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**Neonatal cholestasis can be the first symptom of McCune–Albright syndrome: A case report**

Satomura Y *et al*. McCune–Albright syndrome with neonatal cholestasis

Yoshinori Satomura, Kazuhiko Bessho, Taichi Kitaoka, Shinji Takeyari, Yasuhisa Ohata, Takuo Kubota, Keiichi Ozono

**Yoshinori Satomura, Kazuhiko Bessho, Taichi Kitaoka, Shinji Takeyari, Yasuhisa Ohata, Takuo Kubota, Keiichi Ozono,** Department of Pediatrics, Osaka University Graduate School of Medicine, Osaka 565-0871, Japan

**Author contributions:** Satomura Y designed and wrote the manuscript; Bessho K designed and edited the manuscript; Kitaoka T collected the patient’s clinical data; Takeyari S and Ohata Y extracted genomic DNA and performed genetic studies; Kubota T and Ozono K supervised and edited the manuscript; all authors issued final approval for the version to be submitted.

**Corresponding author: Kazuhiko Bessho, MD, PhD, Associate Professor,** Department of Pediatrics, Osaka University Graduate School of Medicine, 2-2-D5 Yamada-oka, Suita City, Osaka 565-0871, Japan. bessho@ped.med.osaka-u.ac.jp

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**Abstract**

BACKGROUND

McCune–Albright syndrome (MAS) is caused by postzygotic somatic mutations of the *GNAS* gene. It is characterized by the clinical triad of fibrous dysplasia, café-au-lait skin spots, and endocrinological dysfunction. Myriad complications in MAS, including hepatobiliary manifestations, are also reported.

CASE SUMMARY

This is a case of a 4-year-old boy who presented with MAS with neonatal cholestasis. He was suspected to have Alagille syndrome due to neonatal cholestasis with intrahepatic bile duct paucity in liver biopsy, peripheral pulmonary artery stenosis, and renal tubular dysfunction. By the age of 2 years, his cholestatic liver injury gradually improved, but he had repeated left femoral fractures. He did not exhibit endocrinological abnormality or café-au-lait skin spots. However, MAS was suspected due to fibrous dysplasia at the age of 4 years. No mutation was identified in the *GNAS* gene in the DNA isolated from the peripheral blood, but an activating point mutation (c.601C>T, p.Arg201Cys) was observed in the DNA extracted from the affected bone tissue and that extracted from the formalin-fixed paraffin-embedded liver tissue, which was obtained at the age of 1 mo.

CONCLUSION

MAS should be considered as a differential diagnosis for transient cholestasis in infancy.

**Key Words:** McCune–Albright syndrome; *GNAS* gene; Neonatal cholestasis; Alagille syndrome; Bile duct paucity; Case report

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**Core Tip:** McCune–Albright syndrome (MAS) is caused by postzygotic somatic mutations of the *GNAS* gene. It is characterized by the clinical triad of fibrous dysplasia, café-au-lait skin spots, and endocrinological dysfunction. MAS complications other than the triad are also reported. This is the case of a boy with MAS diagnosed with Alagille syndrome in his infancy based on intrahepatic bile duct paucity in liver biopsy, neonatal cholestasis, cardiac manifestation, and renal tubular dysfunction. MAS should be considered as a differential diagnosis for transient cholestasis in infancy.

**INTRODUCTION**

McCune–Albright syndrome (MAS) is a rare sporadic disease characterized by the clinical triad of fibrous dysplasia, café-au-lait skin spots, and endocrinological dysfunction[1,2]. Its estimated prevalence ranges from 1/100000 to 1/1000000[3]. MAS is caused by postzygotic somatic mutations of the *GNAS* gene, which encodes the G protein stimulatory α subunit[4]. MAS complications other than the clinical triad, including hepatobiliary dysfunction, are reported[4-6].

Alagille syndrome (ALGS) is an autosomal dominant disorder with a wide spectrum of clinical variability. The main clinical features and malformations are chronic cholestasis due to intrahepatic bile duct paucity (decreased bile duct-to-portal tract ratio: < 0.4), cardiac disease (particularly peripheral pulmonary artery stenosis), skeletal deformity (particularly butterfly vertebrae), ocular abnormalities (particularly posterior embryotoxon), and characteristic facial features. Additional features include intracranial bleeding, dysplastic kidneys, and bone fractures[7,8]. The majority of cases are caused by *JAG1* gene haploinsufficiency, encoding a ligand jagged1 in the Notch signaling pathway[9,10]. Mutations in *NOTCH2*, a receptor in the same signaling pathway, are identified in some ALGS patients who do not have mutations in *JAG1*[11].

This is a case of a boy who was diagnosed with ALGS in his infancy based on intrahepatic bile duct paucity in liver biopsy, peripheral pulmonary artery stenosis, and renal tubular dysfunction and later with MAS based on radiographic findings of fibrous dysplasia.

**CASE PRESENTATION**

***Chief complaints***

A 4-year-old boy complained of repeated left femoral fractures.

***History of present illness***

The patient had repeated left femoral fractures for four times (at 1 year and 3 mo, 1 year and 11 mo, 2 years and 10 mo, and 4 years and 3 mo old), and the difference in the length of his lower limbs gradually became apparent by the age of 2 years. While repeated femoral fractures were initially considered as bone metabolic disorders associated with ALGS, the serum phosphate levels had remained at the lower limit of the standard for age, and the level of fibroblast growth factor 23 (FGF23) was high as 117 pg/mL (reference range: 15-49 pg/mL[12]). At the age of 4 years and 8 mo, radiographic findings revealed a “ground-glass” appearance in his left femur and tibia and “shepherd’s crook deformity” in his left thigh bone, which were characteristic features of fibrous dysplasia (Figure 1).

***History of past illness***

The patient was born at 40 wk and 6 d’ gestation; with a birth weight of 2726 g. Failure to thrive was noted at 18 d following birth. Further evaluation of this concern revealed hepatomegaly, elevated liver transaminase level [aspartate aminotransferase (AST) 193 U/L, alanine aminotransferase (ALT) 424 U/L], and hyperbilirubinemia (T-Bil 8.0 mg/dL, D-Bil 6.6 mg/dL). Liver biopsy was performed at the age of 1 mo, which revealed bile duct paucity (the ratio of the bile duct to the portal tract was 0.1) (Figure 2). Other than cholestasis, peripheral pulmonary artery stenosis, hypokalemia, and metabolic acidosis due to renal tubular dysfunction were observed. No butterfly vertebrae or ocular abnormalities were found. Although any large deletion and duplication were not observed in the *JAG1* gene by the fluorescence *in situ* hybridization analysis, the patient was clinically suspected to have ALGS and was listed for liver transplantation. Cholestatic liver injury was gradually normalized by the age of 2 years under oral ursodeoxycholic acid and glycyrrhizic acid treatment and did not deteriorate even after both medications were tapered. His DNA was further subjected to a targeted next-generation sequencing that covers 14 genes responsible for cholestatic liver diseases[13], and no pathogenic variants were found in his genes including *JAG1* and *NOTCH2.*

***Personal and family history***

The patient was born to non-consanguineous Japanese parents. The pregnancy had been uncomplicated, and his family history was unremarkable.

***Physical examination***

At the age of 4 years and 9 mo, his height was 101.7cm ( -0.81 SD); body weight, 15.2kg ( -0.82 SD); and arm span, 104 cm. The difference in the length of the lower limbs was 1 cm (right, 53 cm; left, 52 cm). He did not exhibit jaundice or hepatosplenomegaly. He was noted to have a grade 2/6 systolic heart murmur. He did not have café-au-lait skin spots. His testicular capacity was 2 mL, pubic hair had not yet grown, and no precocious puberty was observed.

***Laboratory examinations***

Laboratory examination at the age of 4 years revealed elevated levels of serum alkaline phosphatase (2506 U/L, reference range: 430-1200 U/L), bone alkaline phosphatase (216 U/L, reference range: 59-107 U/L[14]), FGF23 (86 pg/mL), and serum type I collagen cross-linked N-telopeptide (171 nmolBCE/L, reference range: 14-57 nmolBCE/L[15]). No endocrinological abnormalities were found. The transaminase and bilirubin levels were within the reference ranges (AST 28 U/L, ALT 25 U/L, T-Bil 0.6 mg/dL, and D-Bil 0.2 mg/dL).

***Imaging examinations***

Bone scintigraphy with 99 mTc-hydroxymethylene diphosphonate, which was employed to detect lesions with enhanced bone metabolism, revealed multiple lesions with increased uptake in the left skull and upper left limb in addition to the left femur and left tibia (Figure 3).

***Further diagnostic work-up***

For the mutational analysis of the *GNAS* gene, genomic DNA from the peripheral blood was extracted using magLEAD Consumable Kit® (Precision System Science Co., Ltd., Chiba, Japan). In addition, it was polymerase chain reaction (PCR)-amplified for exons 7 to 10 and their splice sites of the *GNAS* gene, where mutation hotspots for MAS were reported. PCRs were conducted using the 5′-TCACTTCCGTTGAGCCTGAC-3′ and 5′-CTTGCACGGGGTTCTTCTCT-3′ primer set designed for detecting the mutation; however, sequencing after PCR did not reveal any mutations (Figure 4A).

Therefore, mutation analysis of the *GNAS* gene was also conducted from bone tissue samples, which were obtained from fibrous dysplastic lesions during a fracture surgery at the age of 5 years and 6 mo. The dissected bone sample was immediately snap-frozen using liquid nitrogen and crushed using 6700 Freezer/Mill (SPEX SamplePrep, NJ, United States). Genomic DNA from the bone tissue was extracted using DNeasy Blood & Tissue Kit (QIAGEN, Hilden, Germany) and was PCR-amplified and sequenced similar to that of the peripheral blood. As a result, an activation point mutation (c.601C>T, p.Arg201Cys)[4] was detected in genomic DNA, and the patient was diagnosed with MAS (Figure 4B).

Furthermore, when he was 6 years old, DNA was extracted from a formalin-fixed paraffin-embedded (FFPE) liver tissue that was collected during the biopsy performed at the age of 1 mo. To isolate genomic DNA from the FFPE liver tissue, Agencourt FormaPure XL Total kit (Agencourt Bioscience Corporation, Beverly, MA, United States) was used. Genomic DNA from the liver tissue was PCR-amplified and sequenced for the corresponding site to the peripheral blood and bone tissue. PCRs and sequencing were conducted using the 5′-TTCGGTTGGCTTTGGTGAGA-3′ and 5′-CACGTCAAACATGCTGGTGG-3′ primer set designed for detecting the mutation. The same mutation from the bone tissue samples was observed (Figure 4C).

**FINAL DIAGNOSIS**

The final diagnosis of the presented case is MAS.

**TREATMENT**

When he was 7 years old, an osteotomy was performed to correct the curvature of the left femur.

**OUTCOME AND FOLLOW-UP**

The patient was followed up for endocrine abnormalities, such as premature puberty and compression optic neuropathy, since bone scintigraphy revealed increased uptake in his skull. Furthermore, although his liver dysfunction did not persist, follow-up was continued with semiannual to annual abdominal ultrasonography for neoplasm in the liver.

**DISCUSSION**

MAS is caused by activating somatic mutations within the *GNAS* gene. These mutations occur in the early postzygotic period. The patient’s somatic cells are mosaic for the mutation; hence, the clinical features are determined by the distribution of the affected cells[4,16,17].

In MAS, hepatobiliary dysfunction is relatively rare, with a frequency of 5%-10%[18,19], and usually develops in the early stage of life as neonatal cholestasis[5,6,16,20,21]. Although cholestasis can be the first symptom of MAS and is sometimes followed by persistent elevation of the levels of serum liver enzymes, natural history has been reported as benign in most patients[5,6], and only a few cases required liver transplantation[20].

The histological findings of the patient in this report revealed intrahepatic bile duct paucity, which suggested ALGS along with characteristic features, such as neonatal cholestasis, peripheral pulmonary artery stenosis, renal tubular dysfunction, and recurrent bone fractures. Giant cell transformation has been the most common finding in the liver histology of MAS[5,22,23]. However, intrahepatic bile duct paucity was also reported in cases with MAS. In such cases, distinguishing MAS from ALGS based on clinical symptoms and pathological features is difficult as in our case, in which the difference in the length of the patient’s legs prompted us to suspect enhanced bone metabolism[6]. MAS should be considered among the differential diagnoses of ALGS when the liver tissue demonstrates intrahepatic bile duct paucity. A recent manuscript reported that combined sequencing of *JAG1* and *NOTCH2* along with copy number variant analysis of *JAG1* did not identify pathogenic variants in 3.2% of patients who met the diagnostic criteria for ALGS[24]. Regarding renal tubular dysfunction and peripheral pulmonary artery stenosis in our case, we did not extract and sequence genomic DNA from renal tubular epithelial cells and pulmonary artery to detect the mutation in the tissues. Although our patient did not meet the classical diagnostic criteria of AGLS which is based on the presence of intrahepatic bile duct paucity on liver biopsy in association with at least three of the major clinical features: chronic cholestasis, cardiac disease, skeletal abnormalities, ocular abnormalities, and characteristic facial features, it is still possible that some other genes than *GNAS* or mutations in *JAG1/NOTCH2* genes that cannot be detected with current methods are involved in AGLS-like renal and pulmonary features in our case.

Due to the somatic mosaic nature of the disease, a negative result of mutation analysis from the peripheral blood does not exclude the possibility of MAS[3,19], and DNA should be isolated from the affected tissues. In this case, *GNAS* gene mutation was detected from the surgical bone specimen and FFPE liver biopsy tissue, which was collected 6 years ago. As in this report, *GNAS* mutations have been detected in the liver tissue obtained from patients with neonatal cholestasis in previous reports[5,16,19,20]. The occurrence and severity of the hepatic phenotype depend on the number and location of the cells with the mutation[5,16]. Whether the patients still keep hepatic cells with the mutation in the *GNAS* gene following amelioration of their hepatic symptoms is unknown.

In most cases, neonatal cholestasis in patients with MAS resolves spontaneously. However, liver dysfunction may persist, and subsequent hepatic lesions may develop and exhibit malignant potential, such as hepatoblastoma and hepatocellular adenomas[6,21]. In this case, liver dysfunction did not persist, and liver lesions were not identified, but we continued to follow-up the patient for serum tumor markers with semiannual to annual abdominal ultrasonography.

We presented a case of a patient with MAS who was suspected of ALGS due to neonatal cholestasis and histological findings that revealed intrahepatic bile duct paucity. No pathogenic variants were noted in the *JAG1* and *NOTCH2* genes, and MAS was suspected from repeated fractures and radiographic findings. The mutation in the *GNAS* gene was detected in the bone and liver tissues, and the patient was diagnosed with MAS. MAS should be considered as a differential diagnosis for cholestasis in infancy.

**CONCLUSION**

Hepatobiliary dysfunction is relatively rare in MAS, but MAS should be considered as a part of the differential diagnosis of neonatal cholestasis with unknown causes, and genetic diagnosis using liver tissue is possible.

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**REFERENCES**

1 **McCune DJ.** Osteitis fibrosa cystica: the case of a nine-year old girl who also exhibits precocious puberty, multiple pigmentation of the skin and hyperthyroidism*. Am J Dis Child* 1936; **52**: 743-744

2 **Albright F,** Butler AM, Hampton AO, Smith P. Syndrome characterized by osteitis fibrosa disseminata, areas of pigmentation and endocrine dysfunction, with precocious puberty in females. *N Engl J Med* 1937; **216**: 727-746 [DOI: 10.1056/NEJM193704292161701]

3 **Dumitrescu CE**, Collins MT. McCune-Albright syndrome. *Orphanet J Rare Dis* 2008; **3**: 12 [PMID: 18489744 DOI: 10.1186/1750-1172-3-12]

4 **Weinstein LS**, Shenker A, Gejman PV, Merino MJ, Friedman E, Spiegel AM. Activating mutations of the stimulatory G protein in the McCune-Albright syndrome. *N Engl J Med* 1991; **325**: 1688-1695 [PMID: 1944469 DOI: 10.1056/NEJM199112123252403]

5 **Silva ES**, Lumbroso S, Medina M, Gillerot Y, Sultan C, Sokal EM. Demonstration of McCune-Albright mutations in the liver of children with high gammaGT progressive cholestasis. *J Hepatol* 2000; **32**: 154-158 [PMID: 10673080 DOI: 10.1016/s0168-8278(00)80202-0]

6 **Johansen L**, Haller W, Thyagarajan M, Kelly D, McKiernan P. Hepatic Lesions Associated With McCune Albright Syndrome. *J Pediatr Gastroenterol Nutr* 2019; **68**: e54-e57 [PMID: 30628989 DOI: 10.1097/MPG.0000000000002266]

7 **Alagille D**, Estrada A, Hadchouel M, Gautier M, Odièvre M, Dommergues JP. Syndromic paucity of interlobular bile ducts (Alagille syndrome or arteriohepatic dysplasia): review of 80 cases. *J Pediatr* 1987; **110**: 195-200 [PMID: 3806290 DOI: 10.1016/S0022-3476(87)80153-1]

8 **Turnpenny PD**, Ellard S. Alagille syndrome: pathogenesis, diagnosis and management. *Eur J Hum Genet* 2012; **20**: 251-257 [PMID: 21934706 DOI: 10.1038/ejhg.2011.181]

9 **Li L**, Krantz ID, Deng Y, Genin A, Banta AB, Collins CC, Qi M, Trask BJ, Kuo WL, Cochran J, Costa T, Pierpont ME, Rand EB, Piccoli DA, Hood L, Spinner NB. Alagille syndrome is caused by mutations in human Jagged1, which encodes a ligand for Notch1. *Nat Genet* 1997; **16**: 243-251 [PMID: 9207788 DOI: 10.1038/ng0797-243]

10 **Oda T**, Elkahloun AG, Pike BL, Okajima K, Krantz ID, Genin A, Piccoli DA, Meltzer PS, Spinner NB, Collins FS, Chandrasekharappa SC. Mutations in the human Jagged1 gene are responsible for Alagille syndrome. *Nat Genet* 1997; **16**: 235-242 [PMID: 9207787 DOI: 10.1038/ng0797-235]

11 **McDaniell R**, Warthen DM, Sanchez-Lara PA, Pai A, Krantz ID, Piccoli DA, Spinner NB. NOTCH2 mutations cause Alagille syndrome, a heterogeneous disorder of the notch signaling pathway. *Am J Hum Genet* 2006; **79**: 169-173 [PMID: 16773578 DOI: 10.1086/505332]

12 **Bacchetta J**, Dubourg L, Harambat J, Ranchin B, Abou-Jaoude P, Arnaud S, Carlier MC, Richard M, Cochat P. The influence of glomerular filtration rate and age on fibroblast growth factor 23 serum levels in pediatric chronic kidney disease. *J Clin Endocrinol Metab* 2010; **95**: 1741-1748 [PMID: 20157196 DOI: 10.1210/jc.2009-1576]

13 **Togawa T**, Sugiura T, Ito K, Endo T, Aoyama K, Ohashi K, Negishi Y, Kudo T, Ito R, Kikuchi A, Arai-Ichinoi N, Kure S, Saitoh S. Molecular Genetic Dissection and Neonatal/Infantile Intrahepatic Cholestasis Using Targeted Next-Generation Sequencing. *J Pediatr* 2016; **171**: 171-7.e1-4 [PMID: 26858187 DOI: 10.1016/j.jpeds.2016.01.006]

14 **Yang L**, Grey V. Pediatric reference intervals for bone markers. *Clin Biochem* 2006; **39**: 561-568 [PMID: 16423337 DOI: 10.1016/j.clinbiochem.2005.11.015]

15 **van der Sluis IM**, Hop WC, van Leeuwen JP, Pols HA, de Muinck Keizer-Schrama SM. A cross-sectional study on biochemical parameters of bone turnover and vitamin d metabolites in healthy dutch children and young adults. *Horm Res* 2002; **57**: 170-179 [PMID: 12053089 DOI: 10.1159/000058378]

16 **Shenker A**, Weinstein LS, Moran A, Pescovitz OH, Charest NJ, Boney CM, Van Wyk JJ, Merino MJ, Feuillan PP, Spiegel AM. Severe endocrine and nonendocrine manifestations of the McCune-Albright syndrome associated with activating mutations of stimulatory G protein GS. *J Pediatr* 1993; **123**: 509-518 [PMID: 8410501 DOI: 10.1016/s0022-3476(05)80943-6]

17 **Völkl TM**, Dörr HG. McCune-Albright syndrome: clinical picture and natural history in children and adolescents. *J Pediatr Endocrinol Metab* 2006; **19 Suppl 2**: 551-559 [PMID: 16789617 DOI: 10.1515/jpem.2006.19.s2.551]

18 **Ringel MD**, Schwindinger WF, Levine MA. Clinical implications of genetic defects in G proteins. The molecular basis of McCune-Albright syndrome and Albright hereditary osteodystrophy. *Medicine (Baltimore)* 1996; **75**: 171-184 [PMID: 8699958 DOI: 10.1097/00005792-199607000-00001]

19 **Lumbroso S**, Paris F, Sultan C; European Collaborative Study. Activating Gsalpha mutations: analysis of 113 patients with signs of McCune-Albright syndrome--a European Collaborative Study. *J Clin Endocrinol Metab* 2004; **89**: 2107-2113 [PMID: 15126527 DOI: 10.1210/jc.2003-031225]

20 **Coles N**, Comeau I, Munoz T, Harrington J, Mendoza-Londono R, Schulze A, Kives S, Kamath BM, Hamilton J. Severe Neonatal Cholestasis as an Early Presentation of McCune-Albright Syndrome *J Clin Res Pediatr Endocrinol* 2019; **11**: 100-103 [PMID: 29991465 DOI: 10.4274/jcrpe.galenos.2018.2018.0110]

21 **Gaujoux S**, Salenave S, Ronot M, Rangheard AS, Cros J, Belghiti J, Sauvanet A, Ruszniewski P, Chanson P. Hepatobiliary and Pancreatic neoplasms in patients with McCune-Albright syndrome. *J Clin Endocrinol Metab* 2014; **99**: E97-101 [PMID: 24170100 DOI: 10.1210/jc.2013-1823]

22 **Ikawa Y**, Yachi Y, Inoue N, Kato A, Okajima M, Yachie A. Neonatal McCune-Albright Syndrome with Giant Cell Hepatitis. *J Pediatr* 2016; **178**: 298 [PMID: 27592093 DOI: 10.1016/j.jpeds.2016.08.009]

23 **Corsi A**, Cherman N, Donaldson DL, Robey PG, Collins MT, Riminucci M. Neonatal McCune-Albright Syndrome: A Unique Syndromic Profile With an Unfavorable Outcome. *JBMR Plus* 2019; **3**: e10134 [PMID: 31485549 DOI: 10.1002/jbm4.10134]

24 **Gilbert MA**, Bauer RC, Rajagopalan R, Grochowski CM, Chao G, McEldrew D, Nassur JA, Rand EB, Krock BL, Kamath BM, Krantz ID, Piccoli DA, Loomes KM, Spinner NB. Alagille syndrome mutation update: Comprehensive overview of JAG1 and NOTCH2 mutation frequencies and insight into missense variant classification. *Hum Mutat* 2019; **40**: 2197-2220 [PMID: 31343788 DOI: 10.1002/humu.23879]

**Footnotes**

**Informed consent statement:** A written informed consent was obtained from the parents of the patient.

**Conflict-of-interest statement:** The authors have no conflicts of interest to declare.

**CARE Checklist (2016) statement:** The authors have read the CARE checklist, and the manuscript was prepared and reviewed according to the guidelines in the “CARE Checklist–2016: Information for writing a case report.”

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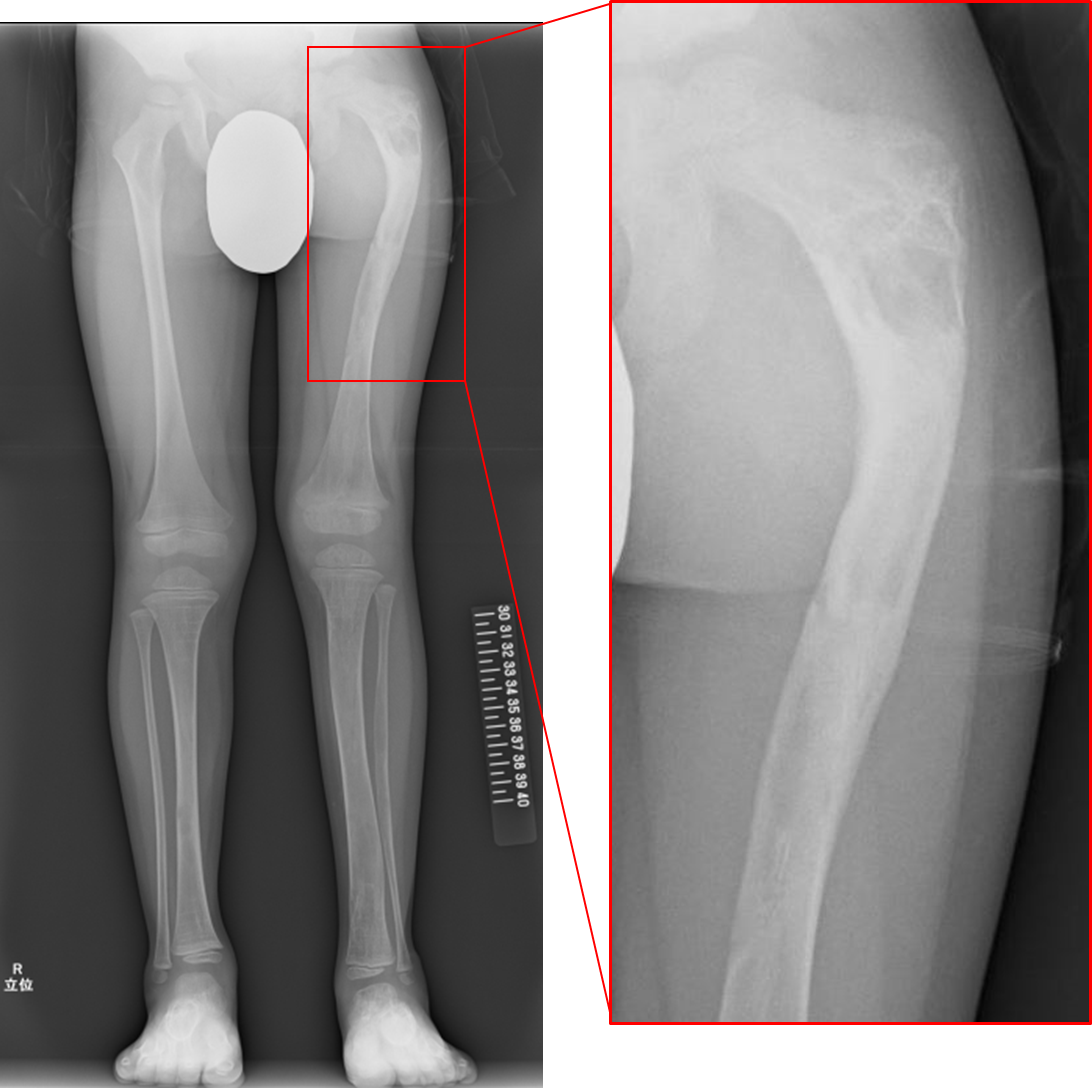
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Grade D (Fair): 0

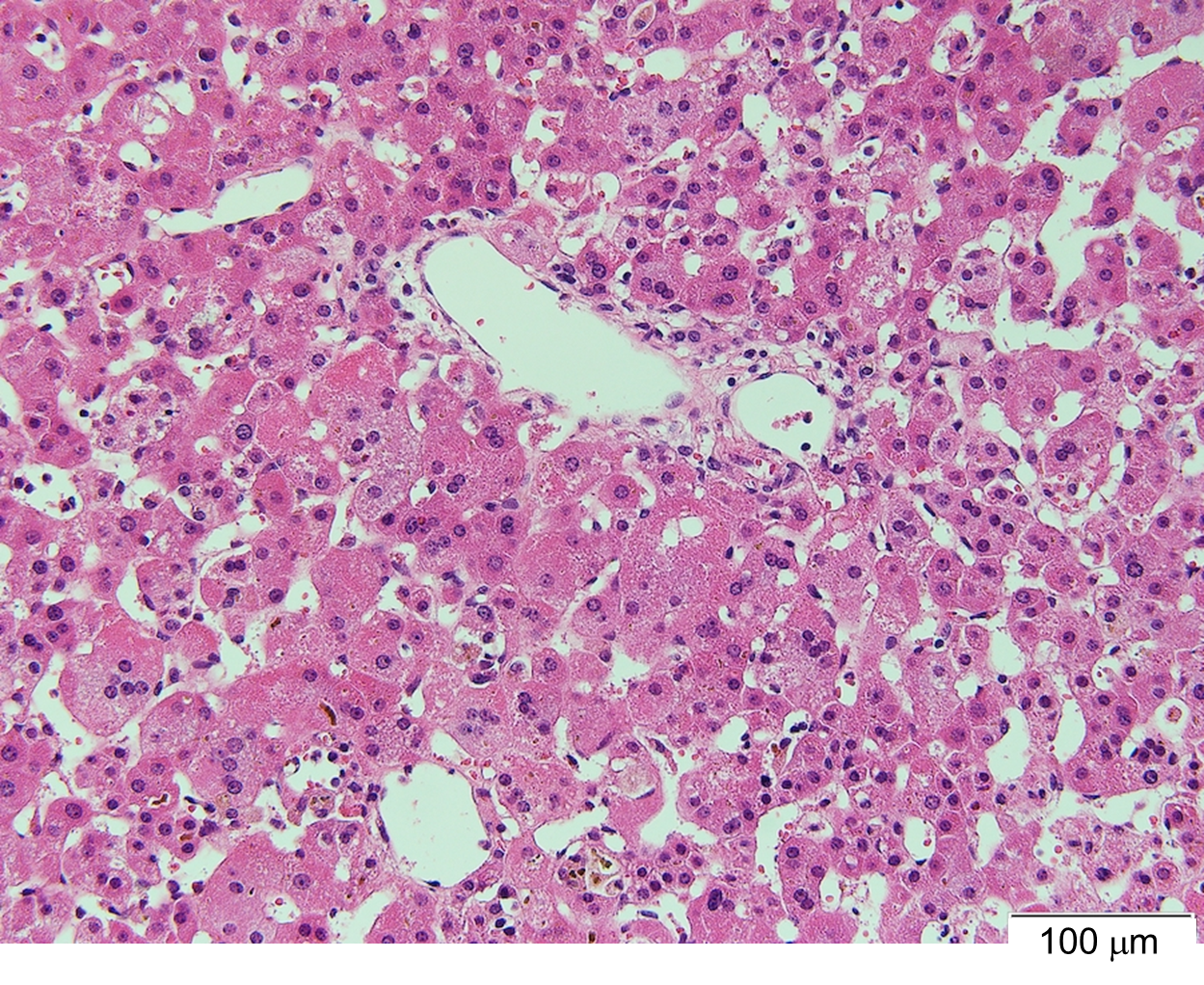
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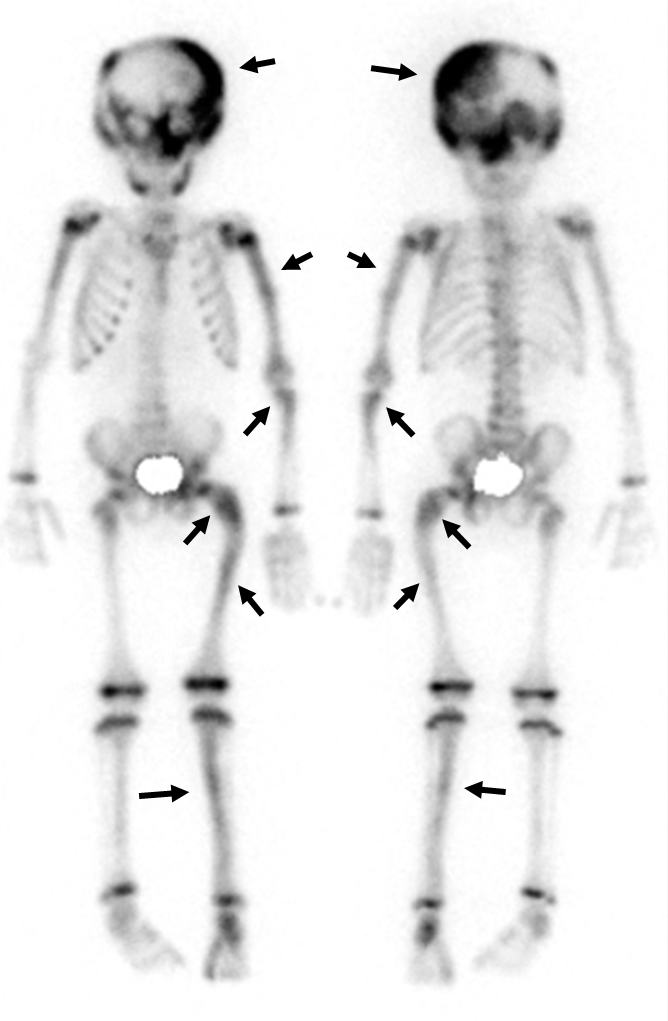
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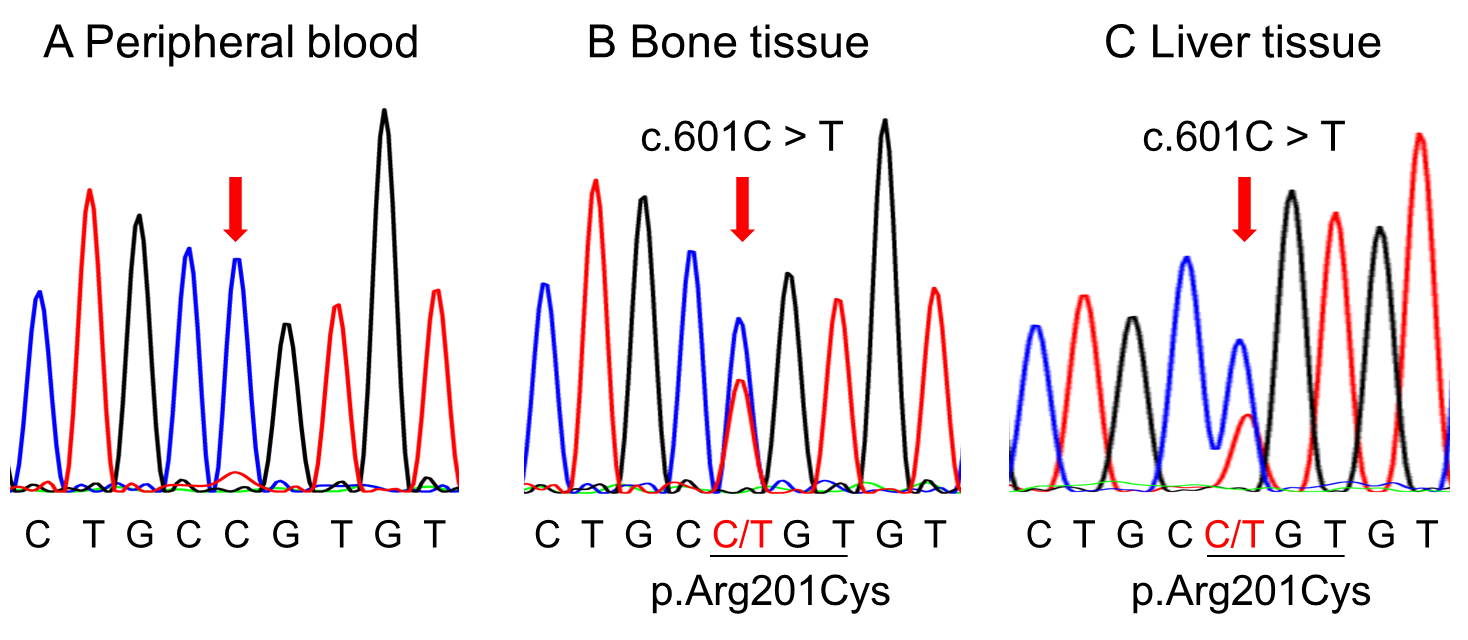
**Figure 1 Radiograph at the age of 4 years and 8 mo.** The radiograph demonstrated a “ground-glass” appearance in his left femur and left tibia and “shepherd’s crook deformity” which is characterized by the presence of proximal femoral varus deformity and retroversion deformity, in his left thigh bone.

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**Figure 2 Liver specimen at the age of 1 mo.** Microscopic examination revealed a lack of bile ducts in the portal area and giant cell transformation of hepatocytes (hematoxylin and eosin staining).

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**Figure 3 Bone scintigraphy with Tc-99 m-hydroxymethylene diphosphonate.** There are multiple hotspots with uptake at the left dominant skull and upper left limb in addition to the left femur and the left tibia.

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**Figure 4 DNA sequencing of the *GNAS* gene.** A: Normal sequencing is shown in the peripheral blood; B and C: Arg201Cys mutation was detected in the bone tissue samples and the liver tissue.