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**Primary myelofibrosis with concurrent *CALR* and *MPL* mutations: A case report**

Zhou FP *et al*. Co-mutation of *CALR* and *MPL* genes

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**Abstract**

BACKGROUND

Primary myelofibrosis (PMF) is a myeloproliferative neoplasm (MPN) characterized by recurrent mutations in the *JAK2*, *CALR*, and *MPL* genes. The *CALR* and *MPL* co-mutation is very rare. To our knowledge, no more than five cases have been reported. Here, we report a case of PMF in which a *CALR* and *MPL* co-mutation was detected by next-generation sequencing (NGS) technology, and a literature review was performed.

CASE SUMMARY

A 73-year-old woman was admitted to our hospital in 2018 due to abdominal distension. The patient had splenomegaly, lymphadenopathy, leukopenia, anemia, and immature granulocytes in peripheral blood. There were dacrocytes and atypical megakaryocytes in bone marrow, and megakaryocytic proliferation was very active, accompanied by reticulin fibrosis grade 2. By NGS analysis of the bone marrow sample, we detected mutations in *CALR*, *MPL,* and *PIK3RI*, while *JAK2* *V617F* and *BCR-ABL* were negative. Therefore, the patient was diagnosed with PMF and received oral ruxolitinib. However, the spleen and hematologic responses were poor. We review the literature, analyze previous reports of the mutation sites in our patient and differences between our patient and other reported cases of co-mutated *CALR* and *MPL* genes, and discuss the reason why the *CALR* and *MPL* co-mutations are rare and possible mechanisms and their impact on the prognosis of patients.

CONCLUSION

*CALR* and *MPL* mutations can be concurrent in MPN, but they are rare. The use of NGS may help to identify more patients with co-mutated *CALR* and *MPL* genes. This will help to further explore the mechanism and its impact on these patients to develop appropriate treatment strategies.

**Key Words:** Primary myelofibrosis; *CALR*; *MPL*; Co-mutation; Next-generation sequencing; Case report

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**Core Tip:** We report a rare case of primary myelofibrosis in which a *CALR* and *MPL* co-mutation was detected by next-generation sequencing technology. It demonstrated that *CALR* and *MPL* mutations can be concurrent in myeloproliferative neoplasm. We review the literature, analyze previous reports of the mutation sites in our patient and differences between our patient and other reported cases of co-mutated *CALR* and *MPL* genes, and discuss the reason why the *CALR* and *MPL* co-mutations are rare and possible mechanisms and their impact on the prognosis of patients.

**INTRODUCTION**

Primary myelofibrosis (PMF) is a myeloproliferative neoplasm (MPN) characterized by recurrent mutations in the *JAK2*, *CALR*, and *MPL* genes. Although these mutations were initially reported to be mutually exclusive in MPN, several studies have reported the presence of *JAK2-CALR* or *JAK2-MPL* co-mutations in PMF patients[1-4], and *CALR* and *MPL* co-mutation cases are still very rarely reported. With the increasingly wide application of next-generation sequencing (NGS) technology, rare myelofibrosis cases with co-mutated *CALR* and *MPL* genes are increasingly being detected. Here, we report a patient with PMF, in which co-mutated *CALR* and *MPL* genes were detected.

**CASE PRESENTATION**

***Chief complaints***

Abdominal distension for 2 mo.

***History of present illness***

A 73-year-old woman was admitted to Sir Run Run Shaw Hospital, Zhejiang University School of Medicine in 2018 due to abdominal distension for 2 mo, especially after eating with no complaints of abdominal pain, diarrhea, vomiting, or reduced anal exhaust and defecation.

***History of past illness***

The patient had a history of a left kidney cyst.

***Personal and family history***

She denied any other specific personal or family history of other diseases.

***Physical examination***

The patient’s conjunctiva and skin were pale, and the sclera was not yellow. Slightly swollen lymph nodes could be felt in the neck and armpit. The abdomen looked a little distended and felt soft, with no tenderness; the spleen under the ribs could be felt, and the texture was slightly hard. The rest of the abdomen did not have any obvious masses. The bowels sounded normal.

***Laboratory examinations***

Peripheral blood tests demonstrated a leukocyte count of 2.9 × 109/L, platelet count of 126 × 109/L, and hemoglobin of 87 g/L, and a smear revealed immature granulocytes of 4%. Liver function tests showed normal levels of transaminases, alkaline phosphatase, gamma-glutamyl transferase, total bilirubin, direct bilirubin, indirect bilirubin, serum creatinine, and potassium. Lactate dehydrogenase was 570 IU/L, and total cholesterol was 2.6 mmol/L.

***Imaging examinations***

An abdominal computed tomography scan showed that the spleen was enlarged and uniform (Figure 1), with a left kidney cyst. Abdominal ultrasound showed that the spleen was enlarged, with a thickness of 5.6 cm and a length of 16.1 cm.

***Pathological examination***

Bone marrow aspiration revealed erythroid and megakaryocytic hyperplasia; dacrocytes were easily observed, and megakaryocytes had atypia (Figure 2). Bone marrow biopsy showed megakaryocytic proliferation and atypia, accompanied by reticulin fibrosis grade 2 (Figure 3).

***Examination for gene mutations***

The bone marrow samples were sequenced by NGS, which showed mutations in *CALR*, *MPL,* and *PIK3RI* (Table 1), while *JAK2* *V617F* and *BCR-ABL* were negative.

**FINAL DIAGNOSIS**

The patient was ultimately diagnosed with PMF (International Prognostic Scoring System score = 3, high risk).

**TREATMENT**

The patient received oral ruxolitinib 15 mg twice per day.

**OUTCOME AND FOLLOW-UP**

Half a year later, the spleen response was poor, and the size of the spleen increased from 16.1 × 5.6 cm to 17.7 × 6.6 cm. Anemia was progressively aggravated in the meantime and developed to blood transfusion dependence, indicating that PMF with co-mutated *MPL* and *CALR* genes cannot benefit from ruxolitinib.

**DISCUSSION**

Most PMF patients carry *JAK2*, *MPL,* or *CALR* driver mutations[5]. Mutations in *MPL* and *CALR* are drivers in the pathogenesis of MPNs, and mutant *CALR* can bind with *MPL* to activate the JAK-STAT signaling pathway. Previous studies have shown that *JAK2*, *MPL,* and *CALR* are mutually exclusively mutated in MPN patients[6,7]. However, studies have also reported the presence of *JAK2-CALR* or *JAK2-MPL* co-mutations in PMF patients[1-4], and cases of *CALR* and *MPL* co-mutations are still very rarely reported. Here, we report the case of a PMF patient with *CALR*-p.364fsand *MPL*-p.X636W mutations. The *CALR*-p.364fsmutation is usually detected in essential thrombocytosis (ET) and PMF patients, while the *MPL*-p.X636Wmutation is not a common mutation and has only been reported in refractory anemia with ring sideroblast patients[8,9]. Furthermore, we also detected a low-burden *PIK3R1* mutation in this case. The *PIK3R1* gene belongs to the Akt signaling pathway and plays a direct role in regulating cell survival, growth, and differentiation. Mutated *PIK3R1* can cause aberrant PI3K signaling, and the p.R228fs mutation site has already been reported in acute lymphocytic leukemia patients[10,11]. To the best of our knowledge, this is the first report of a *CALR* and *MPL* double mutant PMF patient from East Asia.

Earlier research reports show that *CALR* and *MPL* mutations are mutually exclusive in MPN patients. This case demonstrated that *CALR* and *MPL* mutations can coexist in MPN patients. Three MPN cases with co-mutated *CALR* and *MPL* genes have already been reported since 2017[12-14]. Partouche *et al*[12] reported that a post-ET PMF patient carried type I *CALR* and *MPL* *W515R* mutations, with an variant allele frequency (VAF) of 59.1% and 31.7%, respectively. Tashkandi *et al*[13] reported a post-ET PMF patient carrying a *CALR* (p.L367Tfs\*46) mutation at an allele frequency of 47% and two *MPL* (p.S505C and p.W515L) mutations at a lower allele frequency of 4%. Bernal *et al*[14] detected a *CALR* (p.L367fs\*48) mutation at an allele frequency of 30%and a *MPL* (p.Trip515L) mutation at an allele frequency of 11.75% in an ET patient. In contrast to these reported cases, we identified an uncommon mutation in *MPL* (p.X636W) in PMF patients, and its VAF was higher than that of the *CALR* mutation in our patient.

Patients with concurrent *CALR* and *MPL* mutations are truly rare in the clinic. In PMF patients, the frequency of *CALR* (20%-25%) and *MPL* (7%) gene mutations is low[1,6,7]. Before 2017, there were no case reports about *CALR* and *MPL* co-mutations. Currently, a few MPN patients have been detected, which is inseparable from the progress of molecular testing technology. From previous cases, we can see that in MPN patients, a *MPL* mutation VAF is usually low[2,3,15], and some uncommon mutations exist. Traditional Sanger sequencing may miss these mutations because of its test range and sensitivity[7,13-15]. High-throughput sequencing may cover the full loci, and the detection VAF limit was lowered to 1%. Therefore, MPN patients with concurrent *CALR* and *MPL* mutations can be accurately detected.

The *MPL* mutation may have been a molecular event independent of the *CALR* deletion or a secondary event in the *CALR*-containing clone. Some researchers have proven that the *CALR* gene mutation is an early event during disease development, and *MPL* is a passenger event[2,13]. It is speculated that the mutated *CALR* gene can trigger genome instability, which then causes mutations in the *MPL* gene[13]. This is a possible explanation for why *CALR* and *MPL* can co-mutate. However, in our case, the *MPL* mutation was improbable as a secondary event because its VAF was more than 50%, indicating the possibility of a germline mutation. Some research has already proven that MPN patients can carry germline MPN mutations[16-18]. The co-mutation mechanism is not clear and still needs more investigations.

Gene mutations can predict prognosis in PMF patients. *CALR* mutations indicate a good prognosis in ET and PMF patients[19,20]. Compared to *JAK2* or triple-negative patients, *CALR* mutation-positive patients have a longer overall survival and low risk of thrombogenesis[1,7,19,21]. Patients who harbor the *MPL* *W515L/K* mutation have a moderate prognosis and higher risk of [thrombogenesis](file:///C:\Users\acorndx_1005\AppData\Local\youdao\dict\Application\7.5.0.0\resultui\dict\?keyword=thrombogenesis) than *CALR* mutation-positive patients[19]. The prognosis of patients with *MPL* and *CALR* co-mutations is still unknown due to the limited number of patients, but it seems that these patients have more aggressive progression based on our patient and other reported cases[12,13]. Our patient’s disease progression was rapid. In Partouche’s and Tashkandi’s patients, the *CALR* mutation was a pre-existing clone in the ET stage, and the *MPL* mutation was acquired when the disease developed into PMF. Disease acceleration may be associated with *MPL*-mutated clones. Because of the lack of patients with coexisting mutations, further investigations in large cohorts are needed to better quantify the presence and clinical significance.

**CONCLUSION**

In conclusion, we have described a rare case of PMF with concurrent *CALR* and *MPL* mutations. This demonstrates that *CALR* and *MPL* mutations are not mutually exclusive in MPN. The use of NGS may help to identify more patients with *CALR* and *MPL* co-mutations. This will help to further explore the mechanism and its impact on these patients to develop appropriate treatment strategies.

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**Footnotes**

**Informed consent statement:** The patient provided informed written consent during the treatment.

**Conflict-of-interest statement:** The authors declare that they have no conflicts of interest to disclose.

**CARE Checklist (2016) statement:** The authors have read the CARE Checklist (2016), and the manuscript was prepared and revised according to the CARE Checklist (2016).

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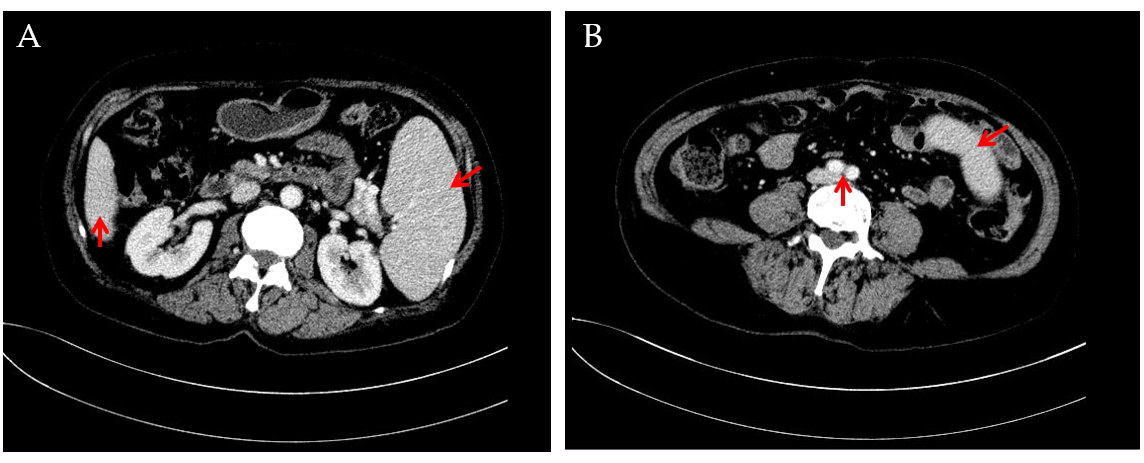
Grade C (Good): C

Grade D (Fair): D

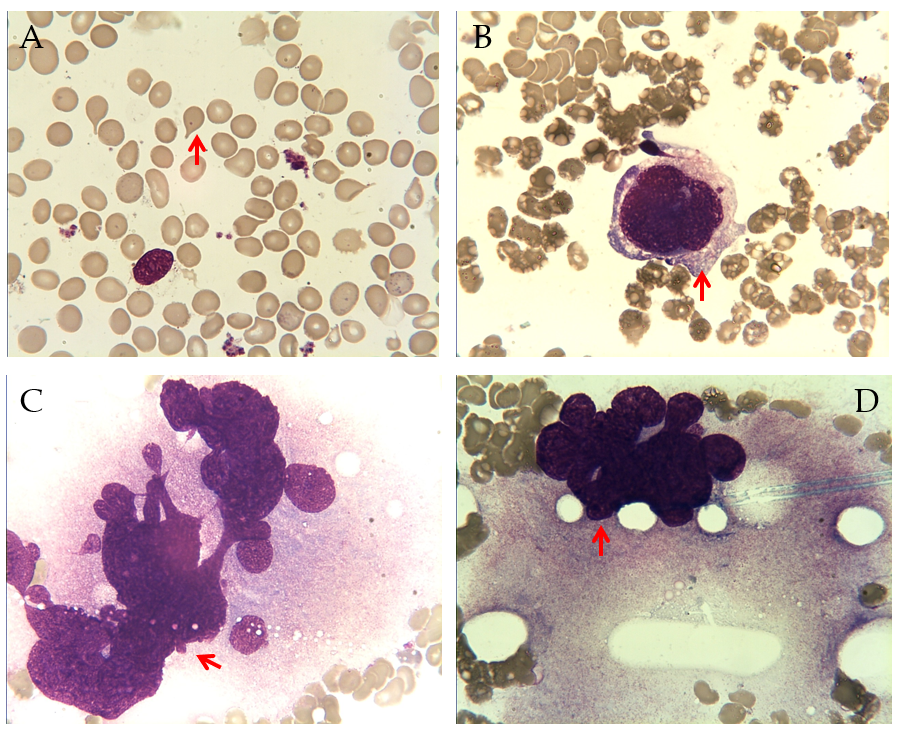
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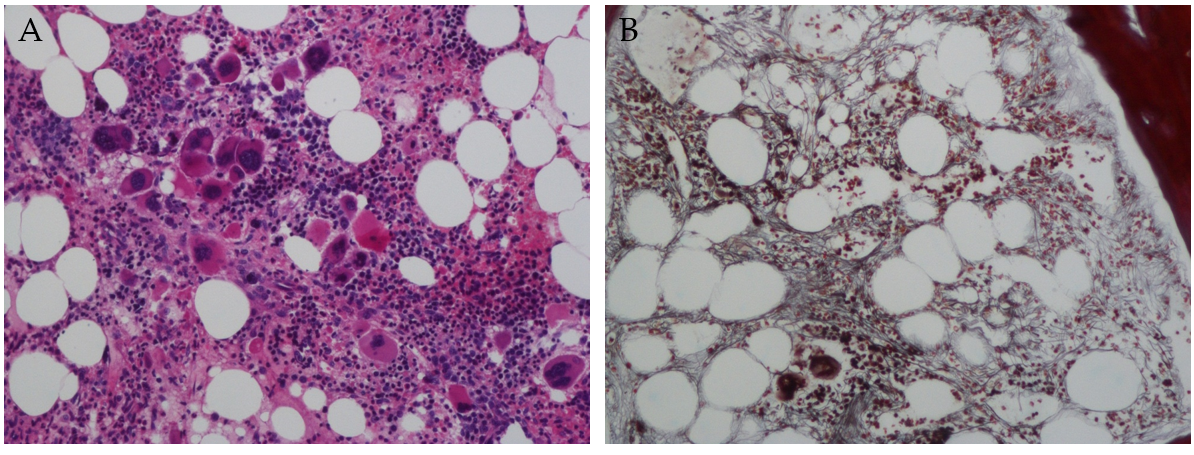
**Figure Legends**



**Figure 1 Computed tomography images showing splenomegaly.** A and B: Abdominal computed tomography images showing that the inferior margin of the spleen was lower than that of the liver (A) and reached the level of abdominal aortic bifurcation (B).



**Figure 2 Bone marrow aspiration findings.** A: Dacrocytes were easily observed; B-D: Images showing the presence of megakaryocyte atypia, including abnormal nuclear-cytoplasmic ratios, malformed nucleus (B), abnormal chromatin clumping with hyperchromatic nucleus (C), and extensive lobulation of the nucleus (D).



**Figure 3 Bone marrow biopsy findings.** A: Hematoxylin and eosin staining showed megakaryocytic proliferation and clustered distribution (magnification, 200 ×); B: Gomori staining showed diffuse and dense increase in reticulin fibres (magnification, 200 ×).

**Table 1 Gene mutation list in the** **primary myelofibrosis patient**

|  |  |  |  |
| --- | --- | --- | --- |
| **Gene** | **VAF** | **HGVS** | **Exonic function** |
| *CALR* | 30.04% | NM\_004343: c.1092\_1143del (p.364fs) | Frameshift deletion |
| *MPL* | 50.11% | NM\_005373: c.1908A>G (p.X636W) | Nonsynonymous SNV |
| *PIK3R1* | 3.18% | NM\_00181523: c.683delG (p.R228fs) | Frameshift deletion |

VAF: Variant allele frequency; HGVS: Human Genome Variation Society; SNV: Single nucleotide variant.