

# World Journal of *Gastroenterology*

*World J Gastroenterol* 2020 November 7; 26(41): 6304-6513



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**ABOUT COVER**

Editorial Board Member of *World Journal of Gastroenterology*, Dr. Ki Mun Kang is a Distinguished Professor at the Gyeongsang National University College of Medicine (Jinju, South Korea). Having received his Bachelor's degree from the College of Medicine of Chosun University in 1990, Dr. Kang undertook his postgraduate training, first at the Catholic College of Medicine, receiving his Master's degree in 1996, and then at Catholic University, receiving his PhD in 2004. He became Assistant Professor in Radiation Oncology at Gyeongsang National University in 2001 and has held the position since. His ongoing research interests include various aspects of immuno-radiobiology for GI and prostate cancers. Currently, he serves as President of the Department of Radiation Oncology, College of Medicine, Gyeongsang National University with Clinical Radiation Oncology and President of the Department of Radiation Oncology, Gyeongsang National University Changwon Hospital. (L-Editor: Filipodia)

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**RESPONSIBLE EDITORS FOR THIS ISSUE**

Production Editor: *Yu-Jie Ma*; Production Department Director: *Xiang Li*; Editorial Office Director: *Ze-Mao Gong*.

**NAME OF JOURNAL**

*World Journal of Gastroenterology*

**ISSN**

ISSN 1007-9327 (print) ISSN 2219-2840 (online)

**LAUNCH DATE**

October 1, 1995

**FREQUENCY**

Weekly

**EDITORS-IN-CHIEF**

Andrzej S Tarnawski, Subrata Ghosh

**EDITORIAL BOARD MEMBERS**

<http://www.wjgnet.com/1007-9327/editorialboard.htm>

**PUBLICATION DATE**

November 7, 2020

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**ARTICLE PROCESSING CHARGE**

<https://www.wjgnet.com/bpg/gerinfo/242>

**STEPS FOR SUBMITTING MANUSCRIPTS**

<https://www.wjgnet.com/bpg/GerInfo/239>

**ONLINE SUBMISSION**

<https://www.f6publishing.com>

## Basic Study

## P-selectin glycoprotein ligand 1 deficiency prevents development of acute pancreatitis by attenuating leukocyte infiltration

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**Supported by** National Natural Science Foundation of China, No. 81670387, No. 31671440, and No. 81800402.

**Institutional review board statement:** This study was reviewed and approved by Ethics Committee of People's Hospital of Zhengzhou University.

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## Abstract

### BACKGROUND

Acute pancreatitis (AP) is rapid-onset pancreatic inflammation that causes local and systemic inflammatory response syndrome (SIRS) with high morbidity and mortality, but no approved therapies are currently available. P-selectin glycoprotein ligand 1 (PSGL-1) is a transmembrane glycoprotein to initiate inflammatory responses. We hypothesized that PSGL-1 may be involved in the development of AP and would be a new target for the treatment of AP.

### AIM

To investigate the role and mechanism of PSGL-1 in the development of AP.

### METHODS

The PSGL-1 expression on leukocytes was detected in peripheral blood of AP patients and volunteers. Pancreatic injury, inflammatory cytokines expression, and inflammatory cell infiltration was measured in AP mouse models induced with PSGL-1 knockout (*PSGL-1<sup>-/-</sup>*) and wild-type (*PSGL-1<sup>+/+</sup>*) mice. Leukocyte-endothelial cell adhesion was measured in a peripheral blood mononuclear cell (PBMC)-endothelial cell coculture system.

### RESULTS

**Institutional animal care and use**

**committee statement:** The animal care and use were approved by the Institute of Laboratory Animal Science, Chinese Academy of Medical Sciences and Peking Union Medical College.

**Conflict-of-interest statement:** We declare that we have no financial and personal relationships with other people or organizations that can inappropriately influence our work, and there is no professional or other personal interest of any nature or kind in any products, service and/or company that could be construed as influencing the position presented in, or the review of, the manuscript.

**Data sharing statement:** No additional data are available.

**ARRIVE guidelines statement:** The authors have read the ARRIVE guidelines, and the manuscript was prepared and revised according to the ARRIVE guidelines.

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**Manuscript source:** Unsolicited manuscript

**Specialty type:** Research and experimental medicine

**Country/Territory of origin:** China

**Peer-review report's scientific quality classification**

Grade A (Excellent): 0  
Grade B (Very good): 0  
Grade C (Good): C

The expression of PSGL-1 on monocytes and neutrophils was significantly increased in AP patients. Compared with *PSGL-1<sup>+/+</sup>* mice, *PSGL-1<sup>-/-</sup>* AP mice induced by caerulein exhibited lower serum amylase, less Interleukin-1beta (IL-1beta) and Interleukin-6 (IL-6) expression, less neutrophil and macrophage infiltration, and reduced peripheral neutrophil and monocyte accounts. PSGL-1 deficiency alleviated leukocyte-endothelial cell adhesion *via* IL-6 but not IL-1beta.

**CONCLUSION**

PSGL-1 deficiency effectively inhibits the development of AP by preventing leukocyte-endothelial cell adhesion *via* IL-6 stimulation and may become a potential therapeutic target for treating AP.

**Key Words:** P-selectin glycoprotein ligand 1; Acute pancreatitis; Inflammation; Leukocyte adhesion; Interleukin-6

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**Core Tip:** The severity of acute pancreatitis (AP) is inversely related to the number of leukocytes that infiltrate pancreatic tissue. P-selectin glycoprotein ligand 1 (PSGL-1) plays an important role in the development of AP by promoting inflammatory cell infiltration initiated by leukocyte and endothelial cell adhesion. PSGL-1 deficiency may protect against the development of AP and may be a new potential target for AP therapy.

**Citation:** Zhang X, Zhu M, Jiang XL, Liu X, Liu P, Wu XX, Yang ZW, Qin T. P-selectin glycoprotein ligand 1 deficiency prevents development of acute pancreatitis by attenuating leukocyte infiltration. *World J Gastroenterol* 2020; 26(41): 6361-6377

**URL:** <https://www.wjnet.com/1007-9327/full/v26/i41/6361.htm>

**DOI:** <https://dx.doi.org/10.3748/wjg.v26.i41.6361>

**INTRODUCTION**

Acute pancreatitis (AP) is rapid-onset pancreatic inflammation that causes local and systemic inflammatory response syndrome (SIRS) with high morbidity and mortality<sup>[1]</sup>. Although most patients with AP experience a mild and self-resolving disease course, 15%–20% of patients develop severe AP that leads to multiple organ dysfunction syndrome and poor prognosis<sup>[2]</sup>. Uncontrolled systemic inflammation in AP results in a high risk of morbidity and mortality, but no approved therapies are currently available<sup>[3]</sup>. Immune cells infiltration such as monocytes and neutrophils is the first step of inflammation and results in pancreatic injury<sup>[4]</sup>. Therefore, blocking immune cell infiltration may be a potential and promising target for AP treatment.

P-selectin glycoprotein ligand 1 (PSGL-1), a transmembrane glycoprotein, is mainly expressed on leukocytes. PSGL-1 regulates leukocyte activation, recruitment, and infiltration *via* binding to E-selectin and P-selectin, which play an important role in initiating inflammatory responses<sup>[5]</sup>. Knockdown of PSGL-1 effectively reduces the numbers of circulating neutrophils and monocytes, improves endothelial damage, and decreases blood pressure<sup>[6]</sup>. Blocking functional PSGL-1 reduces visceral adipose inflammation and ameliorates endothelial dysfunction in atherosclerosis<sup>[7]</sup>. The contribution of PSGL-1 to the inflammatory response has been demonstrated in various inflammation-related diseases<sup>[8]</sup>. However, the role of PSGL-1 in the development of AP has not been tested.

This study tested the hypothesis that PSGL-1 plays a significant role in the development of AP by a clinical experiment and animal experiments using caerulein induced AP models in *PSGL-1<sup>-/-</sup>* and *PSGL-1<sup>+/+</sup>* mice, which would be a new therapeutic target for the treatment of AP.

Grade D (Fair): 0

Grade E (Poor): 0

**Received:** April 14, 2020**Peer-review started:** April 14, 2020**First decision:** April 26, 2020**Revised:** July 13, 2020**Accepted:** September 10, 2020**Article in press:** September 10, 2020**Published online:** November 7, 2020**P-Reviewer:** Takasawa S**S-Editor:** Liu M**L-Editor:** Wang TQ**P-Editor:** Ma YJ

## MATERIALS AND METHODS

### Materials

The mouse brain endothelial cell line (Bend.3) was purchased from Feng Hui Biological Technology Company Co., Ltd. (Changsha, Hunan Province, China). *PSGL-1<sup>+/+</sup>* and *PSGL-1<sup>-/-</sup>* mice were a generous gift from Dr. Xia at the Department of Molecular Biology and Biochemistry, University of Oklahoma Health Science Center. Peripheral blood mononuclear cell (PBMC) extraction kits were purchased from Millipore Sigma Co., Ltd. (St. Louis, MO, United States). Rabbit anti-mouse Interleukin-6 (IL6) and Interleukin-1beta (IL-1beta), as well as the IL-6 inhibitor galiellalactone were purchased from Santa Cruz Biotechnology Co., Ltd (San Francisco, CA, United States). The IL-1beta inhibitor AS101 was purchased from Selleck Chemicals Co., Ltd. (Huston, TX, United States). PerCP/Cyanine5.5-conjugated anti-human CD162 antibody, APC-conjugated anti-human CD14 antibody, FITC-conjugated anti-human CD33 antibody, PE-conjugated anti-human CD45 antibody, PerCP/Cyanine5.5-conjugated anti-mouse/human CD11b antibody, APC/Cyanine7-conjugated anti-mouse CD45 antibody, PE/Cyanine7-conjugated anti-mouse Ly-6G antibody, and FITC-conjugated anti-mouse Ly-6C antibody were purchased from BioLegend Co., Ltd. (San Diego, CA, United States). A PE-conjugated rat anti-Mouse CD162 was purchased from BD Biosciences Co., Ltd. (San Jose, CA, United States). IL-1beta enzyme-linked immunosorbent assay (ELISA) kit and IL-6 ELISA kit were purchased from Shanghai Xin Fan Biological Technology Co., Ltd. (Shanghai, China). TUNEL test kits were purchased from Beyotime Biotechnology Co., Ltd. (Shanghai, China). Caerulein and lipopolysaccharide (LPS) were purchased from Millipore Sigma Co., Ltd. (St. Louis, MO, United States).

### Clinical experiments

Twenty-one patients with AP who were admitted from January 2018 to January 2019 were recruited. The diagnostic criteria for AP were: (1) Abdominal pain consistent with AP (acute, sudden, persistent, severe upper abdominal pain, often radiating to the back); (2) Serum amylase and/or lipase activity at least three times higher than the normal limit; and (3) Enhanced computed tomography/magnetic resonance imaging (CT/MRI) or abdominal ultrasound showing AP-related imaging changes<sup>[9]</sup>. Venous blood was collected from patients on an empty stomach in the early morning of the day after admission. Eight healthy volunteers matched by age and sex were recruited as controls. All patients and volunteers signed an informed consent form, which was approved by the hospital's ethics committee (Approval number: QT19012; Approval date: November 15, 2017).

### Animal experiments

AP was induced in mice by intraperitoneal injection of 0.2 mg/kg caerulein (Millipore Sigma, United States) every hour for 7 h, followed by one injection of 10 mg/kg LPS (Millipore Sigma, United States). Twenty-four hours later, the animals were sacrificed by inhalation of isoflurane (1.5%-2.5%, inhaled). The animals were 6 or 8 wk old at the time of the experiments. Groups ( $n = 4$ ) of age-matched wild-type and/or *PSGL-1<sup>-/-</sup>* mice were used. All experimental protocols were approved by the Animal Care and Use Committee at the Institute of Laboratory Animal Science, Chinese Academy of Medical Sciences and Peking Union Medical College (Approval number: YZW19007; Approval date: December 13, 2019).

### ELISA

The levels of the inflammatory cytokines IL-1beta and IL-6 in the serum or supernatant were tested with an IL-1beta ELISA kit (Shanghai Xin Fan Biological Technology, China) and an IL-6 ELISA kit (Shanghai Xin Fan Biological Technology, China), respectively. Briefly, serum and supernatant samples were added to 96-well plate wells that were coated with IL-1beta or IL-6 mAb, incubated for 2 h at 37 °C, washed with PBS four times for 3 min each time, and then incubated with an horseradish peroxidase (HRP) linked streptavidin solution for 30 min at 37 °C in a darkroom. All samples were tested three times, and the absorbance was measured at 450 nm with a microplate reader.

### Western blot analysis

Total protein was extracted using a Total Protein Extraction kit (Beyotime Biotechnology), and protein concentrations were determined using a Bicinchoninic Acid (BCA) Protein Assay kit (Beyotime Biotechnology), and the amount of protein

was calculated. Twenty micrograms of protein was separated by 12% sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE) at 80 V for 120 min and transferred to a nitrocellulose (NC) membrane, and then the membrane was blocked with 5% skim milk powder at room temperature for 2 h and incubated with a mouse monoclonal IL-6 antibody (Santa Cruz Biotechnology) diluted 1:1000 and an IL-1 $\beta$  antibody (Santa Cruz Biotechnology) diluted 1:1000, at 4 °C overnight. The next day, the membrane was incubated with an HRP-conjugated anti-rabbit antibody (Santa Cruz Biotechnology, United States) diluted 1:5000 at room temperature for 1 h, and the signals were visualized with an electrochemiluminescence kit (Millipore Sigma).

### **Flow cytometry**

Blood samples were lysed with 1 × BD FACS lysing solution (BD Biosciences, United States) and stained with fluorescently labeled antibodies for 30 min in the dark on ice to identify different leukocyte cell populations. Labeled and fixed samples were analyzed by flow cytometry on a FACSCanto III system (BD Biosciences) immediately or within 24 h. Before each run, BD cytometer setup and tracking beads (BD Biosciences) were used for internal calibration. Appropriate controls were prepared for each sample to allow for compensation and detection of nonspecific binding. Cellular fluorescence was quantified as the mean fluorescence intensity or percentage of double-positive cells at each time point. All results were analyzed by using BD FACS Diva software.

### **Immunohistochemistry and terminal deoxynucleotidyl transferase-mediated dUTP-biotin nick end labeling (TUNEL) assay**

Paraffin-embedded tissues were sectioned, dewaxed, immersed in graded ethanol and 3% hydrogen peroxide solution (twice), washed three times with PBS solution, and subjected to high-temperature and high-pressure antigen retrieval in a weak acid solution. Blocking was performed with 10% goat serum to block nonspecific reactions. The samples were incubated with biotin-labeled solution at 37 °C for 60 min in the dark. Then, the sections were incubated with an HRP-labeled secondary antibody, stained with 3,3'-diaminobenzidine staining solution, counterstained with hematoxylin, dehydrated in ethanol dehydration, mounted with gum, and observed under a microscope. To quantify apoptosis in the tissues, a TUNEL assay was performed according to the instructions of the TUNEL kit (Beyotime Biotechnology).

### **Cell culture and adhesion assay**

The pancreatic acinar cell line AR42J was purchased from American Type Culture Collection and cultivated in Dulbecco's modified Eagle's medium (DMEM) with 10% fetal bovine serum (FBS), 100 U/mL penicillin, and 100  $\mu$ g/mL streptomycin at 37 °C and 5% CO<sub>2</sub>. A cell model of AP was established by incubating AR42J cells with 100 nmol/L of caerulein for 24 h after the cells reached 80% confluence. The supernatants were collected and frozen at -80 °C. Bend.3 cells were cultured with DMEM containing 10% FBS, 100  $\mu$ g/mL penicillin, and 10  $\mu$ g/mL streptomycin in 96-well plates. Peripheral blood mononuclear cells (PBMCs) were isolated from the peripheral blood of *PSGL-1*<sup>+/+</sup> and *PSGL-1*<sup>-/-</sup> mice as previously described<sup>[10]</sup>, stained with Green 5-chloromethylformacein diacetate (CMFDA) (Abcam, United Kingdom) for 30 min at 37 °C, and resuspended with normal medium or conditioned medium containing the supernatant from the cell model. Afterwards, the treated PBMCs were incubated with cultured Bend.3 cells grown in monolayers in 96-well plates for 1 h at 37 °C. The IL-1 $\beta$  inhibitor AS101 (Selleck Chemicals, United States) and IL-6 inhibitor galiellalactone (Santa Cruz Biotechnology, United States) were added simultaneously according to the experimental grouping. After the nonadherent PBMCs were removed by washing with PBS, the fluorescence signals were photographed using an inverted fluorescence microscope.

### **Statistical analysis**

The data are expressed as the mean  $\pm$  SD of the mean. Student's *t*-test was used to determine significant differences between the control and experimental groups. *P* < 0.05 was considered statistically significant.

## RESULTS

### **Increased expression of PSGL-1 on monocytes and neutrophils from patients with AP**

We found that there was a significant increase in the number of circulating monocytes and neutrophils in AP patients compared with volunteers (Figure 1A–D). The expression of PSGL-1 on monocytes and neutrophils in AP patients was significantly elevated than that in volunteers (Figure 1E and F), which suggests that PSGL-1 plays an important role in the development of AP.

### **PSGL-1 deficiency alleviates caerulein-mediated inflammatory response and acinar damage**

To further examine the role of PSGL-1 in AP, an AP mouse model was induced by caerulein in *PSGL-1*<sup>-/-</sup> mice and *PSGL-1*<sup>+/+</sup> mice (Figure 2A). Hematoxylin-eosin staining showed that tissue edema and leukocyte infiltration into the pancreas in *PSGL-1*<sup>+/+</sup> but not in *PSGL-1*<sup>-/-</sup> mice (Figure 2B). The level of amylase in *PSGL-1*<sup>+/+</sup> AP mice was significantly higher than that in the control group, while there was no significant increase in amylase in the *PSGL-1*<sup>-/-</sup> AP mice (Figure 2C). The pancreas damage rate was approximately three-fold higher in *PSGL-1*<sup>+/+</sup> mice than in *PSGL-1*<sup>-/-</sup> mice (Figure 2D). The levels of the proinflammatory cytokines IL-1beta and IL-6 were not increased in the pancreatic parenchyma and serum of *PSGL-1*<sup>-/-</sup> AP mice compared with those in *PSGL-1*<sup>+/+</sup> AP mice (Figure 2E–H). Caerulein-induced apoptosis of acinar cells quantified by TUNEL assay was blocked, at least partially, by *PSGL-1* gene knockout (Figure 2I and J).

### **PSGL-1 deficiency attenuates caerulein-mediated leukocyte infiltration in the pancreas**

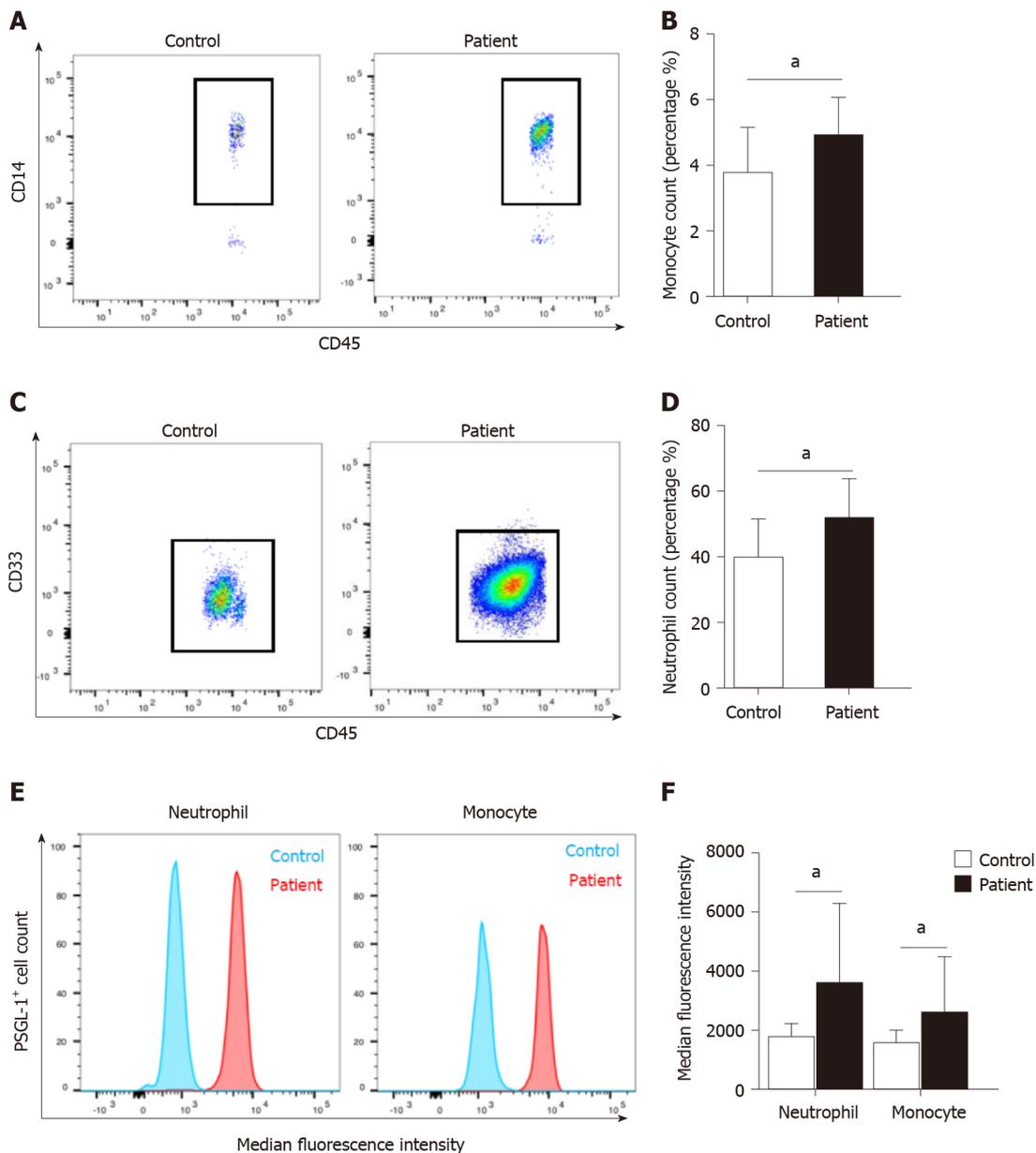
Leukocyte infiltration, especially neutrophils and macrophages, is an important process leading to the occurrence and development of AP. We detected the infiltration of leukocytes in pancreatic tissue from *PSGL-1*<sup>+/+</sup> mice and *PSGL-1*<sup>-/-</sup> mice by immunohistochemistry. We found more pancreatic infiltration of myeloperoxidase-positive neutrophils and F4/80-positive monocytes/macrophages in *PSGL-1*<sup>+/+</sup> AP mice than in *PSGL-1*<sup>-/-</sup> AP mice (Figure 3A–D). However, infiltration of CD3-positive T cells and CD45R-positive B cells was not different (Figure 3E–H) between the two groups of mice. These results indicate that infiltration of neutrophils and macrophages, but not T cells or B cells, is the main pathological change in AP, which is consistent with the results reported in the literature<sup>[11]</sup>. Thus, PSGL-1 deficiency attenuates infiltration of neutrophils and macrophages in the pancreas.

### **PSGL-1 deficiency attenuates the number of circulating leukocytes**

AP occurs early in damaged pancreatic acinar cells, resulting in a local inflammatory response<sup>[12]</sup>. We investigated the role of PSGL-1 in systemic inflammatory response caused by AP, and found that the number of neutrophils (Ly-6G; Figure 4A and C) and monocytes (Ly-6C; Figure 4B and D) in the peripheral blood was significantly increased in *PSGL-1*<sup>+/+</sup> AP mice, with higher expression of PSGL-1 on neutrophils and monocytes cells (Figure 4E and F), compared with *PSGL-1*<sup>-/-</sup> AP mice. The results are consistent with our clinical findings, and indicate that PSGL-1 deficiency significantly prevents the systemic inflammatory response caused by AP and blocks the development of AP.

### **PSGL-1 deficiency alleviates caerulein-induced leukocyte-endothelial cell adhesion**

It has been reported that IL-1beta and IL-6 are important proinflammatory factors in AP<sup>[13,14]</sup>. Therefore, we detected the levels of IL-6 and IL-1beta in the supernatants of AR42J cells treated with caerulein. We found that the change in IL-6 was more obvious than that in IL-1beta (Figure 5A). The result shows that in the early stage of AP, especially within 24 h of onset, IL-6 may play the main promotion role than IL-1beta. We speculated that IL-6 recruits leukocytes and induces leukocyte adhesion and infiltration. We then detected whether PSGL-1 deficiency affects leukocyte-endothelial cell adhesion in an *in vitro* AP model. As shown in Figure 5B–C the number of PBMCs (green) from wild-type mice that adhered to Bend.3 cells was significantly increased when the cells were cultured with the supernatant from the AP model. Compared with the IL-1beta inhibitor, the IL-6 inhibitor significantly decreased the number of adherent cells when the cells were cultured with the supernatant from the AP model. Furthermore, the number of PBMCs (green) from *PSGL-1*<sup>-/-</sup> mice that adhered to



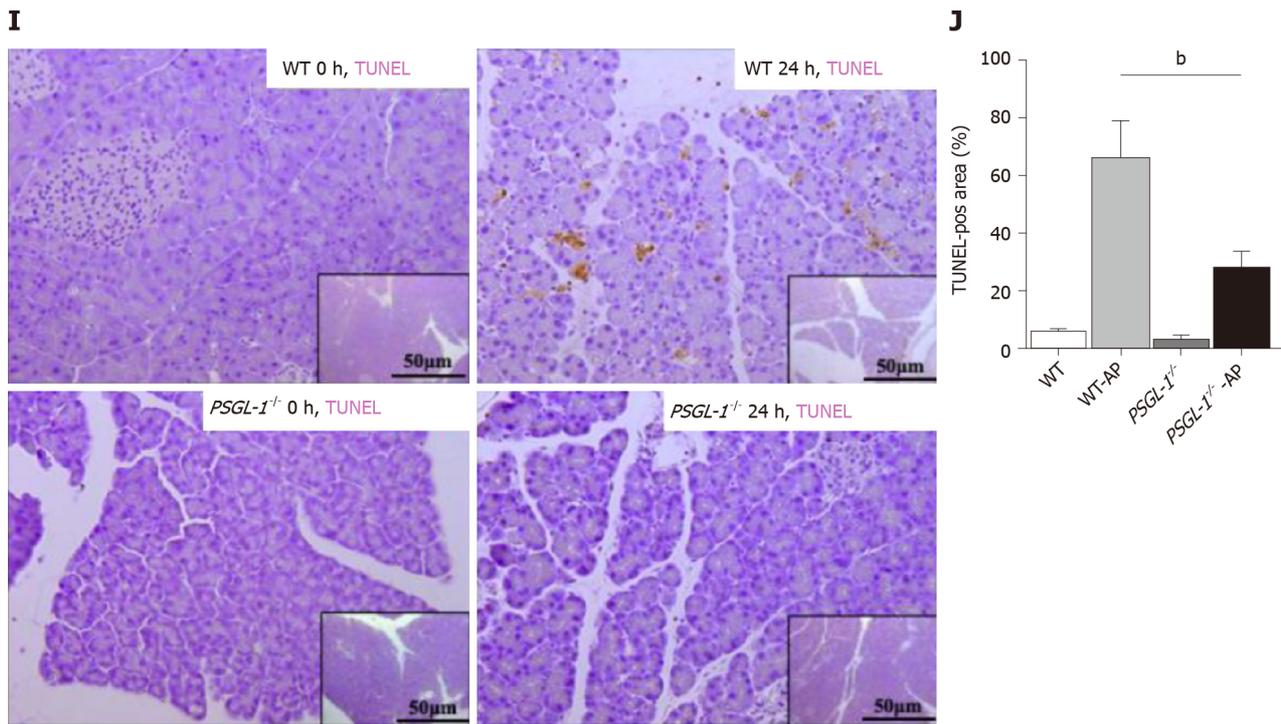
**Figure 1** Increased expression of P-selectin glycoprotein ligand 1 on neutrophils and monocytes from patients with acute pancreatitis. A: Flow cytometry for CD14-positive cells to detect monocytes in the peripheral blood of patients ( $n = 21$ ) with acute pancreatitis (AP) or normal control subjects ( $n = 8$ ); B: Boxplot showing quantification of monocytes; C: Flow cytometry for CD33-positive cells to detect neutrophils in the peripheral blood of patients ( $n = 21$ ) with AP or normal control subjects ( $n = 8$ ); D: Boxplot showing quantification of neutrophils; E: Fluorescence intensity of P-selectin glycoprotein ligand 1 (PSGL-1) on neutrophils and monocytes; F: Boxplot showing quantification of fluorescence intensity of PSGL-1 on neutrophils and monocytes. <sup>a</sup> $P < 0.05$ , Student's *t*-test. PSGL-1: P-selectin glycoprotein ligand 1.

Bend.3 cells obviously decreased when the cells were cultured with the supernatant from the AP model. These results indicate that PSGL-1 plays an important role in caerulein-induced AP by regulating leukocyte-endothelial cell adhesion, which is triggered by IL-6 in the early stage of AP.

## DISCUSSION

Severe AP is an acute abdominal disease with a strong systemic inflammatory response. Immune cell infiltration, along with chemokine and cytokine cascade-mediated inflammation, plays a critical role in the development of this disease<sup>[15,16]</sup>. Although increasing evidence shows that inhibiting the immune cells (neutrophils) or specific adhesion molecules (P-selectin, lymphocyte function antigen-1, and CD11a/CD18) may have a therapeutic effect on excessive inflammatory responses in AP<sup>[17-20]</sup>, no drug that effectively modulates the outcome of AP has been identified<sup>[21]</sup>. It





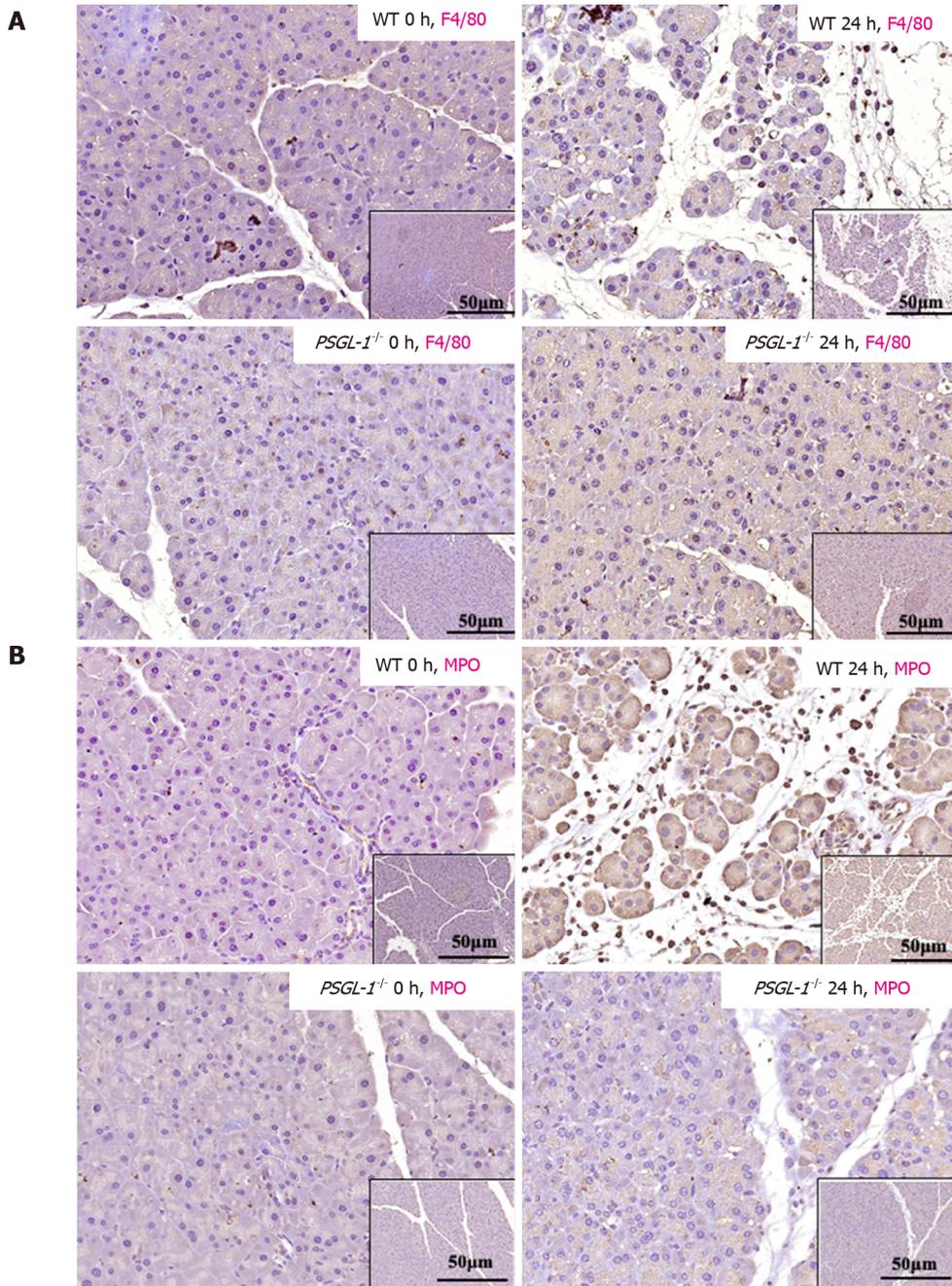
**Figure 2 P-selectin glycoprotein ligand 1 deficiency alleviates caerulein-mediated inflammatory response and acinar damage.** A: Schematic diagram of the induction of acute pancreatitis (AP) showing the frequency of caerulein and lipopolysaccharide injections as well as the sampling time points; B: Hematoxylin-eosin staining of the pancreas of wild-type (upper panels) and P-selectin glycoprotein ligand 1 (*PSGL-1*<sup>-/-</sup>) (lower panels) mice 0 h and 24 h after treatment with caerulein; C: Boxplots showing the expression of amylase ( $n = 4$ ); D: Boxplots showing quantification of acinar cell damage ( $n = 4$ ); E: Expression of IL-6 in the pancreas of wild-type and *PSGL-1*<sup>-/-</sup> mice; F: Expression of IL-1 $\beta$  in the pancreas of wild-type and *PSGL-1*<sup>-/-</sup> mice; G: Expression of IL-6 in sera of wild-type and *PSGL-1*<sup>-/-</sup> mice ( $n = 4$ ); H: Expression of IL-1 $\beta$  in sera of wild-type and *PSGL-1*<sup>-/-</sup> mice ( $n = 4$ ); I: Transferase-mediated dUTP-biotin nick end labeling (TUNEL) assay for apoptosis in the pancreas; J: Boxplots showing quantification of the TUNEL assays ( $n = 4$ ). <sup>a</sup> $P < 0.05$ ; <sup>b</sup> $P < 0.001$ , Student's *t*-test. H&E: Hematoxylin-eosin; TUNEL: Transferase-mediated dUTP-biotin nick end labeling; PSGL-1: P-selectin glycoprotein ligand 1; AP: Acute pancreatitis; WT: Wild type.

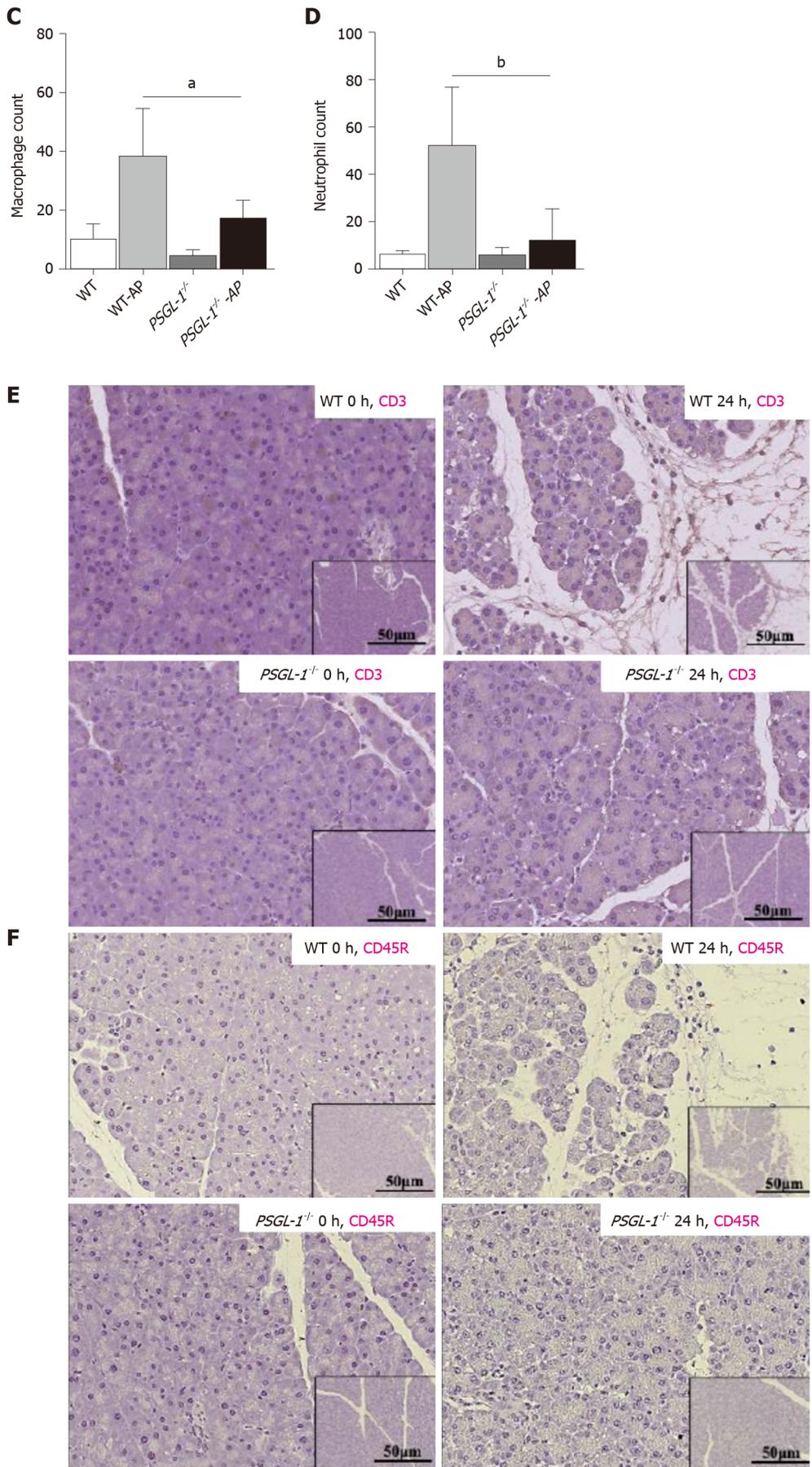
is urgent to find a novel therapeutic target for the treatment of AP.

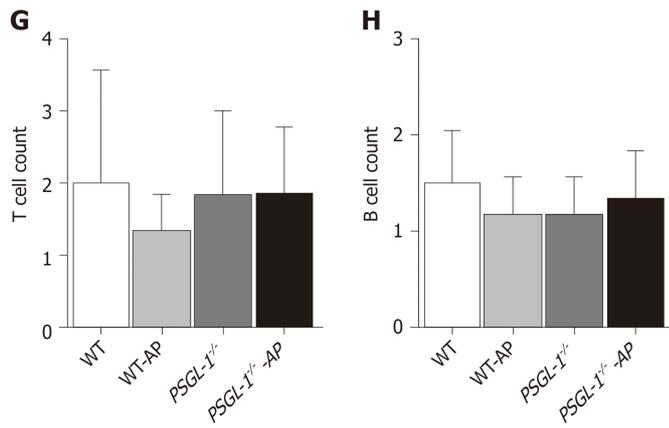
PSGL-1, the main ligand of E-selectin and P-selectin, initiates the capture and rolling steps in leukocyte-endothelial cell adhesion and extravasation into tissues, engaging multiple inflammatory signaling pathways<sup>[22-24]</sup>. Cross-linking between PSGL-1 with P-selectin regulates the rolling and tethering of neutrophils and macrophages/monocytes, plasma B cells, dendritic cells, and T cells during extravasation<sup>[25]</sup>. Evidence shows that patients with pancreatitis have more P-/E-selectin in their serum than healthy individuals, and that these factors are involved in leukocyte adhesion and inflammation<sup>[26]</sup>. However, the role of their ligand, PSGL-1, in the development of AP remains poorly understood. Herein, we explored the role of PSGL-1 in AP and found that PSGL-1 was significantly upregulated on circulating monocytes and neutrophils in patients with AP, which indicated the close relationship between PSGL-1 and AP development.

To better understand the function of PSGL-1, we used *PSGL-1*<sup>-/-</sup> mice to explore the role of PSGL-1 in the progression of AP. We found that caerulein treatment caused amylase upregulation and acinar damage in *PSGL-1*<sup>+/+</sup> mice, but not in *PSGL-1*<sup>-/-</sup> mice. Since acinar cell death is central to AP<sup>[27]</sup>, we also detected whether PSGL-1 deficiency affects acinar cell death by TUNEL assay. We found that PSGL-1 deficiency significantly decreased acinar cell death, indicating the protective effect of PSGL-1 deficiency against acinar cell damage. PSGL-1, as an initiating factor of the leukocyte-endothelial cell adhesion cascade, has been well described to mediate the inflammatory response in immune cells<sup>[28]</sup>. In this study, we found that PSGL-1 deficiency not only attenuated local inflammation, but also attenuated the systemic inflammation. In addition, fewer PBMCs from *PSGL-1*<sup>-/-</sup> mice adhered to endothelial cells than PBMCs from wild-type mice *in vitro*, which was in consistent with a previous study<sup>[10]</sup> and further demonstrated the protective effect of PSGL-1 deficiency on AP development by regulating leukocyte-endothelial adhesion, which might become a promising therapeutic target.

In the initial stage of AP, pancreatic acinar cells are damaged mainly due to abnormal activation of enzymes, and circulating neutrophil and monocyte







**Figure 3 P-selectin glycoprotein ligand 1 deficiency attenuates caerulein-mediated leukocyte infiltration in the pancreas.** A: Immunohistochemistry of pancreatic tissue sections for F4/80 (monocytes/macrophages); B: Immunohistochemistry of pancreatic tissue sections for myeloperoxidase (MPO) (neutrophils); C: Boxplots showing quantification of F4/80-positive monocytes/macrophages in the pancreas ( $n = 4$ ); D: Boxplots showing quantification of MPO-positive neutrophils in the pancreas ( $n = 4$ ); E: Immunohistochemistry of pancreatic tissue sections stained for CD3 (T cells); F: Immunohistochemistry of pancreatic tissue sections stained for CD45R (B cells); G: Boxplots showing quantification of CD3-positive T cells in the pancreas ( $n = 4$ ); H: Boxplots showing quantification of CD45R-positive B cells in the pancreas ( $n = 4$ ); <sup>a</sup> $P < 0.001$ , <sup>b</sup> $P < 0.01$ , Student's *t*-test. PSGL-1: P-selectin glycoprotein ligand 1; MPO: Myeloperoxidase; WT: Wild type; AP: Acute pancreatitis.

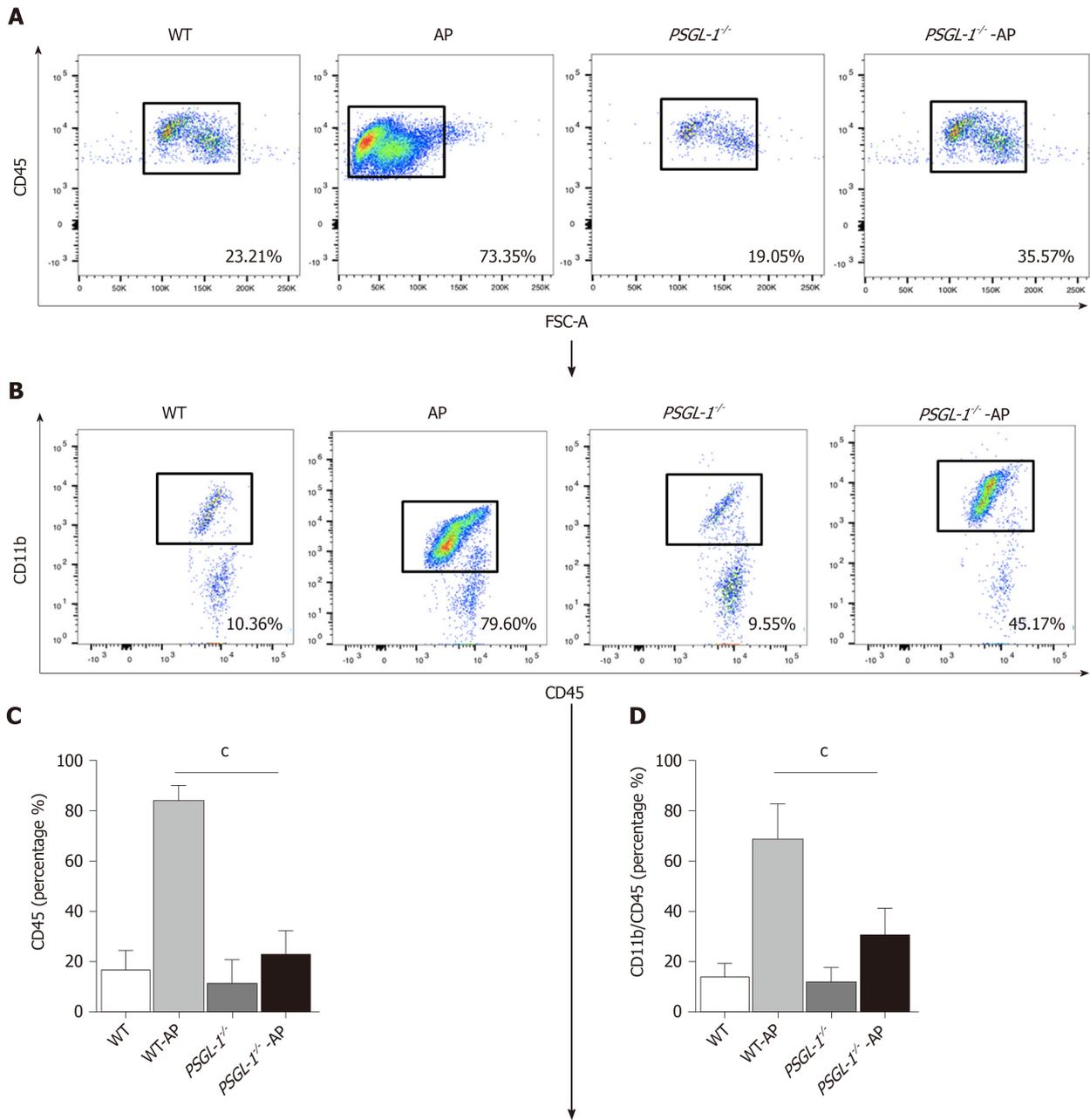
recruitment<sup>[29]</sup> by cytokines and inflammation factors<sup>[30]</sup>, including IL-6 and IL-1beta, thus leading to a vicious cycle<sup>[31]</sup>. In our mechanism study, the expression of IL-6 and IL-1beta was significantly increased in the pancreas of AP mouse models and in cell lines. Evidence shows that IL-6 seems to be an effective regulator during systemic inflammation in the pancreas<sup>[32]</sup>. *IL-6*<sup>-/-</sup> mice have a lower death rate than wild-type mice with AP, while injection of IL-6 causes more lethal AP in mice<sup>[33]</sup>. IL-6 stimulates the phosphorylation of STAT3 and the production of the neutrophil attractant CXCL1 in pancreatic acinar cells<sup>[34]</sup>. Our study demonstrated that IL-6 inhibitor prevented caerulein-induced adhesion of PBMCs-endothelial cells *in vitro*, but not IL-1beta inhibitor treatment. Therefore, this study implied that PSGL-1 may initiate leukocyte adhesion and infiltration and promote the development of AP when IL-6 is activated.

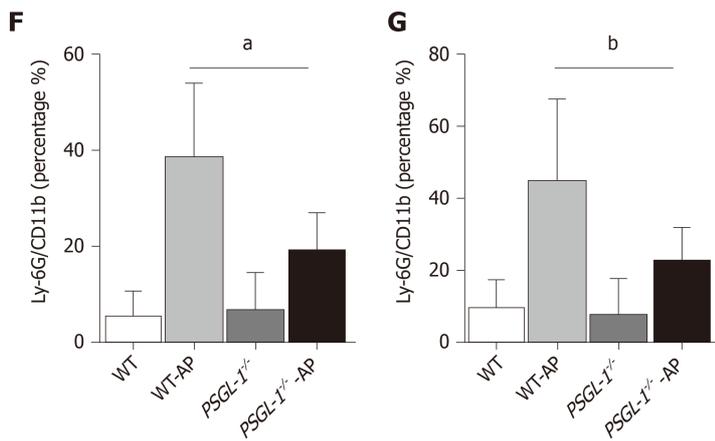
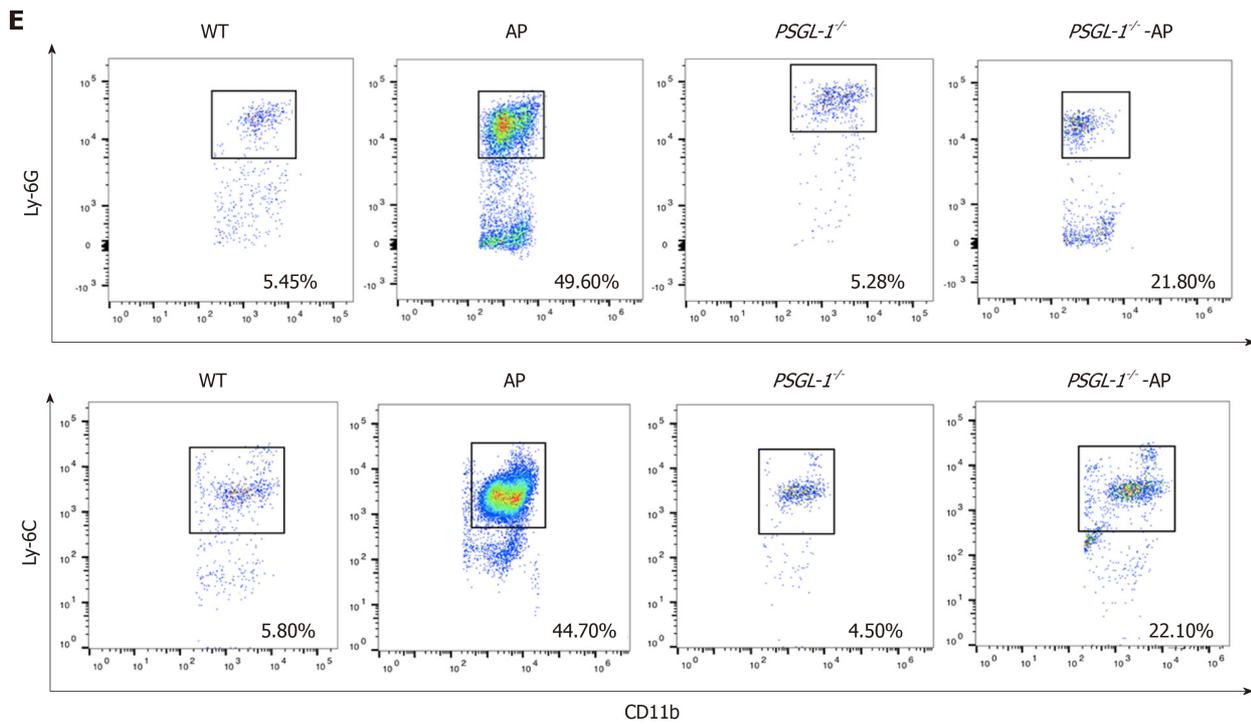
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## CONCLUSION

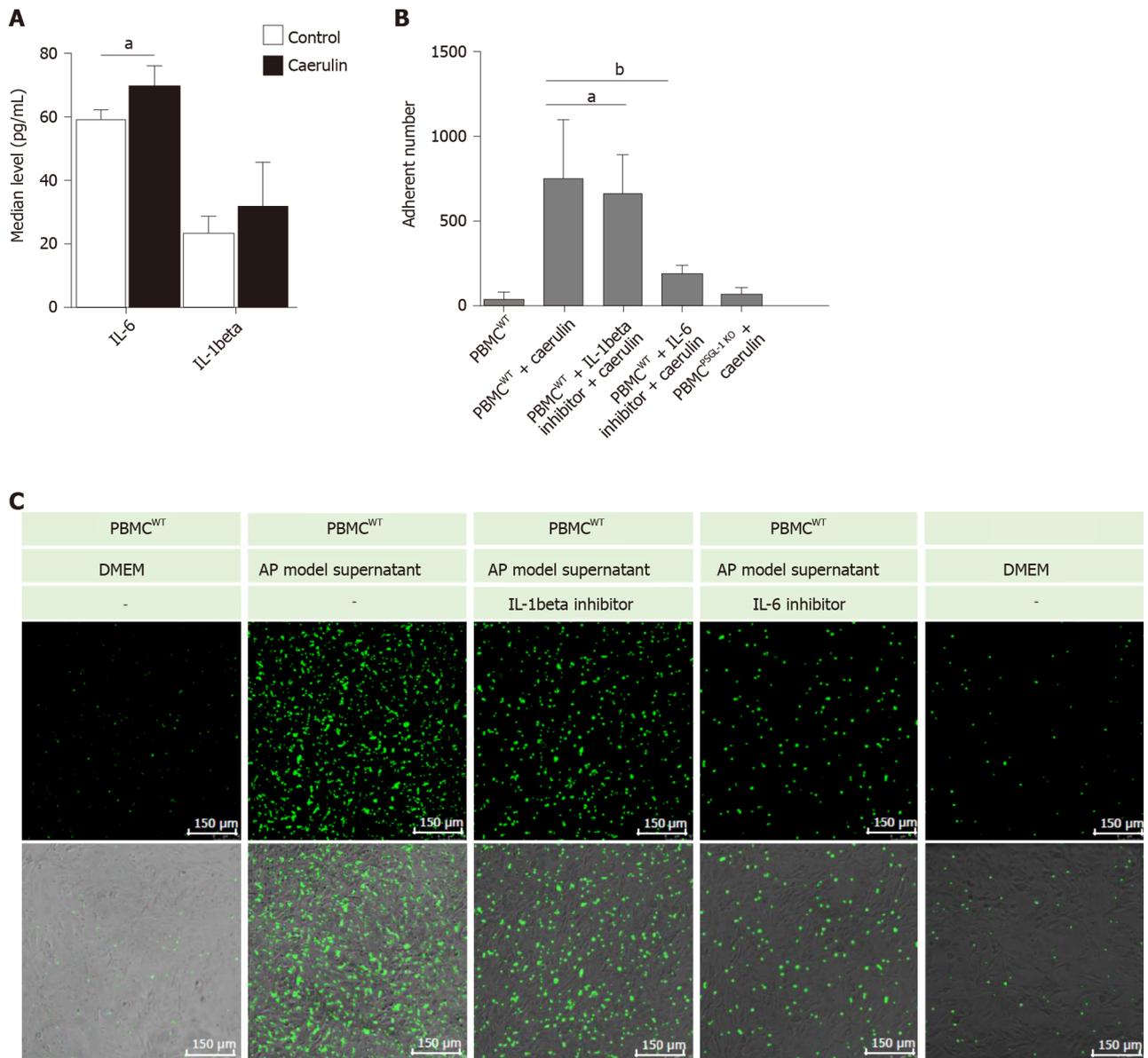
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In conclusion, PSGL-1 plays an important role in the development of AP by promoting inflammatory cell infiltration initiated by leukocyte and endothelial cell adhesion *via* IL-6 stimulation. PSGL-1 deficiency may become a new drug target for AP clinical therapy.





**Figure 4 P-selectin glycoprotein ligand 1 deficiency attenuates the number of circulating leukocytes.** A: Flow cytometry for CD45-positive cells to detect immune cells in the serum; B: Flow cytometry for CD11b-positive cells to detect myeloid cells in the serum; C: Boxplots showing quantification of immune cells in the serum ( $n = 4$ ); D: Boxplots showing quantification of myeloid cells in the serum ( $n = 4$ ); E: Flow cytometry for Ly-6G-positive cells and Ly-6C-positive cells to detect neutrophils and monocytes, respectively, in the serum; F: Boxplots showing quantification of neutrophils in the serum ( $n = 4$ ); G: Boxplots showing quantification of monocytes in the serum ( $n = 4$ ). <sup>a</sup> $P < 0.05$ ; <sup>b</sup> $P < 0.01$ , <sup>c</sup> $P < 0.001$ , Student's  $t$ -test. PSGL-1: P-selectin glycoprotein ligand 1; WT: Wild type; AP: Acute pancreatitis.



**Figure 5 P-selectin glycoprotein ligand 1 deficiency alleviates caerulein-induced leukocyte-endothelial cell adhesion.** A: Enzyme-linked immunosorbent assays for IL-6 and IL-1beta in the supernatants of AR42J cells cultured in caerulein-containing medium ( $n = 4$ ); B: Boxplots showing the quantification of adhered peripheral blood mononuclear cells (PBMCs) ( $n = 4$ ); C: The number of PBMCs that adhered to Bend.3 cells cocultured under different conditions. Bend. 3 cells were cocultured with PBMCs from wild-type or *PSGL-1*<sup>-/-</sup> mice and stained with Green 5-chloromethylformacein diacetate under different conditions. Fluorescent signals were captured using a confocal microscope (Leica, 400 ×). Green represents the number of adhered PBMCs. <sup>a</sup> $P < 0.05$ ; <sup>b</sup> $P < 0.01$ , Student's *t*-test. PSGL-1: P-selectin glycoprotein ligand 1; WT: Wild type; PBMCs: Peripheral blood mononuclear cells; DMEM: Dulbecco's modified Eagle's medium; AP: Acute pancreatitis.

## ARTICLE HIGHLIGHTS

### Research background

Acute pancreatitis (AP)-induced pancreatic injury is positively correlated with the degree of immune cell infiltration. P-selectin glycoprotein ligand 1 (PSGL-1) can regulate leukocyte activation and recruitment in various inflammation-related diseases. Our study showed that the expression of PSGL-1 in leukocytes of AP patients is increased. However, the underlying mechanism has not been fully elucidated.

### Research motivation

Uncontrolled systemic inflammation in AP results in a high risk of morbidity and mortality, but no approved therapies are currently available. Elucidating the mechanism of action of PSGL-1 in AP is expected to help identify new targets for the treatment of pancreatitis.

**Research objectives**

This research aimed to investigate the role and mechanism of PSGL-1 in the inflammatory response during the development of AP.

**Research methods**

We used flow cytometry to detect the expression of PSGL-1 in leukocytes from AP patients and a mouse model of caerulein-induced AP. Next, *PSGL-1<sup>-/-</sup>* mice administered caerulein were used to detect pancreatic injury, inflammatory cytokine expression, and inflammatory cell infiltration. A peripheral blood mononuclear cell-endothelial cell coculture system was used to clarify the mechanism by which PSGL-1 regulates leukocyte adhesion to endothelial cells.

**Research results**

The results of this study indicated that the numbers of monocytes and neutrophils and the expression of PSGL-1 in the peripheral blood of patients were significantly increased. PSGL-1 deficiency reduced serum amylase levels, the expression of IL-1 $\beta$  and IL-6 in the serum and pancreas, the number of infiltrated neutrophils and macrophages in the pancreas, and the number of peripheral circulating neutrophils and monocytes in the AP mouse model. PSGL-1 deficiency alleviated caerulein-induced leukocyte-endothelial cell adhesion.

**Research conclusions**

PSGL-1 deficiency protects against the development of AP by inducing leukocyte-endothelial cell adhesion.

**Research perspectives**

Further research will explore the effect of PSGL-1 on leukocyte function and treatments for AP involving drugs targeting PSGL-1.

**ACKNOWLEDGEMENTS**

Thanks are due to Ya-Ping Zhai and Fang-Yuan Qin for assistance with the experiments.

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