World Journal of *Gastroenterology*

World J Gastroenterol 2021 February 28; 27(8): 666-759





Published by Baishideng Publishing Group Inc

WJG

World Journal of Gastroenterology

Contents

Weekly Volume 27 Number 8 February 28, 2021

OPINION REVIEW

New therapeutic options for persistent low-level viremia in patients with chronic hepatitis B virus 666 infection: Increase of entecavir dosage

Yin GQ, Li J, Zhong B, Yang YF, Wang MR

REVIEW

G protein-coupled receptors as potential targets for nonalcoholic fatty liver disease treatment 677 Yang M, Zhang CY

ORIGINAL ARTICLE

Basic Study

692 Fork head box M1 regulates vascular endothelial growth factor-A expression to promote the angiogenesis and tumor cell growth of gallbladder cancer

Wang RT, Miao RC, Zhang X, Yang GH, Mu YP, Zhang ZY, Qu K, Liu C

Effect of berberine on hyperglycaemia and gut microbiota composition in type 2 diabetic Goto-Kakizaki 708 rats

Zhao JD, Li Y, Sun M, Yu CJ, Li JY, Wang SH, Yang D, Guo CL, Du X, Zhang WJ, Cheng RD, Diao XC, Fang ZH

Prospective Study

725 Endoscopic full-thickness resection using an over-the-scope device: A prospective study Guo JT, Zhang JJ, Wu YF, Liao Y, Wang YD, Zhang BZ, Wang S, Sun SY

META-ANALYSIS

737 Risk factors for lymph node metastasis in T1 esophageal squamous cell carcinoma: A systematic review and meta-analysis

Jiang KY, Huang H, Chen WY, Yan HJ, Wei ZT, Wang XW, Li HX, Zheng XY, Tian D

CASE REPORT

Septic shock due to Granulicatella adiacens after endoscopic ultrasound-guided biopsy of a splenic mass: A 751 case report

Cho SY, Cho E, Park CH, Kim HJ, Koo JY



Contents

Weekly Volume 27 Number 8 February 28, 2021

ABOUT COVER

Marco Milone, MD, PhD, FACS, Professor, Surgeon, Department of Clinical Medicine and Surgery, University of Naples "Federico II", Via Pansini 5, Naples 80131, Italy. milone.marco.md@gmail.com

AIMS AND SCOPE

The primary aim of World Journal of Gastroenterology (WJG, World J Gastroenterol) is to provide scholars and readers from various fields of gastroenterology and hepatology with a platform to publish high-quality basic and clinical research articles and communicate their research findings online. WJG mainly publishes articles reporting research results and findings obtained in the field of gastroenterology and hepatology and covering a wide range of topics including gastroenterology, hepatology, gastrointestinal endoscopy, gastrointestinal surgery, gastrointestinal oncology, and pediatric gastroenterology.

INDEXING/ABSTRACTING

The WJG is now indexed in Current Contents®/Clinical Medicine, Science Citation Index Expanded (also known as SciSearch®), Journal Citation Reports®, Index Medicus, MEDLINE, PubMed, PubMed Central, and Scopus. The 2020 edition of Journal Citation Report[®] cites the 2019 impact factor (IF) for WJG as 3.665; IF without journal self cites: 3.534; 5-year IF: 4.048; Ranking: 35 among 88 journals in gastroenterology and hepatology; and Quartile category: Q2. The WJG's CiteScore for 2019 is 7.1 and Scopus CiteScore rank 2019: Gastroenterology is 17/137.

RESPONSIBLE EDITORS FOR THIS ISSUE

Production Editor: Ji-Hong Liu; Production Department Director: Yun-Xiaojian Wu; Editorial Office Director: Ze-Mao Gong.

NAME OF JOURNAL World Journal of Gastroenterology	INSTRUCTIONS TO AUTHORS https://www.wjgnet.com/bpg/gerinfo/204
ISSN	GUIDELINES FOR ETHICS DOCUMENTS
ISSN 1007-9327 (print) ISSN 2219-2840 (online)	https://www.wjgnet.com/bpg/GerInfo/287
LAUNCH DATE	GUIDELINES FOR NON-NATIVE SPEAKERS OF ENGLISH
October 1, 1995	https://www.wjgnet.com/bpg/gerinfo/240
FREQUENCY	PUBLICATION ETHICS
Weekly	https://www.wjgnet.com/bpg/GerInfo/288
EDITORS-IN-CHIEF	PUBLICATION MISCONDUCT
Andrzej S Tarnawski, Subrata Ghosh	https://www.wjgnet.com/bpg/gerinfo/208
EDITORIAL BOARD MEMBERS	ARTICLE PROCESSING CHARGE
http://www.wjgnet.com/1007-9327/editorialboard.htm	https://www.wjgnet.com/bpg/gerinfo/242
PUBLICATION DATE	STEPS FOR SUBMITTING MANUSCRIPTS
February 28, 2021	https://www.wjgnet.com/bpg/GerInfo/239
COPYRIGHT	ONLINE SUBMISSION
© 2021 Baishideng Publishing Group Inc	https://www.f6publishing.com

© 2021 Baishideng Publishing Group Inc. All rights reserved. 7041 Koll Center Parkway, Suite 160, Pleasanton, CA 94566, USA E-mail: bpgoffice@wjgnet.com https://www.wjgnet.com



WU

World Journal of Gastroenterology

Submit a Manuscript: https://www.f6publishing.com

World J Gastroenterol 2021 February 28; 27(8): 692-707

ISSN 1007-9327 (print) ISSN 2219-2840 (online)

DOI: 10.3748/wjg.v27.i8.692

ORIGINAL ARTICLE

Basic Study Fork head box M1 regulates vascular endothelial growth factor-A expression to promote the angiogenesis and tumor cell growth of gallbladder cancer

Rui-Tao Wang, Run-Chen Miao, Xing Zhang, Gang-Hua Yang, Yi-Ping Mu, Zi-Yun Zhang, Kai Qu, Chang Liu

ORCID number: Rui-Tao Wang 0000-0003-0487-0575: Run-Chen Miao 0000-0003-1651-3970; Xing Zhang 0000-0001-9568-8124; Gang-Hua Yang 0000-0002-9347-5771; Yi-Ping Mu 0000-0001-9155-5973; Zi-Yun Zhang 0000-0003-1272-3268; Kai Qu 0000-0002-1138-3727; Chang Liu 0000-0002-7318-1004.

Author contributions: Wang RT, Miao RC and Zhang X contributed equally to this work; Wang RT and Liu C designed the research; Wang RT, Miao RC and Zhang X wrote the paper; Yang GH, Mu YP and Miao RC collected the patient's clinical data; Yang GH, Zhang ZY and Qu K analyzed the data; Zhang X revised the paper; all authors read and approved the final manuscript.

Supported by Scientific and Technological Development Research Project Foundation of Shaanxi Province of China, No. 2020SF-069.

Institutional review board statement: The study was reviewed and approved by the First Affiliated Hospital of Xi'an Jiaotong University College of Medicine Institutional Review Board

Rui-Tao Wang, Run-Chen Miao, Xing Zhang, Zi-Yun Zhang, Kai Qu, Chang Liu, Department of Hepatobiliary Surgery, The First Affiliated Hospital of Xi'an Jiaotong University, Xi'an 710061, Shaanxi Province, China

Run-Chen Miao, Chang Liu, Department of SICU, The First Affiliated Hospital of Xi'an Jiaotong University, Xi'an 710061, Shaanxi Province, China

Gang-Hua Yang, Department of Geriatric Surgery, The First Affiliated Hospital of Xi'an Jiaotong University, Xi'an 710061, Shaanxi Province, China

Yi-Ping Mu, Department of Medical Information Management Office, The First Affiliated Hospital of Xi'an Jiaotong University, Xi'an 710061, Shaanxi Province, China

Corresponding author: Chang Liu, MD, PhD, Chief Doctor, Professor, Department of Hepatobiliary Surgery, The First Affiliated Hospital of Xi'an Jiaotong University, No. 277 West Yanta Road, Xi'an 710061, Shaanxi Province, China. liuchangdoctor@163.com

Abstract

BACKGROUND

Gallbladder cancer (GBC) is an aggressive type of biliary tract cancer that lacks effective therapeutic targets. Fork head box M1 (FoxM1) is an emerging molecular target associated with tumor progression in GBC, and accumulating evidence suggests that vascular endothelial growth factor (VEGF) promotes various tumors by inducing neoangiogenesis.

AIM

To investigate the role of FoxM1 and the angiogenesis effects of VEGF-A in primary GBC.

METHODS

Using immunohistochemistry, we investigated FoxM1 and VEGF-A expression in GBC tissues, paracarcinoma tissues and cholecystitis tissues. Soft agar, cell invasion, migration and apoptosis assays were used to analyze the malignant phenotype influenced by FoxM1 in GBC. Kaplan-Meier survival analysis was performed to evaluate the impact of FoxM1 and VEGF-A expression in GBC patients. We investigated the relationship between FoxM1 and VEGF-A by



Institutional animal care and use committee statement: This study was reviewed and approved by the Ethics Committee of the Xi'an Jiaotong University.

Conflict-of-interest statement: The authors declared no conflicts of interest.

Data sharing statement: No additional data are available.

ARRIVE guidelines statement: The authors have read the ARRIVE guidelines, and the manuscript was prepared and revised according to the ARRIVE guidelines.

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: htt p://creativecommons.org/License s/by-nc/4.0/

Manuscript source: Unsolicited manuscript

Specialty type: Gastroenterology and hepatology

Country/Territory of origin: China

Peer-review report's scientific quality classification

Grade A (Excellent): 0 Grade B (Very good): B Grade C (Good): C Grade D (Fair): 0 Grade E (Poor): 0

Received: September 9, 2020 Peer-review started: September 9, 2020

First decision: December 4, 2020 Revised: December 16, 2020 Accepted: January 21, 2021 Article in press: January 21, 2021 Published online: February 28, 2021

regulating the level of FoxM1. Next, we performed MTT assays and Transwell invasion assays by knocking out or overexpressing VEGF-A to evaluate its function in GBC cells. The luciferase assay was used to reveal the relationship between FoxM1 and VEGF-A. BALB/c nude mice were used to establish the xenograft tumor model.

RESULTS

FoxM1 expression was higher in GBC tissues than in paracarcinoma tissues. Furthermore, the high expression of Foxm1 in GBC was significantly correlated with a malignant phenotype and worse overall survival. Meanwhile, high expression of FoxM1 influenced angiogenesis; high expression of FoxM1 combined with high expression of VEGF-A was related to poor prognosis. Attenuated FoxM1 significantly suppressed cell proliferation, transfer and invasion in vitro. Knockdown of FoxM1 in GBC cells reduced the expression of VEGF-A. Luciferase assay showed that FoxM1 was the transcription factor of VEGF-A, and knockdown VEGF-A in FoxM1 overexpressed cells could partly reverse the malignancy phenotype of GBC cells. In this study, we found that FoxM1 was involved in regulation of VEGF-A expression.

CONCLUSION

FoxM1 and VEGF-A overexpression were associated with the prognosis of GBC patients. FoxM1 regulated VEGF-A expression, which played an important role in the progression of GBC.

Key Words: Gallbladder neoplasms; Fork head box M1; Vascular endothelial growth factor-A; Angiogenesis; Progression; Prognosis

©The Author(s) 2021. Published by Baishideng Publishing Group Inc. All rights reserved.

Core Tip: Fork head box M1 (FoxM1) is an emerging molecular target associated with early metastasis, drug resistance and poor patient survival in gallbladder cancer. Accumulating evidence suggests that vascular endothelial growth factor (VEGF) plays important roles in many kinds of tumors by inducing neoangiogenesis. In the present study, we found that VEGF-A overexpression was correlated with proliferation, invasion and poor prognosis. The results of Kaplan-Meier analysis showed that both FoxM1 and VEGF-A were associated with survival in gallbladder cancer. VEGF-A expression was regulated by FoxM1, which was the mechanism of FoxM1 enhancing the angiogenesis of gallbladder cancer.

Citation: Wang RT, Miao RC, Zhang X, Yang GH, Mu YP, Zhang ZY, Qu K, Liu C. Fork head box M1 regulates vascular endothelial growth factor-A expression to promote the angiogenesis and tumor cell growth of gallbladder cancer. World J Gastroenterol 2021; 27(8): 692-707 URL: https://www.wjgnet.com/1007-9327/full/v27/i8/692.htm

DOI: https://dx.doi.org/10.3748/wjg.v27.i8.692

INTRODUCTION

Gallbladder cancer (GBC), a frequent and aggressive type of biliary tract cancer, is now the sixth most common gastrointestinal cancer. The vast majority (65% to 90%) of GBCs are adenocarcinomas followed by squamous cell or adenosquamous cell carcinomas (5% to 10%) and undifferentiated carcinomas (5%)^[1]. The incidence rate is 2.2 per 100000 per year worldwide^[1]. Surgery combined with adjuvant chemotherapy and radiotherapy is the general therapy for patients with GBC. However, GBC is known to have an occult onset, rapid progression and abysmal prognosis, with a mean 6-mo survival time and a 5-yr survival rate of 5%^[2]. Furthermore, a low diagnosis rate of GBC has been demonstrated in Japan, with only a 0.011% detection rate from mass screening^[3]. Thus, accurate and specific diagnosis of GBC at the early stage is still urgently needed to improve therapeutic efficacy.

Cancer cells achieve invasion and metastasis through extracellular proteolysis,



P-Reviewer: Merigo F, Zhang LS-Editor: Gao CCL-Editor: FilipodiaP-Editor: Liu JH



which affects cell proliferation, adhesion, migration and angiogenesis. In addition, Fork head box M1 (FoxM1) is characterized as a critical proliferation-associated transcription factor in cell cycle progression^[4]. Inhibiting FoxM1 expression has been reported to induce the loss of the mitotic spindle, resulting in failed cell division. FoxM1 is widely expressed in normal proliferating cells and is also overexpressed in cancer cells, such as ovarian cancer cells, renal cell carcinoma cells, pancreatic cancer cells, breast cancer cells, laryngeal squamous carcinoma cells, glioma cells, prostate carcinoma cells and gastric cancer cells^[5]. Moreover, the overexpression of FoxM1 has an impact on poor patient prognosis and can be regarded as an independent predictor of low survival^[4]. However, in patients with GBC, the clinical correlation of FoxM1 expression remains unclear.

In recent research, investigators have found that the downregulation of FoxM1 inhibits the activity of vascular endothelial growth factor (VEGF), matrix metalloproteinase (MMP)-2, MMP-9, Cyclin B1, CDC25, p21 and p27 proteins, affecting cancer cell proliferation, migration and invasion^[6]. Moreover, FoxM1 was proven to be upregulated at both the mRNA and protein levels^[7]. In glioma, with the increase in FoxM1, cancer cell invasiveness and tumorigenicity are also promoted^[8]. These investigations indicate that FoxM1 plays an important role in the progression of carcinoma and could have the same function in GBC cells. Two investigations also found a correlation between lymph node metastasis and FoxM1. However, the correlation between elevated levels of FoxM1 and clinical specimens has been controversial.

We found that FoxM1 overexpression promoted tumor cell migration and invasion, while FoxM1 downregulation led to an increase in cell apoptosis rates, indicating that FoxM1 might play a key role in disease progression of GBC. Moreover, we found that FoxM1 could be directly involved in the regulation of VEGF-A expression by combining with the VEGF-A promoter and activating VEGF-A transcription. Therefore, we indicated that FoxM1 could promote GBC *progression via* VEGF-A.

MATERIALS AND METHODS

Patient tissues and information

After surgery, 48 cases of fresh frozen GBC tumor tissue were collected from the First Affiliated Hospital of Xi'an Jiaotong University, together with paired tumor-free liver tissue (at least 2 cm from the tumor) from September 2015 to February 2017. GBC was confirmed by histopathological examination, and the study was approved by the Ethics Staff of the First Affiliated Hospital of Xi'an Jiaotong University.

Cell culture and treatment

The human GBC cell line SGC-996 was purchased from the Cell Bank of the Chinese Academy of Sciences (Shanghai branch) and was cultured in RPMI 1640 medium containing 10% fetal bovine serum (Gibco, Grand Island, NY, United States) at 37 °C with 5% CO_2 .

shRNA transfection

At 24 h after cell seeding in the culture dish, the recombinant adenovirus vector containing specific shRNA was transfected into SGC-996 cells with Lipofectamine 2000 (Invitrogen, United States) at different multiplicities of infection. Recombinant lentiviruses containing FoxM1 shRNA and VEGF-A shRNA were purchased from GenePharma (Shanghai, China). A negative control carrying green fluorescent protein, which expresses a scrambled RNA, was constructed as a control. The virus containing the construct was isolated using plaque screening, purification and amplification. The protocol for lentivirus infection was according to the GenePharma Recombinant Lentivirus Operation Manual (http://www.genepharma.com).

The overexpression of VEGF-A and FoxM1 by lentiviral transfection

A FoxM1 expression plasmid was purchased from Sino Biological Inc. (Beijing, China). SGC-996 cells were transfected with the FoxM1 expression vector or an empty control vector (400 ng/well) using HiPerFect. After 48 h of transfection, the cells were collected, and protein levels were analyzed by Western blotting and quantitative reverse transcription (qRT)-PCR. The overexpression of VEGF-A was performed as described above.

Zaishideng® WJG | https://www.wjgnet.com

Western blot analysis

Western blot analysis was performed as previously reported. Briefly, the membranes were probed with the following primary antibodies: FoxM1 (1:1000, Proteintech, Wuhan, China), VEGF-A (1:1000, Proteintech, Wuhan, China) and β -actin (1:3000, Abways, Shanghai, China). After being washed with TBS-T, the membranes were incubated with horseradish peroxidase-conjugated anti-rabbit or anti-mouse secondary antibody (1:10000, Proteintech, Wuhan, China). Mouse anti- β -actin (1:1000, Proteintech, Wuhan, China) with goat anti-mouse (1:10000, Proteintech, Wuhan, China) antibodies were used as a loading control. Chemiluminescence detection was performed with ChemiGlow detection reagents (Bio-Rad Western ECL Substrate). The blots were visualized with a Bio-Rad ChemiDoc MP and quantified with a densitometer using the imager application program (Alpha Innotech, San Leandro, CA, United States).

RNA isolation and RT-PCR

RNA isolation and RT-PCR analysis were performed as previously described. The cDNA for FoxM1, VEGF-A and β -actin were amplified using the Platinum Taq DNA Polymerase kit (Life Technologies) with specific primers. β -actin was used as an internal control. Primer sequences were as follows: FoxM1 Forward, 5'-CAC CCC AGT GCC AAC CGC TAC TTG-3'; FoxM1 Reverse, 5'-AAA GAG GAG CTA TCC CCT CCT CAG-3'; VEGF-A Forward, 5'- CAG ATT ATG CGG ATC AAA CCT CA -3'; VEGF-A Reverse, 5'-CAA GGC CCA CAG GGA CTG CAA -3'.

Immunohistochemical staining

Tissue specimens were fixed in neutral buffered formalin (10% vol/vol formalin in water, pH 7.4) and embedded in paraffin wax. Serial sections of 5 mm thickness were cut and mounted on charged glass slides. Conditions for FoxM1 and VEGF-A were optimized and evaluated by two independent pathologists. The goat monoclonal antibodies against FoxM1 (sc-26688; Santa Cruz Biotechnology, Santa Cruz, CA, United States) and VEGF-A (sc-152; Santa Cruz Biotechnology, Santa Cruz, CA, United States) were used at dilutions of 1:200. The streptavidin-peroxidase kit (SP-9000 Golden Bridge Int., Beijing, China) was used in accordance with the manufacturer's instructions. An irrelevant goat antiserum served as a negative control. Sections were counterstained with Mayer's hematoxylin. The immunohistochemical score and analysis were described as previously reported.

Soft agar cloning assay

Experiments with soft agar were carried out on 6-well plates. Agar was used for the bottom (0.8%) and top (0.4%) containing cells and fed every 4 d in a complete DMEM medium. After 20 d, colonies with diameters greater than 150 µm were scored under a 10-fold fluorescence microscope.

Apoptosis assay

The SGC-996 cells were inoculated into a 6-well plate with a density of 1×10^6 cells per well. Cells at logarithmic growth stage were digested by 0.25% trypsin, and the supernatant was discarded after 800 rpm centrifugation for 5 min. The mixture was resuspended with PBS, centrifuged at 800 rpm for 5 min, and washed twice with PBS. Annexin V (10 μ L) was added at a concentration of 50 μ g/mL and left standing for 40min (Bender Medsystems, Australia). Flow cytometry was used to detect apoptosis (Beckman, United States).

MTT assay for cell viability

SGC-996 cells were seeded at 5×10^3 cells per well in 96-well flat-bottom plates before transfection. For the assay, 20 μ L of MTT solution (5 g/L) was added to each well, and the cells were incubated for 4 h. Then, supernatants were removed, and formazan crystals were dissolved in 200 mL dimethyl sulfoxide. After the insoluble crystals were completely dissolved, the absorbance values at 490 nm were measured using a microplate reader (Bio-Rad, Hercules, CA, United States).

Cell invasion and migration assays

Cell invasion and migration assays were performed using Transwell permeable supports with 8 µm pore size (Costar, Cambridge, MA, United States). Cells were suspended in serum-free medium and seeded into Transwell inserts either uncoated (for the migration assay) or coated (for the invasion assay) with growth factor-reduced



Matrigel (BD Biosciences, Bedford, MA, United States). Bottom wells were filled with complete medium. After 24 h, the invaded cells were fixed with methanol and stained with a crystal violet solution. The number of cells that penetrated the membrane was determined by counting the mean cell number in five randomly selected high-power fields.

Clone formation assay

For the clonogenic assay, cells were plated in the 6-well plate with 1000 cells per well. The samples were fixed with 4% paraformaldehyde for 30 min and stained with 0.05% crystal violet for 20 min. Representative photos were collected and more than 40 colonies were counted.

Endothelial cell tube formation assay

The pre-cooled 96 well plates were coated with matrix gel (35 μ L/well) and then placed in the humidified CO₂ incubator at 37 degrees Celsius for 1 h to solidify the coating. HUVEC cells were starved overnight under 1% bovine serum albumin. Then, the cells were digested with trypsin and collected and inoculated on a 96 well plate with a density of 2.5×10^4 cells per well. SGC-996 cells were cocultured with HUVEC for 4 h. The tube formation of HUVECs was observed at 4 h by a microscope.

Luciferase assay

A pmirGLO Dual luciferase miRNA Target Expression Vector (E1330; Promega Corporation) contained the fragment of FOXM1 3'UTR and VEGF 3'UTR was used to establish luciferase reporter plasmids: WT-FOXM1; MUT-FOXM1; WT-VEGF; and MUT-VEGF. HEK293T cells were cultured in 24-well plates. When the cells reached 70% confluence, cells were transfected with 2 µg WT-FOXM1 or MUT-FOXM1 and 2 µg WT-VEGF or MUT-VEGF using Lipofectamine® 3000 (Invitrogen, Thermo Fisher Scientific, Inc). After 48 h, luciferase activity was detected by the dual-enzyme reporter assay kit (Promega).

Xenotransplantation model in nude mice

Animal experiments were approved by the Animal Ethics Committee of the Medical College of Xi'an Jiaotong University. BALB/c nude mice aged 4-6 wk were purchased from Medical Laboratory Animal Center of Xi'an Jiaotong University. Mice weighing 16 to 20 g were randomly divided into three groups: (1) Lenti-FoxM1-shRNA; (2) cDNA-FoxM1; or (3) control (1×10^7 SGC-996 cells were injected into the back of mice to establish xenograft tumor model). After 1 mo, the mice were killed, and the tumors were collected.

Statistical analysis

All data are expressed as the mean ± SD. To compare the means of normally distributed variables, analysis of variance or Student's t test was applied. P value less than 0.05 was considered significant in all tests. All analyses were performed using SPSS software version 19.0 (SPSS Inc., Chicago, IL, United States).

RESULTS

FoxM1 was overexpressed in GBC tissues

First, the expression of FoxM1 was higher than that in paracarcinoma tissues or cholecystitis tissues (Figure 1A). Moreover, we compared the levels of FoxM1 expression in four paired GBC clinical samples by qRT-PCR and Western blot analysis. The results revealed that the expression of FoxM1 in GBC was significantly higher than that in paracarcinoma tissue (Figure 1B and 1C). Western blot results showed that the protein level of FoxM1 in GBC tissues was higher than the expression of FoxM1 in paracarcinoma tissues (Figure 1D and 1E). This suggested that FoxM1 expression was closely related to the development of GBC.

Overexpression of FoxM1 promoted proliferation and metastasis of GBC cells in vitro

To investigate the function of FoxM1 in GBC, the expression of FoxM1 was suppressed by lenti-FoxM1 shRNA and was elevated by lenti- FoxM1 in the SGC-996 cell line. The protein and mRNA levels of FoxM1 were significantly decreased after lenti-FoxM1 shRNA transfection (Figure 2A-D). The protein and mRNA levels of FoxM1 were





Figure 1 The expression of fork head box M1 in gallbladder cancer. A: The expression of fork head box M1 in gallbladder cancer tissues (immunohistochemical staining, scale bars: 100 µm); B and C: Relative mRNA expression of fork head box M1; D and E: Relative protein expression of fork head box M1. ^a*P* < 0.05, ^b*P* < 0.01. FoxM1: Fork head box M1.

significantly increased after lenti- FoxM1 transfection (Figure 2E-H).

As demonstrated by the soft agar cloning assay, lenti-FoxM1 shRNA significantly decreased proliferation in SGC-996 cells. Conversely, lenti-FoxM1 significantly increased SGC-996 cell proliferation (Figure 2I and 2]). To determine the potential metastatic-promoting effect of FoxM1 in GBC cells, we investigated cell migration and invasion abilities. Compared with that in SGC-996 cells, less cell penetration was observed in lenti-FoxM1 shRNA-transfected cells. However, lenti-FoxM1-transfected cells had stronger invasion abilities in the presence of Matrigel (Figure 3A-D). As shown in the apoptosis assay, knockdown-FoxM1 cells increased apoptotic levels (Figure 3E and 3F). Tube formation also showed that in FoxM1 up-regulated cells vessel generation ability was significantly enhanced (Figure 3G and 3H). These data suggested that FoxM1 plays important roles in GBC progression.

FoxM1 promoted GBC cell proliferation and metastasis via VEGF-A

As shown in Figure 3G and 3H, the high expression level of FoxM1 influenced vessel generation in GBC. Meanwhile, research revealed that VEGF-A was associated with endothelial cell proliferation, division, migration and lumen formation. Therefore, we examined the correlation between FoxM1 and VEGF-A. As shown in Figure 3, FoxM1 and VEGF-A were both associated with GBC, and the expression of VEGF-A was obviously decreased in FoxM1 siRNA-transfected SGC-996 cells. We detected the expression of FoxM1 and VEGF-A by consecutive section immunohistochemistry in 48 GBC tissues. A positive correlation between FoxM1 and VEGF-A was observed (Figure 4A-D). Thus, we hypothesized that VEGF-A is regulated by FoxM1 to promote GBC cell migration and invasion.





Figure 2 Fork head box M1 influenced proliferation of gallbladder cancer cells in vitro. A-H: Relative mRNA and protein expression of fork head box M1; I: Rate of proliferation after inhibition of fork head box M1. ^a*P* < 0.05, ^b*P* < 0.01. FoxM1: Fork head box M1.

First, we investigated whether FoxM1 regulated the protein and mRNA levels of VEGF-A in GBC cells. Therefore, we determined the VEGF-A protein and mRNA expression levels after silencing FoxM1 by Western blot and RT-PCR analyses, respectively. We found that FoxM1 knockdown led to significant downregulation of VEGF-A protein and mRNA expression levels in SGC-996 cells (Figure 4E and 4G). To further prove the link between FoxM1 and VEGF-A expression, we overexpressed FoxM1 by transfecting lenti-FoxM1 in SGC-996 cells and found that VEGF-A expression was significantly increased at the transcriptional and translational levels (Figure 4F and 4H). Moreover, the luciferase assay showed that FoxM1 was the transcription factor of VEGF-A (Figure 4I). Nevertheless, knockdown VEGF-A in FoxM1 overexpressed cells could partly reverse the malignant phenotype of GBC cells

WJG | https://www.wjgnet.com



Figure 3 Fork head box M1 influenced migration and invasion of gallbladder cancer cells *in vitro*. A-D: Lenti-fork head box M1-transfected cells had stronger migration and invasion abilities (cell invasion and migration assays, scale bars: 200 µm); E and F: Knockdown-fork head box M1 cells increased

Caishideng® WJG | https://www.wjgnet.com

apoptotic levels (apoptosis assay); G and H: Upregulated fork head box M1 significantly enhanced vessel generation ability (endothelial cell tube formation assay, scale bars: 100 μ m). ^aP < 0.05, ^bP < 0.01. FoxM1: Fork head box M1.



Figure 4 Fork head box M1 promoted gallbladder cancer cell proliferation and metastasis via vascular endothelial growth factor-A. A-C: Hematoxylin-eosin staining, immunohistochemical staining of fork head box M1 and vascular endothelial growth factor-A (VEGF-A) in gallbladder cancer tissues (scale bars: 100 µm); D: Correlation between fork head box M1 and VEGF-A analyzed according to fluorescence score; E-H: Relative protein expression of VEGF-A; I: Luciferase assay of fork head box M1 and VEGF-A. *P < 0.05, *P < 0.01. FoxM1: Fork head box M1; VEGF-A: Vascular endothelial growth factor-A; NC: Normal control.

> (Figure 5A-I). These results indicated that FoxM1 was involved in the regulation of VEGF-A expression.

FoxM1 enhanced the angiogenesis of GBC by regulating VEGF-A in vivo

Because the in vitro study showed that VEGF-A, a potent vascular active molecule, promoted proliferation, invasion and metastasis regulated by FoxM1, we further





Figure 5 Knockdown vascular endothelial growth factor-A in fork head box M1 overexpressed cells could partly reverse the malignant phenotype of gallbladder cancer cells. A: Relative protein expression of fork head box M1 and vascular endothelial growth factor-A; B and C: Clone formation of gallbladder cancer cells; D-G: Knockdown vascular endothelial growth factor-A in fork head box M1 overexpressed cells influenced migration and invasion abilities (cell invasion and migration assays, scale bars: 200 μ m); H and I: Knockdown vascular endothelial growth factor-A in fork head box M1 overexpressed cells suppressed vessel generation ability (endothelial cell tube formation assay, scale bars: 100 μ m). ^a*P* < 0.05, ^b*P* < 0.01. FoxM1: Fork head box M1; VEGF-A: Vascular endothelial growth factor-A.

investigated the function of the FoxM1-VEGF-A axis on angiogenesis in GBC. As a result, the tumor volume and weight of mice transfected with lenti-FoxM1-shRNA or lenti-FoxM1 were determined (Figure 6A-D). The tumors of mice transfected with lenti-FoxM1 were larger than the control group, and the tumors of mice transfected with lenti-FoxM1-shRNA were smaller than the control group. Next, analysis of hematoxylin-eosin staining was performed to detect the number of tumor foci (Figure 6E). The most tumor foci were in tumor tissues of mice overexpressing FoxM1. Additionally, microvascular density, CD31 and VEGF-A were measured to investigate the angiogenic ability (Figure 6F and 6G). The above results showed that FoxM1 can improve the angiogenesis of GBC by regulating the expression of VEGF-A. Kaplan-

Raishideng® WJG https://www.wjgnet.com



shFoxM1



Snishideng® WJG | https://www.wjgnet.com



Figure 6 Fork head box M1 enhanced the angiogenesis of gallbladder cancer by regulating vascular endothelial growth factor-A in vivo. A: Tumor-bearing mice [(1) Lenti-fork head box M1 (FoxM1)-shRNA, (2) cDNA-FoxM1, (3) control: xenotransplantation model]; B: Tumor from the three groups; C: Body weight of tumor-bearing mice; D: Tumor volume of tumor-bearing mice; E: Hematoxylin-eosin staining of tumor foci; F: Hematoxylin-eosin staining of microvascular density; G: Immunohistochemistry staining of CD31. Five fields per sample were counted; four samples in the control and FoxM1 groups and three samples in the shFoxM1 group were analyzed. Scale bars: 200 µm. ^aP < 0.05, ^bP < 0.01. FoxM1: Fork head box M1.

> Meier analysis was used to assess the impact of FoxM1 and VEGF-A on survival. The results showed that the group with high expression of FoxM1 or VEGF-A showed poor survival compared with the patients with low expression of FoxM1 and VEGF-A (Figure 7).

DISCUSSION

GBC is still a frequent malignancy with a dismal prognosis. Because of the rapid progression and low diagnosis rate of GBC, it is necessary to explore the inherent mechanisms of tumorigenesis. The essential interrelation between FoxM1 and VEGF-A





Figure 7 High expression of fork head box M1 and vascular endothelial growth factor-A were associated with poor prognosis of gallbladder cancer patients. Kaplan-Meier analysis, n = Blue: 12, Green: 14, Red: 20. FoxM1: Fork head box M1; VEGF-A: Vascular endothelial growth factor-A.

provided us with a broader approach to malignancy therapy. In this investigation, we found that FoxM1 played an essential role in the expression of VEGF. Previous studies have indicated that high expression levels of FoxM1 are relevant to adverse prognosis in multiple malignancies^[9]. We proved that the upregulation of FoxM1 in GBC cells influenced the expression of VEGF-A in vivo and in vitro. In GBC patients, VEGF-A expression was closely related to the overexpression of FoxM1 and was highly expressed to promote vascularization. FoxM1 could directly regulate the activity of the VEGF gene promoter to induce its expression. We detected the opposite results in GBC cells with inhibited FoxM1. Our conclusions could provide a novel strategy for treating GBC.

The contribution of FoxM1 on numerous kinds of malignancies has been demonstrated by previous investigations^[8,10-13]. As an important factor of poor prognosis, the expression level of FoxM1 has a negative correlation with survival duration^[14-16]. In our previous study, we found that FoxM1 was upregulated in GBC tissues compared with normal tissues and as a prognostic biomarker had greater relevance to prognosis and survival^[17]. Likewise, in this study, the role of VEGF-A in the progression of GBC was investigated. The same tumor-promoting function through enhanced proliferation, invasion and metastasis was observed in GBC cells. The function of VEGF-A in promoting cancer was also shown in other investigations^[18,19]. A significant decrease in the expression of VEGF-A was detected after knocking down the expression of FoxM1.

Angiogenesis is indispensable to the proliferation of tumor cells, auxesis, migration and invasion. In addition, directly controlling angiogenesis could inhibit the progression of tumors. Angiogenesis is influenced by various factors, which has been shown in many studies^[20]. Oxygen deficiency is one such factor. VEGF expression could be promoted by oxygen deficiency in a mouse model, especially in the adjacent areas of the necrotic regions^[18-20]. For example, as glioma is a hypoxic tumor, the overexpression of VEGF in glioma is associated at least with hypoxia and has been observed in many studies^[21-23]. Moreover, the following two observations were made in our research. The expression of VEGF in GBC cells was significantly higher than that in paracarcinoma tissue. The expression of VEGF in the adjacent areas of the necrotic regions was significantly related to the cells in other normal areas (P < 0.001).

In addition to VEGF, MMP-2, MMP-9 and uPAR are thought to participate in the migration and invasion of tumor cells^[24]. It is generally accepted that VEGF is a predominant facilitator of angiogenesis and the angiogenic signaling cascade^[25]. VEGF-A, a member of the VEGF family, is also considered to be critical for tumor growth, invasion and metastasis^[26,27]. The direct interaction of FoxM1 with the VEGF-A promoter was discovered in our study, which could prove the direct regulation between FoxM1 and VEGF-A. First, the participation of FoxM1 in the regulation of the VEGF promoter was evidenced by our results, and the expression of FoxM1 could activate the VEGF promoter-mediated luciferase expression. Then, we selected two consensus binding sequences on the VEGF-A promoter to which FoxM1 could bind both in vivo and in vitro. After destroying them, the promoter of VEGF-A could not mediate its transcription. The activity of the FoxM1 binding areas could be evaluated.

It is also widely accepted that FoxM1 is a crucial mediator in the cell cycle^[28]. FoxM1 is accumulated in the late G1 and S phases during the cell cycle. It could also be



downregulated by many tumor suppressors, such as p53. FoxM1-siRNA has been used to decrease the proliferation and division of cells by suppressing mRNA and protein related to the cell cycle. As essential components of tumor progression, MMP-2, MMP9, uPA and uPAR were reduced by decreasing FoxM1, which has been proven in numerous studies^[6].

Previous investigations have shown that as a consequence of downregulating FoxM1 with FoxM1-siRNA, the colonies of tumor cells decrease significantly^[29]. Other studies also revealed that downregulation of FoxM1 could decrease the average number of cells crossing the Matrigel-coated membrane in one high power field and the migrating distance^[5,30]. This evidence suggests that FoxM1 plays an essential role in tumor progression. In the present study, we also found that the downregulation of FoxM1 could suppress the viability, migration and invasion of tumor cells. Moreover, decreased FoxM1 has been shown to be relevant to prognosis and survival.

In conclusion, this study indicated that FoxM1 could be directly involved in the regulation of VEGF-A expression by activating and regulating the VEGF-A promoter. The overexpression of FoxM1 leading to excess VEGF-A and angiogenesis was proven to be enhanced in the adjacent areas of the necrotic regions. Therefore, it is conceivable that controlling the expression of FoxM1 could inhibit the angiogenesis and growth of GBC cells with the possibility of decreasing the invasion and metastasis of cancer cells, indicating that FoxM1 may play a key role in the pathogenesis and disease progression of GBC. This is the first work on the effects of FoxM1 and VEGF-A on GBC cells, and we hope the data can offer some reference and evidence to other studies or promote further discussion.

CONCLUSION

In conclusion, FoxM1 and VEGF-A overexpression were associated with prognosis of GBC patients. FoxM1 regulated VEGF-A expression, which played an important role in the progression of GBC.

ARTICLE HIGHLIGHTS

Research background

Gallbladder cancer (GBC), with low detectable diagnosis and high metastasis, has a short survival time and a low 5-yr survival rate. Fork head box M1 (FoxM1) is relevant to poor prognosis and malignant behaviors, including proliferation, invasion and metastasis. Vascular endothelial growth factor-A (VEGF-A) plays an important role in angiogenesis. In GBC, it is meaningful to investigate the functional correlation between FoxM1 and VEGF-A.

Research motivation

FoxM1 is relevant to poor prognosis and malignant behaviors including proliferation, invasion and metastasis. VEGF-A plays an important role in angiogenesis. However, it is unclear whether FoxM1 could regulate VEGF-A.

Research objectives

This study aimed to investigate whether FoxM1 enhanced the angiogenesis of GBC *via* regulating VEGF-A.

Research methods

Using immunohistochemistry, we investigated FoxM1 and VEGF-A expression in GBC tissues, paracarcinoma tissues and cholecystitis tissues. Soft agar, cell invasion, migration and apoptosis assays were used to analyze the malignant phenotype influenced by FoxM1 in GBC. Kaplan-Meier survival analysis was performed to evaluate the impact of FoxM1 and VEGF-A expression in GBC patients. We investigated the relationship between FoxM1 and VEGF-A by regulating the level of FoxM1. Next, we performed MTT assays and Transwell invasion assays by knocking out or overexpressing VEGF-A to evaluate its function in GBC cells. The luciferase assay was used to reveal the relationship between FoxM1 and VEGF-A. BALB/c nude mice were used to establish the xenograft tumor model.

Research results

FoxM1 expression was higher in GBC tissues than in paracarcinoma tissues. Furthermore, the high expression of FoxM1 in GBC was significantly correlated with the malignant phenotype and worse overall survival. Meanwhile, high expression of FoxM1 influenced angiogenesis. Attenuated FoxM1 significantly suppressed cell proliferation, transfer and invasion in vitro. Knockdown of FoxM1 in GBC cells reduced the expression of VEGF-A. The luciferase assay showed that FoxM1 was the transcription factor of VEGF-A. Knockdown VEGF-A in FoxM1 overexpressed cells could partly reverse the malignant phenotype of GBC cells. High expression of FoxM1 combined with high expression of VEGF-A was related to poor prognosis. In this study, we found that FoxM1 was involved in regulation of VEGF-A expression.

Research conclusions

FoxM1 and VEGF-A overexpression were associated with prognosis of GBC patients. FoxM1 upregulated VEGF-A expression, which played an important role in the progression of GBC.

Research perspectives

By comprehending the way FoxM1 induced angiogenesis through regulating VEGF-A in GBC, the present study showed a possible method for treatment strategy of GBC patients with metastasis and in late stage.

ACKNOWLEDGEMENTS

We are indebted to the individuals who have participated in or have helped with this article.

REFERENCES

- Mehrotra R, Tulsyan S, Hussain S, Mittal B, Singh Saluja S, Singh S, Tanwar P, Khan A, Javle M, 1 Hassan MM, Pant S, De Aretxabala X, Sirohi B, Rajaraman P, Kaur T, Rath GK. Genetic landscape of gallbladder cancer: Global overview. Mutat Res 2018; 778: 61-71 [PMID: 30454684 DOI: 10.1016/j.mrrev.2018.08.003]
- 2 Montalvo-Jave EE, Rahnemai-Azar AA, Papaconstantinou D, Deloiza ME, Tsilimigras DI, Moris D, Mendoza-Barrera GE, Weber SM, Pawlik TM. Molecular pathways and potential biomarkers in gallbladder cancer: A comprehensive review. Surg Oncol 2019; 31: 83-89 [PMID: 31541911 DOI: 10.1016/j.suronc.2019.09.006]
- Hickman L, Contreras C. Gallbladder Cancer: Diagnosis, Surgical Management, and Adjuvant 3 Therapies. Surg Clin North Am 2019; 99: 337-355 [PMID: 30846038 DOI: 10.1016/j.suc.2018.12.008
- 4 Choi HJ, Jhe YL, Kim J, Lim JY, Lee JE, Shin MK, Cheong JH. FoxM1-dependent and fatty acid oxidation-mediated ROS modulation is a cell-intrinsic drug resistance mechanism in cancer stem-like cells. Redox Biol 2020; 36: 101589 [PMID: 32521504 DOI: 10.1016/j.redox.2020.101589]
- 5 Nandi D, Cheema PS, Jaiswal N, Nag A. FoxM1: Repurposing an oncogene as a biomarker. Semin Cancer Biol 2018; 52: 74-84 [PMID: 28855104 DOI: 10.1016/j.semcancer.2017.08.009]
- 6 Liao GB, Li XZ, Zeng S, Liu C, Yang SM, Yang L, Hu CJ, Bai JY. Regulation of the master regulator FOXM1 in cancer. Cell Commun Signal 2018; 16: 57 [PMID: 30208972 DOI: 10.1186/s12964-018-0266-6]
- 7 Song X, Fiati Kenston SS, Zhao J, Yang D, Gu Y. Roles of FoxM1 in cell regulation and breast cancer targeting therapy. Med Oncol 2017; 34: 41 [PMID: 28176242 DOI: 10.1007/s12032-017-0888-31
- Kim H, Park KJ, Ryu BK, Park DH, Kong DS, Chong K, Chae YS, Chung YG, Park SI, Kang SH. 8 Forkhead box M1 (FOXM1) transcription factor is a key oncogenic driver of aggressive human meningioma progression. Neuropathol Appl Neurobiol 2020; 46: 125-141 [PMID: 31179553 DOI: 10.1111/nan.12571]
- Liu L, Wu J, Guo Y, Xie W, Chen B, Zhang Y, Li S, Hua Y, Peng B, Shen S. Overexpression of FoxM1 predicts poor prognosis of intrahepatic cholangiocarcinoma. Aging (Albany NY) 2018; 10: 4120-4140 [PMID: 30580327 DOI: 10.18632/aging.101706]
- 10 Wang H, Li H, Hu L, Wang J, Liu Q, Wang D, Sun X. Overexpression of FoxM1 in Sinonasal Inverted Papilloma and Associated Squamous Cell Carcinoma. Am J Rhinol Allergy 2019; 33: 706-715 [PMID: 31333047 DOI: 10.1177/1945892419863193]
- 11 Zhou W, Yu X, Sun S, Zhang X, Yang W, Zhang J, Zhang X, Jiang Z. Increased expression of MMP-2 and MMP-9 indicates poor prognosis in glioma recurrence. Biomed Pharmacother 2019; 118: 109369 [PMID: 31545229 DOI: 10.1016/j.biopha.2019.109369]



- 12 Hu G, Yan Z, Zhang C, Cheng M, Yan Y, Wang Y, Deng L, Lu Q, Luo S. FOXM1 promotes hepatocellular carcinoma progression by regulating KIF4A expression. J Exp Clin Cancer Res 2019; 38: 188 [PMID: 31072351 DOI: 10.1186/s13046-019-1202-3]
- 13 Marchand B, Pitarresi JR, Reichert M, Suzuki K, Laczkó D, Rustgi AK. PRRX1 isoforms cooperate with FOXM1 to regulate the DNA damage response in pancreatic cancer cells. Oncogene 2019; 38: 4325-4339 [PMID: 30705403 DOI: 10.1038/s41388-019-0725-6]
- 14 Cai H, Chen J, He B, Li Q, Li Y, Gao Y. A FOXM1 related long non-coding RNA contributes to gastric cancer cell migration. Mol Cell Biochem 2015; 406: 31-41 [PMID: 25907137 DOI: 10.1007/s11010-015-2421-3]
- 15 Huang W, Chen Z, Zhang L, Tian D, Wang D, Fan D, Wu K, Xia L. Interleukin-8 Induces Expression of FOXC1 to Promote Transactivation of CXCR1 and CCL2 in Hepatocellular Carcinoma Cell Lines and Formation of Metastases in Mice. Gastroenterology 2015; 149: 1053-67. e14 [PMID: 26065367 DOI: 10.1053/j.gastro.2015.05.058]
- Okada K, Fujiwara Y, Takahashi T, Nakamura Y, Takiguchi S, Nakajima K, Miyata H, Yamasaki M, 16 Kurokawa Y, Mori M, Doki Y. Overexpression of forkhead box M1 transcription factor (FOXM1) is a potential prognostic marker and enhances chemoresistance for docetaxel in gastric cancer. Ann Surg Oncol 2013; 20: 1035-1043 [PMID: 23054116 DOI: 10.1245/s10434-012-2680-0]
- Wang R, Song Y, Xu X, Wu Q, Liu C. The expression of Nek7, FoxM1, and Plk1 in gallbladder 17 cancer and their relationships to clinicopathologic features and survival. Clin Transl Oncol 2013; 15: 626-632 [PMID: 23359173 DOI: 10.1007/s12094-012-0978-9]
- 18 Wang Y, Huang Q, Liu J, Wang Y, Zheng G, Lin L, Yu H, Tang W, Huang Z. Vascular endothelial growth factor A polymorphisms are associated with increased risk of coronary heart disease: a metaanalysis. Oncotarget 2017; 8: 30539-30551 [PMID: 28430629 DOI: 10.18632/oncotarget.15546]
- 19 Marisi G, Scarpi E, Passardi A, Nanni O, Ragazzini A, Valgiusti M, Casadei Gardini A, Neri LM, Frassineti GL, Amadori D, Ulivi P. Circulating VEGF and eNOS variations as predictors of outcome in metastatic colorectal cancer patients receiving bevacizumab. Sci Rep 2017; 7: 1293 [PMID: 28465540 DOI: 10.1038/s41598-017-01420-01
- Xie Y, Mansouri M, Rizk A, Berger P. Regulation of VEGFR2 trafficking and signaling by Rab 20 GTPase-activating proteins. Sci Rep 2019; 9: 13342 [PMID: 31527750 DOI: 10.1038/s41598-019-49646-4]
- Wei Y, Sun Q, Zhao L, Wu J, Chen X, Wang Y, Zang W, Zhao G. LncRNA UCA1-miR-507-21 FOXM1 axis is involved in cell proliferation, invasion and G0/G1 cell cycle arrest in melanoma. Med Oncol 2016; 33: 88 [PMID: 27389544 DOI: 10.1007/s12032-016-0804-2]
- Engeland K. Cell cycle arrest through indirect transcriptional repression by p53: I have a DREAM. 22 Cell Death Differ 2018; 25: 114-132 [PMID: 29125603 DOI: 10.1038/cdd.2017.172]
- Jin H, Li XJ, Park MH, Kim SM. FOXM1-mediated downregulation of uPA and MMP9 by 3,3'-23 diindolylmethane inhibits migration and invasion of human colorectal cancer cells. Oncol Rep 2015; 33: 3171-3177 [PMID: 25962429 DOI: 10.3892/or.2015.3938]
- Elaimy AL, Mercurio AM. Convergence of VEGF and YAP/TAZ signaling: Implications for 24 angiogenesis and cancer biology. Sci Signal 2018; 11 [PMID: 30327408 DOI: 10.1126/scisignal.aau1165
- Ramjiawan RR, Griffioen AW, Duda DG. Anti-angiogenesis for cancer revisited: Is there a role for combinations with immunotherapy? Angiogenesis 2017; 20: 185-204 [PMID: 28361267 DOI: 10.1007/s10456-017-9552-v
- 26 Frezzetti D, Gallo M, Maiello MR, D'Alessio A, Esposito C, Chicchinelli N, Normanno N, De Luca A. VEGF as a potential target in lung cancer. Expert Opin Ther Targets 2017; 21: 959-966 [PMID: 28831824 DOI: 10.1080/14728222.2017.1371137]
- 27 Liang L, Yue Z, Du W, Li Y, Tao H, Wang D, Wang R, Huang Z, He N, Xie X, Han Z, Liu N, Li Z. Molecular Imaging of Inducible VEGF Expression and Tumor Progression in a Breast Cancer Model. Cell Physiol Biochem 2017; 42: 407-415 [PMID: 28558368 DOI: 10.1159/000477485]
- Zhang J, Niu Y, Huang C. Role of FoxM1 in the Progression and Epithelial to Mesenchymal 28 Transition of Gastrointestinal Cancer. Recent Pat Anticancer Drug Discov 2017; 12: 247-259 [PMID: 28440206 DOI: 10.2174/1574892812666170424144352]
- O'Regan RM, Nahta R. Targeting forkhead box M1 transcription factor in breast cancer. Biochem 29 Pharmacol 2018; 154: 407-413 [PMID: 29859987 DOI: 10.1016/j.bcp.2018.05.019]
- 30 Laissue P. The forkhead-box family of transcription factors: key molecular players in colorectal cancer pathogenesis. Mol Cancer 2019; 18: 5 [PMID: 30621735 DOI: 10.1186/s12943-019-0938-x]





Published by Baishideng Publishing Group Inc 7041 Koll Center Parkway, Suite 160, Pleasanton, CA 94566, USA Telephone: +1-925-3991568 E-mail: bpgoffice@wjgnet.com Help Desk: https://www.f6publishing.com/helpdesk https://www.wjgnet.com

