

WJG 20th Anniversary Special Issues (14): Pancreatic cancer**Hedgehog signaling pathway as a new therapeutic target in pancreatic cancer**

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Author contributions: Onishi H and Katano M analyzed the data; Onishi H wrote the paper.

Supported by The Japan Society for the Promotion of Science, Kakenhi Grant, No. 24390303

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Received: October 7, 2013 Revised: December 11, 2013

Accepted: January 8, 2014

Published online: March 7, 2014

Core tip: Hedgehog (Hh) signaling is involved in the induction of malignant potential in pancreatic cancer, controlling processes of proliferation, invasiveness and tumorigenesis. This phenotypic change is closely associated with the nuclear factor kappa-light-chain-enhancer of activated B cells transcription factor, both in an autocrine and paracrine manner. Hh signaling is also capable of maintaining pancreatic cancer stem cells, and may be activated under conditions of tumor hypoxia. Thus, the Hh signaling pathway may represent a potential therapeutic target for patients with refractory pancreatic cancer and the use of Hh inhibitors will likely play an important role in future therapeutic strategies.

Abstract

Pancreatic cancer is one of the most aggressive and difficult cancers to treat. Despite numerous research efforts, limited success has been achieved in the therapeutic management of patients with this disease. In the current review, we focus on one component of morphogenesis signaling, Hedgehog (Hh), with the aim of developing novel, effective therapies for the treatment of pancreatic cancer. Hh signaling contributes to the induction of a malignant phenotype in pancreatic cancer and is responsible for maintaining pancreatic cancer stem cells. In addition, we propose a novel concept linking Hh signaling and tumor hypoxic conditions, and discuss the effects of Hh inhibitors in clinical trials. The Hh signaling pathway may represent a potential therapeutic target for patients with refractory pancreatic cancer.

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Key words: Hedgehog signaling pathway; Pancreatic cancer; Cancer stem cells; Hypoxic condition; Therapeutic target

Onishi H, Katano M. Hedgehog signaling pathway as a new therapeutic target in pancreatic cancer. *World J Gastroenterol* 2014; 20(9): 2335-2342 Available from: URL: <http://www.wjgnet.com/1007-9327/full/v20/i9/2335.htm> DOI: <http://dx.doi.org/10.3748/wjg.v20.i9.2335>

INTRODUCTION

Pancreatic cancer remains one of the deadliest cancers, with an overall survival rate of < 5%^[1]. An underlying reason for this may be that few patients undergo curative, surgical operations because of the advanced stage of the cancer at the time of diagnosis. Furthermore, apart from chemotherapy and radiation therapy, there are no effective, alternative therapies for the treatment of refractory pancreatic cancer, and as such, the development of novel therapeutic strategies is urgently required. Recently, it was shown that the Hedgehog (Hh) signaling pathway, which plays a key role in morphogenesis signaling, is re-activated in pancreatic cancer^[2]. Hh signaling contributes to tumor aggressiveness, affecting key tumorigenic processes such as proliferation, invasion

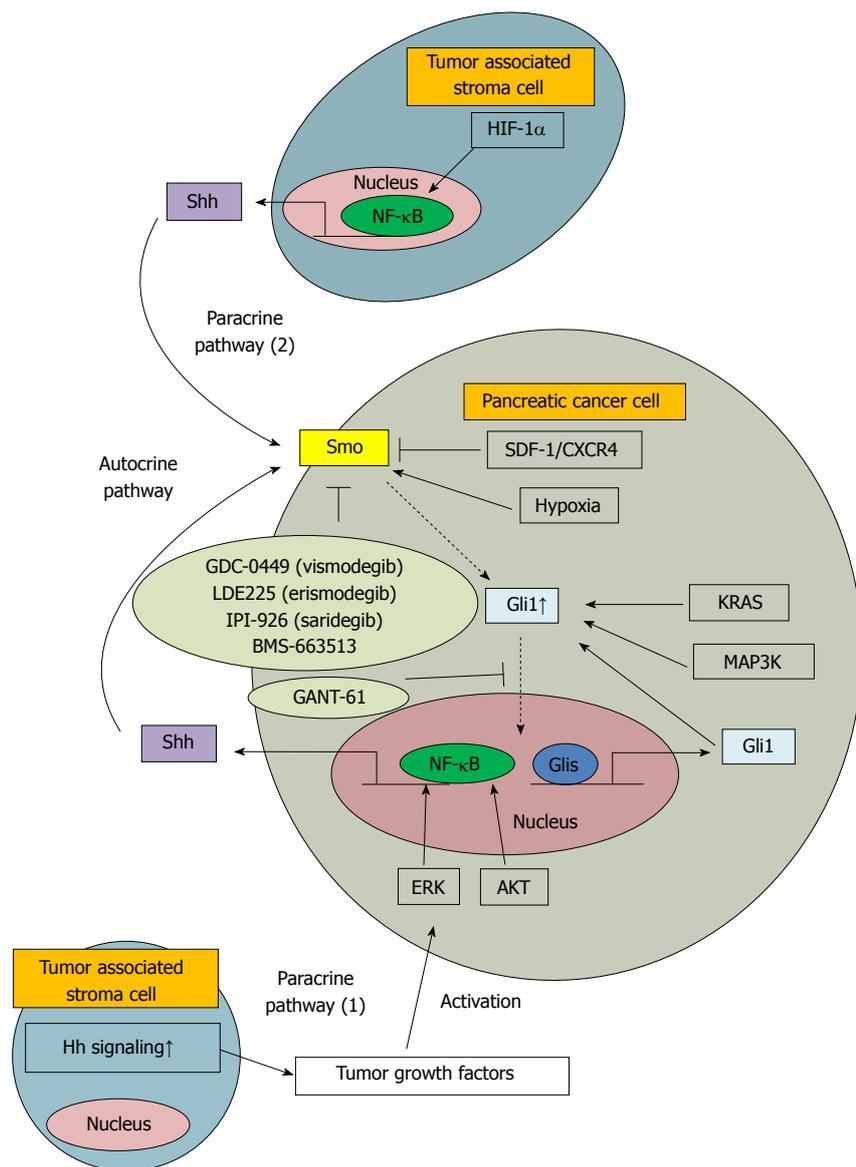


Figure 1 Schematic review. Hedgehog (Hh) signaling is activated in both autocrine and paracrine pathways. Tumor associated stroma cells play a pivotal role in tumor progression related to activation of Hh signaling [paracrine pathways (1) and (2)]. Induction of sonic Hh (Shh) is closely associated with activation of the nuclear factor kappa-light-chain-enhancer of activated B cells transcription factor in pancreatic cancer. Shh is produced by NF-κB activation in pancreatic cancer cells and tumor associated stroma cells. Pathways contributing to Smo and Gli1 activation include SDF-1/CXCR4, hypoxia, KRAS and MAP3K. The effects of Hh inhibitors including GDC-0449 (vismodegib), LDE225 (erismodegib), IPI-926 (saridegib), BMS-663513 and GANT-61 in clinical trials are under investigation. Dotted arrows show components of the Hh signaling pathway in tumor cells focused on in this review.

and progression of cancer cells. Therefore, inhibitors targeting Hh signaling have drawn significant attention as novel, molecularly targeted drugs. Hh signaling components including Patched and Smoothed (Smo) have been detected in almost 70% of human pancreatic cancer specimens and consequently, Hh signaling may play a critical role in the genesis of pancreatic cancer cells^[21]. In this review, we summarize recent efforts in the development of new, therapeutic strategies to treat pancreatic cancer, targeting the Hh signaling pathway.

HH SIGNALING PATHWAY

The Hh signaling pathway plays a pivotal role in em-

bryonic patterning and growth control, acting as a morphogen, mitogen and inducing factor of developing organs^[3-7]. Hh signaling normally ceases after embryogenesis, however in various cancers, including pancreatic cancer, Hh signaling is re-activated^[8]. Therefore, the regulation of Hh signaling in pancreatic cancer likely plays important role in tumorigenesis. The Hh signaling pathway is composed of Hh proteins (sonic Hh; Shh, Indian Hh and Desert Hh), the 12-transmembrane Patched proteins (Patched 1 and Patched 2), the 7-transmembrane protein, Smo and the 5-zinc-finger transcription factors, Gli1, Gli2 and Gli3^[9-11]. In the absence of Hh ligand, Patched suppresses Smo, which is the driving protein for Hh signaling, and Gli2 and Gli3 are cleaved

by ubiquitin ligases to generate transcriptional repressor isoforms^[12-14]. In contrast, in the presence of Hh ligand, inhibition of Smo by Patched is released, Smo is activated, and Gli2 and Gli3 are transmitted to the nucleus as full-length activators leading to the transcription of target genes such as Patched and Gli1^[12-14]. Recent studies demonstrated the existence of primary cilia on the cell surface and showed that Smo moves from the cytoplasm to primary cilia in the process of activation^[15]. One of the target genes of Hh signaling; Ptch and Gli1 regulate the transcription of the Hh responsive genes by themselves^[16]. Other target genes of Hh signaling are the cell cycle regulator Cyclin D1, p21 and N-Myc which plays important role for carcinogenesis and is also typically dysregulated in the cancer cells^[7,17,18]. The Hh signaling pathway is unique because several components of this pathway consist of both oncogenes and cancer suppressor genes.

HH SIGNALING AND THE INDUCTION OF MALIGNANT POTENTIAL IN PANCREATIC CANCER

Originally, the relationship between Hh signaling and tumorigenesis was reported following the association of mutations in genes such as *Gli1*, *Patch* and *Smo* in glioblastoma, basal cell carcinoma and rhabdomyosarcoma^[19-21]. In pancreatic cancer, ligand-dependent activation of Hh signaling, but not genomic mutation, was first reported^[2]. Previous studies have also shown that Shh overexpression is sufficient to initiate pancreatic intraepithelial neoplasia (PanIN)-like precursor lesions^[2,22]. At present, this ligand-dependent pathway is thought to be the major mechanism underlying Hh signaling activation. Two distinct ligand-dependent activation pathways exist; autocrine and paracrine. In addition, association between chronic inflammation and the development of cancer has been recognized for several years^[23-27]. In both autocrine and paracrine pathways, NF- κ B plays a pivotal role. NF- κ B is a transcription factor that controls expression of numerous genes involved in inflammation and immune response processes, including proliferation, invasion, adhesion, angiogenesis and apoptosis^[28]. In the autocrine pathway, Shh is a direct transcriptional target of NF- κ B, and proliferation of pancreatic cancer cells is accelerated *via* overexpression of Shh^[29,30]. In the paracrine paradigm, tumor-associated stroma is important as a microenvironmental factor^[31,32]. In one paracrine pathway, stroma cells surrounding pancreatic ductal adenocarcinoma cells, secrete tumor-growth factors through stromal Hh signaling activation^[31]. This may explain why low concentrations of Hh signaling antagonist are sufficient to inhibit tumor growth [paracrine pathway (1), Figure 1]^[31]. In an alternative paracrine pathway, NF- κ B-activated monocytes located in the tumor stromal area produce Shh, which stimulates the Hh signaling pathway in pancreatic cancer [paracrine pathway (2), Figure 1]^[33]. Inhibition of

Hh signaling targets pancreatic stellate cells in the tumor-associated stroma, specifically reducing pancreatic tumor growth and metastasis^[34,35]. In addition, Singh *et al*^[36] showed that CXCL12/CXCR4 protein signaling induces Shh expression in pancreatic cancer *via* extracellular regulated kinase (ERK) and Akt kinase-mediated activation of NF- κ B. Some other molecules affected by the activation of Hh signaling may also contribute to the induction of malignant potential in pancreatic cancer. Decrease in Cyclin D1 by the inhibition of Hh signaling induces the G₀/G₁ arrest and inhibits cell proliferation^[37]. Matrix metalloproteinase (MMP)-9 and MMP-2 locate the downstream of Gli1 and are involved with the invasiveness in pancreatic cancer^[38,39].

HH SIGNALING AND PANCREATIC CANCER STEM CELLS

Solid tumor cancer stem cells were first identified in breast cancer as CD24^{-/low}CD44⁺ cells^[40]. CD44⁺CD24⁺ epithelial-specific antigen (ESA)⁺ pancreatic cancer cells are reported to exhibit the stem cell characteristics of self-renewal and the ability to produce differentiated progeny^[41]. Most importantly, cancer stem cells (CSCs) are characterized by features of resistance towards conventional chemotherapy and radiotherapy^[42-45]. Pancreatic CSCs exhibit upregulation of Shh^[46]. Recently, inhibition of Hh signaling was reported to inhibit the self-renewal of pancreatic CSCs and reverse chemoresistance^[47]. Subsequent studies demonstrated that various agents were capable of inhibiting pancreatic CSCs *via* suppression of Hh signaling. For example, Tang *et al*^[48] revealed that epigallocatechin-3-gallate, an active compound in green tea, inhibits the self-renewal capacity of pancreatic CSCs *via* inhibition of Hh signaling components including Smo, Ptch, Gli1 and Gli2. Other groups demonstrated that sulforaphane, a component of dietary cruciferous vegetables, decreases pancreatic CSC self-renewal *via* inhibition of Hh signaling components, Smo, Gli1 and Gli2^[49,50]. Han *et al*^[51] has revealed that suppression of Hh signaling by arsenic trioxide leads to the inhibition of the viability of pancreatic CSCs using animal models. A better understanding of the molecular pathways driving CSCs will lead to the development of effective, new therapeutic approaches for the treatment of pancreatic cancer.

As previously discussed, there are numerous reports describing CD44⁺CD24⁺ double positive cells in pancreatic CSCs. However to date, there have been relatively few studies investigating CD24 or CD44 molecules alone as therapeutic targets in pancreatic CSCs. CD24 is a unique molecule because it is described as a marker of pancreatic CSCs, whereas it is expressed at low levels or is absent in breast CSCs. CD24 is thought to act as an adhesion molecule^[52,53]. Recently, truncated Gli1 was shown to induce clinically more aggressive cancer *via* the increased expression of CD24^[54]. Ringel *et al*^[55] showed that constitutive expression of CD44 variants may also be associated with

the malignant state of invasive pancreatic carcinoma. However the precise roles CD24 and CD44 in pancreatic CSCs remain unclear.

HH SIGNALING AND HYPOXIA

Pancreatic cancer is thought to occur under high levels of hypoxia^[56]. Therefore, a detailed understanding of the hypoxic microenvironment is crucial for developing effective therapeutic approaches to treat this malignancy. Previous studies have shown that the oxygen concentration in venous blood and deep tumor environments is 5.3% and 1.3%, respectively^[57,58]. Thus, to accurately analyze the molecular mechanisms underlying pancreatic cancer, experiments performed under hypoxic conditions are required. The relationship between hypoxia and Hh signaling activation was first reported in 2011, with a study showing that hypoxia activates Hh signaling pathway by upregulating Smo transcription^[38]. Thereafter, it was reported that hypoxia induces epithelial to mesenchymal transition (EMT) *via* activation of Hh signaling^[59]. Interestingly, under hypoxic conditions, activation of Hh signaling is independent of hypoxia inducible factor (HIF)-1 α and is also ligand-independent, with no observable increase in Shh^[38,59]. Conversely, Spivak-Kroizman *et al.*^[60] showed that hypoxia and desmoplasia led to more aggressive and therapy-resistant tumors *via* activation of Hh signaling by Shh, due to HIF-1 α activation in the stroma. The mechanisms underlying activation of Hh signaling under hypoxic conditions remains unclear. However, given that Hh signaling is activated under tumor hypoxic conditions, this pathway may represent an important therapeutic target. Indeed, protein-bound polysaccharide decreases invasiveness and proliferation in pancreatic cancer by inhibition of Hh signaling, especially under hypoxia^[39].

HH SIGNALING AND THERAPEUTIC APPROACHES IN PANCREATIC CANCER

Pancreatic cancer is often refractory to standard treatments, and many patients are unable to undergo surgery because of the advanced stage of disease at the time of diagnosis. Chemotherapy using gemcitabine and 5-FU derivatives, Tegafur-Gimeracil-Oteracil Potassium (S-1), are often used in Japan. However, combined use of Hh inhibitors with gemcitabine or 5-FU may induce chemoresistance^[37]. One reason may be that gemcitabine and 5-FU are sensitive to S-phase and that Hh inhibitor often induces G₁ arrest in cancer cells^[37]. Conversely, several groups have shown that combined treatment with Hh inhibitors and gemcitabine has a synergistic effect on tumor growth in a xenograft model^[61]. Combined use of Hh inhibitors and cisplatin, a cell cycle independent drug, may also have a synergistic effect^[57]. Molecular targeting drug is now well established and the combined use of Hh inhibitors and other targeted drugs is currently being

studied and utilized. For example, there is a possible synergistic relationship between Hh and epidermal growth factor receptor (EGFR) signaling pathways in pancreatic cancer^[62-64]. Although combination therapy with Hh inhibitors remains controversial, these findings will be essential for developing new effective therapeutic strategies. Radiation is considered the third therapeutic strategy for the treatment of pancreatic cancer. Recently, focal radiation in combination with Hh inhibitors exhibited synergistic effects on reducing lymph node metastasis in pancreatic cancer^[65]. Immunotherapy is anticipated as the fourth line of therapy after surgery, chemotherapy and radiation. In this approach, activated lymphocytes and dendritic cells (DCs) derived from patients with advanced cancer are often used. Recently, it was reported that Hh signaling is revitalized in activated lymphocytes and DCs derived from patients with advanced cancer and used for immunotherapy, and that this plays a pivotal role in the maintenance of their functions^[66,67]. Therefore, Hh inhibitors may not have a synergistic effect when combined with immunotherapy.

Within the class of Hh inhibitors, recent drug development has focused on Smo inhibitors. Although exact patients' outcome has not been reported yet, Sekulic *et al.*^[68] has shown that the independently assessed response rate was 30% and 43%, and the median duration of response was 7.6 mo using two-cohort study with GDC-0449 (vismodegib) in metastatic and locally advanced basal-cell carcinoma. GDC-0449 and IPI-926 (saridegib) are currently under phase II clinical trials in metastatic, advanced and recurrent pancreatic cancer^[69] and BMS-663513 is under phase I clinical trial^[70]. A recent study demonstrated that LDE225 (erismodegib), a Smo antagonist, suppresses tumor growth and prolongs survival in a murine model of islet cell neoplasms^[71]. Furthermore, GANT-61, a Gli transcription factor inhibitor, has been shown to inhibit pancreatic cancer stem cell growth^[72]. An overview of Hh signaling inhibitors is shown in Figure 1. More recently, inhibition of Hh signaling has received significant attention as an anti-tumor strategy. Based on this, the relationship between Hh signaling and various materials has been reported. For instance, resveratrol, 3,4',5-trihydroxystilbene inhibits proliferation and induces apoptosis *via* Hh signaling in pancreatic cancer^[73]. Curcumin, a phenolic compound extracted from Zingiberaceae turmeric, reverses EMT of pancreatic cancer by inhibiting Hh signaling^[74]. Triparanol, a known cholesterol biosynthesis inhibitor blocking the 24-dehydrocholesterol reductase, suppresses pancreatic cancer tumor growth by deregulation of Hh signaling^[75].

Gli1 is both a transcription factor and a target gene, as shown in previous reviews, and crosstalk between Hh signaling and other pathways has been demonstrated^[8]. Gli1 is activated *via* several kinds of signaling pathways. In pancreatic cancer, various signaling pathways including KRAS^[76], ERK^[36], AKT^[36], MAP3K^[77] and SDF-1/CXCR4^[78] are associated with Hh signaling (Figure 1). Because Gli1 is located downstream in many of these

pathways, it may represent a better therapeutic target.

CONCLUSION

In this review, we have summarized the development of pancreatic cancer treatment, with specific focus on the Hh signaling pathway. The Hh signaling pathway may represent an important therapeutic target in pancreatic cancer because this pathway is activated in the majority of pancreatic cancers and both ligand-dependent and independent inhibitors are effective. Hh inhibitor can successfully inhibit tumor growth and invasiveness *in vitro* and can be a promising drug, however, in clinical trial, it is not easy to verify the effectiveness of Hh signaling inhibitor. This reason may be that the actual function of Hh signaling molecules are not fully understood^[79,80].

Hh signaling inhibitors should be effective in cancers in which Hh components are mutated such as basal cell carcinoma, basal cell nevus syndrome and medulloblastoma because Hh signaling is constitutively activated^[81]. And in these cancers, Hh signaling inhibitors may become the first use drug in future clinical life. However, for other tumors, appropriate combination therapy may be required for the effective therapy. In January 2012, the Smo inhibitor, vismodegib, was clinically approved for the first time by the US Food and Drug Administration, for the treatment of unresectable or metastatic basal cell carcinomas of the skin^[82]. Hh signaling inhibitors will now be used in pancreatic cancer as a monotherapy and in combination therapy with other chemodrugs, molecularly targeted drugs or radiation therapy.

ACKNOWLEDGMENTS

We thank Ms Kaori Nomiya for her skillful technical assistance.

REFERENCES

- 1 **Jemal A**, Siegel R, Ward E, Hao Y, Xu J, Thun MJ. Cancer statistics, 2009. *CA Cancer J Clin* 2009; **59**: 225-249 [PMID: 19474385 DOI: 10.3322/caac.20006]
- 2 **Thayer SP**, di Magliano MP, Heiser PW, Nielsen CM, Roberts DJ, Lauwers GY, Qi YP, Gysin S, Fernández-del Castillo C, Yajnik V, Antoniu B, McMahon M, Warshaw AL, Hebrok M. Hedgehog is an early and late mediator of pancreatic cancer tumorigenesis. *Nature* 2003; **425**: 851-856 [PMID: 14520413 DOI: 10.1038/nature02009]
- 3 **Ingham PW**, McMahon AP. Hedgehog signaling in animal development: paradigms and principles. *Genes Dev* 2001; **15**: 3059-3087 [PMID: 11731473 DOI: 10.1101/gad.938601]
- 4 **McMahon AP**, Ingham PW, Tabin CJ. Developmental roles and clinical significance of hedgehog signaling. *Curr Top Dev Biol* 2003; **53**: 1-114 [PMID: 12509125 DOI: 10.1016/S0070-2153(03)53002-2]
- 5 **Hooper JE**, Scott MP. Communicating with Hedgehogs. *Nat Rev Mol Cell Biol* 2005; **6**: 306-317 [PMID: 15803137 DOI: 10.1038/nrm1622]
- 6 **Mukherjee S**, Frolova N, Sadlonova A, Novak Z, Steg A, Page GP, Welch DR, Lobo-Ruppert SM, Ruppert JM, Johnson MR, Frost AR. Hedgehog signaling and response to cyclopamine differ in epithelial and stromal cells in benign breast and breast cancer. *Cancer Biol Ther* 2006; **5**: 674-683 [PMID: 16855373 DOI: 10.4161/cbt.5.6.2906]
- 7 **Cohen MM**. The hedgehog signaling network. *Am J Med Genet A* 2003; **123A**: 5-28 [PMID: 14556242]
- 8 **Onishi H**, Katano M. Hedgehog signaling pathway as a therapeutic target in various types of cancer. *Cancer Sci* 2011; **102**: 1756-1760 [PMID: 21679342 DOI: 10.1111/j.1349-7006.2011.02010.x]
- 9 **Jiang J**, Hui CC. Hedgehog signaling in development and cancer. *Dev Cell* 2008; **15**: 801-812 [PMID: 19081070 DOI: 10.1016/j.devcel]
- 10 **Bai CB**, Stephen D, Joyner AL. All mouse ventral spinal cord patterning by hedgehog is Gli dependent and involves an activator function of Gli3. *Dev Cell* 2004; **6**: 103-115 [PMID: 14723851 DOI: 10.1016/S1534-5807(03)00394-0]
- 11 **Kasper M**, Regl G, Frischauf AM, Aberger F. GLI transcription factors: mediators of oncogenic Hedgehog signaling. *Eur J Cancer* 2006; **42**: 437-445 [PMID: 16406505 DOI: 10.1016/j.ejca.2005.08.039]
- 12 **Stecca B**, Ruiz I, Altaba A. Context-dependent regulation of the GLI code in cancer by HEDGEHOG and non-HEDGEHOG signals. *J Mol Cell Biol* 2010; **2**: 84-95 [PMID: 20083481 DOI: 10.1093/jmcb/mjp052]
- 13 **Wang B**, Fallon JF, Beachy PA. Hedgehog-regulated processing of Gli3 produces an anterior/posterior repressor gradient in the developing vertebrate limb. *Cell* 2000; **100**: 423-434 [PMID: 10693759 DOI: 10.1016/S0092-8674(00)80678-9]
- 14 **Pan Y**, Bai CB, Joyner AL, Wang B. Sonic hedgehog signaling regulates Gli2 transcriptional activity by suppressing its processing and degradation. *Mol Cell Biol* 2006; **26**: 3365-3377 [PMID: 16611981 DOI: 10.1128/MCB.26.9.3365-3377.2006]
- 15 **Corbit KC**, Aanstad P, Singla V, Norman AR, Stainier DY, Reiter JF. Vertebrate Smoothed functions at the primary cilium. *Nature* 2005; **437**: 1018-1021 [PMID: 16136078 DOI: 10.1038/nature04117]
- 16 **Freeman M**. Feedback control of intercellular signalling in development. *Nature* 2000; **408**: 313-319 [PMID: 11099031 DOI: 10.1038/35042500]
- 17 **Pasca di Magliano M**, Hebrok M. Hedgehog signalling in cancer formation and maintenance. *Nat Rev Cancer* 2003; **3**: 903-911 [PMID: 14737121 DOI: 10.1038/nrc1229]
- 18 **Gill PS**, Rosenblum ND. Control of murine kidney development by sonic hedgehog and its GLI effectors. *Cell Cycle* 2006; **5**: 1426-1430 [PMID: 16855389 DOI: 10.4161/cc.5.13.2928]
- 19 **Kinzler KW**, Bigner SH, Bigner DD, Trent JM, Law ML, O'Brien SJ, Wong AJ, Vogelstein B. Identification of an amplified, highly expressed gene in a human glioma. *Science* 1987; **236**: 70-73 [PMID: 3563490 DOI: 10.1126/science.3563490]
- 20 **Gailani MR**, Bale AE. Developmental genes and cancer: role of patched in basal cell carcinoma of the skin. *J Natl Cancer Inst* 1997; **89**: 1103-1109 [PMID: 9262247 DOI: 10.1093/jnci/89.15.1103]
- 21 **Xie J**, Murone M, Luoh SM, Ryan A, Gu Q, Zhang C, Bonifas JM, Lam CW, Hynes M, Goddard A, Rosenthal A, Epstein EH, de Sauvage FJ. Activating Smoothed mutations in sporadic basal-cell carcinoma. *Nature* 1998; **391**: 90-92 [PMID: 9422511 DOI: 10.1038/34201]
- 22 **Berman DM**, Karhadkar SS, Maitra A, Montes De Oca R, Gerstenblith MR, Briggs K, Parker AR, Shimada Y, Eshleman JR, Watkins DN, Beachy PA. Widespread requirement for Hedgehog ligand stimulation in growth of digestive tract tumours. *Nature* 2003; **425**: 846-851 [PMID: 14520411 DOI: 10.1038/nature01972]
- 23 **Garcea G**, Dennison AR, Steward WP, Berry DP. Role of inflammation in pancreatic carcinogenesis and the implications for future therapy. *Pancreatol* 2005; **5**: 514-529 [PMID: 16110250 DOI: 10.1159/000087493]
- 24 **Farrow B**, Evers BM. Inflammation and the development

- of pancreatic cancer. *Surg Oncol* 2002; **10**: 153-169 [PMID: 12020670 DOI: 10.1016/S0960-7404(02)00015-4]
- 25 **Jura N**, Archer H, Bar-Sagi D. Chronic pancreatitis, pancreatic adenocarcinoma and the black box in-between. *Cell Res* 2005; **15**: 72-77 [PMID: 15686632 DOI: 10.1038/sj.cr.7290269]
- 26 **Luo JL**, Maeda S, Hsu LC, Yagita H, Karin M. Inhibition of NF-kappaB in cancer cells converts inflammation-induced tumor growth mediated by TNFalpha to TRAIL-mediated tumor regression. *Cancer Cell* 2004; **6**: 297-305 [PMID: 15380520 DOI: 10.1016/j.ccr.2004.08.012]
- 27 **Greten FR**, Eckmann L, Greten TF, Park JM, Li ZW, Egan LJ, Kagnoff MF, Karin M. IKKbeta links inflammation and tumorigenesis in a mouse model of colitis-associated cancer. *Cell* 2004; **118**: 285-296 [PMID: 15294155 DOI: 10.1016/j.cell.2004.07.013]
- 28 **Karin M**, Greten FR. NF-kappaB: linking inflammation and immunity to cancer development and progression. *Nat Rev Immunol* 2005; **5**: 749-759 [PMID: 16175180 DOI: 10.1038/nri1703]
- 29 **Nakashima H**, Nakamura M, Yamaguchi H, Yamanaka N, Akiyoshi T, Koga K, Yamaguchi K, Tsuneyoshi M, Tanaka M, Katano M. Nuclear factor-kappaB contributes to hedgehog signaling pathway activation through sonic hedgehog induction in pancreatic cancer. *Cancer Res* 2006; **66**: 7041-7049 [PMID: 16849549]
- 30 **Kasperczyk H**, Baumann B, Debatin KM, Fulda S. Characterization of sonic hedgehog as a novel NF-kappaB target gene that promotes NF-kappaB-mediated apoptosis resistance and tumor growth in vivo. *FASEB J* 2009; **23**: 21-33 [PMID: 18772349 DOI: 10.1096/fj.08-111096]
- 31 **Yauch RL**, Gould SE, Scales SJ, Tang T, Tian H, Ahn CP, Marshall D, Fu L, Januario T, Kallop D, Nannini-Pepe M, Kotkow K, Marsters JC, Rubin LL, de Sauvage FJ. A paracrine requirement for hedgehog signalling in cancer. *Nature* 2008; **455**: 406-410 [PMID: 18754008 DOI: 10.1038/nature07275]
- 32 **Tian H**, Callahan CA, DuPree KJ, Darbonne WC, Ahn CP, Scales SJ, de Sauvage FJ. Hedgehog signaling is restricted to the stromal compartment during pancreatic carcinogenesis. *Proc Natl Acad Sci USA* 2009; **106**: 4254-4259 [PMID: 19246386 DOI: 10.1073/pnas.0813203106]
- 33 **Yamasaki A**, Kameda C, Xu R, Tanaka H, Tasaka T, Chikazawa N, Suzuki H, Morisaki T, Kubo M, Onishi H, Tanaka M, Katano M. Nuclear factor kappaB-activated monocytes contribute to pancreatic cancer progression through the production of Shh. *Cancer Immunol Immunother* 2010; **59**: 675-686 [PMID: 19862523 DOI: 10.1007/s00262-009-0783-7]
- 34 **Hwang RE**, Moore TT, Hattersley MM, Scarpitti M, Yang B, Devereaux E, Ramachandran V, Arumugam T, Ji B, Logsdon CD, Brown JL, Godin R. Inhibition of the hedgehog pathway targets the tumor-associated stroma in pancreatic cancer. *Mol Cancer Res* 2012; **10**: 1147-1157 [PMID: 22859707]
- 35 **Lonardo E**, Frias-Aldeguer J, Hermann PC, Heeschen C. Pancreatic stellate cells form a niche for cancer stem cells and promote their self-renewal and invasiveness. *Cell Cycle* 2012; **11**: 1282-1290 [PMID: 22421149 DOI: 10.4161/cc.19679]
- 36 **Singh AP**, Arora S, Bhardwaj A, Srivastava SK, Kadakia MP, Wang B, Grizzle WE, Owen LB, Singh S. CXCL12/CXCR4 protein signaling axis induces sonic hedgehog expression in pancreatic cancer cells via extracellular regulated kinase- and Akt kinase-mediated activation of nuclear factor kappaB: implications for bidirectional tumor-stromal interactions. *J Biol Chem* 2012; **287**: 39115-39124 [PMID: 22995914 DOI: 10.1074/jbc.M112.409581]
- 37 **Onishi H**, Morifuji Y, Kai M, Suyama K, Iwasaki H, Katano M. Hedgehog inhibitor decreases chemosensitivity to 5-fluorouracil and gemcitabine under hypoxic conditions in pancreatic cancer. *Cancer Sci* 2012; **103**: 1272-1279 [PMID: 22486854 DOI: 10.1111/j.1349-7006.2012.02297.x]
- 38 **Onishi H**, Kai M, Odate S, Iwasaki H, Morifuji Y, Ogino T, Morisaki T, Nakashima Y, Katano M. Hypoxia activates the hedgehog signaling pathway in a ligand-independent manner by upregulation of Smo transcription in pancreatic cancer. *Cancer Sci* 2011; **102**: 1144-1150 [PMID: 21338440 DOI: 10.1111/j.1349-7006.2011.01912.x]
- 39 **Onishi H**, Morisaki T, Nakao F, Odate S, Morisaki T, Katano M. Protein-bound polysaccharide decreases invasiveness and proliferation in pancreatic cancer by inhibition of hedgehog signaling and HIF-1alpha pathways under hypoxia. *Cancer Lett* 2013; **335**: 289-298 [PMID: 23485726 DOI: 10.1016/j.canlet.2013.02.041]
- 40 **Al-Hajj M**, Wicha MS, Benito-Hernandez A, Morrison SJ, Clarke MF. Prospective identification of tumorigenic breast cancer cells. *Proc Natl Acad Sci USA* 2003; **100**: 3983-3988 [PMID: 12629218]
- 41 **Li C**, Heidt DG, Dalerba P, Burant CF, Zhang L, Adsay V, Wicha M, Clarke MF, Simeone DM. Identification of pancreatic cancer stem cells. *Cancer Res* 2007; **67**: 1030-1037 [PMID: 17283135 DOI: 10.1158/0008-5472.CAN-06-2030]
- 42 **Hermann PC**, Huber SL, Herrler T, Aicher A, Ellwart JW, Guba M, Bruns CJ, Heeschen C. Distinct populations of cancer stem cells determine tumor growth and metastatic activity in human pancreatic cancer. *Cell Stem Cell* 2007; **1**: 313-323 [PMID: 18371365 DOI: 10.1016/j.stem.2007.06.002]
- 43 **Bar EE**, Chaudhry A, Lin A, Fan X, Schreck K, Matsui W, Piccirillo S, Vescovi AL, DiMeco F, Olivi A, Eberhart CG. Cyclopamine-mediated hedgehog pathway inhibition depletes stem-like cancer cells in glioblastoma. *Stem Cells* 2007; **25**: 2524-2533 [PMID: 17628016 DOI: 10.1634/stemcells.2007-0166]
- 44 **Mueller MT**, Hermann PC, Witthauer J, Rubio-Viqueira B, Leicht SF, Huber S, Ellwart JW, Mustafa M, Bartenstein P, D'Haese JG, Schoenberg MH, Berger F, Jauch KW, Hidalgo M, Heeschen C. Combined targeted treatment to eliminate tumorigenic cancer stem cells in human pancreatic cancer. *Gastroenterology* 2009; **137**: 1102-1113 [PMID: 19501590 DOI: 10.1053/j.gastro.2009.05.053]
- 45 **Di J**, Duiveman-de Boer T, Figdor CG, Torensma R. Eradicating cancer cells: struggle with a chameleon. *Oncotarget* 2011; **2**: 99-101 [PMID: 21378413]
- 46 **Lee CJ**, Dosch J, Simeone DM. Pancreatic cancer stem cells. *J Clin Oncol* 2008; **26**: 2806-2812 [PMID: 18539958 DOI: 10.1200/JCO.2008.16.6702]
- 47 **Huang FT**, Zhuan-Sun YX, Zhuang YY, Wei SL, Tang J, Chen WB, Zhang SN. Inhibition of hedgehog signaling depresses self-renewal of pancreatic cancer stem cells and reverses chemoresistance. *Int J Oncol* 2012; **41**: 1707-1714 [PMID: 22923052 DOI: 10.3892/ijo.2012.1597]
- 48 **Tang SN**, Fu J, Nall D, Rodova M, Shankar S, Srivastava RK. Inhibition of sonic hedgehog pathway and pluripotency maintaining factors regulate human pancreatic cancer stem cell characteristics. *Int J Cancer* 2012; **131**: 30-40 [PMID: 21796625 DOI: 10.1002/ijc.26323]
- 49 **Li SH**, Fu J, Watkins DN, Srivastava RK, Shankar S. Sulforaphane regulates self-renewal of pancreatic cancer stem cells through the modulation of Sonic hedgehog-GLI pathway. *Mol Cell Biochem* 2013; **373**: 217-227 [PMID: 23129257 DOI: 10.1007/s11010-012-1493-6]
- 50 **Rodova M**, Fu J, Watkins DN, Srivastava RK, Shankar S. Sonic hedgehog signaling inhibition provides opportunities for targeted therapy by sulforaphane in regulating pancreatic cancer stem cell self-renewal. *PLoS One* 2012; **7**: e46083 [PMID: 23029396 DOI: 10.1371/journal.pone.0046083]
- 51 **Han JB**, Sang F, Chang JJ, Hua YQ, Shi WD, Tang LH, Liu LM. Arsenic trioxide inhibits viability of pancreatic cancer stem cells in culture and in a xenograft model via binding to SHH-Gli. *Onco Targets Ther* 2013; **6**: 1129-1138 [PMID: 23990729 DOI: 10.2147/OTT.S49148.]

- 52 **Kristiansen G**, Sammar M, Altevogt P. Tumour biological aspects of CD24, a mucin-like adhesion molecule. *J Mol Histol* 2004; **35**: 255-262 [PMID: 15339045]
- 53 **Aigner S**, Sthoeger ZM, Fogel M, Weber E, Zarn J, Ruppert M, Zeller Y, Vestweber D, Stahel R, Sammar M, Altevogt P. CD24, a mucin-type glycoprotein, is a ligand for P-selectin on human tumor cells. *Blood* 1997; **89**: 3385-3395 [PMID: 9129046]
- 54 **Cao X**, Geradts J, Dewhirst MW, Lo HW. Upregulation of VEGF-A and CD24 gene expression by the tGLI1 transcription factor contributes to the aggressive behavior of breast cancer cells. *Oncogene* 2012; **31**: 104-115 [PMID: 21666711 DOI: 10.1038/onc.2011.219]
- 55 **Ringel J**, Jesnowski R, Schmidt C, Ringel J, Köhler HJ, Rychly J, Batra SK, Löhr M. CD44 in normal human pancreas and pancreatic carcinoma cell lines. *Teratog Carcinog Mutagen* 2001; **21**: 97-106 [PMID: 11135324]
- 56 **Koong AC**, Mehta VK, Le QT, Fisher GA, Terris DJ, Brown JM, Bastidas AJ, Vierra M. Pancreatic tumors show high levels of hypoxia. *Int J Radiat Oncol Biol Phys* 2000; **48**: 919-922 [PMID: 11072146]
- 57 **Caldwell CC**, Kojima H, Lukashev D, Armstrong J, Farber M, Apasov SG, Sitkovsky MV. Differential effects of physiologically relevant hypoxic conditions on T lymphocyte development and effector functions. *J Immunol* 2001; **167**: 6140-6149 [PMID: 11714773]
- 58 **Höckel S**, Schlenger K, Vaupel P, Höckel M. Association between host tissue vascularity and the prognostically relevant tumor vascularity in human cervical cancer. *Int J Oncol* 2001; **19**: 827-832 [PMID: 11562762]
- 59 **Lei J**, Ma J, Ma Q, Li X, Liu H, Xu Q, Duan W, Sun Q, Xu J, Wu Z, Wu E. Hedgehog signaling regulates hypoxia induced epithelial to mesenchymal transition and invasion in pancreatic cancer cells via a ligand-independent manner. *Mol Cancer* 2013; **12**: 66 [PMID: 23786654 DOI: 10.1186/1476-4598-12-66]
- 60 **Spivak-Kroizman TR**, Hostetter G, Posner R, Aziz M, Hu C, Demeure MJ, Von Hoff D, Hingorani SR, Palculict TB, Izzo J, Kiriakova GM, Abdelmelek M, Bartholomeusz G, James BP, Powis G. Hypoxia triggers hedgehog-mediated tumor-stromal interactions in pancreatic cancer. *Cancer Res* 2013; **73**: 3235-3247 [PMID: 23633488 DOI: 10.1158/0008-5472.CAN-11-1433]
- 61 **Bahra M**, Kamphues C, Boas-Knoop S, Lippert S, Esendik U, Schüller U, Hartmann W, Waha A, Neuhaus P, Heppner F, Pietsch T, Koch A. Combination of hedgehog signaling blockage and chemotherapy leads to tumor reduction in pancreatic adenocarcinomas. *Pancreas* 2012; **41**: 222-229 [PMID: 22076568 DOI: 10.1097/MPA.0b013e31822896dd]
- 62 **Qin CF**, Hao K, Tian XD, Xie XH, Yang YM. Combined effects of EGFR and Hedgehog signaling pathway inhibition on the proliferation and apoptosis of pancreatic cancer cells. *Oncol Rep* 2012; **28**: 519-526 [PMID: 22581058 DOI: 10.3892/or.2012.1808]
- 63 **Eberl M**, Klingler S, Mangelberger D, Loipetzberger A, Damhofer H, Zoidl K, Schnidar H, Hache H, Bauer HC, Solca F, Hauser-Kronberger C, Ermilov AN, Verhaegen ME, Bichakjian CK, Dlugosz AA, Nietfeld W, Sibilia M, Lehrach H, Wierling C, Aberger F. Hedgehog-EGFR cooperation response genes determine the oncogenic phenotype of basal cell carcinoma and tumour-initiating pancreatic cancer cells. *EMBO Mol Med* 2012; **4**: 218-233 [PMID: 22294553 DOI: 10.1002/emmm.201100201]
- 64 **Chitkara D**, Singh S, Kumar V, Danquah M, Behrman SW, Kumar N, Mahato RI. Micellar delivery of cyclopamine and gefitinib for treating pancreatic cancer. *Mol Pharm* 2012; **9**: 2350-2357 [PMID: 22780906]
- 65 **Gu D**, Liu H, Su GH, Zhang X, Chin-Sinex H, Hanenberg H, Mendonca MS, Shannon HE, Chiorean EG, Xie J. Combining hedgehog signaling inhibition with focal irradiation on reduction of pancreatic cancer metastasis. *Mol Cancer Ther* 2013; **12**: 1038-1048 [PMID: 23468532 DOI: 10.1158/1535-7163.MCT-12-1030]
- 66 **Onishi H**, Morisaki T, Kiyota A, Koya N, Tanaka H, Umebayashi M, Katano M. The Hedgehog inhibitor cyclopamine impairs the benefits of immunotherapy with activated T and NK lymphocytes derived from patients with advanced cancer. *Cancer Immunol Immunother* 2013; **62**: 1029-1039 [PMID: 23591983 DOI: 10.1007/s00262-013-1419-5]
- 67 **Onishi H**, Morisaki T, Kiyota A, Koya N, Tanaka H, Umebayashi M, Katano M. The Hedgehog inhibitor suppresses the function of monocyte-derived dendritic cells from patients with advanced cancer under hypoxia. *Biochem Biophys Res Commun* 2013; **436**: 53-59 [PMID: 23707943 DOI: 10.1016/j.bbrc.2013.05.057]
- 68 **Sekulic A**, Migden MR, Oro AE, Dirix L, Lewis KD, Hainsworth JD, Solomon JA, Yoo S, Arron ST, Friedlander PA, Marmur E, Rudin CM, Chang AL, Low JA, Mackey HM, Yauch RL, Graham RA, Reddy JC, Hauschild A. Efficacy and safety of vismodegib in advanced basal-cell carcinoma. *N Engl J Med* 2012; **366**: 2171-2179 [PMID: 22670903 DOI: 10.1056/NEJMoa1113713]
- 69 **Kelleher FC**. Hedgehog signaling and therapeutics in pancreatic cancer. *Carcinogenesis* 2011; **32**: 445-451 [PMID: 21186299 DOI: 10.1093/carcin/bgq280]
- 70 **Sandhiya S**, Melvin G, Kumar SS, Dkhar SA. The dawn of hedgehog inhibitors: Vismodegib. *J Pharmacol Pharmacother* 2013; **4**: 4-7 [PMID: 23662017 DOI: 10.4103/0976-500X.107628]
- 71 **Fendrich V**, Wiese D, Waldmann J, Lauth M, Heverhagen AE, Rehm J, Bartsch DK. Hedgehog inhibition with the orally bioavailable Smo antagonist LDE225 represses tumor growth and prolongs survival in a transgenic mouse model of islet cell neoplasms. *Ann Surg* 2011; **254**: 818-823; discussion 823 [PMID: 22042473 DOI: 10.1097/SLA.0b013e318236bc0f]
- 72 **Fu J**, Rodova M, Roy SK, Sharma J, Singh KP, Srivastava RK, Shankar S. GANT-61 inhibits pancreatic cancer stem cell growth in vitro and in NOD/SCID/IL2R gamma null mice xenograft. *Cancer Lett* 2013; **330**: 22-32 [PMID: 23200667 DOI: 10.1016/j.canlet.2012.11.018]
- 73 **Mo W**, Xu X, Xu L, Wang F, Ke A, Wang X, Guo C. Resveratrol inhibits proliferation and induces apoptosis through the hedgehog signaling pathway in pancreatic cancer cell. *Pancreatol* 2011; **11**: 601-609 [PMID: 22301921 DOI: 10.1159/000333542]
- 74 **Sun XD**, Liu XE, Huang DS. Curcumin reverses the epithelial-mesenchymal transition of pancreatic cancer cells by inhibiting the Hedgehog signaling pathway. *Oncol Rep* 2013; **29**: 2401-2407 [PMID: 23563640 DOI: 10.3892/or.2013.2385]
- 75 **Bi X**, Han X, Zhang F, He M, Zhang Y, Zhi XY, Zhao H. Triparanol suppresses human tumor growth in vitro and in vivo. *Biochem Biophys Res Commun* 2012; **425**: 613-618 [PMID: 22877755 DOI: 10.1016/j.bbrc.2012.07.136]
- 76 **Nolan-Stevaux O**, Lau J, Truitt ML, Chu GC, Hebrok M, Fernández-Zapico ME, Hanahan D. GLI1 is regulated through Smoothed-independent mechanisms in neoplastic pancreatic ducts and mediates PDAC cell survival and transformation. *Genes Dev* 2009; **23**: 24-36 [PMID: 19136624 DOI: 10.1101/gad.1753809]
- 77 **An Y**, Cai B, Chen J, Lv N, Yao J, Xue X, Tu M, Tang D, Wei J, Jiang K, Wu J, Li Q, Gao W, Miao Y. MAP3K10 promotes the proliferation and decreases the sensitivity of pancreatic cancer cells to gemcitabine by upregulating Gli-1 and Gli-2. *Cancer Lett* 2013; **329**: 228-235 [PMID: 23178452 DOI: 10.1016/j.canlet.2012.11.005]
- 78 **Li X**, Ma Q, Xu Q, Liu H, Lei J, Duan W, Bhat K, Wang F, Wu E, Wang Z. SDF-1/CXCR4 signaling induces pancreatic cancer cell invasion and epithelial-mesenchymal transition

- in vitro through non-canonical activation of Hedgehog pathway. *Cancer Lett* 2012; **322**: 169-176 [PMID: 22450749 DOI: 10.1016/j.canlet.2012.02.035]
- 79 **Low JA**, de Sauvage FJ. Clinical experience with Hedgehog pathway inhibitors. *J Clin Oncol* 2010; **28**: 5321-5326 [PMID: 21041712 DOI: 10.1200/JCO.2010.27.9943]
- 80 **Merchant AA**, Matsui W. Targeting Hedgehog--a cancer stem cell pathway. *Clin Cancer Res* 2010; **16**: 3130-3140 [PMID: 20530699 DOI: 10.1158/1078-0432.CCR-09-2846]
- 81 **Sahebjam S**, Siu LL, Razak AA. The utility of hedgehog signaling pathway inhibition for cancer. *Oncologist* 2012; **17**: 1090-1099 [PMID: 22851551 DOI: 10.1634/theoncologist.2011-0450]
- 82 **Atwood SX**, Chang AL, Oro AE. Hedgehog pathway inhibition and the race against tumor evolution. *J Cell Biol* 2012; **199**: 193-197 [PMID: 23071148 DOI: 10.1083/jcb.201207140]

P- Reviewers: Ceyhan C, Fan Y **S- Editor:** Wen LL
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ISSN 1007-9327



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