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***Basic Study***

**High expression of *APC* is an** **unfavorable prognostic biomarker** **in** **T4** **gastric cancer patients**

Du WB *et al. APC* in T4 gastric cancer

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**Abstract**

***Background***

Adenoma polyposis coli (*APC*) mutation is associated with tumorigenesis *via* the Wnt signaling pathway.

***Aim***

To investigate the clinical features and mechanism of *APC* expression in gastric cancer (GC).

***Methods***

Based on *APC* expression profile, the related genome-wide mRNA expression, microRNA (miRNA) expression, and methylation profile in GC, the relationship between *APC* and GC, as well as the prognostic significance of *APC* were systematically analyzed by multi-dimensional methods.

***Results***

We found that high expression of *APC* (*APC*high) was significantly associated with adverse outcomes of T4 GC patients. Genome-wide gene expression analysis revealed that varying *APC* expression levels in GC were associated with some important oncogenes, and corresponding cellular functional pathways. Genome-wide miRNA expression analysis indicated that most of miRNAs associated with high *APC* expression were downregulated. The mRNA-miRNA regulatory network analysis revealed that down-regulated miRNAs affected their inhibitory effect on tumor genes. Genome-wide methylation profiles associated with *APC* expression showed that there was differential methylation between the *APC*high and *APC*low groups. The number of hypermethylation sites was larger than that of hypomethylation sites, and most of hypermethylation sites were enriched in CpG islands.

***Conclusion***

Our research demonstrated that high *APC* expression is an unfavorable prognostic factor for T4 GC patients and may be used as a novel biomarker for pathogenesis research, diagnosis, and treatment of GC.

**Key words:** Adenoma polyposis coli; Gastric cancer; Prognosis; mRNA; miRNA; Methylation

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**Core tip:** We found that high expression of adenoma polyposis coli (*APC*) was associated with a poor prognosis in T4 gastric cancer (GC) patients. There was differential expression of mRNAs and miRNAs as well as differential DNA methylation between the high expression of *APC* (*APC*high) and low expression of *APC* (*APC*low) groups. The link between *APC*high and differential expression of mRNAs, miRNAs, and DNA methylation may contribute to the poor prognosis in T4 GC patients, and be involved in the pathogenesis of GC. *APC* could be used as a novel biomarker for clinical diagnosis, therapy, and assessment of prognosis in T4 GC patients, as well as for further research of the pathogenesis of GC.

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**INTRODUCTION**

Gastric cancer (GC) is one of the most common malignant tumors in China, and its morbidity and mortality rank first in digestive tract tumors[1]. As the early detection rate of GC is low, most patients are initially diagnosed at an advanced stage, which seriously affects the prognosis and survival rate of the patients. With the development of endoscopic technique and the improvement of medical technology, the 5-year survival rate of GC has increased to 90%[2, 3]. However, recurrence, metastasis, and drug resistance limit the therapeutic effect. There are still about 350000 deaths due to GC in China every year[4]. Therefore, the early prevention, diagnosis, and anti-tumor comprehensive treatment of GC have attracted more and more attention.

GC is a complex multi-factorial, multi-step progressive process involving coding or non-coding genes and epigenetic change[5]. Many kinds of molecular events, such as DNA mutations[6], aberrant expression of mRNAs[7] and microRNAs (miRNAs)[8], and abnormal proteomes[9], have been found to participate in the pathogenesis of GC. Currently, these molecular events have been extensively studied as biomarkers for the prognosis, diagnosis, and therapy of GC. Li *et al*[10] found that *MUC16* gene mutation was associated with a higher load of tumor mutations and improved prognosis in patients with GC. These findings may have an impact on prognosis prediction and treatment guidance for GC. MiR-1265 inhibits the progression and carcinogenic autophagy of GC by reducing the expression of CAB39 and regulating the AMPK-mTOR signal transduction pathway. Therefore, it was speculated that miR-1265 may represent a potential therapeutic target for GC[11]. DNA methylation is common in non-neoplastic gastric mucosa infected with *Helicobacter pylori*, which is associated with a high risk of GC. Therefore, DNA methylation could be used as a useful diagnostic tool for GC risk assessment[12]. Despite these, the molecular mechanism of GC is currently still unclear, and the identification of biomarkers is of great significance for the clinical prognosis, diagnosis, and treatment of GC.

Adenoma polyposis coli(*APC*) encodes a tumor suppressor protein that can inhibit the Wnt signal pathway in tumor, and mutation and inactivation of *APC* are unique key and early events in tumorigenesis, especially colorectal cancer. The diagnostic and prognostic value of *APC* in colorectal cancer is high[13,14]. Compared with colorectal cancer, there have been much fewer studies on *APC* in GC. However, some successful studies have initially shown that *APC* could be used as a biomarker for GC. Through research and analysis of big data, Bria *et al*[15] defined a risk classification system comprising biomolecular (including *APC*) and clinicopathological predictors, which could divide resected GC patients into three types of risk for guiding treatment. By statistically analyzing a large number of GC samples, Merchant *et al*[16] revealed that the incidence of *APC*-related GC in young Hispanic men has been rising in the United States. And the high expression of *APC* was correlated with advanced stage and low differentiation of GC. Therefore, we speculated that *APC* may have an important role in the pathogenesis and clinical application of GC.

In this study, we demonstrated that *APC* was a biomarker for poor prognosis in T4 GC patient, and identified *APC*-related mRNA and miRNA expression changes, and DNA methylation profile on a genome-wide scale by multi-dimensional methods. Our findings indicated the value of *APC* as a new target for the diagnosis and treatment of T4 GC patients and highlighted the important role of *APC* in the pathogenesis of GC.

**MATERIALS AND METHODS**

***Research data source***

The research data, including high throughput RNA sequencing (RNA-Seq), miRNA-Seq, Illumina Infinium Human Methylation 450, and clinical follow-up information data, which provided all mRNA and miRNA expression and methylation profiles for followed GC patients, were downloaded from the Cancer Genome Atlas (TCGA)[17] on October 31, 2018. Samples with more than 30 d of follow-up were screened from clinical follow-up data to further match the RNAseq expression profile, miRNA expression profile, and methylation profile. We obtained 387 samples and 2173 miRNAs, 335 samples and 19754 mRNAs, and 364 samples and 358418 methylation sites, respectively. The study was approved by the Clinical Research Ethics Committee of College of Medicine, Zhejiang University.

***Statistical analysis***

Overall survival (OS) was defined as the time from randomization to death for any reason, while relapse-free survival rate (RFS) was defined as complete remission (approximately one month after diagnosis) to recurrence or the end of follow-up. In order to evaluate the prognostic value of *APC* in GC samples, we selected the appropriate cut-off value for subdivision. The optimal cut-off was defined by the statistical distribution of *APC* expression. *APC* expression was normally distributed in GC samples, and there was a significantly evident distinction along the median value. Therefore, GC samples were divided into *APC*high and *APC*low groups by the median *APC* expression (Supplemental Figure 1A). Next, the T4 GC patients were classified in to four groups according to quartiles of *APC* expression: Q1, <25%; Q2, 25%–50%; Q3, 50%–75%; Q4, >75%). The results showed that Q4 had the worst prognosis both in OS and RFS. Also, a significant difference was observed between Q2 *vs* Q4 and Q2 *vs* Q4 in OS and RFS (Supplemental Figure 1B and 1C). Then, the relationship between *APC* expression and OS and RFS was evaluated by the Kaplan-Meier (KM) method and further examined by log-rank test. To investigate the association between *APC* and clinical and molecular characteristics, we divided the T4 GC patients into *APC*high and *APC*low groups according to median *APC* expression value and used the *t*-test and chi-square tests to compare the clinical and molecular characteristics between the two groups. The *t*-test and false discovery rate (FDR) were applied to reveal the differential expression of genome-wide genes, miRNAs, and methylation between *APC*high and *APC*low groups. The significantly statistical cut-off values were fold-change (FC) > 2 and FDR < 0.05 for genes, FC > 1.5 or < 0.66 and *P* < 0.05 for miRNAs, and FC > 2 and *P* < 0.05 for methylation. In order to observe the relationship between *APC* expression and cellular functional pathways based on the differential expression of genome-wide genes, we screened all GC samples with gene expression profiles, and used Gene Set Enrichment Analysis (GSEA) tools[18] to perform KEGG pathway enrichment analysis on samples from *APC*high and *APC*low groups with a selection threshold of *P* < 0.05. KEGG pathway enrichment of methylation was performed using R ClusterProfiler with a selection threshold of *P* < 0.05. To observe the relationship between *APC* expression and related miRNAs, we screened miRNAs with expression levels greater than 0 and ratios greater than 50% in each sample, and used the Pearson correlation coefficient to estimate the correlation between miRNA expression profiles and *APC* expression. The significantly statistical cut-off was *P* < 0.05 and a correlation coefficient > 0.3 or < -0.3 to determine the correlation of *APC* with related miRNAs. In additional, we used the miRWalk database to predict miRNA target genes and constructed mRNA-miRNA regulatory networks in conjunction with the most significantly related miRNAs.

**RESULTS**

***APC is related to an unfavorable prognosis*** ***of T4*** ***GC***

Analysis of OS and RFS was performed based on the 335 GC samples to investigate the association between *APC* expression and prognosis. The results showed that there was no significant relationship between *APC* expression and OS (*P* = 0.58) and RFS (*P* = 0.98) in the overall population (Figure 1A and 1B). Subsequently, we analyzed the association between *APC* expression and prognosis in GC of different stages and found that the expression of *APC* in T4 samples was high in the middle and low on both sides with a median value of 10.53 (Figure 1C). According to this median value, the 88 T4 GC samples were divided into either an *APC*high (*n* = 44)or *APC*low group (*n* = 44). It was found that patients with *APC*high GC showed significantly shorter RFS than those with *APC*low GC (log-rank *P* = 0.037) (Figure 1D). These results suggested that high expression of *APC* was associated with a poor prognosis in T4 GC patients.

***Association*** ***of APC*** ***with clinical characteristics and known prognostic genes***

The proportions of GC patients with tumor stages III and IV, tumor grades 2 and 3, and lymph node metastases N2 and N3 were significantly higher in the *APC*high group than in the *APC*low group (*P* = 0.043, 0.047, 0.039, 0.031, and 0.035, respectively). In addition, *APC*high was associated with a greater chance of high expression of some known prognostic biomarkers, such as *KIT*, *PIK3CA*, *KRAS*, and *MLH1* (*P* = 0.031, *P* < 0.001, *P* = 0.038, and *P* = 0.038, respectively), all of which are associated with an unfavorable prognosis (Table 1). *Kit*, a cell surface receptor of stem cell factor, is considered to be a oncogenic signal and has a acquired functional mutation in human gastrointestinal stromal tumors[19]. The frequency of *PIK3CA* mutation was low in GC, but the prognosis of GC patients with *PIK3CA* mutation was poor[20]. Fu *et al*[21]found that *KRAS* mutation was a poor prognostic factor in Chinese CG patients. *MLH1* methylation was associated with oxaliplatin resistance in GC patients[22]. Collectively, these results indicated that *APC* may be a biomarker of poor prognosis in T4 GC patients.

***Differentially expressed mRNAs associated with APC expression***

To observe the association between *APC* expression and other genes, we made a comparison between the *APC*high and *APC*low groups by genome-wide microarray analysis. Twelve significantly differentially expressed mRNAs were revealed to be associated with *APC* expression by the *t*-test and Wilcoxon rank sum test (FC > 2 and FDR < 0.05) (Figure 2A, Table 2). The heat map of these 12 mRNAs is presented in Figure 2B. Previous studies have revealed that most of these 12 mRNAs are closely related to tumorigenesis. *ARID5B* plays a role in the growth and differentiation of B-lymphocyte progenitor cells that are associated with acute lymphoblastic leukemia[23]. *JMY* has strong carcinogenic characteristics, whose expression was elevated in primary colorectal cancer and head and neck cancer in response to DNA damage in cells[24]. *MAP3K14* is involved in the induction of NF-kappaB signaling in cancer cell invasion[25]. *IL6ST* is a signal transduction factor common to many cytokine activators, and it is involved in the STAT3 pathway in sub-groups of primary lung cancer and is associated with tumor progression and a poor prognosis[26]. *Rev3L* plays an important role in regulating the response of cervical cancer cells to cisplatin[27]. Genetic variation of estrogen receptor *PPARGC1B* is important in familial breast cancer[28]. *PARD3* plays an important role in invasiveness and metastasis of lung squamous cell carcinoma because of its repeated inactivation of cell polarity regulator[29]. *PDE4D* acts as a promoter of cell proliferation in prostate cancer[30].

In order to further analyze the cell molecular function of *APC* related genes, we investigated the association between cellular functional pathways of *APC* related genes with *APC* expression. The results revealed that 18 pathways were associated with *APC*high, including 6 upregulated and 12 downregulated pathways (*P* < 0.05) (Table 3). The *APC* related genes were enriched in upregulated pathways such as receptor signaling (Figure 3A), pancreatic cancer (Figure 3B), and prostate cancer of upregulated pathway, whereas downregulated pathways such as ascorbate and aldarate metabolism (Figure 3C), PPAR signaling (Figure 3D), drug metabolism other enzymes, and steroid hormone biosynthesis are closely related to tumor signaling, drug resistance, hormone receptor, and metabolism. These results indicated that *APC*high associated genes are closely related to the occurrence and development of GC.

***Relationship between*** ***APChigh expression and genome-wide miRNA sequencing***

In order to further understand the relationship between biological diversities of GC and different levels of *APC* expression, we conducted a full-genome analysis of miRNA sequencing data to identify the miRNA profiles with a significant correlation with *APC* expression. We found that 674 miRNA expression profiles were correlated with *APC* expression. After Pearson correlation coefficient calculation, a total of 35 miRNAs with a *P*-value< 0.05 and a correlation coefficient > 0.3 or < -0.3 were identified. The relationship between the correlation coefficient and the distribution of *P-*value is shown in Figure 4A. The heatmap of the 35 miRNAs is presented in Figure 4B, from which we can see that most of miRNA were negatively correlated with *APC*, and a few were positively correlated with *APC*. Most of the 35 miRNAs have been previously found to be involved in tumorigenesis mechanisms or be targets for clinical diagnosis and treatment, prognostic markers, and mediators of tumor drug resistance. Hsa-miR-636 may affect the tumorigenesis of hepatocellular carcinoma by downregulating the Rassignaling pathway[31]. The enrichment of hsa-miR-200c-3p secreted in urine was a non-invasive marker for the diagnosis of endometrial carcinoma[32]. The downregulated expression of has-miR-130b-3p was predicted to be associated with cell proliferation, colony formation, anti-apoptosis, and self-renewal in hepatocellular carcinoma[33]. Hsa-miR-339-5p plays a key role in the regulation of lung cancer and may be used as a biomarker to predict the progression of lung cancer[34]. The upregulated expression of miR-451 in papillary thyroid carcinoma with lymph node metastasis is a unique marker for prognosis and progression of thyroid papillary carcinoma[35]. Increasing expression of hsa-miR-103a-3p predicted improved breast cancer survival[36]. Overexpression of miR-940 in breast cancer cells induced extensive osteoblast damage in metastatic bone tumors by promoting osteogenic differentiation of host mesenchymal cells[37]. LncRNA LINC00460 could upregulate IL6 through sponging tumor suppressor gene miR-149-5p to promote the carcinogenesis of nasopharyngeal carcinoma[38]. Ectopic expression of has-let-7c-5p promoted the efficacy of all-trans retinoic acid in the treatment of acute myeloid leukemia cells[39]. Low expression of miR-100-5p endowed lung cancer cells with cisplatin drug resistance[40]. These *APC* related miRNAs, together with *APC*, may be associated with the pathogenesis of GC, which further supports *APC* as a marker for the poor prognosis of T4 GC.

Furthermore, we combined the most significantly related miRNAs, using the miRWalk database, to predict the miRNA target genes to construct the mRNA-miRNA regulatory network (Figure 4C). The regulatory network included 6 upregulated miRNAs, 24 downregulated miRNAs, and 11 genes. It can be seen that some downregulated miRNAs may affect the inhibition of oncogenes, thus aggravating the result of deterioration. For example, hsa-miR-200c-3p, a member of miR-200 family, targets *ARID5B* and plays an important role in inhibiting tumor epithelial-mesenchymal transition (EMT)[41]. Has-miR-940 targeted *PDE4D* and *ZNF81* were downregulated in prostate cancer and can inhibited the migratory and invasive potential of cancer cell[42].In the meantime, some upregulated miRNAs decreased the function of tumor suppressor genes, thus promoting tumorigenesis. For example, *ARID5B* is targeted by has-miR-150-5p, and the upregulated expression of has-miR-150-5p increased the response of multiple myeloma to glucocorticoid specific therapy[43]. Hsa-miR-195-3p, regarded as one of colon cancer metastasis biomarkers, targeted *PARD3B* and other genes that are associated with protein binding, cell adhesion, and cancer metastasis[44]. Taken together, these findings may contribute to understanding why *APC*high could predict a poor prognosis of T4 GC.

***Relationship between*** ***APChigh and genome-wide*** ***DNA methylation sequencing***

DNA methylation is a common and important epigenetic change and plays an important role in the occurrence, development, and prognosis of many malignant tumors. Also, DNA methylation can regulate gene expression and the differentiation of cancer cells to different directions *via* DNA methyltransferase, including DNMT1, DNMT3A, and DNMT3B[45]. Therefore, we analyzed the differences in transcription levels of the three methyltransferase between *APC*high and *APC*low. DNMT1 and DNMT3A were found to be significantly higher in *APC*high than in *APC*low (Figure 5), which suggested that there may be different methylation patterns in *APC*high and *APC*low. Five hundred and forty-three differential methylation sites, including 530 hypermethylation sites and 13 hypomethylation sites, were identified after making a comparison between *APC*high and *APC*low (FC > 2 and *P* < 0.05) (Figure 6A). Since hypermethylation of tumor suppressor genes and hypomethylation of oncogenes were associated with carcinogenesis[46], our finding that there were more hypermethylation sites than hypomethylation sites may suggest that DNA hypermethylationregulated *APC* expression is involved in T4 GC. In addition, we analyzed the distribution of these 543 different methylation sites around CPG islands, and found that there was a significant difference in the distribution of differential DNA methylation sites around CPG islands, *i.e.*, the hypermethylation sites are mainly distributed on the CPG islands (Figure 6B), which is consistent with the view that the methylation of DNA methylation-regulated genes involved in tumorigenesis occurs mainly on CPG islands[47]. Furthermore, we analyzed the distribution of transcription initiation site (TSS) distance between these differential DNA methylation sites and the nearest genes. Most of the DNA methylation sites are located between 600 bp upstream and 1200 bp downstream of TSS (Figure 6C), and the hypermethylation sites tend to be located upstream of TSS, which is related to an important regulatory area. Moreover, the genes corresponding to the nearest TSS of the differential DNA methylation were extracted for KEGG pathway analysis. There were ten significant KEGG pathways, including six upregulated and four downregulated items. The upregulated DNA methylation-related genes were mainly enriched in pathways such as basal cell carcinoma, Hippo signaling pathway, mTOR signaling pathway, and signaling pathways regulating pluripotency of stem cells, which are associated with the pathogenesis of GC (Figure 6D). Whereas, the downregulated DNA methylation-related genes participate in viral carcinogenesis, herpes simplex infection, alcoholism, and systemic lupus erythematosus, which were regarded as inducers of tumorigenesis (Figure 6E).

**DISCUSSION**

The identification of prognostic factors for GC is important for its diagnosis and treatment as well as the development of new targeted therapy. Especially, for precise treatment strategy based on the risk stratification, the prognostic factors for GC can not only provide clinicians with the earliest diagnostic markers and definite prognosis prediction, but also can help them select the best treatment plan. The aim of our research was to identify novel biomarkers for GC prognosis. *APC* encodes an important negative regulator that serves as an antagonist of the Wnt signaling pathway, which is closely related to cancer cell proliferation, metastasis, invasion, adhesion, and activation[48]. Here, our results showed that the expression of *APC* was an independent and useful factor for poor prognosis of T4 GC. KM analysis showed that *APC*high was associated with a shorter RFS in T4 GC patients. In addition, *APC*high was found to be associated with advanced tumor stage and high tumor grade in GC and have significant relevance to some known prognostic biomarkers (*KIT*, *PIK3CA*, *KRAS*,and *MLH1*). However, previous research revealed that *APC* is a tumor suppressor gene, and upregulating *APC* expression can inactivate the Wnt/β-catenin pathway in tumorigenesis[49,50]. However, *APC* was found as a poor prognostic marker in patients with T4 GC, although there was no significant association between *APC* and OS or RFS in the overall GC population. Such discrepancy may be due to a combination of the following reasons. On one hand, *APC*, as a tumor suppressor gene, was verified by experimental evidence, not by statistical analysis of a large number of clinical samples. On the other hand, a study showed that Wnt receptor (Fzd7) can be responsible for mediating Wnt signaling in human GC cells both in the presence and absence of APC mutations[51]. Therefore, we hypothesize that APC is not the only crucial molecule involved in Wnt signaling, and other molecules are also involved in this pathway to promote tumorigenesis. Although the high expression of *APC* has antitumor effects, it is not enough to counteract the carcinogenic effects of other molecules involved in Wnt signaling. However, this hypothesis needs to be confirmed by further research. Our finding that *AP*C is an unfavorable prognostic biomarker for T4 GC suggests that fine stratification of GC will be helpful for clinical precision treatment and prognosis evaluation.

In order to further study the mechanism of the association between the high expression of *APC* and the unfavorable prognosis of T4 GC, we comprehensively analyzed that genome-wide genes, miRNAs, and DNA methylation. Tumorigenesis is the result of the interaction of genetic and epigenetic changes. Genetic changes are mainly regulated by gene expression, while epigenetic changes are mainly regulated by non-coding RNAs and DNA modifications[52]. *APC* associated genome-wide gene analysis revealed that high expression of *APC* was related to 12 genes, which were reported to play important roles in the pathogenesis of cancer or be regarded as biomarkers for cancer diagnosis, treatment, and prognosis. Several studies have found that the interaction between proteins produced by some coding genes and APC is involved in the promotion or inhibition of tumor proliferation *via* the Wnt pathway. Zhan *et al*[53] found that targeting MEK strongly synergized with APC in enhancing Wnt responses to the development of GC. Sox2 as a tumor suppressor gene, restrained stomach adenoma formation through modulation of APC/Wnt-responsive and intestinal genes[54]. We speculate that the interaction between the 12 genes identified in this study and *APC* may promote the occurrence of GC through the Wnt pathway. The further study of 12 genes will be helpful to further explore the pathogenesis of GC.

In addition, *APC* related genes are involved in metabolic pathways, which play an important role in tumorigenesis. Recent research has advanced the understanding of the relationship between metabolic regulation and GC. Also, *APC*-dependent changes in the expression of genes influencing metabolism are linked to gastric carcinogenesis. Gerner *et al*[55] found that *APC* alteration results in dysregulation of the pathway for production of polyamines, which are ubiquitous cations essential for cancer cell growth. Metabolism increases the synthesis of nitric oxide synthase 2, which could enhance the risk of *APC*-dependent gastric carcinogenesis in mouse models[56]. In our study, we identified several metabolism related pathways, *e.g.*, cytochrome P450 and arachidonic acid pathways. A member of the cytochrome P450 family has been found to be overexpressed in a variety of tumors and associated with angiogenesis[57]. The metabolic process of linoleic acid derivatives can regulate tumor-related inflammation[58], and there was a positive correlation between alpha linolenic acid metabolite and invasive prostate cancer[59]. The arachidonic acid pathway is a metabolic process that plays a key role in carcinogenesis, and arachidonic acid pathway enzyme are considered as new targets for the prevention and treatment of cancer[60]. Metabolic reprogramming can contribute to tumor development, and drugs designed according to the metabolic target have become a clinically effective treatment for cancer[61]. Our study suggests that high expression of *APC* may play an important role in tumor metabolism and may be a target for the design of chemotherapy drugs.

In the genome-wide miRNA analysis, we identified several tumorigenesis associated miRNAs that were associated with *APC* expression. Interestingly, most of these miRNAs were negatively correlated with *APC* in theassociational analysis and mRNA-miRNA regulatory network. Previous studies have showed that most of miRNAs identified in our study were downregulated in GC. This finding, together with our result that high expression of the *APC* gene was associated with a poor prognosis in T4 GC patients, suggested that there was a ceRNA mechanism[62] between the *APC* gene and miRNAs. The upregulated expression of upstream lncRNA can sponge miRNAs and thereby upregulate *APC*, thus promoting the proliferation of cancer cells. There is increasing evidence that the imbalance of ceRNA interaction, including miRNAs, cirRNAs, and lncRNAs, promotes the development of GC[63]. The ceRNA mechanism of *APC* and miR-135b-5p as well as *APC* and miR-106a-3p has been reported to promote the carcinogenesis of GC. Our research further proved that *APC*high, together with miRNAs, contributed to the carcinogenesis of GC and that *APC*high was a robust unfavorable prognostic biomarker for T4 GC patients.

Finally, therelationship between *APC*high and genome-wide DNA methylation was explored. We found not only differential methylation between *APC*high and*APC*low, but also different distribution of differential DNA methylation. Our results showed that there were more hypermethylation sites than hypomethylation sites that were associated with *APC* expression. Possibly, hypermethylation of tumor suppressor genes[46] and high expression of *APC* act together to promote the occurrence and deterioration of GC, thereby bring an unfavorable prognosis for T4 GC patients. In addition, the distribution of DNA methylation showed that hypermethylation sites were more frequently located on CpG islands and upstream of TSS. DNA methylation of the CpG islands has been associated with tumor development, cell cycle, DNA repair, angiogenesis, and apoptosis[47], while tumorigenesis was associated with DNA methylated in the region around the TSS[64]. Together, our research suggested that high expression of *APC* was regulated by DNA methylation.

In conclusion, our research demonstrated that *APC*high may be a useful adverse prognostic biomarker for T4 GC patients. The finding that *APC*high is associated with distinctive genome-wide gene/miRNA/methylation expression and related cellular functional pathways could well explain why *APC* acts as an adverse prognostic biomarker for T4 GC patients as well as the mechanism underlying its molecular biological function in GC. As a novel potential biological target, *APC* may be used to study the pathogenesis of GC and guide the clinical diagnosis, treatment, and prognosis evaluation of T4 GC patients.

**Article Highlights**

***Research background***

Adenoma polyposis coli (*APC*) mutation is associated with tumorigenesis *via* the Wnt signaling pathway.

***Research motivation***

*APC* acts as a tumor suppressor gene in Wnt signal pathway, and mutation and inactivation of *APC* are unique key and early events in tumorigenesis, especially colorectal cancer. Compared with colorectal cancer, there have been much fewer studies on *APC* in other cancers. However, some successful studies have initially shown that *APC* could be used as a biomarker for GC. This inspired us to investigate the clinical features and mechanism of *APC* expression in GC.

***Research objectives***

The research objective was to validate that *APC* is associated with the pathogenesis and clinical features of GC. We demonstrated that high expression of *APC* is a biomarker for poor prognosis in T4 GC patient. Our research suggested that *APC* may be used as a novel biomarker for pathogenesis research, diagnosis, and treatment of GC.

***Research methods***

Based on RNA-Seq, miRNA-Seq, Illumina Infinium Human Methylation 450, and clinical follow-up data from TCGA, we systematically analyzed the *APC*-related mRNA and miRNA expression and DNA methylation profile on a genome-wide scale by multi-dimensional methods. The novel application of resource-sharing databases provided us with the opportunity to investigate more reliable biomarkers for GC.

***Research results***

We found that high expression of *APC* was a biomarker for poor prognosis in T4 GC patient. Our research brings new perspective to the *APC* gene in tumors, namely, *APC* is an oncogene in T4 GC patients and the high expression of *APC* indicates a poor prognosis in T4 GC patients. However, why high expression of *APC* predicts a poor prognosis only in T4 GC patients remains to be further studied.

***Research conclusions***

Our research demonstrated that *APC*high may be a useful adverse prognostic biomarker for T4 GC patients. The finding that *APC*high is associated with distinctive genome-wide gene/miRNA/methylation expression and related cellular functional pathways could well explain why *APC* acts as an adverse prognostic biomarker for T4 GC patients as well as the mechanism underlying its molecular biological function in GC.

***Research perspectives***

As a novel potential biological target, *APC* may be used to study the pathogenesis of GC and guide the clinical diagnosis, treatment, and prognosis evaluation of T4 GC patients.

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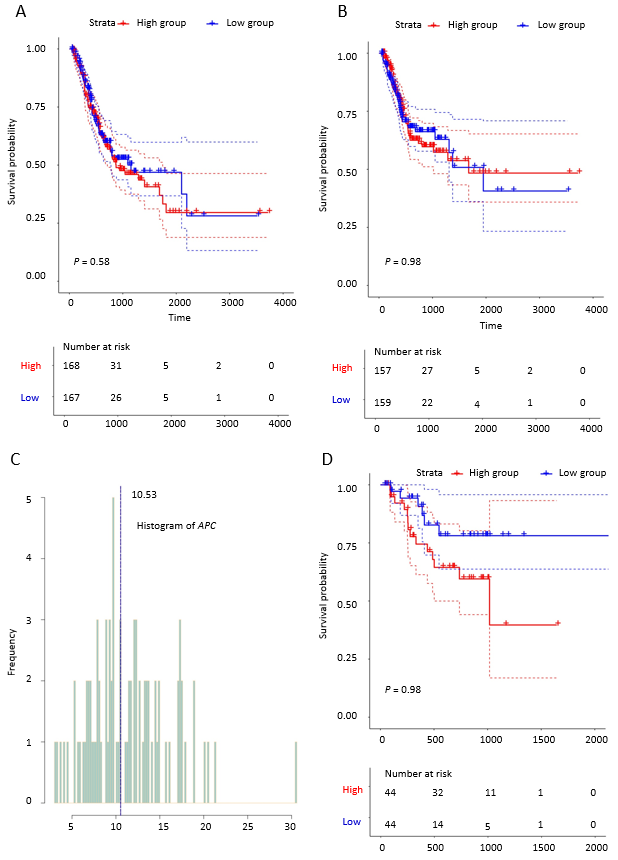
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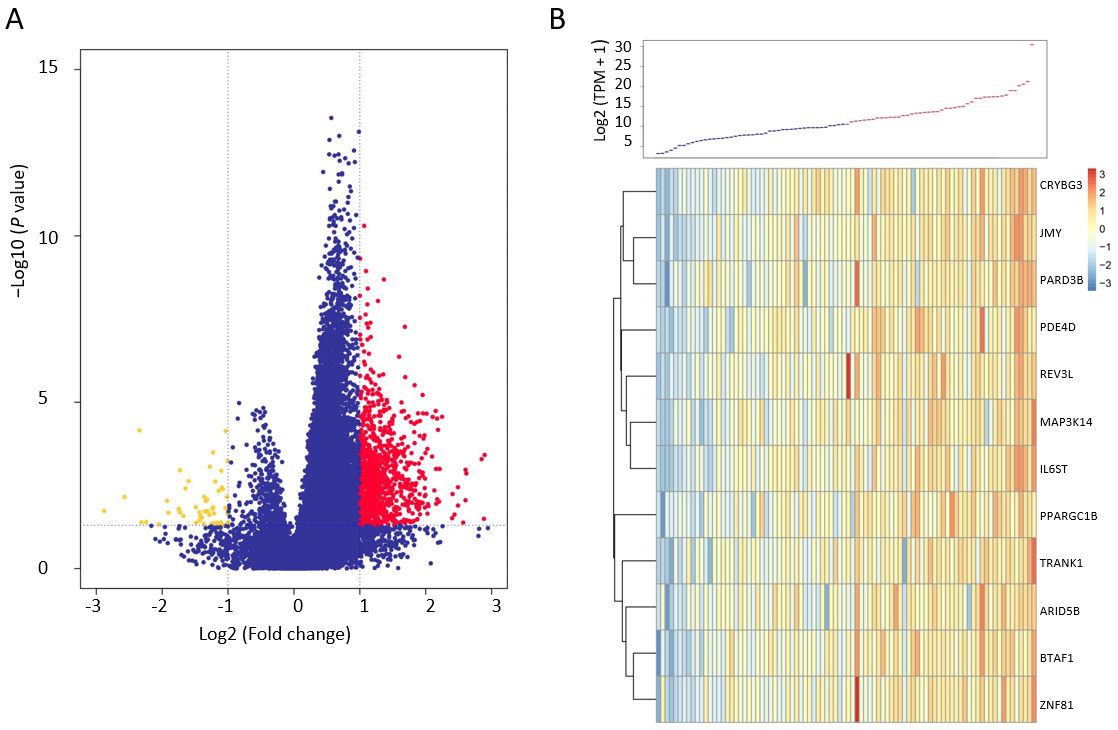
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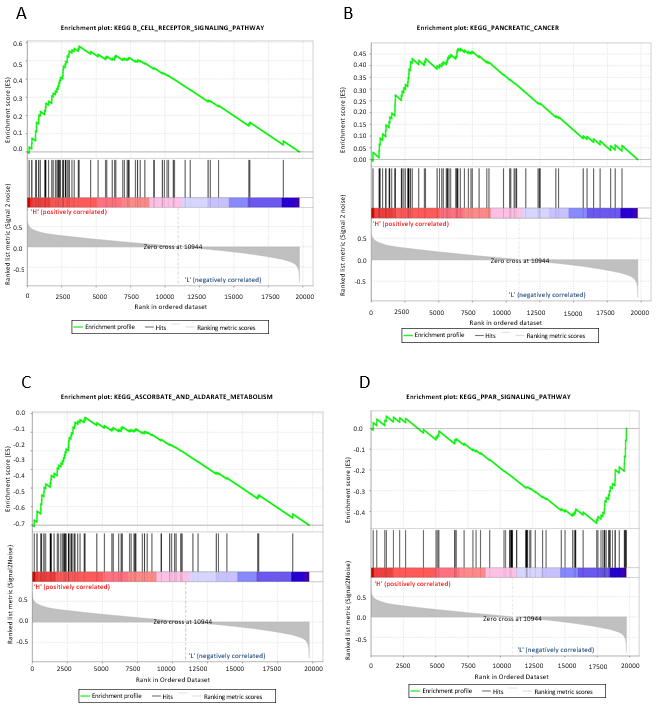
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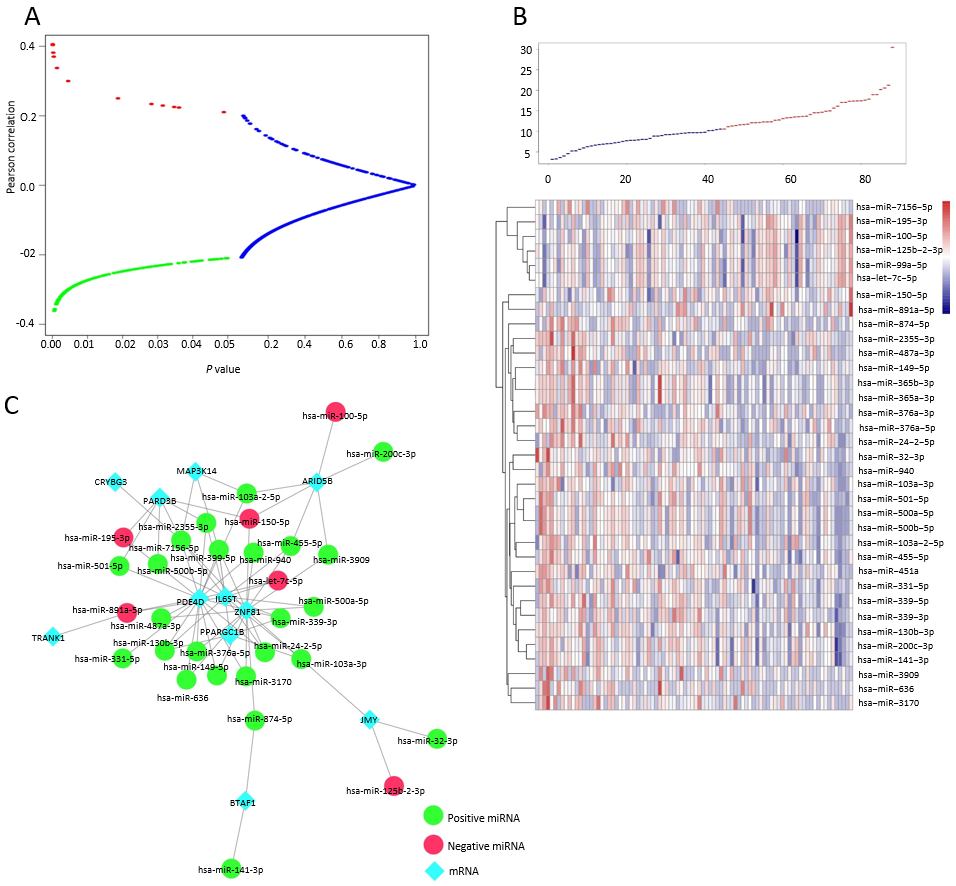
**Figure 1 Prognostic value of *APC* expression in gastric cancer patients.** A: Overall survival of gastric cancer patients with high *APC* expression and those with low expression (based on median value); B: Relapse-free survival of gastric cancer patients with high *APC* expression and those with low expression; C: Distribution of *APC* expression; D: Relapse-free survival of T4 gastric cancer patients with high *APC* expression and those with low expression. *APC*: Adenoma polyposis coli; GC: Gastric cancer.



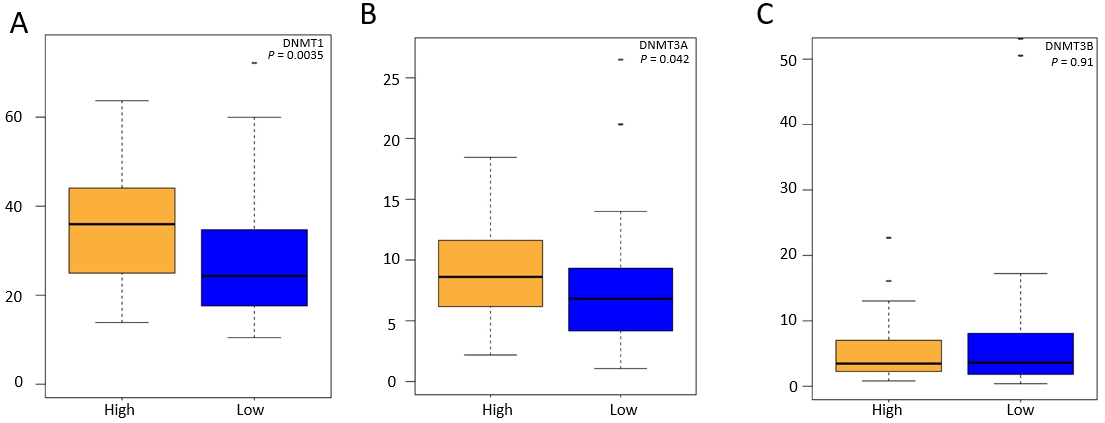
**Figure 2 Differentially expressed mRNAs associated with *APC*high.** A: Volcanic map of differentially expressed mRNAs associated with *APC*high; (B) Heatmap of differentially expressed mRNAs associated with *APC*high. *APC*: Adenoma polyposis coli.



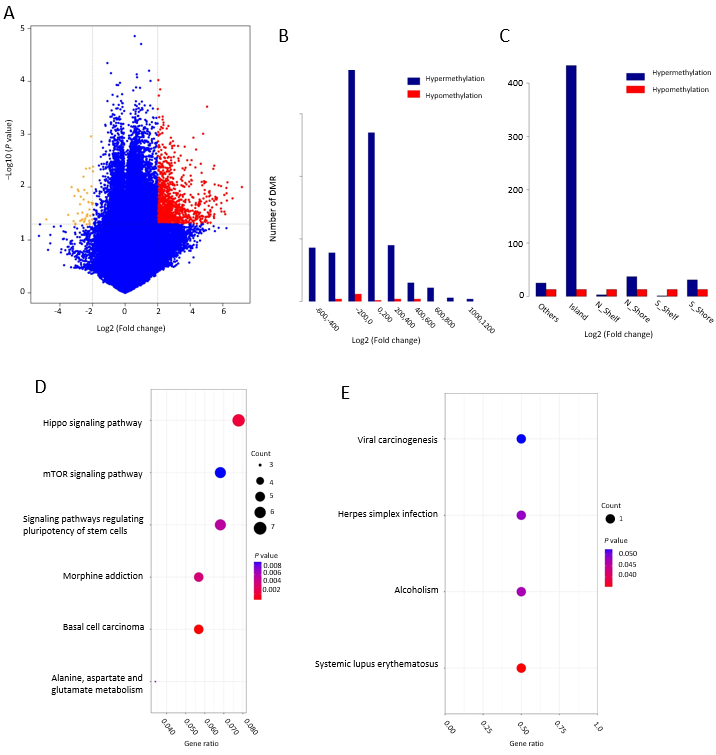
**Figure 3 KEGG pathways of differentially expressed mRNAs associated with *APC*high.** Upregulated pathways of (A) cell receptor signaling and (B) pancreatic cancer and downregulated pathways of (C) ascorbate and aldarate metabolism and (D) PPAR signaling pathway are shown. *APC*: Adenoma polyposis coli.



**Figure 4 Genome-wide analysis of miRNAs associated with *APC*high.** A: Relationship between the correlation coefficient and correlation *P*-value; B: MiRNAs that were significantly positively correlated (red), negatively correlated (green), and had no correlation (blue) with *APC*high, as well as the heatmap of miRNAs associated with *APC*high; C: mRNA-miRNA regulatory network. *APC*: Adenoma polyposis coli.



**Figure 5 Differential expression of DNMT1, DNMT3A, and DNMT3B between *APC*high and *APC*low.** *APC*: Adenoma polyposis coli; DNMT: DNA methyltransferase.



**Figure 6 Analysis of relationship between *APC*high and genome-wide DNA methylation.** A: Volcanic map of differential DNA methylation associated with *APC*high; B: Distribution of DNA methylation associated with *APC*high based on CPG islands; C: Distribution of DNA methylation associated with *APC*high based on transcription initiation site; D: Upregulated KEGG pathways of genes corresponding to the nearest transcription initiation site of differential DNA methylation; E: Downregulated KEGG pathways of genes corresponding to the nearest transcription initiation site of differential DNA methylation. *APC*: Adenoma polyposis coli.

**Table 1 Relationship between *APC*high and clinical characteristic and known prognostic genes**

|  |  |  |  |
| --- | --- | --- | --- |
| **Characteristic** | ***APC*high** | ***APC*low** | ***P-*value** |
| Age | 66 ± 8 | 61 ± 9 | 0.084 |
| Stage |  |  |  |
| I | 5 | 25 | 0.031 |
| II | 10 | 12 | 1 |
| III + IV | 29 | 7 | 0.043 |
| Grade |  |  |  |
| 1 | 4 | 33 | 0.031 |
| 2 | 15 | 5 | 0.047 |
| 3 | 25 | 6 | 0.039 |
| Lymph node metastasis |  |  |  |
| N0 | 7 | 23 | 0.027 |
| N1 | 10 | 12 | 1 |
| N2 | 11 | 4 | 0.031 |
| N3 | 16 | 5 | 0.035 |
| Distant metastasis |  |  |  |
| M0 | 7 | 9 | 0.781 |
| M1 | 37 | 35 | 0.812 |
| Gene |  |  |  |
| *KIT* | 29 | 15 | 0.031 |
| *PIK3CA* | 32 | 12 | <0.001 |
| *KRAS* | 28 | 16 | 0.038 |
| *MLH1* | 28 | 16 | 0.038 |

APC: Adenoma polyposis coli.

**Table 2 Association between differential mRNAs and *APC*high**

|  |  |  |  |  |
| --- | --- | --- | --- | --- |
| **mRNA** | ***P*-value (*t*-test)** | ***P*-value (Wilcoxon test)** | **Fold change** | **Regulation** |
| *BTAF1* | 1.45E-08 | 4.81E-10 | 2.00561 | Up-regulation |
| *ARID5B* | 1.20E-06 | 1.30E-07 | 2.011267 | Up-regulation |
| *ZNF81* | 2.04E-06 | 3.84E-09 | 2.175277 | Up-regulation |
| *JMY* | 1.60E-09 | 4.99E-11 | 2.094035 | Up-regulation |
| *MAP3K14* | 7.83E-07 | 3.00E-07 | 2.092268 | Up-regulation |
| *PDE4D* | 5.58E-07 | 4.33E-08 | 2.151154 | Up-regulation |
| *CRYBG3* | 4.92E-07 | 1.10E-07 | 2.242404 | Up-regulation |
| *PARD3B* | 2.59E-06 | 9.38E-08 | 2.007173 | Up-regulation |
| *PPARGC1B* | 1.76E-08 | 2.04E-09 | 2.576507 | Up-regulation |
| *TRANK1* | 6.01E-07 | 1.15E-08 | 2.165217 | Up-regulation |
| *REV3L* | 6.32E-07 | 1.14E-09 | 2.13368 | Up-regulation |
| *IL6ST* | 3.07E-07 | 1.30E-07 | 2.168223 | Up-regulation |

APC: Adenoma polyposis coli.

**Table 3 *APC*high associated KEGG pathways**

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
| **MSigDB** | **Size** | **ES** | **NES** | ***P-*value** | **Group** |
| KEGG\_B\_CELL\_RECEPTOR\_SIGNALING\_PATHWAY | 74 | 0.579 | 1.712 | 0.012 | High |
| KEGG\_PANCREATIC\_CANCER | 70 | 0.472 | 1.597 | 0.025 | High |
| KEGG\_PROSTATE\_CANCER | 89 | 0.450 | 1.570 | 0.025 | High |
| KEGG\_REGULATION\_OF\_ACTIN\_CYTOSKELETON | 211 | 0.396 | 1.541 | 0.032 | High |
| KEGG\_NEUROTROPHIN\_SIGNALING\_PATHWAY | 126 | 0.437 | 1.533 | 0.046 | High |
| KEGG\_PROGESTERONE\_MEDIATED\_OOCYTE\_MATURATION | 85 | 0.412 | 1.491 | 0.040 | High |
| KEGG\_ASCORBATE\_AND\_ALDARATE\_METABOLISM | 25 | -0.674 | -1.851 | 0.002 | Low |
| KEGG\_PPAR\_SIGNALING\_PATHWAY | 69 | -0.456 | -1.621 | 0.003 | Low |
| KEGG\_DRUG\_METABOLISM\_OTHER\_ENZYMES | 51 | -0.590 | -1.789 | 0.003 | Low |
| KEGG\_STEROID\_HORMONE\_BIOSYNTHESIS | 55 | -0.595 | -1.853 | 0.004 | Low |
| KEGG\_PORPHYRIN\_AND\_CHLOROPHYLL\_METABOLISM | 41 | -0.617 | -1.833 | 0.004 | Low |
| KEGG\_DRUG\_METABOLISM\_CYTOCHROME\_P450 | 70 | -0.569 | -1.776 | 0.012 | Low |
| KEGG\_RETINOL\_METABOLISM | 63 | -0.556 | -1.687 | 0.012 | Low |
| KEGG\_METABOLISM\_OF\_XENOBIOTICS\_BY\_CYTOCHROME\_P450 | 68 | -0.572 | -1.782 | 0.014 | Low |
| KEGG\_PENTOSE\_AND\_GLUCURONATE\_INTERCONVERSIONS | 27 | -0.638 | -1.734 | 0.016 | Low |
| KEGG\_LINOLEIC\_ACID\_METABOLISM | 29 | -0.589 | -1.691 | 0.019 | Low |
| KEGG\_ALPHA\_LINOLENIC\_ACID\_METABOLISM | 19 | -0.565 | -1.662 | 0.020 | Low |
| KEGG\_ARACHIDONIC\_ACID\_METABOLISM | 58 | -0.424 | -1.541 | 0.031 | Low |

APC: Adenoma polyposis coli.