

WJD 5th Anniversary Special Issues (2): Type 2 diabetes**Inflammation in diabetic kidney disease**

Patricia M García-García, María A Getino-Melián, Virginia Domínguez-Pimentel, Juan F Navarro-González

Patricia M García-García, María A Getino-Melián, Virginia Domínguez-Pimentel, Juan F Navarro-González, Nephrology Service and Research Unit, University Hospital Nuestra Señora de Candelaria, 38010 Santa Cruz de Tenerife, Spain

Juan F Navarro-González, Spanish National Coordinator of GEENDIAB (Grupo Español para el Estudio de la Nefropatía Diabética) (RedInRen 12/0021/0019, Instituto de Salud Carlos III, Ministerio de Economía y Competitividad), 38010 Santa Cruz de Tenerife, Spain

Author contributions: García-García PM, Getino-Melián MA and Domínguez-Pimentel V reviewed the literature and wrote the first draft of the manuscript; Navarro-González JF conceived the original idea, reviewed the first draft and revised versions, and finalized the manuscript; all authors approved the final version of the manuscript to be published.

Supported by Ministerio de Ciencia e Innovación (Instituto de Salud Carlos III-Fondo de Investigación Sanitaria), No. PI07/0870 and No. PI10/576; Ministerio de Sanidad y Política Social (Dirección General de Terapias Avanzadas y Trasplante), No. TRA-182; Sociedad Española de Nefrología y ACINEF; Research activity by Navarro-González JF is supported by Programa de Intensificación de la Actividad Investigadora, ISCIII/Canarias

Correspondence to: Juan F Navarro-González, MD, PhD, FASN, Nephrology Service and Research Unit, University Hospital Nuestra Señora de Candelaria, Carretera del Rosario 145, 38010 Santa Cruz de Tenerife, Spain. jnavgon@gobiernodecanarias.org
Telephone: +34-92-2602061 Fax: +34-92-2602349

Received: October 4, 2013 Revised: February 24, 2014

Accepted: June 10, 2014

Published online: August 15, 2014

Abstract

Diabetes mellitus entails significant health problems worldwide. The pathogenesis of diabetes is multifactorial, resulting from interactions of both genetic and environmental factors that trigger a complex network of pathophysiological events, with metabolic and hemodynamic alterations. In this context, inflammation has emerged as a key pathophysiology mechanism. New pathogenic pathways will provide targets for prevention or future treatments. This review will focus on the implications of inflammation in diabetes mellitus, with

special attention to inflammatory cytokines.

© 2014 Baishideng Publishing Group Inc. All rights reserved.

Key words: Diabetes; Diabetic nephropathy; Diabetic kidney disease; Inflammation; Cytokines; Oxidative stress

Core tip: Diabetic kidney disease is the main cause of renal insufficiency. This complication results from interactions of genetic and environmental factors that trigger a complex network of pathophysiological events. Inflammation has emerged as a key pathophysiology mechanism with important implications from a therapeutic perspective.

García-García PM, Getino-Melián MA, Domínguez-Pimentel V, Navarro-González JF. Inflammation in diabetic kidney disease. *World J Diabetes* 2014; 5(4): 431-443 Available from: URL: <http://www.wjgnet.com/1948-9358/full/v5/i4/431.htm> DOI: <http://dx.doi.org/10.4239/wjd.v5.i4.431>

INFLAMMATION IN DIABETIC KIDNEY DISEASE

Diabetes mellitus (DM) is one of the most significant health problems worldwide. According to the projections, the number of adult diabetic patients will be higher than 430 million in 2030. Diabetic kidney disease (DKD) is one of the most prevalent complications, and is now the leading cause of end-stage renal disease (ESRD) in developed countries^[1,2]. In the general population, ESRD rate increases due to the rise of diabetes mellitus. However, a recent study by Burrows *et al*^[3] found that the incidence of ESRD in the diabetic population had shown a reduction, suggesting that the strategies for controlling DKD, including early diagnosis, adequate control targets and follow-up, early initiation of therapy, and the use of

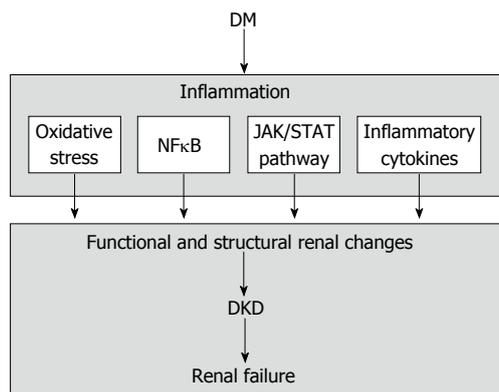


Figure 1 Schematic representation of inflammatory-mediated renal injury in diabetic kidney disease. DM: Diabetes mellitus; NFκB: Nuclear factor κB; DKD: Diabetic kidney disease; JAK/STAT: Janus kinase/signal transducers and activators of transcription.

effective renoprotective therapies, may be efficacious. However, it might be premature to state a real decline in ESRD in diabetes, since other reasons may be possible, such as the lack of enough time to develop ESRD in a large proportion of new diabetic subjects diagnosed in the last 20 years. In addition, the change of the diagnostic criteria for diabetes by the ADA in 1997, may have derived in the diagnosis of diabetes in a earlier stage of the disease, with a much less organ damage, and therefore, when diabetes have a more prolonged evolution, it is possible that this trends in the incidence of ESRD secondary to diabetes may reverse. Finally, another factor is the longer survival of diabetic patients, and thus, these subjects would have an increased risk of developing renal damage and ESRD.

Although kidney biopsy is required to definitively establish the diagnosis of DKD, in clinical practice this is unusual, since the careful screening of patients allow to identify people with DKD. The main criteria to diagnose DKD is the presence of an increased urinary albumin excretion (UAE), which is divided arbitrarily into microalbuminuria and macroalbuminuria, which is associated with an increased risk of decline in glomerular filtration rate (GFR) and a high risk of kidney failure.

DKD has been classically considered as the consequence from the interaction between hemodynamic and metabolic factors. However, renal damage is not completely explained by these factors. Current knowledge indicates that this represents only a partial view of a much more complex scenario. Clear evidence indicates that the pathogenesis of DKD is multifactorial, with the interaction of both genetic and environmental factors that trigger a complex network of pathophysiological events^[4,5]. Clinical observations and epidemiological studies in different ethnic groups have indicated that there is familial aggregation of DKD. Although this information does not allow clearly establishing a model of transmission, diabetic nephropathy has been widely considered as a polygenic disease. There may be many genes, and each has a cumulative genetic effect and interacts with environmental factors in the development of DKD. The

challenge in genetic studies of diabetic nephropathy is to dissect its genetic complexity. Researchers have searched for the genes involved in susceptibility, resistance or progression to DKD. The aim of genetic studies is to provide useful information for better understanding the pathogenesis and further developing novel therapeutic approach in this disease. Genome wide linkage analyses, candidate gene population association, family-based association and genome wide association studies have been used for the identification of the genes in DKD.

In this context, inflammation has become a cardinal pathophysiological mechanism in the development and progression of DKD. This review will focus on the implications of inflammation in DKD, with special attention to inflammatory cytokines.

INFLAMMATION IN DIABETES MELLITUS

Growing evidence indicates that pathogenesis of diabetes mellitus is widely related to the activation of the innate immune system and the presence of a chronic subclinical low-grade inflammatory state^[6,7]. Many studies suggest that individuals who developed DM present characteristics of inflammation several years before the diagnosis of DM^[8,9]. Population-based studies have shown that diverse inflammatory markers, such as cytokines, are strong predictors of the development of diabetes^[10-12]. In addition, inflammatory cytokines have been involved in the pathogenesis of microvascular diabetic complications, including DKD^[13-18].

DKD: AN INFLAMMATORY-BASED COMPLICATION

DM is associated with multiple deviations from normal homeostasis, including hemodynamic and metabolic alterations that produce the activation of diverse transduction pathways in the kidney. At the present time, inflammation is recognized as an important mechanism in the pathogenesis of this complication, through oxidative stress, transcription factors, including nuclear factor κB (NFκB), janus kinase/signal transducers and activators of transcription (JAK/STAT) pathway, and inflammatory cytokines^[13,14] (Figure 1).

OXIDATIVE STRESS

There is solid experimental evidence of a key role for reactive oxygen species (ROS) and oxidative stress and their interplay with the renin-angiotensin-aldosterone system (RAAS) and inflammation, in the pathogenesis of DKD. There is a disproportionate production of ROS secondary to hyperglycemia by different renal cells^[19-25]. Nuclear factor erythroid 2-related factor 2 (Nrf2) is a transcription factor that participates importantly in the regulation of the cellular antioxidant response^[26,27]. Nrf2 appears to counteract renal damage in diabetes, possibly through inhibition of transforming growth factor-β1 (TGF-β1).

In both *in vitro* and *in vivo* experimental studies, Nrf2 ameliorated streptozotocin-induced renal damage. Nrf2(-/-) mice produced greater amounts of ROS and suffered more severe oxidative renal damage compared with wild type mice^[28].

NFκB

NFκB is a transcription factor that controls the expression of genes involved in different processes, such as the immune response, cell differentiation and development, apoptosis, cycle progression, inflammation, and tumorigenesis. Importantly, this factor is activated by many stimuli related to DKD^[29]. Many of the signalling molecules that produce the activation of NFκB may be potential targets for the inhibition of this factor, some of them acting within a network of signals leading to the activation of NFκB.

NFκB is continuously present in cells in an inactive state. In resting cells, NFκB dimers are cloistered by inhibitors of NFκB (IκBs), which prevents the translocation of NFκB to the nucleus. Triggering of the NFκB signalling cascade results in degradation of IκBs, allowing the liberation of NFκB, and thus, this factor translocates to the nucleus and induces transcription. IκB can be classified into several groups: classical IκB (IκBα, IκBβ and IκBε), NFκB precursors (p105 and p100) and nuclear IκB (IκBζ, Bcl-3 and IκBNS). All of them have a central ankyrin repeat domain (ARD), which permits the interaction with NFκB. The activation process of NFκB needs the phosphorylation of IκB, which results in polyubiquitination, a sign for destruction of the IκB by proteasome. The Ser/Thr-specific IκB kinases (IKKs) are the main points for the activation of NFκB. The IKK holo-complex incorporates IKKα or IKKβ, and the protein NEMO (IKKγ or FIP-3). IKK turning on occurs with phosphorylation of the activation loop Ser residues in the canonical MAP kinase kinase consensus motif SxxxS in the kinase domain. NEMO is crucial for the turning on of IKK since in cells without this protein, IKKα and IKKβ cannot be activated by any of the conventional NFκB activators. IKKβ is a key factor for turning on of the canonical NFκB pathway secondary to inflammation, whereas IKKα has a critical function in the non-canonical NFκB pathway through the phosphorylation of p100.

Different extracellular signals initiate the activation of NFκB. After entering the nucleus, this factor interacts with specific sequence motifs (κB sites) on their target genes, resulting in transcriptional turning on. The particular DNA-binding site characteristics of diverse NFκB dimers for a group of related κB sites, and the specific protein-protein binding at target promoters explain the specificity of NFκB signaling. In the majority of instances, turning on of NFκB is temporary and cyclical under the existence of a continuous inducer. This cyclical characteristic is secondary to recurrent destruction and production of IκB and the resulting turning on and inactivation of NFκB, respectively.

NFκB regulates a huge variety of target genes, including those coding for adhesion molecules, chemokines, inflammatory cytokines, nitric oxide synthase, and other molecules related to inflammation and proliferation, all of them involved in the pathogenesis of DKD^[30]. NFκB is activated by a wide variety of stimuli^[31] such as cytokines, oxygen radicals, inhaled particles, ultraviolet irradiation, bacterial or viral products, and metabolic abnormalities. High glucose may produce the activation of NFκB in diverse cells, including endothelial and vascular smooth muscle cells, and cells of the proximal tubule^[32,33]. NFκB is central in the interplay among the different factors, molecules and pathways resulting in structural alterations and functional abnormalities observed in DKD, such as activation of the RAAS, advanced glycation end-products accumulation, and NADPH-dependent oxidative stress^[34]. In experimental models of DKD, it has been established the activation of NFκB in the renal cortical tissue^[35]. Moreover, in human DKD, proteinuria itself, is an important activator of NFκB and it's an important pro-inflammatory stimulus for tubular cells. Chemoattractants and adhesive molecules for inflammatory cells are upregulated by excess ultrafiltered protein load of proximal tubular cells *via* activation of NFκB-dependent and NFκB-independent pathways^[36].

NFκB represents a central factor in inflammation, with the generation of intricated regulatory circuits that include a huge variety of cellular mediators, such as adhesion molecules, intracellular second messengers, microRNA, growth and transcription factors, and cytokines. NFκB system is critical for the flow of biological messages from DNA information to protein synthesis. In addition, these elements have important pathogenic and pathophysiological roles in human disease, including DKD.

JAK/STAT PATHWAY

In animal models and in clinical studies in DKD, it has been demonstrated the enhanced activation of JAK/STAT pathway in the glomeruli and tubulointerstitial cells. The JAK proteins are intracellular, non receptor tyrosine kinases that transduce cytokine-mediated signals. Secondary to the binding of the ligand to the cytokine receptor, the JAK proteins associated with the intracellular domain of the receptor, phosphorylate and activate each other. The autophosphorylation of the JAK proteins induces a conformational modification, allowing the transduction of the intracellular signal by further phosphorylating and activating the STAT transcription factors. The activated STAT molecules dissociated from the receptor and form dimers and translocate to the cell nucleus, where they activate many target genes. The JAK/STAT signaling route is a major connecting system between the receptors located at the cell surface and the transcriptional events occurring within the cell nucleus.

It has been demonstrated the great importance of the JAK/STAT pathway in the pathogenesis of DKD through its participation in several processes, such as the

hypertrophy of mesangial cells induced by angiotensin II (Ang II), and the synthesis of TGF- β , collagen IV and fibronectin. In addition, the high levels of glucose stimulate the production of ROS within the cells, which in turn activates the JAK/STAT pathway.

Although there are several types of JAK proteins, the one primarily studied in renal and vascular tissue is JAK2^[57]. Experimental studies in animal models of diabetic nephropathy have showed that hyperglycemia is able to turning on the JAK2/STAT pathway in renal cells^[38-42]. Moreover, clinical studies in patients with early of advanced stages of DKD have showed an increased expression of JAK/STAT mRNAs and JAK2 protein in the glomerular and tubulointerstitial compartment, with an inverse correlation between JAK2 mRNA levels and estimated GFR in these patients^[43].

The intimate mechanism by which hyperglycemia promotes JAK2 activation has been related to the interaction between JAK2 and ROS caused by high glucose. ROS enhance the activity of JAK2, whereas the use of an inhibitor of ROS formation (diphenylene iodonium) resulted in a marked inhibition of Ang II-induced activation of JAK2. These facts reveal that ROS act as an intracellular activator of the JAK-STAT pathway, and that ROS also act as a second messenger for the regulation of JAK2 activation by Ang II. One of the leading causes of the increased JAK2 tyrosine phosphorylation is the alteration of tyrosine phosphatases (SHP-1 and SHP-2). SHP-1 phosphorylation is abolished under hyperglycemia, whereas SHP-2 phosphorylation is increased under basal and Ang II stimulation, suggesting that JAK2 sustained activation under hyperglycemia is partly due to decreased SHP-1 and increased SHP-2 phosphorylation. In addition, these effects are due to hyperglycemia and not to hyperosmolarity, since no alterations in the tyrosine phosphorylation of both SHP-1 and SHP-2 have been observed under conditions with elevated osmolarity without hyperglycemia^[38-41].

INFLAMMATORY CYTOKINES

Cytokines are low molecular weight polypeptides with autocrine, paracrine and juxtacrine effects, and very complex activities. The classic function of cytokines is related to the regulation of the inflammatory process, but they are also crucial effectors of the immune system. Cytokines often have multiple target cells and multiple pleiotropic actions, and thus a particular cytokine may activate diverse reactions based on the type of cell, the time of action, and the situation and ambience. Moreover, cytokines may share receptor subunits and intracellular signaling pathways, and they can act synergistically in many contexts^[44].

The first studies suggesting that inflammatory cytokines were engaged in the pathogenesis of DKD were published more than 20 years ago by Hasegawa *et al.*^[45,46]. The authors reported that glomerular basement membranes (GBM) obtained from rats after the induction

of diabetes, were able to induce the production of significantly higher quantity of the inflammatory cytokines tumor necrosis factor (TNF)- α and interleukin 1 (IL-1) when were incubated with peritoneal macrophages, as compared with the production of those cytokines when the macrophages were cultured with membranes from normal rats. Later works showed that all types of resident renal cells, as well as infiltrating cells (monocytes, macrophages and lymphocytes) are able to synthesize proinflammatory cytokines^[47,48]. Nowadays, the results of numerous studies support the notion that cytokines play a transcendent role in the pathogenesis of microvascular complications of DM^[13,49,50]. The renal effects of cytokines in DKD are associated with different actions, including intrarenal hemodynamic alterations, modifications of the renal structure with changes in extracellular matrix and basement membranes, abnormalities in the expression of diverse molecules, cellular necrosis and apoptosis, modification in the permeability of glomerular endothelium, and increment in the production of ROS^[50-54].

IL-1

In experimental models of DKD, renal expression of IL-1 is elevated^[55,56], which has been associated with changes in the expression of molecules related to chemotaxis and cellular adhesion. Specifically, IL-1 augments the production of intercellular adhesion molecule 1 (ICAM-1) and vascular cell adhesion molecule 1 by different renal cells, including endothelial, mesangial and tubular epithelial cells. In addition, IL-1 also stimulates the expression of endothelial-leukocyte adhesion molecule 1^[57,58].

IL-1 produces abnormalities of intraglomerular hemodynamics. These effects are secondary to modifications in the synthesis of prostaglandins by mesangial cells. Experimental *in vitro* studies have shown that glomerular mesangial cells incubated with recombinant human IL-1 are stimulated to produce prostaglandin E2 and delivery phospholipase A2^[51]. Furthermore, these cells present an increased secretion of prostaglandin E2 in response to Ang II^[52], whereas the permeability of vascular endothelial cells is enhanced^[59]. Finally, this cytokine raises the production of hyaluronan by epithelial cells of renal proximal tubule^[60], which has been related with the development of hypercellularity in experimental models of diabetes^[61].

IL-6

Clinical studies have shown that IL-6 levels are significantly higher in patients with DKD in comparison with DM patients without nephropathy^[62]. In addition, the histopathological analysis of human renal samples by immunohistochemistry has demonstrated an increased expression of mRNA encoding IL-6 in cells infiltrating the mesangium, interstitium and tubules, with a positive relationship with the severity of mesangial expansion^[63]. Other functional and structural abnormalities related to DKD and progression of renal damage have been associ-

ated with IL-6, including abnormalities in the permeability of glomerular endothelium, expansion of mesangial cells and enhanced expression of fibronectin^[54] and increase in the thickness of the GBM^[64,65]. Our experimental studies have demonstrated an increase in the mRNA levels of IL-6 in the renal cortex of diabetic rats, which is positively associated with the urinary concentration of this cytokine^[56]. In addition, in animal models of diabetes, wet kidney weight, a marker of renal hypertrophy and an early phenomenon in kidney involvement in DM^[66], has been reported to be enhanced, which was related to mRNA gene expression levels and urine concentration of this cytokine^[56].

IL-6 signals through a cell surface receptor, which is formed by the ligand-binding IL-6 receptor (IL-6R)- α chain (CD126) and the signal-transducing component CD130, also called gp130. In addition to the membrane form of the IL-6R, there is a soluble form which is produced by cleavage of the membrane-bound form. These soluble form of the IL-6R comes to the circulation and is able to control the activity of this cytokine. Regarding this regulatory process, it is important to differentiate the actions of soluble CD126 and CD130. In plasma, soluble CD126 binds to IL-6 and results in the increase of the complex half-life, amplifying the bio-activity of this cytokine to tissues that express the membrane form of CD130. On the contrary, soluble form of CD130 in the circulation functions as an IL-6 antagonist. Recent studies have shown that the soluble form of the IL-6R is closely implicated in the evolution from the initial to the final stages of the inflammatory reaction. IL-6 has many biological properties, including the activation of the STAT3 transcription factor, and the induction of the expression of adhesion molecules and other inflammatory cytokines.

IL-18

IL-18, a potent inflammatory cytokine that belongs to the IL-1 superfamily^[67,68], is implicated in different actions, including the release of interferon (IFN)- γ ^[69] (which stimulates functional chemokine receptor expression in human mesangial cells)^[70], the synthesis of other molecules involved in the inflammatory reaction, such as IL-1 and TNF- α , the increase in the expression of ICAM-1, and the apoptotic process of endothelial cells^[71-73]. Tubular renal cells show an increase in the expression of IL-18 in patients with DKD^[74], which has been related to the triggering of mitogen-activated protein kinase (MAPK) pathways secondary to the action of TGF- β ^[75]. Many other cells may also produce this cytokine, such as infiltrating monocytes, macrophages and T cells^[67,68]. High levels of IL-18 has been found in serum and urine of patients with DKD, with an independent relationship with UAE^[76-78]. In addition, serum IL-18 levels are associated with the urine concentration of β -2 microglobulin, a low-weight protein that is used as a marker of tubular dysfunction^[77]. In a recent longitudinal study in patients with type 2 diabetes, serum and urinary levels of IL-18 were direct and independently associated with UAE. In

addition, the concentrations of this cytokine in serum and urine were also significantly associated with changes in albuminuria during the evolution of the study^[77].

TNF- α

TNF- α is a cytokine with prominent proinflammatory effects. It is mainly produced by monocytes, macrophages and T cells, but also intrinsic kidney cells^[47,79-81]. TNF- α exists in the cells as a precursor of the active form. This precursor is transformed in the active form through the action of the TNF- α -converting enzyme^[82]. There are two specific TNF- α receptors: the TNF- α receptor 1 (TNFR1), an epithelial-cell receptor also named p55, and the TNFR2, which is an myeloid-cell receptor (p75). The exact roles of the receptors are not yet completely understood and may differ depending on the organ type^[83]. While TNFR1 modulates the immune response (IL-6 synthesis) and apoptosis (apoptotic signaling kinase 1 and NF κ B of mesangial cells), TNFR2 has been recognized as one of the proinflammatory mediators in glomerulonephritis^[84,85]. After binding to these receptors, the intracellular transduction cascade is activated, leading to the final biological actions of this cytokine^[86], with a potential role in the pathogenesis of DKD. Experimental studies in animal models of diabetes have showed that TNF- α levels and mRNA encoding TNF- α are enhanced in renal glomeruli and tubules^[47,56,80,87-89].

TNF- α may cause direct cytotoxicity to renal cells, inducing direct renal injury^[90], apoptosis and necrotic cell death^[91,92]. It can also produce alterations of intraglomerular blood flow and reduction of glomerular filtration as consequence of the disequilibrium between factors promoting vasoconstriction and vasodilation^[93], in addition to changes in the permeability of endothelial cells. Other actions of this cytokine are the modification in the location of molecules involved in the adhesion process among cells, such as the endothelial-cadherincatenin complexes, as well as the alteration of normal endothelial permeability due to alterations of cellular junctions secondary to the lack of F-actin stress fibers^[94]. In addition, TNF- α is able to directly induce the formation of ROS by renal cells^[95]. Experimental researches has shown that TNF- α induces the activation of NADPH oxidase in isolated rat glomeruli through the activation of the intracellular pathways protein kinase C/phosphatidylinositol-3 kinase and MAPK^[96]. Thus, TNF- α prompts local ROS production, independent of hemodynamic mechanisms, resulting in alterations of the glomerular capillary wall and consequently increased albumin permeability^[53].

An increase in renal size (kidney hypertrophy) and glomerular filtration rate (hyperfiltration) are early and relevant findings of DKD, which are significantly related to TNF- α ^[88,89]. *In vitro* studies demonstrated that TNF- α stimulates the solute uptake in proximal tubular cells secondary to the activation of sodium-dependent cotransporters^[97], whereas *in vivo* studies in diabetic rats found an enhanced urinary excretion of TNF- α excretion, which was related to sodium retention and renal hypertrophy.

All these effects could be blocked by the use of a soluble TNF- α receptor fusion protein^[89,97]. In the renal distal tubule TNF- α activates the epithelial sodium channel resulting in an increased reabsorption of sodium, which can be abrogated by blockers of this renal channel, such as amiloride, and inhibitors of extracellular signal related protein kinase. The increment in renal sodium reabsorption might induce the expression of TFG- β , with the development of renal hypertrophy^[98].

Expression mRNA levels in the renal cortex and urinary TNF- α excretion show a positive and independent correlation with albuminuria^[56,87]. Moreover, microdialysis studies showed that the concentration of TNF- α in the kidney interstitial fluid is elevated, as well as in the urine, with no data of cellular renal infiltration. These findings are observed previously to the detection of an increase in UAE. In addition, there is an elevation in the levels of TNF- α in urine after the increase in UAE, which suggest that the rise of albuminuria has a stimulatory effect in the production of TNF- α by the kidney^[99]. These findings support the intimate relationship between proteinuria and inflammation. Current data indicates that proteinuria per se is an important factor in the development of tubulointerstitial damage, but also by the capacity of activate an inflammatory cellular response *via* chemoattractants, adhesive molecules and proinflammatory cytokines. These changes lead to the renal infiltration by blood circulating cells, with the subsequent damage to renal cells, damage of tubular and interstitial structures, and finally, to the development of renal fibrosis and scarring^[100].

Finally, many clinical studies in patients with DKD have reported that the serum and urinary concentrations of TNF- α are elevated as compared with non-diabetic individuals or with diabetic subjects and kidneys, and that these concentrations increase concomitantly with the progression of DKD. These findings indicate a potential relationship between the elevated levels of this inflammatory cytokine and the development and progression of renal injury in DM^[76,101,102].

In addition to TNF- α , also TNF- α receptors have been related to DKD. In an observational study in type 1 diabetic patients, the serum levels of TNFR1 and TNFR2 were linked with renal function with independence of other variables, such as albuminuria, supporting the important participation of this cytokine in DKD^[103]. In addition, this involvement has also been found in type 2 DM (T2DM). Thus, after more than 10 years of follow-up, the Nurses' Health Study showed that increased concentrations of the soluble TNFR2 were a powerful predictor of the loss of renal function in these patients^[104].

Finally, are also important the findings derived from studies focused on another cytokine within the TNF superfamily, the TNF- α -related apoptosis-inducing ligand (TRAIL). TRAIL participates in diverse cellular processes, including apoptosis, cell expansion and maturity^[105]. Clinical studies in patients with diabetes have shown that the renal expression of this cytokine is enhanced, and more importantly, the grade of expression is directly re-

lated with the seriousness of kidney injury^[106]. Regarding the cell types that express TRAIL, immunohistochemistry studies demonstrated that the renal expression of this cytokine was maximal in tubular epithelial cells. However, it is important to highlight that the expression of TRAIL has been also observed in podocytes^[106,107]. It has been suggested the participation of TRAIL in the pathogenesis of DKD based on the finding that the magnitude of renal tissue staining for this cytokine was directly associated with the grade of tubulointerstitial inflammation, scarring and degeneration.

INFLAMMATION IN DKD: A THERAPEUTIC OPPORTUNITY

Established therapeutic strategies for prevention and treatment of DKD focus on blood pressure and glucose control, RAAS blockade and anti-thrombotic/-inflammatory treatment with aspirin. However, these therapies are insufficient^[108] and new approaches are required^[109].

Oxidative stress

In experimental models, the administration of different antioxidant drugs (tempol, thiol, kallistatin)^[110-112] improved oxidative stress-induced renal injury, decreasing albuminuria and fibrosis. Triterpenoids, synthetic analogues of oleanolic acid with potent anti-inflammatory and antioxidant properties, activate the ARE-Keap1-Nrf2 pathway.

The renoprotective action of bardoxolone methyl, a triterpenoid that reduces oxidative stress and inflammation through Nrf2 activation and inhibition of NF κ B, has been recently explored in humans. A large multicenter double-blind, randomized trial (BEAM study), including 227 patients with moderate-severe CKD and T2DM, showed that administration of bardoxolone was associated with significantly improvement of GFR at 24 wk, but some adverse events were found (mild reversible increase of albuminuria, decreased serum magnesium, muscle spasms, nausea and loss of body weight)^[113]. Later, the BEACON trial, a multinational, multicentric and double-blind randomized, placebo-controlled Phase 3 trial, was designed to determine whether bardoxolone would have beneficial effects on the progression of renal injury and the hazard of ESRD in subjects with T2DM and severe stages of renal disease. Regrettably, the increased risk of heart failure and cardiovascular events observed in the bardoxolone arm of the BEACON study led to the premature ending of this trial^[114].

The most commonly reported serious adverse event in the bardoxolone group was heart failure. The mechanism linking bardoxolone methyl to heart failure is unknown, although some aspects deserve consideration. Firstly, body weight declined significantly in the bardoxolone methyl group, which may suggest a situation of hemodilution secondary to fluid retention, since a reduction in the serum albumin and hemoglobin concentrations was observed. Secondly, it was observed an increase in

blood pressure in the bardoxolone arm, which might result in an elevation of cardiac afterload. This fact, together with the increase in heart preload secondary to fluid retention, combined with a rise in heart rate, result in a situation likely to trigger heart failure. This hypothesis is congruent with the increase in the concentration of B-type natriuretic peptide with bardoxolone methyl, which may reflect an elevated left ventricular wall stress.

NF κ B

The renoprotective effects conferred by blockade of RAAS, provides pleiotropic and anti-inflammatory issues through the suppression of NF κ B-dependent pathways, beyond the control of blood pressure and proteinuria^[115]. In addition, the beneficial effects on the kidney showed by other drugs, such as thiazolidinediones, have been also associated to a suppressive effect on the activation of this transcription factor^[116,117]. In addition, recent experimental studies indicates that suppression of NF κ B activation by various agents, such as 1,25-dihydroxyvitamin D₃^[118], cilostazol^[119], and curcumin^[120], could lead to amelioration of DKD, suggesting the importance of NF κ B as a therapeutic target of DKD.

JAK/STAT pathway

Studies in experimental animal models of DKD have reported that the use of AG490, a specific tyrosine kinase inhibitor of JAK2, was able to abrogate the elevation of systolic blood pressure^[121] and the increase of UAE^[122]. On the other hand, recent studies have highlighted the role of suppressors of cytokine signaling (SOCS) proteins, a group of molecules that bind and interfere with initiating JAK proteins, and act as intracellular negative regulators of JAK/STAT activation in DKD^[37]. Ortiz-Muñoz *et al.*^[123] demonstrated that high concentrations of glucose were associated *in vitro* with activated JAK/STAT/SOCS in human mesangial and tubular cells. Overexpression of SOCS reversed the glucose-induced activation of this pathway, expression of STAT-dependent genes and cell proliferation. On the other hand, the inoculation of recombinant SOCS1 and SOCS3 adenovirus to diabetic rats resulted in an improvement of renal function at 7 wk, and renal lesions such as mesangial expansion, fibrosis or influx of macrophages were also reduced. However, further research into JAK inhibitors, SOCS expression or SOCS mimetics is required, given the critical immunomodulatory role of this pathway, with possible adverse effects^[37].

Inflammatory cytokines

Experimental works using animal models of both types of DM have revealed probable benefits from the use of immunosuppressive drugs. Mycophenolate mofetil (MMF), an immunosuppressive agent with anti-inflammatory properties, was able to avoid the initiation and progression of glomerular damage and albuminuria in rats with streptozotocin-induced diabetes^[124]. Subsequent works demonstrated that MMF produced a marked re-

duction of proteinuria, as well as the amelioration of both renal glomerular and tubulointerstitial scarring^[125]. All these renoprotective effects did not have any relationship with beneficial changes of hemodynamic or metabolic determinants, suggesting that the benefits probably resulted from its immunosuppressive and anti-inflammatory actions. Thus, it was demonstrated that MMF is able to reduce glomerular and tubulointerstitial inflammatory cell infiltration^[126] and abrogate different processes related to the action of TNF- α , such as the expression of ICAM1, the adhesion of neutrophils to the endothelium, as well as the production and discharge of inflammatory cytokines (IL-6 and TNF- α)^[127-129]. Despite these promising experimental results, immunosuppressive treatments actually are not a current clinical therapeutic option in patients with DKD.

Modulation of inflammatory cytokines, mainly TNF- α , has been evaluated in experimental works, as well as in studies with diabetic patients. In experimental studies, the use of etanercept, a recombinant human soluble TNF- α receptor, was associated with the reduction of the urinary excretion of this cytokine and the avoidance of initial kidney structural injury and renal hypertrophy in experimental models of DKD^[88]. Similarly, the use of the monoclonal anti-TNF- α antibody infliximab on rats with DKD led to a significant reduction in the urine excretion of TNF- α and albuminuria^[130]. At the present time, the use of soluble TNF- α receptors or monoclonal antibodies as therapy for DKD have been not tested in clinical trials. However, pentoxifylline (PTF), a drug used in the treatment of peripheral vascular disease, possesses modulating effects on TNF- α , with significant anti-inflammatory properties that has potential clinical applications as a therapy for DKD.

PTF, a methylxanthine derived with non-specific phosphodiesterase activity, possess significant anti-inflammatory properties: this drug is able to abrogate the transcription of the TNF- α gene and hamper the augmentation of TNF- α mRNA^[131,132], regulate IL-1, IL-6 and IFN- γ , and lessen diverse cell actions related to inflammation, such as activation, adherence and phagocytosis^[133,134]. PTF is able to reduce the generation of profibrotic factors (fibronectin and TGF- β) in human mesangial cells caused by elevated glucose levels, and also it protects these cells from the harmful effects of angiotensin II on matrix proteins^[135]. Furthermore, in animal models of DKD, PTF significantly decreased the width of the GMB, the plastering of podocyte foot processes, and the disappearance of the fenestrations of glomerular endothelium^[136]. In addition, PTF prevents the increased renal expression of the inflammatory cytokines TNF- α , IL-1 and IL-6 secondary to diabetes, resulting in a reduction of UAE, the urinary concentration of these cytokines, as well as a decrease of renal hypertrophy and sodium retention^[56,87,88].

Beyond the results from experimental works, a number of clinical studies have showed that PTF is effective to reduce albuminuria and has potential beneficial effects on renal function in diabetic patients^[137-143]. The antipro-

teinuric action of PTF has been straightly associated with its anti-TNF- α activity. This effect has been demonstrated to affect molecules with a high and a low molecular weight, such as IgG, ceruloplasmin, transferrin, albumin, and α 1-antitrypsine, lysozyme and β 2-microglobulin, respectively^[144]. The reduction of proteinuria after PTF administration has been confirmed in various prospective, controlled, randomized clinical studies^[144-146]. Furthermore, PTF has showed beneficial effects on the urinary excretion of markers of tubular damage, such as N-acetylglucosaminidase^[145]. The effectiveness of PTF to reduce urinary protein excretion has been compared with that of angiotensin-converting enzyme inhibitors (ACEI) in T2DM, and the results reveal that PTF is similar to captopril^[144,145]. Moreover, the use of PTF on top of blockade of the RAAS with ACEI or angiotensin II receptor blockers, provide a supplementary and synergistic decrease of albuminuria^[147,148], an effect not related to blood pressure and metabolic control, but positive and directly related with a lowering in the urinary concentration of TNF- α ^[147].

The capacity of PTF to reduce UAE in subjects with DKD has been confirmed by a recent meta-analysis, which highlighted that the anti-inflammatory properties of this drug, with a decrease in the generation of proinflammatory cytokines, was the main potential mechanism to explain its antiproteinuric effect^[149]. A prospective, randomized clinical trial is now ongoing to evaluate the effects of PTF on the renal function of patients with DKD^[150], and new definitive trials (multicentre, adequately powered, prospective, placebo controlled) are needed to give definitive evidence for the use of PTF as a real option for the treatment of DKD.

CONCLUSION

Diabetes mellitus is a major global health problem. DKD is one of the most important complications and constitutes a challenge for physicians. Conventional treatments provide incomplete protection for the development of renal failure. Therefore, new approaches and therapeutic targets are needed. Based on the results of recent studies, nowadays inflammation is acknowledged as a key factor in the development and progression of DKD. Future therapies will focus on modulation of inflammatory pathways, including targets such as inflammatory cytokines, oxidative stress, JAK/STAT pathway, or NF κ B. In addition, further research is needed to understand how inflammatory pathways interact with other pathogenic factors in the context of diabetes.

REFERENCES

- 1 **Ritz E**, Rychlík I, Locatelli F, Halimi S. End-stage renal failure in type 2 diabetes: A medical catastrophe of worldwide dimensions. *Am J Kidney Dis* 1999; **34**: 795-808 [PMID: 10561134]
- 2 **Atkins RC**. The epidemiology of chronic kidney disease. *Kidney Int Suppl* 2005; (**94**): S14-S18 [PMID: 15752232 DOI: 10.1111/j.1523-1755.2005.09403.x]

- 3 **Burrows NR**, Li Y, Geiss LS. Incidence of treatment of end-stage renal disease among individuals with diabetes in the U.S. continue to decline. *Diabetes Care* 2010; **33**: 73-77 [DOI: 10.2337/dc09-0343]
- 4 **Wolf G**. New insights into the pathophysiology of diabetic nephropathy: from haemodynamics to molecular pathology. *Eur J Clin Invest* 2004; **34**: 785-796 [PMID: 15606719 DOI: 10.1111/j.1365-2362.2004.01429.x]
- 5 **Martini S**, Eichinger F, Nair V, Kretzler M. Defining human diabetic nephropathy on the molecular level: integration of transcriptomic profiles with biological knowledge. *Rev Endocr Metab Disor* 2008; **9**: 267-274 [PMID: 18704688]
- 6 **Pickup J**, Crook M. Is type II diabetes mellitus a disease of the innate immune system? *Diabetologia* 1998; **41**: 1241-1248 [DOI: 10.1007/s11154-008-9103-3]
- 7 **Radetti G**, Frizzera S, Castellan C, Mengarda G. [Bone density in swimmers]. *Pediatr Med Chir* 2004; **14**: 521-522 [PMID: 1488310]
- 8 **Bloomgarden ZT**. Inflammation and insulin resistance. *Diabetes Care* 2003; **26**: 1922-1926 [PMID: 12766135 DOI: 10.2337/diacare.26.6.1922]
- 9 **Dandona P**, Aljada A, Bandyopadhyay A. Inflammation: the link between insulin resistance, obesity and diabetes. *Trends Immunol* 2004; **25**: 4-7 [PMID: 14698276 DOI: 10.1016/j.it.2003.10.013]
- 10 **Schmidt MI**, Duncan BB, Sharrett AR, Lindberg G, Savage PJ, Offenbacher S, Azambuja MI, Tracy RP, Heiss G. Markers of inflammation and prediction of diabetes mellitus in adults (Atherosclerosis Risk in Communities study): a cohort study. *Lancet* 1999; **353**: 1649-1652 [PMID: 10335783 DOI: 10.1016/S0140-6736(99)01046-6]
- 11 **Pradhan AD**, Manson JE, Rifai N, Buring J, Ridker P. C-reactive protein, interleukin 6, and risk of developing type 2 diabetes mellitus. *JAMA* 2001; **286**: 327-334 [PMID:11466099 DOI: 10.1001/jama.286.3.327]
- 12 **Spranger J**, Kroke A, Möhlig M, Hoffmann K, Bergmann MM, Ristow M, Boeing H, Pfeiffer AF. Inflammatory cytokines and the risk to develop type 2 diabetes: results of the prospective population-based European Prospective Investigation into Cancer and Nutrition (EPIC)-Potsdam Study. *Diabetes* 2003; **52**: 812-817 [PMID: 12606524 DOI: 10.2337/diabetes.52.3.812]
- 13 **Navarro JF**, Mora C. Role of inflammation in diabetic complications. *Nephrol Dial Transplant* 2005; **20**: 2601-2604 [PMID: 16188894 DOI: 10.1093/ndt/gfi155]
- 14 **Mora C**, Navarro JF. Inflammation and diabetic nephropathy. *Curr Diab Rep* 2006; **6**: 463-468 [PMID: 17118230 DOI: 10.1007/s11892-006-0080-1]
- 15 **Skundric DS**, Lisak RP. Role of neuropoietic cytokines in development and progression of diabetic polyneuropathy: from glucose metabolism to neurodegeneration. *Exp Diabetes Res* 2003; **4**: 303-312 [PMID: 14668051 DOI: 10.1155/edr.2003.303]
- 16 **Jeffcoate WJ**, Game F, Cavanagh PR. The role of proinflammatory cytokines in the cause of neuropathic osteoarthropathy (acute Charcot foot) in diabetes. *Lancet* 2005; **366**: 2058-2061 [PMID: 16338454 DOI: 10.1016/S0140-6736(05)67029-8]
- 17 **Mocan MC**, Kadayifcilar S, Eldem B. Elevated intravitreal interleukin-6 levels in patients with proliferative diabetic retinopathy. *Can J Ophthalmol* 2006; **41**: 747-752 [PMID: 17224958 DOI: 10.3129/i06-070]
- 18 **Demircan N**, Safran BG, Soyulu M, Ozcan AA, Sizmaz S. Determination of vitreous interleukin-1 (IL-1) and tumour necrosis factor (TNF) levels in proliferative diabetic retinopathy. *Eye (Lond)* 2006; **20**: 1366-1369 [PMID: 16284605 DOI: 10.1038/sj.eye.6702138]
- 19 **Debnam ES**, Unwin RJ. Hyperglycemia and intestinal and renal glucose transport: implications for diabetic renal injury. *Kidney Int* 1996; **50**: 1101-1109 [PMID: 8887266 DOI: 10.1038/ki.1996.416]

- 20 **Jeong SO**, Oh GS, Ha HY, Soon Koo B, Sung Kim H, Kim YC, Kim EC, Lee KM, Chung HT, Pae HO. Dimethoxycurcumin, a Synthetic Curcumin Analogue, Induces Heme Oxygenase-1 Expression through Nrf2 Activation in RAW264.7 Macrophages. *J Clin Biochem Nutr* 2009; **44**: 79-84 [PMID: 19177192]
- 21 **Fridlyand LE**, Philipson LH. Oxidative reactive species in cell injury: Mechanisms in diabetes mellitus and therapeutic approaches. *Ann N Y Acad Sci* 2005; **1066**: 136-151 [PMID: 16533924 DOI: 10.1196/annals.1363.019]
- 22 **Kiritoshi S**, Nishikawa T, Sonoda K, Kukidome D, Senokuchi T, Matsuo T, Matsumura T, Tokunaga H, Brownlee M, Araki E. Reactive oxygen species from mitochondria induce cyclooxygenase-2 gene expression in human mesangial cells: potential role in diabetic nephropathy. *Diabetes* 2003; **52**: 2570-2577 [PMID: 14514642 DOI: 10.2337/diabetes.52.10.2570]
- 23 **Lee EA**, Seo JY, Jiang Z, Yu MR, Kwon MK, Ha H, Lee HB. Reactive oxygen species mediate high glucose-induced plasminogen activator inhibitor-1 up-regulation in mesangial cells and in diabetic kidney. *Kidney Int* 2005; **67**: 1762-1771 [PMID: 15840023 DOI: 10.1111/j.1523-1755.2005.00274.x]
- 24 **Thallas-Bonke V**, Thorpe SR, Coughlan MT, Fukami K, Yap FY, Sourris KC, Penfold SA, Bach LA, Cooper ME, Forbes JM. Inhibition of NADPH oxidase prevents advanced glycation end product-mediated damage in diabetic nephropathy through a protein kinase C-alpha-dependent pathway. *Diabetes* 2008; **57**: 460-469 [PMID: 17959934 DOI: 10.2337/db07-1119]
- 25 **Koya D**, Hayashi K, Kitada M, Kashiwagi A, Kikkawa R, Haneda M. Effects of antioxidants in diabetes-induced oxidative stress in the glomeruli of diabetic rats. *J Am Soc Nephrol* 2003; **14**: S250-S253 [PMID: 12874441 DOI: 10.1097/01.asn.0000077412.07578.44]
- 26 **Itoh K**, Ishii T, Wakabayashi N, Yamamoto M. Regulatory mechanisms of cellular response to oxidative stress. *Free Radic Res* 1999; **31**: 319-324 [PMID: 10517536 DOI: 10.1080/10715769900300881]
- 27 **Kensler TW**, Wakabayashi N, Biswal S. Cell survival responses to environmental stresses via the Keap1-Nrf2-ARE pathway. *Annu Rev Pharmacol Toxicol* 2007; **47**: 89-116 [PMID: 16968214 DOI: 10.1146/annurev.pharmtox.46.120604.141046]
- 28 **Jiang T**, Huang Z, Lin Y, Zhang Z, Fang D, Zhang DD. The protective role of Nrf2 in streptozotocin-induced diabetic nephropathy. *Diabetes* 2010; **59**: 850-860 [PMID: 20103708 DOI: 10.2337/db09-1342]
- 29 **Navarro-González JF**, Mora-Fernández C, Muros de Fuentes M, García-Pérez J. Inflammatory molecules and pathways in the pathogenesis of diabetic nephropathy. *Nat Rev Nephrol* 2011; **7**: 327-340 [PMID: 21537349 DOI: 10.1038/nrne-ph.2011.51]
- 30 **Guijarro C**, Egido J. Transcription factor-kappa B (NF-kappa B) and renal disease. *Kidney Int* 2001; **59**: 415-424 [PMID: 11168923 DOI: 10.1046/j.1523-1755.2001.059002415.x]
- 31 **Wada J**, Makino H. Inflammation and the pathogenesis of diabetic nephropathy. *Clin Sci (Lond)* 2013; **124**: 139-152 [PMID: 23075333 DOI: 10.1042/CS20120198]
- 32 **Pieper GM**. Activation of nuclear factor-kappaB in cultured endothelial cells by increased glucose concentration: prevention by calphostin C. *J Cardiovasc Pharmacol* 1997; **30**: 528-532 [PMID: 9335415 DOI: 10.1097/00005344-199710000-00019]
- 33 **Yerneni KK**, Bai W, Khan BV, Medford RM, Natarajan R. Hyperglycemia-induced activation of nuclear transcription factor kappaB in vascular smooth muscle cells. *Diabetes* 1999; **48**: 855-864 [PMID: 10102704 DOI: 10.2337/diabetes.48.4.855]
- 34 **Chuang LY**, Guh JY. Extracellular signals and intracellular pathways in diabetic nephropathy. *Nephrology* 2001; **6**: 165-172 [DOI: 10.1046/j.1440-1797.2001.00043.x]
- 35 **Iwamoto M**, Mizuiri S, Arita M, Hemmi H. Nuclear factor-kappaB activation in diabetic rat kidney: evidence for involvement of P-selectin in diabetic nephropathy. *Tohoku J Exp Med* 2005; **206**: 163-171 [PMID: 15888973 DOI: 10.1620/tjem.206.163]
- 36 **Mezzano S**, Aros C, Droguett A, Burgos ME, Ardiles L, Flores C, Schneider H, Ruiz-Ortega M, Egido J. NF-kappaB activation and overexpression of regulated genes in human diabetic nephropathy. *Nephrol Dial Transplant* 2004; **19**: 2505-2512 [PMID: 15280531 DOI: 10.1093/ndt/gfh207]
- 37 **Brosius FC**, Banes-Berceli A. A new pair of SOCS for diabetic nephropathy. *J Am Soc Nephrol* 2010; **21**: 723-724 [PMID: 20413610 DOI: 10.1681/ASN.2010030286]
- 38 **Li R**, Yang N, Zhang L, Huang Y, Zhang R, Wang F, Luo M, Liang Y, Yu X. Inhibition of Jak/STAT signaling ameliorates mice experimental nephrotic syndrome. *Am J Nephrol* 2007; **27**: 580-589 [PMID: 17823504 DOI: 10.1159/000108102]
- 39 **Marrero MB**, Banes-Berceli AK, Stern DM, Eaton DC. Role of the JAK/STAT signaling pathway in diabetic nephropathy. *Am J Physiol Renal Physiol* 2006; **290**: F762-F768 [PMID: 16527921 DOI: 10.1152/ajprenal.00181.2005]
- 40 **Banes-Berceli AK**, Shaw S, Ma G, Brands M, Eaton DC, Stern DM, Fulton D, Caldwell RW, Marrero MB. Effect of simvastatin on high glucose- and angiotensin II-induced activation of the JAK/STAT pathway in mesangial cells. *Am J Physiol Renal Physiol* 2006; **291**: F116-F121 [PMID: 16449352 DOI: 10.1152/ajprenal.00502.2005]
- 41 **Amiri F**, Shaw S, Wang X, Tang J, Waller JL, Eaton DC, Marrero MB. Angiotensin II activation of the JAK/STAT pathway in mesangial cells is altered by high glucose. *Kidney Int* 2002; **61**: 1605-1616 [PMID: 11967010 DOI: 10.1046/j.1523-1755.2002.00311.x]
- 42 **Wang X**, Shaw S, Amiri F, Eaton DC, Marrero MB. Inhibition of the Jak/STAT signaling pathway prevents the high glucose-induced increase in tgf-beta and fibronectin synthesis in mesangial cells. *Diabetes* 2002; **51**: 3505-3509 [PMID: 12453907 DOI: 10.2337/diabetes.51.12.3505]
- 43 **Berthier CC**, Zhang H, Schin M, Henger A, Nelson RG, Yee B, Boucherot A, Neusser MA, Cohen CD, Carter-Su C, Argentesinger LS, Rastaldi MP, Brosius FC, Kretzler M. Enhanced expression of Janus kinase-signal transducer and activator of transcription pathway members in human diabetic nephropathy. *Diabetes* 2009; **58**: 469-477 [PMID: 19017763 DOI: 10.2337/db08-1328]
- 44 **Vilcek J**. The cytokines: An overview. In: *The Cytokine Handbook*, 4th Ed., Thomson AW, Lotze MT (editors). Academic Press, London, 2003: 3-18 [DOI: 10.1016/B978-012689663-3/50005-3]
- 45 **Hasegawa G**, Nakano K, Sawada M, Uno K, Shibayama Y, Ienaga K, Kondo M. Possible role of tumor necrosis factor and interleukin-1 in the development of diabetic nephropathy. *Kidney Int* 1991; **40**: 1007-1012 [PMID: 1762301 DOI: 10.1038/ki.1991.308]
- 46 **Hasegawa G**, Nakano K, Kondo M. Role of TNF and IL-1 in the development of diabetic nephropathy. *Nefrologia* 1995; **15**: 1-4
- 47 **Sugimoto H**, Shikata K, Wada J, Horiuchi S, Makino H. Advanced glycation end products-cytokine-nitric oxide sequence pathway in the development of diabetic nephropathy: aminoguanidine ameliorates the overexpression of tumour necrosis factor-alpha and inducible nitric oxide synthase in diabetic rat glomeruli. *Diabetologia* 1999; **42**: 878-886 [PMID: 10440132 DOI: 10.1007/s001250051241]
- 48 **Nakamura T**, Fukui M, Ebihara I, Osada S, Nagaoka I, Tomino Y, Koide H. mRNA expression of growth factors in glomeruli from diabetic rats. *Diabetes* 1993; **42**: 450-456 [PMID: 8094359 DOI: 10.2337/diab.42.3.450]
- 49 **Alexandraki K**, Piperi C, Kalofoutis C, Singh J, Alaveras A, Kalofoutis A. Inflammatory process in type 2 diabetes: The role of cytokines. *Ann N Y Acad Sci* 2006; **1084**: 89-117 [PMID: 17151295 DOI: 10.1196/annals.1372.039]
- 50 **Navarro-González JF**, Mora-Fernández C. The role of in-

- flammatory cytokines in diabetic nephropathy. *J Am Soc Nephrol* 2008; **19**: 433-442 [PMID: 18256353]
- 51 **Pfeilschifter J**, Pignat W, Vosbeck K, Märki F. Interleukin 1 and tumor necrosis factor synergistically stimulate prostaglandin synthesis and phospholipase A2 release from rat renal mesangial cells. *Biochem Biophys Res Commun* 1989; **159**: 385-394 [PMID: 2784674 DOI: 10.1016/0006-291X(89)90003-X]
- 52 **Pfeilschifter J**, Muhl H. Interleukin-1 and tumor necrosis factor potentiate angiotensin II- and calcium ionophore-stimulated prostaglandin E2 synthesis in rat renal mesangial cells. *Biochem Biophys Res Commun* 1990; **169**: 585-595 [DOI: 10.1016/0006-291X(90)90371-S]
- 53 **McCarthy ET**, Sharma R, Sharma M, Li JZ, Ge XL, Dileepan KN, Savin VJ. TNF-alpha increases albumin permeability of isolated rat glomeruli through the generation of superoxide. *J Am Soc Nephrol* 1998; **9**: 433-438 [PMID: 9513905]
- 54 **Coleman DL**, Ruef C. Interleukin 6: an autocrine regulator of mesangial cell growth. *Kidney Int* 1992; **41**: 604-606 [DOI: 10.1038/ki.1992.91]
- 55 **Sassy-Prigent C**, Heudes D, Mandet C, Bélair MF, Michel O, Perdureau B, Bariéty J, Bruneval P. Early glomerular macrophage recruitment in streptozotocin-induced diabetic rats. *Diabetes* 2000; **49**: 466-475 [PMID: 10868970 DOI: 10.2337/diabetes.49.3.466]
- 56 **Navarro JF**, Milena FJ, Mora C, León C, García J. Renal proinflammatory cytokine gene expression in diabetic nephropathy: effect of angiotensin-converting enzyme inhibition and pentoxifylline administration. *Am J Nephrol* 2006; **26**: 562-570 [PMID: 17167242 DOI: 10.1159/000098004]
- 57 **Brady HR**. Leukocyte adhesion molecules and kidney diseases. *Kidney Int* 1994; **45**: 1285-1300 [PMID: 8072240 DOI: 10.1038/ki.1994.169]
- 58 **Park CW**, Kim JH, Lee JH, Kim YS, Ahn HJ, Shin YS, Kim SY, Choi EJ, Chang YS, Bang BK. High glucose-induced intercellular adhesion molecule-1 (ICAM-1) expression through an osmotic effect in rat mesangial cells is PKC-NF-kappa B-dependent. *Diabetologia* 2000; **43**: 1544-1553 [PMID: 11151765 DOI: 10.1007/s001250051567]
- 59 **Royall JA**, Berkow RL, Beckman JS, Cunningham MK, Matalon S, Freeman BA. Tumor necrosis factor and interleukin 1 alpha increase vascular endothelial permeability. *Am J Physiol* 1989; **257**: L399-L410 [PMID: 2610269 DOI: 10.1097/0003246-198804000-00075]
- 60 **Jones S**, Jones S, Phillips AO. Regulation of renal proximal tubular epithelial cell hyaluronan generation: implications for diabetic nephropathy. *Kidney Int* 2001; **59**: 1739-1749 [PMID: 11318944 DOI: 10.1046/j.1523-1755.2001.0590051739.x]
- 61 **Mahadevan P**, Larkins RG, Fraser JR, Fosang AJ, Dunlop ME. Increased hyaluronan production in the glomeruli from diabetic rats: a link between glucose-induced prostaglandin production and reduced sulphated proteoglycan. *Diabetologia* 1995; **38**: 298-305 [PMID: 7758876 DOI: 10.1007/BF00400634]
- 62 **Sekizuka K**, Tomino Y, Sei C, Kurusu A, Tashiro K, Yamaguchi Y, Kodera S, Hishiki T, Shirato I, Koide H. Detection of serum IL-6 in patients with diabetic nephropathy. *Nephron* 1994; **68**: 284-285 [PMID: 7830879 DOI: 10.1159/000188281]
- 63 **Suzuki D**, Miyazaki M, Naka R, Koji T, Yagame M, Jinde K, Endoh M, Nomoto Y, Sakai H. In situ hybridization of interleukin 6 in diabetic nephropathy. *Diabetes* 1995; **44**: 1233-1238 [PMID: 7556963]
- 64 **Nosadini R**, Velussi M, Brocco E, Bruseghin M, Abaterusso C, Saller A, Dalla Vestra M, Carraro A, Bortoloso E, Sambataro M, Barzon I, Frigato F, Muollo B, Chiesura-Corona M, Pacini G, Baggio B, Piarulli F, Sfriso A, Fioretto P. Course of renal function in type 2 diabetic patients with abnormalities of albumin excretion rate. *Diabetes* 2000; **49**: 476-484 [PMID: 10868971 DOI: 10.2337/diabetes.49.3.476]
- 65 **Dalla Vestra M**, Mussap M, Gallina P, Bruseghin M, Cernigoi AM, Saller A, Plebani M, Fioretto P. Acute-phase markers of inflammation and glomerular structure in patients with type 2 diabetes. *J Am Soc Nephrol* 2005; **16** Suppl 1: S78-S82 [PMID: 15938041 DOI: 10.1681/ASN.2004110961]
- 66 **Thomson SC**, Deng A, Bao D, Satriano J, Blantz RC, Vallon V. Ornithine decarboxylase, kidney size, and the tubular hypothesis of glomerular hyperfiltration in experimental diabetes. *J Clin Invest* 2001; **107**: 217-224 [PMID: 11160138 DOI: 10.1172/JCI10963]
- 67 **Melnikov VY**, Ecdet T, Fantuzzi G, Siegmund B, Lucia MS, Dinarello CA, Schrier RW, Edelstein CL. Impaired IL-18 processing protects caspase-1-deficient mice from ischemic acute renal failure. *J Clin Invest* 2001; **107**: 1145-1152 [PMID: 11342578 DOI: 10.1172/JCI2089]
- 68 **Melnikov VY**, Faubel S, Siegmund B, Lucia MS, Ljubanovic D, Edelstein CL. Neutrophil-independent mechanisms of caspase-1- and IL-18-mediated ischemic acute tubular necrosis in mice. *J Clin Invest* 2002; **110**: 1083-1091 [PMID: 12393844 DOI: 10.101172/jci0215623]
- 69 **Okamura H**, Tsutsi H, Komatsu T, Yutsudo M, Hakura A, Tanimoto T, Torigoe K, Okura T, Nukada Y, Hattori K. Cloning of a new cytokine that induces IFN-gamma production by T cells. *Nature* 1995; **378**: 88-91 [PMID: 7477296 DOI: 10.1038/378088a0]
- 70 **Schwarz M**, Wahl M, Resch K, Radeke HH. IFN-gamma induces functional chemokine receptor expression in human mesangial cells. *Clin Exp Immunol* 2002; **128**: 285-294 [PMID: 11985519 DOI: 10.1046/j.1365-2249.2002.01829.x]
- 71 **Dai SM**, Matsuno H, Nakamura H, Nishioka K, Yudoh K. Interleukin-18 enhances monocyte tumor necrosis factor alpha and interleukin-1beta production induced by direct contact with T lymphocytes: implications in rheumatoid arthritis. *Arthritis Rheum* 2004; **50**: 432-443 [PMID: 14872485 DOI: 10.1002/art.20064]
- 72 **Mariño E**, Cardier JE. Differential effect of IL-18 on endothelial cell apoptosis mediated by TNF-alpha and Fas (CD95). *Cytokine* 2003; **22**: 142-148 [PMID: 12842762 DOI: 10.1016/S1043-4666(03)00150-9]
- 73 **Stuyt RJ**, Netea MG, Geijtenbeek TB, Kullberg BJ, Dinarello CA, van der Meer JW. Selective regulation of intercellular adhesion molecule-1 expression by interleukin-18 and interleukin-12 on human monocytes. *Immunology* 2003; **110**: 329-334 [PMID: 14632660 DOI: 10.1046/j.1365-2567.2003.01747.x]
- 74 **Fantuzzi G**, Reed DA, Dinarello CA. IL-12-induced IFN-gamma is dependent on caspase-1 processing of the IL-18 precursor. *J Clin Invest* 1999; **104**: 761-767 [PMID: 10491411 DOI: 10.1172/JCI7501]
- 75 **Miyachi K**, Takiyama Y, Honjyo J, Tateno M, Haneda M. Upregulated IL-18 expression in type 2 diabetic subjects with nephropathy: TGF-beta1 enhanced IL-18 expression in human renal proximal tubular epithelial cells. *Diabetes Res Clin Pract* 2009; **83**: 190-199 [PMID: 19110334 DOI: 10.1016/j.diabres.2008.11.018]
- 76 **Moriwaki Y**, Yamamoto T, Shibutani Y, Aoki E, Tsutsumi Z, Takahashi S, Okamura H, Koga M, Fukuchi M, Hada T. Elevated levels of interleukin-18 and tumor necrosis factor-alpha in serum of patients with type 2 diabetes mellitus: relationship with diabetic nephropathy. *Metabolism* 2003; **52**: 605-608 [PMID: 12759891 DOI: 10.1053/meta.2003.50096]
- 77 **Nakamura A**, Shikata K, Hiramatsu M, Nakatou T, Kitamura T, Wada J, Itoshima T, Makino H. Serum interleukin-18 levels are associated with nephropathy and atherosclerosis in Japanese patients with type 2 diabetes. *Diabetes Care* 2005; **28**: 2890-2895 [PMID: 16306550 DOI: 10.2337/diacare.28.12.2890]
- 78 **Wong CK**, Ho AW, Tong PC, Yeung CY, Kong AP, Lun SW, Chan JC, Lam CW. Aberrant activation profile of cytokines and mitogen-activated protein kinases in type 2 diabetic patients with nephropathy. *Clin Exp Immunol* 2007; **149**: 123-131 [PMID: 17425653 DOI: 10.1111/j.1365-2249.2007.03389.x]
- 79 **Jevnikar AM**, Brennan DC, Singer GG, Heng JE, Maslinski W, Wuthrich RP, Glimcher LH, Kelley VE. Stimulated kid-

- ney tubular epithelial cells express membrane associated and secreted TNF alpha. *Kidney Int* 1991; **40**: 203-211 [PMID: 1942768 DOI: 10.1038/ki.1991.201]
- 80 **Nakamura T**, Masai T, Yamauchi T, Higuchi T, Ito H, Toyoshima Y, Sawa Y. Successful surgical management for severe mitral regurgitation unmasked after pericardiectomy for chronic constrictive pericarditis. *Ann Thorac Surg* 2008; **86**: 1994-1996 [PMID: 19022033 DOI: 10.1016/j.athoracsur.2008.05.005]
- 81 **Dong X**, Swaminathan S, Bachman LA, Croatt AJ, Nath KA, Griffin MD. Resident dendritic cells are the predominant TNF-secreting cell in early renal ischemia-reperfusion injury. *Kidney Int* 2007; **71**: 619-628 [PMID: 17311071]
- 82 **Wang X**, Feuerstein GZ, Xu L, Wang H, Schumacher WA, Ogletree ML, Taub R, Duan JJ, Decicco CP, Liu RQ. Inhibition of tumor necrosis factor-alpha-converting enzyme by a selective antagonist protects brain from focal ischemic injury in rats. *Mol Pharmacol* 2004; **65**: 890-896 [PMID: 15044618 DOI: 10.1124/mol.65.4.890]
- 83 **Speeckaert MM**, Speeckaert R, Laute M, Vanholder R, Delanghe JR. Tumor necrosis factor receptors: biology and therapeutic potential in kidney diseases. *Am J Nephrol* 2012; **36**: 261-270 [PMID: 22965073 DOI: 10.1159/000342333]
- 84 **Zhu LJ**, Yang X, Li XY, Liu QH, Tang XQ, Zhou SF, Kong QY, Axelsson J, Yu XQ. Suppression of tumor necrosis factor receptor associated factor (TRAF)-2 attenuates the pro-inflammatory and proliferative effect of aggregated IgG on rat renal mesangial cells. *Cytokine* 2010; **49**: 201-208 [PMID: 19910209 DOI: 10.1016/j.cyto.2009.10.004]
- 85 **Vielhauer V**, Stavarakis G, Mayadas TN. Renal cell-expressed TNF receptor 2, not receptor 1, is essential for the development of glomerulonephritis. *J Clin Invest* 2005; **115**: 1199-1209 [PMID: 15841213 DOI: 10.1172/jci23348]
- 86 **Ortiz A**, Bustos C, Alonso J, Alcázar R, López-Armada MJ, Plaza JJ, González E, Egido J. Involvement of tumor necrosis factor-alpha in the pathogenesis of experimental and human glomerulonephritis. *Adv Nephrol Necker Hosp* 1995; **24**: 53-77 [PMID: 7572422]
- 87 **Navarro JF**, Milena FJ, Mora C, León C, Claverie F, Flores C, García J. Tumor necrosis factor-alpha gene expression in diabetic nephropathy: relationship with urinary albumin excretion and effect of angiotensin-converting enzyme inhibition. *Kidney Int Suppl* 2005; (99): S98-102 [PMID: 16336586]
- 88 **DiPetrillo K**, Coutermarsh B, Gesek FA. Urinary tumor necrosis factor contributes to sodium retention and renal hypertrophy during diabetes. *Am J Physiol Renal Physiol* 2003; **284**: F113-F121 [PMID: 12388406]
- 89 **DiPetrillo K**, Gesek FA. Pentoxifylline ameliorates renal tumor necrosis factor expression, sodium retention, and renal hypertrophy in diabetic rats. *Am J Nephrol* 2004; **24**: 352-359 [PMID: 15205554 DOI: 10.1159/000079121]
- 90 **Bertani T**, Abbate M, Zoja C, Corna D, Perico N, Ghezzi P, Remuzzi G. Tumor necrosis factor induces glomerular damage in the rabbit. *Am J Pathol* 1989; **134**: 419-430 [PMID: 2916653]
- 91 **Laster SM**, Wood JG, Gooding LR. Tumor necrosis factor can induce both apoptotic and necrotic forms of cell lysis. *J Immunol* 1988; **141**: 2629-2634 [PMID: 3171180]
- 92 **Boyle JJ**, Weissberg PL, Bennett MR. Tumor necrosis factor-alpha promotes macrophage-induced vascular smooth muscle cell apoptosis by direct and autocrine mechanisms. *Arterioscler Thromb Vasc Biol* 2003; **23**: 1553-1558 [PMID: 12869351 DOI: 10.1161/01.ATV.0000086961.44581.B7]
- 93 **Baud L**, Perez J, Friedlander G, Ardaillou R. Tumor necrosis factor stimulates prostaglandin production and cyclic AMP levels in rat cultured mesangial cells. *FEBS Lett* 1988; **239**: 50-54 [PMID: 2846348 DOI: 10.1016/0014-5793(88)80543-X]
- 94 **Wójciak-Stothard B**, Entwistle A, Garg R, Ridley AJ. Regulation of TNF-alpha-induced reorganization of the actin cytoskeleton and cell-cell junctions by Rho, Rac, and Cdc42 in human endothelial cells. *J Cell Physiol* 1998; **176**: 150-165 [PMID: 9618155]
- 95 **Radeke HH**, Meier B, Topley N, Flöge J, Habermehl GG, Resch K. Interleukin 1-alpha and tumor necrosis factor-alpha induce oxygen radical production in mesangial cells. *Kidney Int* 1990; **37**: 767-775 [PMID: 2407888 DOI: 10.1038/ki.1990.44]
- 96 **Koike N**, Takamura T, Kaneko S. Induction of reactive oxygen species from isolated rat glomeruli by protein kinase C activation and TNF-alpha stimulation, and effects of a phosphodiesterase inhibitor. *Life Sci* 2007; **80**: 1721-1728 [PMID: 17346751 DOI: 10.1016/j.lfs.2007.02.001]
- 97 **Schreiner GF**, Kohan DE. Regulation of renal transport processes and hemodynamics by macrophages and lymphocytes. *Am J Physiol* 1990; **258**: F761-F767 [PMID: 2184672]
- 98 **Yu HC**, Burrell LM, Black MJ, Wu LL, Dilley RJ, Cooper ME, Johnston CI. Salt induces myocardial and renal fibrosis in normotensive and hypertensive rats. *Circulation* 1998; **98**: 2621-2628 [PMID: 9843472 DOI: 10.1161/01.CIR.98.23.2621]
- 99 **Kalantarinia K**, Awad AS, Siragy HM. Urinary and renal interstitial concentrations of TNF-alpha increase prior to the rise in albuminuria in diabetic rats. *Kidney Int* 2003; **64**: 1208-1213 [PMID: 12969138 DOI: 10.1046/j.1523-1755.2003.0237.x]
- 100 **Abbate M**, Zoja C, Remuzzi G. How does proteinuria cause progressive renal damage? *J Am Soc Nephrol* 2006; **17**: 2974-2984 [PMID: 17035611 DOI: 10.1681/ASN.2006040377]
- 101 **Navarro JF**, Mora C, Maca M, Garca J. Inflammatory parameters are independently associated with urinary albumin in type 2 diabetes mellitus. *Am J Kidney Dis* 2003; **42**: 53-61 [PMID: 12830456 DOI: 10.1016/S0272-6386(03)00408-6]
- 102 **Navarro JF**, Mora C, Muros M, García J. Urinary tumor necrosis factor-alpha excretion independently correlates with clinical markers of glomerular and tubulointerstitial injury in type 2 diabetic patients. *Nephrol Dial Transplant* 2006; **21**: 3428-3434 [PMID: 16935891 DOI: 10.1093/ndt/gfl469]
- 103 **Niewczasz MA**, Ficociello LH, Johnson AC, Walker W, Rosolowski ET, Roshan B, Warram JH, Krolewski AS. Serum concentrations of markers of TNFalpha and Fas-mediated pathways and renal function in nonproteinuric patients with type 1 diabetes. *Clin J Am Soc Nephrol* 2009; **4**: 62-70 [PMID: 19073786]
- 104 **Lin J**, Hu FB, Mantzoros C, Curhan GC. Lipid and inflammatory biomarkers and kidney function decline in type 2 diabetes. *Diabetologia* 2010; **53**: 263-267 [PMID: 19921505]
- 105 **Schneider P**, Thome M, Burns K, Bodmer JL, Hofmann K, Kataoka T, Holler N, Tschopp J. TRAIL receptors 1 (DR4) and 2 (DR5) signal FADD-dependent apoptosis and activate NF-kappaB. *Immunity* 1997; **7**: 831-836 [PMID: 9430228 DOI: 10.1016/S1074-7613(00)80401-X]
- 106 **Lorz C**, Benito-Martín A, Boucherot A, Uceros AC, Rastaldi MP, Henger A, Armelloni S, Santamaría B, Berthier CC, Kretzler M, Egido J, Ortiz A. The death ligand TRAIL in diabetic nephropathy. *J Am Soc Nephrol* 2008; **19**: 904-914 [PMID: 18287563 DOI: 10.1016/18287563]
- 107 **Sanchez-Niño MD**, Sanz AB, Ihalmo P, Lassila M, Holthofer H, Mezzano S, Aros C, Groop PH, Saleem MA, Mathieson PW, Langham R, Kretzler M, Nair V, Lemley KV, Nelson RG, Mervaala E, Mattinzoli D, Rastaldi MP, Ruiz-Ortega M, Martín-Ventura JL, Egido J, Ortiz A. The MIF receptor CD74 in diabetic podocyte injury. *J Am Soc Nephrol* 2009; **20**: 353-362 [PMID: 18842989 DOI: 10.1681/ASN.2008020194]
- 108 **Hostetter TH**. Prevention of end-stage renal disease due to type 2 diabetes. *N Engl J Med* 2001; **345**: 910-912 [PMID: 11565525 DOI: 10.1056/NEJM200109203451209]
- 109 **Williams ME**, Tuttle KR. The next generation of diabetic nephropathy therapies: an update. *Adv Chronic Kidney Dis* 2005; **12**: 212-222 [PMID: 15822057 DOI: 10.1053/j.ackd.2005.01.011]
- 110 **Quiroz Y**, Ferrebuz A, Vaziri ND, Rodriguez-Iturbe B. Effect of chronic antioxidant therapy with superoxide dismutase-

- mimetic drug, tempol, on progression of renal disease in rats with renal mass reduction. *Nephron Exp Nephrol* 2009; **112**: e31-e42 [PMID: 19342872 DOI: 10.1159/000210577]
- 111 **Zhang L**, Fujii S, Igarashi J, Kosaka H. Effects of thiol antioxidant on reduced nicotinamide adenine dinucleotide phosphate oxidase in hypertensive Dahl salt-sensitive rats. *Free Radic Biol Med* 2004; **37**: 1813-1820 [PMID: 15528040 DOI: 10.1016/j.freeradbiomed.2004.08.019]
- 112 **Shen B**, Hagiwara M, Yao YY, Chao L, Chao J. Salutary effect of kallistatin in salt-induced renal injury, inflammation, and fibrosis via antioxidative stress. *Hypertension* 2008; **51**: 1358-1365 [PMID: 18391098]
- 113 **Pergola PE**, Raskin P, Toto RD, Meyer CJ, Huff JW, Grossman EB, Krauth M, Ruiz S, Audhya P, Christ-Schmidt H, Wittes J, Warnock DG. Bardoxolone methyl and kidney function in CKD with type 2 diabetes. *N Engl J Med* 2011; **365**: 327-336 [PMID: 21699484 DOI: 10.1056/NEJMoa1105351]
- 114 **de Zeeuw D**, Akizawa T, Audhya P, Bakris GL, Chin M, Christ-Schmidt H, Goldsberry A, Houser M, Krauth M, Lambers Heerspink HJ, McMurray JJ, Meyer CJ, Parving HH, Remuzzi G, Toto RD, Vaziri ND, Wanner C, Wittes J, Wroblestad D, Chertow GM. Bardoxolone methyl in type 2 diabetes and stage 4 chronic kidney disease. *N Engl J Med* 2013; **369**: 2492-2503 [PMID: 24206459 DOI: 10.1056/NEJMoa1306033]
- 115 **Han SY**, Kim CH, Kim HS, Jee YH, Song HK, Lee MH, Han KH, Kim HK, Kang YS, Han JY, Kim YS, Cha DR. Spiro-nolactone prevents diabetic nephropathy through an anti-inflammatory mechanism in type 2 diabetic rats. *J Am Soc Nephrol* 2006; **17**: 1362-1372 [PMID: 16571782 DOI: 10.1681/ASN.2005111196]
- 116 **Ohga S**, Shikata K, Yozai K, Okada S, Ogawa D, Usui H, Wada J, Shikata Y, Makino H. Thiazolidinedione ameliorates renal injury in experimental diabetic rats through anti-inflammatory effects mediated by inhibition of NF-kappaB activation. *Am J Physiol Renal Physiol* 2007; **292**: F1141-F1150 [PMID: 17190910 DOI: 10.1152/ajprenal.00288.2005]
- 117 **Ko GJ**, Kang YS, Han SY, Lee MH, Song HK, Han KH, Kim HK, Han JY, Cha DR. Pioglitazone attenuates diabetic nephropathy through an anti-inflammatory mechanism in type 2 diabetic rats. *Nephrol Dial Transplant* 2008; **23**: 2750-2760 [PMID: 18388116 DOI: 10.1093/ndt/gfn157]
- 118 **Zhang Z**, Yuan W, Sun L, Szeto FL, Wong KE, Li X, Kong J, Li YC. 1,25-Dihydroxyvitamin D3 targeting of NF-kappaB suppresses high glucose-induced MCP-1 expression in mesangial cells. *Kidney Int* 2007; **72**: 193-201 [PMID: 17507908 DOI: 10.1038/sj.ki.5002296]
- 119 **Lee WC**, Chen HC, Wang CY, Lin PY, Ou TT, Chen CC, Wen MC, Wang J, Lee HJ. Cilostazol ameliorates nephropathy in type 1 diabetic rats involving improvement in oxidative stress and regulation of TGF-Beta and NF-kappaB. *Biosci Biotechnol Biochem* 2010; **74**: 1355-1361 [PMID: 20622454 DOI: 10.1271/bbb.90938]
- 120 **Soetikno V**, Sari FR, Veeraveedu PT, Thandavarayan RA, Harima M, Sukumaran V, Lakshmanan AP, Suzuki K, Kawachi H, Watanabe K. Curcumin ameliorates macrophage infiltration by inhibiting NF-kB activation and proinflammatory cytokines in streptozotocin induced-diabetic nephropathy. *Nutr Metab (Lond)* 2011; **8**: 35 [PMID: 21663638 DOI: 10.1186/1743-7075-8-35]
- 121 **Banes AK**, Shaw S, Jenkins J, Redd H, Amiri F, Pollock DM, Marrero MB. Angiotensin II blockade prevents hyperglycemia-induced activation of JAK and STAT proteins in diabetic rat kidney glomeruli. *Am J Physiol Renal Physiol* 2004; **286**: F653-F659 [PMID: 14678947 DOI: 10.1152/ajprenal.00163.2003]
- 122 **Banes-Berceli AK**, Ketsawatsomkron P, Ogbi S, Patel B, Pollock DM, Marrero MB. Angiotensin II and endothelin-1 augment the vascular complications of diabetes via JAK2 activation. *Am J Physiol Heart Circ Physiol* 2007; **293**: H1291-H1299 [PMID: 17526654 DOI: 10.1152/ajpheart.00181.2007]
- 123 **Ortiz-Muñoz G**, Lopez-Parra V, Lopez-Franco O, Fernandez-Vizcarra P, Mallavia B, Flores C, Sanz A, Blanco J, Mezzano S, Ortiz A, Egido J, Gomez-Guerrero C. Suppressors of cytokine signaling abrogate diabetic nephropathy. *J Am Soc Nephrol* 2010; **21**: 763-772 [PMID: 20185635 DOI: 10.1681/ASN.2009060625]
- 124 **Utamura R**, Fujihara CK, Mattar AL, Malheiros DM, Noronha IL, Zatz R. Mycophenolate mofetil prevents the development of glomerular injury in experimental diabetes. *Kidney Int* 2003; **63**: 209-216 [PMID: 12472785 DOI: 10.1046/j.1523-1755.2003.00736.x]
- 125 **Fernández-López A**, Jiménez-Mejías ME, García-Curiel A, Palomino Nicás J. [Meningitis caused by Clostridium cadaveris]. *Med Clin (Barc)* 1991; **97**: 679 [PMID: 1762467]
- 126 **Rodríguez-Iturbe B**, Quiroz Y, Shahkarami A, Li Z, Vaziri ND. Mycophenolate mofetil ameliorates nephropathy in the obese Zucker rat. *Kidney Int* 2005; **68**: 1041-1047 [PMID: 16105034 DOI: 10.1111/j.1523-1755.2005.00496.x]
- 127 **Durez P**, Appelboom T, Pira C, Stordeur P, Vray B, Goldman M. Antiinflammatory properties of mycophenolate mofetil in murine endotoxemia: inhibition of TNF-alpha and upregulation of IL-10 release. *Int J Immunopharmacol* 1999; **21**: 581-587 [PMID: 10501627 DOI: 10.1016/S0192-0561(99)00037-5]
- 128 **Huang Y**, Liu Z, Huang H, Liu H, Li L. Effects of mycophenolic acid on endothelial cells. *Int Immunopharmacol* 2005; **5**: 1029-1039 [PMID: 15829418 DOI: 10.1111/15829418]
- 129 **Yang YF**, Tan DM, Xie YT, Zhao W, Hou ZH, Zhong YD. Mycophenolate mofetil prevents lethal acute liver failure in mice induced by bacille Calmette-Guérin and lipopolysaccharide. *J Gastroenterol Hepatol* 2008; **23**: 611-618 [PMID: 17944887 DOI: 10.1111/j.1440-1746.2007.05169.x]
- 130 **Moriwaki Y**, Inokuchi T, Yamamoto A, Ka T, Tsutsumi Z, Takahashi S, Yamamoto T. Effect of TNF-alpha inhibition on urinary albumin excretion in experimental diabetic rats. *Acta Diabetol* 2007; **44**: 215-218 [PMID: 17767370 DOI: 10.1007/s00592-007-0007-6]
- 131 **Han J**, Thompson P, Beutler B. Dexamethasone and pentoxifylline inhibit endotoxin-induced cachectin/tumor necrosis factor synthesis at separate points in the signaling pathway. *J Exp Med* 1990; **172**: 391-394 [PMID: 2358784 DOI: 10.1084/jem.172.1.391]
- 132 **Doherty GM**, Jensen JC, Alexander HR, Buresh CM, Norton JA. Pentoxifylline suppression of tumor necrosis factor gene transcription. *Surgery* 1991; **110**: 192-198 [PMID: 1858029]
- 133 **Voisin L**, Breuillé D, Ruot B, Rallièrre C, Rambourdin F, Dalle M, Obled C. Cytokine modulation by PX differently affects specific acute phase proteins during sepsis in rats. *Am J Physiol* 1998; **275**: R1412-R1419 [PMID: 9791055]
- 134 **Cooper A**, Mikhail A, Lethbridge MW, Kemeny DM, Macdougall IC. Pentoxifylline improves hemoglobin levels in patients with erythropoietin-resistant anemia in renal failure. *J Am Soc Nephrol* 2004; **15**: 1877-1882 [PMID: 15213276 DOI: 10.1097/01.ASN.0000131523.17045.56]
- 135 **Bolick DT**, Hatley ME, Srinivasan S, Hedrick CC, Nadler JL. Lisofylline, a novel antiinflammatory compound, protects mesangial cells from hyperglycemia- and angiotensin II-mediated extracellular matrix deposition. *Endocrinology* 2003; **144**: 5227-5231 [PMID: 12960000 DOI: 10.1210/en.2003-0739]
- 136 **Dávila-Esqueda ME**, Vertiz-Hernández AA, Martínez-Morales F. Comparative analysis of the renoprotective effects of pentoxifylline and vitamin E on streptozotocin-induced diabetes mellitus. *Ren Fail* 2005; **27**: 115-122 [PMID: 15717644 DOI: 10.1081/JDI-200042728]
- 137 **Solerte SB**, Fioravanti M, Bozzetti A, Schifino N, Patti AL, Fedele P, Viola C, Ferrari E. Pentoxifylline, albumin excretion rate and proteinuria in type I and type II diabetic patients with microproteinuria. Results of a short-term randomized study. *Acta Diabetol Lat* 1986; **23**: 171-177 [PMID: 3751450 DOI: 10.1007/BF02624677]
- 138 **Tripathi K**, Prakash J, Appaiha D, Srivastava PK. Pent-

- oxifylline in management of proteinuria in diabetic nephropathy. *Nephron* 1993; **64**: 641-642 [PMID: 8366994 DOI: 10.1159/000187415]
- 139 **Guerrero-Romero F**, Rodríguez-Morán M, Paniagua-Sierra JR, García-Bulnes G, Salas-Ramírez M, Amato D. Pentoxifylline reduces proteinuria in insulin-dependent and non insulin-dependent diabetic patients. *Clin Nephrol* 1995; **43**: 116-121 [PMID: 7736673]
- 140 **Gorson DM**. Reduction of macroalbuminuria with pentoxifylline in diabetic nephropathy. Report of three cases. *Diabetes Care* 1998; **21**: 2190-2191 [PMID: 9839116 DOI: 10.2337/diacare.21.12.2190a]
- 141 **Navarro JF**, Mora C. Antiproteinuric effect of pentoxifylline in patients with diabetic nephropathy. *Diabetes Care* 1999; **22**: 1006-1008 [PMID: 10372263 DOI: 10.2337/diacare.22.6.1006]
- 142 **Navarro JF**, Mora C, Rivero A, Gallego E, Chahin J, Macía M, Méndez ML, García J. Urinary protein excretion and serum tumor necrosis factor in diabetic patients with advanced renal failure: effects of pentoxifylline administration. *Am J Kidney Dis* 1999; **33**: 458-463 [PMID: 10070909 DOI: 10.1016/S0272-6386(99)70182-4]
- 143 **Blagosklonnaia IaV**, Mamedov R, Kozlov VV, Emanuéel' VL, Kudriashova MI. [Effect of trental on indices kidney function in diabetes mellitus]. *Probl Endokrinol (Mosk)* 1982; **28**: 3-8 [PMID: 7100130]
- 144 **Rodríguez-Morán M**, González-González G, Bermúdez-Barba MV, Medina de la Garza CE, Tamez-Pérez HE, Martínez-Martínez FJ, Guerrero-Romero F. Effects of pentoxifylline on the urinary protein excretion profile of type 2 diabetic patients with microproteinuria: a double-blind, placebo-controlled randomized trial. *Clin Nephrol* 2006; **66**: 3-10 [PMID: 16878429 DOI: 10.5414/CNP66003]
- 145 **Navarro JF**, Mora C, Muros M, Maca M, Garca J. Effects of pentoxifylline administration on urinary N-acetyl-beta-glucosaminidase excretion in type 2 diabetic patients: a short-term, prospective, randomized study. *Am J Kidney Dis* 2003; **42**: 264-270 [PMID: 12900807 DOI: 10.1016/S0272-6386(03)00651-6]
- 146 **Leyva-Jiménez R**, Rodríguez-Orozco AR, Ortega-Pierres LE, Ramírez-Enríquez J, Gómez-García A, Alvarez-Aguilar C. [Effect of pentoxifylline on the evolution of diabetic nephropathy]. *Med Clin (Barc)* 2009; **132**: 772-778 [PMID: 19464709 DOI: 10.1016/j.medcli.2008.05.024]
- 147 **Navarro JF**, Mora C, Muros M, García J. Additive anti-proteinuric effect of pentoxifylline in patients with type 2 diabetes under angiotensin II receptor blockade: a short-term, randomized, controlled trial. *J Am Soc Nephrol* 2005; **16**: 2119-2126 [PMID: 15917336 DOI: 10.1681/ASN.2005010001]
- 148 **Roosbeh J**, Banihashemi MA, Ghezlou M, Afshariani R, Salari S, Moini M, Sagheb MM. Captopril and combination therapy of captopril and pentoxifylline in reducing proteinuria in diabetic nephropathy. *Ren Fail* 2010; **32**: 172-178 [PMID: 20199178 DOI: 10.3109/08860221003602645]
- 149 **McCormick BB**, Sydor A, Akbari A, Fergusson D, Doucette S, Knoll G. The effect of pentoxifylline on proteinuria in diabetic kidney disease: a meta-analysis. *Am J Kidney Dis* 2008; **52**: 454-463 [PMID: 18433957 DOI: 10.1053/j.ajkd.2008.01.025]
- 150 **Navarro-González JF**, Muros M, Mora-Fernández C, Herrera H, Meneses B, García J. Pentoxifylline for renoprotection in diabetic nephropathy: the PREDIAN study. Rationale and basal results. *J Diabetes Complications* 2011; **25**: 314-319 [PMID: 21144773 DOI: 10.1016/j.jdiacomp.2010.09.003]

P- Reviewer: Cui WP, Ozdemir S, Theilade S **S- Editor:** Qi Y

L- Editor: A **E- Editor:** Liu SQ





Published by **Baishideng Publishing Group Inc**

8226 Regency Drive, Pleasanton, CA 94588, USA

Telephone: +1-925-223-8242

Fax: +1-925-223-8243

E-mail: bpgoffice@wjgnet.com

Help Desk: <http://www.wjgnet.com/esps/helpdesk.aspx>

<http://www.wjgnet.com>

