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***Retrospective Study***

**Accuracy of endoscopic ultrasound-guided needle aspiration specimens for molecular diagnosis of non-small-cell lung carcinoma**

Su W *et al*. EBUS-TBNA and EUS-FNA in NSCLC diagnosis

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**Abstract**

BACKGROUND

Endoscopic ultrasonography-guided fine-needle aspiration (EUS-FNA) and endobronchial ultrasound-guided transbronchial needle aspiration (EBUS-TBNA) are highly sensitive for diagnosing and staging lung cancer. In recent years, targeted therapy has shown great significance in the treatment of non-small cell lung carcinoma (NSCLC). Using these minimally invasive techniques to obtain specimens for molecular testing will provide patients with a more convenient diagnostic approach.

AIM

To evaluate the feasibility and accuracy of tissue samples obtained using EUS-FNA and EBUS-TBNA for molecular diagnosis of NSCLC.

METHODS

A total of 83 patients with NSCLC underwent molecular testing using tissues obtained from EUS-FNA or EBUS-TBNA at the Tianjin Medical University Cancer Hospital from January 2017 to June 2019. All enrolled patients underwent chest computed tomography or positron emission tomography/computed tomography prior to puncture. We detected abnormal expression of *EGFR*, *KRAS*, *MET*, *HER2*, *ROS1* and anaplastic lymphoma kinase protein. Two patients failed to complete molecular testing due to insufficient tumor tissue. The clinical features, puncture records, molecular testing results and targeted treatment in the remaining 81 patients were summarized.

RESULTS

In a total of 99 tissue samples obtained from 83 patients, molecular testing was successfully completed in 93 samples with a sample adequacy ratio of 93.9% (93/99). Biopsy samples from two patients failed to provide test results due to insufficient tumor tissue. In the remaining 81 patients, 62 cases (76.5%) were found to have adenocarcinoma, 11 cases (13.6%) had squamous cell carcinoma, 3 cases (3.7%) had adenosquamous carcinoma and 5 cases (6.2%) had NSCLC-not otherwise specified. The results of molecular testing showed *EGFR* mutations in 21 cases (25.9%), *KRAS* mutations in 9 cases (11.1%), *ROS-1* rearrangement in 1 case (1.2%) and anaplastic lymphoma kinase-positive in 5 cases (6.2%). Twenty-four patients with positive results received targeted therapy. The total effectiveness rate of targeted therapy was 66.7% (16/24), and the disease control rate was 83.3% (20/24).

CONCLUSION

Tissue samples obtained by EUS-FNA or EBUS-TBNA are feasible for the molecular diagnosis of NSCLC and can provide reliable evidence for clinical diagnosis and treatment.

**Key Words:** Endobronchial ultrasound-guided transbronchial needle aspiration; Endoscopic ultrasonography-guided fine-needle aspiration; Non-small cell lung carcinoma; Molecular diagnosis; Targeted therapy

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**Core Tip:** The purpose of this retrospective study was to evaluate the feasibility and reliability of tissue samples obtained using endoscopic ultrasonography-guided fine-needle aspiration and endobronchial ultrasound-guided transbronchial needle aspiration for molecular diagnosis of non-small cell lung carcinoma. In the study, 93.9% of the puncture samples could be used for molecular testing. The proportion of patients with positive results corresponds to the frequency of molecular mutations. Patients receiving targeted therapy responded well to treatment. The samples obtained by the two techniques can be used for molecular diagnosis of lung cancer. They can provide reliable evidence for clinical diagnosis and treatment.

**INTRODUCTION**

According to the latest cancer statistics released in 2018, lung cancer still has the highest morbidity and mortality worldwide. Most patients have locally advanced tumors or distant metastasis at diagnosis. The 5-year survival rate is only 16.1%[1,2]. At present, the treatment of advanced lung cancer mainly depends on comprehensive treatment and individualized treatment strategies that are formulated according to pathological types, molecular genetic characteristics and patient condition[3]. In recent years, targeted therapy for non-small cell lung cancer (NSCLC) has played an important role in clinical applications. Therefore, obtaining tumor tissue or metastatic lymph node tissue for molecular testing before first-line treatment is essential for timely and effective individualized treatment. In China, NSCLC accounts for 80%-85% of all lung cancer cases[1]. The National Comprehensive Cancer Network guidelines clearly indicate that molecular testing should be undertaken in all cases of NSCLC, including lung adenocarcinoma, large cell carcinoma and NSCLC-not otherwise specified[4]. For squamous cell carcinoma, molecular testing is recommended for nonsmokers, patients with small biopsy specimens or mixed histological types[4]. *EGFR*, *KRAS* and anaplastic lymphoma kinase (ALK) should be routinely tested. It has been suggested that more extensive molecular testing is a key factor in NSCLC therapy.

Endoscopic ultrasound-guided minimally invasive diagnosis and treatment technology has been widely used in many diseases in recent years[5-8]. Endoscopic ultrasonography-guided fine-needle aspiration (EUS-FNA) and endobronchial ultrasound-guided transbronchial needle aspiration (EBUS-TBNA) have high sensitivity and specificity for diagnosing and staging lung cancer[9,10] and have become safe and effective methods to establish diagnosis for patients with clinically suspected advanced-stage disease. Due to the different accessibility of lung and mediastinal lesions, the two biopsy methods are considered to be complementary. They can puncture almost all of the mediastinal lymph nodes[11]. The purpose of this retrospective study was to evaluate the feasibility and reliability of EUS-FNA and EBUS-TBNA in obtaining tissue for the molecular diagnosis of NSCLC.

**MATERIALS AND METHODS**

***Patients***

A total of 83 patients with NSCLC underwent molecular testing using tissues obtained from EUS-FNA or EBUS-TBNA at the Tianjin Medical University Cancer Hospital from January 2017 to June 2019. Two patients failed to complete the molecular testing due to insufficient tumor tissue. We summarized the molecular testing results and targeted treatment in the remaining 81 patients. All enrolled patients underwent chest computed tomography or positron emission tomography/computed tomography prior to puncture. The results showed a highly suspicious malignant lung mass or metastatic lymph node. In addition, we reviewed the targeted therapy of patients with positive molecular testing results and evaluated tumor response after 3 mo of targeted therapy. Clinical efficacy was evaluated according to the response evaluation criteria in solid tumors. All patients signed an informed consent before examination. The ethics committee of our institution approved the study.

***EUS-FNA and EBUS-TBNA***

EBUS-TBNA was performed with a convex probe endobronchial ultrasound for line array scanning (BF-UC206FW, Olympus) and a 22-gauge ultrasound bronchial biopsy needle for tissue aspiration (ECHO-HD-22-EBUS-O, Cook). We performed EUS-FNA with a convex ultrasound endoscope (EG-3870UTK, Pentax) and a 22-gauge ultrasound biopsy needle (ECHO-3-22, Cook). The biopsy specimens were collected for cytology, histopathology and molecular testing. Rapid on-site evaluation of cytopathology was not performed.

***Pathology and molecular analysis of specimens***

The collected specimens were fixed with 40 g/L formaldehyde solution and were embedded in paraffin. Processed samples were used for histopathological diagnosis and classification by hematoxylin eosin staining and immunohistochemistry (IHC). If the number of tumor cells exceeded 100 in the paraffin section, they were considered feasible for molecular testing. Following a definite diagnosis, we extracted DNA from paraffin tissue blocks and used Sanger sequencing for mutation analysis of *EGFR* exons 18-21, *KRAS* exon 2, *MET* exon 14 and *HER2* exon 20. The expression of ALK protein was detected by Ventana ALK (D5F3) IHC. The expression of ROS1 protein was preliminarily determined by IHC. Further fluorescence *in situ* hybridization testing was required in patients with IHC results of + to +++. The positive criteria on fluorescence *in situ* hybridization detection were more than 50 tumor cells in the tissue sections and more than 15% of the counted cells showing a separation signal.

***Statistical analysis***

IBM SPSS Statistics (v24.0; IBM Corp., United States) were used for data analysis. Continuous variables are presented as mean ± standard deviation and categorical variables as the frequency (*n*) and percentage (%).

**RESULTS**

***Clinical characteristics of the patients***

In this study, molecular testing was performed in 83 patients. At least one suspected malignant lung lesion or one suspected metastatic lymph node was biopsied in each patient. In a total of 99 lesion samples obtained from 83 patients, molecular testing was successfully performed in 93 samples with a sample adequacy ratio of 93.9% (93/99). Molecular testing failed in two patients due to insufficient biopsied tumor tissue. The clinical characteristics of the remaining 81 patients who completed the molecular testing are summarized in Table 1. Of these patients, 59 were male (72.8%), and 22 were female (27.2%) with a mean age of 59.1 ± 9.4 years (28-76 years). Forty-two patients (51.9%) had a history of smoking, and 39 (48.1%) were nonsmokers. Fifteen patients (18.5%) had a family history of lung cancer. Histopathological analysis revealed that 62 cases (76.5%) had adenocarcinoma, 11 cases (13.6%) had squamous cell carcinoma, 3 cases (3.7%) had adenosquamous carcinoma and 5 cases (6.2%) had NSCLC-not otherwise specified. All patients were at an intermediate or advanced stage (stage III-IV). We performed EUS-FNA on 44 patients (54.3%) and EBUS-TBNA on 37 patients (45.7%).

A total of 97 lesion samples were obtained from 81 patients. One patient underwent biopsy of 3 lesions, and 14 patients underwent biopsy of 2 lesions. The location and size of the lung lesions and lymph nodes sampled are shown in Table 2. The specific conditions of multifocal biopsy are shown in Table 3. Each lesion was punctured 2-4 times with an average of 2.7 ± 0.6 times. In patients with multifocal biopsy, one sample did not provide a reliable malignant diagnosis in two patients. In another two patients, molecular testing was not completed due to insufficient tumor tissue.

***Molecular analysis of the samples***

Of the 81 patients with NSCLC, 21 (25.9%) had *EGFR* mutations, 9 (11.1%) had *KRAS* mutations, 1 (1.2%) had *ROS-1* rearrangements and 5 (6.2%) were ALK positive. However, the detection of *MET* and *HER2* did not yield a positive result. Data analysis showed that genetic mutations were still concentrated in patients with lung adenocarcinoma. Among the 62 patients with lung adenocarcinoma and 3 patients with adenosquamous carcinoma, there were 21 cases (32.3%) of *EGFR* mutations, 8 cases (12.3%) of *KRAS* mutations, 1 case (1.5%) of *ROS-1* rearrangements and 5 cases were ALK positive (7.7%). However, in eleven patients with lung squamous cell carcinoma, only one *KRAS* mutation was detected, and in five patients with NSCLC-not otherwise specified, there were no positive results. Table 4 lists the results of molecular testing.

Of the 15 patients who underwent multifocal sampling, 4 patients had positive results. These four patients underwent biopsy of two lesions. One of the patients had insufficient tumor tissue in one of the samples, and in the other three patients we found that different lesions in the same patient had consistent mutation results (Table 5).

***Evaluation of the efficacy of targeted therapy***

A total of 36 patients had positive molecular testing results, 24 of them received targeted therapy, including 19 cases of *EGFR* mutations, 1 case of *ROS-1* rearrangement and 4 ALK-positive cases. Therapeutic drugs included gefitinib, erlotinib, alectinib and crizotinib. The evaluation of tumor response after 3 mo of treatment is shown in Table 6. The total effectiveness rate of targeted therapy was 66.7% (16/24), and the disease control rate was 83.3% (20/24).

**DISCUSSION**

Due to limited cancer screening and other reasons, approximately 70% of lung cancer patients are diagnosed at a late stage of the disease[12]. Therefore, they have missed or lost the best surgical opportunity. These patients usually have a poor prognosis and a high mortality rate. In recent years, EUS-FNA and EBUS-TBNA have been widely used in the diagnosis and staging of lung cancer, which has greatly shortened the time for treatment decision-making compared with conventional techniques such as thoracoscopic surgery[13]. In addition, because of their high sensitivity and specificity, they can improve the accuracy of lymph node staging, thereby reducing the number of unnecessary surgical interventions and have become the preferred methods for diagnosis and lymph node evaluation in patients with advanced lung cancer[14,15].

With the advancement of targeted therapy for lung cancer, more convenient, rapid and accurate acquisition of patients’ molecular testing results can greatly improve the efficiency of clinical diagnosis and treatment. Based on relevant guidelines and listed targeted drugs, our center provides a molecular testing platform for the corresponding targets. Our research involved the analysis of tumor tissue obtained using EUS-FNA or EBUS-TBNA for molecular testing in NSCLC. We tested *EGFR*, *KRAS*, *MET*, *HER2*, *ROS1* and ALK simultaneously. If the amount of tumor tissue was insufficient and all molecules could not be detected, the sample was considered invalid. A total of 99 tissue samples were obtained from 83 patients, of which 93 samples provided sufficient tumor tissue for molecular detection with a sample adequacy ratio of 93.9%. In a meta-analysis of 28 studies evaluating EBUS-TBNA for the identification of *EGFR* and *ALK* mutations, the pooled probability of obtaining a sufficient sample for the *EGFR* assay was 94.5% [95% confidence interval, 93.2%-96.4%], and the pooled probability was 94.9% for *ALK* mutations (95% confidence interval, 89.4-98.8%)[16]. Folch *et al*[17] obtained lung cancer tissue samples from the hilum pulmonis or mediastinal lymph nodes *via* convex probe-EBUS for *EGFR*, *KRAS* and *ALK* gene detection with a success rate of over 90%. Our research had approximate success rates compared to similar types of studies and completed the detection of multiple molecular targets. Most incomplete molecular diagnostic samples were due to the puncture site containing a large amount of necrotic tumor tissue or blood loss. Research has shown that selecting larger lymph nodes and at least three punctures per lesion may result in a higher success rate in molecular testing[18].

Due to differences in the number of samples, smoking history, ratio of male to female patients and the sensitivity of detection methods, there are differences in the frequencies of abnormal molecules in different studies. Available related studies and reviews concluded that the mutation frequencies of *EGFR*, *KRAS*, *ALK*, *ROS1*, *MET* and *HER2* in Asian lung adenocarcinoma are approximately 30%-50%, 8%-16%, 3%-7%, 1%-3%, 3%-4% and 1%-3%, respectively[19-24]. In the present study, gene mutations mainly occurred in patients with lung adenocarcinoma. In 62 patients with lung adenocarcinoma and 3 patients with adenosquamous carcinoma, there were 21 cases (32.3%) of *EGFR* mutations, 8 cases (12.3%) of *KRAS* mutations, 1 case (1.5%) of *ROS-1* rearrangements and 5 cases were ALK positive (7.7%). These results are broadly similar to those observed in the above studies. In addition, mutation-positive patients responded well to targeted therapy in this study with a total effectiveness rate of 66.7% (16/24) and a disease control rate of 83.3% (20/24). It can be seen that tissue samples obtained by EUS-FNA and EBUS-TBNA for molecular detection are reliable. In addition, we did not obtain positive results for *MET* and *HER2* mutations, and there were no multiple mutations due to an insufficient number of cases.

Of the fifteen patients who underwent multifocal sampling, four patients had positive results. One sample had insufficient tumor tissue, and in the other three patients it was found that different lesions in the same patient had consistent mutation results. Kang *et al*[25] compared the *EGFR* mutation status of lymph node samples obtained by EBUS-TBNA and primary tumor samples and found that the discordance rate in major mutations between matched primary tumor and lymph node specimens was 4.1% (3/74). Ito *et al*[26]compared the *EGFR* and *ALK* mutation status of lymph node samples and primary tumor samples and found that the concordance rate was 96.7%. Due to the limitations of this retrospective study, we did not have enough data on molecular testing to compare matched primary tumor and lymph node specimens. Further research should be performed in the future. In addition, suspected lesions should be sampled as much as possible in clinical applications to obtain a more comprehensive and reliable diagnosis.

**CONCLUSION**

In conclusion, from this analysis, it is believed that the molecular diagnosis of NSCLC patients following the evaluation of tissue samples obtained by EUS-FNA or EBUS-TBNA is reliable. At present, our center provides patients with a next-generation sequencing detection platform, which requires fewer specimens and can detect multiple genes at the same time[27]. EUS-FNA and EBUS-TBNA will provide more efficient help for patients with lung cancer.

**ARTICLE HIGHLIGHTS**

***Research background***

Endoscopic ultrasound-guided needle aspiration technology is applied to the diagnosis of lung cancer, which exempts many patients from undergoing more traumatic examinations. It has been gradually promoted in clinical practice. Whether the limited puncture tissue can be used for pathological diagnosis, molecular diagnosis, *etc.* and obtain reliable results, is the question we wanted to clarify in order to help us understand whether the technology can efficiently assist clinical diagnosis and treatment.

***Research motivation***

To evaluate the value of endoscopic ultrasound-guided needle aspiration in the diagnosis of lung cancer.

***Research objectives***

Through retrospective research to understand the success rate of molecular diagnosis of non-small cell lung carcinoma with puncture specimens. To evaluate the reliability of the diagnosis results through follow-up of clinical treatment effects.

***Research methods***

According to the location of the patient’s lesion, we choose to use endoscopic ultrasonography-guided fine-needle aspiration or endobronchial ultrasound-guided transbronchial needle aspiration. Due to the different accessibility of lung and mediastinal lesions, the two puncture methods are considered to be complementary. We used Sanger sequencing for mutation analysis of *EGFR*, *KRAS*, *MET* and *HER2*. This is not the most advanced detection method, but the test results are reliable. Moreover, it does not require high costs, so more patients can benefit from it.

***Research results***

In this study, 93.9% of the punctured tissues met the molecular test standards. The test results were in line with the mutation frequency of the patient population. The patients who received targeted therapy according to the test results responded well. These results add evidence to support the application value of this technology in diagnosis of non-small cell lung carcinoma. However, this study lacks comparison data between this technique and other methods, so it cannot prove that it is the first choice for clinical diagnosis.

***Research conclusions***

The two puncture methods are considered to be complementary. They can puncture almost all the mediastinal lymph nodes. This technique obtains diseased tissue under minimally invasive conditions, thus reducing unnecessary surgical intervention. The diagnosis results are reliable and can effectively guide clinical treatment.

***Research perspectives***

Advantages and problems of endoscopic ultrasound in diagnosis and treatment of lung cancer and gastrointestinal tumors.

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**REFERENCES**

1 **Chen W**, Sun K, Zheng R, Zeng H, Zhang S, Xia C, Yang Z, Li H, Zou X, He J. Cancer incidence and mortality in China, 2014. *Chin J Cancer Res* 2018; **30**: 1-12 [PMID: 29545714 DOI: 10.21147/j.issn.1000-9604.2018.01.01]

2 **Bray F**, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, Jemal A. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J Clin* 2018; **68**: 394-424 [PMID: 30207593 DOI: 10.3322/caac.21492]

3 **Buettner R**, Wolf J, Thomas RK. Lessons learned from lung cancer genomics: the emerging concept of individualized diagnostics and treatment. *J Clin Oncol* 2013; **31**: 1858-1865 [PMID: 23589544 DOI: 10.1200/JCO.2012.45.9867]

4 **Ettinger DS**, Wood DE, Aisner DL, Akerley W, Bauman J, Chirieac LR, D'Amico TA, DeCamp MM, Dilling TJ, Dobelbower M, Doebele RC, Govindan R, Gubens MA, Hennon M, Horn L, Komaki R, Lackner RP, Lanuti M, Leal TA, Leisch LJ, Lilenbaum R, Lin J, Loo BW Jr, Martins R, Otterson GA, Reckamp K, Riely GJ, Schild SE, Shapiro TA, Stevenson J, Swanson SJ, Tauer K, Yang SC, Gregory K, Hughes M. Non-Small Cell Lung Cancer, Version 5.2017, NCCN Clinical Practice Guidelines in Oncology. *J Natl Compr Canc Netw* 2017; **15**: 504-535 [PMID: 28404761 DOI: 10.6004/jnccn.2017.0050]

5 **Wang G**, Liu X, Wang S, Ge N, Guo J, Sun S. Endoscopic Ultrasound-guided Gastroenterostomy: A Promising Alternative to Surgery. *J Transl Int Med* 2019; **7**: 93-99 [PMID: 31637179 DOI: 10.2478/jtim-2019-0021]

6 **Theerasuwipakorn N**, Tasneem AA, Kongkam P, Angsuwatcharakon P, Ridtitid W, Navicharern P, Kitisin K, Wangrattanapranee P, Rerknimitr R, Kullavanijaya P. Walled-off Peripancreatic Fluid Collections in Asian Population: Paradigm Shift from Surgical and Percutaneous to Endoscopic Drainage. *J Transl Int Med* 2019; **7**: 170-177 [PMID: 32010603 DOI: 10.2478/jtim-2019-0032]

7 **Nunes G**, Marques PP, Patita M, Allen M, Gargaté L. EUS-guided recanalization of complete colorectal anastomotic stenosis using a lumen-apposing metal stent. *Endosc Ultrasound* 2019; **8**: 211-212 [PMID: 30785118 DOI: 10.4103/eus.eus\_62\_18]

8 **Braden B**, Gupta V, Dietrich CF. Therapeutic EUS: New tools, new devices, new applications. *Endosc Ultrasound* 2019; **8**: 370-381 [PMID: 31417067 DOI: 10.4103/eus.eus\_39\_19]

9 **Colella S**, Vilmann P, Konge L, Clementsen PF. Endoscopic ultrasound in the diagnosis and staging of lung cancer. *Endosc Ultrasound* 2014; **3**: 205-212 [PMID: 25485267 DOI: 10.4103/2303-9027.144510]

10 **Liran L**, Rottem K, Gregorio FZ, Avi A, Neville B. A novel, stepwise approach combining conventional and endobronchial ultrasound needle aspiration for mediastinal lymph node sampling. *Endosc Ultrasound* 2019; **8**: 31-35 [PMID: 28879863 DOI: 10.4103/eus.eus\_29\_17]

11 **Mondoni M**, D'Adda A, Terraneo S, Carlucci P, Radovanovic D, DI Marco F, Santus P. Choose the best route: ultrasound-guided transbronchial and transesophageal needle aspiration with echobronchoscope in the diagnosis of mediastinal and pulmonary lesions. *Minerva Med* 2015; **106**: 13-19 [PMID: 27427262]

12 **Molina JR**, Yang P, Cassivi SD, Schild SE, Adjei AA. Non-small cell lung cancer: epidemiology, risk factors, treatment, and survivorship. *Mayo Clin Proc* 2008; **83**: 584-594 [PMID: 18452692 DOI: 10.4065/83.5.584]

13 **Navani N**, Nankivell M, Lawrence DR, Lock S, Makker H, Baldwin DR, Stephens RJ, Parmar MK, Spiro SG, Morris S, Janes SM; Lung-BOOST trial investigators. Lung cancer diagnosis and staging with endobronchial ultrasound-guided transbronchial needle aspiration compared with conventional approaches: an open-label, pragmatic, randomised controlled trial. *Lancet Respir Med* 2015; **3**: 282-289 [PMID: 25660225 DOI: 10.1016/S2213-2600(15)00029-6]

14 **Costache MI**, Iordache S, Karstensen JG, Săftoiu A, Vilmann P. Endoscopic ultrasound-guided fine needle aspiration: from the past to the future. *Endosc Ultrasound* 2013; **2**: 77-85 [PMID: 24949369 DOI: 10.4103/2303-9027.117691]

15 **Dziedzic D**, Peryt A, Szolkowska M, Langfort R, Orlowski T. Evaluation of the diagnostic utility of endobronchial ultrasound-guided transbronchial needle aspiration for metastatic mediastinal tumors. *Endosc Ultrasound* 2016; **5**: 173-177 [PMID: 27386474 DOI: 10.4103/2303-9027.183973]

16 **Labarca G**, Folch E, Jantz M, Mehta HJ, Majid A, Fernandez-Bussy S. Adequacy of Samples Obtained by Endobronchial Ultrasound with Transbronchial Needle Aspiration for Molecular Analysis in Patients with Non-Small Cell Lung Cancer. Systematic Review and Meta-Analysis. *Ann Am Thorac Soc* 2018; **15**: 1205-1216 [PMID: 30011388 DOI: 10.1513/AnnalsATS.201801-045OC]

17 **Folch E**, Yamaguchi N, VanderLaan PA, Kocher ON, Boucher DH, Goldstein MA, Huberman MS, Kent MS, Gangadharan SP, Costa DB, Majid A. Adequacy of lymph node transbronchial needle aspirates using convex probe endobronchial ultrasound for multiple tumor genotyping techniques in non-small-cell lung cancer. *J Thorac Oncol* 2013; **8**: 1438-1444 [PMID: 24128714 DOI: 10.1097/JTO.0b013e3182a471a9]

18 **Zhang Y**, Xie F, Mao X, Zheng X, Li Y, Zhu L, Sun J. Determining factors of endobronchial ultrasound-guided transbronchial needle aspiration specimens for lung cancer subtyping and molecular testing. *Endosc Ultrasound* 2019; **8**: 404-411 [PMID: 31670289 DOI: 10.4103/eus.eus\_8\_19]

19 **Bergethon K**, Shaw AT, Ou SH, Katayama R, Lovly CM, McDonald NT, Massion PP, Siwak-Tapp C, Gonzalez A, Fang R, Mark EJ, Batten JM, Chen H, Wilner KD, Kwak EL, Clark JW, Carbone DP, Ji H, Engelman JA, Mino-Kenudson M, Pao W, Iafrate AJ. ROS1 rearrangements define a unique molecular class of lung cancers. *J Clin Oncol* 2012; **30**: 863-870 [PMID: 22215748 DOI: 10.1200/JCO.2011.35.6345]

20 **Kohno T**, Nakaoku T, Tsuta K, Tsuchihara K, Matsumoto S, Yoh K, Goto K. Beyond ALK-RET, ROS1 and other oncogene fusions in lung cancer. *Transl Lung Cancer Res* 2015; **4**: 156-164 [PMID: 25870798 DOI: 10.3978/j.issn.2218-6751.2014.11.11]

21 **Shi Y**, Au JS, Thongprasert S, Srinivasan S, Tsai CM, Khoa MT, Heeroma K, Itoh Y, Cornelio G, Yang PC. A prospective, molecular epidemiology study of EGFR mutations in Asian patients with advanced non-small-cell lung cancer of adenocarcinoma histology (PIONEER). *J Thorac Oncol* 2014; **9**: 154-162 [PMID: 24419411 DOI: 10.1097/JTO.0000000000000033]

22 **Onozato R**, Kosaka T, Kuwano H, Sekido Y, Yatabe Y, Mitsudomi T. Activation of MET by gene amplification or by splice mutations deleting the juxtamembrane domain in primary resected lung cancers. *J Thorac Oncol* 2009; **4**: 5-11 [PMID: 19096300 DOI: 10.1097/JTO.0b013e3181913e0e]

23 **Cancer Genome Atlas Research Network.**. Comprehensive molecular profiling of lung adenocarcinoma. *Nature* 2014; **511**: 543-550 [PMID: 25079552 DOI: 10.1038/nature13385]

24 **Cooper WA**, Lam DC, O'Toole SA, Minna JD. Molecular biology of lung cancer. *J Thorac Dis* 2013; **5 Suppl 5**: S479-S490 [PMID: 24163741 DOI: 10.3978/j.issn.2072-1439.2013.08.03]

25 **Kang HJ**, Hwangbo B, Lee JS, Kim MS, Lee JM, Lee GK. Comparison of Epidermal Growth Factor Receptor Mutations between Metastatic Lymph Node Diagnosed by EBUS-TBNA and Primary Tumor in Non-Small Cell Lung Cancer. *PLoS One* 2016; **11**: e0163652 [PMID: 27685950 DOI: 10.1371/journal.pone.0163652]

26 **Ito M**, Miyata Y, Hirano S, Kimura S, Irisuna F, Ikeda K, Kushitani K, Kishi N, Tsutani Y, Takeshima Y, Okada M. Synchronicity of genetic variants between primary sites and metastatic lymph nodes, and prognostic impact in nodal metastatic lung adenocarcinoma. *J Cancer Res Clin Oncol* 2019; **145**: 2325-2333 [PMID: 31317326 DOI: 10.1007/s00432-019-02978-0]

27 **Kuwatani M**, Sakamoto N. Evolution and a promising role of EUS-FNA in gene and future analyses. *Endosc Ultrasound* 2020; **9**: 151-153 [PMID: 32584309]

**Footnotes**

**Institutional review board statement:** This study was reviewed and approved by the Ethics Committee of Tianjin Medical University Cancer Institute and Hospital (Approval No. bc2020023).

**Informed consent statement:** Patients were not required to provide informed consent for this study because the analysis used anonymous clinical data that were obtained after patient had agreed to treatment with written consent.

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**Data sharing statement:** No additional data are available.

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**Table 1** **Clinical characteristics of the 81 patients, *n* (%)**

|  |  |
| --- | --- |
| **Characteristics** | **Patients, *n* = 81** |
| Gender |  |
| Male | 59 (72.8) |
| Female | 22 (27.2) |
| Smoking status |  |
| Nonsmoker | 39 (48.1) |
| Former or current smoker | 42 (51.9) |
| Pathological type |  |
| Adenocarcinoma | 62 (76.5) |
| Squamous cell carcinoma | 11 (13.6) |
| Adenosquamous carcinoma | 3 (3.7) |
| NSCLC-NOS | 5 (6.2) |
| Clinical stage |  |
| IIIA | 15 (18.5) |
| IIIB | 28 (34.6) |
| IIIC | 3 (3.7) |
| IV | 35 (43.2) |
| Distant metastasis |  |
| Malignant pleural effusion | 3 (3.7) |
| Pleura | 4 (4.9) |
| Contralateral lung | 9 (11.1) |
| Bone | 14 (17.3) |
| Brain | 9 (11.1) |
| Liver | 3 (3.7) |
| Adrenal gland | 5 (6.2) |

NSCLC-NOS: Non-small cell lung carcinoma-not otherwise specified.

**Table 2 Puncture site and lesion size**

|  |  |  |  |
| --- | --- | --- | --- |
| **Location of sample** | **Patients, *n*** | **Minimum, mm** | **Maximum, mm** |
| Primary tumor |  |  |  |
| Right | 18 | 24 | 65 |
| Left | 7 | 28 | 91 |
| Lymph node |  |  |  |
| 2R | 2 | 10 | 21 |
| 4R | 19 | 9 | 34 |
| 4L | 4 | 27 | 28 |
| 5 | 2 | 19 | 38 |
| 7 | 35 | 15 | 70 |
| 8 | 2 | 12 | 20 |
| 10R | 5 | 24 | 26 |
| 10L | 3 | 23 | 45 |

2R: Right upper paratracheal; 4R: Right lower paratracheal; 4L: Left lower paratracheal; 5: Subaortic; 7: Subcarinal; 8: Paraesophageal; 10R: Right hilar; 10L: Left hilar.

**Table 3 Puncture of multiple lesions**

|  |  |
| --- | --- |
| **Location of sample** | **Patients, *n*** |
| 4R, 4L, 7 | 1 |
| 4R, 7 | 7 |
| 5, 7 | 1 |
| 7, 8 | 1 |
| Left lung mass, 7 | 1 |
| Right lung mass, 4R | 2 |
| Right lung mass, 7 | 2 |

4R: Right lower paratracheal; 4L: Left lower paratracheal; 5: Subaortic; 7: Subcarinal; 8: Paraesophageal.

**Table 4 Molecular analysis of all patients**

|  |  |  |  |  |
| --- | --- | --- | --- | --- |
|  | **Adenocarcinoma, *n* = 62** | **Squamous cell carcinoma, *n* = 11** | **Adenosquamous carcinoma, *n* = 3** | **NSCLC-NOS, *n* = 5** |
| ***EGFR* mutation** |  |  |  |  |
| Exon 19 |  |  |  |  |
| E746-A750del (1) | 2 | 0 | 0 | 0 |
| E746-A750del (2) | 4 | 0 | 0 | 0 |
| L747-T751del | 2 | 0 | 0 | 0 |
| L747-S752del | 1 | 0 | 0 | 0 |
| L747-A750del | 1 | 0 | 0 | 0 |
| Exon 21 |  |  | 0 |  |
| L858R | 8 | 0 | 1 | 0 |
| L861Q | 2 | 0 | 0 | 0 |
| ***KRAS* mutation** |  |  |  |  |
| Exon 2 |  |  |  |  |
| G12C | 3 | 1 | 0 | 0 |
| G12D | 2 | 0 | 0 | 0 |
| G12V | 2 | 0 | 0 | 0 |
| G12L | 1 | 0 | 0 | 0 |
| ***ROS-1* (FISH) positive** | 1 | 0 | 0 | 0 |
| **ALK (Ventana IHC) positive** | 5 | 0 | 0 | 0 |

NSCLC-NOS: Non-small cell lung carcinoma-not otherwise specified; ALK: Anaplastic lymphoma kinase; FISH: Fluorescence *in situ* hybridization; IHC: Immunohistochemistry.

**Table 5 Molecular analysis of patients with multiple lesions**

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
|  | **Type** | **Puncture site** | **Mutation** | **Puncture site** | **Mutation** |
| 1 | Adenocarcinoma | 5 | Insufficient sample | 7 | *EGFR*, exon 21 L858R |
| 2 | Adenocarcinoma | Right lung mass | *EGFR*, exon 21 L858R | 7 | *EGFR*, exon 21 L858R |
| 3 | Adenocarcinoma | 4R | *EGFR*, exon 21 L858R | 7 | *EGFR*, exon 21 L858R |
| 4 | Squamous cell carcinoma | 4R | *KRAS*, exon 2 G12C | 7 | *KRAS*, exon 2 G12C |

4R: Right lower paratracheal; 5: Subaortic; 7: Subcarinal.

**Table 6 Response to targeted therapy**

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
|  | **Drugs** | **CR, *n*** | **PR, *n*** | **SD, *n*** | **PD, *n*** |
| *EGFR* mutation | Gefitinib, erlotinib, extinib | 0 | 13 | 3 | 3 |
| *ROS-1* positive | Crizotinib | 0 | 0 | 1 | 0 |
| ALK positive | Crizotinib | 0 | 3 | 0 | 1 |

CR: Complete response; PR: Partial response; SD: Stable disease; PD: Progressive disease.