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***Basic Study***

**Identification of the circRNA-miRNA-mRNA regulatory network and its prognostic effect in colorectal cancer**

Yin TF *et al*. CircRNA-miRNA-mRNA network in CRC

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**Abstract**

BACKGROUND

The high morbidity and mortality of colorectal cancer (CRC) have posed great threats to human health. CircRNA and miRNA, acting as competing endogenous RNAs (ceRNAs), have been found to play vital roles in carcinogenesis. However, the biological function of ceRNAs in CRC pathogenesis and prognosis remains largely unexplored.

AIM

To identify the CRC-specific circRNA-miRNA-mRNA regulatory network and uncover the subnetwork associated with its prognosis.

METHODS

CircRNAs, miRNAs and mRNAs differentially expressed (DE) in CRC tissues were selected by expression file analysis in the Group on Earth Observations (GEO) database, and the downstream target molecules of circRNAs and miRNAs were predicted. Then, the intersection of differentially expressed RNA molecules with the predicted targets was determined to obtain a ceRNA network. GO and KEGG pathway analyses were conducted to elucidate the possible mechanism of pathogenesis. A survival analysis using the gene profiles and clinical information in The Cancer Genome Atlas (TCGA) database was performed to identify the mRNAs associated with the clinical outcome of CRC patients and construct a prognostic subnetwork.

RESULTS

We downloaded three datasets (GSE126095, GSE41655 and GSE41657) of large-scale CRC samples from the GEO database. There were 55 DEcircRNAs, 114 DEmiRNAs and 267 DEmRNAs in CRC tissues compared with normal tissues. After intersecting these molecules with predicted targets, 19 circRNAs, 13 miRNAs and 28 mRNAs were chosen to develop a circRNA-miRNA-mRNA network. GO and KEGG functional enrichment analyses indicated that the retinol metabolic process, leukocyte chemotaxis, extracellular matrix remodeling, endoplasmic reticulum stress, alcohol dehydrogenase activity, gastric acid secretion, nitrogen metabolism and NOD-like receptor signaling pathway might participate in the tumorigenesis of CRC. After verifying the identified mRNA effect in the TCGA database, we finally recognized 3 mRNAs (*CA2, ITLN1* and *LRRC19*) that were significantly associated with the overall survival of CRC patients and constructed a ceRNA subnetwork including 5 circRNAs (hsa\_circ\_0080210, hsa\_circ\_0007158, hsa\_circ\_0000375, hsa\_circ\_0018909 and hsa\_circ\_0011536) and 3 miRNAs (hsa-miR-601, hsa-miR-671-5p and hsa-miR-765), which could contain innovative and noninvasive indicators for the early screening and prognostic prediction of CRC.

CONCLUSION

We proposed a circRNA-miRNA-mRNA regulatory network closely associated with the progression and clinical outcome of CRC that might include promising biomarkers for carcinogenesis and therapeutic targets.

**Key Words:** CircRNA; miRNA; Network; Colorectal cancer; Prognosis; Biomarkers

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**Core Tip:** The biological functions of circRNA and miRNA interactions and their potential as noninvasive biomarkers have not been well elucidated in colorectal cancer (CRC). In this study, we constructed a circRNA-miRNA-mRNA regulatory network with 19 circRNAs, 13 miRNAs and 28 mRNAs. GO and KEGG analyses indicated several signaling pathways probably involved in tumorigenesis. After being combined with survival analysis, a prognostic subnetwork was constructed including 5 circRNAs, 3 miRNAs and 3 mRNAs, which may represent novel diagnostic and prognostic candidate biomarkers, as well as therapeutic targets of CRC.

**INTRODUCTION**

Colorectal cancer (CRC) is one of the most common tumors worldwide. The global cancer burden report in 2018 showed that the incidence and mortality of CRC ranked third and second, respectively[1]. Conventional therapeutic options for CRC, such as chemotherapy and radiotherapy, cannot satisfy the ever-rising demand for overall and disease-free survival. At present, the high morbidity and mortality of CRC pose a great threat to the health of humans. Early screening and prevention should be actively and urgently carried out. The pathogenesis of CRC remains largely unexplored, and gene regulation disorders may play an important role in it.

In recent years, circRNAs and miRNAs, acting as noncoding RNAs, have attracted considerable research attention in a variety of diseases. These RNA molecules could regulate gene expression through complex mechanisms and interactions. CircRNA is a newly identified class of single-stranded circular, noncoding RNA molecules without 5′ poly-A and 3′ cap ends, which makes it resistant to degradation by RNA exonucleases and more stable than the linear RNA class[2]. Emerging evidence has proven that circRNAs are widely expressed in eukaryotic cells and can be implicated in physiological and pathological processes[3]. Salmena *et al*[4] first hypothesized that competing endogenous RNAs (ceRNAs) contain adequate miRNA response elements and may act as miRNA sponges to bind and compete with corresponding miRNAs, thereby sequestering miRNAs and regulating mRNA expression at the posttranslational level. MiRNAs are small, noncoding RNAs of approximately 20-22 nucleotides that can play a vital role in the regulation of gene expression, such as decreasing mRNA stability in various biological and pathological processes[5].

Accumulating evidence has revealed that the circRNA-miRNA-mRNA network could play a significantly important role in the development and progression of many diseases, especially cancer. For example, Song and Fu[6] discovered that the hsa\_circ\_00001666/has-mir-1229/*CXCR5* axis could participate in the pathogenesis of CRC and act as a promising biomarker for targeted treatment. A related study indicated that hsa\_circ\_0005100 has pivotal value in the progression of CRC *via* the miR-1182/*hTERT* axis[7]. Hsa\_circ\_000984 could sequester miR-106b and consequently intensify the proliferation and migration of CRC cell lines[8]. These studies indicate that dysregulated circRNAs, miRNAs and mRNAs are closely related to the progression and prognosis of CRC and could be used as potential CRC-specific predictors, but the competitive regulatory pattern and biological function mechanism among circRNAs, miRNAs and mRNAs are still complicated and need further verification.

Along with the enormous advancement of RNA-sequencing technology, many public databases, such as Group on Earth Observations (GEO) and The Cancer Genome Atlas (TCGA), have been established, and a surge of large-scale RNA sequence data are available. Recently, bioinformatics analysis has been widely used to help screen key genes, construct regulatory models, and select therapeutic targets that may participate in tumor development and prognosis as well as provide guidance for basic and clinical research, which is helpful to clarify the pathogenesis of tumors and guide clinical treatment options.

The objective of this study was to explore a competitive regulatory model among circRNAs, miRNAs and mRNAs and discover promising indicators for driving factors and mechanisms that induce the progression and affect the prognosis of CRC. To achieve this goal, we downloaded three datasets (GSE126095, GSE41655 and GSE41657) of large-scale CRC samples in the GEO database, selected differentially expressed (DE) circRNAs, DEmiRNAs and DEmRNAs in CRC tissues compared with normal controls, and predicted the downstream target molecules of circRNAs and miRNAs. After intersection with the differentially expressed RNA molecules and predicted targets, a circRNA-miRNA-mRNA network was identified. Functional enrichment analyses were conducted to identify the underlying mechanism involved in the pathogenesis of CRC. To verify the prognostic effect of the mRNAs found above, we performed survival analysis using the gene profiles and clinical information in the TCGA database. Finally, survival-related genes were determined, and a prognostic subnetwork was developed. Our results revealed the circRNA-miRNA-mRNA regulatory interaction and provided guidance for expanding the understanding of CRC progression, prognosis and therapeutic options. The flow chart of the procedure in our study is illustrated in Figure 1.

**MATERIALS AND METHODS**

***Dataset retrieval***

The expression profiles of three datasets (GSE126095, GSE41655 and GSE41657) were downloaded from the GEO database (http://www.ncbi.nlm.nih.gov/geo). Referring to the annotation information on the platform, probes were transformed to corresponding gene symbols. The GSE126095 dataset included 10 CRC and 10 normal tissues. The GSE41655 dataset contained 33 CRC tissues, 15 normal tissues and 59 colorectal adenomas, and GSE41657 included 25 CRC tissues, 12 normal tissues and 51 colorectal adenomas. CRC and normal tissue data in the two datasets above were chosen for comprehensive analysis. Moreover, clinical information and mRNA expression profiles were obtained from the TCGA database (https://portal.gdc.cancer.gov/). A total of 530 samples of patients (488 CRC tissues and 42 normal tissues), which had complete clinical characteristics, including age, sex, stage, survival time and survival state, and survival or follow-up time ≥ 30 d, were selected for further analysis. This study was approved by the Ethics Committee of China-Japan Friendship Hospital (No. 2018-116-K85-1). An informed consent statement was not necessary because all data were acquired from the GEO and TCGA databases and available to the public.

***Identification of differentially expressed circRNAs, miRNAs and mRNAs***

After downloading data from the GEO database, the “limma” package was utilized for background correction and normalization of the raw read counts of circRNA, miRNA and mRNA as well as identification of DEcircRNAs, DEmiRNAs and DEmRNAs between tumor samples and normal controls. The cut-off criteria of circRNA and mRNA were set at an adjusted *P* < 0.05 and |log2 fold change (FC)| > 2, while the cut-off value of miRNA was set at an adjusted *P* < 0.05 and |log2 FC|>1.

***Target prediction and intersection for ceRNA network construction***

In the circBase database (http://www.circb ase.org/), specific information on the DEcircRNAs was available, and then the Cancer-Specific CircRNA database (CSCD) (http://gb.whu.edu.cn/CSCD/) was used to obtain structural patterns of circRNAs and predict the binding relationship between circRNAs and miRNAs. After intersection with the DEmiRNAs, promising miRNAs were finally identified, and their target mRNAs were predicted in the TargetScan and miRDB databases. When both databases supported mRNAs as candidate targets, these targets and DEmRNAs were intersected to obtain the final mRNA. After removing unconnected nodes, the circRNA-miRNA-mRNA network was developed based on the results above and visualized using Cytoscape 3.7.2.

***GO and KEGG functional enrichment analysis***

To reveal the pathophysiological processes and critical signaling pathways involved in the carcinogenesis of CRC, two widely used bioinformatics analysis methods, GO and KEGG, were conducted. The “clusterProfiler” package in R/Bioconductor was used for analyzing GO term and KEGG pathway enrichment. An adjusted *P* < 0.05 was regarded as a statistical criterion.

***Survival analysis and prognostic subnetwork construction***

To assess the value of the identified ceRNA network and determine the mRNAs related to prognosis, the mRNA expression profile and clinical information for CRC patients were downloaded from the TCGA database. Kaplan–Meier curves were generated for survival analysis. The log-rank test was utilized for statistical analysis. The cut-off criterion was a *P* < 0.05. Finally, a ceRNA subnetwork was developed on the basis of the above verified mRNAs.

**RESULTS**

***Identification of differentially expressed and intersecting circRNAs, miRNAs and mRNAs***

The basic characteristics of the three GEO datasets (GSE126095, GSE41655 and GSE41657) and the TCGA is shown in Table 1. In the circRNA profile data of GSE126095, there were a total of 55 DEcircRNAs between the CRC samples and normal controls, among which 22 upregulated and 33 downregulated in CRC tissues (Figure 2). After removing 6 circRNAs that did not have details in the CSCD database, the structural models of 49 DEcircRNAs obtained from CSCD database are depicted in Figure 3, and the basic information of the these circRNAs is listed in Supplementary Table 1. Based on this, we successfully predicted that 1602 miRNAs might be the targets of these 49 circRNAs. For miRNAs with aberrant expression in GSE41655, 114 miRNAs were investigated to be differentially expressed in CRC, among which 58 were overexpressed and 56 were downregulated (Supplementary Table 2). A total of 25 miRNAs were identified from the intersection of 114 DEmiRNAs and 1602 circRNA targets predicted by the CSCD database (Supplementary Table 3). According to the TargetScan and miRDB databases, 7190 potential target genes for the 25 intersecting miRNAs were found. In the GSE41657 mRNA expression profile, 267 mRNAs were differentially expressed, among which 112 were highly expressed and 155 were expressed at low levels in CRC compared with normal tissues (Supplementary Table 4). A total of 77 intersecting mRNAs were generated (Supplementary Table 5). We utilized Venn diagrams to illustrate the intersecting states of miRNA and mRNA (Figure 4A and B) and took the 20 miRNAs and 20 mRNAs expressing the most significant upregulation and downregulation, respectively, to draw heat maps (Figure 4C and D).

***Construction of the CRC-specific ceRNA network***

Based on the recognized 49 circRNAs, 25 miRNAs and 77 mRNAs, we removed unconnected nodes and chose 19 circRNAs (hsa\_circ\_0000520, hsa\_circ\_0000519, hsa\_circ\_0001955, hsa\_circ\_0028198, hsa\_circ\_0080210, hsa\_circ\_0007158, hsa\_circ\_0000375, hsa\_circ\_0000026, hsa\_circ\_0023685, hsa\_circ\_0000370, hsa\_circ\_0061817, hsa\_circ\_0005927, hsa\_circ\_0072088, hsa\_circ\_0018909, hsa\_circ\_0013912, hsa\_circ\_0071681, hsa\_circ\_0011536, hsa\_circ\_0043278, and hsa\_circ\_0006220), 13 miRNAs (hsa-miR-423-5p, hsa-miR-532-3p, hsa-miR-765, hsa-miR-1224-5p, hsa-miR-650, hsa-miR-769-5p, hsa-miR-671-5p, hsa-miR-1290, hsa-miR-125a-3p, hsa-miR-601, hsa-miR-198, hsa-miR-1202, and hsa-miR-1182) and 28 mRNAs (*RNF43, DSG3, AZGP1, SST, DES, TCF21, MFAP4, EREG, BCAS1, C1QA, SPARCL1, CXCL3, EPHB3, TRAF3IP3, TRPM2, CA2, LRRC19, SCG2, C16orf89, ADH1A, MZB1, HAPLN1, S100A2, GPR34, MS4A12, ITLN1, DHRS9,* and *CHGB*) in to construct a circRNA-miRNA-mRNA regulatory network utilizing Cytoscape 3.7.2 (Figure 5). The expression levels of these RNA molecules are shown in Figure 6.

***GO and KEGG functional enrichment analysis***

GO analysis revealed that the enrichments of the identified mRNAs in the ceRNA network were mainly in the ‘retinoic acid metabolic process’, ‘retinol metabolic process’, ‘digestive tract morphogenesis’, and ‘leukocyte chemotaxis’ (biological processes; *P* < 0.005) (Figure 7A); ‘collagen-containing extracellular matrix’, ‘endoplasmic reticulum lumen’ and ‘endoplasmic reticulum chaperone complex’ (cellular components; *P* < 0.02) (Figure 7B); and ‘alcohol dehydrogenase (ADH) [NAD(P)+] activity’, ‘retinol dehydrogenase activity’ and ‘oxidoreductase activity’ (molecular functions; *P* < 0.02) (Figure 7C). The results of KEGG pathway analysis involving the ceRNA network indicated that the target genes were mainly enriched in ‘retinol metabolism’, ‘gastric acid secretion’, ‘nitrogen metabolism’, ‘NOD-like receptor signaling pathway’, ‘proximal tubule bicarbonate reclamation’, ‘collecting duct acid secretion’ and ‘tyrosine metabolism’ (*P* < 0.05) (Figure 7D). According to these results, the retinol metabolic process, leukocyte chemotaxis, extracellular matrix remodeling, endoplasmic reticulum stress, ADH activity, gastric acid secretion, nitrogen metabolism and NOD-like receptor signaling pathway might participate in the tumorigenesis of CRC.

***Survival analysis and construction of the prognostic ceRNA subnetwork***

We obtained mRNA profiles and clinical information of CRC patients in the TCGA database and performed survival analysis for each mRNA in the obtained ceRNA network. Finally, we recognized that 3 mRNAs (*CA2, ITLN1,* and *LRRC19*) were significantly correlated with the clinical outcome of CRC patients (Figure 8). The patients with the upregulation of *CA2* (*P* = 0.002), *ITLN1* (*P* = 0.001) and *LRRC19* (*P* = 0.032) had a better prognosis than the corresponding group with low expression. Considering the 3 mRNAs identified above, we successfully constructed and visualized a ceRNA subnetwork including 5 circRNAs (hsa\_circ\_0080210, hsa\_circ\_0007158, hsa\_circ\_0000375, hsa\_circ\_0018909 and hsa\_circ\_0011536) and 3 miRNAs (hsa-miR-601, hsa-miR-671-5p and hsa-miR-765) (Figure 9).

**DISCUSSION**

We identified the intersection of CRC-specific DEcircRNAs, DEmiRNAs and DEmRNAs in the GEO database with the target molecules of circRNAs and miRNAs predicted by relevant databases. Then, 19 circRNAs, 13 miRNAs and 28 mRNAs were identified to develop a circRNA-miRNA-mRNA regulatory network that may play a pivotal role in the progression of CRC. Subsequently, we conducted GO and KEGG pathway analyses for the 28 mRNAs to expand our understanding of the vital pathophysiological process of CRC initiation and progression. Finally, we verified the differential expression of the identified mRNAs in the TCGA database, screened prognosis-related mRNAs by conducting a survival analysis, and constructed a prognostic subnetwork using 5 circRNAs, 3 miRNAs and 3 mRNAs.

CircRNAs, as stable, abundant and conserved ceRNAs that act as miRNA sponges, could be identified as valuable indicators for the diagnosis and pathogenesis of CRC. Similar studies also supported the evidence that DEcircRNAs identified in our study could be vital components in the ceRNA network, which modulate crucial gene expression in the initiation and progression of cancer, especially CRC. For example, the dysregulation of hsa\_circ\_0000520 could affect the tumorigenesis of cervical cancer and breast cancer through the miRNA-mRNA axis[9,10]. The hsa\_circ\_0072088 in the circRNA-miRNA-mRNA network was identified as being related to CRC and lung cancer progression[11,12]. The hsa\_circ\_0001955 was found to mediate a ceRNA network in CRC by bioinformatics analysis and experimental validation[13]. The hsa\_circ\_0005927 was verified in gastric cancer and could be a biomarker for gastric cancer screening[14]. The hsa\_circ\_0000026 was found to be expressed at low levels in gastric cancer and may correlate with the progression of CRC[15]. The hsa\_circ\_0000370 in plasma showed diagnostic value for CRC and might be involved in tumorigenesis[16]. All of these studies indicate that circRNAs may participate in CRC progression and could be vital biomarkers for diagnosis as well as therapeutic targets. The majority of 55 DEcircRNAs identified in our study were innovative biomarkers in CRC and still require further investigation in the future.

To further explore signaling pathways that might play an important role in the tumorigenesis and progression of CRC, we conducted GO and KEGG analyses of 28 identified genes in the ceRNA network. Growing studies have confirmed that the signaling pathways uncovered in our study participate in crucial pathological processes in many kinds of cancer. Bhattacharya *et al*[17] reported that the inhibition of retinoic acid signaling, a key regulator of intestinal immunity, could promote the tumorigenesis of CRC by cytotoxic T cells, and retinoic acid catabolizing enzyme was a promising negative predictor for the prognosis of CRC patients. Retinol dehydrogenase 16, one of the isoforms of the rate-limiting enzyme of the retinol cycle, was reported to increase the level of retinoic acid, and associate with the tumor size of hepatocellular carcinoma and poor overall survival of patients as well[18]. Mesenchymal cells in the intestine, called cancer-associated fibroblasts, exert critical functions to regulate a variety of activities, including intestinal inflammation, epithelial proliferation, extracellular matrix remodeling and metastasis, which could affect the microenvironment and promote CRC development and progression[19]. A recent study revealed that leukocyte chemotaxis and adhesion were distinctly reduced in the vasculature of CRC[20], and some miRNAs, including miR-15A and miR-16-1, were able to modulate the pathway of immune regulatory B cell chemotaxis in CRC, which could affect the tumor growth and survival time[21]. Endoplasmic reticulum stress is one of the pivotal processes in carcinogenesis and could represent an innovative therapeutic target in resistant tumors[22]. Cheng *et al*[23] found that endoplasmic reticulum stress participated in the apoptosis and autophagy in CRC induced by apatinib, a novel tyrosine kinase inhibitor. The consumption of alcohol was reported to increase the risk of colorectal adenomas, and ADH might modify the correlation between alcohol consumption and colorectal adenomas[24]. ADH expression has the potential to be a prognostic marker of pancreatic adenocarcinoma[25], and acetaldehyde is recognized to elevate the possibility of chemically induced rectal carcinogenesis[26]. A recent meta-analysis revealed that gastric acid suppressant use showed a significant correlation with poor survival for patients receiving oral chemotherapy for gastrointestinal tract cancer, supporting a possible negative impact of gastric acid suppressants on the survival outcome of CRC[27]. As one of the most fundamental requirements for biosynthesis, nitrogen metabolism is utilized and modulated to sustain the increased demand for nitrogen sources in cancer proliferation[28]. The regulation of nitrogen metabolism participates in the process of obesity-associated pancreatic cancer, small cell lung cancer and CRC metastasis[29-31]. The NOD-like receptor signaling pathway was one of the enriched pathways of genes with aberrant expression among pancreatic, thyroid, and renal cancer compared with healthy controls through bioinformatics analyses[32-34]. Furthermore, KEGG website (http://www.kegg.jp/) was utilized to analyze the specific steps of these biological processes affected by identified mRNAs in ceRNA network. It is well-known that CA2 participates in the processes of combining water and carbon dioxide to generate carbonic acid, acting as one of the key enzymes in the proximal tubule bicarbonate reclamation and collecting duct acid secretion. According to the pathway diagrams in KEGG website, CA2 also involves in gastric acid secretion and arginine biosynthesis in nitrogen metabolism. CXCXL3 is one of the downstream chemokines of NOD-nuclear factor-kappaB pathway in NOD-like receptor signaling pathway. ADH, as one of the key enzymes for the mutual transformation of all-trans-retinal and all-trans-retinol (vitamin A), participates in the final metabolic process of dopamine to 3-methoxy-4-hydroxy-phenylethylene-glycol in tyrosine metabolism. In summary, the retinol metabolic process, leukocyte chemotaxis, extracellular matrix remodeling, endoplasmic reticulum stress, ADH activity, gastric acid secretion, nitrogen metabolism and NOD-like receptor signaling pathway might represent essential signaling pathways involved in the pathogenesis of CRC. However, the molecular mechanism of tumorigenesis and progression is quite complicated and still requires further exploration.

We verified the differential expression of the mRNAs in the ceRNA network using the TCGA database, combined the results of the survival analysis, screened prognosis-related mRNAs, and finally used 5 circRNAs, 3 miRNAs and 3 mRNAs to construct a prognostic subnetwork. The roles of the 5 circRNAs (hsa\_circ\_0080210, hsa\_circ\_0007158, hsa\_circ\_0000375, hsa\_circ\_0018909 and hsa\_circ\_0011536) involved in tumorigenesis require investigation, which implies that these circRNAs might have the potential to become novel indicators for CRC diagnosis and targeted treatment. Some studies were consistent with our result that 3 miRNAs (hsa-miR-601, hsa-miR-671-5p and hsa-miR-765) might represent promising biomarkers of cancer progression and prognosis. MiR-601 was identified to hold diagnostic value for CRC with 69.2% sensitivity and 72.4% specificity for CRC diagnosis[35], and might suppress the proliferation and invasion of esophageal squamous cell carcinoma and breast cancer[36,37]. Some studies revealed that miR-671-5p could promote and maintain the oncogenesis and progression of various cancers, such as esophageal cancer, breast cancer, glioblastoma and melanoma[38-41], and act as a prognostic predictor of locally advanced rectal cancer due to the significant upregulation in pathological response to neoadjuvant chemoradiotherapy[42]. The expression level of hsa-miR-765 could be utilized to independently predict overall survival and disease-free survival and correlate with the tumor stage, clinical stage and lymph node metastasis in esophageal squamous cell carcinoma[43]. Hsa-miR-765 could also promote the aggressiveness of hepatocellular carcinoma and osteosarcoma[44,45]. To date, although some studies have focused attention on the roles of these 3 miRNAs in many types of cancer, hsa-miR-671-5p and hsa-miR-765 have not been recognized as promising biomarkers of CRC before our study.

We found that the upregulation of *CA2, ITLN1* and *LRRC19* might be related to better clinical outcomes in CRC patients. *CA2*, the gene that encodes carbonic anhydrase Ⅱ, was validated to be downregulated in CRC tissue and cell lines compared with healthy controls through experimental assays, and the overexpression of *CA2* suppressed tumor cell growth *in vitro* and *in vivo* and elevated the sensitivity of CRC cells to chemotherapy drugs[46]. *CA2* might also be useful for the survival prediction of CRC patients[47]. Other carbonic anhydrase isoforms, such as *CA1*, *CA4*, *CA9* and *CA12*, could exert effects on favorable outcomes or rescue the tumor progression of CRC[48-50]. In summary, *CA2* could be a beneficial predictor for CRC diagnosis and prognosis. The expression of *ITLN1*, which encodes intelectin-1 (also known as omentin-1), presented a sequentially descending trend with the mucosa-adenoma-carcinoma process[51]. The downregulation of *ITLN1* was reported to be related to poor prognosis among CRC patients at advanced stages[52]. *ITLN1* could alsosuppress tumorigenesis and correlate with a better prognosis probability of neuroblastoma and ovarian cancer[53,54]. The low expression of *LRRC19* (leucine-rich repeat containing 19) was identified as an independent risk factor and prognostic biomarker of kidney renal clear cell carcinoma and was involved in selenium adjuvant therapy[55]. In addition, *LRRC19* could have predictive power for sensitivity to AZD0530 in pancreatic tumor and promote the elimination of pathogenic bacteria from the kidneys[56,57]. However, whether *LRRC19* could be regarded as an indicator of oncogenesis and the clinical outcome of CRC has not been illustrated before. In summary, *CA2, ITLN1* and *LRRC19* might have the potential to become novel biomarkers for CRC diagnosis, especially early screening, as well as patient prognosis. At present, the hsa\_circ\_0011536/hsa-miR-601/*CA2* axis and the complex interaction among circRNAs (hsa\_circ\_0080210, hsa\_circ\_0007158, hsa\_circ\_0000375 and hsa\_circ\_0018909), miRNAs (hsa-miR-671-5p and hsa-miR-765) and mRNAs (*ITLN1* and *LRRC19*) could provide novel insights into the pathogenesis of and therapeutic options for CRC and need further investigation.

Collectively, we have provided a deeper understanding of the circRNA-related ceRNA mechanism of CRC by developing a circRNA-miRNA-mRNA regulatory network. Through GO and KEGG functional enrichment analysis, the retinol metabolic process, leukocyte chemotaxis, extracellular matrix remodeling, endoplasmic reticulum stress, ADH activity, gastric acid secretion, nitrogen metabolism and the NOD-like receptor signaling pathway might play critical roles in the initiation and progression of CRC. After being combined with survival analysis, a prognostic subnetwork was constructed, including 5 circRNAs, 3 miRNAs and 3 mRNAs, which could be novel candidate biomarkers for the clinical outcome of CRC. Our research still has some shortcomings. First, molecular-level verification in clinical samples and CRC cell lines should be applied to validate biomarkers and clarify the actual significance of the regulatory ceRNA network and prognostic subnetwork. In addition, the lack of research on the downstream target molecules of the ceRNA network makes it difficult to completely elucidate the specific mechanism in the occurrence and development of CRC. Moreover, due to the sample size of the datasets, the sample size of this study is not too large.

**CONCLUSION**

In summary, we constructed a CRC-specific circRNA-miRNA-mRNA regulatory network and performed functional enrichment analysis, which may assist in revealing the mechanism of carcinogenesis. A prognostic ceRNA subnetwork was successfully developed, thereby identifying several RNA molecules that could serve as innovative and noninvasive indicators for the RNA-based early screening and prognostic prediction of CRC. Comprehensive experimental studies are still required to enrich the understanding of ceRNAs, which is essential to illustrate the pathogenesis and prognosis of CRC and provide new opportunities for targeted therapeutics.

**ARTICLE HIGHLIGHTS**

***Research background***

Colorectal cancer (CRC) is one of the most common malignant tumors worldwide. CircRNA and miRNA, acting as competing endogenous RNAs (ceRNAs), have been investigated to play vital roles in carcinogenesis. Accumulating evidence highlights that it is necessary to further explore the biological function of circRNA and miRNA in CRC pathogenesis and prognosis.

***Research motivation***

Dysregulated circRNAs, miRNAs and mRNAs could closely associate with the progression and prognosis of CRC and act as potential CRC-specific predictors, but the competitive regulatory pattern and biological function mechanism among circRNAs, miRNAs and mRNAs are still complicated and have not yet been elucidated.

***Research objectives***

This study aimed to uncover a CRC-specific competitive regulatory model among circRNAs, miRNAs and mRNAs and explore the subnetwork associated with CRC prognosis.

***Research methods***

Expression profiles of circRNAs, miRNAs and mRNAs were downloaded from the Group on Earth Observations (GEO) database. Differentially expressed (DE) circRNAs, miRNAs and mRNAs in CRC tissues and the predicted target molecules of circRNAs and miRNAs were intersected to obtain a CRC-specific ceRNA network. GO and KEGG pathway analyses were conducted to explore the mechanism of CRC pathogenesis. Based on the survival analysis using the gene profiles and clinical information in The Cancer Genome Atlas (TCGA) database, the mRNAs significantly associated with the clinical outcome of CRC patients were identified and a prognostic subnetwork was constructed.

***Research results***

There were 55 DEcircRNAs, 114 DEmiRNAs and 267 DEmRNAs of CRC in three datasets (GSE126095, GSE41655 and GSE41657) from GEO database. After intersected with predicted targets, 19 circRNAs, 13 miRNAs and 28 mRNAs were chosen to develop ceRNA network. Go and KEGG analyses indicated that several signaling pathways might participate in the tumorigenesis. After verifying effect in TCGA database by survival analysis, we finally recognized 3 mRNAs (*CA2, ITLN1* and *LRRC19*) associated with prognosis, and constructed a ceRNA subnetwork including 5 circRNAs (hsa\_circ\_0080210, hsa\_circ\_0007158, hsa\_circ\_0000375, hsa\_circ\_0018909 and hsa\_circ\_0011536) and 3 miRNAs (hsa-miR-601, hsa-miR-671-5p and hsa-miR-765).

***Research conclusions***

A circRNA-miRNA-mRNA regulatory network closely associated with the progression and clinical outcome of CRC was identified, which might include promising biomarkers for carcinogenesis and therapeutic targets.

***Research perspectives***

We have provided a deeper understanding of the circRNA-related ceRNA mechanism of CRC by developing a circRNA-miRNA-mRNA regulatory network. Comprehensive experimental studies are still required to confirm our findings and provide new opportunities for targeted therapeutics.

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**REFERENCES**

1 **Bray F**, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, Jemal A. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J Clin* 2018; **68**: 394-424 [PMID: 30207593 DOI: 10.3322/caac.21492]

2 **Su M**, Xiao Y, Ma J, Tang Y, Tian B, Zhang Y, Li X, Wu Z, Yang D, Zhou Y, Wang H, Liao Q, Wang W. Circular RNAs in Cancer: emerging functions in hallmarks, stemness, resistance and roles as potential biomarkers. *Mol Cancer* 2019; **18**: 90 [PMID: 30999909 DOI: 10.1186/s12943-019-1002-6]

3 **Memczak S**, Jens M, Elefsinioti A, Torti F, Krueger J, Rybak A, Maier L, Mackowiak SD, Gregersen LH, Munschauer M, Loewer A, Ziebold U, Landthaler M, Kocks C, le Noble F, Rajewsky N. Circular RNAs are a large class of animal RNAs with regulatory potency. *Nature* 2013; **495**: 333-338 [PMID: 23446348 DOI: 10.1038/nature11928]

4 **Salmena L**, Poliseno L, Tay Y, Kats L, Pandolfi PP. A ceRNA hypothesis: the Rosetta Stone of a hidden RNA language? *Cell* 2011; **146**: 353-358 [PMID: 21802130 DOI: 10.1016/j.cell.2011.07.014]

5 **Lu TX**, Rothenberg ME. MicroRNA. *J Allergy Clin Immunol* 2018; **141**: 1202-1207 [PMID: 29074454 DOI: 10.1016/j.jaci.2017.08.034]

6 **Song W**, Fu T. Circular RNA-Associated Competing Endogenous RNA Network and Prognostic Nomogram for Patients With Colorectal Cancer. *Front Oncol* 2019; **9**: 1181 [PMID: 31781492 DOI: 10.3389/fonc.2019.01181]

7 **Li Y**, Li C, Xu R, Wang Y, Li D, Zhang B. A novel circFMN2 promotes tumor proliferation in CRC by regulating the miR-1182/hTERT signaling pathways. *Clin Sci (Lond)* 2019; **133**: 2463-2479 [PMID: 31738400 DOI: 10.1042/CS20190715]

8 **Xu XW**, Zheng BA, Hu ZM, Qian ZY, Huang CJ, Liu XQ, Wu WD. Circular RNA hsa\_circ\_000984 promotes colon cancer growth and metastasis by sponging miR-106b. *Oncotarget* 2017; **8**: 91674-91683 [PMID: 29207676 DOI: 10.18632/oncotarget.21748]

9 **Zhang J**, Cai R, Zhang Y, Wang X. Involvement of a novel circularRNA, hsa\_circ\_0000520, attenuates tumorigenesis of cervical cancer cell through competitively binding with miR-146b-3p. *J Cell Mol Med* 2020; **24**: 8480-8490 [PMID: 32592222 DOI: 10.1111/jcmm.15414]

10 **Zang H**, Li Y, Zhang X, Huang G. Blocking circ\_0000520 Suppressed Breast Cancer Cell Growth, Migration and Invasion Partially *via* miR-1296/SP1 Axis Both *in vitro* and in vivo. *Cancer Manag Res* 2020; **12**: 7783-7795 [PMID: 32922078 DOI: 10.2147/CMAR.S251666]

11 **Liang L**, Zhang L, Zhang J, Bai S, Fu H. Identification of circRNA-miRNA-mRNA Networks for Exploring the Fundamental Mechanism in Lung Adenocarcinoma. *Onco Targets Ther* 2020; **13**: 2945-2955 [PMID: 32308427 DOI: 10.2147/OTT.S235664]

12 **Bian L**, Zhi X, Ma L, Zhang J, Chen P, Sun S, Li J, Sun Y, Qin J. Hsa\_circRNA\_103809 regulated the cell proliferation and migration in colorectal cancer *via* miR-532-3p / FOXO4 axis. *Biochem Biophys Res Commun* 2018; **505**: 346-352 [PMID: 30249393 DOI: 10.1016/j.bbrc.2018.09.073]

13 **Ding B**, Yao M, Fan W, Lou W. Whole-transcriptome analysis reveals a potential hsa\_circ\_0001955/hsa\_circ\_0000977-mediated miRNA-mRNA regulatory sub-network in colorectal cancer. *Aging (Albany NY)* 2020; **12**: 5259-5279 [PMID: 32221048 DOI: 10.18632/aging.102945]

14 **Ding HX**, Xu Q, Wang BG, Lv Z, Yuan Y. MetaDE-Based Analysis of circRNA Expression Profiles Involved in Gastric Cancer. *Dig Dis Sci* 2020; **65**: 2884-2895 [PMID: 31894486 DOI: 10.1007/s10620-019-06014-6]

15 **Huang YS**, Jie N, Zou KJ, Weng Y. Expression profile of circular RNAs in human gastric cancer tissues. *Mol Med Rep* 2017; **16**: 2469-2476 [PMID: 28737829 DOI: 10.3892/mmr.2017.6916]

16 **Ye DX**, Wang SS, Huang Y, Chi P. A 3-circular RNA signature as a noninvasive biomarker for diagnosis of colorectal cancer. *Cancer Cell Int* 2019; **19**: 276 [PMID: 31700498 DOI: 10.1186/s12935-019-0995-7]

17 **Bhattacharya N**, Yuan R, Prestwood TR, Penny HL, DiMaio MA, Reticker-Flynn NE, Krois CR, Kenkel JA, Pham TD, Carmi Y, Tolentino L, Choi O, Hulett R, Wang J, Winer DA, Napoli JL, Engleman EG. Normalizing Microbiota-Induced Retinoic Acid Deficiency Stimulates Protective CD8(+) T Cell-Mediated Immunity in Colorectal Cancer. *Immunity* 2016; **45**: 641-655 [PMID: 27590114 DOI: 10.1016/j.immuni.2016.08.008]

18 **Zhu YH**, Li JB, Wu RY, Yu Y, Li X, Li ZL, Zhang HL, Feng GK, Deng R, Zhu XF. Clinical significance and function of RDH16 as a tumor-suppressing gene in hepatocellular carcinoma. *Hepatol Res* 2020; **50**: 110-120 [PMID: 31661588 DOI: 10.1111/hepr.13432]

19 **Koliaraki V**, Pallangyo CK, Greten FR, Kollias G. Mesenchymal Cells in Colon Cancer. *Gastroenterology* 2017; **152**: 964-979 [PMID: 28111227 DOI: 10.1053/j.gastro.2016.11.049]

20 **Bessa X**, Elizalde JI, Mitjans F, Piñol V, Miquel R, Panés J, Piulats J, Piqué JM, Castells A. Leukocyte recruitment in colon cancer: role of cell adhesion molecules, nitric oxide, and transforming growth factor beta1. *Gastroenterology* 2002; **122**: 1122-1132 [PMID: 11910362 DOI: 10.1053/gast.2002.32369]

21 **Liu R**, Lu Z, Gu J, Liu J, Huang E, Liu X, Wang L, Yang J, Deng Y, Qian J, Luo F, Wang Z, Zhang H, Jiang X, Zhang D, Qian J, Liu G, Zhu H, Qian Y, Liu Z, Chu Y. MicroRNAs 15A and 16-1 Activate Signaling Pathways That Mediate Chemotaxis of Immune Regulatory B cells to Colorectal Tumors. *Gastroenterology* 2018; **154**: 637-651.e7 [PMID: 29031499 DOI: 10.1053/j.gastro.2017.09.045]

22 **Salaroglio IC**, Panada E, Moiso E, Buondonno I, Provero P, Rubinstein M, Kopecka J, Riganti C. PERK induces resistance to cell death elicited by endoplasmic reticulum stress and chemotherapy. *Mol Cancer* 2017; **16**: 91 [PMID: 28499449 DOI: 10.1186/s12943-017-0657-0]

23 **Cheng X**, Feng H, Wu H, Jin Z, Shen X, Kuang J, Huo Z, Chen X, Gao H, Ye F, Ji X, Jing X, Zhang Y, Zhang T, Qiu W, Zhao R. Targeting autophagy enhances apatinib-induced apoptosis *via* endoplasmic reticulum stress for human colorectal cancer. *Cancer Lett* 2018; **431**: 105-114 [PMID: 29859300 DOI: 10.1016/j.canlet.2018.05.046]

24 **Tiemersma EW**, Wark PA, Ocké MC, Bunschoten A, Otten MH, Kok FJ, Kampman E. Alcohol consumption, alcohol dehydrogenase 3 polymorphism, and colorectal adenomas. *Cancer Epidemiol Biomarkers Prev* 2003; **12**: 419-425 [PMID: 12750236]

25 **Liao X**, Huang R, Liu X, Han C, Yu L, Wang S, Sun N, Li B, Ning X, Peng T. Distinct prognostic values of alcohol dehydrogenase mRNA expression in pancreatic adenocarcinoma. *Onco Targets Ther* 2017; **10**: 3719-3732 [PMID: 28769575 DOI: 10.2147/OTT.S140221]

26 **Seitz HK**, Simanowski UA, Garzon FT, Rideout JM, Peters TJ, Koch A, Berger MR, Einecke H, Maiwald M. Possible role of acetaldehyde in ethanol-related rectal cocarcinogenesis in the rat. *Gastroenterology* 1990; **98**: 406-413 [PMID: 2295396 DOI: 10.1016/0016-5085(90)90832-l]

27 **Indini A**, Petrelli F, Tomasello G, Rijavec E, Facciorusso A, Grossi F, Ghidini M. Impact of Use of Gastric-Acid Suppressants and Oral Anti-Cancer Agents on Survival Outcomes: A Systematic Review and Meta-Analysis. *Cancers (Basel)* 2020; **12** [PMID: 32325628 DOI: 10.3390/cancers12040998]

28 **Kurmi K**, Haigis MC. Nitrogen Metabolism in Cancer and Immunity. *Trends Cell Biol* 2020; **30**: 408-424 [PMID: 32302552 DOI: 10.1016/j.tcb.2020.02.005]

29 **Zaytouni T**, Tsai PY, Hitchcock DS, DuBois CD, Freinkman E, Lin L, Morales-Oyarvide V, Lenehan PJ, Wolpin BM, Mino-Kenudson M, Torres EM, Stylopoulos N, Clish CB, Kalaany NY. Critical role for arginase 2 in obesity-associated pancreatic cancer. *Nat Commun* 2017; **8**: 242 [PMID: 28808255 DOI: 10.1038/s41467-017-00331-y]

30 **Gmeiner WH**, Hellmann GM, Shen P. Tissue-dependent and -independent gene expression changes in metastatic colon cancer. *Oncol Rep* 2008; **19**: 245-251 [PMID: 18097602]

31 **Kodama M**, Oshikawa K, Shimizu H, Yoshioka S, Takahashi M, Izumi Y, Bamba T, Tateishi C, Tomonaga T, Matsumoto M, Nakayama KI. A shift in glutamine nitrogen metabolism contributes to the malignant progression of cancer. *Nat Commun* 2020; **11**: 1320 [PMID: 32184390 DOI: 10.1038/s41467-020-15136-9]

32 **Yang Y**, Zheng Y, Liu X, Ji R, Chen Z, Guo Q, Wu G, Wang Y, Zhou Y. Comprehensive analysis of gene regulation network and immune signatures of prognostic biomarker YAP1 in pancreatic cancer. *J Cancer* 2020; **11**: 6960-6969 [PMID: 33123286 DOI: 10.7150/jca.49117]

33 **Shen Y**, Lai Y, Xu D, Xu L, Song L, Zhou J, Song C, Wang J. Diagnosis of thyroid neoplasm using support vector machine algorithms based on platelet RNA-seq. *Endocrine* 2020 [PMID: 33179221 DOI: 10.1007/s12020-020-02523-x]

34 **Zhou L**, Li Y, Li Z, Huang Q. Mining therapeutic and prognostic significance of STATs in renal cell carcinoma with bioinformatics analysis. *Genomics* 2020; **112**: 4100-4114 [PMID: 32640276 DOI: 10.1016/j.ygeno.2020.06.032]

35 **Lee K**, Ferguson LR. MicroRNA biomarkers predicting risk, initiation and progression of colorectal cancer. *World J Gastroenterol* 2016; **22**: 7389-7401 [PMID: 27672263 DOI: 10.3748/wjg.v22.i33.7389]

36 **Liu C**, Tian X, Sun HB, Wang ZF, Jiang LF, Li ZX. MiR-601 inhibits the proliferation and metastasis of esophageal squamous cell carcinoma (ESCC) by targeting HDAC6. *Eur Rev Med Pharmacol Sci* 2019; **23**: 1069-1076 [PMID: 30779074 DOI: 10.26355/eurrev\_201902\_16995]

37 **Hu JY**, Yi W, Wei X, Zhang MY, Xu R, Zeng LS, Huang ZJ, Chen JS. miR-601 is a prognostic marker and suppresses cell growth and invasion by targeting PTP4A1 in breast cancer. *Biomed Pharmacother* 2016; **79**: 247-253 [PMID: 27044835 DOI: 10.1016/j.biopha.2016.02.014]

38 **Li X**, Nie C, Tian B, Tan X, Han W, Wang J, Jin Y, Li Y, Guan X, Hong A, Chen X. miR-671-5p Blocks The Progression Of Human Esophageal Squamous Cell Carcinoma By Suppressing FGFR2. *Int J Biol Sci* 2019; **15**: 1892-1904 [PMID: 31523191 DOI: 10.7150/ijbs.32429]

39 **Tan X**, Li Z, Ren S, Rezaei K, Pan Q, Goldstein AT, Macri CJ, Cao D, Brem RF, Fu SW. Dynamically decreased miR-671-5p expression is associated with oncogenic transformation and radiochemoresistance in breast cancer. *Breast Cancer Res* 2019; **21**: 89 [PMID: 31391072 DOI: 10.1186/s13058-019-1173-5]

40 **Li X**, Diao H. Circular RNA circ\_0001946 acts as a competing endogenous RNA to inhibit glioblastoma progression by modulating miR-671-5p and CDR1. *J Cell Physiol* 2019; **234**: 13807-13819 [PMID: 30663767 DOI: 10.1002/jcp.28061]

41 **Gartner JJ**, Parker SC, Prickett TD, Dutton-Regester K, Stitzel ML, Lin JC, Davis S, Simhadri VL, Jha S, Katagiri N, Gotea V, Teer JK, Wei X, Morken MA, Bhanot UK; NISC Comparative Sequencing Program, Chen G, Elnitski LL, Davies MA, Gershenwald JE, Carter H, Karchin R, Robinson W, Robinson S, Rosenberg SA, Collins FS, Parmigiani G, Komar AA, Kimchi-Sarfaty C, Hayward NK, Margulies EH, Samuels Y. Whole-genome sequencing identifies a recurrent functional synonymous mutation in melanoma. *Proc Natl Acad Sci USA* 2013; **110**: 13481-13486 [PMID: 23901115 DOI: 10.1073/pnas.1304227110]

42 **Della Vittoria Scarpati G**, Falcetta F, Carlomagno C, Ubezio P, Marchini S, De Stefano A, Singh VK, D'Incalci M, De Placido S, Pepe S. A specific miRNA signature correlates with complete pathological response to neoadjuvant chemoradiotherapy in locally advanced rectal cancer. *Int J Radiat Oncol Biol Phys* 2012; **83**: 1113-1119 [PMID: 22172905 DOI: 10.1016/j.ijrobp.2011.09.030]

43 **Jiang B**, Xu G, Lv HQ, Huang M, Li Z. Up-regulation of miR-765 predicts a poor prognosis in patients with esophageal squamous cell carcinoma. *Eur Rev Med Pharmacol Sci* 2018; **22**: 3789-3794 [PMID: 29949154 DOI: 10.26355/eurrev\_201806\_15261]

44 **Xie BH**, He X, Hua RX, Zhang B, Tan GS, Xiong SQ, Liu LS, Chen W, Yang JY, Wang XN, Li HP. Mir-765 promotes cell proliferation by downregulating INPP4B expression in human hepatocellular carcinoma. *Cancer Biomark* 2016; **16**: 405-413 [PMID: 27062697 DOI: 10.3233/CBM-160579]

45 **Lv DB**, Zhang JY, Gao K, Yu ZH, Sheng WC, Yang G, Gao YZ. MicroRNA-765 targets MTUS1 to promote the progression of osteosarcoma *via* mediating ERK/EMT pathway. *Eur Rev Med Pharmacol Sci* 2019; **23**: 4618-4628 [PMID: 31210288 DOI: 10.26355/eurrev\_201906\_18040]

46 **Zhou R**, Huang W, Yao Y, Wang Y, Li Z, Shao B, Zhong J, Tang M, Liang S, Zhao X, Tong A, Yang J. CA II, a potential biomarker by proteomic analysis, exerts significant inhibitory effect on the growth of colorectal cancer cells. *Int J Oncol* 2013; **43**: 611-621 [PMID: 23727877 DOI: 10.3892/ijo.2013.1972]

47 **Viikilä P**, Kivelä AJ, Mustonen H, Koskensalo S, Waheed A, Sly WS, Pastorek J, Pastorekova S, Parkkila S, Haglund C. Carbonic anhydrase enzymes II, VII, IX and XII in colorectal carcinomas. *World J Gastroenterol* 2016; **22**: 8168-8177 [PMID: 27688658 DOI: 10.3748/wjg.v22.i36.8168]

48 **Mori M**, Staniunas RJ, Barnard GF, Jessup JM, Steele GD Jr, Chen LB. The significance of carbonic anhydrase expression in human colorectal cancer. *Gastroenterology* 1993; **105**: 820-826 [PMID: 8359652 DOI: 10.1016/0016-5085(93)90900-w]

49 **Zhang J**, Tsoi H, Li X, Wang H, Gao J, Wang K, Go MY, Ng SC, Chan FK, Sung JJ, Yu J. Carbonic anhydrase IV inhibits colon cancer development by inhibiting the Wnt signalling pathway through targeting the WTAP-WT1-TBL1 axis. *Gut* 2016; **65**: 1482-1493 [PMID: 26071132 DOI: 10.1136/gutjnl-2014-308614]

50 **Zengin Kurt B**, Sonmez F, Ozturk D, Akdemir A, Angeli A, Supuran CT. Synthesis of coumarin-sulfonamide derivatives and determination of their cytotoxicity, carbonic anhydrase inhibitory and molecular docking studies. *Eur J Med Chem* 2019; **183**: 111702 [PMID: 31542715 DOI: 10.1016/j.ejmech.2019.111702]

51 **Wu Z**, Liu Z, Ge W, Shou J, You L, Pan H, Han W. Analysis of potential genes and pathways associated with the colorectal normal mucosa-adenoma-carcinoma sequence. *Cancer Med* 2018; **7**: 2555-2566 [PMID: 29659199 DOI: 10.1002/cam4.1484]

52 **Maeda K**, Saigo C, Kito Y, Sakuratani T, Yoshida K, Takeuchi T. Expression of TMEM207 in Colorectal Cancer: Relation between TMEM207 and Intelectin-1. *J Cancer* 2016; **7**: 207-213 [PMID: 26819645 DOI: 10.7150/jca.13732]

53 **Li D**, Mei H, Pu J, Xiang X, Zhao X, Qu H, Huang K, Zheng L, Tong Q. Intelectin 1 suppresses the growth, invasion and metastasis of neuroblastoma cells through up-regulation of N-myc downstream regulated gene 2. *Mol Cancer* 2015; **14**: 47 [PMID: 25889839 DOI: 10.1186/s12943-015-0320-6]

54 **Au-Yeung CL**, Yeung TL, Achreja A, Zhao H, Yip KP, Kwan SY, Onstad M, Sheng J, Zhu Y, Baluya DL, Co NN, Rynne-Vidal A, Schmandt R, Anderson ML, Lu KH, Wong STC, Nagrath D, Mok SC. ITLN1 modulates invasive potential and metabolic reprogramming of ovarian cancer cells in omental microenvironment. *Nat Commun* 2020; **11**: 3546 [PMID: 32669559 DOI: 10.1038/s41467-020-17383-2]

55 **Zhang Y**, Wang J, Liu X. LRRC19-A Bridge between Selenium Adjuvant Therapy and Renal Clear Cell Carcinoma: A Study Based on Datamining. *Genes (Basel)* 2020; **11** [PMID: 32316597 DOI: 10.3390/genes11040440]

56 **Payton S**. Infection: LRRC19 mediates elimination of uropathogenic bacteria from kidney. *Nat Rev Urol* 2014; **11**: 482 [PMID: 25091007 DOI: 10.1038/nrurol.2014.199]

57 **Rajeshkumar NV**, Tan AC, De Oliveira E, Womack C, Wombwell H, Morgan S, Warren MV, Walker J, Green TP, Jimeno A, Messersmith WA, Hidalgo M. Antitumor effects and biomarkers of activity of AZD0530, a Src inhibitor, in pancreatic cancer. *Clin Cancer Res* 2009; **15**: 4138-4146 [PMID: 19509160 DOI: 10.1158/1078-0432.CCR-08-3021]

**Footnotes**

**Institutional review board statement:** This study was approved by the Ethics Committee of China-Japan Friendship Hospital, No. 2018-116-K85-1.

**Conflict-of-interest statement:** There are no conflicts of interest to report.

**Data sharing statement:** No additional data are available.

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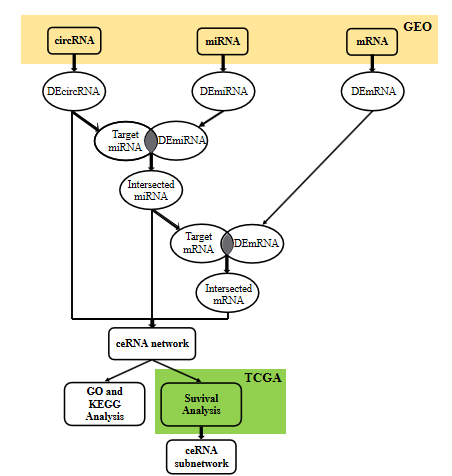
Grade C (Good): 0

Grade D (Fair): 0

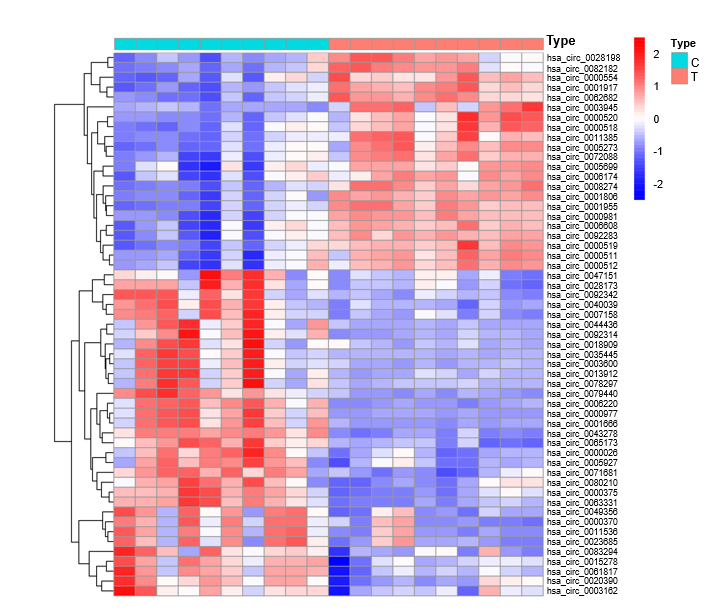
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**P-Reviewer:** Koumarianou A **S-Editor:** Fan JR **L-Editor: P-Editor:**

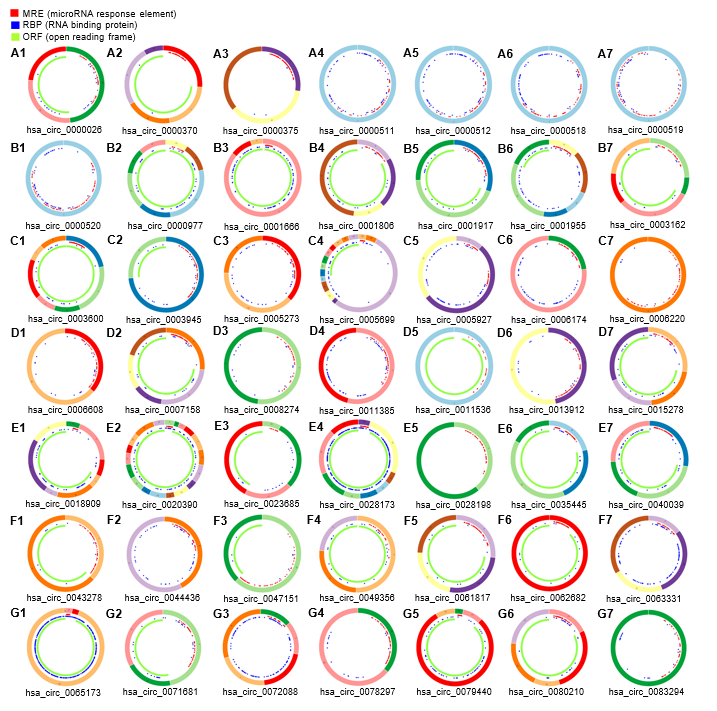
**Figure Legends**



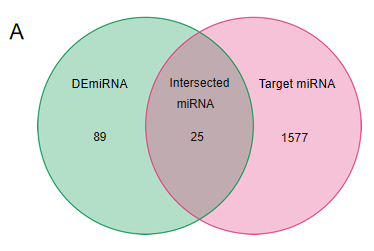
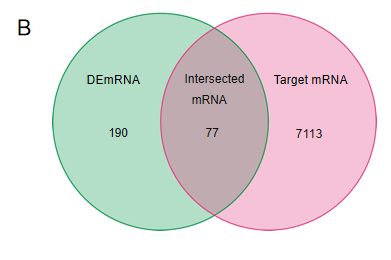
**Figure 1 Flow chart of the procedure applied in this study.** We identified the differentially expressed (DE) circRNAs, DEmiRNAs and DEmRNAs and then took the intersection of DEmiRNAs and DEmRNAs with the predicted targets of DEcircRNAs and intersected miRNA to obtain the competing endogenous RNA network in the Group on Earth Observations database, performed GO and KEGG enrichment analyses, and constructed a prognostic subnetwork based on survival analysis in The Cancer Genome Atlas database. DE: Differentially expressed; TCGA: The Cancer Genome Atlas; ceRNA: Competing endogenous RNA.

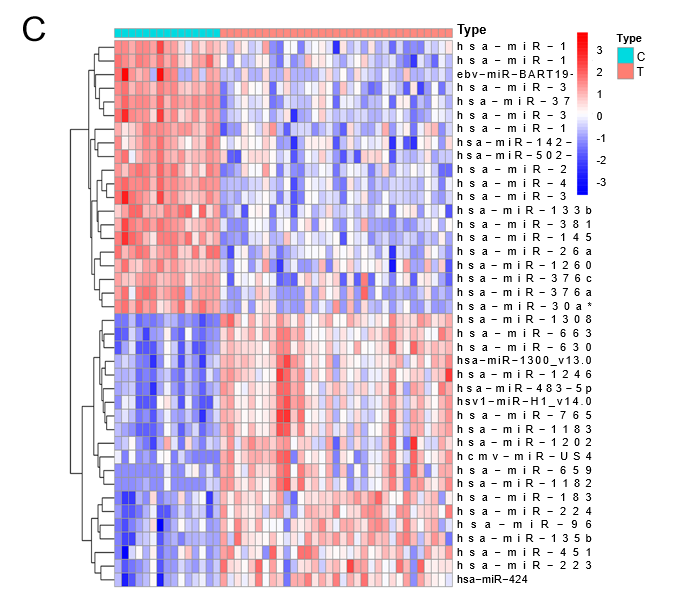


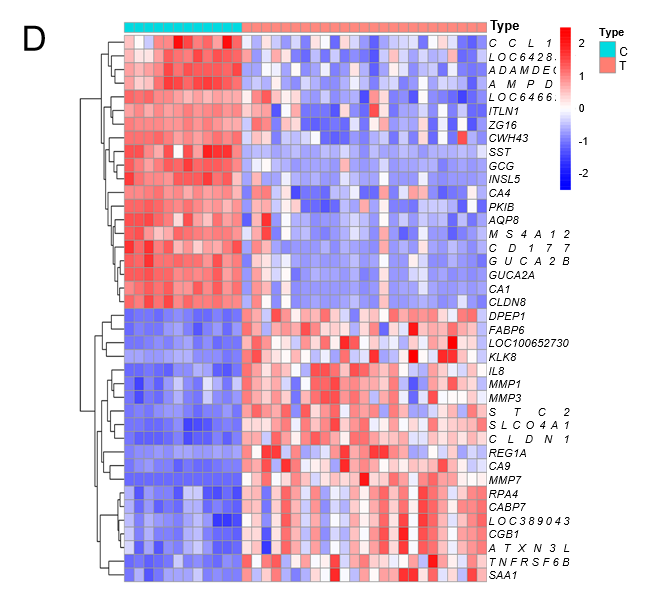
**Figure 2 Heat map of circRNAs differentially expressed in GSE126095.** By analyzing the circRNA profile, 55 differentially expressed circRNAs were obtained. Red indicates high expression, while blue indicates low expression. A deeper color indicates a more significant difference. The cut-off value was a |log2 fold change| > 2 and an adjusted *P* < 0.05.



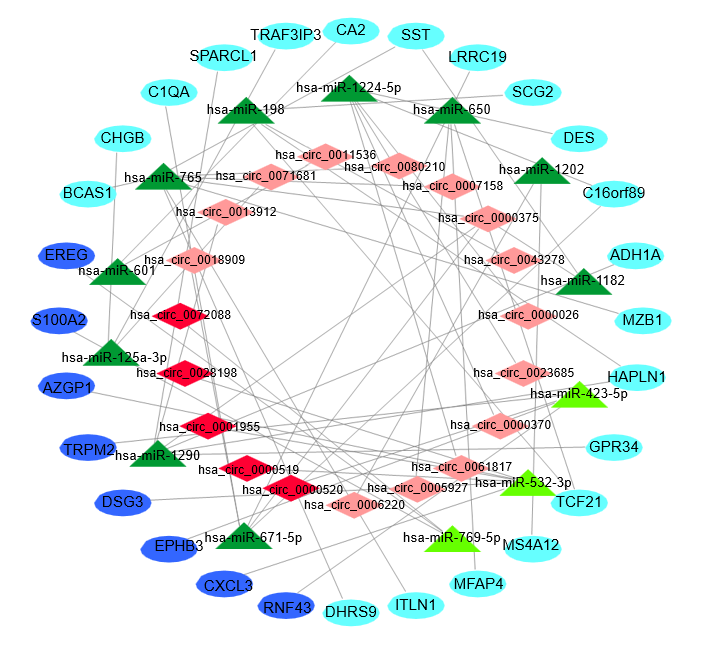
**Figure 3 Structural pattern of 49 differentially expressed circRNAs obtained from Cancer-Specific CircRNA Database.** Red indicates the miRNA response element, blue indicates the RNA binding protein, and green indicates the open reading frame. CSCD: Cancer-Specific CircRNA Database (http://gb.whu.edu.cn/CSCD/).

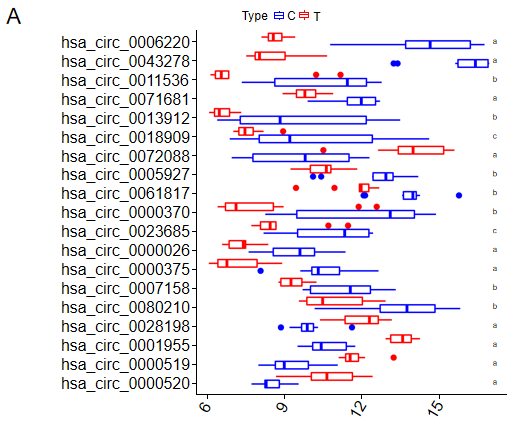


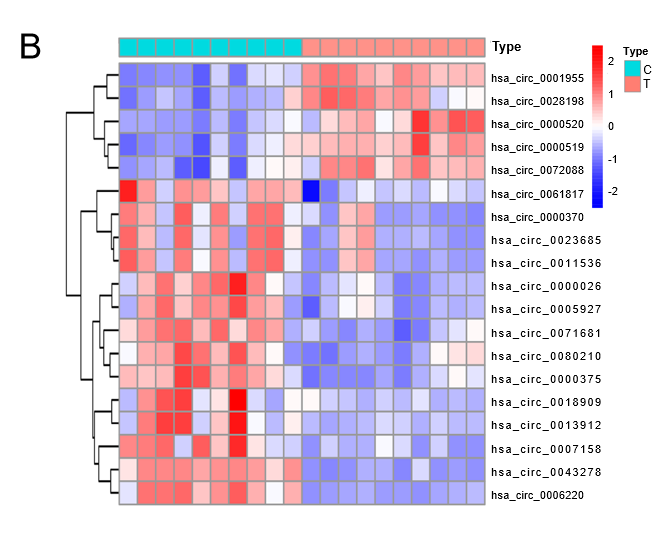


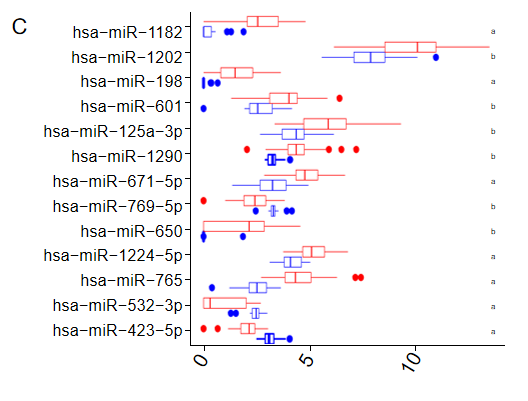
**Figure 4 Venn diagrams and heat maps of miRNAs and mRNAs.** A: Twenty-five miRNAs were identified from the intersection of 114 differentially expressed (DE) miRNAs with 1602 circRNA targets predicted by the Cancer-Specific CircRNA Database; B: 77 mRNAs were obtained from the intersection of 267 DEmRNAs and 7190 miRNA targets predicted using the TargetScan and miRDB databases; C: Each 20 DEmiRNAs expressing the most significant upregulation and downregulation, respectively; D. Each 20 DEmRNAs expressing the most significant upregulation and downregulation, respectively. |log2 fold change (FC)| > 1 and an adjusted *P* < 0.05 were considered the statistical criteria for DEmiRNA; adjusted *P* < 0.05 and |log2 FC| > 2 were considered the statistical criteria for DEmRNA. DE: Differentially expressed.

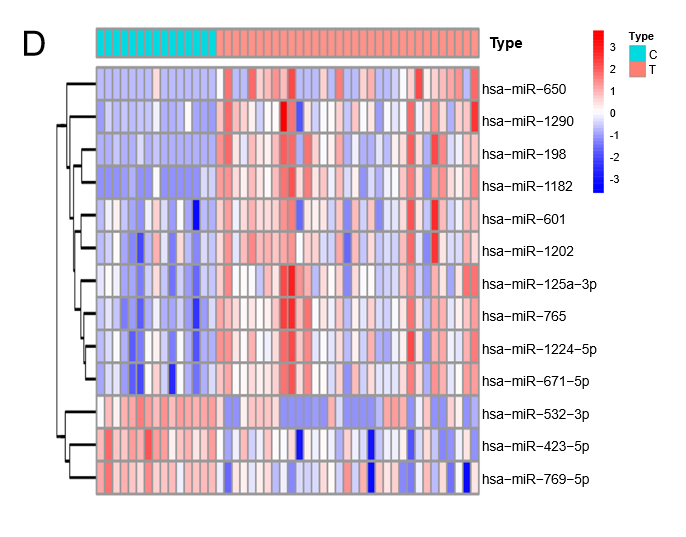


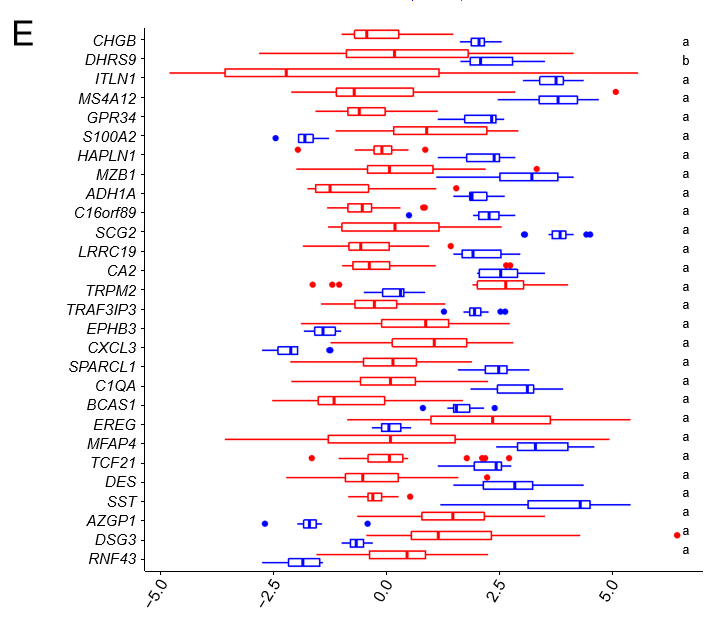
**Figure 5 CircRNA-miRNA-mRNA regulatory network of colorectal cancer.** CircRNAs, miRNAs and mRNAs are represented by red diamonds, green triangles and blue ellipses, respectively. Dark colors represent overexpression, and light colors represent low expression.

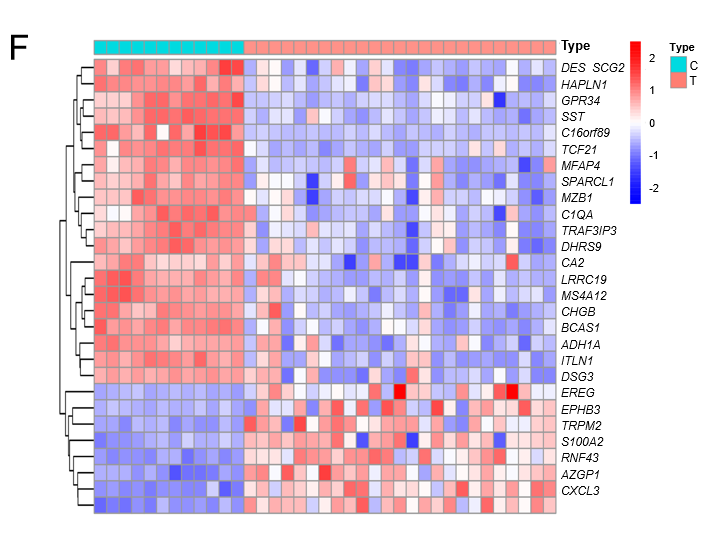




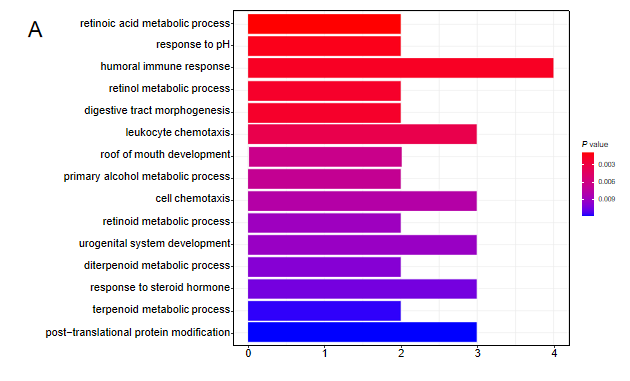


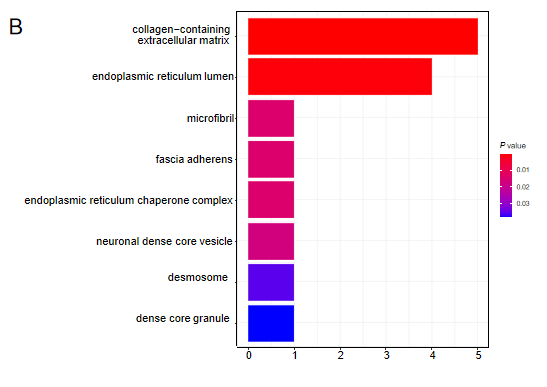


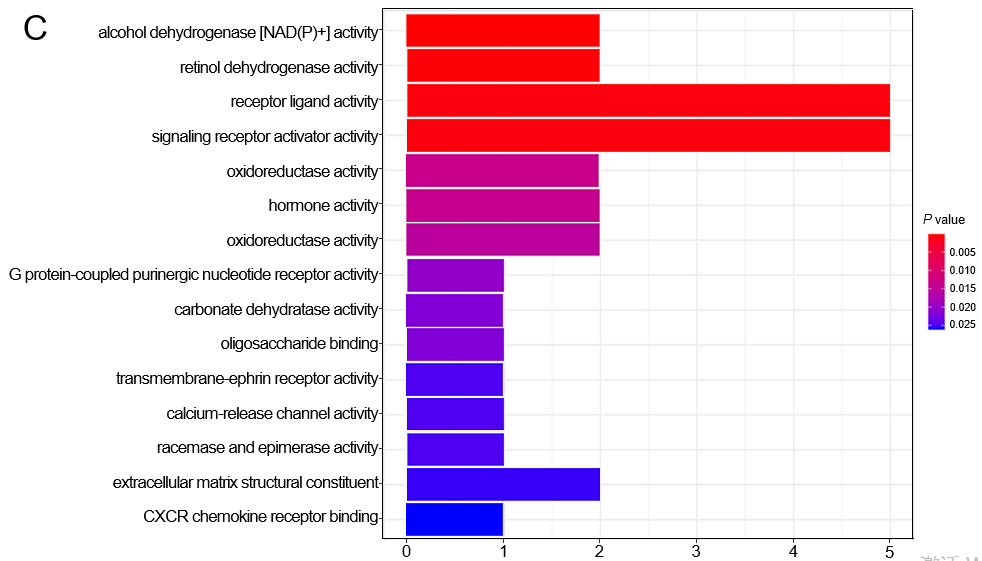


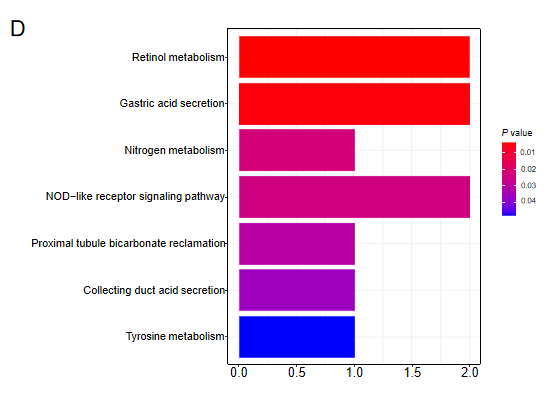


**Figure 6 The expression levels of circRNAs, miRNAs and mRNAs in the** competing endogenous RNA **network.** A and B: Expression levels of 19 circRNAs in the competing endogenous RNA (ceRNA) network; C and D: Expression levels of 13 miRNAs in the ceRNA network; E and F: Expression levels of 25 mRNAs in the ceRNA network. a*P* < 0.001; b*P* < 0.01; c*P* < 0.05.

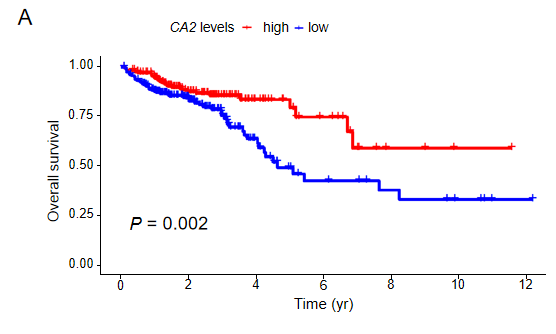
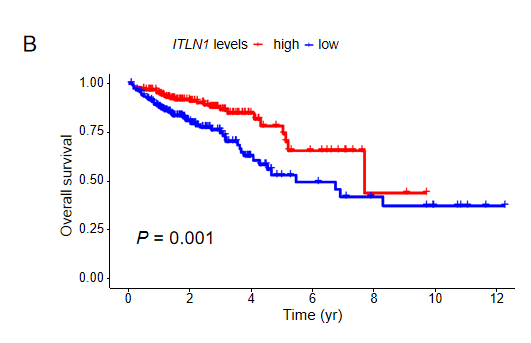


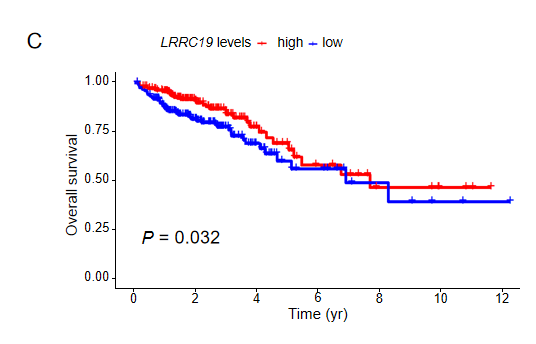




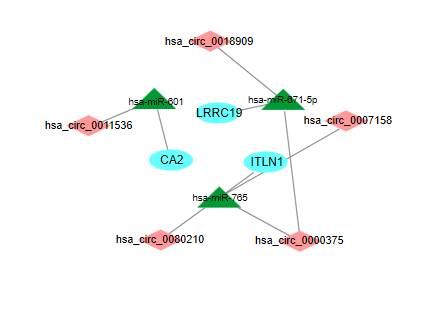


**Figure 7 GO and KEGG pathway analyses of the identified mRNAs in the competing endogenous RNA network.** A: GO analysis of biological processes; B: GO analysis of cellular components; C: GO analysis of molecular functions; D: KEGG pathway analysis. *P* < 0.05 were considered significant.



**Figure 8 Survival analysis of the identified mRNAs in the circRNA-miRNA-mRNA regulatory network using the cancer genome atlas database**. A: Comparison between patients with upregulation or downregulation of *CA2* (*P* = 0.002); B: Comparison between patients with upregulation or downregulation of *ITLN1* (*P* = 0.001); C: Comparison between patients with upregulation or downregulation of *LRRC19* (*P* = 0.032).



**Figure 9 The competing endogenous RNA subnetwork associated with colorectal cancer prognosis.** Based on mRNAs with potential for the clinical outcome prediction of colorectal cancer, a prognostic competing endogenous RNA subnetwork was successfully developed. Diamonds, triangles and ellipses represent circRNAs, miRNAs and mRNAs, respectively.

**Table 1 Basic characteristics of 4 microarray datasets in Group on Earth Observations and The Cancer Genome Atlas**

|  |  |  |  |  |
| --- | --- | --- | --- | --- |
| **Data source** | **Platform** | **Series** | **Sample size** | |
| **Tumor** | **Control** |
| circRNA | GPL19978 | GSE126095 | 10 | 10 |
| miRNA | GPL11487 | GSE41655 | 33 | 15 |
| mRNA | GPL6480 | GSE41657 | 25 | 12 |
| mRNA | TCGA | None | 488 | 42 |

TCGA: The Cancer Genome Atlas.