of BMD[47,48]. In particular, in CD the involvement of the terminal ileum may affect the bile salt enterohepatic circulation thus leading to a reduced absorption of vitamin D. Chronic inflammation seems to have a key role in the reduction of BMD as suggested in some studies that show that rat models with colitis have a drastic loss of trabecular bone and a suppression of bone formation. At resolution of colitis with mucosal healing, the bone formation regresses to normal levels. During gut inflammation some mediators that alter the deposition of new bone matrix mediated by osteoblasts are produced, such as IL-6 and RANKL[49].

Untreated inflammation may be the main determinant for the loss of BMD in IBDs. In fact, a study by Ghosh et al showed that CD patients have very low T-scores at the diagnosis before any drug therapy[43]. Several studies in last years have also demonstrated that the usage of anti-TNF agents seems to have a positive effect on BMD[50] (see below). Recent studies suggest a role for the inflammatory process in the alteration of bone metabolism through the involvement of immune system cells, however it is not clear whether the inflammation is directly involved in the loss of BMD or if other factors contribute to the decline of BMD in IBD patients.

Bone alterations in IBD population appear to have a multifactorial etiology: genetic factors, gut-bone immune signaling interaction, inflammation-related bone resorption, multiple intestinal resections, microbiota and pathogenic micro-organisms interaction, and dietary malabsorption of minerals[51,52] (Figure 2).

However, the main factor that seems to directly affect bone metabolism, with consequent reduction in BMD values, is corticosteroid treatment. In the following paragraphs we will summarize the evidence linking these factors to BMD deterioration in IBD patients.

***Main risk factors***

**Genetic factors:** The literature concerning the genetic predisposition to OP in IBD patients appears rather contradictory. A number of polymorphic sites apparently associated with the increased risk of bone loss in IBD patients such as *IL-6* and *IL-1ra* have already been described[53]. Todhunter *et al*[54] showed that the polymorphisms identified in some genes such as *COL1A1* and *IL-6* seems to influence BMD in IBD patients, particularly those with CD. Nemetz et al. showed also an increased risk of bone loss in patients with *IL1B* polymorphism (*IL1B-511\*2*) associated with hypersecretion of *IL1B*[55].

One of the latest genetic factors associated with OP development is OPG encoded by the gene *TNFRS11B*, where the polymorphism c.-223C>T in 5’ UTR region was identified as strongly related to OP in postmenopausal women. However, the genotyping analysis did not show unequivocal association between c.-223C>T of *TNFRS11B* and a predisposition to OP in IBD patients, although this polymorphism is more frequent in IBD patients than in healthy controls[56]. Krela-Kaźmierczak *et al*[56] studied the relationship between different polymorphic variants of *TGFB*1 and bone loss in IBD, finding no significant differences in BMD values ​​or in the risk of fragility fractures between UC and CD patients and healthy controls with different polymorphic variants of the *TGFB1* gene. Moreover, no association between the 29T>C polymorphic variant of *TGFB1* and BMD of spongy bone and cortical bones was found[57].

The gene encoding for bone morphogenetic protein 2 (BMP2) was analyzed using restriction fragments length polymorphisms (RFLPs) to determine the association among the incidence of 570 A>T polymorphism, BMD alterations, and the incidence of fractures in IBD patients. The analysis revealed no significant association between this polymorphism and changes in bone metabolism in both UC and CD patients[58].

An interesting and recent theory correlates the presence of genetic alterations that might affect the ability to respond to endoplasmic reticulum (ER) stress and normal bone tissue physiology in IBD patients. A recent meta-analysis focused on the association between the genes involved in the response to unfolded proteins (UPR) and ER stress, which could directly correlate with the pathogenesis of IBDs[59]. It was suggested that the same defects in the Paneth cells inherent in UPR may also be present in bone cells, both osteoblasts and osteoclasts (although this has not yet been confirmed in IBD)[60]. Although its polymorphic variant seems to have no direct association with BMD, BMP2 activates the UPR during osteogenesis[61] allowing the production of RANKL[62]. In conclusion, during osteoblast differentiation ER stress is induced and activates also the PERK-eIF2α-ATF4 pathway that appears as a potential target against bone diseases[61]. To date, the association between genetic factors and bone alterations in IBD patients is not clear and other risk factors must be taken into account, such as those related to nutrition and lifestyle, as well as the ethno-demographic characteristics.

**Gut-bone immune signaling:** Bone structure depends on the balance between osteoblasts activity, specialized in the deposition of new bone matrix and osteoclasts, responsible for the resorption of bone tissue. Mounting evidence suggests an immunological involvement in the alteration of bone metabolism[63]. Activated CD4+ cells appear to be important actors in the bone loss related to IBD. In fact, in mice models, bone marrow CD4+ cells producing IL-17 and TNF-α migrate into the bone marrow during the inflammation, promoting the recruitment of monocytes as osteoclast progenitors, thus contributing to the bone loss[64]. Ashcroft et al. reported that activated T cells, producing RANKL, are accumulated in the bone marrow during intestinal inflammation[65].

As already discussed, osteoclastogenesis is guided by the RANK-RANKL pathway and by the RANKL/OPG ratio. OPG produced by osteoblasts works as a decoy receptor for RANKL, thus interfering with osteoclast activation. Li *et al*[66] suggested that lymphocytes might work as key regulators of bone metabolism by interfering with this pathway. In particular, they found that over 60% of OPG is produced by B lymphocytes and that T lymphocytes stimulate OPG production by osteoblasts via CD40L/CD40 co-stimulation. Finally, they showed that B-cells as well as CD40 or CD40L knockout mice developed OP and OPG deficiency[66].

Current knowledge strongly suggests a dynamic interplay between skeletal and immune system which is referred to as osteoimmunology. IL-17-producing helper T cells [T(H)17] induce RANKL, stimulating osteoclastogenesis through nuclear factor of activated T cells cytoplasmic 1 (NFATc1)[67]. There is evidence of activation of NFAT2 in lamina propria mononuclear cells of subjects with UC[68].

All these studies seem to show, in IBD, a direct involvement of immune system in the bone loss, mainly due to CD4+ cells that appear osteoclastogenic during inflammation.

**Microbiota and pathogenic micro-organisms:** The human microbiota consists of a set of about 100 trillion of commensal micro-organisms belonging to different species, which express a genome of about a hundred times greater than that expressed by the host’s cells. There is evidence that alterations of the microbiota composition influence the healthy state of the host[69].

In particular, an immune response to an altered intestinal microbiota or an alteration of the immune response leading to its activation in the face of a normal intestinal microbiota both leading to a sustained inflammatory process, have been suggested as the main pathogenic mechanisms for the development of IBD[70].

Several studies suggest a key role for microbiota in the alteration of BMD. Irwin *et al*[71] reported a close correlation between *H. hepaticus* infection and bone loss. McCabe *et al*[72] showed that treating healthy male mice with *Lactobacillus reuteri* enhanced bone density and suppressed basal TNFα mRNA levels in male mice, but not in females. They also showed that probiotics increased male trabecular bone parameters, as mineral density in the distal femur metaphyseal region as well as in the lumbar vertebrae and increased osteoblast serum markers in male mice, although no effect on bone parameters in females was found[72]. Furthermore, Schepper *et al*[73] investigated the effect of antibiotic treatment on gut and bone health in mice models. They found an increase in the Firmicutes/Bacteroidetes ratio, in the intestinal permeability, and a reduction of femoral trabecular bone volume. Treating the mice with *Lactobacillus reuteri* reduced the post-antibiotic elevation of the Firmicutes/Bacteroidetes ratio and prevented bone loss. Antibiotic-induced dysbiosis was associated with decreased osteoblast and increased osteoclast activities[73].

Recently, Naser *et al*[74] showed that the inflammation associated to *Mycobacterium avium* *subspecies paratuberculosis* (MAP) infection results in elevation of undercarboxylated osteocalcin (ucOC) and downregulation of active osteocalcin (OC) in CD patients. This suggest that MAP infection may serve as a trigger factor in the development of OP in CD patients.

**Nutrition and vitamin deficiency:** Nutritional alterations and vitamins or minerals deficiency due to inadequate diet intake and/or malabsorption correlate with a low BMD and may contribute to the development of osteopenia and OP both in UC and CD patients. Lim *et al*[75] assessed the nutritional status of 41 IBD patients with or without malnutrition and showed significantly higher serum C-reactive protein (CRP) and lower serum calcium in the malnourished group. No significant differences between malnourished and normal group were found as to BMD although lower bone density was more frequent in malnourished group. Also, Azzopardi et al. found a significant correlation between body mass index (BMI) and BMD of IBD patients[47,76].

Many studies measured calcium daily intake in IBD patients. Silvennoinen *et al*[77] assessed calcium intake and measured BMD in IBD patients and controls. They found that, although the daily intake of calcium was lower in IBD subjects than in controls, especially in male patients, there was no significant correlations between calcium deficiency and BMD[77]. Vernia *et al*[78] analyzed by means of a questionnaire the dietary calcium intake in 187 IBD patients. They confirmed that calcium intake is frequently lower in IBD patients; moreover, most of the patients adopted some arbitrary dietary restrictions (*i.e.*, avoidance of milk or dairy products) which increased the risk of OP[78]. Calcium supplements as well as vitamin D administration showed an improvement of BMD at lumbar spine in osteoporotic patients with IBD. On the contrary, fluoride supplementation does not seem to provide any benefit to IBD patients[79].

Vitamin D has systemic functions; it acts on regulation of the innate and adaptive immune responses and modulates calcium homeostasis involved in bone metabolism. In IBD patients a vitamin D deficiency negatively affects the immune system inducing dysregulation and inflammation-associated loss of BMD[80]. Vitamin D deficiency is more frequent in IBD patients than in the general population[81]. Del Pinto *et al*[82], in a meta-analysis involving 14 studies, with 938 IBD patients and 953 controls, showed that 64% of IBD patients had lower vitamin D serum levels than controls. Interestingly, UC appeared to be associated with more than double the odds of vitamin D deficiency compared to healthy controls. A recent study, aimed at evaluating the absorption of orally administered vitamin D in CD patients compared with healthy controls, showed a great variability in the bioavailability of vitamin D(2) in CD patients although no significant differences between patients with different location of disease or among those with or without previous surgery were found. Moreover, 24 h after an oral load of vitamin D(2), the authors reported that the ability of absorption in CD patients was on average 30% lower than in normal subjects (*P* < 0.001)[83]. Based on most of the studies, it seems reasonable that measurement of serum vitamin D levels should be included in the follow up of IBD patients both adults and children. In fact, in pediatric IBD patients a vitamin D deficiency may enhance the odds of developing osteopenia or OP[84].

IBD patients might also have a reduced absorption of vitamin K, especially CD patients with an involvement of distal ileum. Besides playing a major role in coagulation processes vitamin K prevents bone resorption, inhibiting the production of prostaglandin E2 by osteoclasts, so its deficiency may affect BMD, as reported in both adults and pediatric patients[80]. However, the role of vitamin K in bone metabolism is controversial and the routine supplementation is not widely accepted yet[85].

**Glucocorticoid therapy:** A major risk factor associated to bone metabolism alterations in IBD patients is GC therapy, which in several occasions is administered without additional vitamin D or calcium supplementation[4].GCs are still largely used in IBD patients with moderate or severe disease[85]. GC therapy causes a biphasic bone loss, firstly with a rapid decrease of BMD of about 6% to 12% in the first year, and after, with an annual loss of about 3% for as long as the therapy is administered[87] associated with an increase of the fracture risk in the first 3 mo, which can reach a percentage as high as 75%. The fracture risk then decreases in the first 3 mo after GC withdrawal, before any significant improvement in BMD values[88]. GCs cause a reduction in the cortical thickness and an increased cortical porosity in mice models, associated with increased osteoclast number at the endocortical surface. Osteoclast formation in trabecular bone depends on the production of RANKL by osteocytes as well as by the increase in cortical bone resorption induced by mechanical unloading or by dietary calcium deficiency. *In vitro* models showed that GC therapy directly increases the production of RANKL and reduces OPG expression levels in stromal cells and osteoblasts[89]. Also, Hofbauer et al. reported that RANKL inhibition prevented GC-induced bone loss[90].

Piemontese *et al*[91] examined the effects of prednisolone on cortical bone in mice lacking RANKL production in osteocytes. Prednisolone increased osteoclast number at the endocortical surface, increased cortical porosity, and reduced cortical thickness in control mice, but none of these effects were found in mice lacking RANKL in osteocytes. Moreover, in cortical bone organ cultures and primary osteoblasts, dexamethasone suppressed OPG without any variation of RANKL levels. Therefore, based on these observations, OPG, rather than RANKL, seems to play a major role in the endocortical resorption.

OPG-mediated bone loss prevention acts through the inhibition of RANK-RANKL pathway and reduction of osteocytes apoptosis, induced by GCs. Weinstein et al., both *in vivo* and *in vitro*, studied the effect of OPG administration, with or without the fragment crystallizable region of Ig heavy chains (OPG-Fc), on the bone loss and on the apoptosis of osteocytes, with or without GCs administration. They showed that in mice treated with prednisolone combined with OPG-Fc or only with OPG-Fc there was a decreased expression of both receptors of cathepsin K and OC, which are markers of osteoclast number. Moreover, OPG-Fc administration preserved the BMD at spine compared with animals who received only prednisolone. The authors also reported an increase of vertebral strength of about 29% in mice receiving OPG-Fc compared to those receiving OPG-Fc combined with prednisone. Finally, OPG-Fc administration, alone or combined with prednisolone, decreased the number of osteoclasts of about 7% and 5% respectively, compared with placebo group. Prednisolone also induced an increase of the osteocytes apoptosis of about 335%, which was prevented by OPG-Fc administration. This supports the concept that OPG administration may prevent the reduction of BMD, of vertebral cortical thickness, and of osteocytes viability induced by GCs[41].

In conclusion, GC therapy is a major determinant of bone mass alteration in IBD patients. New generation GCs, such as budesonide or beclometasone, which show a very efficient hepatic first pass metabolism may represent a valid alternative to conventional GCs in order to try to minimize GC detrimental effect in general and on bone structure in particular[92,93].

DISEASE SEVERITY/ACTIVITY AND BONE MINERAL DENSITY

Calcium is an essential ion for bone formation and its only source is diet. The absorption of dietary calcium is a vitamin D-dependent process[94]. In the distal part of the intestine, 70%-80% of the ingested calcium is absorbed (mostly in the ileum)[95] through the action of Vitamin D receptor which is expressed in all segments of the small and large intestine with the highest levels reported in the cecum and colon. In patients with extensive intestinal resection, calcium absorption has been reported to be significantly higher when the colon is preserved[96]. In IBD, different segments of the intestine may be involved by the inflammatory process and the subsequent alterations of the absorptive processes have been suggested to be an important determinant of bone loss[97,98]. Several studies evaluated the correlation between disease extension or activity according to the Montreal classification for IBD[99] and BMD. In 1997, Bjarnason *et al*[29] reported that there were no significant differences in T scores for spine or hip within the patient subgroups according to disease location. Also, Jahnsen *et al*[30] assessed BMD in 60 patients with CD in 60 patients with UC and in 60 healthy subjects. Patients with CD had similar BMD, independently on whether the colon or the small intestine was involved. Furthermore, no differences were described between patients with CD with or without small bowel resection. In addition, no correlations between the length of small bowel resected and BMD were found. Finally, in the UC patients there was no influence of disease location and extension on MBD. More recently, the same results have been described by Vázquez *et al*[34] on 107 patients with IBDs (53 with CD and 54 with UC) with different location of the disease. The extension and the location of the disease did not seem to influence BMD loss or vertebral fractures prevalence. In a study conducted in 99 consecutive CD outpatients, Cravo *et al*[100] assessed disease activity by Harvey–Bradshaw Index (HBI). With a multivariate analysis, they described a direct and significant association between age (above 40 years), chronic active disease (HBI 4), previous colonic surgeries and the presence of OP. Both small bowel and colonic resection were similarly associated to OP. This might be explained by the major impact of inflammation due to disease severity in respect to the reduction of mineral absorption induced by short bowel syndrome, as initially hypothesized. Moreover, to support the role of chronic inflammation in the pathogenesis of bone loss, patients with active disease (HBI > 4) and those with a penetrating or structuring disease, which are usually more aggressive phenotypes, were also those with the highest rate of OP.

More recently, Lima *et al*[101] evaluated the correlation between disease severity and BMD in 68 patients with UC and 60 with CD of 17-40 years of age. About half CD patients had an ileocolonic disease (53.3%) while 29 subjects (48.3%) had non-stricturing non-penetrating disease and 33.3% had perianal disease, according to Montreal classification. In the UC group, 29 patients (44.6%) had extensive UC according to the Montreal classification[99]. The authors described a higher incidence of osteopenia in UC and CD patients than controls (OR = 14.93/OR = 24.38, respectively). At multivariate analysis in CD group, low BMD was associated with sex (M > F), perianal disease, penetrating behavior and age at diagnosis > 40 years, while, no association was described between BMD and disease activity. In the UC group, low BMD was significantly associated to sex (M > F) and left colitis. Therefore, disease activity does not seem to be a major determinant of bone density alteration in CD and UC patients, while disease severity seems to be associated with osteopenia in IBD patients. It must be emphasized that only 0.02% of UC patients and 0.26% of CD patients had active disease at the time DEXA was performed, and, therefore, the remission of the disease might have been be associated with an increase in BMD.

TNF-α/ANTI-TNF-α THERAPY AND BONE DENSITY ALTERATIONS

IBD is an immune-mediated inflammatory condition characterized by activation of different inflammatory pathways and abnormal secretion of different cytokines such as TNF-α[102].

Anti-TNF-α is the first available biologic therapy for IBD and, currently, its effects on BMD are not known. Moreover, it is not clear whether the effects of anti-TNFα agents on bone health are the consequence of a direct interference with the process of bone modeling or if these effects are simply due to a decreased disease activity and subsequent improvement of mineral absorption.

As previously mentioned, two members of TNF superfamily, RANKLOPG, are the key regulators of bone remodeling. RANKL, derived by osteoblasts, stimulates formation of mature osteoclasts while OPG, produced by osteoblasts, is a competitor that inhibits the interaction between RANKL and its receptor[103]. TNF-α is a main actor of osteoclastogenesis by inducing activation of NF-κ B transcription and, also, reducing bone formation through the inhibition of osteoblast differentiation[104]. Moreover, it increases the survival of osteoclasts by protecting them against apoptosis[105] while it induces apoptosis of osteoblasts to reduce bone formation[106]. Therefore, TNF-α not only plays a central role in the pathogenesis of IBD but is also involved in bone metabolism, promoting bone resorption through regulation of osteoclast activity (Figure 1). As described by Azuma *et al*[107], TNF-α directly induces the differentiation of osteoclast progenitors into mature osteoclast playing an important role in local osteolysis in chronic inflammatory diseases. Based on this, many studies have evaluated the effect of infliximab, a chimeric (*i.e.*, half human and half murine) anti-TNF agent, on bone metabolism investigating serum bone marker, BMD or incidence of bone fractures. Only one study assessed the impact of adalimumab, another all human anti-TNF agent, on bone metabolism. In a Belgian study[108], authors evaluated markers of bone formation and resorption at eight weeks from the beginning of infliximab therapy in comparison with healthy controls. In their cohort, regardless of patients’ clinical response, anti-TNF-α increased bone formation and, in the majority of patients, strongly decreased bone resorption. In a one year follow up study[109], after starting therapy, the mean BMD resulted to be increased significantly in CD patients without any correlation with concurrent corticosteroid therapy. So, the authors suggested that amongst the factors inducing bone loss in CD, the inflammatory disease process might be predominant over the effects of treatment with prednisone. This might be especially relevant in CD patients who need to continue steroid therapy despite concurrent immunomodulatory therapy.

In 24 patients with active CD treated with infliximab, Ryan *et al*[110] described a significant increase of bone alkaline phosphatase, a marker of bone formation, and OC, a bone specific calcium-binding protein produced by osteoblasts, which persisted up to 4 wk after the end of treatment. As underlined by other authors, the benefits occurred independently of the clinical response of CD to biological treatment. On the other side, Miheller *et al*[111] dosed serum OC and CrossLaps (bCL), a degradation product of collagen, in 27 patients with fistulizing CD treated with anti-TNF-α. In the group of patients who responded to therapy, but not in those who did not, serum bCL concentrations were significantly decreased from week 0 to week 6, while a statistically significant increase was described for OC, thus suggesting that the beneficial effect of anti-TNF therapy was related to the amelioration of the underlying inflammatory process.

More recently, a 7-year follow-up longitudinal prospective cohort study by Maldonado-Pérez et al[112] evaluated the role of anti-TNF-α in decreasing fracture risk or modifying BMD in IBD patients. The authors described no difference in the incidence of vertebral fracture and value of bone mass between the group of patients treated with anti-TNF-α and the control group which did not receive biological treatment. Despite the biological-treated patients had received GC therapy for a longer period of time compared to the control group, new fractures were more common and more severe in the control, nonbiological-treated group. After 7 years of follow-up, bone mass increased significantly in the spine and in the femoral neck in patients treated with anti-TNF-α, compared to subjects who did not receive biological therapy.

Only one study[113] evaluated the impact of adalimumab therapy on bone metabolism. Parathyroid hormone, vitamin D, bone formation and resorption marker, pro- and anti-inflammatory OPG, and sRANKL were measured in healthy controls and in CD patients pre- and post-treatment with adalimumab. Moreover, viability and differentiation of human osteoblasts (hFOB 1.19) cells after exposure to sera from CD patients pre- and post-adalimumab treatment was also analyzed. Following adalimumab therapy, a rapid increase in bone formation markers (OC and procollagen type 1 N-terminal pro-peptide) and a not significant decrease of a bone resorption marker (C-telopeptide of type-1 collagen) were observed. In the *in vitro* study, osteoblasts exposed to sera of CD patients before adalimumab therapy showed consistently higher levels of viability and lower levels of ALP compared to control group suggesting a greater viability of osteoblasts associated to a lower osteoblast function likely due to an inflammatory-driven response. After treatment, serum of CD patients induced higher levels of ALP in hFOB cells probably due to an improvement of their functionality.

In conclusion, anti-TNF seems to improve BMD in IBD patients both through a direct beneficial effect on bone metabolism and through the improvement in the underlying intestinal inflammatory process. Whether other biologic agents now available for the treatment of IBD, such as vedolizumab or Uutekinumab, have any effect on bone metabolism needs to be determined.

DIAGNOSIS OF OSTEOPOROSIS

Diagnosis of OP should be based on patient clinical history, physical examination, BMD measurements, and laboratory investigations[17]. In particular, because changes in bone metabolism are frequently associated with the evolution of IBD and may have a negative impact on the patient’s quality of life, assessment of BMD in all IBD patients is essential to prevent and treat appropriately MBDs. The gold standard for its assessment is DXA. Changes in BMD values are key determinants to evaluate treatment efficacy at follow-up. Moreover, FRAX algorithm by combining all the fracture risks with the hip BMD value can quantify the 10-year risk of experiencing a fragility fracture[114]. Laboratory tests are necessary not only to exclude secondary forms of OP but also for the bone metabolism assessment. They should include biochemical markers of bone turnover and vitamin D status that might provide additional information regarding the patient fracture risk.

Moreover, increase in the incidence of BMD loss supports the recommendation to screen patients with IBD at an early stage of the disease. Screening recommendation of European Crohn and Colitis Organization (ECCO)[115] does not differ from those for the general population. It considers risk factors such as postmenopausal state, ongoing corticosteroid treatment, cumulative corticosteroid use > 3 mo, history of low-trauma fracture and age. Moreover, annual DXA scans is recommended in patients receiving long-term steroid therapy (in particular when there are others risk factors) if the T-score approaches the threshold for treatment with bisphosphonates (BPs) (T-score < −1.5 SD)[116].

THE PHARMACOLOGICAL MANAGEMENT OF OP

The aim of the management of OP is to reduce the risk of fragility fractures in individuals at high-risk. Therefore, pharmacological intervention thresholds should be based on the assessment of this risk deriving from the integration of densitometric data with other important clinical factors as determined by the FRAX[116]. Approved pharmacological treatments for the management of OP can be classified into two categories: anti-resorption (or anti-catabolic) and anabolic drugs. Among anti-catabolic drugs, biphosphonates (BPs) blocking the osteoclastic activity, manage to reduce the bone remodeling process with a consequent increase in bone density. Alendronate and risedronate are the most commonly used for the prevention of vertebral and non-vertebral fractures (including hip) based on strong scientific evidence of efficacy[117]. However, they have reduced compliance and persistence to prolonged therapy, due to daily or weekly administration regimens and possible gastro-intestinal adverse events. Zoledronic acid is a BP intravenously administered with documented efficacy in reducing the risk of vertebral, non-vertebral and hip fractures[117]. A meta-analysis of studies on BPs use in IBD patients showed that these drugs are effective in case of low BMD reducing the risk of vertebral but not of non-vertebral fracture[118]; so, the use of BPs should be recommended for fracture prevention in IBD patients taking always into account the possible adverse effects of treatment.

A powerful inhibitor of bone resorption is denosumab, a human monoclonal antibody capable of neutralizing RANKL, a cytokine that interacts with the RANK receptor on the membrane of preosteoclasts and mature osteoclasts, affecting their recruitment, maturation and survival. A dose of 60 mg subcutaneously every 6 months is sufficient to strongly inhibit osteoclastic activity and to reduce the risk of vertebral and non-vertebral fractures (including hip). Unlike BPs, discontinuation of denosumab is followed by a sharp increase in bone turnover and a rapid loss of BMD. Therefore, discontinuation of denosumab generally requires the patient to initiate BPs treatment at an appropriate dosage as soon as possible[119].

Among anabolic drugs, teriparatide, the active fragment of PTH (1-34 PTH) is the most widely used. It can stimulate both bone formation and resorption, with a predominant effect on the neoformation (anabolic window) which is evident above all during the first 12 mo of treatment. It is generally used as a second line anti-osteoporotic drug in case of intolerance or resistance to other anti-resorption agents and as first choice in case of severe OP in patients with multiple fragility fractures[17].

All clinical guidelines agree that the pharmacological therapy of OP, independently of the prescribed treatment, should always be supplemented by the administration of vitamin D and, in case of nutritional deficient intake, of calcium[120].

ECCO guidelines[115] suggest, some recommendations concerning the management of bone alterations in IBD population (Table 1).

CONCLUSION

The prevalence of OP and/or fragility fractures in IBD patients is controversial because of different factors, such as different study population and study design, and location of the disease. Moreover, some aspects still need to be clarified, particularly the correlation between the increased risk of fragility fractures in subjects affected by IBD.

Changes in bone metabolism are frequently associated with the evolution of IBD and may have a negative impact on the patient’s quality of life.

In this context, even understanding the pathophysiological milieu seems to be quite challenging. For example, the genetic background predisposing to the development of osteopenia and OP, specific for IBD patients, since the strongly multifactorial nature of these diseases, does not allow to evaluate its pathogenic role without considering other factors, such as nutrition, lifestyle, or more simply, pharmacological therapy for IBD. On the other side, increasing evidence suggests a gut-bone signaling pathway, which is responsible for a close cross-talk between the musculoskeletal and the GI system, and whose alteration may potentially correlate with the evolution of this type of EIMs. Furthermore, because of the emerging role of intestinal microbiota in the pathogenesis of IBDs, a direct impact of dysbiotic commensal microflora on bone metabolism, as shown in the healthy population, seems to be possible. A schematic diagram summarizing the pathophysiologic mechanism, including molecular mechanisms, underlying BMD alteration in IBD patients is shown in Figure 3.

The nutritional aspects, always considered among the main factors capable of triggering bone alterations, appear once again crucial, especially regarding the intake of calcium and vitamin D, the lack of which, both in adults and children, shows a direct correlation with the increased probability of developing bone fragility, specifically linked to osteoporomalacic findings. For this reason, it is important to include the evaluation of serum vitamin D levels and of nutritional status in IBD patients, both in active phase and remission of disease, in order to avoid the establishment of malnutrition that may increase the onset of comorbidity.

Pharmacotherapy of IBD might play a major role in bone metabolism. GCs are the main determinants of bone alterations and their prolonged use is associated with OP, osteopenia and increased risk of fractures. Therefore, their use should be limited and, whenever possible, new generation corticosteroids with a safer profile should be used. Finally, anti-TNF agents seem to improve bone health in IBD patients both by directly interfering with the metabolic pathways involved in bone modeling and by decreasing the disease activity and severity. Whether new biologic agents exert any beneficial effect on bone tissue in IBD patients remains to be determined.

Finally, and more important, a thorough evaluation of bone metabolism including serological markers should be part of the follow-up of IBD patients in order to prevent and/or promptly treat any bone alteration which may alter their quality of life and increase the risk of fractures.

**REFERENCES**

1 **Ferreira PVALS**, Cavalcanti AS, Silva GAPD. Linear growth and bone metabolism in pediatric patients with inflammatory bowel disease. *J Pediatr (Rio J)* 2019; **95 Suppl 1**: 59-65 [PMID: 30562479 DOI: 10.1016/j.jped.2018.11.002]

2 **Sheth T**, Pitchumoni CS, Das KM. Musculoskeletal manifestations in inflammatory bowel disease: a revisit in search of immunopathophysiological mechanisms. *J Clin Gastroenterol* 2014; **48**: 308-317 [PMID: 24492406 DOI: 10.1097/MCG.0000000000000067]

3 **Vavricka SR**, Rogler G, Gantenbein C, Spoerri M, Prinz Vavricka M, Navarini AA, French LE, Safroneeva E, Fournier N, Straumann A, Froehlich F, Fried M, Michetti P, Seibold F, Lakatos PL, Peyrin-Biroulet L, Schoepfer AM. Chronological Order of Appearance of Extraintestinal Manifestations Relative to the Time of IBD Diagnosis in the Swiss Inflammatory Bowel Disease Cohort. *Inflamm Bowel Dis* 2015; **21**: 1794-1800 [PMID: 26020601 DOI: 10.1097/MIB.0000000000000429]

4 **Schüle S**, Rossel JB, Frey D, Biedermann L, Scharl M, Zeitz J, Freitas-Queiroz N, Kuntzen T, Greuter T, Vavricka SR, Rogler G, Misselwitz B; Swiss IBD cohort study. Widely differing screening and treatment practice for osteoporosis in patients with inflammatory bowel diseases in the Swiss IBD cohort study. *Medicine (Baltimore)* 2017; **96**: e6788 [PMID: 28562531 DOI: 10.1097/MD.0000000000006788]

5 Consensus development conference: diagnosis, prophylaxis, and treatment of osteoporosis. *Am J Med* 1993; **94**: 646-650 [PMID: 8506892 DOI: 10.1016/0002-9343(93)90218-E]

6 **Kanis JA**, McCloskey EV, Johansson H, Oden A, Melton LJ 3rd, Khaltaev N. A reference standard for the description of osteoporosis. *Bone* 2008; **42**: 467-475 [PMID: 18180210 DOI: 10.1016/j.bone.2007.11.001]

7 World Health Organization – WHO Criteria for Diagnosis of Osteoporosis [Internet]. 4BoneHealth 2018 Available from: http://www.4bonehealth.org/education/world-health-organization-criteria-diagnosis-osteoporosis/

8 **Baim S**, Binkley N, Bilezikian JP, Kendler DL, Hans DB, Lewiecki EM, Silverman S. Official Positions of the International Society for Clinical Densitometry and executive summary of the 2007 ISCD Position Development Conference. *J Clin Densitom* 2008; **11**: 75-91 [PMID: 18442754 DOI: 10.1016/j.jocd.2007.12.007]

9 **Maggi S**, Noale M, Giannini S, Adami S, Defeo D, Isaia G, Sinigaglia L, Filipponi P, Crepaldi G; ESOPO Study Group. Quantitative heel ultrasound in a population-based study in Italy and its relationship with fracture history: the ESOPO study. *Osteoporos Int* 2006; **17**: 237-244 [PMID: 16142503 DOI: 10.1007/s00198-005-1985-2]

10 **World Health Organization (WHO)**. Guidelines for preclinical evaluation and clin-ical trials in osteoporosis [Internet]. 1998 Available from: http://www.who.int/iris/handle/10665/42088

11 **Tarantino U**, Iolascon G, Cianferotti L, Masi L, Marcucci G, Giusti F, Marini F, Parri S, Feola M, Rao C, Piccirilli E, Zanetti EB, Cittadini N, Alvaro R, Moretti A, Calafiore D, Toro G, Gimigliano F, Resmini G, Brandi ML. Clinical guidelines for the prevention and treatment of osteoporosis: summary statements and recommendations from the Italian Society for Orthopaedics and Traumatology. *J Orthop Traumatol* 2017; **18**: 3-36 [PMID: 29058226 DOI: 10.1007/s10195-017-0474-7]

12 **Khosla S**. Update in male osteoporosis. *J Clin Endocrinol Metab* 2010; **95**: 3-10 [PMID: 20056806 DOI: 10.1210/jc.2009-1740]

13 **Johnell O**, Kanis JA. An estimate of the worldwide prevalence and disability associated with osteoporotic fractures. *Osteoporos Int* 2006; **17**: 1726-1733 [PMID: 16983459 DOI: 10.1007/s00198-006-0172-4]

14 **Cooper C**, Cole ZA, Holroyd CR, Earl SC, Harvey NC, Dennison EM, Melton LJ, Cummings SR, Kanis JA; IOF CSA Working Group on Fracture Epidemiology. Secular trends in the incidence of hip and other osteoporotic fractures. *Osteoporos Int* 2011; **22**: 1277-1288 [PMID: 21461721 DOI: 10.1007/s00198-011-1601-6]

15 **Svedbom A**, Hernlund E, Ivergård M, Compston J, Cooper C, Stenmark J, McCloskey EV, Jönsson B, Kanis JA; EU Review Panel of IOF. Osteoporosis in the European Union: a compendium of country-specific reports. *Arch Osteoporos* 2013; **8**: 137 [PMID: 24113838 DOI: 10.1007/s11657-013-0137-0]

16 **Piscitelli P**, Iolascon G, Argentiero A, Chitano G, Neglia C, Marcucci G, Pulimeno M, Benvenuto M, Mundi S, Marzo V, Donati D, Baggiani A, Migliore A, Granata M, Gimigliano F, Di Blasio R, Gimigliano A, Renzulli L, Brandi ML, Distante A, Gimigliano R. Incidence and costs of hip fractures vs strokes and acute myocardial infarction in Italy: comparative analysis based on national hospitalization records. *Clin Interv Aging* 2012; **7**: 575-583 [PMID: 23269863 DOI: 10.2147/CIA.S36828]

17 **Ali T**, Lam D, Bronze MS, Humphrey MB. Osteoporosis in inflammatory bowel disease. *Am J Med* 2009; **122**: 599-604 [PMID: 19559158 DOI: 10.1016/j.amjmed.2009.01.022]

18 **Boubaker J**, Feki M, Hsairi M, Fekih M, Kaabachi N, Filali A, Mebazaa A. [Osteoporosis and inflammatory bowel disease: prevalence and risk factors in Tunisian patients]. *Gastroenterol Clin Biol* 2003; **27**: 901-907 [PMID: 14631305]

19 **Dumitrescu G**, Mihai C, Dranga M, Prelipcean CC. Bone mineral density in patients with inflammatory bowel disease from north-eastern Romania. *Rev Med Chir Soc Med Nat Iasi* 2013; **117**: 23-28 [PMID: 24505888]

20 **Naito T**, Yokoyama N, Kakuta Y, Ueno K, Kawai Y, Onodera M, Moroi R, Kuroha M, Kanazawa Y, Kimura T, Shiga H, Endo K, Nagasaki M, Masamune A, Kinouchi Y, Shimosegawa T. Clinical and genetic risk factors for decreased bone mineral density in Japanese patients with inflammatory bowel disease. *J Gastroenterol Hepatol* 2018; **33**: 1873-1881 [PMID: 29603369 DOI: 10.1111/jgh.14149]

21 **Targownik LE**, Bernstein CN, Leslie WD. Inflammatory bowel disease and the risk of osteoporosis and fracture. *Maturitas* 2013; **76**: 315-319 [PMID: 24139749 DOI: 10.1016/j.maturitas.2013.09.009]

22 **Komaki Y**, Komaki F, Micic D, Ido A, Sakuraba A. Risk of Fractures in Inflammatory Bowel Diseases: A Systematic Review and Meta-Analysis. *J Clin Gastroenterol* 2019; **53**: 441-448 [PMID: 29672437 DOI: 10.1097/MCG.0000000000001031]

23 **Lopes LH**, Sdepanian VL, Szejnfeld VL, de Morais MB, Fagundes-Neto U. Risk factors for low bone mineral density in children and adolescents with inflammatory bowel disease. *Dig Dis Sci* 2008; **53**: 2746-2753 [PMID: 18351466 DOI: 10.1007/s10620-008-0223-0]

24 **Bryant RV**, Schultz CG, Ooi S, Goess C, Costello SP, Vincent AD, Schoeman SN, Lim A, Bartholomeusz FD, Travis SPL, Andrews JM. Obesity in Inflammatory Bowel Disease: Gains in Adiposity despite High Prevalence of Myopenia and Osteopenia. *Nutrients* 2018; **10** [PMID: 30200405 DOI: 10.3390/nu10091192]

25 **Nobile S**, Grand RJ, Pappa HM. Risk factors for low bone mineral density in pediatric inflammatory bowel disease: the positive role of physical activity. *Eur J Gastroenterol Hepatol* 2018; **30**: 471-476 [PMID: 29438136 DOI: 10.1097/MEG.0000000000001076]

26 **Ward LM**, Ma J, Rauch F, Benchimol EI, Hay J, Leonard MB, Matzinger MA, Shenouda N, Lentle B, Cosgrove H, Scharke M, Konji VN, Mack DR. Musculoskeletal health in newly diagnosed children with Crohn's disease. *Osteoporos Int* 2017; **28**: 3169-3177 [PMID: 28791436 DOI: 10.1007/s00198-017-4159-0]

27 **Huber AM**, Gaboury I, Cabral DA, Lang B, Ni A, Stephure D, Taback S, Dent P, Ellsworth J, LeBlanc C, Saint-Cyr C, Scuccimarri R, Hay J, Lentle B, Matzinger M, Shenouda N, Moher D, Rauch F, Siminoski K, Ward LM; Canadian Steroid-Associated Osteoporosis in the Pediatric Population (STOPP) Consortium. Prevalent vertebral fractures among children initiating glucocorticoid therapy for the treatment of rheumatic disorders. *Arthritis Care Res (Hoboken)* 2010; **62**: 516-526 [PMID: 20391507 DOI: 10.1002/acr.20171]

28 **Sylvester FA**, Gordon CM, Thayu M, Burnham JM, Denson LA, Essers J, Ferrari S, Gupta N, Hewison M, Koletzko S, McCabe L, Pappa H, Sanderson I, Ward L, Zanotti S. Report of the CCFA pediatric bone, growth and muscle health workshop, New York City, November 11-12, 2011, with updates. *Inflamm Bowel Dis* 2013; **19**: 2919-2926 [PMID: 23974992 DOI: 10.1097/MIB.0b013e3182a5a004]

29 **Bjarnason I**, Macpherson A, Mackintosh C, Buxton-Thomas M, Forgacs I, Moniz C. Reduced bone density in patients with inflammatory bowel disease. *Gut* 1997; **40**: 228-233 [PMID: 9071937 DOI: 10.1136/gut.40.2.228]

30 **Jahnsen J**, Falch JA, Aadland E, Mowinckel P. Bone mineral density is reduced in patients with Crohn's disease but not in patients with ulcerative colitis: a population based study. *Gut* 1997; **40**: 313-319 [PMID: 9135518 DOI: 10.1136/gut.40.3.313]

31 **Best WR**, Becktel JM, Singleton JW, Kern F Jr. Development of a Crohn's disease activity index. National Cooperative Crohn's Disease Study. *Gastroenterology* 1976; **70**: 439-444 [PMID: 1248701 DOI: 10.1016/S0016-5085(76)80163-1]

32 **TRUELOVE SC**, WITTS LJ. Cortisone in ulcerative colitis; final report on a therapeutic trial. *Br Med J* 1955; **2**: 1041-1048 [PMID: 13260656 DOI: 10.1136/bmj.2.4947.1041]

33 **de Silva AP**, Karunanayake AL, Dissanayaka TG, Dassanayake AS, Duminda HK, Pathmeswaran A, Wickramasinghe AR, de Silva HJ. Osteoporosis in adult Sri Lankan inflammatory bowel disease patients. *World J Gastroenterol* 2009; **15**: 3528-3531 [PMID: 19630109 DOI: 10.3748/wjg.15.3528]

34 **Vázquez MA**, Lopez E, Montoya MJ, Giner M, Pérez-Temprano R, Pérez-Cano R. Vertebral fractures in patients with inflammatory bowel disease compared with a healthy population: a prospective case-control study. *BMC Gastroenterol* 2012; **12**: 47 [PMID: 22584049 DOI: 10.1186/1471-230X-12-47]

35 **Lacey DL**, Boyle WJ, Simonet WS, Kostenuik PJ, Dougall WC, Sullivan JK, San Martin J, Dansey R. Bench to bedside: elucidation of the OPG-RANK-RANKL pathway and the development of denosumab. *Nat Rev Drug Discov* 2012; **11**: 401-419 [PMID: 22543469 DOI: 10.1038/nrd3705]

36 **Moester MJ**, Papapoulos SE, Löwik CW, van Bezooijen RL. Sclerostin: current knowledge and future perspectives. *Calcif Tissue Int* 2010; **87**: 99-107 [PMID: 20473488 DOI: 10.1007/s00223-010-9372-1]

37 **Robling AG**, Niziolek PJ, Baldridge LA, Condon KW, Allen MR, Alam I, Mantila SM, Gluhak-Heinrich J, Bellido TM, Harris SE, Turner CH. Mechanical stimulation of bone in vivo reduces osteocyte expression of Sost/sclerostin. *J Biol Chem* 2008; **283**: 5866-5875 [PMID: 18089564 DOI: 10.1074/jbc.M705092200]

38 **Gaudio A**, Pennisi P, Bratengeier C, Torrisi V, Lindner B, Mangiafico RA, Pulvirenti I, Hawa G, Tringali G, Fiore CE. Increased sclerostin serum levels associated with bone formation and resorption markers in patients with immobilization-induced bone loss. *J Clin Endocrinol Metab* 2010; **95**: 2248-2253 [PMID: 20305005 DOI: 10.1210/jc.2010-0067]

39 **Gifre L**, Ruiz-Gaspà S, Monegal A, Nomdedeu B, Filella X, Guañabens N, Peris P. Effect of glucocorticoid treatment on Wnt signalling antagonists (sclerostin and Dkk-1) and their relationship with bone turnover. *Bone* 2013; **57**: 272-276 [PMID: 23981659 DOI: 10.1016/j.bone.2013.08.016]

40 **Humphrey EL**, Williams JH, Davie MW, Marshall MJ. Effects of dissociated glucocorticoids on OPG and RANKL in osteoblastic cells. *Bone* 2006; **38**: 652-661 [PMID: 16298558 DOI: 10.1016/j.bone.2005.10.004]

41 **Weinstein RS**. Clinical practice. Glucocorticoid-induced bone disease. *N Engl J Med* 2011; **365**: 62-70 [PMID: 21732837 DOI: 10.1056/NEJMcp1012926]

42 **Briot K**, Geusens P, Em Bultink I, Lems WF, Roux C. Inflammatory diseases and bone fragility. *Osteoporos Int* 2017; **28**: 3301-3314 [PMID: 28916915 DOI: 10.1007/s00198-017-4189-7]

43 **Ghosh S**, Cowen S, Hannan WJ, Ferguson A. Low bone mineral density in Crohn's disease, but not in ulcerative colitis, at diagnosis. *Gastroenterology* 1994; **107**: 1031-1039 [PMID: 7926456 DOI: 10.1016/0016-5085(94)90227-5]

44 **McCabe LR,** Parameswaran N. Understanding the gut-bone Signaling Axis: Mechanism and therapeutics implications. 2017 [DOI: 10.1007/978-3-319-66653-2]

45 **Rossini M**, Adami S, Bertoldo F, Diacinti D, Gatti D, Giannini S, Giusti A, Malavolta N, Minisola S, Osella G, Pedrazzoni M, Sinigaglia L, Viapiana O, Isaia GC. Guidelines for the diagnosis, prevention and management of osteoporosis. *Reumatismo* 2016; **68**: 1-39 [PMID: 27339372 DOI: 10.4081/reumatismo.2016.870]

46 **Van Assche G**, Dignass A, Reinisch W, van der Woude CJ, Sturm A, De Vos M, Guslandi M, Oldenburg B, Dotan I, Marteau P, Ardizzone A, Baumgart DC, D'Haens G, Gionchetti P, Portela F, Vucelic B, Söderholm J, Escher J, Koletzko S, Kolho KL, Lukas M, Mottet C, Tilg H, Vermeire S, Carbonnel F, Cole A, Novacek G, Reinshagen M, Tsianos E, Herrlinger K, Oldenburg B, Bouhnik Y, Kiesslich R, Stange E, Travis S, Lindsay J; European Crohn's and Colitis Organisation (ECCO). The second European evidence-based Consensus on the diagnosis and management of Crohn's disease: Special situations. *J Crohns Colitis* 2010; **4**: 63-101 [PMID: 21122490 DOI: 10.1016/j.crohns.2009.09.009]

47 **Azzopardi N**, Ellul P. Risk factors for osteoporosis in Crohn's disease: infliximab, corticosteroids, body mass index, and age of onset. *Inflamm Bowel Dis* 2013; **19**: 1173-1178 [PMID: 23511037 DOI: 10.1097/MIB.0b013e31828075a7]

48 **Jahnsen J**, Falch JA, Mowinckel P, Aadland E. Bone mineral density in patients with inflammatory bowel disease: a population-based prospective two-year follow-up study. *Scand J Gastroenterol* 2004; **39**: 145-153 [PMID: 15000276 DOI: 10.1080/00365520310007873]

49 **Bernstein CN**, Leslie WD, Leboff MS. AGA technical review on osteoporosis in gastrointestinal diseases. *Gastroenterology* 2003; **124**: 795-841 [PMID: 12612917 DOI: 10.1053/gast.2003.50106]

50 **Veerappan SG**, O'Morain CA, Daly JS, Ryan BM. Review article: the effects of antitumour necrosis factor-α on bone metabolism in inflammatory bowel disease. *Aliment Pharmacol Ther* 2011; **33**: 1261-1272 [PMID: 21521250 DOI: 10.1111/j.1365-2036.2011.04667.x]

51 **Yang BR**, Choi NK, Kim MS, Chun J, Joo SH, Kim H, Lee J. Prevalence of extraintestinal manifestations in Korean inflammatory bowel disease patients. *PLoS One* 2018; **13**: e0200363 [PMID: 29990326 DOI: 10.1371/journal.pone.0200363]

52 **Von Tirpitz C**, Pischulti G, Klaus J, Rieber A, Brückel J, Böhm BO, Adler G, Reinshagen M. [Pathological bone density in chronic inflammatory bowel diseases--prevalence and risk factors]. *Z Gastroenterol* 1999; **37**: 5-12 [PMID: 10091278]

53 **Schulte CM**, Dignass AU, Goebell H, Röher HD, Schulte KM. Genetic factors determine extent of bone loss in inflammatory bowel disease. *Gastroenterology* 2000; **119**: 909-920 [PMID: 11040178 DOI: 10.1053/gast.2000.18158]

54 **Todhunter CE**, Sutherland-Craggs A, Bartram SA, Donaldson PT, Daly AK, Francis RM, Mansfield JC, Thompson NP. Influence of IL-6, COL1A1, and VDR gene polymorphisms on bone mineral density in Crohn's disease. *Gut* 2005; **54**: 1579-1584 [PMID: 16009674 DOI: 10.1136/gut.2005.064212]

55 **Nemetz A**, Tóth M, García-González MA, Zágoni T, Fehér J, Peña AS, Tulassay Z. Allelic variation at the interleukin 1beta gene is associated with decreased bone mass in patients with inflammatory bowel diseases. *Gut* 2001; **49**: 644-649 [PMID: 11600466 DOI: 10.1136/gut.49.5.644]

56 **Krela-Kaźmierczak I**, Kaczmarek-Ryś M, Szymczak A, Michalak M, Skrzypczak-Zielińska M, Drwęska-Matelska N, Marcinkowska M, Eder P, Łykowska-Szuber L, Wysocka E, Linke K, Słomski R. Bone Metabolism and the c.-223C &gt; T Polymorphism in the 5'UTR Region of the Osteoprotegerin Gene in Patients with Inflammatory Bowel Disease. *Calcif Tissue Int* 2016; **99**: 616-624 [PMID: 27639566 DOI: 10.1007/s00223-016-0192-9]

57 **Krela-Kaźmierczak I**, Michalak M, Wawrzyniak A, Szymczak A, Eder P, Łykowska-Szuber L, Kaczmarek-Ryś M, Drwęska-Matelska N, Skrzypczak-Zielińska M, Linke K, Słomski R. The c.29T&gt;C polymorphism of the transforming growth factor beta-1 (TGFB1) gene, bone mineral density and the occurrence of low-energy fractures in patients with inflammatory bowel disease. *Mol Biol Rep* 2017; **44**: 455-461 [PMID: 28993955 DOI: 10.1007/s11033-017-4131-2]

58 **Krela-Kazmierczak I**, Wawrzyniak A, Szymczak A, Eder P, Lykowska-Szuber L, Michalak M, Drweska-Matelska N, Kaczmarek-Rys M, Skrzypczak-Zielinska M, Szalata M, Slomski R. Bone mineral density and the 570A&gt;T polymorphism of the bone morphogenetic protein 2 (BMP2) gene in patients with inflammatory bowel disease: a cross-sectional study. *J Physiol Pharmacol* 2017; **68**: 757-764 [PMID: 29375051]

59 **Cleynen I**, Boucher G, Jostins L, Schumm LP, Zeissig S, Ahmad T, Andersen V, Andrews JM, Annese V, Brand S, Brant SR, Cho JH, Daly MJ, Dubinsky M, Duerr RH, Ferguson LR, Franke A, Gearry RB, Goyette P, Hakonarson H, Halfvarson J, Hov JR, Huang H, Kennedy NA, Kupcinskas L, Lawrance IC, Lee JC, Satsangi J, Schreiber S, Théâtre E, van der Meulen-de Jong AE, Weersma RK, Wilson DC; International Inflammatory Bowel Disease Genetics Consortium, Parkes M, Vermeire S, Rioux JD, Mansfield J, Silverberg MS, Radford-Smith G, McGovern DP, Barrett JC, Lees CW. Inherited determinants of Crohn's disease and ulcerative colitis phenotypes: a genetic association study. *Lancet* 2016; **387**: 156-167 [PMID: 26490195 DOI: 10.1016/S0140-6736(15)00465-1]

60 **Wu Y**, Yang M, Fan J, Peng Y, Deng L, Ding Y, Yang R, Zhou J, Miao D, Fu Q. Deficiency of osteoblastic Arl6ip5 impaired osteoblast differentiation and enhanced osteoclastogenesis via disturbance of ER calcium homeostasis and induction of ER stress-mediated apoptosis. *Cell Death Dis* 2014; **5**: e1464 [PMID: 25321471 DOI: 10.1038/cddis.2014.427]

61 **Saito A**, Ochiai K, Kondo S, Tsumagari K, Murakami T, Cavener DR, Imaizumi K. Endoplasmic reticulum stress response mediated by the PERK-eIF2(alpha)-ATF4 pathway is involved in osteoblast differentiation induced by BMP2. *J Biol Chem* 2011; **286**: 4809-4818 [PMID: 21135100 DOI: 10.1074/jbc.M110.152900]

62 **Tohmonda T**, Yoda M, Mizuochi H, Morioka H, Matsumoto M, Urano F, Toyama Y, Horiuchi K. The IRE1α-XBP1 pathway positively regulates parathyroid hormone (PTH)/PTH-related peptide receptor expression and is involved in pth-induced osteoclastogenesis. *J Biol Chem* 2013; **288**: 1691-1695 [PMID: 23235147 DOI: 10.1074/jbc.C112.424606]

63 **Arron JR**, Choi Y. Bone versus immune system. *Nature* 2000; **408**: 535-536 [PMID: 11117729 DOI: 10.1038/35046196]

64 **Ciucci T**, Ibáñez L, Boucoiran A, Birgy-Barelli E, Pène J, Abou-Ezzi G, Arab N, Rouleau M, Hébuterne X, Yssel H, Blin-Wakkach C, Wakkach A. Bone marrow Th17 TNFα cells induce osteoclast differentiation, and link bone destruction to IBD. *Gut* 2015; **64**: 1072-1081 [PMID: 25298539 DOI: 10.1136/gutjnl-2014-306947]

65 **Ashcroft AJ**, Cruickshank SM, Croucher PI, Perry MJ, Rollinson S, Lippitt JM, Child JA, Dunstan C, Felsburg PJ, Morgan GJ, Carding SR. Colonic dendritic cells, intestinal inflammation, and T cell-mediated bone destruction are modulated by recombinant osteoprotegerin. *Immunity* 2003; **19**: 849-861 [PMID: 14670302 DOI: 10.1016/S1074-7613(03)00326-1]

66 **Li Y**, Toraldo G, Li A, Yang X, Zhang H, Qian WP, Weitzmann MN. B cells and T cells are critical for the preservation of bone homeostasis and attainment of peak bone mass in vivo. *Blood* 2007; **109**: 3839-3848 [PMID: 17202317 DOI: 10.1182/blood-2006-07-037994]

67 **Nakashima T**, Takayanagi H. Osteoimmunology: crosstalk between the immune and bone systems. *J Clin Immunol* 2009; **29**: 555-567 [PMID: 19585227 DOI: 10.1007/s10875-009-9316-6]

68 **Shih TC**, Hsieh SY, Hsieh YY, Chen TC, Yeh CY, Lin CJ, Lin DY, Chiu CT. Aberrant activation of nuclear factor of activated T cell 2 in lamina propria mononuclear cells in ulcerative colitis. *World J Gastroenterol* 2008; **14**: 1759-1767 [PMID: 18350607 DOI: 10.3748/wjg.14.1759]

69 **Fukuda S**, Ohno H. Gut microbiome and metabolic diseases. *Semin Immunopathol* 2014; **36**: 103-114 [PMID: 24196453 DOI: 10.1007/s00281-013-0399-z]

70 **Tomasello G**, Tralongo P, Damiani P, Sinagra E, Di Trapani B, Zeenny MN, Hussein IH, Jurjus A, Leone A. Dismicrobism in inflammatory bowel disease and colorectal cancer: changes in response of colocytes. *World J Gastroenterol* 2014; **20**: 18121-18130 [PMID: 25561781 DOI: 10.3748/wjg.v20.i48.18121]

71 **Irwin R**, Lee T, Young VB, Parameswaran N, McCabe LR. Colitis-induced bone loss is gender dependent and associated with increased inflammation. *Inflamm Bowel Dis* 2013; **19**: 1586-1597 [PMID: 23702805 DOI: 10.1097/MIB.0b013e318289e17b]

72 **McCabe LR**, Irwin R, Schaefer L, Britton RA. Probiotic use decreases intestinal inflammation and increases bone density in healthy male but not female mice. *J Cell Physiol* 2013; **228**: 1793-1798 [PMID: 23389860 DOI: 10.1002/jcp.24340]

73 **Schepper JD**, Collins FL, Rios-Arce ND, Raehtz S, Schaefer L, Gardinier JD, Britton RA, Parameswaran N, McCabe LR. Probiotic Lactobacillus reuteri Prevents Postantibiotic Bone Loss by Reducing Intestinal Dysbiosis and Preventing Barrier Disruption. *J Bone Miner Res* 2019; **34**: 681-698 [PMID: 30690795 DOI: 10.1002/jbmr.3635]

74 **Naser A**, Qasem A, Naser SA. Mycobacterial infection influences bone biomarker levels in patients with Crohn's disease. *Can J Physiol Pharmacol* 2018; **96**: 662-667 [PMID: 29638140 DOI: 10.1139/cjpp-2017-0700]

75 **Lim H**, Kim HJ, Hong SJ, Kim S. Nutrient intake and bone mineral density by nutritional status in patients with inflammatory bowel disease. *J Bone Metab* 2014; **21**: 195-203 [PMID: 25247157 DOI: 10.11005/jbm.2014.21.3.195]

76 **Leslie WD**, Miller N, Rogala L, Bernstein CN. Body mass and composition affect bone density in recently diagnosed inflammatory bowel disease: the Manitoba IBD Cohort Study. *Inflamm Bowel Dis* 2009; **15**: 39-46 [PMID: 18623166 DOI: 10.1002/ibd.20541]

77 **Silvennoinen J**, Lamberg-Allardt C, Kärkkäinen M, Niemelä S, Lehtola J. Dietary calcium intake and its relation to bone mineral density in patients with inflammatory bowel disease. *J Intern Med* 1996; **240**: 285-292 [PMID: 8946811 DOI: 10.1046/j.1365-2796.1996.25862000.x]

78 **Vernia P**, Loizos P, Di Giuseppantonio I, Amore B, Chiappini A, Cannizzaro S. Dietary calcium intake in patients with inflammatory bowel disease. *J Crohns Colitis* 2014; **8**: 312-317 [PMID: 24090907 DOI: 10.1016/j.crohns.2013.09.008]

79 **Abitbol V**, Mary JY, Roux C, Soulé JC, Belaiche J, Dupas JL, Gendre JP, Lerebours E, Chaussade S; Groupe D'etudes Thérapeutiques des Affections Inflammatoires Digestives (GETAID). Osteoporosis in inflammatory bowel disease: effect of calcium and vitamin D with or without fluoride. *Aliment Pharmacol Ther* 2002; **16**: 919-927 [PMID: 11966500 DOI: 10.1046/j.1365-2036.2002.01247.x]

80 **Ghishan FK**, Kiela PR. Vitamins and Minerals in Inflammatory Bowel Disease. *Gastroenterol Clin North Am* 2017; **46**: 797-808 [PMID: 29173522 DOI: 10.1016/j.gtc.2017.08.011]

81 **Zhang YZ**, Li YY. Inflammatory bowel disease: pathogenesis. *World J Gastroenterol* 2014; **20**: 91-99 [PMID: 24415861 DOI: 10.3748/wjg.v20.i1.91]

82 **Del Pinto R**, Pietropaoli D, Chandar AK, Ferri C, Cominelli F. Association Between Inflammatory Bowel Disease and Vitamin D Deficiency: A Systematic Review and Meta-analysis. *Inflamm Bowel Dis* 2015; **21**: 2708-2717 [PMID: 26348447 DOI: 10.1097/MIB.0000000000000546]

83 **Farraye FA**, Nimitphong H, Stucchi A, Dendrinos K, Boulanger AB, Vijjeswarapu A, Tanennbaum A, Biancuzzo R, Chen TC, Holick MF. Use of a novel vitamin D bioavailability test demonstrates that vitamin D absorption is decreased in patients with quiescent Crohn's disease. *Inflamm Bowel Dis* 2011; **17**: 2116-2121 [PMID: 21910173 DOI: 10.1002/ibd.21595]

84 **Levin AD**, Wadhera V, Leach ST, Woodhead HJ, Lemberg DA, Mendoza-Cruz AC, Day AS. Vitamin D deficiency in children with inflammatory bowel disease. *Dig Dis Sci* 2011; **56**: 830-836 [PMID: 21222159 DOI: 10.1007/s10620-010-1544-3]

85 **Palermo A**, Tuccinardi D, D'Onofrio L, Watanabe M, Maggi D, Maurizi AR, Greto V, Buzzetti R, Napoli N, Pozzilli P, Manfrini S. Vitamin K and osteoporosis: Myth or reality? *Metabolism* 2017; **70**: 57-71 [PMID: 28403946 DOI: 10.1016/j.metabol.2017.01.032]

86 **Dubois-Camacho K**, Ottum PA, Franco-Muñoz D, De la Fuente M, Torres-Riquelme A, Díaz-Jiménez D, Olivares-Morales M, Astudillo G, Quera R, Hermoso MA. Glucocorticosteroid therapy in inflammatory bowel diseases: From clinical practice to molecular biology. *World J Gastroenterol* 2017; **23**: 6628-6638 [PMID: 29085208 DOI: 10.3748/wjg.v23.i36.6628]

87 **LoCascio V**, Bonucci E, Imbimbo B, Ballanti P, Adami S, Milani S, Tartarotti D, DellaRocca C. Bone loss in response to long-term glucocorticoid therapy. *Bone Miner* 1990; **8**: 39-51 [PMID: 2306553 DOI: 10.1016/0169-6009(91)90139-Q]

88 **Van Staa TP**, Laan RF, Barton IP, Cohen S, Reid DM, Cooper C. Bone density threshold and other predictors of vertebral fracture in patients receiving oral glucocorticoid therapy. *Arthritis Rheum* 2003; **48**: 3224-3229 [PMID: 14613287 DOI: 10.1002/art.11283]

89 **Hofbauer LC**, Gori F, Riggs BL, Lacey DL, Dunstan CR, Spelsberg TC, Khosla S. Stimulation of osteoprotegerin ligand and inhibition of osteoprotegerin production by glucocorticoids in human osteoblastic lineage cells: potential paracrine mechanisms of glucocorticoid-induced osteoporosis. *Endocrinology* 1999; **140**: 4382-4389 [PMID: 10499489 DOI: 10.1210/endo.140.10.7034]

90 **Hofbauer LC**, Zeitz U, Schoppet M, Skalicky M, Schüler C, Stolina M, Kostenuik PJ, Erben RG. Prevention of glucocorticoid-induced bone loss in mice by inhibition of RANKL. *Arthritis Rheum* 2009; **60**: 1427-1437 [PMID: 19404943 DOI: 10.1002/art.24445]

91 **Piemontese M**, Xiong J, Fujiwara Y, Thostenson JD, O'Brien CA. Cortical bone loss caused by glucocorticoid excess requires RANKL production by osteocytes and is associated with reduced OPG expression in mice. *Am J Physiol Endocrinol Metab* 2016; **311**: E587-E593 [PMID: 27460899 DOI: 10.1152/ajpendo.00219.2016]

92 **Rutgeerts P**, Löfberg R, Malchow H, Lamers C, Olaison G, Jewell D, Danielsson A, Goebell H, Thomsen OO, Lorenz-Meyer H. A comparison of budesonide with prednisolone for active Crohn's disease. *N Engl J Med* 1994; **331**: 842-845 [PMID: 8078530 DOI: 10.1056/NEJM199409293311304]

93 **D'Haens G**, Verstraete A, Cheyns K, Aerden I, Bouillon R, Rutgeerts P. Bone turnover during short-term therapy with methylprednisolone or budesonide in Crohn's disease. *Aliment Pharmacol Ther* 1998; **12**: 419-424 [PMID: 9663720 DOI: 10.1046/j.1365-2036.1998.00321.x]

94 **Christakos S**, Dhawan P, Porta A, Mady LJ, Seth T. Vitamin D and intestinal calcium absorption. *Mol Cell Endocrinol* 2011; **347**: 25-29 [PMID: 21664413 DOI: 10.1016/j.mce.2011.05.038]

95 **Wasserman RH.** Vitamin D and intestinal absorption of calcium: a view and over-view. In: P JW, Feldman D, Glorieux F, editors. Vitamin D. San Diego, 2005: 411–428 [DOI: 10.1016/B978-012252687-9/50027-9]

96 **Xue Y**, Fleet JC. Intestinal vitamin D receptor is required for normal calcium and bone metabolism in mice. *Gastroenterology* 2009; **136**: 1317-1327, e1-e2 [PMID: 19254681 DOI: 10.1053/j.gastro.2008.12.051]

97 **Vogelsang H**, Ferenci P, Woloszczuk W, Resch H, Herold C, Frotz S, Gangl A. Bone disease in vitamin D-deficient patients with Crohn's disease. *Dig Dis Sci* 1989; **34**: 1094-1099 [PMID: 2743850 DOI: 10.1007/BF01536381]

98 **Tromm A**, Rickels K, Hüppe D, Wiebe V, May B. [Osteopenia in chronic inflammatory bowel diseases. Results of a cross-sectional study using quantitative computerized tomography]. *Leber Magen Darm* 1994; **24**: 23-26, 29-30 [PMID: 8145623]

99 **Satsangi J**, Silverberg MS, Vermeire S, Colombel JF. The Montreal classification of inflammatory bowel disease: controversies, consensus, and implications. *Gut* 2006; **55**: 749-753 [PMID: 16698746 DOI: 10.1136/gut.2005.082909]

100 **Cravo M**, Guerreiro CS, dos Santos PM, Brito M, Ferreira P, Fidalgo C, Tavares L, Pereira AD. Risk factors for metabolic bone disease in Crohn's disease patients. *Inflamm Bowel Dis* 2010; **16**: 2117-2124 [PMID: 20848459 DOI: 10.1002/ibd.21297]

101 **Lima CA**, Lyra AC, Mendes CMC, Lopes MB, Coqueiro FG, Rocha R, Santana GO. Bone mineral density and inflammatory bowel disease severity. *Braz J Med Biol Res* 2017; **50**: e6374 [PMID: 29069227 DOI: 10.1590/1414-431X20176374]

102 **Holleran G**, Lopetuso L, Petito V, Graziani C, Ianiro G, McNamara D, Gasbarrini A, Scaldaferri F. The Innate and Adaptive Immune System as Targets for Biologic Therapies in Inflammatory Bowel Disease. *Int J Mol Sci* 2017; **18** [PMID: 28934123 DOI: 10.3390/ijms18102020]

103 **Takahashi N**, Udagawa N, Suda T. A new member of tumor necrosis factor ligand family, ODF/OPGL/TRANCE/RANKL, regulates osteoclast differentiation and function. *Biochem Biophys Res Commun* 1999; **256**: 449-455 [PMID: 10080918 DOI: 10.1006/bbrc.1999.0252]

104 **Theill LE**, Boyle WJ, Penninger JM. RANK-L and RANK: T cells, bone loss, and mammalian evolution. *Annu Rev Immunol* 2002; **20**: 795-823 [PMID: 11861618 DOI: 10.1146/annurev.immunol.20.100301.064753]

105 **Kaji K**, Katogi R, Azuma Y, Naito A, Inoue JI, Kudo A. Tumor necrosis factor alpha-induced osteoclastogenesis requires tumor necrosis factor receptor-associated factor 6. *J Bone Miner Res* 2001; **16**: 1593-1599 [PMID: 11547829 DOI: 10.1359/jbmr.2001.16.9.1593]

106 **Tsuboi M**, Kawakami A, Nakashima T, Matsuoka N, Urayama S, Kawabe Y, Fujiyama K, Kiriyama T, Aoyagi T, Maeda K, Eguchi K. Tumor necrosis factor-alpha and interleukin-1beta increase the Fas-mediated apoptosis of human osteoblasts. *J Lab Clin Med* 1999; **134**: 222-231 [PMID: 10482306 DOI: 10.1016/S0022-2143(99)90201-9]

107 **Azuma Y**, Kaji K, Katogi R, Takeshita S, Kudo A. Tumor necrosis factor-alpha induces differentiation of and bone resorption by osteoclasts. *J Biol Chem* 2000; **275**: 4858-4864 [PMID: 10671521 DOI: 10.1074/jbc.275.7.4858]

108 **Franchimont N**, Putzeys V, Collette J, Vermeire S, Rutgeerts P, De Vos M, Van Gossum A, Franchimont D, Fiasse R, Pelckmans P, Malaise M, Belaiche J, Louis E. Rapid improvement of bone metabolism after infliximab treatment in Crohn's disease. *Aliment Pharmacol Ther* 2004; **20**: 607-614 [PMID: 15352908 DOI: 10.1111/j.1365-2036.2004.02152.x]

109 **Bernstein M**, Irwin S, Greenberg GR. Maintenance infliximab treatment is associated with improved bone mineral density in Crohn's disease. *Am J Gastroenterol* 2005; **100**: 2031-2035 [PMID: 16128948 DOI: 10.1111/j.1572-0241.2005.50219.x]

110 **Ryan BM**, Russel MG, Schurgers L, Wichers M, Sijbrandij J, Stockbrugger RW, Schoon E. Effect of antitumour necrosis factor-alpha therapy on bone turnover in patients with active Crohn's disease: a prospective study. *Aliment Pharmacol Ther* 2004; **20**: 851-857 [PMID: 15479356 DOI: 10.1111/j.1365-2036.2004.02097.x]

111 **Miheller P**, Muzes G, Zagoni T, Toth M, Racz K, Tulassay Z. Infliximab therapy improves the bone metabolism in fistulizing Crohn's disease. *Dig Dis* 2006; **24**: 201-206 [PMID: 16699279 DOI: 10.1159/000091299]

112 **Maldonado-Pérez MB**, Castro-Laria L, Caunedo-Álvarez A, Montoya-García MJ, Giner-García M, Argüelles-Arias F, Romero-Gómez M, Vázquez-Gámez MÁ. Does the Antitumor Necrosis Factor-α Therapy Decrease the Vertebral Fractures Occurrence in Inflammatory Bowel Disease? *J Clin Densitom* 2019; **22**: 195-202 [PMID: 30205986 DOI: 10.1016/j.jocd.2018.07.010]

113 **Veerappan SG**, Healy M, Walsh BJ, O'Morain CA, Daly JS, Ryan BM. Adalimumab Therapy Has a Beneficial Effect on Bone Metabolism in Patients with Crohn's Disease. *Dig Dis Sci* 2015; **60**: 2119-2129 [PMID: 25732718 DOI: 10.1007/s10620-015-3606-z]

114 **Kanis JA**, Johnell O, Oden A, Johansson H, McCloskey E. FRAX and the assessment of fracture probability in men and women from the UK. *Osteoporos Int* 2008; **19**: 385-397 [PMID: 18292978 DOI: 10.1007/s00198-007-0543-5]

115 **Harbord M**, Annese V, Vavricka SR, Allez M, Barreiro-de Acosta M, Boberg KM, Burisch J, De Vos M, De Vries AM, Dick AD, Juillerat P, Karlsen TH, Koutroubakis I, Lakatos PL, Orchard T, Papay P, Raine T, Reinshagen M, Thaci D, Tilg H, Carbonnel F; European Crohn’s and Colitis Organisation. The First European Evidence-based Consensus on Extra-intestinal Manifestations in Inflammatory Bowel Disease. *J Crohns Colitis* 2016; **10**: 239-254 [PMID: 26614685 DOI: 10.1093/ecco-jcc/jjv213]

116 **Scott EM**, Gaywood I, Scott BB. Guidelines for osteoporosis in coeliac disease and inflammatory bowel disease. British Society of Gastroenterology. *Gut* 2000; **46 Suppl 1**: i1-i8 [PMID: 10647595 DOI: 10.1136/gut.46.suppl\_1.I1]

117 **Black DM**, Rosen CJ. Postmenopausal Osteoporosis. *N Engl J Med* 2016; **374**: 2096-2097 [PMID: 27223157 DOI: 10.1056/NEJMc1602599]

118 **Melek J**, Sakuraba A. Efficacy and safety of medical therapy for low bone mineral density in patients with inflammatory bowel disease: a meta-analysis and systematic review. *Clin Gastroenterol Hepatol* 2014; **12**: 32-44.e5 [PMID: 23981521 DOI: 10.1016/j.cgh.2013.08.024]

119 **Papapoulos S**, Chapurlat R, Libanati C, Brandi ML, Brown JP, Czerwiński E, Krieg MA, Man Z, Mellström D, Radominski SC, Reginster JY, Resch H, Román Ivorra JA, Roux C, Vittinghoff E, Austin M, Daizadeh N, Bradley MN, Grauer A, Cummings SR, Bone HG. Five years of denosumab exposure in women with postmenopausal osteoporosis: results from the first two years of the FREEDOM extension. *J Bone Miner Res* 2012; **27**: 694-701 [PMID: 22113951 DOI: 10.1002/jbmr.1479]

120 **Nuti R**, Brandi ML, Checchia G, Di Munno O, Dominguez L, Falaschi P, Fiore CE, Iolascon G, Maggi S, Michieli R, Migliaccio S, Minisola S, Rossini M, Sessa G, Tarantino U, Toselli A, Isaia GC. Guidelines for the management of osteoporosis and fragility fractures. *Intern Emerg Med* 2019; **14**: 85-102 [PMID: 29948835 DOI: 10.1007/s11739-018-1874-2]

**P-Reviewer:** Poturoglu S, Fan H **S-Editor:** Dou Y **L-Editor:**A **E-Editor:** Wang J

**Specialty type:** Medicine, Research and Experimental

**Country of origin:** Italy

**Peer-review report classification**

Grade A (Excellent): 0

Grade B (Very good): B

Grade C (Good): C

Grade D (Fair): 0

Grade E (Poor): 0

**Table 1 European Crohn and Colitis Organization guidelines for the management of bone alterations in inflammatory bowel diseases population**

|  |
| --- |
| **ECCO guidelines for the management of bone alterations in IBD population**  |
| Life style recommendations | Physical exercise; Stopping smoking; and adequate dietary calcium (1 g/daily) |
| Vitamin and mineral supplements | Calcium (500-1000 mg/daily); Vitamin D (dose of ~1000 IU daily, or higher dose if known vitamin D deficiency) supplement for prophylaxis in patients receiving systemic steroid therapy; Calcium and vitamin D supplement if the T score is lower than -1.5 |
| Treatment recommendations | More intensive treatment in patients with a history of pre-existing fracture; Regular use of BPs and other therapies in subjects with underlying disease activity particularly in young and postmenopausal women or those with previous spontaneous fractures |

ECCO: European Crohn and Colitis Organization; IBD: Inflammatory bowel diseases.

**Figure 1 Modulation of osteoclast differentiation by serum TNF-α and anti-TNF-α treatment.** A: TNF-α influences osteoclast precursor differentiation and bone resorption activity inducing RANKL expression on osteoblast cells and preventing the binding of OPG; B: Anti-TNF-α treatment reduces RANKL expression resulting in decrease of osteoclast differentiation and bone resorption.

**Figure 2 Gut-bone immune signaling: interplay between different factors which may affect bone metabolism in patients with inflammatory bowel diseases.**

**Figure 3 Diagramatic representation of the pathogenic mechanisms involved in alteration of bone mineral density in inflammatory bowel diseases.**