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***Observational Study***

**Polymorphisms in HIF-1a gene are not associated with diabetic retinopathy in China**

Liu YH *et al*. HIF-1a gene polymorphisms in diabetic retinopathy

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**Abstract**

BACKGROUND

Ithas been reported that vascular endothelial growth factor (VEGF) is a susceptibility gene for both type 2 diabetes mellitus (T2DM) and diabetic retinopathy (DR). In response to hypoxia, VEGF mRNA levels are increased, which is mainly mediated by the binding of hypoxia-inducible factor-1 (HIF-1) and hypoxia response element upstream of the transcriptional start site of VEGF. Therefore, HIF-1a is supposed to be involved in pathology of DR.

AIM

To investigate whether the polymorphisms in HIF-1a gene are associated with DR.

METHODS

Two hundred and ninety-nine type 2 diabetic patients (128 males and 171 females) and 144 healthy volunteers were recruited. Mean age was 56.04 ± 21.05 years. According to the results of fundus fluorescein angiography and examination of ophthalmoscopy, patients were divided into two groups, DNR group (diabetes without retinopathy) and DR group (diabetes with retinopathy). There are 150 cases in DNR group and 149 cases in DR group. Two single nucleotide polymorphisms (SNP) of the HIF-1a gene were tested using matrix-assisted laser desorption/Ionization time of flight mass spectrometry. The frequency of genotypes and alleles, and odds ratio were measured.

RESULTS

The mean age of the cases with diabetes was 55.84 ± 3.66 years, the mean age of the cases with DR was 55.97 ± 4.66 years and that of controls was 56.32 ± 4.70 years. Two variations were found in 76 patients. Rs11549465 is the change of C-T base, rs11549467 is the change of G-A base. The rs11549467 G/A genotype was 5.33% in diabetes and 6.04% in DR patients, respectively. The rs11549465 C/T genotype was 10% and 12.75% in patients with diabetes and DR. The rs11549467 A allele frequencies and rs11549465 T frequencies was similar to that of controls. Paired SNP linkage disequilibrium analysis indicated that rs11549467 was in linkage disequilibrium with rs11549465. Haplotype association analysis denoted that the haplotype association exhibited similar distribution in the patients compared to the normal controls.

CONCLUSION

This study suggests that there is no relationship between the genetic variations of HIF1a and diabetes or DR.

**Key Words:** Diabetic retinopathy; Single nucleotide polymorphisms; HIF1a; Vascular endothelial growth factor; Diabetes; Mutation

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**Core Tip:** Vascular endothelial growth factor (VEGF), a potent endothelial cell mitogen, plays a crucial role in neovascularization of proliferative diabetic retinopathy (DR). The human VEGF gene encoding VEGF was identified to be the first target genes of hypoxia-inducible factor-1 (HIF-1). Therefore, HIF-1a is supposed to be involved in pathology of DR. Here, we investigate whether the polymorphisms in HIF-1a gene are associated with DR.

**INTRODUCTION**

Microvascular complications are the principal causes of early mortality in diabetes[1,2]. Most diabetic patients develop diabetic retinopathy (DR) with the progression of diabetes, which remains the major cause of new-onset blindness among diabetic adults[3]. Long duration of diabetes and hypertension along with hyperglycemia are widely considered as main risk factors for the development of DR[4,5]. However, accumulated evidences have revealed that genetic factors may also have a critical impact on the development and progression of DR[6,7]. A great quantity of studies have explored regions of the genome that may contain genes associated with the etiology of DR through using a genome-wide association approach[8-11].

DR is characterized by increased vascular permeability, ischemia and neovascularization[12]. Vitreous hemorrhage and fibrosis, as well as tractional retinal detachment led to a high risk of blindness in the course of neovascularization. Vascular endothelial growth factor (VEGF), a key factor in angiogenesis, can facilitate angiogenesis, increase the permeability of microvessels and promote the formation of collateral vessels[13]. Meanwhile, VEGF plays an essential role in determining mass and property of ß-cell by stimulating angiogenesis of islets in the developmental stage of pancreas[14]. The expression of VEGF mRNA is increased with the activation of transcription in response to hypoxia. This increase is mainly mediated by the binding of hypoxia-inducible factor-1 (HIF-1) and a hypoxia response element upstream of the transcriptional initiation site of VEGF[15,16]. It has been reported that VEGF is a susceptibility gene for both type 2 diabetes mellitus (T2DM) and DR[12].

HIF-1, a transcription factor found in mammalian cells cultured in hypoxic environment, is a heterodimer composed of a HIF-1a subunit complexed with a HIF-1ß subunit[17]. The HIF-1a gene is located at chromosome 14q21-q24[18]. The transcriptional activity and protein expression of HIF-1a are strictly regulated in response to oxygen concentration. Thus, oxygen-regulated expression of the HIF-1a controlled the activity of HIF-1. HIF-1 plays an important role in systemic homeostatic and the increase in expression of VEGF in response to hypoxia.

Therefore, the purpose of the present study was to examine whether there is association of HIF-1a polymorphisms in DR and to assess whether the combined effects of these genetic variations have an influence on the risk for DR.

**MATERIALS AND METHODS**

***Study subjects***

The study was approved by the Clinical Research Ethics Committee of Harbin Medical University and conducted according to the principles in the Declaration of Helsinki. All subjects gave written informed consent. Two hundred and ninety-nine patients with T2DM who were enrolled consecutively between January 2012 and August 2013 were recruited from the Second Affiliated Hospital Ophthalmic Clinic of Harbin Medical University. Of this group, 149 patients were diagnosed with DR. Diabetes was diagnosed according to American Diabetes Association criteria. Control subjects (*n* = 144) were healthy volunteers with negative family history of diabetes and normal fasting glucose (Table 1). Briefly, the patients received a standard questionnaire that include questions about family history, the age of diabetes diagnosis, treatment methods, smoking status and other medical issues. All patients received a complete ophthalmic examination at least once a year, including fundus photography, ophthalmoscopy and corrected visual acuity. Retinopathy was diagnosed based on the Early Treatment Diabetic Retinopathy Study (ETDRS) criteria[19]. Fundoscopic findings were assessed by retinal specialists. In this study, patients were not stratified according to the severity of retinopathy. Patients and controls were matched on gender and age (*χ*2 test, *P* = 0.84).

***Laboratory measurements***

Fasting blood samples were collected in the morning between 9:00 and 10:00 h after overnight fasting. Serum insulin and C-peptide were measured by Commercial RIA kits. Autoanalyzer was used to measure the level of glucose and lipids.

***Deoxyribonucleic acid isolation and genotyping***

Ten milliliter samples of venous blood were collected from the control subjects and the patients with T2DM in EDTA vacutainers (BD, San Jose, CA, United States). Blood Genome DNA Extraction Kits (TAKARA BIOTECHNOLOGY, DALIAN, China ) was used to extract genome DNA from peripheral blood leukocytes. The primers (Table 1) for polymerase chain reaction were designed according to the reference sequences in NCBI Gene database by using Primer 5. Detection of SNP gene of the selected genotypes using matrix - assisted laser desorption / Ionization time of flight mass spectrometry (MALDI-TOF-MS) method (Beijing bo’ao biotechnology company).

***Statistical analysis***

Data of quantitative characteristics are represented as means ± SD. Data of qualitative characteristics are showed as absolute numbers or percent values. Comparisons between groups were carried out *χ2* test (nominal data) or Student’s *t*-test (interval data). The value of *P* < 0.05 was considered as statistical significant. Hardy–Weinberg equilibrium was conducted by the *χ2* method. Univariate and multivariable logistic regression for the association with DR was performed using the SAS program.

**RESULTS**

***Characteristics of participants***

One hundred and fifty healthy controls, 144 patients with DR and 149 patients with diabetes were recruited in this study. The mean age of the cases with diabetes was 55.84 ± 3.66 years, the mean age of the cases with DR was 55.97 ± 4.66 years and that of controls was 56.32 ± 4.70 years. In addition, the patients and controls were matched on gender (Table 1). Age differences were adjusted with logistic regression.

***Identification of single nucleotide polymorphisms and novel mutations***

Two novel nucleotide changes were detected. Rs11549465 is the change of C-T base, rs11549467 is the change of G-A base. These two loci are identical with SNP (rs11549465 and rs11549467) of HIF1A gene in NCBI gene bank (Figure 1). The distributions of all SNPs were in accordance with the Hardy-Weinberg equilibrium (HWE) in the control group (Table 2). Compared with the healthy controls, the rs11549467 G/G genotype and G/A allele was showed with slightly lower frequency in patients with diabetes and DR, while this decrease failed to achieve statistical significance. The rs11549465 C/C genotype and C/T was found with mildly higher frequencies in the patients with diabetes and DR, however, this was not significant different. Table 3 exhibits the distribution of the genotype and allele frequency of rs11549467 and rs11549465.

***Paired single nucleotide polymorphism linkage disequilibrium analysis and haplotype analysis***

As shown in Figure 2, paired SNP linkage disequilibrium analysis demonstrated that rs11549467 was in linkage disequilibrium with rs11549465. The A allele of rs11549467 and the T allele of rs11549465 was found in one diabetic group, however, this failed to achieve statistically significant due to the low frequency. As shown in Table 4, haplotype association analysis denoted that the haplotype association exhibited similar distribution in the patients compared to the normal controls.

**DISCUSSION**

VEGF, a potent endothelial cell mitogen, plays an essential role in neovascularization in proliferative DR. The human VEGF gene encoding VEGF that is essential for the angiogenic process was identified to be the first target genes of HIF-1[20]. The level of VEGF mRNA increases due to the increased transcriptional activation. This increase is mainly mediated by the binding of HIF-1 to a hypoxia response element[21]. VEGF facilitates angiogenesis, increases the permeability of microvessels and promotes the formation of collateral vessels[13]. Besides, it has been reported that VEGF is a susceptible gene of T2DM and DR, leading us to evaluate the association between HIF-1 and T2DM as well as retinopathy[12].

HIF-1, a transcription factor found in mammalian cells cultured in hypoxic environment, act as a heterodimer comprised of a 120-kDa HIF-1a subunit complexed with a 91- to 94-kDa HIF-1β subunit[17]. oxygen-regulated expression of the HIF-1a controlled the activity of HIF-1[22]. HIF-1 plays an important role in systemic homeostatic and the increased expression of VEGF in response to hypoxia.

Expression of HIF-1 Leads to a variety of effects. Some of them may protect against peripheral artery disease, such as formation of collateral vessels or promotion of erythropoiesis. On the other hand, HIF-1 is involved in the vascularization and destabilization of atherosclerotic plaques and may be regarded as a potential risk factor for peripheral artery disease[23,24]. Subsequent gain- or loss-of-function experiments showed that the expression of other angiogenic growth factors, including platelet-derived growth factor B, stromal-derived factor 1, ANGPT2, and placental growth factor was also controlled by HIF-1[25]. Moreover, loss of HIF-1 blunts translocation of GLUT4 and leads to the decrease in glucose uptake by the skeletal muscle cells[26].

Indeed, there is a large number of evidences supporting that the deregulation of HIF-1 in diabetes is related to the failure of cellular adaptation to hypoxia and demonstrating that low oxygen levels by high glucose concentrations results in HIF-1 instability[27]. Catrina observed that high glucose reduced hypoxia-induced function and stabilization of HIF- 1a in human dermal microvascular endothelial cells in culture[28]. Another important founding is that HIF-1β mRNA level was decreased in pancreatic islets in patients with T2DM compared with non-diabetic patients, suggesting that dysfunction of HIF-1 is involved in the etiology of diabetes[29]. In the past years, it has demonstrated that human a HIF-1a P582S genetic polymorphism has protective effects on DR and delays the development of severe DR[30]. Although the molecular mechanisms underlying disorders of HIF-1 in diabetes are still unknown, some studies suppose that diabetes may result in the down-regulation of HIF-1[31]. Additionally, insulin can stimulate HIF- 1a expression through a signaling pathway dependent on the activation of PI3K and the target of rapamycin[32]. This could underlie the amelioration of diabetic complications arising from a blunt cellular response to low oxygen levels after insulin administration. Furthermore, Lin demonstrated that HIF-1a is a key mediator of the main pathological changes in DR, including vascular inflammation, vascular leakage and retinal neovascularization[33].

In this study, we investigate the relationship between HIF-1 genetic polymorphisms and the risk of diabetes and DR in the Chinese population. We did not find any difference in the HIF1 polymorphism frequencies between the diabetes, DR and normal controls. It suggested that functional polymorphisms in the HIF1 gene did not contribute to susceptibility to diabetes or DR. Meanwhile, paired SNP linkage disequilibrium analysis showed that rs11549467 was in linkage disequilibrium with rs11549465 and the haplotype association including the G allele of rs11549467 and the C allele of rs11549465, though exhibited similar distribution in patients with controls.

**CONCLUSION**

This study investigated the association between HIF-1, diabetes and DR in Chinese population. No effects of above two polymorphisms have been observed on diabetes nor DR. However, further studies are needed to elucidate the mechanism underlying HIF-1 deregulation in diabetes and DR.

**ARTICLE HIGHLIGHTS**

***Research background***

Vascular endothelial growth factor (VEGF) mRNA levels are increased response to hypoxia, which is mainly mediated by the binding of hypoxia-inducible factor-1 (HIF-1) and hypoxia response element upstream of the transcriptional start site of VEGF. Therefore, HIF-1a is supposed to be involved in pathology of diabetic retinopathy (DR).

***Research motivation***

In order to find causes associated with DR.

***Research objectives***

In order to investigate whether the polymorphisms in HIF-1a gene are associated with DR.

***Research methods***

Two hundred and ninety-nine type 2 diabetic patients (128 males and 171 females) and 144 healthy volunteers were recruited to 2 groups, DNR group (diabetes without retinopathy) and DR group (diabetes with retinopathy). There are 150 cases in DNR group and 149 cases in DR group. Two single nucleotide polymorphisms of the HIF-1a gene were tested using matrix-assisted laser desorption/Ionization time of flight mass spectrometry. The frequency of genotypes and alleles, and odds ratio were measured.

***Research results***

There is no difference in the HIF1 polymorphism frequencies between the diabetes, DR and normal controls. Haplotype association exhibited similar distribution in the patients compared to the normal controls.

***Research conclusions***

This study suggests that there is no relationship between the genetic variations of HIF1a and diabetes or DR.

***Research perspectives***

Further research is needed to illustrate the mechanism underlying deregulation in HIF-1 in diabetes.

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**Footnotes**

**Institutional review board statement:** The study was reviewed and approved by the Medical Ethics Review Committee of The Second Affiliated Hospital of Harbin Medical University.

**Informed consent statement:** All study participants, or their legal guardian, provided informed written consent prior to study enrollment.

**Conflict-of-interest statement:** The authors declare no competing interests.

**Data sharing statement:** Participants gave informed consent for data sharing.

**STROBE statement:** The authors have read the STROBE Statement-checklist of items, and the manuscript was prepared and revised according to the STROBE Statement-checklist of items.

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**Figure Legends**



**Figure 1 Polar plot of HIF-1a.**



**Figure 2 Paired single nucleotide polymorphism.** Linkage disequilibrium analysis. Paired single nucleotide polymorphisms linkage disequilibrium analysis revealed that rs11549467 was in linkage disequilibrium with rs11549465 (D` > 0.75).

**Table 1 Baseline characteristics of the study subjects**

|  |  |  |  |  |
| --- | --- | --- | --- | --- |
| **Group** | **Total** | **Male (%)** | **Female (%)** | **Mean age (yr)** |
| Diabetes | 150 | 66 | 84 | 55.84 ± 3.66 |
| Diabetic retinopathy | 149 | 62 | 87 | 55.97 ± 4.66 |
| Controls | 144 | 67 | 77 | 56.32 ± 4.70 |
|  |  |  |  | *P* = 0.84 |

**Table 2 Hardy-Weinberg analysis of rs11549467 and rs11549465**

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
| **Group** | **SNP loci** | **Genotype** | **Actual cases** | **Examples of H-W equilibrium theory** | ***P* value** |
| Diabetes | 11549467 | GG|GA|AA | 142|8|0 | 142.11|7.79|0.11 | 0.7372 |
| 11549465 | CC|CT|TT | 135|15|0 | 135.38|14.25|0.38 | 0.5192 |
| Diabetic retinopathy | 11549467 | GG|GA|AA | 140|9|0 | 140.14|8.73|0.14 | 0.7038 |
| 11549465 | CC|CT|TT | 130|19|0 | 130.61|17.79|0.61 | 0.4058 |
| Controls | 11549467 | GG|GA|AA | 137|7|0 | 137.09|6.83|0.09 | 0.7650 |
| 11549465 | CC|CT|TT | 125|19|0 | 125.63|17.75|0.63 | 0.3967 |

SNP: Single nucleotide polymorphisms.

**Table 3 Distribution of the genotype and allele frequencies of rs11549467 and rs115469465 in the diabetes, diabetic retinopathy and control group**

|  |  |  |  |
| --- | --- | --- | --- |
| **SNP** | **Patient/control** | **Genotype, *n* (%)** | **Allele, *n* (%)** |
| rs11549467 |  | G/G | G/A | A/A | G | A |
|  | Diabetes | 142 (94.67) | 8 (5.33) | 0 (0) | 292 (97.33) | 8 (2.67) |
|  | Diabetic retinopathy | 140 (93.96) | 9 (6.04) | 0 (0) | 289 (96.98) | 9 (3.02) |
|  | Controls | 137 (95.14) | 7 (4.86) | 0 (0) | 281 (97.57) | 7 (2.43) |
|  |  |  |  |  |  | *P* > 0.05 |
| rs11549465 |  | C/C | C/T | T/T | C | T |
|  | Diabetes | 135 (90) | 15 (10) | 0 (0) | 285 (95) | 15 (5) |
|  | Diabetic rentinopathy | 130 (87.25) | 19 (12.75) | 0 (0) | 279 (93.62) | 19 (6.38) |
|  | Controls | 125 (86.81) | 19 (13.19) | 0 (0) | 269 (93.4) | 19 (6.6) |
|  |  |  |  |  |  | *P* > 0.05 |

SNP: Single nucleotide polymorphisms.

**Table 4 Haplotype association analysis**

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
| **rs11549467** | **rs11549465** | **Diabetes frequency** | **Diabetic retinopathy frequency** | **Control frequency** | ***P* value** |
| G | C | 92.535 | 90.604 | 90.972 | - |
| G | T | 4.798 | 6.376 | 6.597 | 0.13 |
| A | C | 2.465 | 3.020 | 2.431 | 0.29 |