

World Journal of *Gastrointestinal Oncology*

World J Gastrointest Oncol 2020 April 15; 12(4): 365-513



REVIEW

- 365 Efficacy of mesenchymal stem cells in the treatment of gastrointestinal malignancies
Li JN, Li W, Cao LQ, Liu N, Zhang K

ORIGINAL ARTICLE**Basic Study**

- 383 Potential microRNA panel for the diagnosis and prediction of overall survival of hepatocellular carcinoma with hepatitis B virus infection
Zhang Q, Xu HF, Song WY, Zhang PJ, Song YB
- 394 LINC00511 promotes gastric cancer cell growth by acting as a ceRNA
Sun CB, Wang HY, Han XQ, Liu YN, Wang MC, Zhang HX, Gu YF, Leng XG

Retrospective Cohort Study

- 405 Primary tumor location and survival in colorectal cancer: A retrospective cohort study
Aggarwal H, Sheffield KM, Li L, Lenis D, Sorg R, Barzi A, Miksad R
- 424 Robotic- vs laparoscopic-assisted proctectomy for locally advanced rectal cancer based on propensity score matching: Short-term outcomes at a colorectal center in China
Ye SP, Zhu WQ, Liu DN, Lei X, Jiang QG, Hu HM, Tang B, He PH, Gao GM, Tang HC, Shi J, Li TY

Retrospective Study

- 435 Diagnostic ability of multi-detector spiral computed tomography for pathological lymph node metastasis of advanced gastric cancer
Jiang ZY, Kinami S, Nakamura N, Miyata T, Fujita H, Takamura H, Ueda N, Kosaka T
- 447 Nomogram using F-18 fluorodeoxyglucose positron emission tomography/computed tomography for preoperative prediction of lymph node metastasis in gastric cancer
Song BI
- 457 Perineural invasion of hilar cholangiocarcinoma in Chinese population: One center's experience
Li CG, Zhou ZP, Tan XL, Zhao ZM
- 467 Prognostic significance of systemic immune-inflammation index in patients with intrahepatic cholangiocarcinoma undergoing hepatic resection
Li H, Wang JJ, Zhang M, Ren B, Li JX, Xu L, Wu H

Observational Study

- 483** Evaluation of the value of multiparameter combined analysis of serum markers in the early diagnosis of gastric cancer
Zhang ZG, Xu L, Zhang PJ, Han L

Prospective Study

- 492** Expression and significance of miR-654-5p and miR-376b-3p in patients with colon cancer
Li P, Cai JX, Han F, Wang J, Zhou JJ, Shen KW, Wang LH

EVIDENCE-BASED MEDICINE

- 503** Adjuvant chemotherapy in curatively resected rectal cancer: How valid are the data?
Manzini G, Hapke F, Hines IN, Henne-Bruns D, Kremer M

ABOUT COVER

Editorial Board Member of *World Journal of Gastrointestinal Oncology*, Ki-Tae Ha, MD, PhD, Professor, Department of Korean Medical Science, School of Korean Medicine, Pusan National University, Yangsan 50612, Gyeongnam, South Korea

AIMS AND SCOPE

The primary aim of *World Journal of Gastrointestinal Oncology (WJGO, World J Gastrointest Oncol)* is to provide scholars and readers from various fields of gastrointestinal oncology with a platform to publish high-quality basic and clinical research articles and communicate their research findings online.

WJGO mainly publishes articles reporting research results and findings obtained in the field of gastrointestinal oncology and covering a wide range of topics including islet cell adenoma, liver cell adenoma, adenomatous polyposis coli, appendiceal neoplasms, bile duct neoplasms, biliary tract neoplasms, hepatocellular carcinoma, islet cell carcinoma, pancreatic ductal carcinoma, cecal neoplasms, colonic neoplasms, colorectal neoplasms, hereditary nonpolyposis colorectal neoplasms, common bile duct neoplasms, duodenal neoplasms, esophageal neoplasms, gallbladder neoplasms, etc.

INDEXING/ABSTRACTING

The *WJGO* is now indexed in Science Citation Index Expanded (also known as SciSearch®), PubMed, and PubMed Central. The 2019 edition of Journal Citation Reports® cites the 2018 impact factor for *WJGO* as 2.758 (5-year impact factor: 3.220), ranking *WJGO* as 52 among 84 journals in gastroenterology and hepatology (quartile in category Q3), and 131 among 229 journals in oncology (quartile in category Q3).

RESPONSIBLE EDITORS FOR THIS ISSUE

Responsible Electronic Editor: *Li-Li Qi*
 Proofing Production Department Director: *Xiang Li*
 Responsible Editorial Office Director: *Jin-Lai Wang*

NAME OF JOURNAL

World Journal of Gastrointestinal Oncology

ISSN

ISSN 1948-5204 (online)

LAUNCH DATE

February 15, 2009

FREQUENCY

Monthly

EDITORS-IN-CHIEF

Monjur Ahmed, Rosa M Jimenez Rodriguez, Pashtoon Kasi

EDITORIAL BOARD MEMBERS

<https://www.wjgnet.com/1948-5204/editorialboard.htm>

PUBLICATION DATE

April 15, 2020

COPYRIGHT

© 2020 Baishideng Publishing Group Inc

INSTRUCTIONS TO AUTHORS

<https://www.wjgnet.com/bpg/gerinfo/204>

GUIDELINES FOR ETHICS DOCUMENTS

<https://www.wjgnet.com/bpg/GerInfo/287>

GUIDELINES FOR NON-NATIVE SPEAKERS OF ENGLISH

<https://www.wjgnet.com/bpg/gerinfo/240>

PUBLICATION ETHICS

<https://www.wjgnet.com/bpg/GerInfo/288>

PUBLICATION MISCONDUCT

<https://www.wjgnet.com/bpg/gerinfo/208>

ARTICLE PROCESSING CHARGE

<https://www.wjgnet.com/bpg/gerinfo/242>

STEPS FOR SUBMITTING MANUSCRIPTS

<https://www.wjgnet.com/bpg/GerInfo/239>

ONLINE SUBMISSION

<https://www.f6publishing.com>

Basic Study

Potential microRNA panel for the diagnosis and prediction of overall survival of hepatocellular carcinoma with hepatitis B virus infection

Qi Zhang, Hai-Feng Xu, Wen-Yue Song, Peng-Jun Zhang, Yong-Bo Song

ORCID number: Qi Zhang (0000-0001-5710-2401); Hai-Feng Xu (0000-0003-0023-3894); Wen-Yue Song (0000-0003-4483-6507); Peng-Jun Zhang (0000-0002-7391-2495); Yong-Bo Song (0000-0002-9548-433X).

Author contributions: Zhang Q, Xu HF, Song YB and Zhang PJ designed the study; Zhang Q performed the research; Xu HF and Song WY analyzed the data; Zhang Q and Xu HF wrote the paper; Song YB and Zhang PJ revised the manuscript for final submission; Zhang Q and Xu HF contributed equally to this study; Song YB and Zhang PJ are the co-corresponding authors.

Supported by the National Key R&D Program of China, No. 2016YFC0106604.

Institutional review board

statement: The study was reviewed and approved by the Peking University Cancer Hospital & Institute review board. All study participants or their legal guardian provided written informed consent prior to study enrollment.

Conflict-of-interest statement: We declare that we have no financial or personal relationships with other individuals or organizations that can inappropriately influence our work and that there is no professional or other personal interest of any nature in any product, service and/or company that could be construed as influencing the position presented in or the review of the manuscript.

Qi Zhang, Wen-Yue Song, Yong-Bo Song, School of Life Science and Biopharmaceutics, Shenyang Pharmaceutical University, Shenyang 110016, Liaoning Province, China

Hai-Feng Xu, Peng-Jun Zhang, Key Laboratory of Carcinogenesis and Translational Research (Ministry of Education/Beijing), Interventional Therapy Department, Peking University Cancer Hospital and Institute, Beijing 100142, China

Corresponding author: Yong-Bo Song, PharmD, Associate Professor, School of Life Science and Biopharmaceutics, Shenyang Pharmaceutical University, No. 103 Wenhua Road, Shenhe District, Shenyang 110016, Liaoning Province, China. songyongbo@syphu.edu.cn

Abstract**BACKGROUND**

In hepatocellular carcinoma (HCC), abnormal expression of multiple microRNAs (miRNAs) has been shown to be involved in the malignant biological behavior of liver cancer. The vast majority of liver cancer cases in China are closely related to hepatitis B virus (HBV) infection, but there are few studies on the changes of miRNA expression in the progression from HBV infection to hepatoma.

AIM

To explore the role of miRNAs in the progression of HBV infection to cirrhosis and even to liver cancer.

METHODS

We screened differentially expressed miRNAs in 40 HBV cirrhosis, 40 normal and 15 HCC tissues by using a TaqMan Low Density Array and real time quantitative polymerase chain reaction. To evaluate the power of the selected miRNAs to predict disease, we calculated the area under the receiver-operating-characteristic curves. The overall survival of HBV cirrhosis patients was analyzed via Kaplan-Meier analysis.

RESULTS

The levels of miR-375, miR-122 and miR-143 were significantly lower in HBV cirrhosis tissues, while miR-224 was significantly higher than in the controls ($P < 0.0001$). The area under the curves of the receiver-operating-characteristic curve for the 4-miRNA panel was 0.991 (95%CI: 0.974-1). Patients with a lower expression level of miR-224 or higher expression levels of miR-375, miR-122 and miR-143 had longer overall survival.

CONCLUSION

Data sharing statement: No additional data are available.

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

Manuscript source: Unsolicited manuscript

Received: December 22, 2019

Peer-review started: December 22, 2019

First decision: January 19, 2020

Revised: February 6, 2020

Accepted: March 23, 2020

Article in press: March 23, 2020

Published online: April 15, 2020

P-Reviewer: Abd el Moety HA, Campanale M, Sogabe I

S-Editor: Wang JL

L-Editor: A

E-Editor: Qi LL



The four miRNAs (miR-375, miR-122, miR-143 and miR-224) may be helpful for early diagnosis of HBV infection, HBV cirrhosis, and prediction of its overall survival.

Key words: Hepatitis B virus infection; Hepatocellular carcinoma; MicroRNAs; Cirrhosis; Biomarker; Tissue

©The Author(s) 2020. Published by Baishideng Publishing Group Inc. All rights reserved.

Core tip: Abnormal expression of microRNAs (miRNAs) may lead to an abnormal physiological state and disease, such as, kinds of cancers. We detect the levels of miR-375, miR-122, miR-143 and miR-224. By combination of the miRNA panels, the area under the curves of the receiver-operating-characteristic curve was 0.991. In addition, the four miRNAs (miR-375, miR-122, miR-143 and miR-224) may be helpful for early detection and prognosis of hepatocellular carcinoma.

Citation: Zhang Q, Xu HF, Song WY, Zhang PJ, Song YB. Potential microRNA panel for the diagnosis and prediction of overall survival of hepatocellular carcinoma with hepatitis B virus infection. *World J Gastrointest Oncol* 2020; 12(4): 383-393

URL: <https://www.wjgnet.com/1948-5204/full/v12/i4/383.htm>

DOI: <https://dx.doi.org/10.4251/wjgo.v12.i4.383>

INTRODUCTION

Hepatitis B virus (HBV) is known as the smallest double-stranded DNA virus that infects humans, and HBV infection has become a global problem. The World Health Organization reported that an estimated 257 million people are living with HBV infection, defined as hepatitis B surface (HBsAg) antigen positivity, and it resulted in 887000 deaths in 2015, mostly from complications [including cirrhosis and hepatocellular carcinoma (HCC)]^[1]. Despite the availability of effective vaccines, the virus causes approximately 780000 deaths every year. It is estimated that approximately 15% to 30% of HBV carriers have a risk of developing cirrhosis^[2]. For diagnosis of hepatitis B, the incubation period is long, and the disease cannot be diagnosed in the incubation period, which is from the invasion of the hepatitis virus to the initial clinical symptoms and can last 75 d on average but can vary from 30 to 180 d^[1]. HBsAg, which is the main marker of HBV infection, can be detected in serum 2 to 6 wk before alanine aminotransferase elevation. DNA detection of HBV is sensitive to low-level HBV virus in vivo by amplifying viral nucleic acid. This assay is commonly used to assess viral replication, indicating HBV replication and contagiousness.

MicroRNAs (miRNAs) are a class of noncoding single-stranded RNA molecules of approximately 22 nt in size. They regulate the expression of the mRNA and protein of other genes by binding to target mRNAs to participate in various physiological processes, such as growth and development, inflammation, tumors and physiological and pathological processes^[3-5]. Abnormal expression of miRNAs may lead to an abnormal physiological state and disease. In many cancers, the expression levels of miRNAs will change significantly, which may affect proto-oncogenes and tumor suppressor genes^[9-12]. In HCC, abnormal expression of multiple miRNAs has been shown to be involved in the malignant biological behavior of liver cancer^[13-17]. The vast majority of liver cancer cases in China are closely related to HBV infection, but there are few studies on the changes of miRNA expression in the progression from HBV infection to hepatoma.

In this study, TaqMan Low Density Array (TLDA) and real time quantitative polymerase chain reaction (RT-qPCR) were used to characterize the profile of miRNAs in chronic hepatitis B, HCC and normal control tissues to explore the role of miRNAs in the development of chronic hepatitis B to liver cancer.

MATERIALS AND METHODS

Samples

Liver tissues from patients undergoing liver cancer resection or liver biopsy from July 2011 to January 2013 were collected, including 40 HBV cirrhosis, 40 normal, and 15 HCC tissues. HBsAg and/or HBV-DNA positivity for more than 6 months was considered to be chronic HBV infection, while the normal liver tissue was from the determination of no liver disease during the past and during hospitalization. All samples had no other basic liver disease and were confirmed by pathology. After the tissue was placed in liquid nitrogen for the first time, it was stored at -80 °C for later use.

The patients were followed up for 5 years, and their overall survival was recorded. For patients who survived 5 years later, their overall survival time was considered to be 5 years. All subjects had signed informed consent, and the study was approved by the Ethics Committee of Beijing Cancer Hospital.

RNA isolation

Total RNA from 5 µg of normal liver and HBV cirrhosis samples was extracted according to the steps of the TRIzol reagent manual. The absorbance values of A260 and A280 of total RNA were determined by a UV spectrophotometer to calculate the concentration of RNA. The A260 value was used to detect the purity of RNA, and the value of A260/A280 was calculated to further test the quality of total RNA.

TLDA

The TLDA (TaqMan Array Human MicroRNA A+B Cards Set v3.0, Life Technologies) was used to profile the 754 different human miRNAs described in previous literature^[18]. To increase the sensitivity of the TLDA, we performed a pre-amplification using the QuantStudio 7 Flex RT-PCR System (Applied Biosystems)^[19]. The threshold cycle (Cq) values showed the concentrations of miRNAs, which were normalized to an internal control. The fold changes of miRNA expression were calculated by the equation $2^{-\Delta\Delta Cq}$.

Individual RT-qPCR assays

According to the manufacturer's instructions (QuantStudio 7 Flex RT PCR System; Applied Biosystems) with slight modifications, hydrolysis probe-based qRT-PCR was performed. Reverse transcription was carried out as previously described^[18]. All experiments were carried out in triplicate. An endogenous control, the combination of let-7d, let-7g and let-7i (let-7d/g/i) in this experiment, is important for normalizing qRT-PCR data^[18,20]. Relative levels of miRNAs were normalized to let-7d/g/i and were calculated using the $2^{-\Delta\Delta Cq}$ method^[18,21].

Statistical analysis

Statistical analyses were performed with the Statistical Analysis System software SPSS 16.0, and data are presented as the mean ± SE for miRNAs or mean ± SD for other variables. With Student's *t*-test and two-sided χ^2 test, we compared the differences between the two groups, and the *P* value must be < 0.05, which will be considered statistically significant. The receiver operating characteristic (ROC) curves and the area under the ROC curves (AUC) were calculated to evaluate the predictive power of the selected miRNAs. Furthermore, risk score analysis was performed to evaluate the associations between miRNAs and HBV cirrhosis as previously described^[20,22]. To indicate miRNAs' contribution to the risk score function, the regression coefficient was used for the risk score as the weight^[23,24]. Samples were divided into a high-risk group, predicting HBV cases, and a low-risk group, predicting control individuals, according to their risk score function, and then, we found the appropriate cutoff point. Analysis of patient survival was performed by Kaplan-Meier analysis.

RESULTS

Expression profile of miRNAs in liver tissue by TLDA

A multiphase case control study was designed to investigate the differences in miRNA expression profiles between normal liver and HBV cirrhosis (Figure 1, Table 1). Using TLDA, we analyzed miRNA expression in three random pairs of samples of normal liver and HBV cirrhosis. The Cq values of the miRNAs were all < 25, and the concentrations of the miRNAs all showed > 2-fold differences between normal liver and HBV cirrhosis, which was defined as differential expression. Nine miRNAs, including miR-125b, miR-602, miR-210, miR-224, miR-129, miR-99a, miR-141, miR-342

and miR-145, were upregulated, while 14 miRNAs, including miR-122, miR-143, miR-199a, miR-375, miR-27a, miR-34b, miR-130a, miR-625, miR-142, miR-193a, miR-140, miR-100, miR-342 and miR-29c, were downregulated among the 754 miRNAs in HBV cirrhosis patients compared to normal liver tissue.

Expression of miRNAs by RT-qPCR analysis

To verify the accuracy of differentially expressed miRNAs in the above TLDA results, we performed RT-qPCR analysis at the individual sample level.

In the training set, miRNAs were measured in a separate set of individual tissue samples from 25 HBV cirrhosis patients and 25 normal liver controls, and only miRNAs with a mean change of 2-fold and a *P* value of 0.001 were selected for further analysis. We used these criteria to generate a list of 4 miRNAs (miR-224, miR-375, miR-122 and miR-143), which showed significantly different miRNA patterns between HBV cirrhosis patients and normal controls.

Furthermore, RT-qPCR was performed to verify the expression of the 4 miRNAs chosen above with another 15 HBV cirrhosis patients and 15 normal liver controls. The results showed that miR-224 was increased, while miR-375, miR-122 and miR-143 were significantly decreased in the tissues of HBV cirrhosis patients compared with the controls (at least *P* < 0.005), which was the same as the former cohort (Table 2, Figure 2A-D).

Diagnostic ability of the selected miRNAs

Subsequently, to evaluate the ability of the selected four tissue miRNAs to distinguish HBV cirrhosis from normal controls, we performed ROC curve analysis for each miRNA. For 40 cases of HBV cirrhosis and 40 normal control tissue samples, the AUC values of miR-224, miR-375, miR-122 and miR-143 were 0.938, 0.932, 0.923 and 0.915, respectively (Figure 3A-D). To further assess the diagnostic value of miRNAs in distinguishing between HBV cirrhosis and normal controls, we performed a risk score analysis of the dataset and used this risk scoring method to predict HBV cirrhosis and normal controls. The results showed that the best cutoff value (in this cutoff value, sensitivity + specificity is the largest) was 2.016, and 6 normal controls showed a risk score > 2.016, while 37 of the 40 HBV cirrhosis patients exhibited a risk score > 2.016 (Table 3). Furthermore, we integrated the 4-miRNA signature into a single biomarker using the risk score functions and evaluated the diagnostic accuracy of the miRNA signatures as HBV cirrhosis fingerprints. As expected, we obtained an AUC value of 0.991 (95% CI: 0.974-1) by combining miR-224, miR-375, miR-122 and miR-143 to differentiate HBV cirrhosis patients from healthy controls (Figure 3E).

Relationship between miRNA and overall survival of HBV cirrhosis patients

After statistical analysis of the follow-up of these 40 HBV cirrhosis patients, the overall survival of patients with different expression levels of the 4 miRNAs was determined and is shown in Table 4. Log rank analysis showed that patients with a lower expression level of miR-224 and higher expression levels of miR-375, miR-122 and miR-143 had longer overall survival (Figure 4) than those with the opposite expression pattern (*P* < 0.01). According to Cox analysis, miR-224, miR-375, miR-122 and miR-143 are important factors affecting overall survival.

DISCUSSION

In this study, we used TLDA and RT-qPCR validation to systematically detect miRNA expression in HBV cirrhosis and found a new miR-panel (miR-224, miR-375, miR-122 and miR-143) that can effectively distinguish HBV cirrhosis patients from controls. Furthermore, we examined the expression of the 4 miRNAs in the tumor tissues of patients with HCC and found that the results were consistent with those in HBV cirrhosis. Compared with normal controls, in the tissue/serum of patients with HBV cirrhosis, miR-375, miR-92a, miR-10a, miR-223, miR-423, miR-23b/a, miR-342-3p, miR-150, let-7c, miR-99a, miR-125b, miR-22, miR-720, miR-1275, miR-486-3p, miR-1908, miR-675, and miR-1231 were significantly upregulated^[25-29]. However, this 4-miRNA combination in our study has not been reported, and the combination has a high ROC curve AUC of 0.991, suggesting a strong ability to distinguish HBV cirrhosis from normal controls.

Simultaneously, we analyzed the relationship of each miRNA with overall survival and found that patients with lower abnormal miRNA expression will have a longer overall survival. MiR-224 can offset the effects on the reduction of tumor growth and cell proliferation of glycine N-methyltransferase (GNMT) by targeting GNMT, which is a tumor suppressor for HCC^[30]. The receptor tyrosine-protein kinase erbB-2, a direct target gene of miR-375, was associated with human liver cancer growth, and the

Table 1 Clinical features of the hepatitis B virus cirrhosis and normal controls, *n* (%)

Variables	HBV cirrhosis (<i>n</i> = 40)	Normal controls (<i>n</i> = 40)	<i>P</i> value
Average age (yr)	42.38 ± 8.93	44.58 ± 11.79	0.356 ¹
Sex			0.133 ²
Male	26 (65.0)	32 (80.0)	
Female	14 (35.0)	8 (20.0)	
Alcohol consumption			0.056 ²
Ever or current	31 (77.5)	23 (57.5)	
Never	9 (22.5)	17 (42.5)	

¹Student-*t* test.²Two-sided χ^2 test. HBV: Hepatitis B virus.

upregulation of miR-375 can inhibit human liver cancer cell growth by regulating its cell apoptosis^[31]. The overexpression of GATA-binding factor 6, which is a downstream target of miR-143 in HCC, significantly increased cell proliferation and invasion rates in HCC, suggesting that miR-143 may suppress the malignancy of HCC by targeting GATA-binding factor 6^[32]. Liver-specific miR-122, which is essential for metabolic homeostasis, suppresses glucose-6-phosphate-dehydrogenase (G6PD) expression by directly interacting with its 3'UTR to achieve its anti-HCC efficacy. G6PD is the rate-limiting enzyme of the pentose phosphate pathway, which is often activated in human malignancies to generate precursors for nucleotide and lipid synthesis^[33]. MiR-122 overexpression inhibited the epithelial-mesenchymal transition by targeting Snail1 and Snail2 and regulated their expression levels to inhibit cell proliferation, colony formation and cell invasion in HCC cells^[34]. In summary, the high expression of miRNA-224 and low expression of miR-375, miR-122, and miR143 in HBV cirrhosis tissue may promote the development of liver cirrhosis to liver cancer. This may be the reason that smaller miRNA expression differences result in longer survival.

When performing RT-qPCR, it is critical to use a stably expressed gene as an internal standard for standardization. To date, no consensus has been established for endogenous controls in the study of circulating miRNAs. There are many different internal controls, such as RNU6B, RNU44, RNU48 and miR-16^[35,36], and as expected, the results are not the same. In this study, we chose the combination of let-7d, let-7g and let-7i as an internal reference, which is statistically superior and can better correct experimental changes^[20]. In addition, some other levels detection method also have been combined with miRNA, such as, CRISPER/Cas 9 system^[37,38], DNA methylation^[39], proteomics^[40], and metabolics, to involve the development of HCC.

Overall, we found a new miRNA group (miR-122, miR-375, miR-224 and miR-143) for the initial diagnosis of HBV cirrhosis and HBV infection, and compared with normal controls, patients with HBV cirrhosis had high expression of miR-224 and low expression of miR-375, miR-122 and miR-143. In addition, miR-224 low expression/miR-375, miR-122 and miR-143 high expression patients had a longer overall survival. In short, we identified four miRNAs as potential biomarkers for early diagnosis of HBV infection, HBV cirrhosis, and prediction of its overall survival.

Table 2 The relative concentration of miRNAs in hepatitis B virus cirrhosis samples and control samples¹ (*n* = 40)

miRNA	HBV cirrhosis	Normal control	Fold change ²	P value
miR-224	14.85	6.67	2.23	< 0.0001
miR-122	38.87	81.44	0.48	< 0.0001
miR-375	24.75	52.53	0.47	< 0.0001
miR-143	7.32	15.10	0.48	< 0.0001

¹miRNA concentrations are presented as the mean (SE).

²Hepatitis B virus cirrhosis/normal control. HBV: Hepatitis B virus.

Table 3 Risk score analysis of hepatitis B virus cirrhosis patients and normal controls

Score	0-2.016	> 20.16	PPV	NPV
Normal subject (<i>n</i> = 40)	34	6		0.92
HBV cirrhosis (<i>n</i> = 40)	3	37	0.86	
Total	37	43		

PPV: Positive predictive value; NPV: Negative predictive value; HBV: Hepatitis B virus.

Table 4 The overall survival of patients with different expression level (mo)

	miR-224	miR-122	miR-375	miR-143
Higher expression	34.45 ± 13.09	42.19 ± 12.98	40.75 ± 14.12	39.38 ± 15.08
Lower expression	41.72 ± 13.42	34.75 ± 13.39	34.70 ± 12.60	35.89 ± 11.77

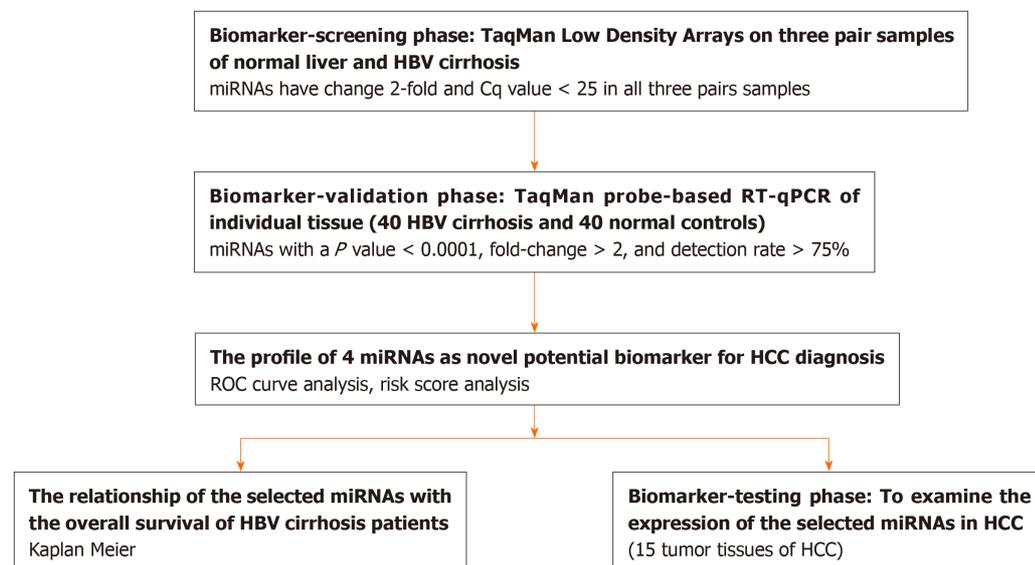


Figure 1 A flow chart of the experimental design. HCC: Hepatocellular carcinoma; HBV: Hepatitis B virus; RT-qPCR: Real time quantitative polymerase chain reaction.

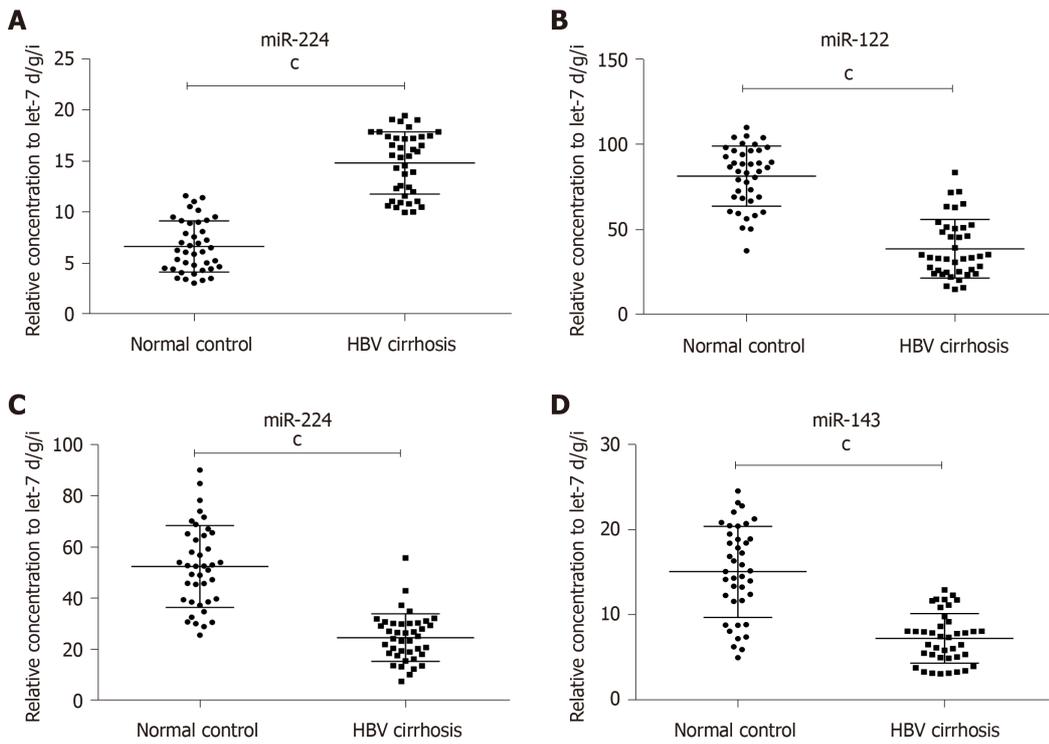


Figure 2 The relative concentration of the selected four miRNAs in the tissue from hepatitis B virus cirrhosis patients and normal controls. A: miR-224; B: miR-122; C: miR-375; D: miR-143. $^cP < 0.001$.

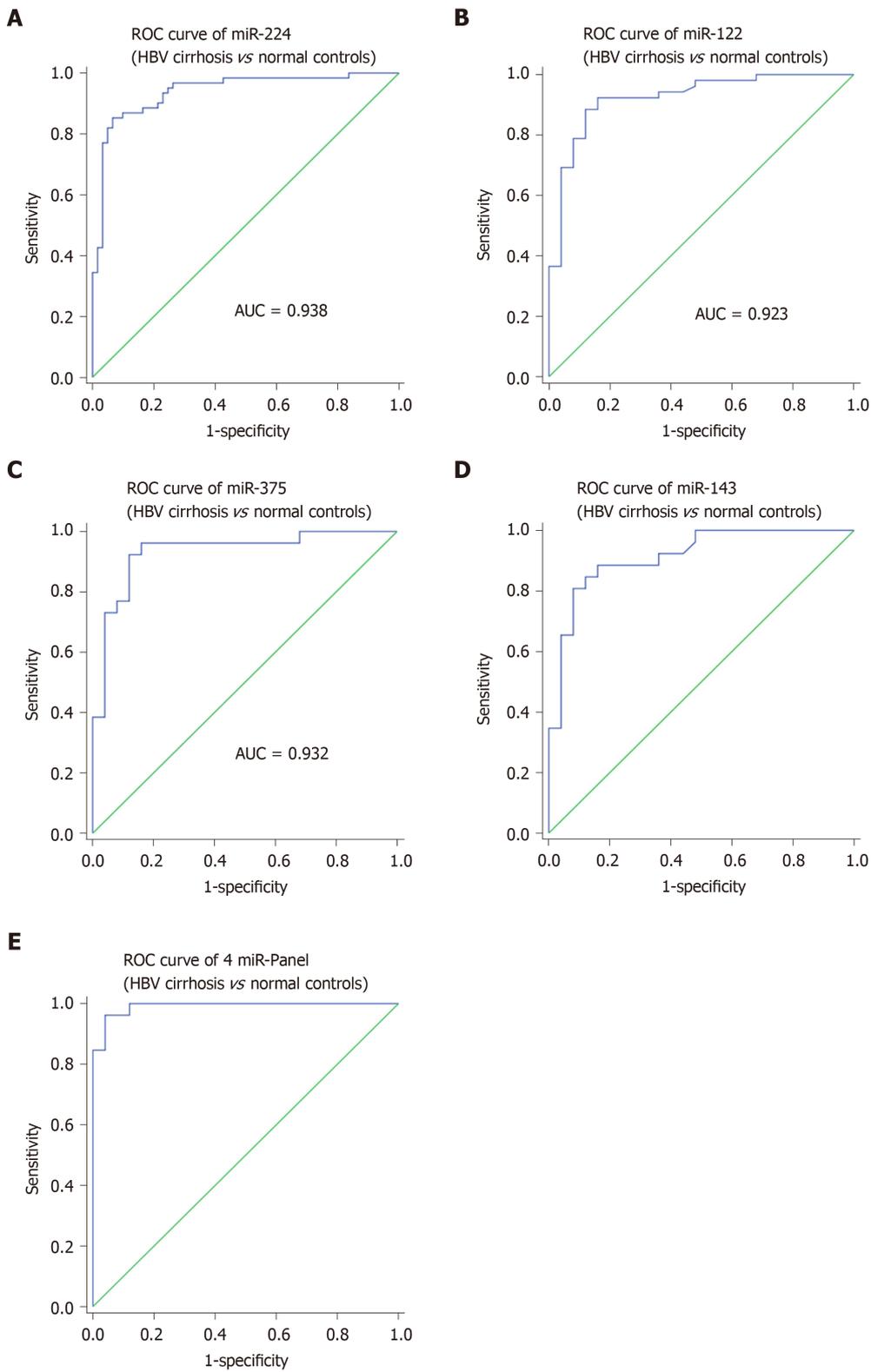


Figure 3 Sensitivity and specificity of the four-miRNA and their panel. A: Receiver operating characteristic (ROC) curve of miR-224; B: ROC curve of miR-122; C: ROC curve of miR-375; D: ROC curve of miR-143; E: ROC curve of 4 miR-Panel. ROC: Receiver operating characteristic curves; AUC: Area under curves; HBV: Hepatitis B virus.

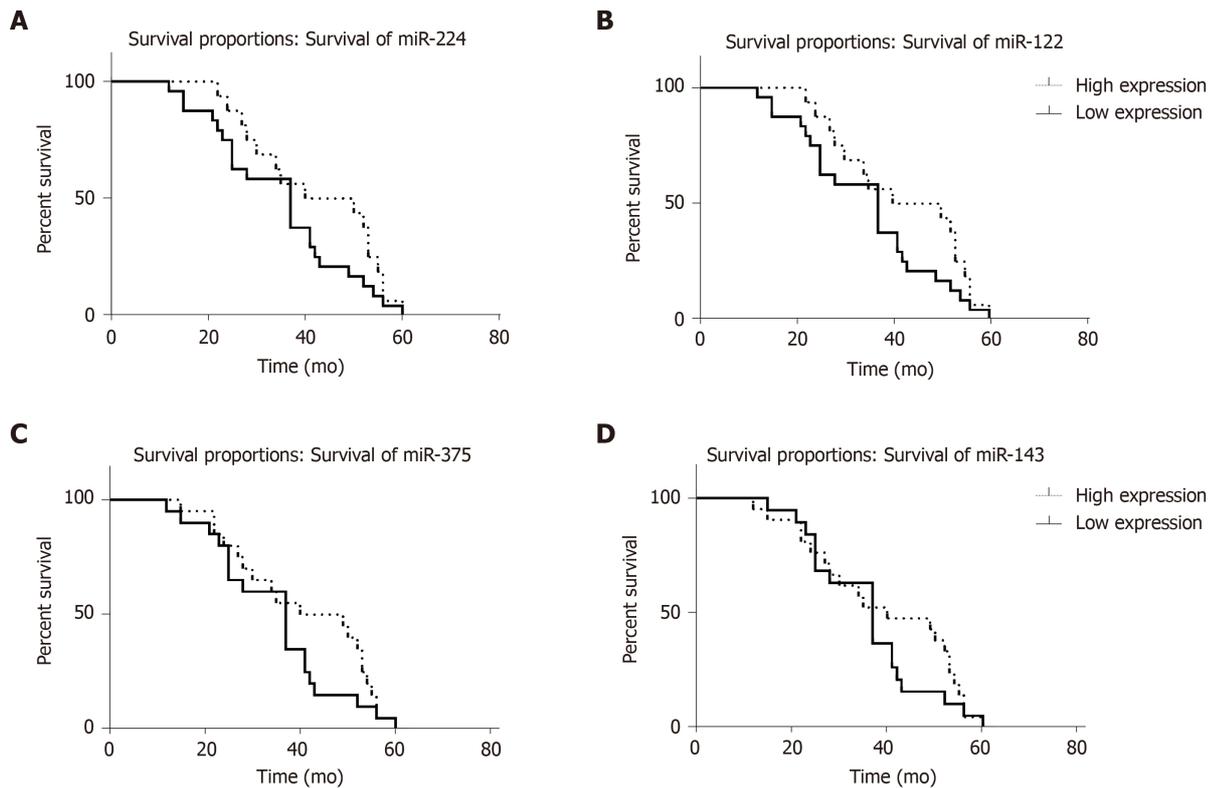


Figure 4 Kaplan-Meier curve of relationship between the expression of 4 miRNAs and overall survival of hepatitis B virus cirrhosis. A: Survival of miR-224; B: Survival of miR-122; C: Survival of miR-375; D: Survival of miR-143.

ARTICLE HIGHLIGHTS

Research background

The vast majority of liver cancer cases in China are closely related to hepatitis B virus (HBV) infection, but there are few studies on the changes of microRNAs (miRNA) expression in the progression from HBV infection to hepatoma.

Research motivation

In this study, TaqMan Low Density Array and real time quantitative polymerase chain reaction were used to characterize the profile of miRNAs in chronic hepatitis B, HCC and normal control tissues.

Research objectives

This study aimed to explore the role of miRNAs in the progression of HBV infection to cirrhosis and even to liver cancer.

Research methods

The authors screened differentially expressed miRNAs in 40 HBV cirrhosis, 40 normal and 15 HCC tissues. Authors calculated the area under the receiver-operating-characteristic curves.

Research results

The levels of miR-375, miR-122 and miR-143 were significantly lower in HBV cirrhosis tissues, while miR-224 was significantly higher than in the controls. The area under the curves of the receiver-operating-characteristic curve for the 4-miRNA panel was 0.991. Patients with a lower expression level of miR-224 or higher expression levels of miR-375, miR-122 and miR-143 had longer overall survival.

Research conclusions

The miR-375, miR-122, miR-143 and miR-224 may be helpful for early diagnosis of HBV infection, HBV cirrhosis, and prediction of its overall survival.

REFERENCES

- 1 **World Health Organization.** Global Hepatitis Report 2017. Geneva: World Health Organization, 2017. Available from: <http://apps.who.int/iris/bitstream/handle/10665/255016/9789241565455-eng.pdf;jsessionid=>

- sionid=7E29131834B6D97C84AC00E8742A0BFE?sequence=1
- 2 **Han Y**, Zeng A, Liao H, Liu Y, Chen Y, Ding H. The efficacy and safety comparison between tenofovir and entecavir in treatment of chronic hepatitis B and HBV related cirrhosis: A systematic review and Meta-analysis. *Int Immunopharmacol* 2017; **42**: 168-175 [PMID: 27915131 DOI: 10.1016/j.intimp.2016.11.022]
 - 3 **Iorio MV**, Ferracin M, Liu CG, Veronese A, Spizzo R, Sabbioni S, Magri E, Pedriali M, Fabbri M, Campiglio M, Ménard S, Palazzo JP, Rosenberg A, Musiani P, Volinia S, Nenci I, Calin GA, Querzoli P, Negrini M, Croce CM. MicroRNA gene expression deregulation in human breast cancer. *Cancer Res* 2005; **65**: 7065-7070 [PMID: 16103053 DOI: 10.1158/0008-5472.CAN-05-1783]
 - 4 **He H**, Jazdzewski K, Li W, Liyanarachchi S, Nagy R, Volinia S, Calin GA, Liu CG, Franssila K, Suster S, Kloos RT, Croce CM, de la Chapelle A. The role of microRNA genes in papillary thyroid carcinoma. *Proc Natl Acad Sci USA* 2005; **102**: 19075-19080 [PMID: 16365291 DOI: 10.1073/pnas.0509603102]
 - 5 **Chan JA**, Krichevsky AM, Kosik KS. MicroRNA-21 is an antiapoptotic factor in human glioblastoma cells. *Cancer Res* 2005; **65**: 6029-6033 [PMID: 16024602 DOI: 10.1158/0008-5472.CAN-05-0137]
 - 6 **Szafrańska AE**, Davison TS, John J, Cannon T, Sipos B, Maghnoji A, Labourier E, Hahn SA. MicroRNA expression alterations are linked to tumorigenesis and non-neoplastic processes in pancreatic ductal adenocarcinoma. *Oncogene* 2007; **26**: 4442-4452 [PMID: 17237814 DOI: 10.1038/sj.onc.1210228]
 - 7 **Lin SL**, Chiang A, Chang D, Ying SY. Loss of mir-146a function in hormone-refractory prostate cancer. *RNA* 2008; **14**: 417-424 [PMID: 18174313 DOI: 10.1261/rna.874808]
 - 8 **Lei KJ**, Lin YM, An GY. miR156 modulates rhizosphere acidification in response to phosphate limitation in Arabidopsis. *J Plant Res* 2016; **129**: 275-284 [PMID: 26659856 DOI: 10.1007/s10265-015-0778-8]
 - 9 **Matullo G**, Naccarati A, Pardini B. MicroRNA expression profiling in bladder cancer: the challenge of next-generation sequencing in tissues and biofluids. *Int J Cancer* 2016; **138**: 2334-2345 [PMID: 26489968 DOI: 10.1002/ijc.29895]
 - 10 **Xian X**, Tang L, Wu C, Huang L. miR-23b-3p and miR-130a-5p affect cell growth, migration and invasion by targeting CB1R via the Wnt/ β -catenin signaling pathway in gastric carcinoma. *Oncotargets Ther* 2018; **11**: 7503-7512 [PMID: 30498363 DOI: 10.2147/OTT.S181706]
 - 11 **Jiménez-Wences H**, Martínez-Carrillo DN, Peralta-Zaragoza O, Campos-Viguri GE, Hernández-Sotelo D, Jiménez-López MA, Muñoz-Camacho JG, Garzón-Barrientos VH, Illades-Aguier B, Fernández-Tilapa G. Methylation and expression of miRNAs in precancerous lesions and cervical cancer with HPV16 infection. *Oncol Rep* 2016; **35**: 2297-2305 [PMID: 26797462 DOI: 10.3892/or.2016.4583]
 - 12 **Bagheri A**, Khorshid HRK, Tavallaie M, Mowla SJ, Sherafatian M, Rashidi M, Zargari M, Boroujeni ME, Hosseini SM. A panel of noncoding RNAs in non-small-cell lung cancer. *J Cell Biochem* 2019; **120**: 8280-8290 [PMID: 30485511 DOI: 10.1002/jcb.28111]
 - 13 **Murakami Y**, Yasuda T, Saigo K, Urashima T, Toyoda H, Okanou T, Shimotohno K. Comprehensive analysis of microRNA expression patterns in hepatocellular carcinoma and non-tumorous tissues. *Oncogene* 2006; **25**: 2537-2545 [PMID: 16331254 DOI: 10.1038/sj.onc.1209283]
 - 14 **Jiang J**, Gusev Y, Aderca I, Mettler TA, Nagorney DM, Brackett DJ, Roberts LR, Schmittgen TD. Association of MicroRNA expression in hepatocellular carcinomas with hepatitis infection, cirrhosis, and patient survival. *Clin Cancer Res* 2008; **14**: 419-427 [PMID: 18223217 DOI: 10.1158/1078-0432.CCR-07-0523]
 - 15 **Huang YS**, Dai Y, Yu XF, Bao SY, Yin YB, Tang M, Hu CX. Microarray analysis of microRNA expression in hepatocellular carcinoma and non-tumorous tissues without viral hepatitis. *J Gastroenterol Hepatol* 2008; **23**: 87-94 [PMID: 18171346 DOI: 10.1111/j.1440-1746.2007.05223.x]
 - 16 **Budhu A**, Jia HL, Forgues M, Liu CG, Goldstein D, Lam A, Zanetti KA, Ye QH, Qin LX, Croce CM, Tang ZY, Wang XW. Identification of metastasis-related microRNAs in hepatocellular carcinoma. *Hepatology* 2008; **47**: 897-907 [PMID: 18176954 DOI: 10.1002/hep.22160]
 - 17 **Wang Y**, Lee AT, Ma JZ, Wang J, Ren J, Yang Y, Tantoso E, Li KB, Ooi LL, Tan P, Lee CG. Profiling microRNA expression in hepatocellular carcinoma reveals microRNA-224 up-regulation and apoptosis inhibitor-5 as a microRNA-224-specific target. *J Biol Chem* 2008; **283**: 13205-13215 [PMID: 18319255 DOI: 10.1074/jbc.M707629200]
 - 18 **Wu C**, Wang C, Guan X, Liu Y, Li D, Zhou X, Zhang Y, Chen X, Wang J, Zen K, Zhang CY, Zhang C. Diagnostic and prognostic implications of a serum miRNA panel in oesophageal squamous cell carcinoma. *PLoS One* 2014; **9**: e92292 [PMID: 24651474 DOI: 10.1371/journal.pone.0092292]
 - 19 **Luo Y**, Wang C, Chen X, Zhong T, Cai X, Chen S, Shi Y, Hu J, Guan X, Xia Z, Wang J, Zen K, Zhang CY, Zhang C. Increased serum and urinary microRNAs in children with idiopathic nephrotic syndrome. *Clin Chem* 2013; **59**: 658-666 [PMID: 23344497 DOI: 10.1373/clinchem.2012.195297]
 - 20 **Chen X**, Liang H, Guan D, Wang C, Hu X, Cui L, Chen S, Zhang C, Zhang J, Zen K, Zhang CY. A combination of Let-7d, Let-7g and Let-7i serves as a stable reference for normalization of serum microRNAs. *PLoS One* 2013; **8**: e79652 [PMID: 24223986 DOI: 10.1371/journal.pone.0079652]
 - 21 **Livak KJ**, Schmittgen TD. Analysis of relative gene expression data using real-time quantitative PCR and the 2(-Delta Delta C(T)) Method. *Methods* 2001; **25**: 402-408 [PMID: 11846609 DOI: 10.1006/meth.2001.1262]
 - 22 **Yang C**, Wang C, Chen X, Chen S, Zhang Y, Zhi F, Wang J, Li L, Zhou X, Li N, Pan H, Zhang J, Zen K, Zhang CY, Zhang C. Identification of seven serum microRNAs from a genome-wide serum microRNA expression profile as potential noninvasive biomarkers for malignant astrocytomas. *Int J Cancer* 2013; **132**: 116-127 [PMID: 22674182 DOI: 10.1002/ijc.27657]
 - 23 **Vandesompele J**, De Preter K, Pattyn F, Poppe B, Van Roy N, De Paepe A, Speleman F. Accurate normalization of real-time quantitative RT-PCR data by geometric averaging of multiple internal control genes. *Genome Biol* 2002; **3**: RESEARCH0034 [PMID: 12184808]
 - 24 **Andersen CL**, Jensen JL, Ørntoft TF. Normalization of real-time quantitative reverse transcription-PCR data: a model-based variance estimation approach to identify genes suited for normalization, applied to bladder and colon cancer data sets. *Cancer Res* 2004; **64**: 5245-5250 [PMID: 15289330 DOI: 10.1158/0008-5472.CAN-04-0496]
 - 25 **Zhang GL**, Li YX, Zheng SQ, Liu M, Li X, Tang H. Suppression of hepatitis B virus replication by microRNA-199a-3p and microRNA-210. *Antiviral Res* 2010; **88**: 169-175 [PMID: 20728471 DOI: 10.1016/j.antiviral.2010.08.008]
 - 26 **Kohno T**, Tsuge M, Murakami E, Hiraga N, Abe H, Miki D, Imamura M, Ochi H, Hayes CN, Chayama K. Human microRNA hsa-miR-1231 suppresses hepatitis B virus replication by targeting core mRNA. *J Viral Hepat* 2014; **21**: e89-e97 [PMID: 24835118 DOI: 10.1111/jvh.12240]
 - 27 **Yang X**, Li H, Sun H, Fan H, Hu Y, Liu M, Li X, Tang H. Hepatitis B Virus-Encoded MicroRNA

- Controls Viral Replication. *J Virol* 2017; **91** [PMID: 28148795 DOI: 10.1128/JVI.01919-16]
- 28 **Li LM**, Hu ZB, Zhou ZX, Chen X, Liu FY, Zhang JF, Shen HB, Zhang CY, Zen K. Serum microRNA profiles serve as novel biomarkers for HBV infection and diagnosis of HBV-positive hepatocarcinoma. *Cancer Res* 2010; **70**: 9798-9807 [PMID: 21098710 DOI: 10.1158/0008-5472.CAN-10-1001]
- 29 **Akamatsu S**, Hayes CN, Tsuge M, Miki D, Akiyama R, Abe H, Ochi H, Hiraga N, Imamura M, Takahashi S, Aikata H, Kawaoka T, Kawakami Y, Ohishi W, Chayama K. Differences in serum microRNA profiles in hepatitis B and C virus infection. *J Infect* 2015; **70**: 273-287 [PMID: 25452043 DOI: 10.1016/j.jinf.2014.10.017]
- 30 **Hung JH**, Li CH, Yeh CH, Huang PC, Fang CC, Chen YF, Lee KJ, Chou CH, Cheng HY, Huang HD, Chen M, Tsai TF, Lin AM, Yen CH, Tsou AP, Tyan YC, Chen YA. MicroRNA-224 down-regulates Glycine N-methyltransferase gene expression in Hepatocellular Carcinoma. *Sci Rep* 2018; **8**: 12284 [PMID: 30115977 DOI: 10.1038/s41598-018-30682-5]
- 31 **Li L**, Jia L, Ding Y. Upregulation of miR-375 inhibits human liver cancer cell growth by modulating cell proliferation and apoptosis via targeting ErbB2. *Oncol Lett* 2018; **16**: 3319-3326 [PMID: 30127930 DOI: 10.3892/ol.2018.9011]
- 32 **Xue F**, Yin J, Xu L, Wang B. MicroRNA-143 inhibits tumorigenesis in hepatocellular carcinoma by downregulating GATA6. *Exp Ther Med* 2017; **13**: 2667-2674 [PMID: 28587328 DOI: 10.3892/etm.2017.4348]
- 33 **Barajas JM**, Reyes R, Guerrero MJ, Jacob ST, Motiwala T, Ghoshal K. The role of miR-122 in the dysregulation of glucose-6-phosphate dehydrogenase (G6PD) expression in hepatocellular cancer. *Sci Rep* 2018; **8**: 9105 [PMID: 29904144 DOI: 10.1038/s41598-018-27358-5]
- 34 **Jin Y**, Wang J, Han J, Luo D, Sun Z. MiR-122 inhibits epithelial-mesenchymal transition in hepatocellular carcinoma by targeting Snail1 and Snail2 and suppressing WNT/ β -cadherin signaling pathway. *Exp Cell Res* 2017; **360**: 210-217 [PMID: 28890291 DOI: 10.1016/j.yexcr.2017.09.010]
- 35 **Chang KH**, Mestdagh P, Vandesompele J, Kerin MJ, Miller N. MicroRNA expression profiling to identify and validate reference genes for relative quantification in colorectal cancer. *BMC Cancer* 2010; **10**: 173 [PMID: 20429937 DOI: 10.1186/1471-2407-10-173]
- 36 **Kroh EM**, Parkin RK, Mitchell PS, Tewari M. Analysis of circulating microRNA biomarkers in plasma and serum using quantitative reverse transcription-PCR (qRT-PCR). *Methods* 2010; **50**: 298-301 [PMID: 20146939 DOI: 10.1016/j.ymeth.2010.01.032]
- 37 **Gao W**, Long L, Tian X, Xu F, Liu J, Singh PK, Botella JR, Song C. Genome Editing in Cotton with the CRISPR/Cas9 System. *Front Plant Sci* 2017; **8**: 1364 [PMID: 28824692 DOI: 10.3389/fpls.2017.01364]
- 38 **Guo J**, Li K, Jin L, Xu R, Miao K, Yang F, Qi C, Zhang L, Botella JR, Wang R, Miao Y. A simple and cost-effective method for screening of CRISPR/Cas9-induced homozygous/biallelic mutants. *Plant Methods* 2018; **14**: 40 [PMID: 29872452 DOI: 10.1186/s13007-018-0305-8]
- 39 **Sun Q**, Qiao J, Zhang S, He S, Shi Y, Yuan Y, Zhang X, Cai Y. Changes in DNA methylation assessed by genomic bisulfite sequencing suggest a role for DNA methylation in cotton fruiting branch development. *PeerJ* 2018; **6**: e4945 [PMID: 29915693 DOI: 10.7717/peerj.4945]
- 40 **Yu J**, Zhang Y, Liu J, Wang L, Liu P, Yin Z, Guo S, Ma J, Lu Z, Wang T, She Y, Miao Y, Ma L, Chen S, Li Y, Dai S. Proteomic discovery of H₂O₂ response in roots and functional characterization of PutGLP gene from alkaligrass. *Planta* 2018; **248**: 1079-1099 [PMID: 30039231 DOI: 10.1007/s00425-018-2940-8]



Published By Baishideng Publishing Group Inc
7041 Koll Center Parkway, Suite 160, Pleasanton, CA 94566, USA
Telephone: +1-925-3991568
E-mail: bpgoffice@wjgnet.com
Help Desk: <https://www.f6publishing.com/helpdesk>
<https://www.wjgnet.com>

