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**Fluorescence guided intraluminal endoscopy in the gastrointestinal tract: A systematic review**

Mortensen OE *et al*. Intraluminal endoscopy in the gastrointestinal tract

Olivia Engholt Mortensen, Nikolaj Nerup, Morten Thorsteinsson, Morten Bo Søndergaard Svendsen, Hironari Shiwaku, Michael Patrick Achiam

**Olivia Engholt Mortensen, Nikolaj Nerup, Morten Thorsteinsson, Michael Patrick Achiam,** Department of Surgical Gastroenterology, Copenhagen University Hospital, Copenhagen 2100, Denmark

**Morten Bo Søndergaard Svendsen,** Copenhagen Academy for Medical Education and Simulation, Copenhagen University Hospital, Copenhagen 2100, Denmark

**Hironari Shiwaku,** Department of Surgical Gastroenterology, Fukuoka University Faculty of Medicine, Fukuoka 814-0133, Japan

**Author contributions:** Mortensen OE, Achiam MP, Nerup N, Shiwaku H, and Svendsen MBS contributed to the idea, conceptualized, and designed the review; Mortensen OE, Achiam MP, and Thorsteinsson M carried out the screening process and data extraction; Mortensen OE drafted the initial manuscript; all authors reviewed and approved the final manuscript as submitted.

**Corresponding author: Olivia Engholt Mortensen, BSc, Research Fellow,** Department of Surgical Gastroenterology, Copenhagen University Hospital, Inge Lehmanns Vej 7, Copenhagen 2100, Denmark. olivia.mortensen@yahoo.com

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**Abstract**

BACKGROUND

Conventional endoscopy is based on full spectrum white light. However, different studies have investigated the use of fluorescence based endoscopy systems where the white light has been supplemented by infrared light and the use of relevant fluorophores. Fluorescence endoscopy utilizes the fluorescence emitted from a fluorophore, visualizing what is not visible to the naked eye.

AIM

To explore the feasibility of fluorescence endoscopy and evaluate its use in diagnosing and evaluating gastrointestinal disease.

METHODS

We followed the PRISMA guidelines for this systematic review. The research covered five databases; PubMed, Scopus, Web of Science, Embase, and the Cochrane Collection, including only studies in English and Scandinavian languages. Authors screened title and abstract for inclusion, subsequently full-text for inclusion according to eligibility criteria listed in the protocol. The risk of bias was assessed for all studies according to the Newcastle-Ottawa Scale. The authors extracted the data and reported the results in both text and tables.

RESULTS

We included seven studies in the systematic review after screening a total of 2769 papers. The most prominent fluorophore was indocyanine green (*n* = 6), and whereas one study (*n* = 1) used Bevacizumab 800-CW. Three studies investigated fluorescence endoscopy in detecting varices, adenomas in patients with familial adenomatous polyposis and neoplasms in the gastrointestinal tract. Four studies evaluated the usefulness of fluorescence endoscopy in assessing tumor invasion. Three of the four studies reported an exceptional diagnostic accuracy (93%, 89% and 88%) in assessing tumor invasion, thus representing better visualization and more correct diagnosis by fluorescence endoscopy compared with the conventional endoscopy. The relationship between the endoscopic findings, tumor invasion, and tumor vascularity was evaluated in two studies showing a significant correlation (d*P* < 0.05 and b*P* < 0.01).

CONCLUSION

The use of fluorescence endoscopy is a promising method adding diagnostic value in the detection of neoplasia, adenomas, and assessment of tumor invasion within the gastrointestinal tract. More studies are needed to utilize the feasibility of fluorescence endoscopy compared with other endoscopic methods.

**Key words:** Fluorescence endoscopy; Gastroscopy; Gastrointestinal tract; Gastrointestinal diseases; Infrared light; Fluorophore; Indocyanine green

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**Core tip:** In the evaluation of tumor invasion, detection of neoplasia and adenomas, studies on fluorescence endoscopy reports promising results.

**INTRODUCTION**

Gastrointestinal diseases are the third most common cause of death with gastrointestinal cancer as the leading cause; in 2018 gastric cancer was estimated to cause 738000 deaths worldwide[1]. The high prevalence is, among others, correlated to multifactorial reasons like lifestyle, physical inactivity, stress, and genetics[2,3]. Conventional endoscopy is widely used for gastrointestinal diseases because it is a minimally invasive and potentially curative procedure, facilitating diagnosis, staging, and treatment. The method of flexible conventional endoscopy is based on the visualization by white light. Thus, allowing the surgeon to visualize the gastrointestinal tract from the inside[4,5]. Recently, studies have examined flexible endoscopy in combination with infrared light, and administration of a fluorophore[6].

Fluorescence arises when a fluorophore is in circulation, and the tissue of interest is exposed to light in a wavelength, that the fluorophore absorbs. When the fluorophore absorbs the photons from the light, an excitation happens where the electrons are shifted to a higher state of energy. Spontaneously, the electrons will shift back to their state of energy releasing the extra energy (emission) as light at another wavelength seen as fluorescence[7,8] (Figure 1). Fluorescence guided flexible intraluminal endoscopy is based on the principle of fluorescence and the spectrum of infrared (IR) light, including near-infrared light. IR light has a wavelength of about 780 nm to 1000 nm. IR light has a limited scattering when it reaches the tissue and a low absorption by water and hemoglobin, thus facilitating a less obstructed penetration through tissue compared with standard white light[9]. The mucosal and submucosal vessels are not visible to the naked eye (in white light), but after intravenous injection of a fluorophore and illumination by IR light, profound structures can be visualized. As angiogenesis and neovascularization are essential factors in carcinogenesis and tumor invasion, visualization of mucosal and submucosal vessels may increase the diagnostic value of the endoscopy[10,11].

Conceptually, the endoscope consists of a light source and an imaging plane–light fibers within the endoscope, with an external camera chip on the tip of the distal end of the camera. The light entering the endoscope for illumination can be white light for standard visualization, whereas when in fluorescence mode, the light primarily consists of the excitatory wavelengths of the fluorophore used. Still in fluorescence mode, after reaching the tissue, the total amount of light reenters the endoscope at the tip. Before reaching the camera chip, the excitatory light needs to be filtered by an optical filter (Figure 1)[12,13]. A frequently used fluorophore is Indocyanine green (ICG), which is excited at the wavelength at 805 nm. Intravenously administered ICG binds to the lipoproteins in the circulation[7]; however, several kinds of other fluorophores exist. The IRDye-800CW is another cyanine fluorophore used for specific protein labeling *e.g.*, Bevacizumab-800CW[13-15]. The aim of this systematic review was to evaluate the diagnostic and therapeutic value of fluorescence-guided flexible intraluminal endoscopy.

**MATERIALS AND METHODS**

The protocol, flow diagram, and the present manuscript adhered to the PRISMA guidelines for systematic reviews[16]. The protocol was submitted for PROSPERO with the registration number CRD42020147516[17].

***Criteria and outcomes***

The eligibility criteria for this systematic review was made according to the principals of participants, interventions, comparison, and outcome. Only human studies examining gastrointestinal diseases and surgical advantages, in general, were included. The studies should use fluorescence endoscopy and compare this method with the use of standard endoscopy or endoscopic expert knowledge, or histopathological examinations. Outcomes of interest were a result representing an increase or decrease in the diagnostic or therapeutic value of fluorescence endoscopy. According to the study design, animal studies and other reviews were excluded. We included randomized controlled trials, case-series with more than five subjects, and prospective/retrospective cohort studies independent of the year of publication and the publication status. Only studies written in English or Scandinavian languages were included.

***Search strategy***

The search string was built in PubMed (Table 1) and adapted to Scopus, Web of Science, Embase, and the Cochrane Collection to identify all the relevant articles for this systematic review. The search string covers all organs from the mouth to the anus, but it does not include the accessory glandular organs. The key words used in the search strategy is shown in Table 1. The database search was performed on June 9th, 2019. Titles and abstracts were screened using an online tool Rayyan[18,19] by four authors (Mortensen OE, Achiam MP, Nerup N, and Thorsteinsson M) to meet the inclusion and exclusion criteria. Consecutively, with two of the authors performing a full-text screening. Subsequently, the reference lists of the included studies were screened to find additional studies. If any discrepancies about inclusion or exclusion, the full-text studies were brought to a meeting and re-examined until consensus. The authors used the web application Rayyan to manage all the data in the screening process. Two authors (Mortensen OE and Thorsteinsson M) performed a data extraction. The handling of data and data from the studies have been extracted from the studies without any modifications and statistical measurements. We extracted data about patients, patient characteristics, diagnosis, fluorophores and dosage used, adverse events, endoscopic findings, diagnostic accuracy, vessel count, and conclusions. No additional analyses were performed.

***Quality assessment***

The Newcastle-Ottawa scale for cohort studies was used to assess the risk of bias of the included studies[20]. The risk of bias assessment focused on the three main subjects; selection, comparability, and outcome (Table 2).

**RESULTS**

The authors screened 2769 articles in Rayyan and added one study from the reference lists of other studies. The authors screened 2069 articles after the removal of duplicates, of those 2052 articles were excluded after the screening of title and abstract. Seventeen articles were assessed for full-text screening, where additional ten studies were excluded due to wrong study design or if full-text versions were not available. Finally, seven studies were included comprising a total of 190 patients (Table 3), selected according to the criteria listed. The full screening process is shown in the PRISMA flow diagram (Figure 2).

***Quality assessment***

The studies were rated for bias according to the Newcastle-Ottawa scale and reported according to their quality (Table 2). All studies were assessed as poor quality due to the lack of comparability and missing control groups. No risk of bias was made across the studies because of the limited number of studies included.

***Studies and definitions***

All the included studies used a system from Olympus (Tokyo, Japan). Intravenous injection of the fluorophore was done in all included studies visualizing the vascularity of the tissue of interest. Six of the seven studies investigated the diagnostic value of fluorescence endoscopy in patients with previously diagnosed adenomas, neoplasms, or cancer (*n* = 170)[12,15,21], and one study investigated the use in detecting esophageal varices (*n* = 20)[22-25].

All studies categorized and evaluated the endoscopic findings differently according to the observed fluorescence appearance. Two studies classified the fluorescence staining asno tumor stain, homogeneous tumor stain, inhomogeneous tumor stain, or pooling of the dye[12,25], while another study categorized the staining as no stain, faint stain, dense stain, homogeneous stain, and pooling of the dye. The definitions were as follows; no stain:A decreased dye accumulation in the tumor compared to surrounding mucosa, homogeneous stain: A diffusely increased dye accumulation in the tumor compared with the surrounding mucosa, inhomogeneous stain: A scattered dye accumulation in the tumor, and pooling of the dye: A substantial dye accumulation in the tumor[24]. In another two studies, they categorized the pooling of the dye/fluorescence categorized as positive or negative[21,23]. The staining definitions and diagnostic values accordingly are shown in Table 3.

***Fluorophores***

Six of seven studies used ICG as a fluorophore[12,21-25]. The dose of ICG ranged from 0.001 to 5 mg/kg bodyweight varying between a fixed dose or different doses of ICG. Four studies reported no adverse events according to ICG[21-24], and the remaining two did not report the frequency or absence of adverse events[12,25]. One study made a dose-response test for Bevacizumab-800CW, which was used as a fluorophore labeling Vascular Endothelial Growth Factor A present in colorectal adenomas and reported no adverse events according to the injections and doses (Table 3)[15].

***Inter- and intraobserver examination***

Three studies assessed inter- or intraobserver agreement in the infrared fluorescence endoscopic examination. One study reported 90% in interobserver agreement[23], while another study reported a 97% interobserver agreement[21]. The third study reported 97% (kappa 0.97) in intraobserver agreement and a 85% (kappa 0.85) in interobserver agreement[25].

***Tumor invasion and neoplasms***

Five studies reported infrared fluorescence endoscopy as useful to assess tumor invasion or detect neoplasia[12,21,23-25]. In a retrospective study of 30 patients with depressed gastric cancers, the authors reported that 21 of 23 intramucosal and submucosal tumors smaller than 1 mm were observed with no stain or faint stain. Seven of seven submucosal tumors larger than 1 mm and more invasive tumors were observed with dense staining or pooling of the dye. Consequently, 28 of 30 both mucosal and submucosal tumors were correctly diagnosed (diagnostic accuracy 93%, Table 3). Additionally, 18 of 19 (accuracy 95%) of tumors with ulcerative changes were correctly diagnosed. Diagnostic accuracy was described as the level of compliance for endoscopic findings by using IR-light and a fluorophore compared with the histopathological examinations[12].

Iseki *et al*[25] (*n* = 37) reported that 16 of 18 mucosal tumors were observed with no stain or homogeneous tumor stain. Seventeen of 19 submucosal or deeper tumors were observed with inhomogeneous tumor stain or pooling of the dye. Consequently, 33 of 37 mucosal and submucosal tumors correctly diagnosed (diagnostic accuracy 89%, Table 3). Additionally, 33 of 37 (accuracy 89%) tumors correctly diagnosed as depressed or ulcerative. The study compared the diagnostic accuracy of fluorescence endoscopy and chromoendoscopy in assessing tumor invasion. Chromoendoscopy had a diagnostic accuracy at 68%, compared with fluorescence endoscopy (89%, a*P* < 0.02). Furthermore, the authors reported that tumor invasion assessed by fluorescence endoscopy was strongly correlated to the degree of tumor vascularity (b*P* < 0.01).

The study of Mataki *et al*[21](*n* = 33) reported all eight gastric adenomas (accuracy 100%) negative for pooling of dye as in contrast to 20 of 25 (80%) for both mucosal and submucosal tumors which were positive for pooling of dye (Table 3) (c*P* < 0.03 for mucosal and submucosal). The authors suggested the fluorescence endoscopy as a diagnostic staging tool to determine if a tumor was eligible to make an endoscopic mucosal resection.

Kimura *et al*[23](*n* = 30) reported one of 20 gastric adenomas or intramucosal tumors as being positive in fluorescence, and eight of ten submucosal tumors as being positive in fluorescence. The study did not state diagnostic accuracy, but the numbers correspond to a sensitivity of 80% and specificity of 95%. Also, a significant correlation between the invasiveness of the tumor, fluorescence, and vessel count was found (d*P* < 0.05).

One study examined early neoplastic lesions within Barrett’s esophagus in 23 cases[24]. Seven cases showed no stain, and histology showed less than high-grade dysplasia in five of those seven cases. Eighteen of 23 showed staining, and histology showed at least high-grade dysplasia, intramucosal carcinoma or submucosal carcinoma. Diagnostic accuracy was 88% (Table 3), sensitivity 90%, specificity 83%, and negative predictive value 71% in identifying the high-grade dysplasia or more advanced histopathology.

***Dose-response***

Two studies made a dose-response examination[15,22]. Okamoto *et al*[22]investigated esophageal varices (*n* = 20) with two studies-a clinical study, and an experimental study to evaluate tissue permeability. The clinical study suggested the optimal dose range of ICG between 0.005-0.01 mg/kg bodyweight based on their evaluation of the fluorescent signal to differentiate between normal mucosa and varices.

One study made a dose-response study with another fluorophore, Bevacizumab-800CW, investigating patients with Familial Adenomatous Polyposis (*n* = 17). Colorectal adenomas were detected with all doses of the fluorophore; 4.5 mg, 10 mg, and 25 mg, whereas normal mucosa showed no fluorescence[15].

**DISCUSSION**

In this systematic review, we identified seven studies using fluorescence endoscopy to assess and evaluate tumor invasion, detect neoplasms, adenomas and esophageal varices. Although fluorescence endoscopy was first described many years ago, this method with interesting results has become even more promising for therapeutic and diagnostic purposes with the recent advances within the field of fluorescence-guided surgery and cancer-specific imaging[26].

***Tumor development and invasion***

Six studies evaluated fluorescence endoscopy according to tumor development and invasion. In one study, fluorescence endoscopy was compared with chromoendoscopy, which is another method used to visualize and detect neoplasia in the gastrointestinal tract. The authors found a significantly higher diagnostic accuracy using fluorescence endoscopy (68% *vs* 89%, a*P* < 0.02)[25]. Furthermore, the authors reported a significant correlation between tumor invasion and tumor vascularity when using fluorescence endoscopy (b*P* < 0.01) as tumors with a tumor stain had significantly more vessels than did tumors without a tumor stain[25-27]. Additionally, the vessels were more varied in size in tumors showing inhomogeneous stain than tumors with a homogeneous stain. The authors suggested that tumor invasion to the submucosa will induce new, permeable vessels, which will result in extravasation of blood observed as pooling of the dye. The association between tumor invasion, fluorescence and vessel count was reproduced in another study with a significant correlation (d*P* < 0.05)[23]. The association of vascularity and tumor invasion was also demonstrated in a study of 44 patients which reported a color change in the endoscopic findings based on the tumor vascularity. The study assessed the tumor vascularity with an endoscopic quantitative analysis of the hemoglobin index[28]. Additionally, another study of 25 specimens from resections of early gastric cancer investigated color changes appearing during endoscopy. They suggested that blood flow, angiogenesis, and the microvasculature in tumors as factors responsible for the endoscopic findings[29]. Nevertheless, these mechanisms are not fully understood and need further assessment.

***Indocyanine green***

For evaluating vascularity, ICG has been used for many years, first for photography, later for angiography in 1969[30]. The contrast has been commercially available for many years, as it has a high level of safety and a very low incidence of adverse events has been reported[31,32]. In this systematic review, four of six included studies (*n* = 106) using ICG specifically reported that no adverse effects occurred[21-24], while the remaining two studies reporting nothing on adverse events. Usually, the recommended dosage of ICG is 0.2-0.25 mg/kg, which must not exceed 2 mg/kg in total[33]. One study included in this review reported an optimal dose of ICG at 0.005 to 0.01 mg/kg body weight[22], while another study reported a very high dosage of ICG at 2-5 mg/kg body weight[25]. However, no consensus about the ICG dosage exists in the studies.

***Cancer-specific probes***

Another subject of emerging clinical interest is the potentially cancer-specific fluorescent probes. Studies investigating the cancer-specific probes reflect the need for developing cancer-specific, optically detectable imaging agents to detect cancers and to add diagnostic and therapeutic value to fluorescence endoscopy. Both cancer-specific probes and the fluorescence endoscopy has been validated by several studies[15,34-37].

Recently, several studies have investigated the urokinase-type plasminogen activator receptor (uPAR) as a cancer-specific probe[38-41]. Using uPAR as a probe, one study subsequently demonstrated the feasibility of uPAR-coupled fluorescent probes. The promising results pointed towards a future using ICG-coupled uPAR probes for imaging and image-guided surgery as the tumor-targeted fluorophores may improve the discrimination between normal and neoplastic tissue. Cancer-specific fluorescent probes may also enable fluorescence-guided endoscopic resection with real-time assessment of the tumor margins, as well as prove to be a novel tool in response evaluation of tumors after chemoradiotherapy. The latter being possible by evaluation of fluctuations in fluorescence intensity caused by changes in tumor vascularity[42].

***Quantitative examination***

Fluorescence endoscopy is still lacking a method to quantify the fluorescent signal to decrease the subjectivity and increase objectivity, sensitivity, and specificity of the method. Some studies have investigated methods to quantitate the fluorescent signal. In the studies included in this review, the fluorescent signal was judged qualitative, meaning visually subjectively, except for one study which quantified the fluorescent signal *ex* *vivo*[15]. In a series of animal studies[43-45], a new method named quantitative-ICG for quantification of perfusion using ICG fluorescence was presented and validated. The quantification of the fluorescent signal will add an important factor to all technologies using fluorescence as a diagnostic marker.

***Limitations***

This systematic review with a focus on human studies using fluorescence endoscopy led to 2769 articles screened, but only seven studies included in the final review, which reflects the limited research within the field. Notwithstanding the limited number of studies, seven of seven studies were rated as poor quality in the Newcastle Ottawa Scale for bias assessment. The low score reflects potential unreliability within the studies, as they all lacked control groups and non-exposed cohorts, thus indicating that this method needs further investigation. However, less strict criteria may have led to more heterogeneous studies included and a more challenging comparison of the endoscopic findings. The exclusion criteria were to keep a homogeneity in the studies and to reflect high clinical applicability of this systematic review.

**CONCLUSION**

In conclusion, this systematic review found that fluorescence endoscopy may add both diagnostic and therapeutic value within the field of gastrointestinal diseases. The majority of the studies included investigated the value within tumor staging, and the detection of adenomas, and neoplasms, thus indicating this method as an opportunity for a more precise diagnosis in the early development of neoplasms and tumors. More studies are needed to examine the usefulness of fluorescence endoscopy compared with other endoscopic methods. Furthermore, the combination of fluorescence endoscopy, quantification of the fluorescent signal, and cancer-specific fluorescent probes has the potential to improve the endoscopic diagnosis, monitoring and therapy of gastrointestinal diseases.

**ARTICLE HIGHLIGHTS**

***Research background***

Different studies have investigated the use of fluorescence based endoscopy systems where the white light has been supplemented by infrared light and the use of relevant fluorophores. Fluorescence endoscopy is among the recent advances within the field of fluorescence-guided surgery and cancer-specific imaging.

***Research motivation***

The aim of this systematic review was to evaluate both the diagnostic and therapeutic value of fluorescence-guided flexible intraluminal endoscopy. Angiogenesis and neovascularization are important factors in tumor invasion, and as mucosal and submucosal vessels are not visible to the naked eye, but after intravenous injection of a fluorophore and illumination by infrared light, profound structures can be visualized.

***Research objectives***

Fluorescence endoscopy can be used within the detection the early development of neoplasms and tumors, adenomas, assessment of tumor invasion within the gastrointestinal tract. Those qualities are a part of the recent advances within the field of fluorescence-guided surgery and cancer-specific imaging.

***Research methods***

The research method was a data analysis. We followed the PRISMA guidelines for this systematic review. The research covered five databases; PubMed, Scopus, Web of Science, Embase, and the Cochrane Collection. Authors screened title and abstract for inclusion, subsequently full-text for inclusion according to eligibility criteria listed in the protocol. The risk of bias was assessed for all studies according to the Newcastle-Ottawa Scale. The authors extracted the data and reported the results in both text and tables.

***Research results***

We included seven studies in the systematic review after screening a total of 2769 papers. Four studies evaluated the usefulness of fluorescence endoscopy in assessing tumor invasion. Three of the four studies reported an exceptional diagnostic accuracy in assessing tumor invasion, thus representing better visualization and more correct diagnosis by fluorescence endoscopy compared with the conventional endoscopy. The relationship between the endoscopic findings, tumor invasion, and tumor vascularity was evaluated in two studies showing a significant correlation. The use of fluorescence endoscopy is a promising method.

***Research conclusions***

This systematic review explored the diagnostic and therapeutic value of fluorescence endoscopy. This study proposes fluorescence endoscopy as a method, which can increase those values, in the context of what is already known. This systematic review reflects a high clinical applicability, and fluorescence endoscopy is a method, that builds on the approach of tumor vascularity. This is the hypothesis of this systematic review and how this cooperate with the diagnostic and therapeutic value.

***Research perspectives***

More studies are needed to utilize the feasibility of fluorescence endoscopy compared with other endoscopic methods exploring the diagnostic and therapeutic value in different clinical issues.

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**Footnotes**

**Conflict-of-interest statement:** All authors state that they have no conflict of interest.

**PRISMA 2009 Checklist statement:** The guidelines of the PRISMA 2009 statement have been adopted.

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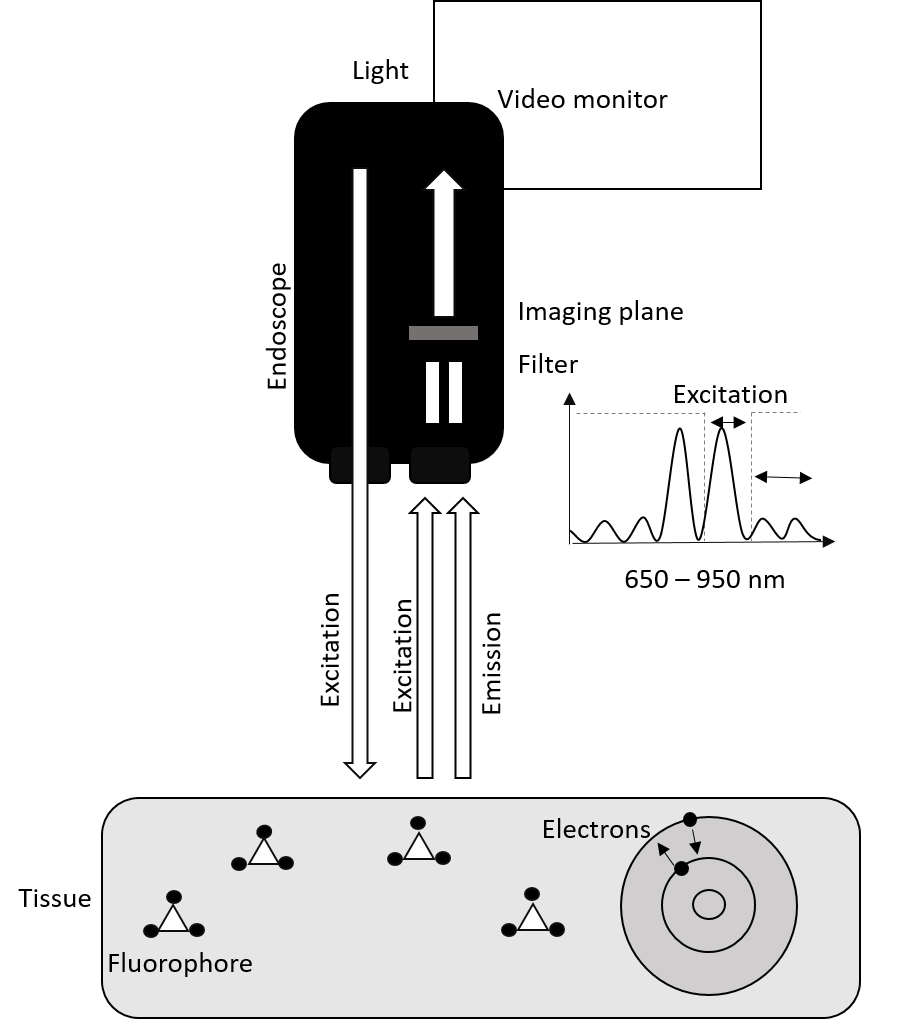
Grade C (Good): 0

Grade D (Fair): 0

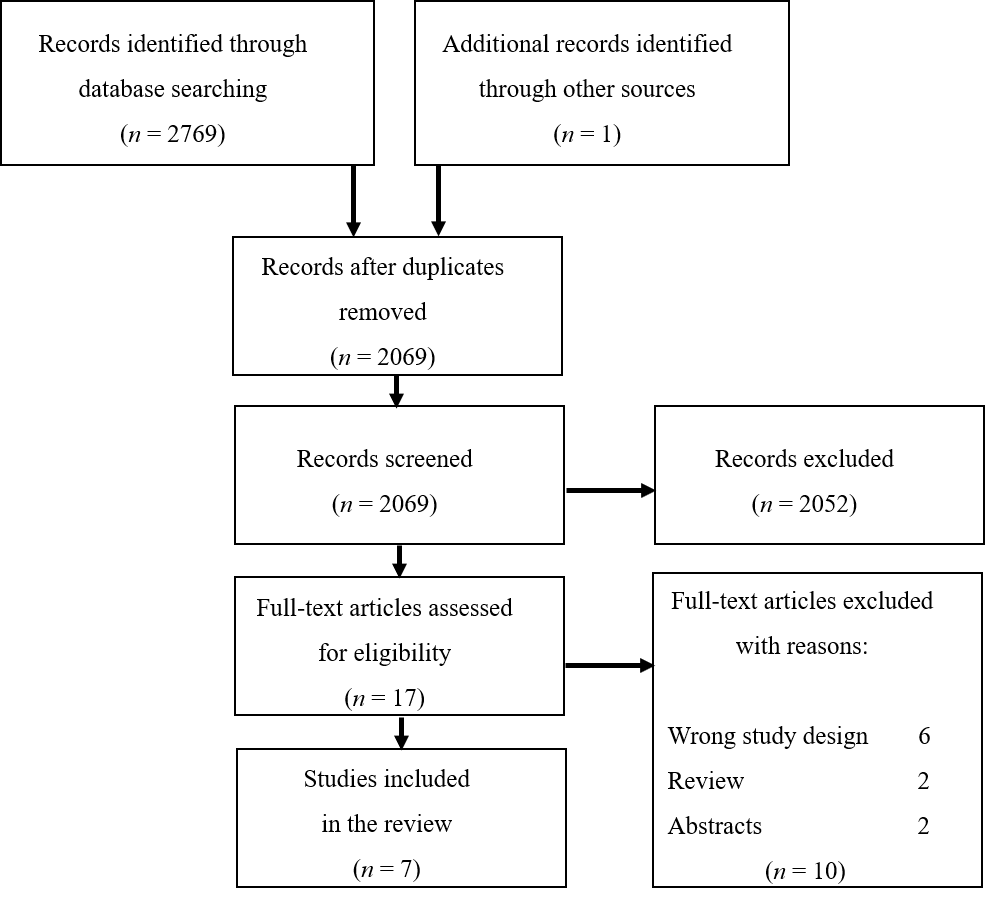
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**Figure Legends**



**Figure 1 The endoscope emits light in the excitation spectrum of the fluorophore injected.** The electrons of the fluorophore will shift from one state of energy to another (excitation), and back, releasing energy as light (fluorescence) at another wavelength (emission). The imaging plane and the filter receive the signal and separate the signals of excitation and emission, only allowing the excitation light to pass.



**Figure 2 The screening process for the systematic review according to the PRISMA flow diagram.**

**Table 1 Search string in Pubmed, Embase, Scopus, Web of Science and Cochrane**

|  |
| --- |
| **Classification** |
| (Endoscop OR Esophagoscop OR Gastroscop OR Gastroscopic Surgical Procedure OR Gastroscopic Surgical Procedures OR Colonoscop OR Colonoscopic Surgical Procedure OR Colonoscopic Surgical Procedures OR Surgery Gastroscopic OR Surgery Colonoscopic) AND (Indocyanine green fluorescence OR Indocyanine Green OR ICG OR fluorescent OR fluorescent dye OR fluorescence OR fluorescein OR near-infrared OR near infrared) AND (Upper Gastrointestinal Tract OR Lower Gastrointestinal Tract OR Upper gastrointestinal disease OR Lower gastrointestinal disease OR Upper gastrointestinal diseases OR Lower gastrointestinal diseases OR gastrointestinal tract OR gastrointestinal diseases OR GI diseases OR GI-diseases OR Upper GI-Diseases OR Lower GI-diseases) |

ICG: Indocyanine green; GI: Gastrointestinal.

**Table 2 Newcastle Ottawa quality assessment scale**

|  |  |  |  |  |  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- |
| **Ref.** | **Selection** | | | | | **Comparability** | | **Outcome** | | | | **Total score** |
| **1** | **2** | **3** | **4** | **Score** | **1** | **Score** | **1** | **2** | **3** | **Score** |
| Iseki *et al*[25],2000 | a | b | a | a | ★☆★★ | - | ☆☆ | a | a | a | ★★★ | Poor quality |
| Mataki *et al*[21], 2003 | a | b | a | b | ★☆★☆ | - | ☆☆ | a | a | a | ★★★ | Poor quality |
| Okamoto *et al*[22], 2005 | a | b | a | b | ★☆★☆ | - | ☆☆ | c | a | a | ☆★★ | Poor quality |
| Ishihara *et al*[12], 2006 | a | b | a | a | ★☆★★ | - | ☆☆ | a | a | a | ★★★ | Poor quality |
| Kimura *et al*[23], 2007 | a | b | a | b | ★☆★☆ | - | ☆☆ | a | a | a | ★★★ | Poor quality |
| Ortiz-  Fernandez-Sordo *et al*[24], 2018 | a | b | a | a | ★☆★★ | - | ☆☆ | a | a | a | ★★★ | Poor quality |
| Hartmans *et al*[15], 2018 | a | b | a | a | ★☆★★ | - | ☆☆ | c | a | a | ☆★★ | Poor quality |

★/a: One star rewarded; ☆/b/c: No star rewarded.

**Table 3 Included studies**

|  |  |  |  |  |  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- |
| **Ref.** | **Study design** | **Patients (*n*)** | **Age (yr)** | **Gender (M/W)** | **Diagnosis** | **Contrast** | **Dosage (mg/kg)** | **Adverse events** | **Endoscopic findings** | **Diagnostic accuracy (%)** | **Vessel count** | **Applicability** |
| Iseki *et al*[25],2000 | Retrospective | 37 | 59 (me) | 25/12 | Gastric cancer | ICG | 2-5 | N/A | 16/18 M tumors: No stain or homogeneous stain. 17/19 SM or more invasive tumors: Inhomogeneous stain or pooling of the dye | 89 | Yes | Tumor invasion |
| Mataki *et al*[21],2003 | Retrospective | 33 | N/A | N/A | Early stage gastric cancer and gastric adenoma | ICG | 1 | None | 0/8 adenomas: + fluorescence. 9/14 M tumors: + fluorescence.  11/11 SM tumors: + fluorescence | N/A | N/A | Tumor invasion |
| Okamoto *et al*[22],2005 | Retrospective | 20 | 65 (me) | 12/8 | Varices | ICG | 2, 0.1, 0.01, 0.005 or 0.001 | None | Clear fluorescence with doses of ICG in 0.005 to 0.01 mg/kg | N/A | N/A | Detection of varices |
| Ishihara *et al*[12],2006 | Retrospective | 30 | N/A | N/A | Gastriccancer | ICG | 2 | N/A | 21/23 M or SM tumors < 1 mm: No stain or homogeneous stain*.* 7/7 SM tumors > 1 mm:Inhomogeneous stain or pooling of the dye | 93 | N/A | Tumor invasion |
| Kimura *et al*[23],2007 | Retrospective | 30 | 71.5 (me) | 20/10 | Early stage gastric cancer and gastric adenoma | ICG | 0.01 | None | 1/20 M tumors: + fluorescence. 8/10 SM tumors: + fluorescence | N/A | Yes | Tumor invasion |
| Ortiz-  Fernandez-Sordo *et al*[24],2018 | Pilot study | 23 | 69 (49-85) (med) | 20/3 | Early neoplastic lesions within Barrett’s esophagus | ICG | 2 | None | 7/23 tumors: No stain (5/7 were less than HGD) 18/23 tumors: Stain(17/18 were at least HGD, MC or SMC) | 88 | N/A | Detection of neoplasms |
| Hartmans *et al*[15],2018 | Retrospective | 17 | 42 (20-65) (med) | 5/12 | FAP | Bevacizumab800CW | 4.5, 10 or 25 mg | None | Colorectal adenomas detected at all doses by fluorescence | N/A | N/A | Detection of colorectal adenomas |

N/A: Not applicable; FAP: Familial adenomatous polyposis; M: Mucosal; SM: Submucosal; me: Mean; med: Median; No stain: Decreased dye accumulation in the tumor compared to surrounding mucosa; Homogeneous stain: Diffuse increased dye accumulation in the tumor compared to surrounding mucosa; Inhomogeneous stain: Scattered dye accumulation in the tumor; Pooling of the dye: Strong dye accumulation in the tumor; HGD: High grade dysplasia; MC: Mucosal carcinoma; SMC: Submucosal carcinoma.