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Contents

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REVIEW

- 1 Perspectives of pluripotent stem cells in livestock
Kumar D, Talluri TR, Selokar NL, Hyder I, Kues WA
- 30 Adipose-derived stem cells: Pathophysiologic implications *vs* therapeutic potential in systemic sclerosis
Rosa I, Romano E, Fioretto BS, Matucci-Cerinic M, Manetti M
- 49 Mesenchymal stem cell-derived small extracellular vesicles in the treatment of human diseases: Progress and prospect
Shi J, Zhao YC, Niu ZF, Fan HJ, Hou SK, Guo XQ, Sang L, Lv Q

MINIREVIEWS

- 64 Pancreatic β cell regeneration induced by clinical and preclinical agents
Wang KL, Tao M, Wei TJ, Wei R

ORIGINAL ARTICLE

Basic Study

- 78 Stem cell transplantation and/or adenoviral glial cell line-derived neurotrophic factor promote functional recovery in hemiparkinsonian rats
Tsai MJ, Hung SC, Weng CF, Fan SF, Liou DY, Huang WC, Liu KD, Cheng H
- 91 Vascularization and osteogenesis in ectopically implanted bone tissue-engineered constructs with endothelial and osteogenic differentiated adipose-derived stem cells
Najdanović JG, Cvetković VJ, Stojanović ST, Vukelić-Nikolić MD, Živković JM, Najman SJ
- 115 Proliferation and tenogenic differentiation of bone marrow mesenchymal stem cells in a porous collagen sponge scaffold
Zhang BY, Xu P, Luo Q, Song GB
- 128 Transcriptomic alterations underline aging of osteogenic bone marrow stromal cells
Cheng YH, Liu SF, Dong JC, Bian Q

ABOUT COVER

Editorial Board Member of *World Journal of Stem Cells*, Dr. Yong-Can Huang received his Bachelor's and Master's degrees from Sichuan University (China) in 2007 and 2010, respectively; in 2015, he obtained his PhD from the University of Hong Kong. Currently, he is Associate Professor at Peking University Shenzhen Hospital in Shenzhen (China). His major research interests include bone marrow- and placenta-derived stem cells for musculoskeletal regeneration, and intervertebral disc transplantation. Dr. Huang is member of the International Society of Orthopaedic Surgery and Traumatology, the International Chinese Musculoskeletal Research Society, and the Tissue Engineering and Regenerative Medicine International Society. (L-Editor: Filipodia)

AIMS AND SCOPE

The primary aim of *World Journal of Stem Cells* (*WJSC*, *World J Stem Cells*) is to provide scholars and readers from various fields of stem cells with a platform to publish high-quality basic and clinical research articles and communicate their research findings online. *WJSC* publishes articles reporting research results obtained in the field of stem cell biology and regenerative medicine, related to the wide range of stem cells including embryonic stem cells, germline stem cells, tissue-specific stem cells, adult stem cells, mesenchymal stromal cells, induced pluripotent stem cells, embryonal carcinoma stem cells, hemangioblasts, lymphoid progenitor cells, *etc.*

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Basic Study

Proliferation and tenogenic differentiation of bone marrow mesenchymal stem cells in a porous collagen sponge scaffold

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Abstract

BACKGROUND

Collagen is one of the most commonly used natural biomaterials for tendon tissue engineering. One of the possible practical ways to further enhance tendon repair is to combine a porous collagen sponge scaffold with a suitable growth factor or cytokine that has an inherent ability to promote the recruitment, proliferation, and tenogenic differentiation of cells. However, there is an incomplete understanding of which growth factors are sufficient and optimal for the tenogenic differentiation of rat bone marrow mesenchymal stem cells (BMSCs) in a collagen sponge-based 3D culture system.

AIM

To identify one or more ideal growth factors that benefit the proliferation and tenogenic differentiation of rat BMSCs in a porous collagen sponge scaffold.

METHODS

We constructed a 3D culture system based on a type I collagen sponge scaffold. The surface topography of the collagen sponge scaffold was observed by scanning electron microscopy. Primary BMSCs were isolated from Sprague-Dawley rats. Cell survival on the surfaces of the scaffolds with different growth factors was assessed by live/dead assay and CCK-8 assay. The mRNA and protein expression levels were confirmed by quantitative real-time polymerase chain reaction and Western blot, respectively. The deposited collagen was assessed by Sirius Red staining.

RESULTS

Transforming growth factor β 1 (TGF- β 1) showed great promise in the tenogenic differentiation of BMSCs compared to growth differentiation factor 7 (GDF-7) and

CQUPT2018016).

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insulin-like growth factor 1 (IGF-1) in both the 2D and 3D cultures, and the 3D culture enhanced the differentiation of BMSCs into tenocytes well beyond the level of induction in the 2D culture after TGF- β 1 treatment. In the 2D culture, the proliferation of the BMSCs showed no significant changes compared to the control group after TGF- β 1, IGF-1, or GDF-7 treatment. However, TGF- β 1 and GDF-7 could increase the cell proliferation in the 3D culture. Strangely, we also found more dead cells in the BMSC-collagen sponge constructs that were treated with TGF- β 1. Moreover, TGF- β 1 promoted more collagen deposition in both the 2D and 3D cultures.

CONCLUSION

Collagen sponge-based 3D culture with TGF- β 1 enhances the responsiveness of the proliferation and tenogenic differentiation of rat BMSCs.

Key Words: Bone marrow mesenchymal stem cells; Collagen sponge; Transforming growth factor β 1; Tenogenic differentiation; Proliferation; Collagen deposition

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Core Tip: Growth factor supplementation of stem cells facilitates cell differentiation and/or cell growth, but there is an incomplete understanding of which growth factors are optimal in 3D culture. We compared different growth factors for the proliferation and tenogenic differentiation of bone marrow mesenchymal stem cells (BMSCs) in a monolayer culture (2D) and in a collagen sponge-based 3D culture. We found that transforming growth factor β 1 (TGF- β 1) showed great promise in the tenogenic differentiation of BMSCs compared to growth differentiation factor 7 and insulin-like growth factor 1 in both 2D and 3D cultures, and the 3D culture enhanced the tenogenic differentiation of BMSCs well beyond the level of induction in the 2D culture after TGF- β 1 treatment.

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INTRODUCTION

Tendon is a tough band of fibrous connective tissue that connects muscle to bone and is capable of withstanding cyclic tension. In daily life, excessive physical training, heavy physical labor, or an unexpected accident often leads to tendon injury. The therapeutic strategies for the treatment of injured tendons usually can be broadly classified into nonsurgical (conservative) and surgical. Conservative treatments are usually used for chronic tendon injuries through physical or chemical means and those that need to take a long time to repair^[1]. Surgical treatments are used for acute tendon rupture where damaged tendons are sutured together with or without the use of autografts, allografts, xenografts, or prosthetic devices^[2]. Nonetheless, there are some drawbacks that seem difficult to overcome in surgical treatments, such as immunological rejection, donor site morbidity, and the risk of disease transmission^[3].

With the development of cell culture techniques, medical biomaterials, and implantation techniques, tissue engineering techniques show great potential for tendon repair and regeneration^[4,5]. Along with scaffolds, the use of growth factors for the differentiation of stem/progenitor cells towards the tenogenic lineage before transplantation is considered a very crucial step in tendon tissue engineering^[6]. A growth factor is a naturally occurring substance capable of regulating cellular growth, differentiation, and production of the extracellular matrix (ECM)^[7]. In recent decades, extensive studies have demonstrated that growth factors, including transforming growth factor (TGF)- β family members (TGF- β 1, TGF- β 2, TGF- β 3, *etc.*)^[8,9], growth differentiation factor (GDF) family members (GDF-5, GDF-6, and GDF-7)^[10-12], basic



fibroblast growth factor (bFGF)^[13], and connective tissue growth factor^[14,15], can promote the tenogenic differentiation of stem/progenitor cells in a 2D culture. However, using tissue engineering techniques to restore the injured tendon requires a 3D scaffold incorporated with cells to form a functional tendon construct *in vitro*. The interaction between cells, the transmission of nutrients/growth factors/cytokines, and the combination of cells and matrix are clearly different in a 3D culture environment than in a 2D culture^[16]. Furthermore, there is a considerable variety of scaffold materials that have different origins and diverse properties. Therefore, the ideal growth factors for tenogenic differentiation of stem/progenitor cells in different scaffolds still need to be identified.

Type I collagen is a popular material used for tendon tissue engineering due to its ease of extraction, and it is the main component of normal tendon^[17,18]. Insoluble collagen can be processed into various forms, such as tubes, sponges, films, and gels, and the form of sponge is the most commonly used. Although the mechanical strength of collagen sponge scaffolds is not as good as synthetic polymeric scaffolds, it possesses excellent biocompatibility and biodegradability. However, there is an incomplete understanding of which growth factors are sufficient and optimal in a type I collagen sponge-based 3D culture.

In this study, we constructed a 3D culture system based on a type I collagen sponge scaffold. We compared different growth factors for the proliferation and the tenogenic differentiation of rat bone marrow mesenchymal stem cells (BMSCs) in a monolayer culture and in a collagen sponge-based 3D culture under the same conditions. Our ultimate goal was to confirm one or more ideal growth factors that benefit the construction of the tissue-engineered tendon.

MATERIALS AND METHODS

Cell isolation and cultivation

Primary BMSCs from Sprague-Dawley rats (either gender, 4 wk old, weight 130 ± 10 g, from the Laboratory Animal Center, Chongqing Medical University, China) were explanted and cultured according to our previous study^[19]. All of the procedures were approved by the Chongqing Science and Technology Commission, Chongqing, China, and performed in accordance with the United States National Institutes of Health (NIH, 8th edition, 2011). Briefly, the femurs and tibias from rats were sawed open, and the gelatinous bone marrow was extracted under sterile conditions. Rat BMSCs were obtained by density gradient centrifugation with 1.073 g/mL Percoll (Sigma-Aldrich, Saint Louis, MO, United States) at 2500 rpm/min for 30 min and then maintained in Dulbecco's modified Eagle's medium (DMEM) (Dingguo Biotech, Beijing, China) supplemented with 10% fetal bovine serum (FBS; HyClone, Logan, UT, United States), 100 U/mL penicillin, and 100 μ g/mL streptomycin in a standard incubator under a humidified atmosphere of 5% CO₂ and 95% air at 37 °C. The culture medium was replaced every 3 d. Subculturing was performed when the cells achieved an 80% confluency. Cells at passages 3-5 were used for the experiments.

Construct preparation

Type I collagen medical sponge (Kejibang, Wuxi, China) was used in our study to create constructs (Figure 1). First, the collagen sponge was rinsed three times with phosphate-buffered saline (PBS) and steeped with 15% FBS for 1 h. The cells were then seeded at a density of 1×10^5 cells/cm² into the type I collagen sponge and allowed to adhere, infiltrate, and proliferate for 3 d. After 3 d of incubation, the BMSC-collagen sponge constructs were cultured in DMEM without FBS for 12 h to synchronize the BMSCs. Finally, the BMSC-collagen sponge constructs were exposed to TGF- β 1 (10 ng/mL), insulin-like growth factor 1 (IGF-1, 10 ng/mL), or GDF-7 (100 ng/mL) (PeproTech, NJ, United States) for 3 or 12 d. As controls, the cells were cultured in a collagen sponge under the same conditions but were not subjected to any stimulation.

Cell survival assay

Cell survival on the surfaces of scaffolds with different growth factors was assessed by live/dead assay (Calcein-AM/PI Double Stain Kit, YEASEN, Shanghai, China). Briefly, after exposure to TGF- β 1 (10 ng/mL), IGF-1 (10 ng/mL), or GDF-7 (100 ng/mL) (PeproTech, NJ, United States) for 12 d, the BMSC-collagen sponge constructs were harvested and incubated with 2 μ mol/L calcein AM and 4.5 μ mol/L propidium iodide (PI) for 30 min in the dark at 37 °C. After rinsing three times with PBS, the fluorescence was visualized using a fluorescence microscope (Olympus, Japan).

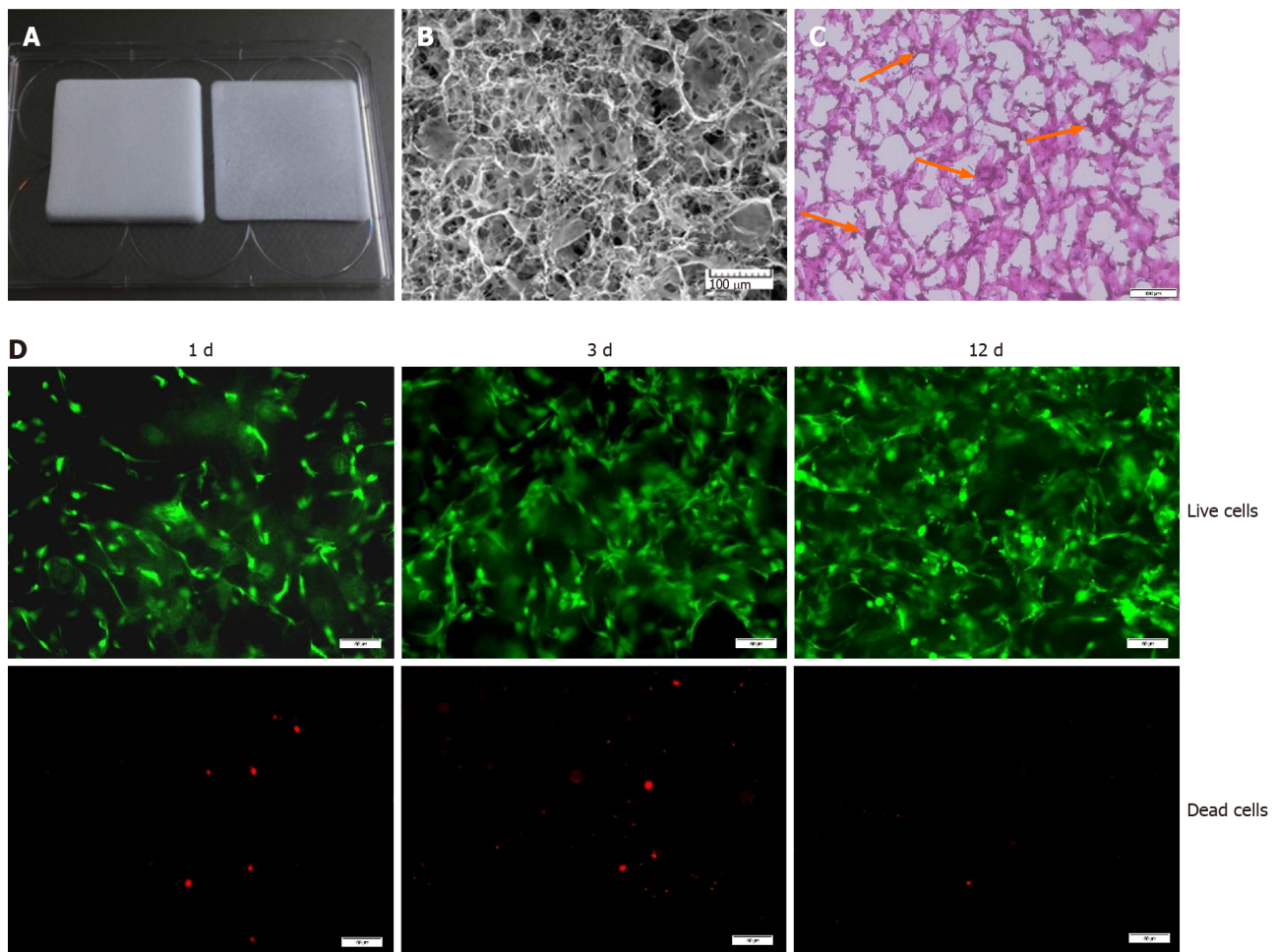


Figure 1 A 3D culture system based on a type I collagen sponge scaffold. A: Type I collagen sponge scaffold; B: Surface topography of the collagen sponge scaffold observed by scanning electron microscopy. Bars = 100 µm; C: Hematoxylin and eosin staining of sectioned bone marrow mesenchymal stem cell-collagen sponge constructs. Orange arrows indicate the cells. Bars = 100 µm; D: Live/dead assay of the scaffolds during the cultivation process. Green indicates the live cells, and red indicates the dead cells. Bars = 100 µm.

CCK-8 assay

A CCK-8 assay was performed to evaluate the overall viability of the cells according to the manufacturer's instructions. For the 2D culture, the BMSCs were seeded at a density of 3×10^3 cells/well in a 96-well culture plate. After adherence, the cells were serum-starved overnight and treated with growth factors for 3 or 12 d, after which CCK-8 (0.5 mg/mL, Sigma-Aldrich, St. Louis, MO, United States) was added into the 96-well culture plate and incubated at 37 °C for 2 h. For the 3D culture, after exposure to the growth factors, the culture medium was replaced with a serum-free culture medium containing CCK-8. After 2 h of culturing, 100 µL of culture was taken from each well and then transferred to a new 96-well plate. The luminescence was then quantified using a microplate reader (Model 680, Bio-Rad, Hercules, CA, United States). The relative cell viability was determined by normalizing the optical density (OD) value of the experimental group to that of the control group.

RNA preparation and real-time polymerase chain reaction

For the 3D culture, after exposure to the growth factors, the BMSC-collagen sponge constructs were digested with collagenase (1 mg/mL; Solarbio, Beijing, China) for 15 min at 37 °C. The BMSCs were then harvested by centrifugation. Total RNA was isolated from the BMSCs using the RNeasy Mini Kit (BioTeke, Beijing, China) according to the manufacturer's instructions. Total RNA (500 ng) was reverse transcribed in a 20 µL system using a reverse transcriptase kit (Takara, Japan). Quantitative real-time polymerase chain reaction (qRT-PCR) analysis of eight genes, namely, *Col1a1*, *Col3a1*, *Tnc*, *Scx*, *Tnmd*, *PPARγ*, *Runx2*, and *GAPDH*, was performed with a CFX96 Touch RT-PCR System (Bio-Rad, Hercules, CA, United States) using a reagent kit (Takara, Japan). The primer sequences used in this study are listed in

Table 1. The results were analyzed using CFX Manager Software (Bio-Rad, Hercules, CA, United States). *GAPDH* was used as an endogenous reference gene. The data are presented as the fold change relative to the expression level of the control (untreated BMSCs).

Western blot analysis

After exposure to TGF- β 1 (10 ng/mL), IGF-1 (10 ng/mL), or GDF-7 (100 ng/mL) for 12 d in the 2D and 3D cultures, the BMSCs were harvested and washed with ice-cold PBS. The proteins were extracted using cell lysis buffer. The protein concentration was determined by Lowry's method using bovine serum albumin as a standard. After electrophoretic separation by SDS-polyacrylamide gel electrophoresis, the proteins were electrotransferred onto polyvinylidene fluoride membranes (Millipore, Billerica, MA, United States). The membranes were then blocked for 1 h at room temperature in Tris-buffered saline containing 0.1% Tween-20 (TBST) and 5% skim milk. Col1a1 goat pAb (Santa Cruz, CA, United States), Col3a1 mouse mAb (Santa Cruz, CA, United States), Tnc rabbit mAb (Cell Signaling Technology, Danvers, MA, United States), Scx rabbit pAb (Abcam, Cambridge, MA, United States), Tnmd rabbit pAb (Abcam, Cambridge, MA, United States), and GAPDH mouse mAb (ZSGB-Bio, Beijing, China) were used according to the manufacturers' protocols, and the membranes were incubated overnight with these antibodies at 4 °C with slight shaking. The membranes were then washed three times in TBST and further incubated with an HRP-conjugated antibody (MultiSciences, Beijing, China) for 1 h at room temperature. Finally, signal detection was performed with an enhanced chemiluminescence kit (Thermo, Waltham, MA, United States). A semiquantitative evaluation of the bands was performed by densitometry (VersaDoc, Bio-Rad, Hercules, CA, United States). The protein levels of Col1a1, Col3a1, Tnc, Scx, and Tnmd were determined through their normalization to GAPDH.

Scanning electron microscopy

The BMSC-collagen sponge constructs were collected, washed twice in PBS, fixed with 2.5% glutaraldehyde in PBS for 4 h at room temperature, rinsed in PBS, and frozen at -80 °C overnight. After lyophilization, the samples were fixed to the sample stage, sputter-coated with platinum for 20 s, and observed by scanning electron microscopy (SEM) (VEGA3, TESCAN, Brno, Czech).

Sirius Red staining

The BMSCs were seeded at a density of 5×10^4 cells/well in a 6-well culture plate. After adherence, the cells were serum-starved overnight and treated with growth factors for 12 d. The conditioned medium was then removed, and the cells were washed with PBS. Before Sirius Red staining, the cells were fixed with 70% ethanol for 30 min and washed with PBS three times. The deposited collagen was assessed by Sirius Red staining (Solarbio, Beijing, China) according to the manufacturer's instructions. Finally, the samples were photographed using a microscope (Olympus, Japan).

Histological analysis

Histological analyses were performed to investigate the cell distribution and collagen deposition within the constructs. The BMSC-collagen sponge constructs were quick-frozen, embedded in tissue freezing medium (ZSGB-Bio, Beijing, China), and sectioned into 8 μ m-thick transverse sections using a freezing microtome (CM1900, Leica, Solms, Germany). The samples were then stained using hematoxylin and eosin (H&E, Solarbio, Beijing, China) and Masson's trichrome (MT, Sigma-Aldrich, Saint Louis, MO, United States) according to the manufacturer's instructions. Finally, the samples were photographed using a microscope (Olympus, Japan).

Statistical methods

All quantitative data are expressed as the mean \pm SD. The Student's *t*-test was performed to analyze the differences between two groups. Bonferroni post hoc tests were used when the *P* value indicated a significant difference between the groups. *P* < 0.05 was deemed to be statistically significant.

Table 1 Primer sequences utilized for real-time polymerase chain reaction

Gene	5'-3'	Primer sequence	Product size (bp)	Reference sequence
<i>Col1a1</i>	Forward Reverse	GGAGAGAGTGCCAACTCCAG; GTGCTTTGGAAAATGGTGCT	182	NM_053304.1
<i>Col3a1</i>	Forward Reverse	TCCCAGAACATTACATACCACT; GCTATTTCCTTCAGCCTTGA	126	NM_032085.1
<i>Tnc</i>	Forward Reverse	AGATGCTACTCCAGACGGTTTC; CACGGCTTATTCCATAGAGTTCA	200	NM_053861.1
<i>Scx</i>	Forward Reverse	AACACGGCCTTCACTGCGCTG; CAGTAGCACGTGCCCAGGTG	123	NM_001130508.1
<i>Tnmd</i>	Forward Reverse	GTGGTCCCACAAGTGAAGGT; GTCTTCCTCGCTTGCTTGTC	60	NM_022290.1
<i>PPARγ</i>	Forward Reverse	CAGGCTTGCTGAACGTGAAG; ACGTGCTCTGTGACAATCTGC	157	NM_001145366.1
<i>Runx2</i>	Forward Reverse	CAGACCAGCAGCACTCCATA; CAGCGTCAACACCATCATTC	178	NM_001278483.1
<i>GAPDH</i>	Forward Reverse	AAGTTCAACGGCACAGTCAAGG; CGCCAGTAGACTCCACGACATA	139	NM_017008.4

RESULTS

Collagen sponge-based 3D cell culture system

In our study, we used a medical type I collagen sponge to construct a 3D culture system. The surface topography of the collagen sponge scaffold was observed by SEM. As shown in [Figure 1B](#), the collagen sponge scaffold was isotropic, with a homogeneous distribution of rounded pores within the scaffold. H&E staining showed that many cells could adhere to the surface of the material ([Figure 1C](#)). Furthermore, the live/dead assay showed that the collagen sponge scaffold had a good biocompatibility. The collagen sponge scaffold allowed the adhesion and spreading of cells and only a few dead cells were observed on the surfaces of the scaffold after 12 d of culture ([Figure 1D](#)).

Effect of different growth factors on tenogenic differentiation of BMSCs in the 2D and 3D cultures

The concentrations of growth factors were chosen based on published studies and on our screening results^[8,20]. As shown in [Figure 2](#), TGF- β 1 increased *Col1a1*, *Col3a1*, *Tnc*, *Scx*, and *Tnmd* mRNA expression levels compared to those of the other groups in the both 2D and 3D cultures. IGF-1 and GDF-7 had no significant effect on *Col1a1*, *Col3a1*, *Tnc*, *Scx*, and *Tnmd* mRNA expression levels within a short time (day 3) in both the 2D and 3D cultures. Similar to expression at the mRNA levels, the protein levels of *Col1a1*, *Col3a1*, *Tnc*, and *Tnmd* were all significantly upregulated in cells treated with TGF- β 1 for 12 d in 3D culture ([Figure 3](#)). Moreover, we also detected the effect of different growth factors on the mRNA expression levels of *PPAR γ* (adipogenic marker) and *Runx2* (osteoblast-specific marker). The results showed that TGF- β 1 could efficiently inhibit the mRNA expression level of *PPAR γ* in both the 2D and 3D cultures ([Figure 2F](#)). TGF- β 1 had no effect on the mRNA expression of *Runx2* in the 2D culture but could inhibit the mRNA expression of *Runx2* in the 3D culture. The inhibitory effect gradually disappeared with the extended culture time ([Figure 2G](#)).

Effect of different growth factors on collagen synthesis in BMSCs in the 2D and 3D cultures

The expression levels of *Col1a1* and *Col3a1*, confirmed by qRT-PCR and western blotting, in the TGF- β 1-treated group were significantly higher than those in the other groups in both the 2D and 3D cultures ([Figures 2 and 3](#)). Moreover, Sirius Red staining showed that more collagen fibers were formed in the TGF- β 1-treated group compared to the other groups in the 2D culture, and MT staining showed that more collagen fibers were formed in the TGF- β 1-treated group compared to the other groups in the 3D culture ([Figure 4](#)).

Effect of different growth factors on proliferation of BMSCs in the 2D and 3D cultures

In the 2D culture, the viability of BMSCs showed no significant changes compared to that of the control group in 3 d after TGF- β 1, IGF-1, or GDF-7 treatment, but the viability of the TGF- β 1-treated BMSCs was reduced with the culture time extension (day 12) ([Figure 5](#)). In the 3D culture, the results also showed that TGF- β 1, IGF-1, or GDF-7 had no significant effect on the proliferation of the BMSCs in a relatively short

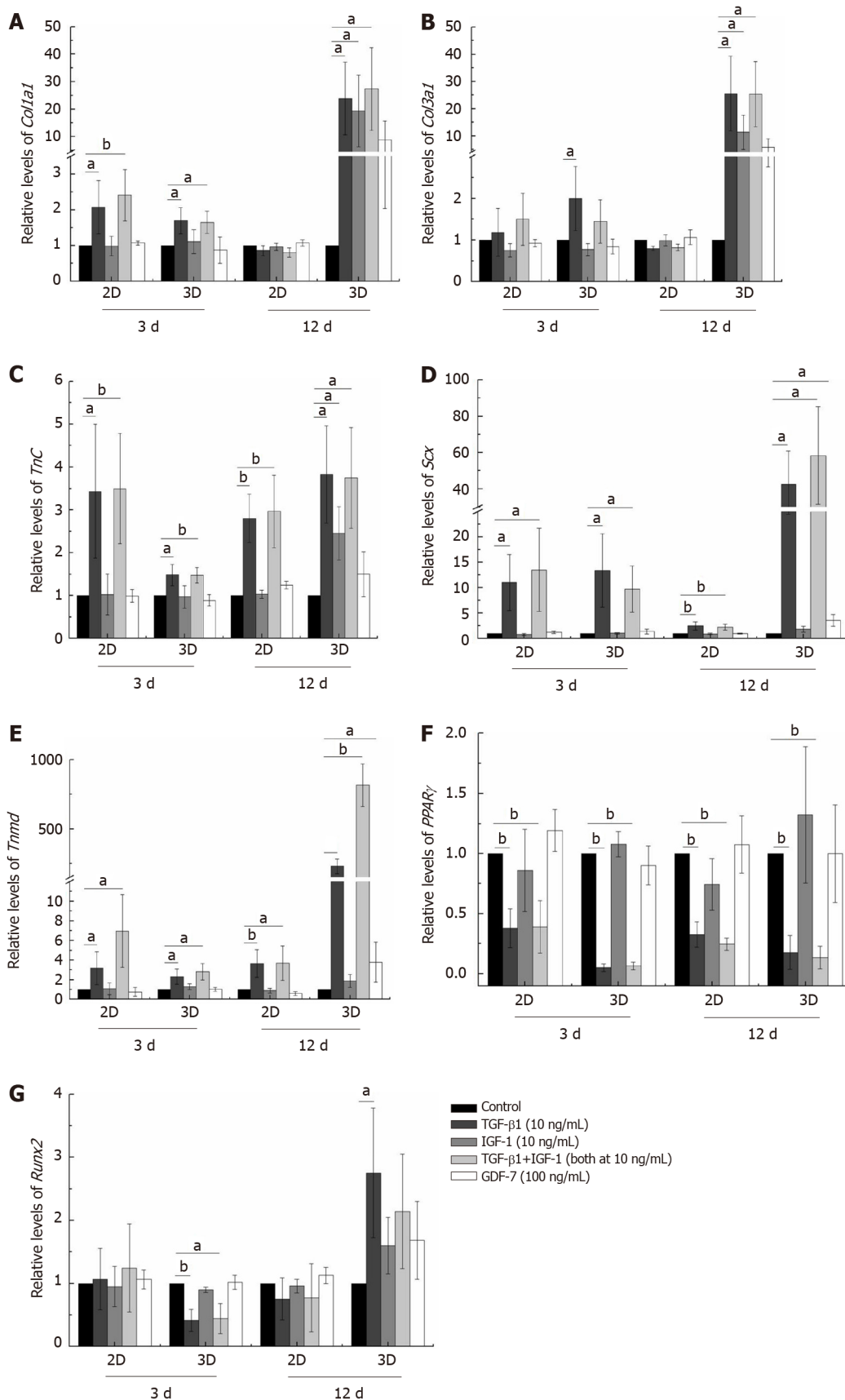


Figure 2 Real-time PCR analysis of gene expression levels after stimulation with transforming growth factor beta 1 (10 ng/mL), insulin-like growth factor 1 (10 ng/mL), growth differentiation factor 7 (100 ng/mL), or transforming growth factor beta 1 plus insulin-like growth factor 1 (both at 10 ng/mL) for 3 or 12 d in the 2D and 3D cultures. A: *Col1a1*; B: *Col3a1*; C: *Tnc*; D: *Scx*; E: *Tnmd*; F: *PPARγ*; G: *Runx2*. The results of the target gene expression levels were normalized to those of *GAPDH*. The data are expressed as the mean \pm SD; $n = 3$. ^a $P < 0.05$; ^b $P < 0.01$. TGF-β1: Transforming growth factor beta 1; IGF-1: Insulin-like growth factor 1; GDF-7: Growth differentiation factor 7.

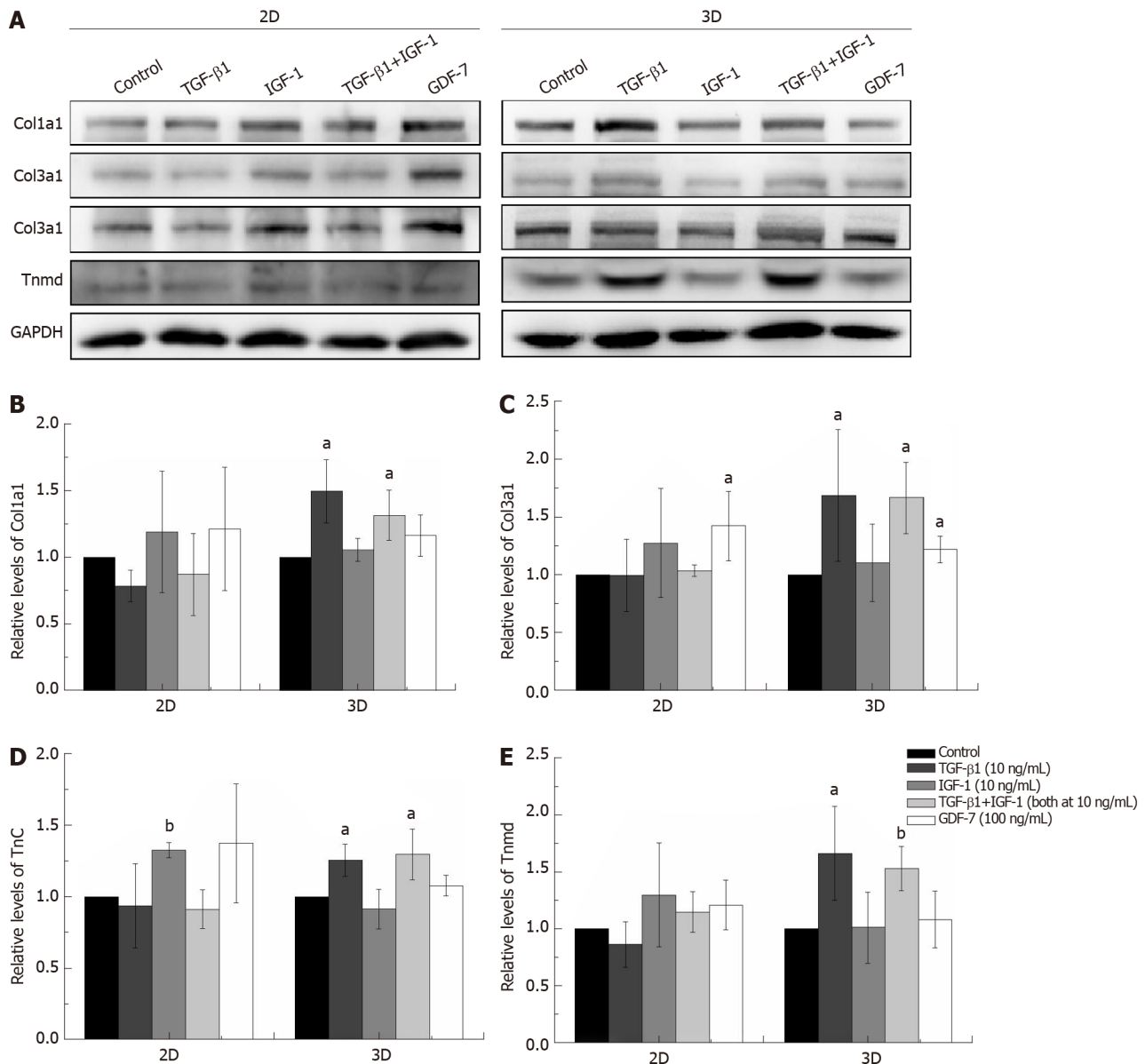


Figure 3 Effect of stimulation with transforming growth factor $\beta 1$ (10 ng/mL), insulin-like growth factor 1 (10 ng/mL), growth differentiation factor 7 (100 ng/mL), or transforming growth factor beta 1 plus insulin-like growth factor 1 (both at 10 ng/mL) on the differentiation of bone marrow mesenchymal stem cells in the 2D and 3D cultures. A: The protein expression levels of Col1a1, Col3a1, Tnc, and Tnmd detected by Western blot after growth factor stimulation for 12 d; B-E: The results of the densitometric analysis of the target protein expression levels were normalized to those of GAPDH. The data are expressed as the mean \pm SD; $n = 3$. ^a $P < 0.05$; ^b $P < 0.01$. TGF- $\beta 1$: Transforming growth factor $\beta 1$; IGF-1: Insulin-like growth factor 1; GDF-7: Growth differentiation factor 7.

time (day 3). However, TGF- $\beta 1$ and GDF-7 could promote the cell proliferation in 12 d. Strangely, we also found more dead cells in the 3D culture where cells were treated with TGF- $\beta 1$ (Figure 6).

DISCUSSION

Growth factor supplementation of MSCs facilitates tenogenic differentiation, but there is an incomplete understanding of which growth factors are sufficient and optimal. In this study, we investigated the proliferation of rat BMSCs after treatment with different growth factors in a monolayer culture and in a collagen sponge-based 3D culture. In the 2D culture, TGF- $\beta 1$, IGF-1, or GDF-7 all showed no significant effect on the proliferation of BMSCs in a short time (day 3), and TGF- $\beta 1$ even inhibited cell growth with the extension of culture time. Interestingly, in the 3D culture, the results also showed that TGF- $\beta 1$, IGF-1, or GDF-7 had no effect on the proliferation of the

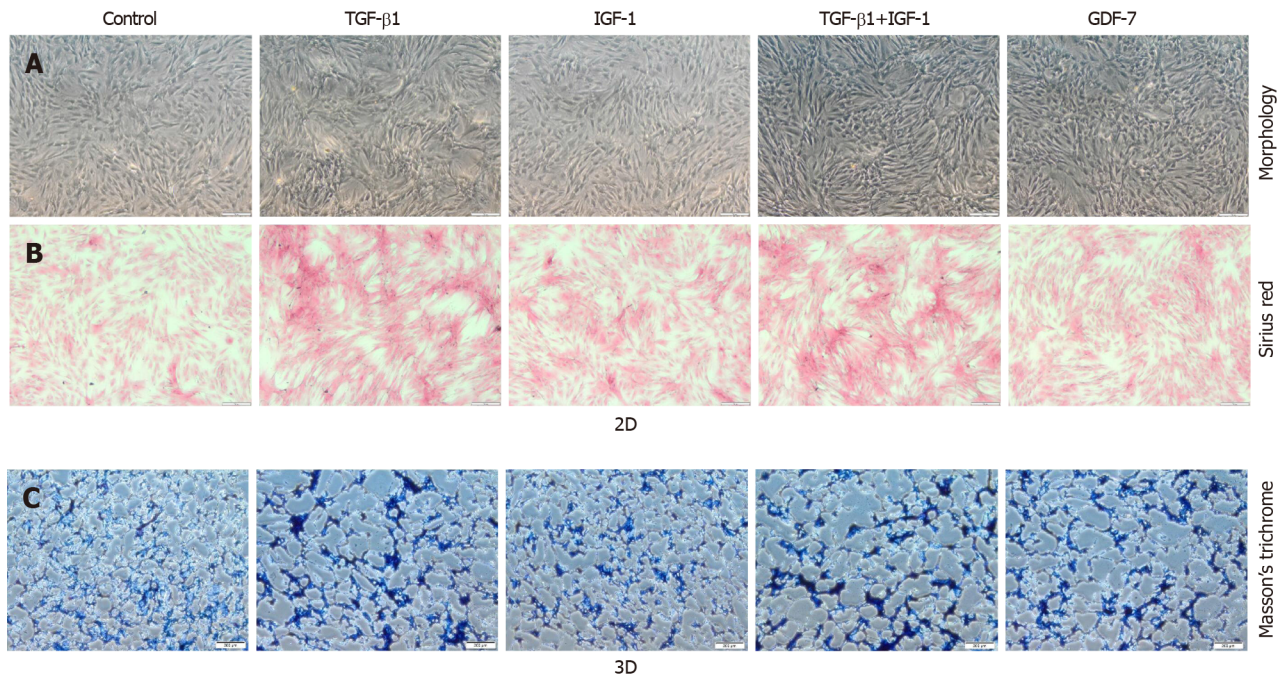


Figure 4 Effect of transforming growth factor $\beta 1$ (10 ng/mL), insulin-like growth factor 1 (10 ng/mL), growth differentiation factor 7 (100 ng/mL), and transforming growth factor $\beta 1$ or insulin-like growth factor 1 (both at 10 ng/mL) on collagen synthesis in the bone marrow mesenchymal stem cells in the 2D and 3D cultures. A: Morphologic appearance of the bone marrow mesenchymal stem cells (BMSCs) in culture supplemented with various growth factors at day 3; B: Sirius Red staining for collagen deposition evaluation in the BMSCs cultured in the absence of growth factor (control) or the presence of growth factors at day 3. Scale bars = 200 μ m; C: Masson's trichrome staining of sectioned BMSC-collagen sponge constructs after growth factor stimulation for 3 d. Bars = 200 μ m. TGF- $\beta 1$: Transforming growth factor $\beta 1$; IGF-1: Insulin-like growth factor 1; GDF-7: Growth differentiation factor 7.

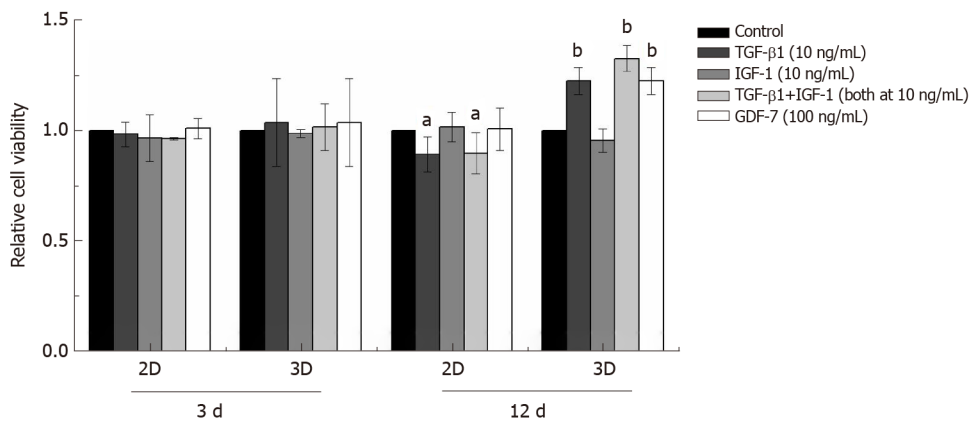


Figure 5 Effect of transforming growth factor $\beta 1$ (10 ng/mL), insulin-like growth factor 1 (10 ng/mL), growth differentiation factor 7 (100 ng/mL), or transforming growth factor $\beta 1$ combined with insulin-like growth factor 1 (both at 10 ng/mL) on the viability of bone marrow mesenchymal stem cells in the 2D and 3D cultures. The viability of all the bone marrow mesenchymal stem cells was evaluated by CCK-8 assay after growth factor stimulation for 3 or 12 d. The data are expressed as the mean \pm SD; $n = 3$. ^a $P < 0.05$ and ^b $P < 0.01$ compared with the control (non-treated). TGF- $\beta 1$: Transforming growth factor $\beta 1$; IGF-1: Insulin-like growth factor 1; GDF-7: Growth differentiation factor 7.

BMSCs in 3 d, but TGF- $\beta 1$ and GDF-7 could increase the overall viability in 12 d.

The expression levels of Col1a1 and Col3a1, confirmed by qRT-PCR and Western blot, in the TGF- $\beta 1$ -treated group were significantly higher than those of the other groups in both the 2D and 3D cultures (Figures 2 and 3). Sirius Red and MT staining also showed that more collagen fibers were formed in the TGF- $\beta 1$ -treated group compared to the other groups in the 2D and 3D cultures, respectively (Figure 4). A high collagen production is required as a basic constituent of the ECM for repairing injured tendons^[21]. Many earlier studies have demonstrated that TGF- $\beta 1$ exerts strong effects on collagen production^[22,23]. Our results showed that TGF- $\beta 1$ has great promise for the tenogenic differentiation of BMSCs compared to GDF-7 and IGF-1 in a collagen sponge-based 3D culture. However, Rajpar and Barrett claimed that BMP-12 (GDF-7)

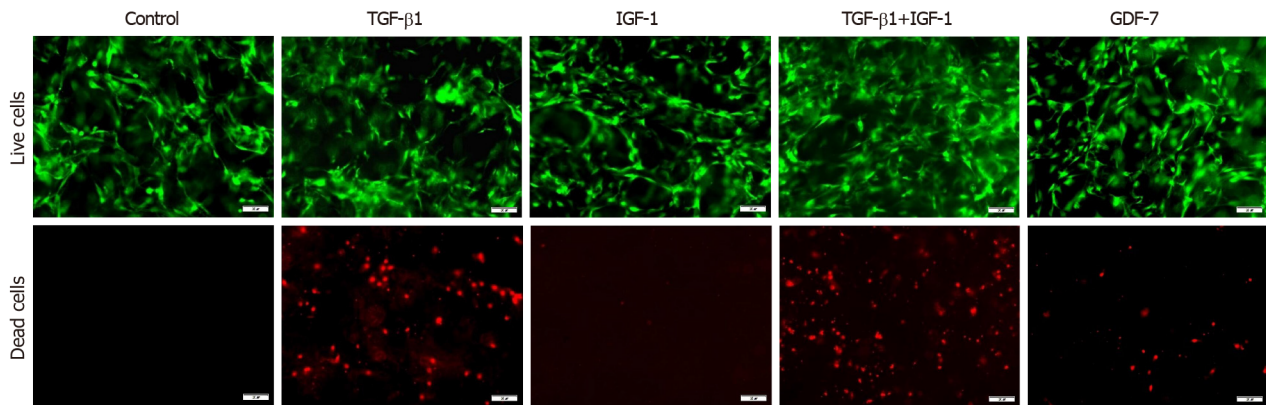


Figure 6 Effect of transforming growth factor $\beta 1$ (10 ng/mL), insulin-like growth factor 1 (10 ng/mL), growth differentiation factor 7 (100 ng/mL), or transforming growth factor $\beta 1$ combined with insulin-like growth factor 1 (both at 10 ng/mL) on the survival of bone marrow mesenchymal stem cells in the collagen sponge-based 3D culture. The live/dead assay showed that more dead cells were observed in the bone marrow mesenchymal stem cells-collagen sponge constructs treated with transforming growth factor $\beta 1$ (TGF- $\beta 1$) (TGF- $\beta 1$ combined with insulin-like growth factor 1) for 3 d. Bars = 100 μ m. Green indicates the live cells, and red indicates the dead cells. TGF- $\beta 1$: Transforming growth factor $\beta 1$; IGF-1: Insulin-like growth factor 1; GDF-7: Growth differentiation factor 7.

is primarily a tendon differentiation factor in a collagen type I hydrogel-based 3D culture, whereas FGF-2, TGF- $\beta 1$, and IGF-1 may be better described as inducers of matrix synthesis and/or cell proliferation^[6].

TGF- $\beta 1$ is a secreted protein that performs many cellular functions, including the control of cell growth, proliferation, differentiation, apoptosis, migration, and production of the ECM in many cell types^[24,25]. A number of studies have illustrated the role of TGF- $\beta 1$ in the proliferation and differentiation of BMSCs. Although this seems straightforward, the reality is complicated, as many contradictory reports have been published. TGF- $\beta 1$ can regulate the osteogenic differentiation of BMSCs. Studies have shown that low concentrations of TGF- $\beta 1$ (0.1–1 ng/mL) promote the osteogenic differentiation of BMSCs, while high concentrations of TGF- $\beta 1$ (10 ng/mL) inhibit the osteogenic differentiation of BMSCs^[26,27]. Gong *et al*^[28] reported that TGF- $\beta 1$ at a concentration of 0.1–10 ng/mL inhibited BMSC proliferation but promoted smooth muscle cell differentiation^[28]. In our study, the effect of TGF- $\beta 1$ (10 ng/mL) on the tenogenic differentiation of BMSCs was gradually reduced as the treatment time increased in the 2D culture but became more obvious in the collagen sponge-based 3D culture (Figure 2A–E). We also detected the effect of TGF- $\beta 1$ (10 ng/mL) on the mRNA expression levels of *PPAR γ* and *Runx2*. The results showed that TGF- $\beta 1$ (10 ng/mL) could efficiently inhibit the mRNA expression of *PPAR γ* in both the 2D and 3D cultures (Figure 2F), which is consistent with other studies^[29]. TGF- $\beta 1$ (10 ng/mL) had no effect on the mRNA expression of *Runx2* in the 2D culture but could inhibit the mRNA expression of *Runx2* in the 3D culture, whereas the inhibitory effect gradually disappeared with the extended culture time (Figure 2G). The 3D environment is complicated, and the transmission of growth factors in a 3D culture environment is clearly different from that in the 2D culture. In our study, the collagen sponge scaffold can provide more space for cell proliferation and affect the shape of cells, and this may promise the further enhancement of the responsiveness of the cells to growth factor treatment. Wang *et al*^[30] demonstrated that elongation of cultured human dermal fibroblasts can induce predominant tenocyte lineage differentiation *via* a synergistic effect of TGF- $\beta 1$ and cytoskeletal signaling^[30]. Rajpar and Barrett used collagen type I hydrogels to culture MSCs, and the structure and physical properties of the collagen type I hydrogel are different from those of the collagen type I sponge^[6]. Therefore, we hypothesized that the composition, structure, surface characteristics, and mechanical properties of the scaffold may also affect the cells' response to the growth factors.

CONCLUSION

We showed that collagen sponge-based 3D culture is more favorable for cells to respond to growth factors and demonstrated that TGF- $\beta 1$ is more effective in the tenogenic differentiation of BMSCs compared to GDF-7 and IGF-1 in the collagen sponge-based 3D culture. Together, our results indicated that TGF- $\beta 1$ is a powerful

chemical factor that can be used in tendon tissue engineering. These findings are helpful towards a better understanding of the role of growth factors in regulating the tenogenic differentiation of BMSCs in tendon tissue engineering.

ARTICLE HIGHLIGHTS

Research background

How to manage the damaged tendon is still one of the most challenging problems in orthopedics. With the development of cell culture techniques, medical biomaterials, and implantation techniques, tissue engineering techniques show great potential for tendon repair and regeneration. Type I collagen is a popular natural materials used for tendon tissue engineering. One of the possible practical ways to further enhance tendon repair is to combine a porous collagen sponge scaffold with a suitable growth factor that has an inherent ability to promote the recruitment, proliferation, and tenogenic differentiation of cells.

Research motivation

At present, there is an incomplete understanding of which growth factors are sufficient and optimal for the tenogenic differentiation of stem cells in a porous collagen sponge scaffold.

Research objectives

To identify one or more ideal growth factors that benefit the proliferation or/and tenogenic differentiation of stem cells in a porous collagen sponge scaffold, which will help us better understand the role of growth factors in tendon tissue engineering.

Research methods

We constructed a 3D culture system based on a type I collagen sponge which is a porous scaffold material. The surface topography of the collagen sponge was observed by scanning electron microscopy. Cell survival on the surfaces of the collagen sponge was assessed by live/dead assay, and the activity of cells was assessed by CCK-8 assay. The mRNA and protein expression levels of related molecules were confirmed by quantitative real-time polymerase chain reaction and Western blot, respectively. The deposited collagen was assessed by Sirius Red staining. Histological analyses were performed to investigate the cell distribution and collagen deposition, which can help us visually understand the changes within the constructs.

Research results

Medical type I collagen sponge had a good biocompatibility. Transforming growth factor $\beta 1$ (TGF- $\beta 1$) had great promise for the tenogenic differentiation and proliferation of bone marrow mesenchymal stem cells (BMSCs) compared to growth differentiation factor 7 (GDF-7) and insulin-like growth factor 1 (IGF-1) in a monolayer culture (2D) and in a collagen sponge-based 3D culture. Moreover, TGF- $\beta 1$ promoted more collagen deposition in both the 2D and 3D cultures. In the 2D culture, the proliferation of the BMSCs showed no significant changes compared to the control group after TGF- $\beta 1$, IGF-1, or GDF-7 treatment. However, TGF- $\beta 1$ and GDF-7 could increase the cell proliferation in the collagen sponge-based 3D culture. Strangely, we also found more dead cells in the 3D culture where cells were treated with TGF- $\beta 1$.

Research conclusions

TGF- $\beta 1$ shows great promise for BMSCs compared to GDF-7 and IGF-1 in both 2D and 3D cultures, and the collagen sponge-based 3D culture enhances the tenogenic differentiation of BMSCs well beyond the level of induction in the 2D culture after TGF- $\beta 1$ treatment.

Research perspectives

Based on our results, we believe that TGF- $\beta 1$ is an ideal growth factor that benefits the tenogenic differentiation of BMSCs in a porous collagen sponge scaffold.

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