

World Journal of *Clinical Cases*

World J Clin Cases 2020 December 26; 8(24): 6213-6545



MINIREVIEWS

- 6213 Role of gut microbiome in regulating the effectiveness of metformin in reducing colorectal cancer in type 2 diabetes

Huang QY, Yao F, Zhou CR, Huang XY, Wang Q, Long H, Wu QM

ORIGINAL ARTICLE

Retrospective Cohort Study

- 6229 Impact factors of lymph node retrieval on survival in locally advanced rectal cancer with neoadjuvant therapy

Mei SW, Liu Z, Wang Z, Pei W, Wei FZ, Chen JN, Wang ZJ, Shen HY, Li J, Zhao FQ, Wang XS, Liu Q

Retrospective Study

- 6243 Three-year follow-up of Coats disease treated with conbercept and 532-nm laser photocoagulation

Jiang L, Qin B, Luo XL, Cao H, Deng TM, Yang MM, Meng T, Yang HQ

- 6252 Virus load and virus shedding of SARS-CoV-2 and their impact on patient outcomes

Chen PF, Yu XX, Liu YP, Ren D, Shen M, Huang BS, Gao JL, Huang ZY, Wu M, Wang WY, Chen L, Shi X, Wang ZQ, Liu YX, Liu L, Liu Y

- 6264 Risk factors for *de novo* hepatitis B during solid cancer treatment

Sugimoto R, Furukawa M, Senju T, Aratake Y, Shimokawa M, Tanaka Y, Inada H, Noguchi T, Lee L, Miki M, Maruyama Y, Hashimoto R, Hisano T

- 6274 Cause analysis and reoperation effect of failure and recurrence after epiblepharon correction in children

Wang Y, Zhang Y, Tian N

Clinical Trials Study

- 6282 Effects of different acupuncture methods combined with routine rehabilitation on gait of stroke patients

Lou YT, Yang JJ, Ma YF, Zhen XC

Observational Study

- 6296 Application of endoscopic submucosal dissection in duodenal space-occupying lesions

Li XY, Ji KY, Qu YH, Zheng JJ, Guo YJ, Zhang CP, Zhang KP

- 6306 Early renal injury indicators can help evaluate renal injury in patients with chronic hepatitis B with long-term nucleos(t)ide therapy

Ji TT, Tan N, Lu HY, Xu XY, Yu YY

Prospective Study

- 6315** Neoadjuvant chemoradiotherapy plus surgery in the treatment of potentially resectable thoracic esophageal squamous cell carcinoma
Yan MH, Hou XB, Cai BN, Qu BL, Dai XK, Liu F

CASE REPORT

- 6322** Uterine rupture in patients with a history of multiple curettages: Two case reports
Deng MF, Zhang XD, Zhang QF, Liu J
- 6330** Pleural effusion and ascites in extrarenal lymphangiectasia caused by post-biopsy hematoma: A case report
Lin QZ, Wang HE, Wei D, Bao YF, Li H, Wang T
- 6337** Eighty-year-old man with rare chronic neutrophilic leukemia caused by CSF3R T618I mutation: A case report and review of literature
Li YP, Chen N, Ye XM, Xia YS
- 6346** Sigmoid colon duplication with ectopic immature renal tissue in an adult: A case report
Namgung H
- 6353** Paraplegia from spinal intramedullary tuberculosis: A case report
Qu LM, Wu D, Guo L, Yu JL
- 6358** Confocal laser endomicroscopy distinguishing benign and malignant gallbladder polyps during choledochoscopic gallbladder-preserving polypectomy: A case report
Tang BF, Dang T, Wang QH, Chang ZH, Han WJ
- 6364** Sclerosing stromal tumor of the ovary with masculinization, Meig's syndrome and CA125 elevation in an adolescent girl: A case report
Chen Q, Chen YH, Tang HY, Shen YM, Tan X
- 6373** Primary pulmonary malignant melanoma diagnosed with percutaneous biopsy tissue: A case report
Xi JM, Wen H, Yan XB, Huang J
- 6380** SRY-negative 45,X/46,XY adult male with complete masculinization and infertility: A case report and review of literature
Wu YH, Sun KN, Bao H, Chen YJ
- 6389** Refractory case of ulcerative colitis with idiopathic thrombocytopenic purpura successfully treated by Janus kinase inhibitor tofacitinib: A case report
Komeda Y, Sakurai T, Sakai K, Morita Y, Hashimoto A, Nagai T, Hagiwara S, Matsumura I, Nishio K, Kudo M
- 6396** Immunotherapies application in active stage of systemic lupus erythematosus in pregnancy: A case report and review of literature
Xiong ZH, Cao XS, Guan HL, Zheng HL

- 6408** Minimally invasive maxillary sinus augmentation with simultaneous implantation on an elderly patient: A case report
Yang S, Yu W, Zhang J, Zhou Z, Meng F, Wang J, Shi R, Zhou YM, Zhao J
- 6418** Congenital nephrogenic diabetes insipidus due to the mutation in *AVPR2* (c.541C>T) in a neonate: A case report
Lin FT, Li J, Xu BL, Yang XX, Wang F
- 6425** Primary gastric melanoma in a young woman: A case report
Long GJ, Ou WT, Lin L, Zhou CJ
- 6432** Extreme venous letting and cupping resulting in life-threatening anemia and acute myocardial infarction: A case report
Jang AY, Suh SY
- 6437** Novel conservative treatment for peritoneal dialysis-related hydrothorax: Two case reports
Dai BB, Lin BD, Yang LY, Wan JX, Pan YB
- 6444** Clinical characteristics of pulmonary cryptococcosis coexisting with lung adenocarcinoma: Three case reports
Zheng GX, Tang HJ, Huang ZP, Pan HL, Wei HY, Bai J
- 6450** Fracture of the scapular neck combined with rotator cuff tear: A case report
Chen L, Liu CL, Wu P
- 6456** Synchronous colonic mucosa-associated lymphoid tissue lymphoma found after surgery for adenocarcinoma: A case report and review of literature
Li JJ, Chen BC, Dong J, Chen Y, Chen YW
- 6465** Novel mutation in the *ASXL3* gene in a Chinese boy with microcephaly and speech impairment: A case report
Li JR, Huang Z, Lu Y, Ji QY, Jiang MY, Yang F
- 6473** Recurrent thrombosis in the lower extremities after thrombectomy in a patient with polycythemia vera: A case report
Jiang BP, Cheng GB, Hu Q, Wu JW, Li XY, Liao S, Wu SY, Lu W
- 6480** Status epilepticus as an initial manifestation of hepatic encephalopathy: A case report
Cui B, Wei L, Sun LY, Qu W, Zeng ZG, Liu Y, Zhu ZJ
- 6487** Delayed diagnosis of prosopagnosia following a hemorrhagic stroke in an elderly man: A case report
Yuan Y, Huang F, Gao ZH, Cai WC, Xiao JX, Yang YE, Zhu PL
- 6499** Oral myiasis after cerebral infarction in an elderly male patient from southern China: A case report
Zhang TZ, Jiang Y, Luo XT, Ling R, Wang JW
- 6504** Rare case of drain-site hernia after laparoscopic surgery and a novel strategy of prevention: A case report
Gao X, Chen Q, Wang C, Yu YY, Yang L, Zhou ZG

- 6511** Extracorporeal shock wave therapy treatment of painful hematoma in the calf: A case report
Jung JW, Kim HS, Yang JH, Lee KH, Park SB
- 6517** Takotsubo cardiomyopathy associated with bronchoscopic operation: A case report
Wu BF, Shi JR, Zheng LR
- 6524** Idiopathic adulthood ductopenia with elevated transaminase only: A case report
Zhang XC, Wang D, Li X, Hu YL, Wang C
- 6529** Successful endovascular treatment with long-term antibiotic therapy for infectious pseudoaneurysm due to *Klebsiella pneumoniae*: A case report
Wang TH, Zhao JC, Huang B, Wang JR, Yuan D
- 6537** Primary duodenal tuberculosis misdiagnosed as tumor by imaging examination: A case report
Zhang Y, Shi XJ, Zhang XC, Zhao XJ, Li JX, Wang LH, Xie CE, Liu YY, Wang YL

ABOUT COVER

Peer-Reviewer of *World Journal of Clinical Cases*, Dr. Adonis Protopapas is a gastroenterology Resident at the first Propaedeutic Department of Internal Medicine of the Aristotle University of Thessaloniki (Greece), located at the A.H.E.P.A Hospital. He earned his Bachelor's degree in 2015 from the Democritus University of Thrace, followed by three Master's of Science degrees, with specializations in clinic pharmacology, medical research methodology, and healthcare management. His research interests are mainly focused on the area of hepatology, although he also participates in various projects related to endoscopy and inflammatory bowel disease. He is particularly fascinated by research on cirrhosis and its complications. (L-Editor: Filipodia)

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Role of gut microbiome in regulating the effectiveness of metformin in reducing colorectal cancer in type 2 diabetes

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Abstract

The prevalence of colorectal cancer (CRC) and type 2 diabetes mellitus (T2DM) is increasing globally. It is rarely noticed that the incidence of CRC is higher in patients with T2DM. What needs to be mentioned is that metformin, a commonly used clinical drug for T2DM, attracts scholars' attention because of its benefits in lowering the risk of developing CRC. Hence, we try to find the common grounds of initiation of T2DM and CRC and the reason why metformin reduces the risk of CRC in patients with T2DM. We noticed consistent changes of gut microbiota, such as elevated *Bacteroides*, *Prevotella* and *Bifidobacterium* and depressed *Firmicutes* and *Lactobacillus*. Furthermore, many studies in recent years have proved that the efficacy of metformin, such as improving blood glucose, depends on the gut microbiota. Coincidentally, the progression of CRC is inseparable from the contributions of gut microbiota. Therefore, we first proposed the concept of the metformin-gut microbiota-CRC (in T2DM) axis to explain the effect of metformin in reducing CRC in patients with T2DM. In this review, we elaborated the new concept and its potential clinical application value.

Key Words: Metformin; Colorectal cancer; Gut microbiota; Type 2 diabetes mellitus

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Core Tip: Metformin has been found to reduce colorectal cancer in patients with type 2 diabetes, but the mechanism is unknown. Studies have confirmed that gut microbiota is

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not only closely related to type 2 diabetes and colorectal cancer, but also mediates the effects of metformin. Therefore, we proposed the concept of the metformin-gut microbiota-colorectal cancer axis in type 2 diabetes mellitus. The concept also provides new ideas for clinical drug use, clinical cancer treatment, and clinical application of gut microbiota.

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INTRODUCTION

Several decades ago, colorectal cancer (CRC) was rarely diagnosed. To date, it has become the fourth deadliest cancer in the world, killing an average of 90000 people every year. The accumulation of environmental and genetic factors leading to genetic mutations and epigenetic changes are the factors in the carcinogenesis of intestinal epithelial cells^[1]. Nevertheless, recent studies have shown that the occurrence of CRC is closely related to the gut microbiota, and the alterations of gut microbiota are inconsistent at different stages^[2,3]. **Table 1** summarizes the main alterations of gut microbiota in human studies related to CRC from 2014 to 2019.

There are more than 547 million patients with diabetes mellitus in the world, of which 373 million have been diagnosed. The number of young patients aged 20-39 exceeded 60 million in 2013^[4]. Recent studies have shown that type 2 diabetes mellitus (T2DM) is also closely related to changes in gut microbiota. **Table 2** summarizes the main alterations of gut microbiota in patients with T2DM. Interestingly, metformin (systematic name: 1-carbamimidamido-N,N-dimethylmethanimidamide, C₄H₁₁N₅), as one of the most well-known drugs to treat T2DM, has been shown to have a positive pharmacological effect on restraining CRC in T2DM patients. Moreover, more and more studies have proved that its pharmacological function is mediated by gut microbiota.

Epidemiological evidence suggests that CRC is predisposed in patients with T2DM^[5]. However, after comparing and analyzing the data of current research, the consistent changes of some gut microbiota were observed in patients with T2DM and patients with CRC. **Table 2** summarizes the similarities in changing gut microbiota in patients with T2DM and patients with CRC. Therefore, based on the facts above, we set up a new concept called the metformin-gut microbiota-CRC axis (in T2DM) to explain that metformin reduces the incidence of CRC in patients with T2DM. More importantly, the new concept can be extended to be the drug-gut microbiota-diseases axis. If this theory is proven to be effective, it will provide new ideas for the use of clinical drugs and the application of gut microbiota for the treatment of clinical diseases.

METFORMIN-GUT MICROBIOTA-CRC AXIS IN T2DM

Metformin suppresses CRC in T2DM

To date, epidemiological evidence suggests that T2DM is one of the risk factors for CRC. However, the latest meta-analysis of observational studies displayed insulin therapy significantly increased the risk of CRC [risk ratio (95% confidence interval (CI)): 1.69 (1.25, 2.27)]^[6]. Compared to this, the latest meta-analysis from observational studies demonstrated that metformin therapy was associated with a significantly lower overall survival [pooled risk ratio = 0.75, 95%CI: 0.66-0.86]^[7], lower CRC-specific survival [hazard ratio (95%CI): 0.66 (0.50-0.87)]^[8], and a lower risk of colorectal neoplasm [risk ratio (95%CI): 0.63 (0.50-0.79)] in patients with T2DM^[9]. *In vivo*, Tomimoto *et al*^[10] suggested administration of metformin significantly reduced the number of tumors larger than 2 mm in diameter in Apc (Min/+) mice. Subsequent studies have shown that metformin suppressed colorectal aberrant crypt foci in

Table 1 Summary of the alteration of gut microbiota in patients with colorectal cancer from 2014 to 2019

Tumor types	Status	Alteration of gut microbiota
Adenomas	Increased ¹	Families: <i>Ruminococcaceae</i> , <i>Porphyromonadaceae</i> Genera: <i>Clostridium</i> , <i>Pseudomonas</i> Species: <i>Fusobacterium nucleatum</i> , <i>Atopobium parvulum</i> , <i>Actinomyces odontolyticus</i>
	Decreased ¹	Order: <i>Clostridiales</i> Family: <i>Lachnospiraceae</i> Genera: <i>Bacteroides</i> , <i>Clostridium</i> Species: <i>Bifidobacterium animalis</i> , <i>Streptococcus thermophilus</i>
Carcinoma	Increased ¹	Classes: <i>Bacilli</i> , <i>Gammaproteobacteria</i> Order: <i>Enterobacteriales</i> Genera: <i>Bacteroides</i> , <i>Parabacteroides</i> , <i>Fusobacterium</i> , <i>Alistipes</i> , <i>Escherichia</i> , <i>Parvimonas</i> , <i>Bilophila</i> , <i>Porphyromonas</i> , <i>Actinomyces</i> , <i>Streptococcus</i> Species: <i>Alistipes putredinis</i> , <i>Bilophila wadsworthia</i> , <i>Lachnospiraceae</i> bacterium, <i>Enterobacteriaceae</i> , <i>Fusobacterium nucleatum</i> spp., <i>Actinomyces odontolyticus</i> , <i>Atopobium parvulum</i>
	Decreased ¹	Classes: <i>Erysipelotrichi</i> , <i>Clostridia</i> Order: <i>Clostridiales</i> Families: <i>Lachnospiraceae</i> , <i>Ruminococcaceae</i> , <i>Clostridiaceae</i> Genera: <i>Bacteroides</i> Species: <i>Bifidobacterium animalis</i> , <i>Streptococcus thermophilus</i>
CRC	Increased ¹	Phyla: <i>Firmicutes</i> , <i>Fusobacteria</i> , <i>Bacteroidetes</i> Families: <i>Erysipelotrichaceae</i> , <i>Prevotellaceae</i> , <i>Coriobacteriaceae</i> Genus: <i>Peptostreptococcus</i> Species: <i>Colinsella aerofaciens</i> , <i>Dorea longicatena</i> , <i>Porphyromonas uenonis</i> , <i>Selenomonas sputigena</i> , <i>Streptococcus anginosus</i> , <i>Desulfovibrio vietnamensis</i> , <i>Bilophila wadsworthia</i> , <i>Fusobacterium nucleatum</i> , <i>Parvimonas micra</i> , <i>Peptostreptococcus anaerobius</i> , <i>Solobacterium moorei</i> , <i>Eubacterium ventriosum</i> , <i>Clostridium hathewayi</i> , <i>Bacteroides clarus</i> , <i>Roseburia intestinalis</i> Others: <i>clbA+</i> bacteria, <i>Clostridium symbiosum</i>
	Decreased ¹	Genus: <i>Bifidobacterium</i> Species: <i>Lachnospira multipara</i> , <i>Eubacterium eligens</i>

¹Compared with healthy subjects. A simultaneous increase or decrease of a certain bacteria in the same stage of colorectal cancer means that the current research on this bacteria is still controversial. CRC: Colorectal cancer.

humans^[11,12] and an azoxymethane-induced animal mode of CRC^[13]. In patients with T2DM, it has been reported that metformin has a positive prophylactic effect on colorectal adenomas^[14] and CRC^[15]. In a multicenter double-blind, placebo-controlled, randomized phase 3 trial, Higurashi *et al*^[16] proved the administration of low-dose metformin for 1 year to patients was safe and reduced the prevalence and number of metachronous adenomas or polyps after polypectomy.

At the cellular level, the effect of metformin is mainly manifested in inhibiting the proliferation of CRC cells as a result of AMPK activation and increased reactive oxygen species production^[17,18] on the one hand and accelerating the apoptosis of CRC cells induced by immune cells and cytokines on the other hand^[19]. Complementarily, metformin may go through a variety of pathways or mechanisms, such as by alteration of cellular responses to oxidative stress, protection of mitochondrial structures *via* suppressing reactive oxygen species production and NF- κ B activity^[20-22], and repression of epithelial-mesenchymal transition induced by IL-6 or TGF- β ^[19,23]. At the genetic level, it has been shown that the upregulation of adenosine A1 receptor^[24], blocking of encoding DNA replication proteins and protooncogene protein synthesis^[25,26], and regulation of the SNAIL/miR-34:ZEB/miR-200 system^[27] are the targets of metformin. In addition, key sites for metformin action have been shown in multiple signaling pathways, such as attenuation of cell stemness *via* inhibiting the

Table 2 Summary of the alteration of gut microbiota in patients with type 2 diabetes mellitus and the common alteration with both type 2 diabetes mellitus and colorectal cancer from 2012 to 2019

Status	Classification	T2DM	T2DM and CRC
Increased ¹	Phyla	Verrucomicrobia, Proteobacteria, Actinobacteria, Euryarchaeota, Firmicutes	Firmicutes
	Order	Lactobacillales, Bacteroidales	
	Families	Lachnospiraceae, Erysipelotrichaceae	Lachnospiraceae, Erysipelotrichaceae
	Genera	Ruminococcus, Enterobacteriaceae, Eggerthella, Desulfovibrio, Escherichia, Haemophilus, Clostridium, Eubacterium, Subdoligranulum, Bacteroides, Parabacteroides, Bifidobacterium spp., Lactobacillus	Enterobacteriaceae, Desulfovibrio, Escherichia, Clostridium, Eubacterium, Bacteroides, Parabacteroides, Bifidobacterium spp. Species: Alistipes,
	Species	Akkermansia, Alistipes, Parabacteroides	
Decreased ¹	Phyla	Tenericutes, Fusobacteria, Firmicutes, Bacteroidetes, Verrucomicrobia, Proteobacteria, Elusimicrobia	
	Families	Lachnospiraceae, Peptostreptococcaceae, Flavobacteriaceae, Clostridiaceae	Lachnospiraceae, Clostridiaceae
	Genera	Bacteroides, Anaerostipes, Roseburia, Faecalibacterium, Bifidobacterium	Bacteroides, Bifidobacterium
	Species	A. muciniphila, F. prausnitzii	

¹Compared with healthy volunteers. A simultaneous increase or decrease of a certain bacteria means that the current research on this bacteria is still controversial. CRC: Colorectal cancer; T2DM: Type 2 diabetes mellitus.

Wnt3a/ β -catenin^[28] and AMPK/PI3K/Akt pathways^[29].

To sum up, in both humans and animals or at both the individual and cellular level, metformin has shown an effect of inhibiting the development of CRC. However, as a treatment for T2DM, the underlying mechanism has not been fully elucidated.

Metformin's action depends on gut microbiota

The changes of gut microbiota have been investigated to be associated with metformin administration, contributing to the beneficial therapeutic effects^[30,31]. Decreased profiles of *Intestinibacter* spp. and *Clostridium* spp. were observed upon administration of metformin to health young men with a corresponding increase of *Escherichia/Shigella* spp. and *Bilophila wadsworthia*^[32]. To date, there are many studies focusing on the role of the gut microbiota that mediate the effect of metformin in regard to improving the metabolic phenotype.

In diet-induced obese mice, an increase in the population of *Akkermansia*, a mucin degradation bacteria, induced the improved glucose homeostasis in the cases of metformin administration^[33]. Subsequent work focused on the effects of improving metabolism and delaying T2DM progression in animal models^[34,35]. The study addressed by Forslund *et al*^[36] supported the opinion from the perspective of the human gut genome that metformin could reduce the consumption of short-chain fatty acid-producing groups in T2DM and improve the transfer of functional microbial groups. Through the implementation of 16S rRNA gene sequencing, it was observed that the participating microbiota were mainly *Akkermansia muciniphila* and short-chain fatty acid-producing bacteria, including *Butyrivibrio*, *Bifidobacterium bifidum*, *Megasphaera*, and *Prevotella* after the metformin intervention^[37,38].

Overall, at the phylum level, the main shifts by metformin in patients with T2DM are reflected in the increase of *Bacteroidetes*, *Actinobacteria*, and *Proteobacteria* with a corresponding decrease of *Firmicutes* and *Verrucomicrobia*. At the genus level, the increased bacteria were mainly *Bacteroides*, *Streptococcus*, *Collinsella*, *Escherichia*, *Clostridium*, and *Subdoligranulum*, while the decreased bacteria were *Faecalibacterium* and *Ruminococcus*^[39,40]. Finally, through a comparative analysis of currently available data, we found that the alteration of gut microbiota involved in the inhibition of CRC by metformin in T2DM may be mainly the increase of *Firmicutes* and decrease of *Bacteroidetes*, *Fusobacteria*, and *Bacteroidetes* at the phyla levels. At the genus level, it is mainly manifested as the increase of *Bifidobacterium* and decrease of *Fusobacterium* (*F. nucleatum*).

In addition, Bauer *et al*^[41], Sun *et al*^[42], and Pryor *et al*^[43] confirmed that metformin improved metabolic dysfunction, such as hyperglycemia, *via* the glucose-SGLT1-sensing glucoregulatory pathway, *B. fragilis*–GUDCA–intestinal FXR axis, and the model of host-microbe-drug-nutrient interactions, respectively. These three new findings are profound to help understand how metformin and gut bacteria together

affect disease progression. Table 3 lists their details and the involved types of gut microbiota. Not only that, with the participation of gut microbiota, other fields have gradually revealed the role of metformin, including anti-inflammatory effects, downregulation of interleukin expression, and ameliorating polycystic ovary syndrome in an animal model^[44]. Moreover, metformin accelerated fatty acid oxidation by regulating host metabolism and longevity under the action of regulating a multipressure metabolic system^[43].

Therefore, it can be confidently said that the gut microbiota, such as *Akkermansia*, *B. fragilis* and *E. coli*, mediate the pharmacological effects of metformin in different fields separately.

Gut microbiota affects the occurrence and development of CRC

At present, many studies have confirmed that gut microbiota affect the occurrence and development of CRC. In humans, Yachida *et al*^[2] used metagenomics analysis techniques to analyze the changes of gut microbiota in different stages of CRC on samples from a large cohort of 616 participants. They found the relative abundance of *Fusobacterium nucleatum* spp. was significantly elevated continuously from intramucosal carcinoma to more advanced stages. In addition, *Atopobium parvulum* and *Actinomyces odontolyticus*, which co-occurred in intramucosal carcinomas, were significantly increased only in multiple polypoid adenomas and/or intramucosal carcinomas. Moreover, Yu *et al*^[45] has shown that *F. nucleatum* was abundant in CRC tissues in patients with recurrence post chemotherapy. *F. nucleatum* promoted CRC resistance to chemotherapy by targeting TLR4 and MYD88 innate immune signaling and specific microRNAs to activate the autophagy pathway. By 16S rRNA gene sequencing of stool samples in taxon-based analysis, stool of conventional adenoma patients was depleted in a network of *Clostridia* operational taxonomic units from families *Ruminococcaceae*, *Clostridiaceae*, and *Lachnospiraceae* and enriched in the classes *Bacilli* and *Gammaproteobacteria*, order *Enterobacteriales*, and genera *Actinomyces* and *Streptococcus*^[46].

In animal models through fecal bacteria transplantation, the intestinal microbiota of CRC patients promoted the progression of intestinal adenomas in *Apc^{min/+}* mice^[47]. Donohoe *et al*^[48] proved that dietary fiber protects against colorectal tumorigenesis in a microbiota- and butyrate-dependent manner by a gnotobiotic mouse model. Zhu *et al*^[49] found that editing of the gut microbiota reduced carcinogenesis in colitis-associated CRC. Qin *et al*^[50] suggested that the gut microbiome, under specific dietary exposures, stimulates a reprogramming of the enhancer landscape in the colon with downstream effects on transcription factors, which may be associated with CRC development. In addition, the immune system plays an important role in the effect of gut microbiota on CRC. Cremonesi *et al*^[51] demonstrated that gut microbiota modulated T cell trafficking in human CRC. Wang *et al*^[52] has shown that a purified membrane protein from *Akkermansia muciniphila* or pasteurized bacterium blunts colitis-associated tumorigenesis by modulation of CD8⁺ T cells. Meanwhile, Long *et al*^[53] has also shown that *Peptostreptococcus anaerobius* promoted the occurrence of CRC and regulated tumor-related immunity. At the cellular level, Belcheva *et al*^[54] demonstrated that gut microbial metabolism drives transformation of MSH2-deficient colon epithelial cells.

To sum up, in both humans and animals, gut microbiota is closely related to CRC, and this relationship refers to the immune system. In humans, studies have elaborated changes in the types of gut microbiota, and animal and cell experiments have further elaborated the possible mechanisms by which gut microbiota affects CRC. In Wong *et al*^[55]'s review, the gut microbiota is thought to influence colorectal carcinogenesis *via* microbial-derived factors such as metabolites or genotoxins, promotion of cancer, or proliferating as opportunistic microorganisms in the tumor-associated micro-environment and the activation of procarcinogenic signaling pathways.

Changes in different gut microbiota by metformin at different stages of CRC

As we described before, the changes in the gut microbiota are inconsistent at different stages of development of CRC. According to the interaction between metformin and gut microbiota, does metformin mediate the changes of different gut microbiota at different stages of the development of CRC?

Based on the eighth Union for International Cancer Control Tumor Node Metastasis Classification of Malignant Tumors, at the phylum level in stage 0/pTis, the enriched bacteria are mainly *Actinobacteria* accompanied by a certain amount of *Proteobacteria*, *Firmicutes*, and *Fusobacteria*. In stage SI/II, the bacteria that are enriched are *Proteobacteria*, *Firmicutes*, *Fusobacteria*, and *Bacteroidetes*, and they show roughly the same abundance. In the stage SIII/IV, the enrichment of *Firmicutes* and *Bacteroidetes* is more

Table 3 Summary of alteration of gut microbiota by metformin in patients with type 2 diabetes mellitus that may affect colorectal cancer from 2014 to 2020

Status	Classification	Metformin-related	CRC-related			
			Multiple polypoid	Stage 0/pTis	Stage SI/II	Stage SIII/IV
Increased ¹	Phyla	<i>Bacteroidetes</i> , <i>Actinobacteria</i> , <i>Proteobacteria</i>	<i>Proteobacteria</i>	<i>Firmicutes</i> , <i>Fusobacteria</i> <i>Bacteroidetes</i>	<i>Firmicutes</i> , <i>Fusobacteria</i> <i>Bacteroidetes</i>	<i>Firmicutes</i> , <i>Fusobacteria</i> <i>Bacteroidetes</i>
	Genus	<i>Bacteroides</i> , <i>Streptococcus</i> , <i>Collinsella</i> , <i>Escherichia</i> , <i>Clostridium</i> , <i>Subdoligranulum</i>		<i>F. nucleatum</i>	<i>F. nucleatum</i>	<i>F. nucleatum</i>
	Species	<i>Escherichia</i> spp., <i>Anaerostipes</i> , <i>Blautia</i> , <i>Akkermansia muciniphila</i>	<i>Atopobium parvulum</i>	<i>Solobacterium moorei</i> , <i>Lactobacillus sanfranciscensis</i> , <i>Gemella morbillorum</i> , <i>Actinomyces odontolyticus</i> , <i>Desulfovibrio longreachensis</i> , <i>Phascolarctobacterium succinatutens</i>	<i>Solobacterium moorei</i> , <i>Peptostreptococcus stomatis</i> , <i>Peptostreptococcus anaerobius</i> , <i>Parvimonas micra</i> , <i>Gemella morbillorum</i>	<i>Solobacterium moorei</i> , <i>Peptostreptococcus stomatis</i> , <i>Peptostreptococcus anaerobius</i> , <i>Lactobacillus sanfranciscensis</i> , <i>Parvimonas micra</i> , <i>Gemella morbillorum</i>
Decreased ¹	Phyla	<i>Firmicutes</i> , <i>Verrucomicrobia</i>				
	Genus	<i>Faecalibacterium</i> , <i>Ruminococcus</i>		<i>Bifidobacterium</i>		
	Species	<i>Intestinibacter</i> spp., <i>Intestinibacter bartlettii</i> , <i>Alistipes</i> , <i>Oscillibacter</i> , un-Ruminococcaceae				
Other axes	Increased ¹	<i>Lactobacillus</i> (Glucose-SGLT1-Sensing Glucoregulatory Pathway) <i>E. coli</i> , <i>Proteobacteria</i> (Host-GUDCA-intestinal FXR axis)				
	Decreased ¹	<i>Bacteroides fragilis</i> (B. fragilis–GUDCA–intestinal FXR axis)				

¹Compared with metformin untreated. CRC: Colorectal cancer; T2DM: Type 2 diabetes mellitus.

obvious. *Proteobacteria*, *Actinobacteria*, and *Fusobacteria* are also present, but their abundance is not very high and is roughly equal^[2]. In a nonblinded, one-armed intervention study, the relative abundance of 11 bacterial genera significantly changed during metformin intervention but returned to baseline levels after treatment cessation^[32], which demonstrated that the effects of metformin on the gut microbiota are transient. However, for patients with T2DM, this condition is nothing to worry about because they require long-term medication. Moreover, there was a significant decrease of the phylum *Firmicutes* and of the ratio of *Firmicutes* to *Bacteroidetes* after taking metformin^[39,56]. The other result of the microbiome analyses indicated a shift in the bacterial distribution in fish exposed to metformin, leading to an increase of *Proteobacteria* and a reduction of *Firmicutes* and *Actinobacteria*^[57]. Compared with the previous data, metformin use from intramucosal cancer to advanced stage is likely to mediate its anticancer effect by provoking different gut microbiota. In stage 0/pTis, *Actinobacteria* may be the main target of the

action of metformin, which is specifically manifested by attenuating its abundance. In stage SI/II and SIII/IV, the decrease of the phylum *Firmicutes* and of the ratio of *Firmicutes* to *Bacteroidetes* may be the main function of metformin. Of course, these processes may also be accompanied by the regulation of other flora. Table 3 details the changes. Therefore, the use of metformin to interfere with different types of gut microbiota at different stages of the development of CRC may be a means to improve the survival rate and prognosis of CRC in the future.

Therapeutic effectiveness of metformin and changes in gut microbiome in recurrent CRC

Patients with stage III colon cancer have a risk of recurrence ranging between 15% and 50%^[58]. Interestingly, the administration of metformin is associated with decreased recurrence of carcinoma^[59], covering the colorectal adenoma (hazard ratio 0.572, 95%CI 0.385-0.852) in diabetic patients with previous colorectal adenoma^[60].

So, how does the gut microbiota influence CRC recurrence? A recent study showed that higher fear of cancer recurrence was associated with lower relative abundance of *Firmicutes* and higher relative abundance of *Bacteroidetes* at the phylum level and higher relative abundance of *Bacteroides* and lower relative abundance of *Lachnospiraceae* and *Ruminococcus* at the genus level. The chemotherapy-induced changes in gut microbiota influence the fear of cancer recurrence^[61]. In addition, colorectal adenoma resection gave rise to a significant increase of *Parabacteroides* postoperatively. The microbiota signature of *Parabacteroides*, *Streptococcus*, and *Ruminococcus* showed an optimal discriminating performance of postoperative status^[62]. These changes suggested that gut microbiota may be used in the future to prevent postoperative recurrence of CRC. Finally, it has been stressed that the intervention of metformin can directly affect the gut microbiota profile. In other words, metformin may inhibit the recurrence of CRC by changing the abundance structure of the corresponding gut microbiota.

The recurrence of CRC is resistant to routine chemotherapy and thought to be due to the enrichment of cancer stem cells. It has been reported that long term treatment of metformin impedes development of chemoresistance by regulating cancer stem cell differentiation in cancer cells^[63]. Combination metformin with chemotherapeutics showed an overall modest but intriguing activity in patients with refractory CRC^[64] and a synergistic inhibitory cytotoxicity in human CRC cells^[65]. Moreover, metformin can act as an alternative radiosensitizing agent to 5-fluorouracil during neoadjuvant treatment for rectal cancer^[66]. In addition, a large number of studies have confirmed that metformin, as a cancer stem cell-targeting agent in ovarian cancer, has a direct inhibitory effect on cancer stem cells^[63,67]. The metformin intervention reduced expression of cancer stem cell markers and stemness-related genes in primary oral cancer cells^[68].

Interestingly, microbiota also have a substantial role in the effectiveness of chemotherapy, chemoresistance, and related side effects^[69]. For instance, the presence of specific bacterial species such as *Slackia* and *Blautia obeum* may act as microbial markers associated with drug resistance monitoring^[70]. The inhibition of the growth of *F. nucleatum* significantly augments the efficiency of first-line chemotherapy treatments of CRC^[71]. In addition, chemotherapy-induced changes in gut microbiota impact chemotherapy *via* influencing fear of cancer recurrence^[61]. Moreover, in terms of cancer stem cells, gut microbiota regulate tumor metastasis *via* impacting circular RNA expression to regulate levels of corresponding miRNAs^[72].

All in all, in the recurrence of CRC due to both the chemoresistance and the enrichment of cancer stem cells the gut microbiota plays a vital role, which may be one of the reasons why metformin can inhibit the recurrence of CRC in T2DM patients.

Metformin-gut microbiota-CRC (in T2DM) axis

Metformin, known as a “magic drug”, reduces the incidence of CRC in people with T2DM. Moreover, the gut microbiota has been shown to mediate the pharmacological effects of metformin and the progression of CRC. Finally, the shift of gut microbiota has similarity in the occurrence and development of T2DM and CRC. Therefore, a network of metformin-gut microbiota-CRC (in type 2 diabetes) axis could be assumed. In the network, the gut microbiota plays a key role as a bridge that affects not only the pharmacological effects of metformin but also the occurrence of CRC in T2DM patients.

We have hypothesized the potential mechanism of action of this axis around the gut microbiota. On the one hand, these mechanisms will pave the way for us to understand how gut microbiota can participate in drug therapy. On the other hand,

they will provide ideas for the prospective clinical application and transformation of gut microbiota. More importantly, this axis can be expanded into a profound network of gut microbiota-drug-disease to explain the crosstalk relationship between gut microbiota, drugs, and diseases.

UNDERLYING INVOLVED MECHANISMS

The above narrations provide an elaborate explanation about the metformin-gut microbiota-CRC axis (in T2DM), but we need to know more details about its underlying mechanisms. **Figure 1** provides an exhaustive overview. There are two aspects: (1) Metformin may change the gut microbiota after entering the intestine. The changed gut microbiota has several cascade reactions to affect tumorigenesis, including reduction of inflammation, production of metabolites, and regulation of immunity; and (2) Metformin may play a pharmacological role to influence cancer-related systems in colorectal epithelial cells in the presence of gut microbiota.

Metformin entering the intestine changes the composition of the gut microbiota leading to the cascade anticancer effects

Noticeably, before oral drugs are absorbed into our bloodstream, they must pass through our intestines for catabolism. In this process, the drugs are likely to first react with the intestinal microorganisms colonized in the intestinal epithelium, one of which is gut microbiota. Therefore, the drug may cause a cascade-like reaction related to gut microbiota by changing the composition and abundance of the gut microbiota. The drug may also be metabolized into other secondary products under the action of gut microbiota to exert subsequent efficacy. But the causation and deep interaction mechanism still remain exclusive. We have mentioned above that the gut microbiota involved in the inhibition of CRC by metformin in T2DM may be mainly *Bacteroides*, *Ruminococcus*, *Clostridium*, *Firmicute*, *Lactobacillus*, and *E. coli*. After these gut microbiota are changed, it may cause subsequent cancer suppression *via* regulation of inflammation and immunity and reduction of the production of genotoxic metabolites, *etc.*

Intestinal inflammation and related pathways are closely related to the initiation of CRC. In terms of regulating inflammation, after the mice were treated with metformin, the level of inflammatory markers TNF- α , IL-6, and IL-17 α in plasma was reduced with a corresponding increase of the level of IL-10, which was accompanied by a reduction in *Helicobacter pylori*. Lee *et al.*^[44] suggested that fecal microbiota transplantation using metformin-treated mouse fecal material can upregulate the expression of GLP-1 and pattern recognition receptors TLR1 and TLR4. It has been shown that CRC-enriched genotoxic polyketide synthase (*pks*) + *E. coli*, *E. faecalis*, and *A. fingoldii* and TLR2 and/or TLR4 pathway-related bacteria, such as *F. nucleatum* and *Peptostreptococcus anaerobius*, are related closely to intestinal inflammation^[53]. In our previous discussion, these bacteria are likely to be the targets of metformin, which provides a theoretical basis for our conjecture.

In terms of innate immune response, it has been proven that metformin specifically affects the accumulation of IFN- γ producing NK cells, IL-4, and IL-17 producing NKT cells and IL-12 producing macrophages/dendritic cells in the intestine in animal models^[73]. In addition, the latest studies have shown that metformin could upregulate protein molecules involved in the classical immune pathway such as PMK-1/p38MAPK^[74], RAGE ligands^[75], and TLRs^[76]. It should be mentioned that this process is likely to be mediated by gut microbiota. *F. nucleatum* has been shown to activate autophagy *via* the TLR4 and MYD88 pathways closely related to innate immunity^[45].

In terms of adaptive immune response, it has been mentioned that metformin regulates adaptive immune cell infiltration^[77,78], the expression of related immune factors, such as IFN- γ ^[79], IL-2^[80], and IL-6^[23,81,82], and T cell metabolic reprogramming in tumor tissues^[83]. Nevertheless, it must be pointed out that these effects may also be manifested in the presence of gut microbiota. Cremonesi *et al.*^[51] proved that gut microbiota modulated T cell trafficking into human CRC including CCL5, CXCL9, and CXCL10 for cytotoxic T lymphocytes and T helper (Th) 1 cells, CCL17, CCL22, and CXCL12 for Th1 and regulatory T cells, CXCL3 for follicular Th cells, and CCL20 and CCL17 for IL-17-producing Th cells. Sethi *et al.*^[84] showed that gut microbiota depletion significantly reduced tumor burden in subcutaneous and liver metastasis models of pancreatic cancer, colon cancer, and melanoma. However, the effect was counteracted in Rag1-knockout mice that lacked mature T and B cells. In fact, gut microbiota is likely more complicated in the mechanisms of drug-mediated immunity affecting the

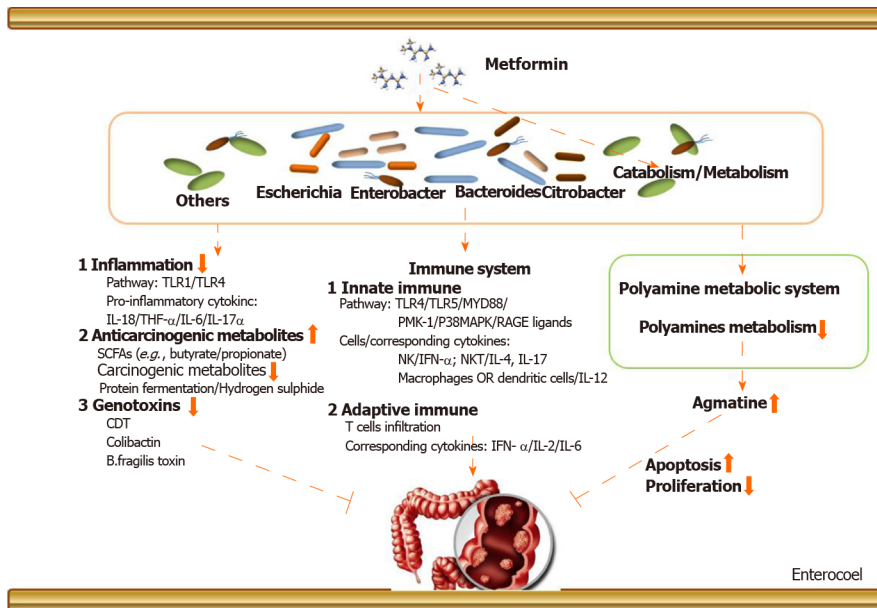


Figure 1 The potential metformin-gut microbiota-colorectal cancer axis in type 2 diabetes mellitus. (1) Metformin may change the gut microbiota after entering the intestine. The changed gut microbiota has cascade reactions to affect tumorigenesis, including reducing inflammation, regulating immunity, and producing metabolites, such as bile acid and genetic toxins. (2) Metformin may play a pharmacological role to influence cancer-related systems in colorectal epithelial cells in the presence of gut microbiota. CDT: Cytolethal distending toxin; SCFAs: Short-chain fatty acids.

development of CRC. A lot of work needs to be done to supplement this field.

In addition, it should be mentioned that after metformin changes the abundance of gut microbiota, the metabolites of the gut microbiota such as carcinogenic-related metabolites and genotoxicity will also change accordingly. Some of these metabolites are cancer-promoting, such as products of protein fermentation, hydrogen sulfide, bile acid metabolism, and ethanol, but some show protective effects. For example, short-chain fatty acids (butyrate and propionate) are anti-inflammatory molecules. Butyrate can inhibit histone deacetylase in colon epithelial cells and immune cells to downregulate proinflammatory cytokines and induce apoptosis in CRC cell lines. Therefore, the reduction of gut microbiota that produces procarcinogenic metabolites and the increase of gut microbiota that produces anti-inflammatory metabolites are likely to be the reason of metformin's action. Similarly, metformin can directly slow down the abundance of *E. coli*, restraining to the intestinal bacteria's carcinogenic effects. On the other hand, metformin may also promote some floras that are beneficial for anticancer effects to become a dominant flora. This is consistent with the changes in gut microbiota caused by the metformin intervention mentioned in Table 3.

Role of polyamine metabolism system in the inhibition of CRC by metformin in the presence of gut microbiota

Polyamines are a class of compounds containing two or more amino groups. The raw materials for their synthesis are ornithine and arginine. The key enzymes are ornithine decarboxylase and arginine decarboxylase. It has been proven that they can regulate cell proliferation and apoptosis, and their biosynthesis is closely related to the formation and metastasis of cancer^[85]. There is a correlation between the expression of ornithine decarboxylase and the prediction of cancer risk and treatment response in certain epithelial cancers. However, it must be pointed out that the function of the polyamine system is affected by metformin in the presence of several pathogens, including *Shigella flexneri*, *Streptococcus pneumoniae*, *Salmonella enterica* subsp., *enterica* serovar *Typhimurium*, and *Helicobacter pylori*^[86]. Therefore, metformin may inhibit the development of CRC by regulating the polyamine system under the action of certain gut microbiota.

Agmatine, a biogenic amine, is a polyamine that is converted from arginine by the action of arginine decarboxylase on the mitochondrial membrane of cells, and it has vasomotor implication and regulation of anti-inflammation^[87]. Interestingly, the ability of bacteria to produce agmatine was enhanced under the administration of metformin in T2DM in a nutrient-dependent manner^[43]. In addition, it must be emphasized that agmatine has a directly inhibitory effect on the proliferation of tumor cells with the corresponding acceleration of apoptosis^[88]. As for CRC, agmatine has been confirmed

to inhibit the proliferation of six types of human intestinal tumor cell lines in a concentration-dependent manner^[89]. It is able to arrest proliferation in cell lines by depleting intracellular polyamine levels and enter mammalian cells *via* the polyamine transport system.

To date, there are few reports on the use of agmatine metabolism as a direct therapeutic target in clinical research, but there are studies affirming that the agmatine metabolism system has the potential to treat tumors. The role of metformin and gut microbiota is like a fuse in the antitumor effect mediated by the agmatine metabolic system. Moreover, Pryor *et al.*^[43] suggested that the *Escherichia*, *Bacteroides*, *Enterobacter*, and *Citrobacter* genera produce the most agmatine in the intestine, and these bacteria were abundant in the gut of patients after metformin intervention. Furthermore, this inhibitory effect of agmatine on tumors was probably attributable to an interaction between agmatine and the intracellular polyamine metabolism system^[90,91]. For example, agmatine regulates the functional gene delivery system of spermidine/spermine acetyltransferase in cancer cell^[90]. Therefore, the exploration of a metformin-gut microbiota-cell polyamine metabolism system-cancer chain helps us to understand how metformin inhibits CRC by regulating the agmatine metabolism system in the presence of gut microbiota. From the perspective of drugs-gut microbiota-pharmacological effects-diseases, this will be an interesting and profound finding.

To sum up, metformin may indirectly restrain clinical colorectal tumors by regulating the agmatine metabolism system in the presence of gut microbiota. In future studies, more attention should be attached to the drug's interaction with gut microbiota to the intracellular polyamine system because the intervention of the polyamine system may be one of the important methods for clinical cancerous treatment.

FUTURE EXPECTATIONS

One clear goal moving forward is to explore how to better apply the gut microbiota in clinical treatment. Gut microbiota may directly induce metformin's anti-CRC effect *via* the structure of its own composition, the production of secondary metabolites, and the regulation of the immune system after interplay with metformin. As a consequence, accumulated information has proven that metformin influences colorectal tissue carcinogenesis, and this effect is dependent on gut microbiota. Then we need to figure it out that what types of gut microbiota play key roles in this process.

Herein, we advocate to build a complete network of relationships of drugs-gut microbiota-diseases. This concept will provide a new treatment strategy for the diagnosis and treatment of clinical diseases by using gut microbiota. However, in order to achieve this goal, the relationship between changes in the structure and function of the gut microbiota and the pharmacological effects of drugs needs to be thoroughly explained. For example, composition of gut microbiota is a determinant for development of gastrointestinal adverse effects following metformin intake^[32]. Wu *et al.*^[92] investigated the pharmacodynamic and pharmacokinetic effects of metformin mediated by the gut microbiota *in vivo*. The pharmacodynamic indexes were evaluated, and metformin concentrations were measured with a validated liquid chromatography-tandem mass spectrometry method after oral administration. They described the pharmacodynamic differences between metformin in sterile diabetic rats and conventional diabetic rats. Compared to conventional diabetic rats, fasting blood glucose and *c_{max}* in pseudosterile diabetic rats were significantly increased with a corresponding reduction in oral glucose and *t_{1/2α}*. All of these elaborated that these differences in pharmacodynamics and pharmacokinetics may be due to the decrease in the expression of *Oct1* in the liver, contributing to the changes in liver uptake of metformin^[92]. Napolitano *et al.*^[93] suggested that metformin has complex effects that alter bile acid recycling and gut microbiota (positive correlation between the microbiota abundance of the phylum *Firmicutes* and changes in cholic acid and conjugates, negative correlation in *Bacteroidetes*) due to gut-based pharmacology, which might provide insights into novel therapeutic approaches to treat T2DM and associated metabolic diseases. But the explanation of these pharmacodynamics and pharmacokinetic mechanisms is not enough for the determination of clinical drugs. In addition, the current majority of research focuses on the relationship between the changes in the profiles of gut microbiota and the effects of drugs on diseases (*e.g.*, metformin on CRC).

To date, the research in respect to the role of metformin to promote the formation of CRC in T2DM were mainly carried out in the observational studies of populations. In

future work, more experiments need to be conducted to explore deeper mechanisms in individuals and in animal models, which can also verify the effect of an anti-hyperglycemia pharmaceutical for anticancer treatment. Furthermore, metformin, a biguanide derivative, has pleiotropic effects beyond glucose reduction, including antitumor, antihaze-induced pneumonia, and anti-aging. Other pharmacological effects and corresponding mechanisms of metformin need to be further discovered. For instance, we elaborated that metformin elicits its role in tumors *via* regulating the function of the immune system and how the effect is performed in the presence of gut microbiota based on the fact that the occurrence of tumors is closely related to the body's immunity.

Overall, CRC is an urgent research topic undertaken by many researchers all over the world. Nevertheless, the diagnosis and treatment of CRC still has many unresolved issues, but the research on the gut microbiota has opened a new window. Through this window, we could see the emergence of gut microbiota as a novel way to fight tumors. However, the current research is limited to the changes in its quantity and distribution due to the limitations of current scientific research technologies and methods. When metformin is regarded as an anticancer drug in the intestine, the intestinal microbiota will change or affect the production of certain anticancer substances. Hence, it must be stressed again that establishing a complete drug-gut microbiota-disease relationship network is necessary for the potential medical application of gut microbiota. In addition, novel methods for microbiological research should be systematically carried out in order to explore the function and structure of gut microbiota in the future: structure functions, genomics, transcriptomics, proteomics, and metabolomics. These efforts will benefit the prevention, diagnosis, and treatment of clinical CRC. Advances in this field will also promote the progress of other microbial-related diseases and provide new ideas for the use of clinical drugs.

CONCLUSION

In summary, metformin can inhibit the initiation of CRC in T2DM patients by changing the abundance of gut microbiota or under the participation of gut microbiota. Further research is needed to confirm this hypothesis and explore the mechanisms behind it. Moreover, in order to understand thoroughly the function and structure of gut microbiota, more cutting-edge scientific research methods and technologies are needed to implement.

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