**Name of journal:** *World Journal of Diabetes*

**ESPS Manuscript NO: 5746**

**Columns: TOPIC HIGHLIGHT**

**WJD 5th Anniversary Special Issues (2): 2 diabetes**

**Therapeutic effects of sericin on diabetic keratopathy in Otsuka Long-Evans Tokushima Fatty rat**

**Nagai N *et al*.** Therapeutic effects of sericin on diabetic keratopathy

Noriaki Nagai, Yoshimasa Ito

**Noriaki Nagai, Yoshimasa Ito,** Faculty of Pharmacy, Kinki University, Osaka 577-8502, Japan

**Author contributions:** Nagai N performed research, and analyzed the data; Nagai N and Ito Y wrote the paper.

**Correspondence to: Yoshimasa Ito, PhD,** Faculty of Pharmacy, Kinki University, Higashi-Osaka, Osaka 577-8502, Japan. itoyoshi@phar.kindai.ac.jp

**Telephone:** +81-6-43073638 **Fax:** +81-6-43073638

**Received:** September 14, 2013  **Revised:** October 19, 2013

**Accepted:** November 15, 2013

**Published online:**

**Abstract**

An Otsuka Long-Evans Tokushima Fatty (OLETF) rat provides a useful model for studies to develop corneal wound-healing drugs for use in diabetic keratopathy resulting from type 2 diabetes mellitus. We investigated the effects of sericin on corneal wound healing in OLETF rats. Corneal wounds were prepared by removal of the corneal epithelium, and documented using a TRC-50X. Sericin was instilled into the eyes of rats five times a day following corneal abrasion. The plasma levels of glucose, triglycerides, cholesterol and insulin in 38-wk-old OLETF rats were significantly higher than in normal control rats (LETO rats), and the rate of corneal wound healing in OLETF rats is slower than in normal rat, probably due to the suppression of cell migration and proliferation caused by high plasma glucose levels. The corneal wounds of OLETF rats instilled with saline showed almost complete healing by 72 h after corneal epithelial abrasion. On the other hand, the instillation of sericin has a potent effect in promoting wound healing and wound-size reduction in OLETF rats, and the wounds showed almost complete healing at 48 h after abrasion. The sericin may provide an effective and safe drug to promote corneal wound healing in diabetic keratopathy.

© 2013 Baishideng Publishing Group Co., Limited. All rights reserved.

**Key words:** Sericin; Diabetic keratopathy; Cornea; Type 2 diabetes mellitus; Otsuka Long-Evans Tokushima Fatty rat

**Core tip:** Otsuka Long-Evans Tokushima Fatty (OLETF) rats provide a useful model for studies to develop corneal wound-healing drugs for use in diabetic keratopathy resulting from type 2 diabetes mellitus, and the present study demonstrates that the rate of corneal wound healing in OLETF rats is slower than in normal rat, probably due to the suppression of cell migration and proliferation caused by high plasma glucose levels. The instillation of sericin has a potent effect in promoting wound healing and wound-size reduction in OLETF rats. The sericin may provide an effective and safe drug to promote corneal wound healing in diabetic keratopathy.

Nagai N, Ito Y. Therapeutic effects of sericin on diabetic keratopathy in Otsuka Long-Evans Tokushima Fatty rat. *World J Diabetes* 2013;

**Available from:** URL: http://www.wjgnet.com/esps/

**DOI:** http://dx.doi.org/10.4239/wjd.v0.i0.0000

**INTRODUCTION**

Diabetes mellitus is a common metabolic disorder, affecting more than 190 million people worldwide[1,2]. The prevalence of type 2 diabetes mellitus is increasing rapidly and is affecting the health of millions of humans, and will continue to do so in the near future. Among the factors responsible for the increasing prevalence of this disease are obesity, the consumption of energy-dense diets and low levels of physical activity[3].The development of type 2 diabetes mellitus is associated with pancreatic β-cells dysfunction occurring together with insulin resistance. Normal β-cellss can compensate for insulin resistance by increasing insulin secretion[4], but insufficient compensation leads to the onset of glucose intolerance. Once hyperglycemia becomes apparent, β-cells function progressively deteriorates: glucose-induced insulin secretion becomes further impaired and degranulation of β-cells becomes evident, often accompanied by a decrease in the number of β-cells[5-8]. Recently, it was reported that the production of reactive oxygen species (ROS) mediated by glycation reduces insulin gene transcription and decreases the number of β-cellss through apoptosis[9,10]. Once β-cells are exposed to ROS under diabetic conditions they are likely to be profoundly damaged due to their low levels of antioxidant enzyme expression[11].

Ocular complications secondary to type 2 diabetes mellitus are a well-known cause of diabetic keratopathy[12]. Diabetic keratopathy is an entity that includes slow healing or loose adhesion of the corneal epithelium after wounding in diabetic patients. Histologically, it involves a thickening of the corneal epithelial basement membrane and morphologic changes in the corneal epithelium and endothelium[13-18]. Clinically, the damage to the corneal epithelium during vitreous surgery and retinal photocoagulation sometimes induces vision-threatening corneal complications, such as persistent epithelial defects in diabetic patients[19]. It has been reported that such diabetic keratopathy is experienced by 50% or more of diabetic patients[20].

The corneal wound repair process involves cell adhesion, migration, proliferation, matrix deposition and tissue remodeling[21]. Many of these biological processes are mediated by growth factors, cytokines and other mediators released in injured tissues or cells[22]. These growth factors have been recognized as important mediators of proper wound repair[23], and treatment with growth factors such as platelet-derived growth factor-BB, and recombinant human epidermal growth factor and fibronectin has been shown to be beneficial for patients with chronic pressure ulcers or non-healing diabetic ulcers[24-30]. However, these autologous serum eye drops have problems in terms safety and stability. In addition, for reasons of effectiveness, safety and stable supply, a potent wound-healing agent for human corneal wounds has not yet been introduced. Therefore, a potent corneal wound-healing agent for human corneal wounds that avoids these problems is highly anticipated. In this study, we attempted the development of corneal wound-healing drugs against type 2 diabetic mellitus.

**MODEL FOR STUDY OF TYPE 2 DIABETES MELLITUS**

In the development of corneal wound-healing drugs against type 2 diabetic mellitus, the selection of the experimental animal is very important. Animal models used to investigate diabetic mellitus include streptozotocin-induced diabetic rats as an animal model for insulin-dependent diabetes mellitus[31,32]. Although, hyperglycemia is present in streptozotocin-induced diabetic rats, the general pathophysiology of these models differs from that of non-insulin- dependent diabetes mellitus in humans. On the other hand, the general pathophysiology of Goto-Kakizaki (GK) rats, which are used as a model animal for type 2 diabetes mellitus, is similar[33]. However, GK rats develop type 2 diabetes mellitus without metabolic syndrome. The Otsuka Long-Evans Tokushima Fatty (OLETF) rat is an established model of human type 2 diabetes[34]. Nearly 100% of male OLETF rats develop a diabetic syndrome by 25 wk of age, and hyperglycemia and hyperinsulinemia are exhibited in the early phases of the disease as a result of islet cell hyperplasia and peripheral insulin resistance[35-38], and a close relationship was observed between the plasma glucose and insulin levels (*y* = -51*x* + 335, *r* = -0.9094) in 38-wk-old OLETF rats (Table 1)[39]. With continued aging, the rats eventually develop hypoinsulinemia as a result of the deterioration of islet beta cells (Table 1)[36,38,39]. These changes are similar to those in GK rats, which are used as a model animal for type 2 diabetic mellitus[40]. In addition, OLETF rats develop type 2 diabetic mellitus *via* a metabolic syndrome. Therefore, the changes in the biological characteristics of OLETF rats show an obvious correspondence to those that take place in human type 2 diabetes mellitus, indicating that OLETF rats provide a better model than GK rats for studies to clarify the effects of corneal wound-healing drugs for type 2 diabetic mellitus[39].

**KINETIC ANALYSIS OF THE RATE OF CORNEAL WOUND HEALING**

The maintenance of corneal epithelial cell mass can be viewed as the result of three separate, independent phenomena[41]. Thoft and Friend have termed these: *X*, the proliferation of basal epithelial cells; *Y*, the contribution to the cell mass of the centripetal migration of peripheral cells; and *Z*, epithelial cell loss from the surface. Corneal epithelial maintenance thus can be defined by the equation: *X* + *Y* = *Z*, which simply states that if the corneal epithelium is to be maintained, cell loss must be balanced by cell replacement[42]. The corneal wound healing process is divided into three sequential and partially overlapping steps: epithelial cell loss from the surface (*Z)* reduces and eventually covers the wound surface (*Y*), while cell proliferation (*X*) provides cells to rebuild the tissue and tissue remodeling to restore the stratified epithelium[33,43-48]. Therefore, it is important to development of the evaluation method of cell proliferation (*X*) and migration (*Y*) in the study for corneal wound healing using rat debrided corneal epithelium.

It has been reported that the early stages of epithelial wound closure rely predominately on cell migration rather than cell proliferation; cell proliferation starts approximately 24 h after corneal epithelial injury, after which tissue remodeling to restore the stratified epithelium occurs[49,50]. Furthermore, Zagon *et al*[49] showed that the corneal wounds of Sprague-Dawley rats (250-300 g) were covered due to cell migration by 24 h corneal epithelial injury. After that, the interior damage was healed by cell proliferation[49,51]. On the other hand, the wound surfaces in 38-wk-old normal and OLETF rats were not completely covered by cell migration 24 h corneal epithelial injury. Corneal wound healing in 38-wk-old OLETF rats takes place in clear two phases. The second-phase of healing, involving cell migration and proliferation, may be the source of the delay in corneal wound healing in the 38-wk-old OLETF rats. Therefore, we analyzed the two phases of corneal wound healing in 38-wk-old normal and OLETF rats was analyzed by kinetic analysis. The rate of corneal wound healing is represented by the corneal wound healing rate constant (*α* and *β*, /h), and the corneal wound healing rate constant was calculated from the following equations (Eq. 1 and 2) and the iterative nonlinear least-squares regression procedure MULTI[52].

Corneal wound (%) = wound area 0-72 h / wound area 0 h × 100 Eq. 1 Eq. 2



where *t* is time (0-72 h) after corneal abrasion, and *W*t is the percentage of corneal wound (%) at the corresponding time. *W*0 is the percentage of corneal wound (%) at time 0. *α* and *β* show the corneal wound healing rate constants in the first and second-phases, respectively. *A* and *B* are the corneal wound areas (%) in the *α*- and *β*-phases, respectively. In addition, the represents cell migration, which is the main wound healing process up to 24 h after corneal epithelial abrasion, while the *β*-phase, which takes place 18-72 h after corneal epithelial abrasion, represents cell proliferation[52]. The *β* values for 38-wk-old OLETF rats were significantly lower than those for normal rats at corresponding ages. The contribution ratio *A* of the corneal wound healing process to the *β*-phase in 38-wk-old normal rats was significantly higher than the ratio *B* of the corneal wound healing process for the second-phase. The contribution ratio *B* of the corneal wound healing process for the **-phase in 38-week-old OLETF rats tended to increase. Therefore, a deficit in cell proliferation is predominantly responsible for the delay in corneal wound healing in this model (OLETF rat). On the other hand, it was known that corneal wound healing in normal rats was delayed with aging, and the corneal wound healing rate constant of 38-wk-old normal rat was lower than that of 7-wk-old normal rats. The *α* and *β* values for 38-wk-old normal rats were similar, and the contribution to the corneal wound healing process of ratio *A* for the *β*-phase is significantly higher than the *B* for the *β*-phase. This result suggests that the corneal wounds of old normal rats are repaired by both cell migration and proliferation, and the rate of healing wound becomes equivalent once cell proliferation begins[52].

It is important to understand the mechanisms underlying the delay in corneal wound healing in type 2 diabetic mellitus. In diabetes, the levels of glucose in the cornea and tears are increased. Glucose levels in the corneal epithelium have been reported to be 6-fold higher (1.8 to 12.2 μmol/g *dw*) in diabetic patients than in normal controls[53], and large increases in the glucose content of tears (range 2.16-9.55 mg/dL and 14.69-27.02 mg/dL for normal and diabetic patients, respectively) have also been reported[54-56]. March *et al*[57] reported that the glucose content of tears is approximately 10% the plasma glucose level, and that the glucose content of tears follows changes in plasma glucose levels. High glucose levels suppress the cellular behavior (cell migration and proliferation) of human corneal epithelial cells[58]. In addition, it has been reported that the instillation of insulin normalizes the delay in corneal wound healing in streptozotocin rats[20]. Furthermore, a close relationship was observed between the *α* and *β* values and glucose levels in 38-wk-old OLETF rats, unlike the progression of type 2 diabetic mellitus[52]. These reports indicate that the decrease in corneal wound healing in diabetic keratopathy is caused by a suppression of cell migration and proliferation due to high glucose levels in tears. In addition, this result supports the previous findings for human diabetic keratopathy[58].

**HEALING EFFECT OF SERICIN ON DIABETIC KERATOPATHY**

Proteins such as fibroin and sericin are the main constituents of silk, with fibroin contributing 70% to 80% and sericin 20% to 30% of the total cocoon weight[59]. When cocoons or raw silk are used for textiles, the sericin is mostly removed from the cocoon and disposed of unused. However, sericin has recently been investigated for its activities in biotechnological fields (Figure 1A)[60,61]. Terada *et al*[62] found growth promotion in several human cell lines and mouse hybridomas when sericin was added to the culture media. It is possible that sericin may be applied as eye drops for corneal wound repair. The sericin solutions used in this study were prepared by adding sericin to saline (pH 6.5-7.5, Figure 1), and the instillation of 10% sericin (30 kDa, Seiren Co, Ltd, Fukui, Japan) produces no observable neovascularization or inflammation[51]. In addition, the instillation of 10% sericin resulted in a significantly greater rate of corneal wound size reduction and healing than the instillation of 0.1% hyaluronic acid in Wistar rat eyes, probably by increasing cell migration and proliferation (Figure 2)[51]. In this study, the rates of corneal wound healing in both Long-Evans Tokushima Otsuka rats (LETO rat, normal control) and OLETF rat eyes were faster following the instillation of sericin than in the case of saline instillation, and the rate constant increased with increasing sericin concentration. The rates of corneal wound healing in OLETF rat eyes instilled with 5% or 10% sericin solution were similar to those of LETO rats treated similarly. In addition, the instillation of sericin did not affect glucose levels in the OLETF rats (saline instillation, 213.0 ± 19.7 mg/dL; sericin instillation, 221.1 ± 13.9 mg/dL; mean ± SE of 4 independent OLETF rats). The instillation of 5% or 10% sericin increased the corneal wound healing in OLETF rats 12 h after corneal epithelial abrasion (Figures 3 and 4). Furthermore, we reported that sericin increases the adhesion and proliferation[51].Taken together, the instillation of sericin solution counter the decreases in the rates of cell migration and proliferation, thus preventing the delay in corneal wound healing in OLETF rats (Figure 5).

Next, we investigated the effects of the sericin on cell adhesion and proliferation in a human cornea epithelial cell line (HCE-T), since the effect of sericin in orneal wound healing is result from the increase in cell migration and proliferation. The adhesion and proliferation of HCE-T cells reached a maximum when treated with 0.1%-0.2% sericin solution; the levels of adhesion and proliferation of 1.0% sericin-treated HCE-T cells did not differ significantly from those of control HCE-T cells (Figure 6). On the other hand, the instillation of high concentration sericin solutions (1%-10%) promoted enhanced wound healing in the corneal wound rat model. It is known that the concentration drugs administered in eye drops is diluted to approximately 20% by lacrimal fluids, and that the components of eye drops are excreted though the nasolacrimal duct into the mouth[34]. Thus, our findings suggest that the optimum concentration of the sericin solutions in the *in vivo* instillation experiment, which involves a short residence time, is higher than in the *in vitro* experiment. These findings provide information significant for designing further studies to develop potent drugs to improve the corneal wound-healing ability of diabetic patients.

**CONCLUSION**

OLETF rats provide a useful model for studies to develop corneal wound-healing drugs for use in diabetic keratopathy resulting from type 2 diabetes mellitus, and the present study demonstrates that the rate of corneal wound healing in OLETF rats is slower than in LETO rats, probably due to the suppression of cell migration and proliferation caused by high plasma glucose levels. The instillation of sericin solution has a potent effect in promoting wound healing and wound-size reduction in LETO and OLETF rats. The sericin may provide an effective and safe drug to promote corneal wound healing in diabetic keratopathy.

**REFERENCES**

1 **King H**, Aubert RE, Herman WH. Global burden of diabetes, 1995-2025: prevalence, numerical estimates, and projections. *Diabetes Care* 1998; **21**: 1414-1431 [PMID: 9727886 DOI: 10.2337/diacare.21.9.1414]

2 **Yoon KH**, Lee JH, Kim JW, Cho JH, Choi YH, Ko SH, Zimmet P, Son HY. Epidemic obesity and type 2 diabetes in Asia. *Lancet* 2006; **368**: 1681-1688 [PMID: 17098087 DOI: 10.1016/S0140-6736(06)69703-1]

3 **Schrauwen P**, Hesselink MK. Reduced tricarboxylic acid cycle flux in type 2 diabetes mellitus? *Diabetologia* 2008; **51**: 1694-1697 [PMID: 18587560 DOI: 10.1007/s00125-008-1069-x]

4 **DeFronzo RA**, Ferrannini E. Insulin resistance. A multifaceted syndrome responsible for NIDDM, obesity, hypertension, dyslipidemia, and atherosclerotic cardiovascular disease. *Diabetes Care* 1991; **14**: 173-194 [PMID: 2044434 DOI: 10.2337/diacare.14.3.173]

5 **Porte D**. Banting lecture 1990. Beta-cells in type II diabetes mellitus. *Diabetes* 1991; **40**: 166-180 [PMID: 1991568 DOI: 10.2337/diabetes.40.2.166]

6 **DeFronzo RA**, Bonadonna RC, Ferrannini E. Pathogenesis of NIDDM. A balanced overview. *Diabetes Care* 1992; **15**: 318-368 [PMID: 1532777 DOI: 10.2337/diacare.15.3.318]

7 **Yki-Järvinen H**. Glucose toxicity. *Endocr Rev* 1992; **13**: 415-431 [PMID: 1425483]

8 **Vinik A**. Advancing therapy in type 2 diabetes mellitus with early, comprehensive progression from oral agents to insulin therapy. *Clin Ther* 2007; **29 Spec No**: 1236-1253 [PMID: 18046925 DOI: 10.1016/j.clinthera.2007.07.005]

9 **Matsuoka T**, Kajimoto Y, Watada H, Kaneto H, Kishimoto M, Umayahara Y, Fujitani Y, Kamada T, Kawamori R, Yamasaki Y. Glycation-dependent, reactive oxygen species-mediated suppression of the insulin gene promoter activity in HIT cells. *J Clin Invest* 1997; **99**: 144-150 [PMID: 9011569 DOI: 10.1172/JCI119126]

10 **Kaneto H**, Fujii J, Myint T, Miyazawa N, Islam KN, Kawasaki Y, Suzuki K, Nakamura M, Tatsumi H, Yamasaki Y, Taniguchi N. Reducing sugars trigger oxidative modification and apoptosis in pancreatic beta-cells by provoking oxidative stress through the glycation reaction. *Biochem J* 1996; **320 ( Pt 3)**: 855-863 [PMID: 9003372]

11 **Tiedge M**, Lortz S, Drinkgern J, Lenzen S. Relation between antioxidant enzyme gene expression and antioxidative defense status of insulin-producing cells. *Diabetes* 1997; **46**: 1733-1742 [PMID: 9356019 DOI: 10.2337/diabetes.46.11.1733]

12 **Schultz RO**, Van Horn DL, Peters MA, Klewin KM, Schutten WH. Diabetic keratopathy. *Trans Am Ophthalmol Soc* 1981; **79**: 180-199 [PMID: 7342400]

13 **Taylor HR**, Kimsey RA. Corneal epithelial basement membrane changes in diabetes. *Invest Ophthalmol Vis Sci* 1981; **20**: 548-553 [PMID: 7216670]

14 **Azar DT**, Spurr-Michaud SJ, Tisdale AS, Gipson IK. Decreased penetration of anchoring fibrils into the diabetic stroma. A morphometric analysis. *Arch Ophthalmol* 1989; **107**: 1520-1523 [PMID: 2803103 DOI: 10.1001/archopht.1989.01070020594047]

15 **Friend J**, Ishii Y, Thoft RA. Corneal epithelial changes in diabetic rats. *Ophthalmic Res* 1982; **14**: 269-278 [PMID: 7133621 DOI: 10.1159/000265202]

16 **Schultz RO**, Matsuda M, Yee RW, Edelhauser HF, Schultz KJ. Corneal endothelial changes in type I and type II diabetes mellitus. *Am J Ophthalmol* 1984; **98**: 401-410 [PMID: 6486211 DOI: 10.1016/0002-9394(84)90120-X]

17 **Tsubota K**, Chiba K, Shimazaki J. Corneal epithelium in diabetic patients. *Cornea* 1991; **10**: 156-160 [PMID: 2019126 DOI: 10.1097/00003226-199103000-00011]

18 **Hosotani H**, Ohashi Y, Yamada M, Tsubota K. Reversal of abnormal corneal epithelial cell morphologic characteristics and reduced corneal sensitivity in diabetic patients by aldose reductase inhibitor, CT-112. *Am J Ophthalmol* 1995; **119**: 288-294 [PMID: 7872388]

19 **Perry HD**, Foulks GN, Thoft RA, Tolentino FI. Corneal complications after closed vitrectomy through the pars plana. *Arch Ophthalmol* 1978; **96**: 1401-1403 [PMID: 678179 DOI: 10.1001/archopht.1978.03910060155011]

20 **Zagon IS**, Klocek MS, Sassani JW, McLaughlin PJ. Use of topical insulin to normalize corneal epithelial healing in diabetes mellitus. *Arch Ophthalmol* 2007; **125**: 1082-1088 [PMID: 17698755 DOI: 10.1001/archopht.125.8.1082]

21 **Martin P**. Wound healing--aiming for perfect skin regeneration. *Science* 1997; **276**: 75-81 [PMID: 9082989 DOI: 10.1126/science.276.5309.75]

22 **Werner S**, Grose R. Regulation of wound healing by growth factors and cytokines. *Physiol Rev* 2003; **83**: 835-870 [PMID: 12843410]

23 **Imanishi J**, Kamiyama K, Iguchi I, Kita M, Sotozono C, Kinoshita S. Growth factors: importance in wound healing and maintenance of transparency of the cornea. *Prog Retin Eye Res* 2000; **19**: 113-129 [PMID: 10614683 DOI: 10.1016/S1350-9462(99)00007-5]

24 **Hong JP**, Jung HD, Kim YW. Recombinant human epidermal growth factor (EGF) to enhance healing for diabetic foot ulcers. *Ann Plast Surg* 2006; **56**: 394-38; discussion 394-38; [PMID: 16557070 DOI: 10.1097/01.sap.0000198731.12407.0c]

25 **Mustoe TA**, Cutler NR, Allman RM, Goode PS, Deuel TF, Prause JA, Bear M, Serdar CM, Pierce GF. A phase II study to evaluate recombinant platelet-derived growth factor-BB in the treatment of stage 3 and 4 pressure ulcers. *Arch Surg* 1994; **129**: 213-219 [PMID: 8304833 DOI: 10.1001/archsurg.1994.01420260109015]

26 **Robson MC**, Phillips LG, Thomason A, Altrock BW, Pence PC, Heggers JP, Johnston AF, McHugh TP, Anthony MS, Robson LE. Recombinant human platelet-derived growth factor-BB for the treatment of chronic pressure ulcers. *Ann Plast Surg* 1992; **29**: 193-201 [PMID: 1524367 DOI: 10.1097/00000637-199209000-00001]

27 **Smiell JM**, Wieman TJ, Steed DL, Perry BH, Sampson AR, Schwab BH. Efficacy and safety of becaplermin (recombinant human platelet-derived growth factor-BB) in patients with nonhealing, lower extremity diabetic ulcers: a combined analysis of four randomized studies. *Wound Repair Regen* 1999; **7**: 335-346 [PMID: 10564562 DOI: 10.1046/j.1524-475X.1999.00335.x]

28 **Nishida T**, Nakagawa S, Nishibayashi C, Tanaka H, Manabe R. Fibronectin enhancement of corneal epithelial wound healing of rabbits in vivo. *Arch Ophthalmol* 1984; **102**: 455-456 [PMID: 6703996 DOI: 10.1001/archopht.1984.01040030369040]

29 **Ho PC**, Davis WH, Elliott JH, Cohen S. Kinetics of corneal epithelial regeneration and epidermal growth factor. *Invest Ophthalmol* 1974; **13**: 804-809 [PMID: 4414039]

30 **Nakamura M**, Hikida M, Nakano T. Concentration and molecular weight dependency of rabbit corneal epithelial wound healing on hyaluronan. *Curr Eye Res* 1992; **11**: 981-986 [PMID: 1451529 DOI: 10.3109/02713689209033496]

31 **Sybulski S**, Maughan GB. Use of streptozotocin as diabetic agent in pregnant rats. *Endocrinology* 1971; **89**: 1537-1540 [PMID: 5120649 DOI: 10.1210/endo-89-6-1537]

32 **Aerts L**, Holemans K, Van Assche FA. Impaired insulin response and action in offspring of severely diabetes rats. In: Shafir E, editor. Frontiers in Diabetes Research. Lessons from Animal Diabetes III. London: Slmith Godon, 1990: 561-566

33 **Saika S**, Shiraishi A, Liu CY, Funderburgh JL, Kao CW, Converse RL, Kao WW. Role of lumican in the corneal epithelium during wound healing. *J Biol Chem* 2000; **275**: 2607-2612 [PMID: 10644720 DOI: 10.1074/jbc.275.4.2607]

34 **Kawano K**, Hirashima T, Mori S, Saitoh Y, Kurosumi M, Natori T. Spontaneous long-term hyperglycemic rat with diabetic complications. Otsuka Long-Evans Tokushima Fatty (OLETF) strain. *Diabetes* 1992; **41**: 1422-1428 [PMID: 1397718 DOI: 10.2337/diabetes.41.11.1422]

35 **Ishida K**, Mizuno A, Min Z, Sano T, Shima K. Which is the primary etiologic event in Otsuka Long-Evans Tokushima Fatty rats, a model of spontaneous non-insulin-dependent diabetes mellitus, insulin resistance, or impaired insulin secretion? *Metabolism* 1995; **44**: 940-945 [PMID: 7616855 DOI: 10.1016/0026-0495(95)90249-X]

36 **Ito A**, Uriu K, Inada Y, Qie YL, Takagi I, Ikeda M, Hashimoto O, Suzuka K, Eto S, Tanaka Y, Kaizu K. Inhibition of neuronal nitric oxide synthase ameliorates renal hyperfiltration in streptozotocin-induced diabetic rat. *J Lab Clin Med* 2001; **138**: 177-185 [PMID: 11528370 DOI: 10.1067/mlc.2001.116843]

37 **Sato T**, Asahi Y, Toide K, Nakayama N. Insulin resistance in skeletal muscle of the male Otsuka Long-Evans Tokushima Fatty rat, a new model of NIDDM. *Diabetologia* 1995; **38**: 1033-1041 [PMID: 8591816 DOI: 10.1007/BF00402172]

38 **Yabuki A**, Tahara T, Taniguchi K, Matsumoto M, Suzuki S. Neuronal nitric oxide synthase and cyclooxygenase-2 in diabetic nephropathy of type 2 diabetic OLETF rats. *Exp Anim* 2006; **55**: 17-25 [PMID: 16508208 DOI: 10.1538/expanim.55.17]

39 **Nagai N**, Murao T, Ito Y, Okamoto N, Sasaki M. Enhancing effects of sericin on corneal wound healing in Otsuka Long-Evans Tokushima fatty rats as a model of human type 2 diabetes. *Biol Pharm Bull* 2009; **32**: 1594-1599 [PMID: 19721238]

40 **Wakuta M**, Morishige N, Chikama T, Seki K, Nagano T, Nishida T. Delayed wound closure and phenotypic changes in corneal epithelium of the spontaneously diabetic Goto-Kakizaki rat. *Invest Ophthalmol Vis Sci* 2007; **48**: 590-596 [PMID: 17251454 DOI: 10.1167/iovs.05-1168]

41 **Zhang X**, Tseng H. Basonuclin-null mutation impairs homeostasis and wound repair in mouse corneal epithelium. *PLoS One* 2007; **2**: e1087 [PMID: 17971852 DOI: 10.1371/journal.pone.0001087]

42 **Thoft RA**, Friend J. The X, Y, Z hypothesis of corneal epithelial maintenance. *Invest Ophthalmol Vis Sci* 1983; **24**: 1442-1443 [PMID: 6618809]

43 **Danjo Y**, Gipson IK. Actin 'purse string' filaments are anchored by E-cadherin-mediated adherens junctions at the leading edge of the epithelial wound, providing coordinated cell movement. *J Cell Sci* 1998; **111 ( Pt 22)**: 3323-3332 [PMID: 9788874]

44 **Chen JJ**, Tseng SC. Corneal epithelial wound healing in partial limbal deficiency. *Invest Ophthalmol Vis Sci* 1990; **31**: 1301-1314 [PMID: 1694836]

45 **Stepp MA**, Spurr-Michaud S, Gipson IK. Integrins in the wounded and unwounded stratified squamous epithelium of the cornea. *Invest Ophthalmol Vis Sci* 1993; **34**: 1829-1844 [PMID: 8473121]

46 **Chung EH**, Hutcheon AE, Joyce NC, Zieske JD. Synchronization of the G1/S transition in response to corneal debridement. *Invest Ophthalmol Vis Sci* 1999; **40**: 1952-1958 [PMID: 10440248]

47 **Zieske JD**. Corneal development associated with eyelid opening. *Int J Dev Biol* 2004; **48**: 903-911 [PMID: 15558481 DOI: 10.1387/ijdb.041860jz]

48 **Saika S**, Muragaki Y, Okada Y, Miyamoto T, Ohnishi Y, Ooshima A, Kao WW. Sonic hedgehog expression and role in healing corneal epithelium. *Invest Ophthalmol Vis Sci* 2004; **45**: 2577-2585 [PMID: 15277480 DOI: 10.1167/iovs.04-0001]

49 **Zagon IS**, Sassani JW, Ruth TB, McLaughlin PJ. Cellular dynamics of corneal wound re-epithelialization in the rat. III. Mitotic activity. *Brain Res* 2000; **882**: 169-179 [PMID: 11056196 DOI: 10.1016/S0006-8993(00)02864-X]

50 **Zieske JD**, Gipson IK. “Agents that affect corneal wound healing: modulation of structure and function.” eds by Albert DM, Jakobiec FA. Principles and Practice of Ophthalomology, Saunders Press, Philadelphia, 1994: 1093-1109

51 **Nagai N**, Murao T, Ito Y, Okamoto N, Sasaki M. Enhancing effects of sericin on corneal wound healing in rat debrided corneal epithelium. *Biol Pharm Bull* 2009; **32**: 933-936 [PMID: 19420767 DOI: 10.1248/bpb.32.933]

52 **Nagai N**, Murao T, Okamoto N, Ito Y. Kinetic analysis of the rate of corneal wound healing in Otsuka long-evans Tokushima Fatty rats, a model of type 2 diabetes mellitus. *J Oleo Sci* 2010; **59**: 441-449 [PMID: 20625236 DOI: 10.5650/jos.59.441]

53 **Foulks GN**, Thoft RA, Perry HD, Tolentino FI. Factors related to corneal epithelial complications after closed vitrectomy in diabetics. *Arch Ophthalmol* 1979; **97**: 1076-1078 [PMID: 444136 DOI: 10.1001/archopht.1979.01020010530002]

54 **Gasset AR**, Braverman LE, Fleming MC, Arky RA, Alter BR. Tear glucose detection of hyperglycemia. *Am J Ophthalmol* 1968; **65**: 414-420 [PMID: 5643199]

55 **McDermott AM**, Kern TS, Murphy CJ. The effect of elevated extracellular glucose on migration, adhesion and proliferation of SV40 transformed human corneal epithelial cells. *Curr Eye Res* 1998; **17**: 924-932 [PMID: 9746440 DOI: 10.1076/ceyr.17.9.924.5133]

56 **Sen DK**, Sarin GS. Tear glucose levels in normal people and in diabetic patients. *Br J Ophthalmol* 1980; **64**: 693-695 [PMID: 7426593 DOI: 10.1136/bjo.64.9.693]

57 **March WF**, Mueller A, Herbrechtsmeier P. Clinical trial of a noninvasive contact lens glucose sensor. *Diabetes Technol Ther* 2004; **6**: 782-789 [PMID: 15684630 DOI: 10.1089/dia.2004.6.782]

58 **Fujita H**, Morita I, Takase H, Ohno-Matsui K, Mochizuki M. Prolonged exposure to high glucose impaired cellular behavior of normal human corneal epithelial cells. *Curr Eye Res* 2003; **27**: 197-203 [PMID: 14562170 DOI: 10.1076/ceyr.27.4.197.16598]

59 **Kato N**, Sato S, Yamanaka A, Yamada H, Fuwa N, Nomura M. Silk protein, sericin, inhibits lipid peroxidation and tyrosinase activity. *Biosci Biotechnol Biochem* 1998; **62**: 145-147 [PMID: 9501526 DOI: 10.1271/bbb.62.145]

60 **Tsubouchi K**, Igarashi Y, Takasu Y, Yamada H. Sericin enhances attachment of cultured human skin fibroblasts. *Biosci Biotechnol Biochem* 2005; **69**: 403-405 [PMID: 15725668 DOI: 10.1271/bbb.69.403]

61 **Minoura N**, Aiba S, Gotoh Y, Tsukada M, Imai Y. Attachment and growth of cultured fibroblast cells on silk protein matrices. *J Biomed Mater Res* 1995; **29**: 1215-1221 [PMID: 8557723 DOI: 10.1002/jbm.820291008]

62 **Terada S**, Nishimura T, Sasaki M, Yamada H, Miki M. Sericin, a protein derived from silkworms, accelerates the proliferation of several mammalian cell lines including a hybridoma. *Cytotechnology* 2002; **40**: 3-12 [PMID: 19003099 DOI: 10.1023/A: 1023993400608]

**P-Reviewers:** Liu SH, Sakata N, Shafrir E **S-Editor:** Gou SX

**L-Editor: E-Editor:**

**Figure 1 Picture of sericin and protocol in this study.** A: Picture of sericin; B: Picture of sericin solution. The sericin solutions used in this study were prepared by adding sericin to saline (pH 6.5-7.5); C: Protocol for instillation of sericin. Saline, sericin or hyaluronic acid solutions were instilled into the eyes of rats five times a day.

**Figure 2 Corneal Images of Wistar rats with or without the instillation of sericin solutions.** The corneal epithelium was removed with a BD Micro-SharpTM, and the resulting corneal wounds were dyed with 1% fluorescein solution. Saline, sericin or hyaluronic acid solutions were instilled into the eyes of rats five times a day. a*P* < 0.05 *vs* saline-instilled rat.

**Figure 3 Corneal images.** A: Long-Evans Tokushima Otsuka (LETO) rats; B: Otsuka Long-Evans Tokushima Fatty (OLETF) rats with or without the instillation of sericin solutions.The photograph have been reported in reference 39. The corneal epithelium was removed with a BD Micro-SharpTM, and the resulting corneal wounds were dyed with 1% fluorescein solution. Saline or sericin solutions were instilled into the eyes of rats five times a day.

**Figure 4 Effect of sericin solutions on corneal wound healing.** A: Long-Evans Tokushima Otsuka (LETO) rats; B: Otsuka Long-Evans Tokushima Fatty (OLETF) rats eyes. The data have been reported in reference 39. Saline or sericin solutions were instilled into the eyes of rats five times a day. The data are presented as mean ± SE of 3-5 independent rat corneas. a*P* < 0.05 *vs* saline-instilled rat.

**Figure 5** **The function of cell migration and proliferation in corneal wound healing in 38-wk-old Otsuka Long-Evans Tokushima Fatty rats with or without the instillation of sericin solutions.** The movement of superficial cells shows cell migration, and the number of basal cells represent cell proliferation. OLETF: Otsuka Long-Evans Tokushima Fatty.

**Figure 6 Effect of sericin on the adhesion (A) and growth (B) of human cornea epithelial cell line.** The data have been reported in reference 51. Human cornea epithelial cell line (HCE-T) cells were cultured in Dulbecco's modified Eagle’s medium/Ham’s F12 containing 5% (v/v) heat-inactivated fetal bovine serum, 0.1 mg/mL streptomycin and 1000 IU/mL penicillin. Cell growth was calculated by TetraColor One. The amount of cell adhesion and growth were represented by the following equation: cell adhesion or growth (%) = *Abss*ericin treatment/*Abs*control ×100. The data are presented as mean ± SE of 5-25 experiments. a*P* < 0.05 *vs* control HCE-T cells.

**Table 1 Body weight and some blood test values for diabetes mellitus in 38- and 60-wk-old normal and Otsuka Long-Evans Tokushima Fatty rats**

|  |  |  |
| --- | --- | --- |
|  | **38-wk-old** | **60-wk-old** |
|  | **Normal** | **OLETF** | **Normal** | **OLETF** |
| Weight (g) | 488.6 ± 14.2 | 621.3 ± 19.7a | 526.3 ± 33.0 | 416.3 ± 17.4c |
| Glucose (mg/dL) | 119.3 ± 4.9 | 213.5 ± 15.7a | 140.8 ± 3.6 | 244.3 ± 23.9c |
| Triglycerides (mg/dL) | 128.0 ± 9.3 | 419.8 ± 22.2a | 150.0 ± 14.4 | 335.8 ± 11.0c |
| Totalcholesterol (mg/dL) | 101.4 ± 11.4 | 209.2 ± 11.1a | 83.6 ± 14.3 | 274.5 ± 26.2c |
| Insulin (ng/dL) | 105.5 ± 11.6 | 237.4 ± 26.6a | 111.1 ± 6.7 | 83.0 ± 7.2c |

The data are presented as mean ± SE of 4 independent rats. a*P* < 0.05 *vs* 38-wk-old normal rats for each category; c*P* < 0.05 *vs* 60-wk-old normal rats for each category.OLETF: Otsuka Long-Evans Tokushima Fatty.