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**Allyl isothiocyanate ameliorates lipid accumulation and inflammation in nonalcoholic fatty liver disease *via* the Sirt1/AMPK and NF-κB signaling pathways**

Li CX *et al*. Allyl isothiocyanate ameliorates NAFLD

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**Abstract**

BACKGROUND

Allyl isothiocyanate (AITC), a classic anti-inflammatory and antitumorigenic agent, was recently identified as a potential treatment for obesity and insulin resistance. However, little is known about its direct impact on the liver.

***AIM***

To investigate the effect and underlying mechanism of AITC in nonalcoholic fatty liver disease (commonly referred to as NAFLD).

METHODS

To establish a mouse and cellular model of NAFLD, C57BL/6 mice were fed a high fat diet (HFD) for 8 wk, and AML-12 cells were treated with 200 μM palmitate acid for 24 h. For AITC treatment, mice were administered AITC (100 mg/kg/d) orally and AML-12 cells were treated with AITC (20 μmol/L).

RESULTS

AITC significantly ameliorated HFD-induced weight gain, hepatic lipid accumulation and inflammation *in vivo*. Furthermore, serum alanine aminotransferase and aspartate aminotransferase levels were markedly reduced in AITC-treated mice. Mechanistically, AITC significantly downregulated the protein levels of sterol regulatory element­binding protein 1 (SREBP1) and its lipogenesis target genes and upregulated the levels of proteins involved in fatty acid β-oxidation, as well as the upstream mediators Sirtuin 1 (Sirt1) and AMP-activated protein kinase α (AMPKα), in the livers of HFD-fed mice. AITC also attenuated the nuclear factor kappa B (NF-κB) signaling pathway. Consistently, AITC relieved palmitate acid-induced lipid accumulation and inflammation in AML-12 cells *in vitro* through the Sirt1/AMPK and NF-κB signaling pathways. Importantly, further studies showed that the curative effect of AITC on lipid accumulation was abolished by siRNA-mediated knockdown of either Sirt1 or AMPKα in AML-12 cells.

CONCLUSION

AITC significantly ameliorates hepatic steatosis and inflammation by activating the Sirt1/AMPK pathway and inhibiting the NF-κB pathway. Therefore, AITC is a potential therapeutic agent for NAFLD.

**Key words:** Allyl isothiocyanate; Nonalcoholic fatty liver disease; Hepatic steatosis; Liver inflammation

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**Core tip:** Nonalcoholic fatty liver disease (NAFLD) is rapidly prevalent as a remarkable problem worldwide. We aimed to investigate the therapeutic role of allyl isothiocyanate (AITC) in lipid accumulation and inflammation during NAFLD development in mice fed a high fat diet and AML-12 cells treated with palmitate acid. Our study for the first time demonstrates that AITC ameliorates hepatic steatosis and inflammation by activating the Sirt1/AMPK and IKK/NF-κB signaling pathway. This study reveals role for AITC as a potential therapeutic agent for NAFLD.

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**INTRODUCTION**

Nonalcoholic fatty liver disease (NAFLD) is currently one of the most epidemic chronic liver diseases worldwide and has become an enormous clinical and economic burden. NAFLD affects approximately one-fourth of adults globally[1]. NAFLD represents a wide spectrum of disease stages, ranging from simple steatosis to nonalcoholic steatohepatitis (commonly known as NASH), which is characterized by hepatocellular injury and inflammation and may eventually progress to cirrhosis and hepatocellular carcinoma[2]. Furthermore, NAFLD is a strong risk factor for type 2 diabetes, atherosclerosis, cardiovascular disease and chronic kidney disease[3-5]. However, its pathogenesis remains unclear, and current therapeutic options are relatively limited. No treatments for NAFLD other than lifestyle modifications leading to weight loss and increased physical activity are currently approved[6]. Therefore, there is an urgent need to develop effective medical treatments for NAFLD.

Hepatic steatosis, characterized by excessive triglyceride (TG) accumulation in hepatocytes, is strongly associated with chronic hepatic inflammation and insulin resistance[7]. Furthermore, the I*κ*B kinase (IKK)/nuclear factor kappa B (NF-κB) signaling pathway plays a crucial role in the development of metabolic disorders, including NAFLD, and especially in hepatic inflammation[8-10].

Sirtuin 1 (Sirt1) is a highly conserved nicotinamide adenine dinucleotide-dependent protein deacetylase that regulates a wide variety of biological functions in mammals, including lipid metabolism and energy homeostasis[11,12]. AMP-activated protein kinase (AMPK) functions as an energy switch that controls several cellular processes, such as lipid metabolism, by inhibiting hepatic lipogenesis and stimulating fatty acid oxidation[13]. Previous studies revealed that Sirt1 is an important regulator of hepatic lipogenesis and fatty acid oxidation through multiple nutrient sensors, including sterol regulatory element­binding protein 1 (SREBP1), peroxisome proliferator-activated receptor gamma coactivator1α (PGC1α) and peroxisome proliferator-activated receptor α (PPARα)[14-16]. Furthermore, Sirt1 was shown to regulate AMPK activation in NAFLD, resulting in enhanced lipolysis and β-oxidation, as well as ameliorated hepatic steatosis[17-19].

Allyl isothiocyanate (AITC) is derived from its precursor sinigrin, which is present in many common cruciferous vegetables and is widely consumed by humans[20]. Myrosinase in the intestinal microflora catalyzes the hydrolysis of sinigrin to AITC in both humans and animals[20,21]. Previous studies have shown that AITC exhibits anti-inflammatory and anticancer activities[22-24]. Recently, AITC was identified as a potential novel treatment for diet-induced obesity and insulin resistance through its modulation of mitochondrial dysfunction[25]. Another study revealed that AITC can augment basal and epinephrine-induced lipolysis in adipocytes and intensify hydrolysis of TG in the blood serum of rats[26]. Moreover, a previous study showed AITC effectively inhibits adipogenic differentiation of 3T3-L1 preadipocytes and suppresses expression of genes up-regulated during adipogenesis[27]. However, little is known about its direct impact on the liver or its underlying mechanism.

Herein, we conducted both *in vivo* and *in vitro* experiments to explore the effect of AITC on NAFLD, focusing on its role in hepatic steatosis and inflammatory responses, and to elucidate its mechanism of action.

**MATERIALS AND METHODS**

***Animal experiments***

All experiments were conducted with approval of the First Afﬁliated Hospital of Zhejiang University Institutional Animal Care and Use Committee (Permit number: 2016-231). Six-week-old male C57BL/6 mice were purchased from B&K Laboratory Animal Corp., Ltd. (Shanghai, China). After acclimatization for 2 wk with free access to food and water, mice were fed a standard chow diet (SCD) or high fat diet (HFD) (60% fat-derived calories, 20% carbohydrate-derived calories, and 20% protein-derived calories; D12492, Research Diets, New Brunswick, NJ, United States). In general, mice were given SCD or HFD feeding for a total of 8 wk, and from the 5th wk, SCD-fed mice began to receive corn oil (control) (*n* = 10), and HFD-fed mice were randomly divided into two groups to receive 100 mg/kg/d AITC (99.7%; Sigma-Aldrich, St. Louis, MO, United States) (*n* = 10) or corn oil (*n* = 9) daily by gavage for an additional 4 wk while remaining on SCD or HFD.

***Cell culture and treatments***

The established immortalized AML-12 mouse hepatocyte cell line was purchased from the Type Culture Collection of the Chinese Academy of Sciences (Shanghai, China). AML-12 cells were cultured in DMEM/F12 (1:1) medium supplemented with 10% FBS, 100 U/mL penicillin, 100 µg/mL streptomycin, 0.1 µmol/L dexamethasone, and 1% insulin-transferrin-selenium Liquid Media Supplement (I3146; Sigma-Aldrich). To establish a cellular model of NAFLD, palmitate acid (PA) (Sigma-Aldrich) was dissolved in bovine serum albumin (Sangon Biotech, Shanghai, China), and then AML-12 cells were exposed to 200 μM PA for 24 h. To investigate the effect of AITC on lipid deposition *in vitro*, PA-stimulated AML-12 cells in serum-free conditions were treated with AITC (20 μmol/L) or dimethyl sulfoxide (commonly known as DMSO) (vehicle) for 24 h.

AML-12 cells were transfected with *Sirt1* small interfering RNA (siRNA) #1 (target sequence 5’-GATGAAGTTGACCTCCTCA-3’), *Sirt1* siRNA #2 (target sequence 5’-CCGATGGACTCCTCACTAA-3’), *Sirt1* siRNA #3 (target sequence 5’-GGTTGTTAATGAAGCTATA-3’), *AMPKα* siRNA #1 (target sequence 5’-GCAGAAGATTCGGAGCCTT-3’), *AMPKα* siRNA #2 (target sequence 5’-GCACACCCTGGATGAATTA-3’), *AMPKα* siRNA #3 (target sequence 5’-GCAGAAGTTTGTAGAGCAA-3’) or the corresponding scrambled control (RIBOBIO, Guangzhou, China) using Lipofectamine RNAiMAX (Invitrogen, Shanghai, China) according to the manufacturer’s protocol. After 48 h, the cells were incubated in medium containing PA with or without AITC for an additional 24 h.

***Hepatic and cellular TG assay***

Hepatic and cellular TG contents were measured using a commercial kit (Applygen Technologies Inc., Beijing, China) according to the manufacturer’s protocol.

***Hematoxylin–eosin and oil red O staining***

Mouse liver tissues were rapidly harvested, fixed in 10% formalin, embedded in paraffin, sectioned and stained with hematoxylin and eosin (commonly known as H&E) for histological examination. Frozen liver sections (8 μm) and cells in 6-well plates were stained with oil red O (Sigma-Aldrich) to assess lipid accumulation.

***Metabolic measurements***

Plasma alanine aminotransferase (ALT), aspartate aminotransferase (AST), total cholesterol and uric acid levels were determined with a Hitachi 7600 autoanalyzer (Hitachi, Tokyo, Japan) according to the manufacturer’s instructions.

***Quantitative real-time PCR***

Total mRNA was extracted from liver tissues or cultured cells using RNA plus (Takara, Dalian, China) and reverse transcribed into cDNA using a PrimeScript® RT reagent kit (Takara, Japan) according to the manufacturer’s protocol. Real-time PCR was performed on an ABI Prism 7500 Sequence Detection System (Applied Biosystems, Foster City, CA, United States) using SYBR Green (Takara) to quantify PCR amplification. Relative mRNA expression levels of target genes were normalized to β-actin or GAPDH mRNA levels for each sample.

***Western blot analysis***

Liver tissue samples and cells were lysed using RIPA buffer (Applygen Technologies Inc.) supplemented with protease and phosphatase inhibitors (Sigma). Equal amounts of extracted proteins were separated by SDS-PAGE and transferred to PVDF membranes (Millipore, Inc., Darmstadt, Germany). Membranes were blocked with 5% nonfat milk in TBST and then incubated overnight at 4°C with the following primary antibodies: anti-Sirt1 (8469), anti-AMPKα (2603), anti-phosphorylated (p) AMPKα(p-*AMPKα*; 2535), anti-NF-κB p65 (6956), anti-p-NF-κB p65 (3033), anti-p-IKK (2697), anti-*IKKα* (2682), anti-*IKKβ* (8943), anti-p- inhibitor of nuclear factor kappa B alpha (IκBα) (2859), anti-IκBα (4812), anti-glyceraldehyde-3-phosphate dehydrogenase (GAPDH) (2118), and β-actin (3700) (Cell Signaling Technology); and anti-PGC1α (ab54481), anti-PPARα (ab8934), anti-carnitine palmitoyltransferase 1 α (CPT1α) (ab128568), anti-SREBP1 (ab3259), anti-fatty acid synthase (FAS) (ab128856), anti-stearoyl coenzyme A desaturase 1 (SCD1) (ab19862), and anti-tubulin (ab6160) (Abcam, Cambridge, United Kingdom). Proteins levels were evaluated using an enhanced ECL kit (Lianke, Hangzhou, China).

***Statistical analysis***

The statistical methods of this study were reviewed by Hong Zhang from the Department of Statistics and Finance, School of Management, University of Science and Technology of China. All statistical analyses were performed with SPSS 22 (IBM, Chicago, IL, United States). Data are presented as the average ± S.D. Statistical analysis was carried out using Student’s two-tailed *t*-test and one-way ANOVA with Tukey’s post-test. *P*-values less than 0.05 indicated statistical signiﬁcance.

**RESULTS**

***AITC reduces body weight, ameliorates hepatic steatosis and attenuates liver injury in an HFD mouse model***

Supplementary Figure 1 shows the chemical structure of AITC. To explore the effect of AITC on NAFLD, an HFD model, which is quite similar to but does not completely mirror human NAFLD, was adopted. HFD-fed mice exhibited higher body weight (Figure 1A and B) and liver weight (Figure 1D), more serious lipid accumulation (Figure 1C), much higher TG (Figure 1E), ALT (Figure 1F), AST (Figure 1G), cholesterol (Figure 1H) and uric acid (Figure 1I) levels compared to SCD-fed mice, showing characteristics of NAFLD *in vivo*. The mice exhibited a significant decrease in body weight upon 100 mg/kg/d AITC administration (Figure 1A and B) and a slight but nonsignificant decrease in liver weight (Figure 1D). Notably, hepatic steatosis was improved after AITC treatment, as demonstrated by decreased hepatic TG levels (Figure 1E) and reduced lipid accumulation (H&E and oil red O staining; Figure 1C). As shown in figure 1F-I, AITC attenuated HFD-induced liver injury, as evidenced by markedly decreased serum ALT and AST levels. These findings demonstrate that AITC ameliorates body weight, hepatic steatosis and liver injury in HFD-fed mice.

***AITC attenuates de novo lipogenesis and promotes fatty acid*** ***β-oxidation by activating the Sirt1/AMPK pathway in vivo***

To investigate the mechanisms by which AITC ameliorates hepatic steatosis, the expression levels of hepatic lipid metabolism-related genes were measured. *SREBP1* is the central transcription factor that enhances the expression of genes required for hepatic fatty acid synthesis and TG synthesis[28]. As a result, AITC treatment *in vivo* significantly downregulated *SREBP1* protein levels and its target genes, including *SCD1* and *FAS* (Figure 2A). In contrast, AITC increased the expression levels of proteins involved in fatty acid β-oxidation, such as *PGC1α*, *PPARα* and *CPT1α*, in the livers of HFD-fed mice (Figure 2A).

Previous studies revealed that *Sirt1* activation attenuates hepatic steatosis in mice with diet- or genetics-induced obesity by regulating hepatic lipogenesis and fatty acid oxidation[16,29,30]. To investigate whether Sirt1/AMPK are involved in the amelioration of the HFD-induced dysregulation of hepatic lipid homeostasis by AITC, we evaluated *Sirt1* and p-*AMPKα* expression levels *in vivo*. As shown in figure 2B, AITC treatment markedly elevated hepatic protein levels of Sirt1 and p-AMPKα. Collectively, these results suggest that AITC attenuates *de novo* lipogenesis and promotes fatty acid β-oxidation by activating the Sirt1/AMPKsignaling pathway *in vivo* in the liver tissues of HFD-fed mice.

***AITC decreases HFD-induced inflammation by inhibiting the NF-κB signaling pathway in vivo***

In addition to its protective role in lipogenesis and fatty acid β-oxidation, AITC treatment substantially decreased the transcription of proinflammatory cytokines [tumor necrosis factor α (TNFα), interleukin (IL)-1β, and IL-6] in the liver tissues of HFD-fed mice (Figure 3A).

To investigate whetherIKK/NF-κBare involved in the amelioration of hepatic inflammation by AITC, the expression levels of IKK/NF-κB signaling pathway components were detected. As shown in figure 3B, AITC treatment upregulated *IκBα* protein levels and downregulated IKK, IκBα, and p65 phosphorylation. These results indicate that AITC decreases inflammation by inhibiting the IKK/NF-κB signaling pathway *in vivo*.

***AITC alleviates PA-induced lipid accumulation in hepatocytes***

Our above data revealed that AITC could ameliorate hepatic steatosis *in vivo*. To investigate the effect of AITC on PA-induced lipid deposition *in vitro*, we used AML-12 cells, which have been well documented as cellular models of NAFLD[31,32]. First, we evaluated the viability of AML-12 cells after AITC treatment. The cultured hepatocytes were treated with different concentrations of AITC for 24 h, and cell viability was measured using cholecystokinin-8 and lactate dehydrogenase release assays. Finally, we chose 20 μM as the optimal AITC concentration for subsequent experiments because this concentration did not significantly affect cell viability (Figure 4A and B). Then, PA-stimulated AML-12 cells were treated with AITC (20 μmol/L) or vehicle for 24 h. As shown in figure 4C and D, AITC relieved the PA-induced increases in intracellular TG levels and lipid accumulation in AML-12 cells. These data demonstrate that AITC directly alleviates PA-induced lipid accumulation in hepatocytes.

***AITC attenuates de novo lipogenesis and promotes fatty acid β-oxidation by activating the Sirt1/AMPK signaling pathway in vitro***

Consistent with the *in vivo* findings, AITC treatment significantly decreased the protein levels of SREBP1 and its target genes, including *SCD1*, *FAS* and acetyl-CoA carboxylase (*ACC*), in PA-treated AML-12 cells (Figure 5A). In addition, AITC enhanced *PGC1α* expression in PA-treated AML-12 cells (Figure 5C).

As shown in figure 5B *in vitro*, AITC upregulated *Sirt1* and p-*AMPKα* levels, consistent with the *in vivo* results (Figure 6A). These results indicate that AITC attenuates lipogenesis and promotes fatty acid β-oxidation by activating the Sirt1/AMPK signaling pathway *in vitro*.

***AITC attenuates inflammation by inhibiting the NF-κB signaling pathway in vitro***

Consistent with the *in vivo* findings, AITC treatment significantly decreased *TNFα* and *IL-6* mRNA levels in PA-treated AML-12 cells (Figure 6A). As shown in figure 6B, AITC upregulatedIκBα protein levels and downregulated IKK, IκBα, and p65 phosphorylation in PA-stimulated AML-12 cells. Taken together, these findings clearly indicate that AITC may attenuate inflammation by inhibiting the *NF-κB* signaling pathway *in vitro*.

***Sirt1 or AMPKα knockdown abolishes the ability of AITC to mitigate TG levels***

To investigate whether AITC can alleviate lipid accumulation through the Sirt1/AMPK pathway, Sirt1 expression in AML-12 cells was selectively knocked down by siRNA transfection (Figure 7A). Sirt1 knockdown abolished the ability of AITC to ameliorate TG accumulation and p-AMPKα and PGC1α upregulation induced by PA in AML-12 cells (Figure 7B and C). Consistently, AMPKα knockdown significantly reversed the effects of AITC on PA-induced intracellular TG accumulation in AML-12 cells (Figure 7D and E). Taken together, these findings verify that AITC ameliorates lipid accumulation by activating the Sirt1/AMPK signaling pathway.

**DISCUSSION**

In the current study, we investigated the effects and mechanisms of AITC on NAFLD development (summarized in Figure 8A). Our results demonstrate that AITC significantly ameliorates hepatic lipid accumulation by activating the *Sirt1*/*AMPK* pathway and alleviates hepatic inflammatory responses by inhibiting the *NF-κB* pathway both *in vivo* and *in vitro*.

Some studies suggested that AITC leads to better metabolic outcome. Kim YJ demonstrated that AITC effectively suppresses the expression of genes that are up-regulated during adipogenesis, such as PPARγ, C/EBPα and FAS[27]. Miyata S showed that AITC reduces *de novo* synthesis of both fatty acids and cholesterol in human hepatoma Huh-7 cells[33]. These results indicate a physiological function of AITC in lipid metabolism regulation.

A previous study by Anh et al. demonstrated that AITC protects against HFD-induced obesity and insulin resistance in mice and that the protective effect of AITC may be partly mediated through the modulation of mitochondrial dysfunction in skeletal muscle cells and the liver[25]. However, the mechanism underlying the beneficial effects of AITC on hepatic steatosis and liver inflammation has not been clearly defined. Our results suggested that AITC ameliorates lipid accumulation by activating the Sirt1/AMPKα signaling pathway and improves inflammation in hepatocytes both *in vivo* and *in vitro*.

A series of recent studies revealed that Sirt1 plays a critical role in various cellular and physiological processes, including lipid metabolism and energy homeostasis. A previous study demonstrated that Sirt1 transgenic mice are protected from HFD-induced metabolic damage *via* the upregulation of *PGC1α* and the inhibition of the *NF-κB* pathway[29]. Consistently, another study showed that hepatic overexpression of *Sirt1* attenuates hepatic steatosis, possibly by inhibiting endoplasmic reticulum stress in the liver of obese mice[30]. On the other hand, liver-specific *Sirt1*-knockout mice develop hepatic steatosis, hepatic inflammation, and endoplasmic reticulum stress, which are associated with decreased hepatic fatty acid oxidation and increased lipogenesis[14,15]. In addition, these findings were supported by later reports that pharmacological activation of *Sirt1* protects against HFD-induced metabolic disorders[17,34]. Sirt1 regulates lipid homeostasis through multiple nutrient sensors such as SREBP1, AMPK, PGC1α, PPARα and the hepatocyte-derived hormone fibroblast growth factor 21[14-16,35]. In this study, we found that AITC significantly ameliorated hepatocyte lipid accumulation both *in vivo* and *in vitro* by activating theSirt1/AMPK signaling pathway, resulting in decreased expression of lipogenesis genes, such as *SREBP1*, *SCD1* and *FAS*, and increased expression of fatty acid β-oxidation genes, including *PGC1α*, *PPARα* and *CPT1α*. Furthermore, another study showed that sulforaphane-induced *Sirt1* activation inhibits endoplasmic reticulum (commonly referred to as ER) stress and prevents cardiomyocytes from hypoxia/reoxygenation injury *in vitro*[36]. As we have detected *Sirt1* activation after AITC application, further studies should be carried out to explore mechanisms of ER stress in this model.

Chronic inflammation is characterized by the abnormal production of proinflammatory cytokines, including TNFα, IL-1β and IL-6, and the activation of inflammatory signaling pathways, such as the NF-κB and JNK pathways; moreover, it has been proposed to play a crucial role in the pathogenesis of NAFLD[7,37]. A number of recent studies have demonstrated a key role for the IKK/NF-κB signaling pathway in NAFLD development[8,9]. In response to numerous inflammatory stimuli, *IKK* complex activation induces *IκB* phosphorylation and subsequent degradation, which releases *NF-κB* and allows it to translocate into the nucleus[38]. A previous finding indicated that hepatic lipid accumulation activates the IKK/NF-κB pathway, promoting downstream proinflammatory cytokine production and subacute inflammation[39]. In this study, we demonstrated that AITC treatment upregulatedIκBα protein levels and downregulated IKK, IκBα, and p65 phosphorylation in both the NAFLD mouse and cellular models. Furthermore, AITC treatment substantially decreased hepatic proinflammatory cytokine levels *in vivo* and *in vitro*. In addition, HFD-induced liver injury was attenuated in AITC-treated mice, as evidenced by markedly decreased serum levels of ALT and AST.

Although several isothiocyanates have been proposed as chemopreventive agents for cancers, AITC has been reported to exhibit both carcinogenic and anticarcinogenic potential. A previous study demonstrated that AITC has the ability to cause Cu(II)-mediated DNA damage and induce 8-oxo-7,8-dihydro-29-deoxyguanosine (8-oxodG) formation, leading to carcinogenesis in human myelogenous leukemic cell lines[40]. Moreover, impaired copper availability in obesity-related NAFLD was shown to predict early atherosclerosis as a main cardiovascular risk[41]. On the other hand, another study demonstrated that AITC could inhibit proliferation of human prostate cancer cells through inducing G2/M arrest and apoptosis[22]. Our studies mentioned above have proposed many different mechanisms of AITC-induced amelioration of hepatic steatosis and inflammation in both i*n vivo* and *in vitro* models of NAFLD, but none have explored whether AITC took part in Cu(II)-mediated DNA damage or cancer development. Further studies are required to clarify whether AITC induces oxidative DNA damage in our model and investigate the mechanism of AITC on cancer.

In conclusion, our study demonstrates that AITC ameliorates hepatic steatosis and inflammation by activating the Sirt1/AMPK and inhibiting the IKK/NF-κB signaling pathway, respectively. This study indicates that AITC may be a potential therapeutic agent for the progress of NAFLD.

**Article Highlights**

***Research background***

Nonalcoholic fatty liver disease (NAFLD) is an unmet medical need with no approved therapies. Recent studies have shown that allyl isothiocyanate (AITC) has a potential protective effect on obesity and insulin resistance. The evaluation of the effect of AITC on NAFLD as well as the mechanism of action may provide a new therapeutic trend.

***Research motivation***

Emerging evidence suggests a beneficial role for AITC in inflammation, cancer, diet-induced obesity and insulin resistance. Enhanced lipolysis in adipocytes and intensified hydrolysis of triglyceride in the serum of rats treated with MS-275 was also reported. As little is known about its direct impact on liver or its underlying mechanism, it is imperative to characterize the potential effect of AITC on NAFLD.

***Research objectives***

To validate the effect of AITC on NAFLD and clarify the possible mechanism of action.

***Research methods***

C57BL/6 mice were fed a high fat diet (HFD) for 8 wk, and AML-12 cells were treated with 200 μmol/L palmitate acid (PA) for 24 h to establish *in vivo* and *in vitro* models of hepatic steatosis. Mice were administered AITC (100 mg/kg/d) orally and AML-12 cells were treated with AITC (20 μmol/L) to detect the effect of AITC on NAFLD.

***Research results***

Our results show that AITC significantly ameliorates HFD-induced weight gain, hepatic lipid accumulation, inflammation, and PA-induced lipid accumulation as well as inflammation in AML-12 cells*,*accompanied by activated Sirt1/AMPK and inhibited NF-κB signaling pathways. The curative effect of AITC on lipid accumulation is abolished by siRNA-mediated knockdown of either Sirt1 or AMPKα in AML-12 cells.

***Research conclusions***

AITC treatment protects against HFD and PA-induced lipid accumulation and inflammation *in vivo* and *in vitro.* These effects are associated with Sirt1/AMPK and NF-κB signaling pathways.

***Research perspectives***

Plant compounds such as AITC should be further explored for their potential effective activity in NAFLD.

**References**

1 **Younossi ZM**, Koenig AB, Abdelatif D, Fazel Y, Henry L, Wymer M. Global epidemiology of nonalcoholic fatty liver disease-Meta-analytic assessment of prevalence, incidence, and outcomes. *Hepatology* 2016; **64**: 73-84 [PMID: 26707365 DOI: 10.1002/hep.28431]

2 **Cohen JC**, Horton JD, Hobbs HH. Human fatty liver disease: old questions and new insights. *Science* 2011; **332**: 1519-1523 [PMID: 21700865 DOI: 10.1126/science.1204265]

3 **Chalasani N**, Younossi Z, Lavine JE, Diehl AM, Brunt EM, Cusi K, Charlton M, Sanyal AJ. The diagnosis and management of non-alcoholic fatty liver disease: practice Guideline by the American Association for the Study of Liver Diseases, American College of Gastroenterology, and the American Gastroenterological Association. *Hepatology* 2012; **55**: 2005-2023 [PMID: 22488764 DOI: 10.1002/hep.25762]

4 **Sinn DH**, Cho SJ, Gu S, Seong D, Kang D, Kim H, Yi BK, Paik SW, Guallar E, Cho J, Gwak GY. Persistent Nonalcoholic Fatty Liver Disease Increases Risk for Carotid Atherosclerosis. *Gastroenterology* 2016; **151**: 481-488.e1 [PMID: 27283259 DOI: 10.1053/j.gastro.2016.06.001]

5 **Musso G**, Gambino R, Tabibian JH, Ekstedt M, Kechagias S, Hamaguchi M, Hultcrantz R, Hagström H, Yoon SK, Charatcharoenwitthaya P, George J, Barrera F, Hafliðadóttir S, Björnsson ES, Armstrong MJ, Hopkins LJ, Gao X, Francque S, Verrijken A, Yilmaz Y, Lindor KD, Charlton M, Haring R, Lerch MM, Rettig R, Völzke H, Ryu S, Li G, Wong LL, Machado M, Cortez-Pinto H, Yasui K, Cassader M. Association of non-alcoholic fatty liver disease with chronic kidney disease: a systematic review and meta-analysis. *PLoS Med* 2014; **11**: e1001680 [PMID: 25050550 DOI: 10.1371/journal.pmed.1001680]

6 **Younossi ZM**, Loomba R, Rinella ME, Bugianesi E, Marchesini G, Neuschwander-Tetri BA, Serfaty L, Negro F, Caldwell SH, Ratziu V, Corey KE, Friedman SL, Abdelmalek MF, Harrison SA, Sanyal AJ, Lavine JE, Mathurin P, Charlton MR, Chalasani NP, Anstee QM, Kowdley KV, George J, Goodman ZD, Lindor K. Current and future therapeutic regimens for nonalcoholic fatty liver disease and nonalcoholic steatohepatitis. *Hepatology* 2018; **68**: 361-371 [PMID: 29222911 DOI: 10.1002/hep.29724]

7 **Samuel VT**, Shulman GI. Mechanisms for insulin resistance: common threads and missing links. *Cell* 2012; **148**: 852-871 [PMID: 22385956 DOI: 10.1016/j.cell.2012.02.017]

8 **Wang XA**, Zhang R, She ZG, Zhang XF, Jiang DS, Wang T, Gao L, Deng W, Zhang SM, Zhu LH, Guo S, Chen K, Zhang XD, Liu DP, Li H. Interferon regulatory factor 3 constrains IKKβ/NF-κB signaling to alleviate hepatic steatosis and insulin resistance. *Hepatology* 2014; **59**: 870-885 [PMID: 24123166 DOI: 10.1002/hep.26751]

9 **Arkan MC**, Hevener AL, Greten FR, Maeda S, Li ZW, Long JM, Wynshaw-Boris A, Poli G, Olefsky J, Karin M. IKK-beta links inflammation to obesity-induced insulin resistance. *Nat Med* 2005; **11**: 191-198 [PMID: 15685170 DOI: 10.1038/nm1185]

10 **Wang PX**, Zhang XJ, Luo P, Jiang X, Zhang P, Guo J, Zhao GN, Zhu X, Zhang Y, Yang S, Li H. Hepatocyte TRAF3 promotes liver steatosis and systemic insulin resistance through targeting TAK1-dependent signalling. *Nat Commun* 2016; **7**: 10592 [PMID: 26882989 DOI: 10.1038/ncomms10592]

11 **Picard F**, Kurtev M, Chung N, Topark-Ngarm A, Senawong T, Machado De Oliveira R, Leid M, McBurney MW, Guarente L. Sirt1 promotes fat mobilization in white adipocytes by repressing PPAR-gamma. *Nature* 2004; **429**: 771-776 [PMID: 15175761 DOI: 10.1038/nature02583]

12 **Cohen HY**, Miller C, Bitterman KJ, Wall NR, Hekking B, Kessler B, Howitz KT, Gorospe M, de Cabo R, Sinclair DA. Calorie restriction promotes mammalian cell survival by inducing the SIRT1 deacetylase. *Science* 2004; **305**: 390-392 [PMID: 15205477 DOI: 10.1126/science.1099196]

13 **Herzig S**, Shaw RJ. AMPK: guardian of metabolism and mitochondrial homeostasis. *Nat Rev Mol Cell Biol* 2018; **19**: 121-135 [PMID: 28974774 DOI: 10.1038/nrm.2017.95]

14 **Li Y**, Wong K, Giles A, Jiang J, Lee JW, Adams AC, Kharitonenkov A, Yang Q, Gao B, Guarente L, Zang M. Hepatic SIRT1 attenuates hepatic steatosis and controls energy balance in mice by inducing fibroblast growth factor 21. *Gastroenterology* 2014; **146**: 539-49.e7 [PMID: 24184811 DOI: 10.1053/j.gastro.2013.10.059]

15 **Purushotham A**, Schug TT, Xu Q, Surapureddi S, Guo X, Li X. Hepatocyte-specific deletion of SIRT1 alters fatty acid metabolism and results in hepatic steatosis and inflammation. *Cell Metab* 2009; **9**: 327-338 [PMID: 19356714 DOI: 10.1016/j.cmet.2009.02.006]

16 **Ponugoti B**, Kim DH, Xiao Z, Smith Z, Miao J, Zang M, Wu SY, Chiang CM, Veenstra TD, Kemper JK. SIRT1 deacetylates and inhibits SREBP-1C activity in regulation of hepatic lipid metabolism. *J Biol Chem* 2010; **285**: 33959-33970 [PMID: 20817729 DOI: 10.1074/jbc.M110.122978]

17 **Feige JN**, Lagouge M, Canto C, Strehle A, Houten SM, Milne JC, Lambert PD, Mataki C, Elliott PJ, Auwerx J. Specific SIRT1 activation mimics low energy levels and protects against diet-induced metabolic disorders by enhancing fat oxidation. *Cell Metab* 2008; **8**: 347-358 [PMID: 19046567 DOI: 10.1016/j.cmet.2008.08.017]

18 **Dogra S**, Kar AK, Girdhar K, Daniel PV, Chatterjee S, Choubey A, Ghosh S, Patnaik S, Ghosh D, Mondal P. Zinc oxide nanoparticles attenuate hepatic steatosis development in high-fat-diet fed mice through activated AMPK signaling axis. *Nanomedicine* 2019; **17**: 210-222 [PMID: 30708053 DOI: 10.1016/j.nano.2019.01.013]

19 **Liou CJ**, Wei CH, Chen YL, Cheng CY, Wang CL, Huang WC. Fisetin Protects Against Hepatic Steatosis Through Regulation of the Sirt1/AMPK and Fatty Acid β-Oxidation Signaling Pathway in High-Fat Diet-Induced Obese Mice. *Cell Physiol Biochem* 2018; **49**: 1870-1884 [PMID: 30235452 DOI: 10.1159/000493650]

20 **Zhang Y**. Allyl isothiocyanate as a cancer chemopreventive phytochemical. *Mol Nutr Food Res* 2010; **54**: 127-135 [PMID: 19960458 DOI: 10.1002/mnfr.200900323]

21 **Getahun SM**, Chung FL. Conversion of glucosinolates to isothiocyanates in humans after ingestion of cooked watercress. *Cancer Epidemiol Biomarkers Prev* 1999; **8**: 447-451 [PMID: 10350441]

22 **Xiao D**, Srivastava SK, Lew KL, Zeng Y, Hershberger P, Johnson CS, Trump DL, Singh SV. Allyl isothiocyanate, a constituent of cruciferous vegetables, inhibits proliferation of human prostate cancer cells by causing G2/M arrest and inducing apoptosis. *Carcinogenesis* 2003; **24**: 891-897 [PMID: 12771033 DOI: 10.1093/carcin/bgg023]

23 **Jeong WS**, Kim IW, Hu R, Kong AN. Modulatory properties of various natural chemopreventive agents on the activation of NF-kappaB signaling pathway. *Pharm Res* 2004; **21**: 661-670 [PMID: 15139523]

24 **Wagner AE**, Boesch-Saadatmandi C, Dose J, Schultheiss G, Rimbach G. Anti-inflammatory potential of allyl-isothiocyanate--role of Nrf2, NF-(κ) B and microRNA-155. *J Cell Mol Med* 2012; **16**: 836-843 [PMID: 21692985 DOI: 10.1111/j.1582-4934.2011.01367.x]

25 **Ahn J**, Lee H, Im SW, Jung CH, Ha TY. Allyl isothiocyanate ameliorates insulin resistance through the regulation of mitochondrial function. *J Nutr Biochem* 2014; **25**: 1026-1034 [PMID: 25034503 DOI: 10.1016/j.jnutbio.2014.05.006]

26 **Okulicz M**. Multidirectional time-dependent effect of sinigrin and allyl isothiocyanate on metabolic parameters in rats. *Plant Foods Hum Nutr* 2010; **65**: 217-224 [PMID: 20809411 DOI: 10.1007/s11130-010-0183-3]

27 **Kim YJ**, Lee DH, Ahn J, Chung WJ, Jang YJ, Seong KS, Moon JH, Ha TY, Jung CH. Pharmacokinetics, Tissue Distribution, and Anti-Lipogenic/Adipogenic Effects of Allyl-Isothiocyanate Metabolites. *PLoS One* 2015; **10**: e0132151 [PMID: 26317351 DOI: 10.1371/journal.pone.0132151]

28 **Shimano H**, Sato R. SREBP-regulated lipid metabolism: convergent physiology - divergent pathophysiology. *Nat Rev Endocrinol* 2017; **13**: 710-730 [PMID: 28849786 DOI: 10.1038/nrendo.2017.91]

29 **Pfluger PT**, Herranz D, Velasco-Miguel S, Serrano M, Tschöp MH. Sirt1 protects against high-fat diet-induced metabolic damage. *Proc Natl Acad Sci U S A* 2008; **105**: 9793-9798 [PMID: 18599449 DOI: 10.1073/pnas.0802917105]

30 **Li Y**, Xu S, Giles A, Nakamura K, Lee JW, Hou X, Donmez G, Li J, Luo Z, Walsh K, Guarente L, Zang M. Hepatic overexpression of SIRT1 in mice attenuates endoplasmic reticulum stress and insulin resistance in the liver. *FASEB J* 2011; **25**: 1664-1679 [PMID: 21321189 DOI: 10.1096/fj.10-173492]

31 **Yu J**, Chu ES, Wang R, Wang S, Wu CW, Wong VW, Chan HL, Farrell GC, Sung JJ. Heme oxygenase-1 protects against steatohepatitis in both cultured hepatocytes and mice. *Gastroenterology* 2010; **138**: 694-704, 704.e1 [PMID: 19818781 DOI: 10.1053/j.gastro.2009.09.058]

32 **Li S**, Dou X, Ning H, Song Q, Wei W, Zhang X, Shen C, Li J, Sun C, Song Z. Sirtuin 3 acts as a negative regulator of autophagy dictating hepatocyte susceptibility to lipotoxicity. *Hepatology* 2017; **66**: 936-952 [PMID: 28437863 DOI: 10.1002/hep.29229]

33 **Miyata S**, Inoue J, Shimizu M, Sato R. Allyl isothiocyanate suppresses the proteolytic activation of sterol regulatory element-binding proteins and de novo fatty acid and cholesterol synthesis. *Biosci Biotechnol Biochem* 2016; **80**: 1006-1011 [PMID: 26822063 DOI: 10.1080/09168451.2015.1132154]

34 **Lagouge M**, Argmann C, Gerhart-Hines Z, Meziane H, Lerin C, Daussin F, Messadeq N, Milne J, Lambert P, Elliott P, Geny B, Laakso M, Puigserver P, Auwerx J. Resveratrol improves mitochondrial function and protects against metabolic disease by activating SIRT1 and PGC-1alpha. *Cell* 2006; **127**: 1109-1122 [PMID: 17112576 DOI: 10.1016/j.cell.2006.11.013]

35 **Hou X**, Xu S, Maitland-Toolan KA, Sato K, Jiang B, Ido Y, Lan F, Walsh K, Wierzbicki M, Verbeuren TJ, Cohen RA, Zang M. SIRT1 regulates hepatocyte lipid metabolism through activating AMP-activated protein kinase. *J Biol Chem* 2008; **283**: 20015-20026 [PMID: 18482975 DOI: 10.1074/jbc.M802187200]

36 **Li YP**, Wang SL, Liu B, Tang L, Kuang RR, Wang XB, Zhao C, Song XD, Cao XM, Wu X, Yang PZ, Wang LZ, Chen AH. Sulforaphane prevents rat cardiomyocytes from hypoxia/reoxygenation injury in vitro via activating SIRT1 and subsequently inhibiting ER stress. *Acta Pharmacol Sin* 2016; **37**: 344-353 [PMID: 26775664 DOI: 10.1038/aps.2015.130]

37 **Hotamisligil GS**. Inflammation and metabolic disorders. *Nature* 2006; **444**: 860-867 [PMID: 17167474 DOI: 10.1038/nature05485]

38 **Perkins ND**. Integrating cell-signalling pathways with NF-kappaB and IKK function. *Nat Rev Mol Cell Biol* 2007; **8**: 49-62 [PMID: 17183360 DOI: 10.1038/nrm2083]

39 **Cai D**, Yuan M, Frantz DF, Melendez PA, Hansen L, Lee J, Shoelson SE. Local and systemic insulin resistance resulting from hepatic activation of IKK-beta and NF-kappaB. *Nat Med* 2005; **11**: 183-190 [PMID: 15685173 DOI: 10.1038/nm1166]

40 **Murata M**, Yamashita N, Inoue S, Kawanishi S. Mechanism of oxidative DNA damage induced by carcinogenic allyl isothiocyanate. *Free Radic Biol Med* 2000; **28**: 797-805 [PMID: 10754276]

41 **Tarantino G**, Porcu C, Arciello M, Andreozzi P, Balsano C. Prediction of carotid intima-media thickness in obese patients with low prevalence of comorbidities by serum copper bioavailability. *J Gastroenterol Hepatol* 2018; **33**: 1511-1517 [PMID: 29405466 DOI: 10.1111/jgh.14104]

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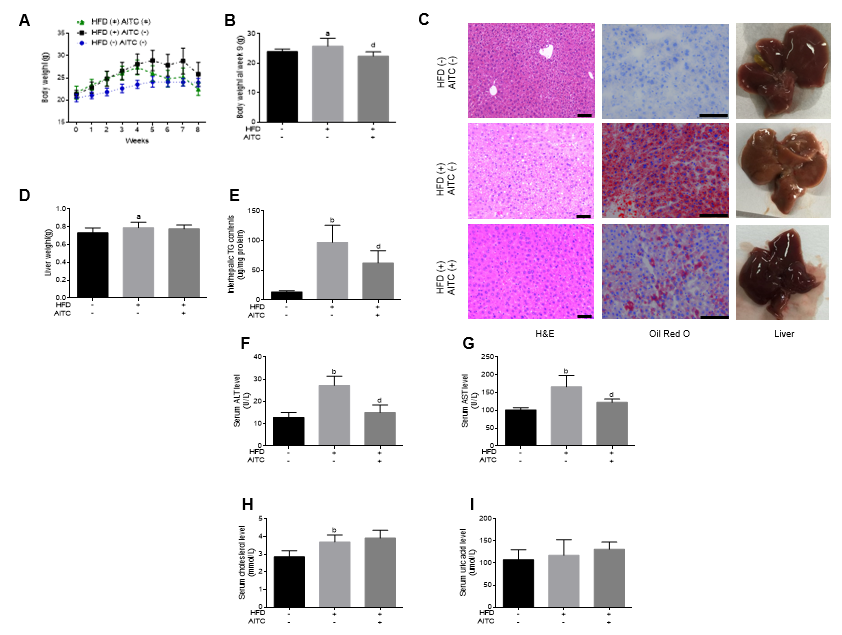
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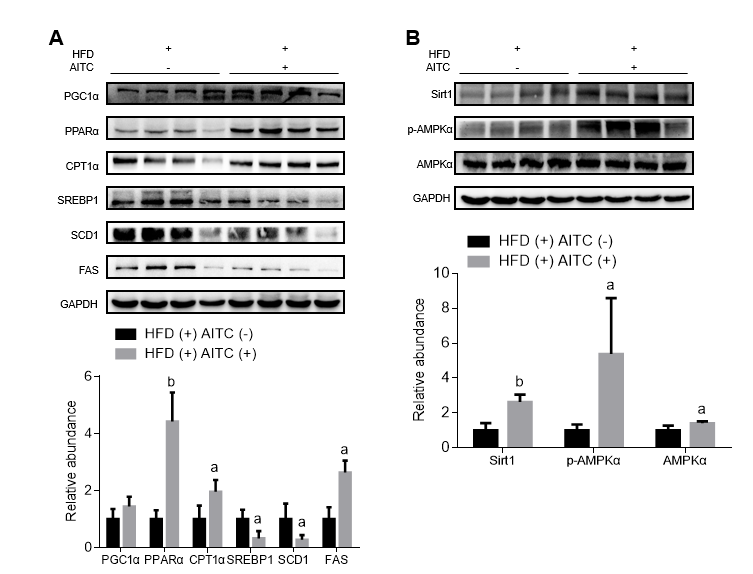
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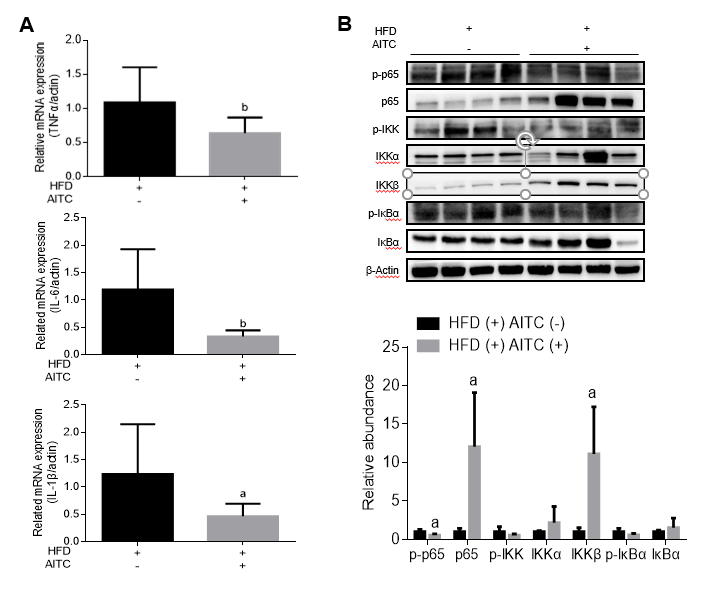
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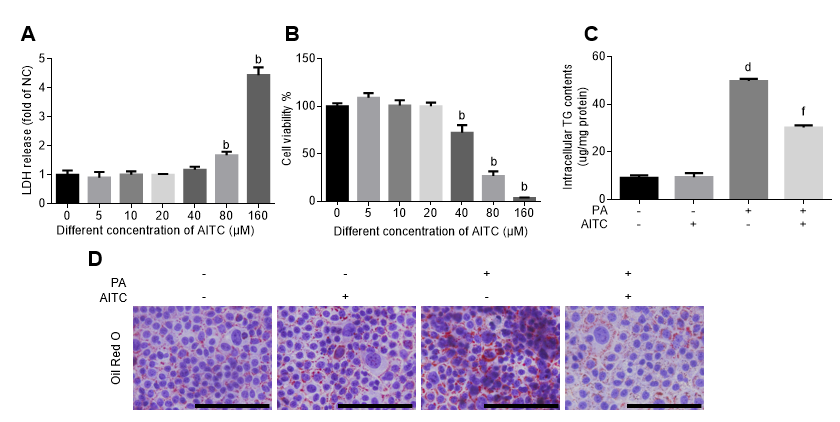
**Figure 1 Allyl isothiocyanate reduces body weight, ameliorates hepatic steatosis and attenuates liver injury in high fat diet-fed mice.** A: Body weight evaluated weekly; B: Body weight at week 8; C: Representative liver sections stained with hematoxylin and eosin (H&E) (left panel) or oil red O (middle panel) and macroscopic pictures of livers (right panel). D: Liver weight; E: Intrahepatic triglyceride (TG) content. Liver function was evaluated by detecting serum levels of alanine aminotransferase (ALT) (F), aspartate aminotransferase (AST) (G), total cholesterol (H) and uric acid (I). Scale bar in panel represents 100 μm. Data are presented as the mean ± S.D. a*P* < 0.05, b*P* < 0.01 *vs* HFD(-) AITC(-), d*P* < 0.01 *vs* HFD(+) AITC(-). HFD: High fat diet; AITC: Allyl isothiocyanate; TG: Triglyceride; H&E: Hematoxylin and eosin; ALT: Alanine aminotransferase; AST: Aspartate aminotransferase.



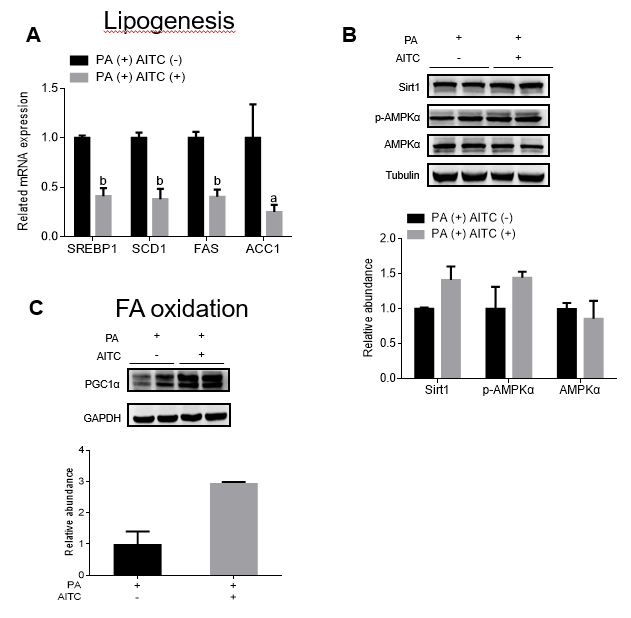
**Figure 2 Allyl isothiocyanate upregulates the expression of proteins involved in fatty acid β-oxidation, downregulates the protein levels of lipogenesis genes, and activates the Sirtuin 1/AMP-activated protein kinase signaling pathway in the liver tissues of high fat diet-fed mice.** A: The protein expression of sterol regulatory element­binding protein 1 (SREBP1), its lipogenesis target genes (*SCD1* and *FAS*), and genes involved in fatty acid β-oxidation, such as proliferator-activated receptor gamma coactivator 1α (PGC1α), peroxisome proliferator-activated receptor α (PPARα) and carnitine palmitoyltransferase 1 α (CPT1α), was detected by western blot analysis. B: Sirtuin 1 (Sirt1), total and phosphorylated AMP-activated protein kinase α (AMPKα) protein expression was detected by western blot analysis. a*P* < 0.05, b*P* < 0.01 *vs* HFD(+) AITC(-). PGC1α: Proliferator-activated receptor gamma coactivator 1α; PPARα: Peroxisome proliferator-activated receptor α; CPT1α: Carnitine palmitoyltransferase 1 α; SREBP1: Sterol regulatory element­binding protein 1; SCD1: Stearoyl coenzyme A desaturase 1; FAS: Fatty acid synthase; Sirt1: Sirtuin 1; AMPKα: AMP-activated protein kinase α; p-AMPKα: Phosphorylated AMP-activated protein kinase α; HFD: High fat diet; AITC: Allyl isothiocyanate.



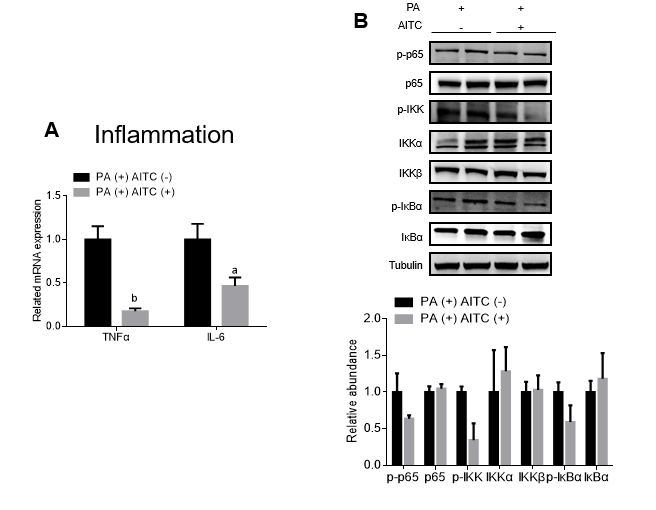
**Figure 3 Allyl isothiocyanate attenuates hepatic inflammation and inhibits the I*κB* kinase /nuclear factor kappa B signaling pathway in the liver tissues of high fat diet-fed mice.** A: The mRNA levels of proinflammatory cytokines in the liver of high fat diet (HFD)-fed control (*n* = 9) and HFD-fed allyl isothiocyanate (AITC)-treated mice (*n* = 10) were measured by quantitative real-time PCR. B: The protein expression of phosphorylated p65, p65, phosphorylated IκB kinase (IKK), IKKα, IKKβ, total and phosphorylated inhibitor of nuclear factor kappa B α (IκB α) in the liver was detected by western blot analysis. Data are presented as the mean ± S.D. a*P* < 0.05, b*P* < 0.01 *vs* HFD(+) AITC(-). TNFα: Tumor necrosis factor α; IL-6: Interleukin-6; IL-1β: Interleukin-1β; HFD: High fat diet; AITC: Allyl isothiocyanate; p-p65: Phosphorylated p65; IKK: IκB kinase; IκBα: Inhibitor of nuclear factor kappa B α.



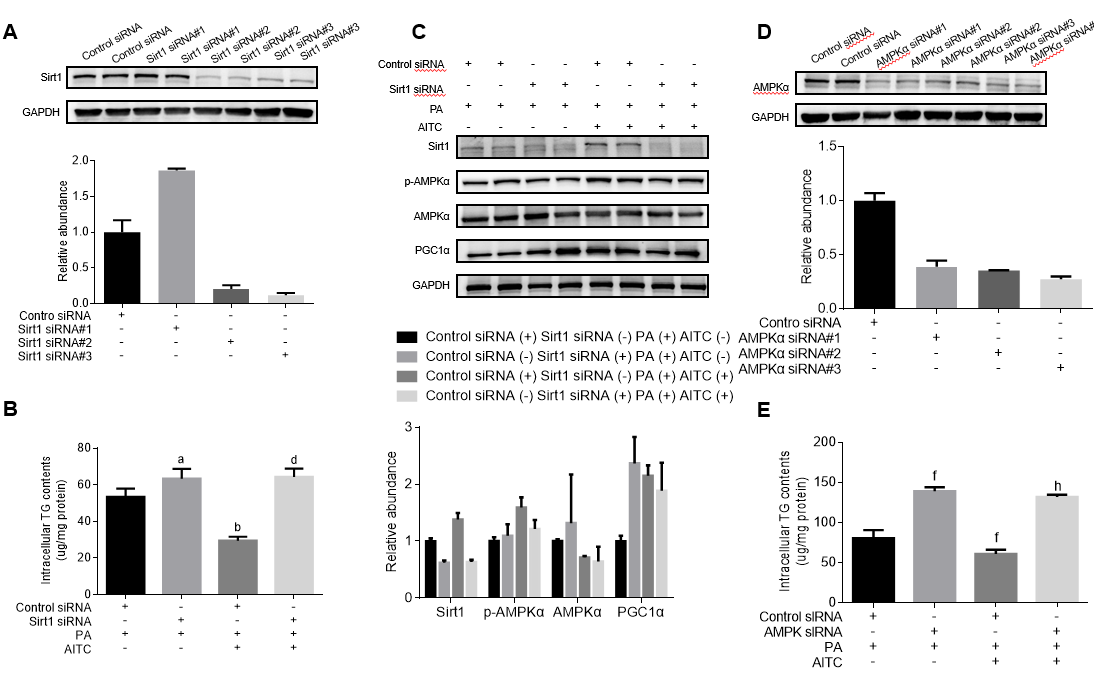
**Figure 4 Allyl isothiocyanate alleviates palmitate acid-induced lipid accumulation *in vitro*. Palmitate acid (200 μmo/L)-stimulated AML-12 cells were treated with allyl isothiocyanate (20 μmo/L) or dimethyl sulfoxide (vehicle) for 24 h.** A: Cytotoxicity was measured by the lactate dehydrogenase (LDH) release method in AML-12 cells (*n* = 3/group). B: Cell viability was measured using the cholecystokinin-8 (CCK-8) assay in AML-12 cells (*n* = 3/group). C: Intracellular triglyceride (TG) content in AML-12 cells (*n* = 3/group). And D: Representative image of oil red O staining of AML-12 cells in different groups. Scale bar in panel represents 100 μm. Data are presented as the mean ± SD. b*P* < 0.01 *vs* 0 μmol/L AITC, d*P* < 0.01 *vs* PA(-) AITC(-), f*P* < 0.01 *vs* PA(+) AITC(-). LDH: Lactate dehydrogenase; CCK-8: Cholecystokinin-8; TG: Triglyceride; NC: Negative control; PA: Palmitate acid: AITC: Allyl isothiocyanate.



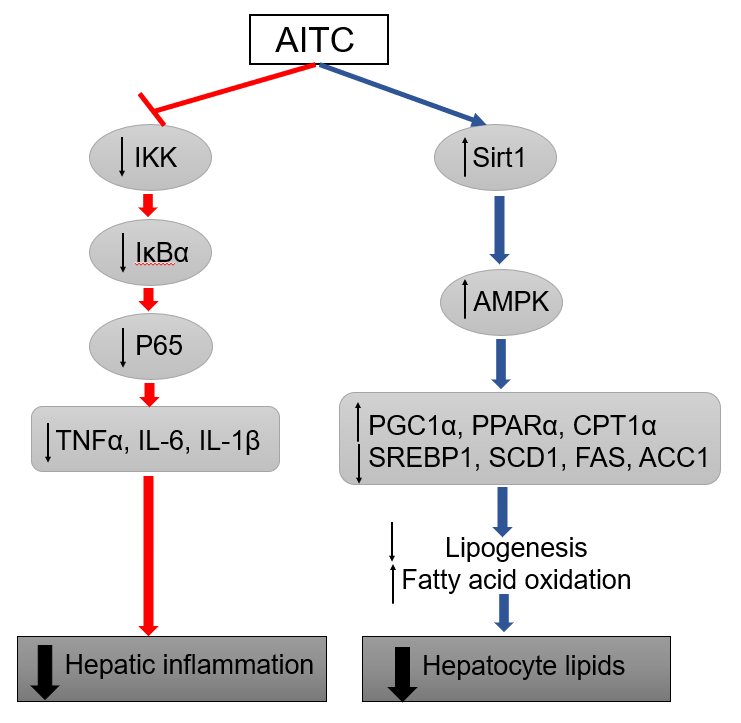
**Figure 5 Allyl isothiocyanate downregulates the mRNA levels of genes involved in lipogenesis, upregulates the mRNA levels of genes involved in fatty acid β-oxidation and activates the Sirtuin 1/AMP-activated protein kinase signaling pathway *in vitro*.** A: Palmitate acid (PA) (200 μmol/L)-stimulated AML-12 cells were treated with allyl isothiocyanate (AITC) (20 μmol/L) or dimethyl sulfoxide (DMSO) (vehicle) for 6 h (*n* = 3/group). The mRNA levels of sterol regulatory element­binding protein 1 (SREBP1) and its lipogenesis target genes, including stearoyl coenzyme A desaturase 1 (SCD1), fatty acid synthase (FAS) and acetyl-CoA carboxylase 1 (ACC1), were determined. B: PA (200 μmol/L)-stimulated AML-12 cells were treated with AITC (20 μmol/L) or DMSO (vehicle) for 24 h. The protein expression of Sirtuin 1 (Sirt1), total and phosphorylated AMP-activated protein kinase α (AMPKα) was detected by western blot analysis. C: PA (200 μmol/L)-stimulated AML-12 cells were treated with AITC (20 μmol/L) or DMSO (vehicle) for 24 h. The protein level ofproliferator-activated receptor gamma coactivator1α (PGC1α), which is involved in fatty acid β-oxidation, was detected by western blot analysis. Data are presented as the mean ± SD. a*P* < 0.05, b *P* < 0.01 *vs* PA(+) AITC(-). PA: Palmitate acid; AITC: Allyl isothiocyanate; PGC1α: Proliferator-activated receptor gamma coactivator 1α; SREBP1: Sterol regulatory element­binding protein 1; SCD1: Stearoyl coenzyme A desaturase 1; FAS: Fatty acid synthase; ACC1: Acetyl-CoA carboxylase 1; Sirt1: Sirtuin 1; AMPKα: AMP-activated protein kinase α; p-AMPKα: Phosphorylated AMP-activated protein kinase α.



**Figure 6 Allyl isothiocyanate downregulates the mRNA levels of proinflammatory markers and inhibits the I*κB* kinase /nuclear factor kappa B signaling pathway *in vitro*.** A: Palmitate acid (PA) (200 μmol/L)-stimulated AML-12 cells were treated with allyl isothiocyanate (AITC) (20 μmol/L) or dimethyl sulfoxide (DMSO) (vehicle) for 6 h (*n* = 3/group). The mRNA levels of proinflammatory cytokines were measured by quantitative real-time PCR. (B) PA (200 μmol/L)-stimulated AML-12 cells were treated with AITC (20 μmol/L) or DMSO (vehicle) for 24 h. The protein expression of phosphorylated p65, p65, phosphorylated IκB kinase (IKK), IKKα, IKKβ, total and phosphorylated inhibitor of nuclear factor kappa B α (IκB α) was detected by western blot analysis. Data are presented as the mean ± S.D. a*P* < 0.05, b *P* < 0.01 *vs* PA(+) AITC(-). TNFα: Tumor necrosis factor α; IL-6: Interleukin-6; PA: Palmitate acid; AITC: Allyl isothiocyanate; p-p65: Phosphorylated p65; IKK: IκB kinase; IκBα: Inhibitor of nuclear factor kappa B α.



**Figure 7 Allyl isothiocyanate ameliorates lipid accumulation by activating the Sirt1/AMPKα signaling pathway.** After transfection with Sirtuin 1 (Sirt1) small interfering RNA (siRNA), AMP-activated protein kinase α (AMPKα) siRNA or the corresponding scrambled control for 48 h, AML-12 cells were incubated in normal medium or medium containing palmitate acid (PA) with or without allyl isothiocyanate (AITC) for 24 h. A: Three different Sirt1 siRNA sequences were used, and Sirt1 protein levels were examined by western blot analysis. In the following experiments, we selected Sirt1 siRNA #3. B: Intracellular triglyceride (TG) content in AML-12 cells (*n* = 4/group). C: Sirt1, phosphorylated AMPKα and proliferator-activated receptor gamma coactivator1α (PGC1α) protein expression was detected by western blot analysis. D: Three different AMPKα siRNA sequences were used, and AMPKα protein levels were examined by western blot analysis. In the subsequent experiments, we selected AMPKα siRNA #3. (E) Intracellular TG content in AML-12 cells (*n* = 4/group). Data are presented as the mean ± SD. a*P* < 0.05, b*P* < 0.01 *vs* Control siRNA + PA, d*P* < 0.01 *vs* Control siRNA + PA + AITC, f*P* < 0.01 *vs* Control siRNA + PA, h*P* < 0.01 *vs* Control siRNA + PA + AITC. PA: Palmitate acid; AITC: Allyl isothiocyanate; PGC1α: Proliferator-activated receptor gamma coactivator 1α; Sirt1: Sirtuin 1; AMPKα: AMP-activated protein kinase α; p-AMPKα: Phosphorylated AMP-activated protein kinase α; TG: Triglyceride.



**Figure 8 Model of allyl isothiocyanate action.** Schematic diagram: allyl isothiocyanate ameliorates hepatic lipid accumulation and hepatic inflammation by activating theSirt1/AMPK signaling pathway and inhibiting the NF-κBpathway. AITC: Allyl isothiocyanate; IKK: IκB kinase; IκBα: Inhibitor of nuclear factor kappa B α; TNFα: Tumor necrosis factor α; IL-6: Interleukin-6; IL-1β: Interleukin-1β; Sirt1: Sirtuin 1; AMPKα: AMP-activated protein kinase α; PGC1α: Proliferator-activated receptor gamma coactivator 1α; PPARα: Peroxisome proliferator-activated receptor α; CPT1α: Carnitine palmitoyltransferase 1 α; SREBP1: Sterol regulatory element­binding protein 1; SCD1: Stearoyl coenzyme A desaturase 1; FAS: Fatty acid synthase; ACC1: Acetyl-CoA carboxylase 1.