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***Basic Study***

**Estrogen augmented visceral pain and colonic neuron modulation in a double-hit model of prenatal and adult stress**

Chen JH *et al*. Estrogen re-enhanced visceral hyperalgesia

Jing-Hong Chen, Ying Sun, Pei-Jun Ju, Jin-Bao Wei, Qing-Jie Li, John H Winston

**Jing-Hong Chen, Ying Sun, Pei-Jun Ju, Jin-Bao Wei,** Shanghai Key Laboratory of Psychotic Disorders, Shanghai Mental Health Center, Shanghai Jiao Tong University School of Medicine, Shanghai 200030, China

**Jing-Hong Chen, Qing-Jie Li, John H Winston,** Department of Internal Medicine, University of Texas Medical Branch, Galveston, TX 77555, United States

**Author contributions:** Chen JH and Winston JH contributed to the concept and design of the experiments; Chen JH performed the experiments; Chen JH, Sun Y, Ju PJ, Wei JB, Li QJ and Winston JH analyzed the data and prepared the paper.

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**Corresponding author: John H Winston, PhD, Professor,** Department of Internal Medicine, University of Texas Medical Branch, 8-104 Med Res Bldg, Galveston, TX 77555, United States. jhwinsto@utmb.edu

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**Abstract**

BACKGROUND

Chronic stress during pregnancy may increase visceral hyperalgesia of offspring in a sex-dependent way. Combining adult stress in offspring will increase this sensitivity. Based on the evidence implicating estrogen in exacerbating visceral hypersensitivity in female rodents in preclinical models, we predicted that chronic prenatal stress (CPS) + chronic adult stress (CAS) will maximize visceral hyperalgesia; and that estrogen plays an important role in colonic hyperalgesia.

AIM

The aim was to illuminate the role of estrogen in colonic hyperalgesia and its underlying mechanisms.

METHODS

We established a CPS plus CAS rodent model in which the balloon was used to distend the colorectum. The single-fiber recording *in vivo* and patch clamp experiments *in vitro* were used to monitor the colonicneuron’s activity. The reverse transcription-polymerase chain reaction, western blot, and immunofluorescence were used to study the effects of CPS and CAS on colon primary afferent sensitivity. We used ovariectomy and letrozole to reduce estrogen levels of female rats respectively in order to assess the role of estrogen in female-specific enhanced primary afferent sensitization.

RESULTS

Spontaneous activity and single fiber activity were significantly greater in females than in males. The enhanced sensitization in female rats mainly came from low-threshold neurons. CPS significantly increased single-unit afferent fiber activity in L6-S2 dorsal roots in response. Activity was further enhanced by CAS. In addition, the excitability of colon-projecting dorsal root ganglion (DRG) neurons increased in CPS + CAS rats and was associated with a decrease in transient A-type K+ currents. Compared with ovariectomy, treatment with the aromatase inhibitor letrozole significantly reduced estrogen levels in female rats, confirming the gender difference. Moreover, mice treated with letrozole had decreased colonic DRG neuron excitability. The intrathecal infusion of estrogen increased brain-derived neurotrophic factor (BDNF) protein levels and contributed to the response to visceral pain. Western blotting showed that nerve growth factor protein was upregulated in CPS + CAS mice.

CONCLUSION

This study adds to the evidence that estrogen-dependent sensitization of primary afferent colon neurons is involved in the development of chronic stress-induced visceral hypersensitivity in female rats.

**Key Words:** Chronic prenatal stress; Estrogen; Visceral pain; Neuronal sensitization; Excitability; Letrozole

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**Core Tip:** We investigated whether estrogen re-enhanced visceral hyperalgesia in chronic prenatal stress plus chronic adult stress models. After using physical ovariectomy or chemical inhibition with letrozole to reduce estrogen levels, we found that visceral hyperalgesia, colonic afferent neuronal excitability, nerve growth factor and brain-derived neurotrophic factor, and estrogen were all increased. The findings indicate that chronic stress-induced visceral hypersensitivity was estrogen dependent and that the hypersensitivity was mediated by estrogen-dependent sensitization of primary afferent colon neurons. The preclinical models provide key scientific evidence in support of developing gender-based visceral pain management.

**INTRODUCTION**

Visceral pain of colonic origin is the most prominent symptom in irritable bowel syndrome (IBS) patients[1]. Female IBS patients report more severe pain that occurs more frequently and with longer episodes than in male patients[1,2]. The ratio of female to male IBS is about 2:1 among patients seen in medical clinics[3]. Moreover, females have a higher prevalence of IBS co-morbidities such as anxiety and depression[4,5] and are more vulnerable to stress-induced exacerbation of IBS symptoms compared with males[3,6,7].

Clinical studies show that early life adverse experiences are risk factors for the development of IBS symptoms, including visceral pain and ongoing chronic stress, especially abdominal pain[8-10]. These factors contribute to the development of visceral hypersensitivity, a key component of the IBS symptom complex and one that may be responsible for symptoms of pain[11,12]. Our previous research found that the female offspring of mothers subjected to chronic prenatal stress (CPS) had a markedly greater visceral sensitivity than their male littermates following challenge by another chronic adult stress (CAS) protocol. A critical molecular event in the development of this female-enhanced visceral hypersensitivity is upregulation of brain-derived neurotrophic factor (BDNF) expression in the lumbar-sacral spinal cord of female CPS + CAS rats[13]. However, the neurophysiological changes underlying the enhanced female-specific visceral hypersensitivity and the role of hormone in the development of stress-induced visceral hypersensitivity are not well understood.

Visceral hypersensitivity in IBS involves abnormal changes in neurophysiology throughout the brain-gut axis. In IBS, there is evidence for sensitization of primary afferents to jejunal distention and electrical stimulation[14], and there is evidence for increased sensitivity of lumbar splanchnic afferents[15,16]. In animal models of either early life adverse events or adult stress-induced visceral hypersensitivity[17], there is evidence of colon primary afferent sensitization. However, the studies were performed in male rodents. Therefore, in this study, we established a CPS and CAS rodent model to analyze the impact on female colon afferent neuron function and the role of estrogen. Our hypothesis was that female CPS offspring subjected to chronic stress as adults would exhibit greater colonic dorsal root ganglion (DRG) neuron sensitization compared with their male littermates, and that the enhanced visceral sensitization and primary afferent sensitization in females was estrogen dependent.

**MATERIALS AND METHODS**

***Animals***

The Institutional Animal Care and Use Committee of the University of Texas Medical Branch at Galveston, TX approved all animal procedures. Experiments were performed on pregnant Sprague Dawley rats and their 8-wk-old to 16-wk-old male and female offspring. Rats were housed individual cages with access to food and water in a room with controlled conditions (22 ± 2 °C, relative humidity of 50% ± 5%), and a 12 h light/12 h dark cycle.

***CPS and CAS models***

Pregnant dams were subjected to a CPS protocol that consisted of a random sequence of twice-daily applications of one of three stress sessions, a 1-h water-avoidance, 45-min cold-restraint, or a 20-min forced swim starting on day 6 and continuing until delivery on day 21. Male and female offspring of the stressed dams were designated CPS rats. Control dams received sham stress and their offspring were designated control rats. As adults at 8-16 wk of age, control and prenatally stressed offspring were challenged by the same CAS protocol for 9 d. Ovariectomy (OVX) or sham surgery was performed on female prenatal-stress offspring on day 56. Daily letrozole treatment was initiated on day 49, 2 wk prior to initiation of adult stress. Treatment was continued through the stress protocol. A schematic diagram of the study procedures is shown in Figure 1A.

***Rat treatment***

Before OVX or letrozole treatment, vaginal smears were used to identify the estrus cycle phase. OVX or sham surgery was performed on female prenatal-stress offspring on day 56. The aromatase inhibitor letrozole [4,4’-(1H-1,2,4-triazol-l-yl-methylene)-bis-benzonitrile], (Novartis) 1.0 mg/kg was orally administered in the experimental group and vehicle (hydroxypropyl cellulose 0.3% in water) was given in the control group once daily for 14 d. Direct transcutaneous intrathecal injections of estrogen and letrozole were performed as described by Mestre *et al*[18].

***In vivo single fiber recording of L6-S2 DRG rootlets***

Multiunit afferent discharges were recorded from the distal ends of L6-S2 dorsal rootlets decentralized close to their entry into the spinal cord. A bundle of multiunit fibers was distinguished into 2-6 single units off-line using wave mark template matching in Spike 2 software that differentiates spikes by shape and amplitude. Colonic afferent fibers were identified by their response to graded colorectal distention (CRD). A balloon was used to distend the colorectum. Isoflurane, 2.5%, followed by 50 mg/kg intraperitoneal sodium pentobarbital induced general anesthesia that was maintained by infusing a mixture of pentobarbital sodium + pancuronium bromide + saline by intravenous infusion through the tail vein. The adequacy of anesthesia was confirmed by the absence of corneal and pupillary reflexes and stability of the end-tidal CO2 level. A tracheotomy tube connected to a ventilator system provided a mixture of room air and oxygen. Expired CO2 was monitored and maintained at 3.5%. Body temperature was monitored and maintained at 37 °C by a servo-controlled heating blanket. A laminectomy from T12 to S2 exposed the spinal cord. The head was stabilized in a stereotaxic frame.

***In vitro patch clamp recordings in colonic DRG neurons***

**Retrograde fluorescence label injections:** Labeling of colon-projecting DRG neurons was performed as previously described[13]. Under general 2% isoflurane anesthesia, the lipid soluble fluorescent dye, 1,1’-dioleyl-3,3,3’,3’-tetramethylindocarbocyanine methane-sulfonate (9-DiI, Invitrogen, Carlsbad, CA) was injected (50 mg/mL) into the muscularis externa on the exposed distal colon in 8 to 10 sites (2 μL each site). To prevent leakage, the needle was kept in place for 1 min following each injection.

**Dissociation and culture of DRG neurons:** Rats were deeply anesthetized with isoflurane followed by decapitation. Lumbosacral (L6–S2) DRGs were collected in ice cold and oxygenated dissecting solution, containing (in mM) 130 NaCl, 5 KCl, 2 KH2PO4, 1.5 CaCl2, 6 MgSO4, 10 glucose, and 10 HEPES, pH 7.2 (305 mOsm). After removal of the connective tissue, the ganglia were transferred to a 5 mL dissecting solution containing collagenase D (1.8 mg/mL; Roche) and trypsin (1.0 mg/mL; Sigma, St Louis, MO), and incubated for 1.5 h at 34.5 °C. DRGs were then taken from the enzyme solution, washed, and put in 0.5-2 mL of the dissecting solution containing DNase (0.5 mg/mL; Sigma). Cells were subsequently dissociated by gentle trituration 10 to 15 times with fire-polished glass pipettes and placed on acid-cleaned glass coverslips. The dissociated DRG neurons were kept in 1 mL DMEM (with 10% FBS) in an incubator (95% O2/5% CO2) at 37 *°*C overnight.

**Whole-cell patch clamp recordings from dissociated DRG neurons:** Before each experiment, a glass coverslip with DRG neurons was transferred to a recording chamber perfused (1.5 mL/min) with external solution containing (10 mM): 130 NaCl, 5 KCl, 2 KH2PO4, 2.5 CaCl2, 1 MgCl2, 10 HEPES, and 10 glucose, pH adjusted to pH 7.4 with NaOH (300 mOsm) at room temperature. Recording pipettes, pulled from borosilicate glass tubing, with resistance of 1-5 MΩ, were filled with solution containing (in mM): 100 KMeSO3, 40 KCl, and 10 HEPES, pH 7.25 adjusted with KOH (290 mOsm). DiI-labeled neurons were identified by fluorescence microscopy. Whole-cell currents and voltage were recorded from DiI-labeled neurons using a Dagan 3911 patch clamp amplifier. Data were acquired and analyzed by pCLAMP 9.2 (Molecular Devices, Sunnyvale, CA). The currents were filtered at 2–5 kHz and sampled at 50 or 100 s *per* point. While still under voltage clamp, the Clampex Membrane Test program (Molecular Devices) was used to determine membrane capacitance (Cm) and membrane resistance (Rm), during a 10 ms, 5 mV depolarizing pulse form a holding potential of −60 mV. The configuration was then switched to current clamp (0 pA) to determine other electrophysiological properties. After stabilizing for 2–3 min, the resting membrane potential was measured. The minimum acceptable resting membrane potential was −40 mV. Spontaneous activity (SA) was then recorded over two 30 s periods separated by 60 s without recording, as described by Bedi *et al*[19].

**Transient A-type K+ current (IA) recording method in patch studies:** To record voltage-gated K+ current (Kv), Na+ in control external solution was replaced with equimolar choline and the Ca2+ concentration was reduced to 0.03 mM to suppress Ca2+ currents and to prevent Ca2+ channels becoming Na+ conducting. The reduced external Ca2+ would also be expected to suppress Ca2+-activated K+ currents. The current traces of Kv in DRG neurons were measured at different holding potentials. The membrane potential was held at −100 mV and voltage steps were from −40 to +30 mV to record the total Kv. The membrane potential was held at −50 mV to record the sustained Kv. The IA currents were calculated by subtracting the sustained current from the total current. The current density (in pA/pF) was calculated by dividing the current amplitude by cell membrane capacitance.

***Real time reverse transcription-polymerase chain reaction (RT-PCR)***

Total RNA was extracted using RNeasy Mini Kits (QIAGEN, Valencia, CA). One microgram of total RNA was reverse-transcribed using the SuperScriptTM III First-Strand Synthesis System. PCR assays were performed on a StepOnePlus thermal cycler with 18 s as the normalizer using Applied Biosystems primer/probe set Rn02531967\_s1 directed against the translated exon IX. Fold-change relative to control was calculated using the ΔΔCt method (Applied Biosystems).

***Western blot***

Samples were lysed in RIPA buffer containing protease inhibitor cocktail and phenylmethanesulfonyl fluoride. Lysates were incubated for 30 min on ice and then centrifuged at 10 000 × *g* for 10 min at 4 °C. The protein concentration in the supernatant was determined using bicinchoninic acid (BCA) assay kits with bovine serum albumin as a standard. Equal amounts of protein (30 μg per lane) were separated by 10% sodium dodecyl sulfate polyacrylamide gel electrophoresis (SDS-PAGE) and then transferred to nitrocellulose membranes (Bio-Rad, United States). The membranes were blocked in Li-Cor blocking buffer for 1 h at room temperature and then incubated with primary antibodies. BDNF antibody (Santa Cruz Biotechnologies, Santa Cruz, CA) was used at 1:200 dilution; nerve growth factor (NGF)antibody (Abcam, MA) was used at 1:1000 dilution; β-actin antibody (Sigma Aldrich, St Louis, MO) was used at 1:5000 dilution. The secondary antibodies were donkey anti-rabbit Alexa Fluor 680 (Invitrogen) and goat anti-mouse IRDye 800 (Rockland). Images were acquired and band intensities measured using a Li-Cor Odyssey system (Li-Cor, Lincoln, NE).

***Immunofluorescence***

Frozen sections of colon tissue from control, CAS, CPS and CPS + CAS female rats were mounted on glass slides, and rehydrated in phosphate buffered saline at room temperature. The slides were treated for antigen retrieval and blocked with 10% normal goat serum diluted in 0.3% phosphate buffered saline-Triton for 1 h, and then incubated with NGF primary antibody in antibody diluent (Renoir Red, Biocare Medical, Concord, CA) at 4 °Covernight. The slides were exposed to fluorescent dye-conjugated secondary antibody for 2 h at room temperature, counterstained with 4',6-diamidino-2-phenylindole and coverslipped. Images were taken in fluorescence mode on an Olympus laser scanning confocal microscope and the average signal intensity was calculated by the bundled software.

***Serum estradiol and norepinephrine levels***

Serum estradiol, adrenocorticotropic hormone (ACTH), and norepinephrine levels were measured using specific enzyme-linked immunosorbent assay kits for each analyte (CSB-E05110r, CSB-E06875r, CSB-E07022, Cusabio Bioteck CO., United States) following the manufacturer’s instructions.

***Data analysis***

Single fiber responses (impulses/second) to CRD were calculated by subtracting SA from the mean 30 s maximal activity during distension. Fibers were considered responsive if CRD increased their activity to 30% greater than the baseline value. Mechanosensitive single units were classified as high threshold (> 20 mmHg) or low threshold (≤ 20 mmHg) on the basis of their response threshold and profile during CRD. Single fiber activity data were analyzed by analysis of variance with repeated measures; CRD intensity was the repeated factor and the experimental group was the between-group factor. If significant main effects were present, the individual means were compared using the Fisher post-hoc test. All authors had access to the study data and reviewed and approved the final manuscript.

**RESULTS**

***Effects of CPS plus CAS on primary afferent responses to CRD in male and female rats***

The basal activity of a spinal afferent fiber was defined as the average number of action potentials persecond (impulses/sec) in the 60 s period before the onset of a distention stimulus. In male controls, 66% of the afferent fibers under study displayed SA and SA was significantly higher in female controls than in male controls (0.71 ± 0.21 *vs* 1.24 ± 0.20 imp/sec; Figure 1B). The average single fiber activity in response to CRD was significantly higher in female control rats compared with male controls (Figure 1C). We found that the enhanced sensitization in female rats mainly came from the low-threshold fibers (Figure 1D and E).

To assess the effects of CPS + CAS on colon afferent fiber activities, we compared average single colon afferent fiber activities projecting from dorsal roots S1-L6 in response to CRD in male and female control, CPS, control + CAS and CPS + CAS rats recorded approximately 24 h after the last stressor. In females, CPS significantly increased single-unit afferent activity in response to CRD *vs* control female rats (Figure 1F). CAS alone enhanced single-unit activity compared with control. The increase in average afferent responses after CAS in prenatally stressed female rats (44.0%) was significantly greater than the increase in female control rats (39.3%). In males, CPS had no significant effect on primary afferent responses (Figure 1G). When we compared males to females within each experimental group, we found that the average single fiber activity was significantly higher in female compared with male CPS + CAS rats (Figure 1F, G). The increased activity may contribute to the enhanced female visceral hypersensitivity previously reported in this model. Average single-fiber activities were significantly greater in control and CPS and CAS females than in their corresponding male experimental groups (Figure 1F and G). Both CAS and CPS + CAS rats had significantly increased primary afferent responses compared with control and CPS rats. Thus, our CPS and CAS protocols sensitized colon-projecting primary afferent fibers, with the greatest effects produced by the combination of CPS + CAS in both males and females.

***Increase in excitability of colon-projecting lumbosacral DRG neurons in female CPS + CAS rats***

To elucidate the electrophysiological basis of enhanced stress-induced primary afferent activity in female rats, we performed patch clamp studies on acutely dissociated retrograde-labeled colon-projecting neurons from the L6-S2 DRGs in control, prenatal stress, adult stress only, and CPS + CAS female rats isolated 24 h after the last adult stressor (Figure 2A). Input resistance (Figure 2B) and rheobase (Figure 2C) were significantly decreased in neurons from CPS + CAS rats compared with the other three groups. The number of action potentials elicited at either 2 × or 3 × the rheobase were significantly greater in adult stress and CPS + CAS neurons compared with control and to CPS neurons (Figure 2D, E). CAS significantly increased action potential overshoot with or without CPS (Figure 2F), but it did not significantly alter other electrophysiological characteristics, such as number of spontaneous spikes, membrane capacitance (pF), resting membrane potential, cell diameter, time constant, and DRG neuron action potential amplitude and duration (Table 1).

The percentage of neurons with SA in was significantly greater in CPS + CAS rats than in control or CPS only rats (Figure 2G). Under voltage clamp conditions (Figure 2H), neurons from female CPS + CAS, CAS, CPS and control groups had IA and sustained outward rectifier K+ currents (IK). Compared with the other three groups, DRG neurons from CPS + CAS rats had significantly reduced average IA (*P* < 0.05). The average IK density was decreased but the change was not significant.

***Effects of CPS and/or CAS on plasma estrogen concentration***

We did a vaginal smear test to identify the estrus cycle phases by identifying the vaginal cytological cell types. Estrogen concentration was significantly higher in the CPS proestrus/estrus phase compared with control diestrus, control proestrus/estrus, and CPS diestrus proestrus (*P* < 0.05; Figure 3A). Comparison of the plasma estrogen concentrations in control, CAS, CPS, CPS + CAS showed that CPS significantly increased plasma estrogen levels compared with the control rats and that CAS increased plasma estrogen level compared with the control and CPS rats (Figure 3B).

To determine whether estrogen contributed to stress-induced visceral hypersensitivity in prenatal stressed females, we reduced plasma estrogen levels by either OVX or letrozole treatment. OVX significantly lowered serum estradiol levels before and after CAS (Figure 3C). Treatment was continued throughout CAS. After treatment with letrozole, serum estradiol levels were significantly reduced (Figure 3D). To study the effects of gender and stress on norepinephrine and ACTH levels, we measured plasma norepinephrine levels in female rats from all four experimental groups. CAS alone significantly increased plasma norepinephrine levels compared with both the controls and with CPS alone (Figure 3E) Plasma norepinephrine levels were significantly increased in CPS + CAS rats compared with CAS alone as well as with controls and CPS. Plasma ACTH levels were significantly increased in CPS + CAS rats compared with controls. (Figure 3F).

***Effects of Letrozole treatment on colon DRG neuron excitability***

We performed patch clamp experiments on acutely isolated retrograde-labeled DRG neurons from CPS + CAS females with or without letrozole treatment 24 h after the last adult stressor. Letrozole treatment significantly increased rheobase (Figure 4A), and significantly reduced input resistance (Figure 4B). Action potential overshoot (Figure 4C) and the number of action potentials elicited by a current injection at either 2 × or 3 × rheobase were significantly reduced by letrozole treatment (Figure 4D). Other electrophysiological properties were not significantly altered (Table 2). We also recorded electromyographic activity to determine whether the reduction in visceral sensitivity in female CPS + CAS rats caused by OVX or systemic letrozole treatment reduced visceromotor responses. The findings demonstrated a significant decrease in excitability of colon-projecting L6-S2 neurons.

***Spinal cord BDNF levels regulated by estrogen***

To investigate the effect of estrogen on BDNF expression, we measured BDNF mRNA and protein levels in the lumbar-sacral spinal cords of OVX and Sham CPS + CAS female rats. Systemic estradiol administration to naïve cycling females produced significant increases in plasma estrogen (Figure 5A), lumbar-sacral spinal cord BDNF mRNA (Figure 5B), and protein (Figure 5C). We also measured BDNF mRNA and protein levels in the lumbar-sacral spinal cords of OVX and Sham CPS + CAS female rats. BDNF mRNA and protein expression were significantly suppressed by OVX compared with sham rats. Another experiment showed that intrathecal infusion of estrogen into naïve female rats significantly increased BDNF protein levels, which proved that estrogen reversed the experimental results and contributed to the response to visceral pain[13].

***Peripheral NGF level increased in CPS + CAS female rats***

We examined NGF expression in the colons of females from all four experimental groups by immunohistochemistry (Figure 6A). Morphometric analysis showed that CAS and CPS + CAS significantly increased NGF levels in the colon wall, with the increase in CPS + CAS significantly greater that of CAS alone (Figure 6B). Western blotting showed that NGF protein was significantly upregulated in CPS + CAS rats compared with controls (Figure 6C).

**DISCUSSION**

Enhanced CPS-induced visceral hypersensitivity in female rats was associated with an increase in the responses of lumbosacral nerve fibers to CRD in both male and female offspring. These findings are further supported by data showing increased excitability of colon-projecting DRG neurons from females in patch clamp studies. The magnitude of the sensitization was the greatest in female CPS + CAS rats, suggesting that it made a major contribution to the observed enhanced female visceral hypersensitivity in our model.

Chronic stress is known to increase the excitability of colon-projecting DRG neurons in rats and mice. In adult male Sprague Dawley rats, colon DRG neuron sensitization was shown to be driven by increases in NGF expression in the colon muscularis externa[13]. In our model, we also observed a significant increase in colon NGF, but its potential role in primary afferent sensitization and visceral hypersensitivity was not investigated.. Other studies in male mice showed that stress, in the form of water avoidance, significantly increased the excitability of colon-projecting DRG neurons and that the combined activity of the stress mediators corticosterone and norepinephrine increased DRG neuron excitability in vitro[20,21]. We previously found significant increases in the serum levels of norepinephrine in CPS + CAS females. However, daily systemic treatment with adrenergic antagonists during the adult stress protocol failed to reduce visceral hypersensitivity in female neonatal + adult stress rats[13] suggesting that norepinephrine did not play a major role in the acquisition of enhanced female visceral hypersensitivity or primary afferent sensitization in our model.

When we tested on lumbar-sacral afferent fibers and dissociated neurons in patch clamp studies, we found significant decreases in transient potassium IA currents in neurons isolated from CPS + CAS females compared with the other three experimental groups. Declines in A-type Kv currents in DRG neurons have been associated with persistent pain in multiple chronic pain models[22]. Whether the decline was caused by changes in channel properties or expression was not investigated in this study. However, another study demonstrated that estrogen significantly shifted the activation curve of IA currents in the hyperpolarizing direction and that estrogen inhibited Kv (+) channels in mouse DRG neurons through a membrane ER-activated nongenomic pathway[23].

Our results showed that the excitability of colon-projecting neurons in CPS + CAS females was significantly reduced by systemic letrozole treatment, suggesting that estrogen contributed to the sensitization process. Previous studies show that estrogen receptors expressed on primary afferent neurons contributed to enhanced sensitivity in various pain models[24-26]. One study found no decline in the responses of colon-projecting nerve fibers to CRD following OVX and found no detectable estrogen receptor alpha immunoreactivity in colon-projecting DRG neurons[27]. The reasons for the differing results are not clear, but local production of estrogen in DRG neurons could be sufficient to sustain sensitization.

NGF and its receptors play important roles in the mechanism of visceral pain and hyperalgesia in women. For example, endometriosis is estrogen dependent and is commonly diagnosed. The main symptoms are various types of pelvic pain that have a serious effect on physical and mental health, but the mechanisms of abdominal pain are still unclear. Studies have shown NGF to be an inflammatory mediator and modulator of pain in adulthood[28].

**CONCLUSION**

In this study, we examined the sex differences and effects of estrogen on the acquisition of enhanced visceral hypersensitivity in the offspring of rats in a model of prenatal and adult stress as shown in Figure 7. Our study shows that estrogen acted in the spinal cord and the primary afferent neurons to enhance visceral nociception. Acute blockade of the endogenous synthesis of estrogens in rat spinal cord significantly reduced visceral hypersensitivity, suggesting that locally produced estrogen in the central nervous system can regulate nociceptive neurons to modulate visceral hypersensitivity. The chronic stress-estrogen-BDNF axis sensitized visceral hypersensitivity in the offspring of females subjected to CPS. The development of chronic stress-induced visceral hypersensitivity in female rats was estrogen dependent. A key component of this hypersensitivity was estrogen-dependent sensitization of primary afferent colon neurons. Our findings provide key scientific evidence in a preclinical model in support of developing gender-based treatment for abdominal pain in IBS.

**ARTICLE HIGHLIGHTS**

***Research background***

Chronic stress during pregnancy may increase visceral hyperalgesia in the offspring. Combining adult stress in offspring will increase this sensitivity. Therefore, based on the evidence implicating estrogen exacerbates visceral hypersensitivity in female rodents in preclinical models, we predicted that chronic prenatal stress (CPS) + chronic adult stress (CAS) would maximize visceral hyperalgesia and that estrogen has an important role in colonic hyperalgesia.

***Research motivation***

The mechanisms of visceral hypersensitivity are not well defined. Understanding the neurophysiological mechanisms driving visceral hypersensitivity will spur the development of female pain-specific therapies.

***Research objectives***

The objective was to identify the enhancement of visceral hypersensitivity in a CPS + CAS model and explain the role of estrogen in that process.

***Research methods***

A CPS + CAS rodent model was established. Single fiber recording *in vivo* and patch clamp experiments *in vitro* were used to monitor the activity of colonic neurons. Reverse transcription-polymerase chain reaction (RT-PCR), western blots, and immunofluorescence were used to study the effects of CPS and CAS on colon primary afferent sensitivity. We used ovariectomy (OVX) and letrozole to reduce estrogen levels in female rats in order to assess the role of estrogen in female-specific enhanced primary afferent sensitization.

A CPS + CAS rodent model was established. Single fiber recording *in vivo* and patch clamp experiments were used to monitor the colonic neuron activity in vitro. RT-PCR, western blots, and immunofluorescence were used to study the effects of CPS and CAS on colon primary afferent sensitivity. We used OVX and letrozole to reduce estrogen levels of female rats in order to assess the role of estrogen in female-specific enhanced primary afferent sensitization.

***Research results***

Spontaneous activity and single fiber activity were significantly greater in females than in males. The enhanced sensitization in female rats mainly came from the low-threshold neurons. CPS significantly increased single-unit afferent fiber activity in the L6-S2 dorsal roots in response. Activity was further enhanced by CAS. In addition, the increased excitability of colon-projecting DRG neurons in CPS + CAS rats was associated with a decrease in transient A-type K+ currents. Compared with OVX, letrozole treatment further reduced the estrogen levels of female rats, which confirmed the gender difference. Moreover, rats treated with letrozole had decreased colonic DRG neuron excitability. The intrathecal infusion of estrogen increased BDNF protein levels and contributed to the response to visceral pain. Western blots showed that nerve growth factor protein was upregulated in CPS + CAS rats.

***Research conclusions***

This study adds to the evidence of the development of chronic stress-induced visceral hypersensitivity in females, and that it involves estrogen-dependent sensitization of primary afferent colon neurons.

***Research perspectives***

This study demonstrated that CAS + CPS induced visceral hypersensitivity and that estrogen played a role in the process. Understanding the molecular and neurophysiological mechanisms driving this response will spur the development of female pain-specific therapies.

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**Footnotes**

**Institutional review board statement:** The study was undertaken with the approval of the Institutional Animal Care and Use Committee of the University of Texas Medical Branch at Galveston, TX, United States.

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**Conflict-of-interest statement:** The authors declare that they have no conflicting interests.

**Data sharing statement:** No additional data are available.

**ARRIVE guidelines statement:** The authors have read the ARRIVE guidelines, and the manuscript was prepared and revised according to the ARRIVE guidelines.

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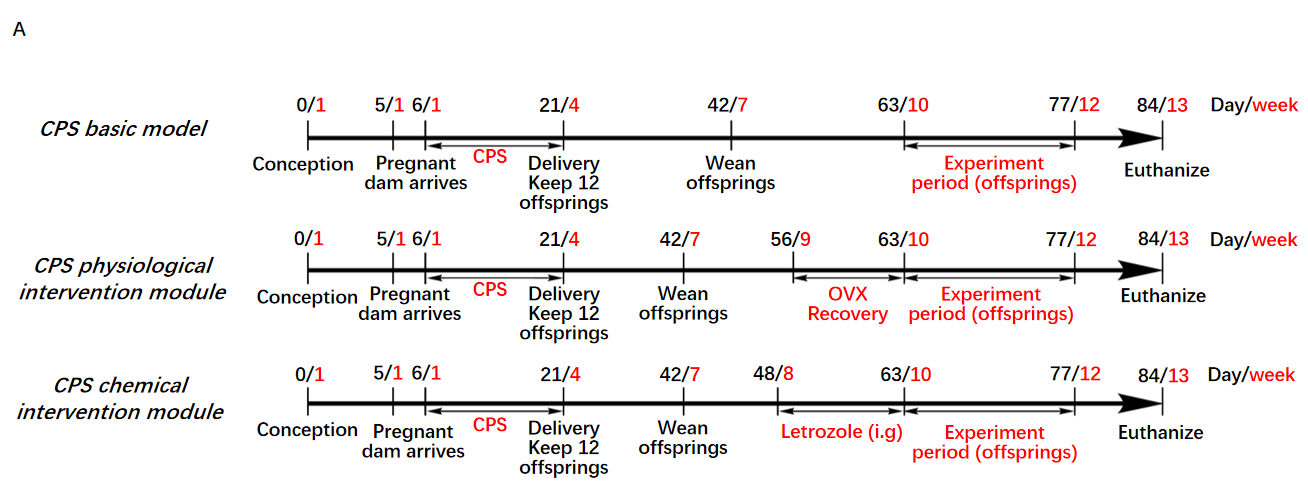
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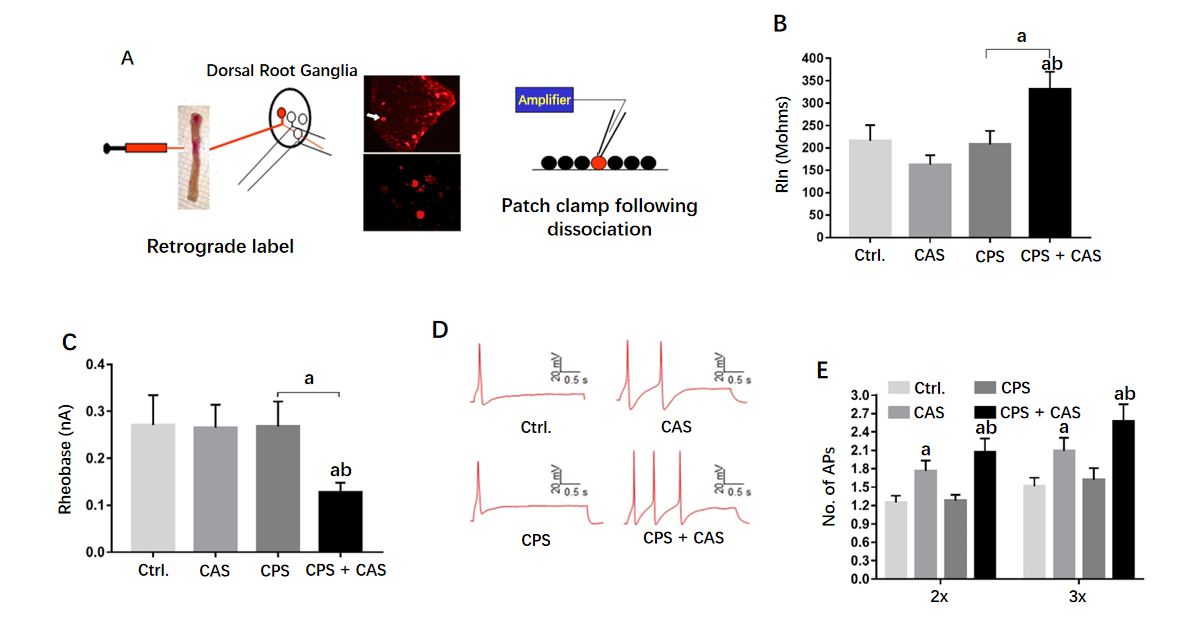
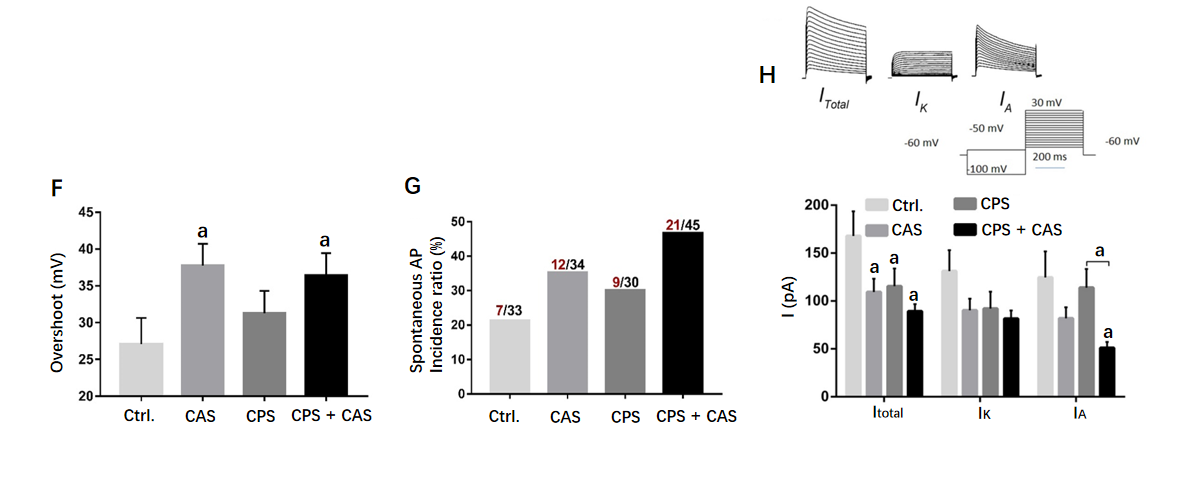
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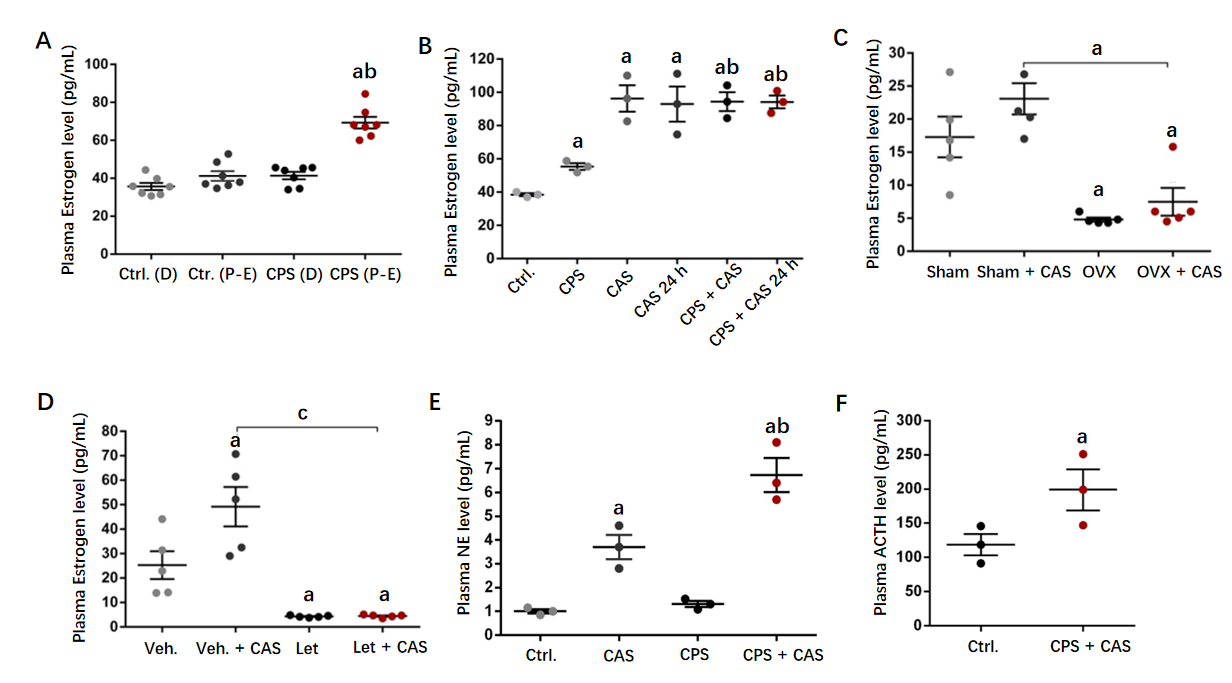
**Figure Legends**

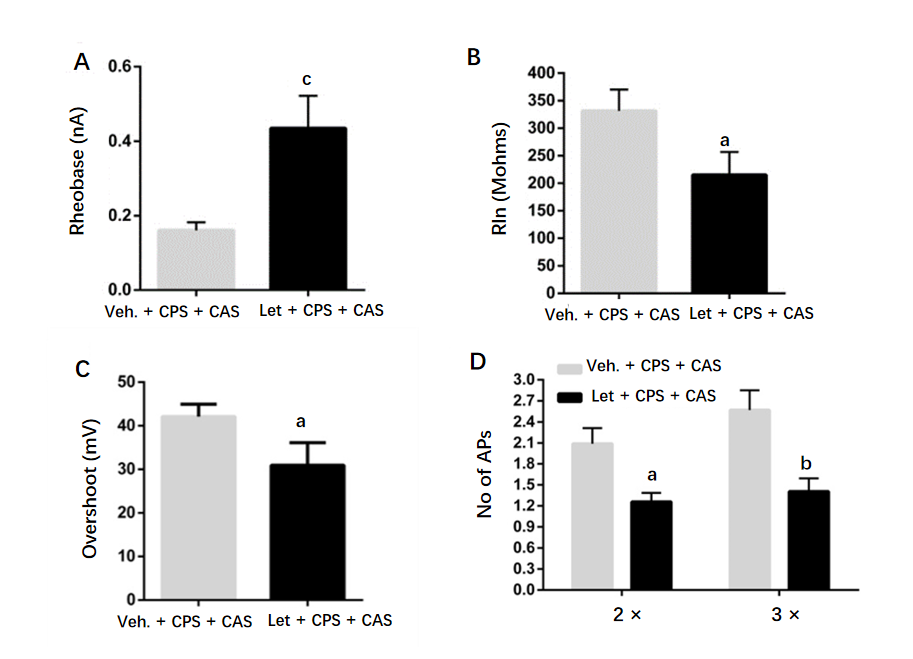
**Figure 1 Primary afferent responses to colorectal distention.** A: Chronic prenatal stress (CPS) plus chronic adult stress (CAS) model. Pregnant dams were subjected to prenatal stress from on day 11 of gestation. Ovariectomy (OVX) or sham surgery was performed on female prenatal-stress offspring on day 56. Daily Letrozole was initiated on day 49, 2 wk prior to initiation of adult stress. Treatment was continued through the stress protocol; B: Spontaneous activity (SA) of single afferent units in male and female control rats (*n* = 70 fibers in 6 rats in each group, *t*-test, a*P* < 0.05); C: Average response to graded colorectal distention (CRD) of 56 afferent fibers in 6 male and 70 afferent fibers in 6 female control rats; two-way analysis of variance (ANOVA; a*P* < 0.05 *vs* the same pressure male group); D: Responses of low-threshold (LT) fibers to CRD in 42 fibers in 6 male rats and 40 fibers in 6 female control rats (ANOVA, a*P* < 0.05 *vs* the same pressure male group); E: Responses of high-threshold (HT) afferent fibers to CRD in 14 fibers in 6 male and 29 fibers in 6 female control rats; F: Effects of CAS on afferent fiber responses to CRD from 59 fibers in 6 control and 99 fibers in 6 CPS female rats; (two-way ANOVA, a*P* < 0.05 *vs* the same pressure control group, b*P* < 0.05 *vs* the same pressure CPS group); G: Effects of CAS on afferent fiber responses to CRD in control and CPS male rats (*n* = 6 rats, 57 fibers for control and 95 fibers for CPS female group; two-way ANOVA, a*P* < 0.05 *vs* the same pressure-control group).

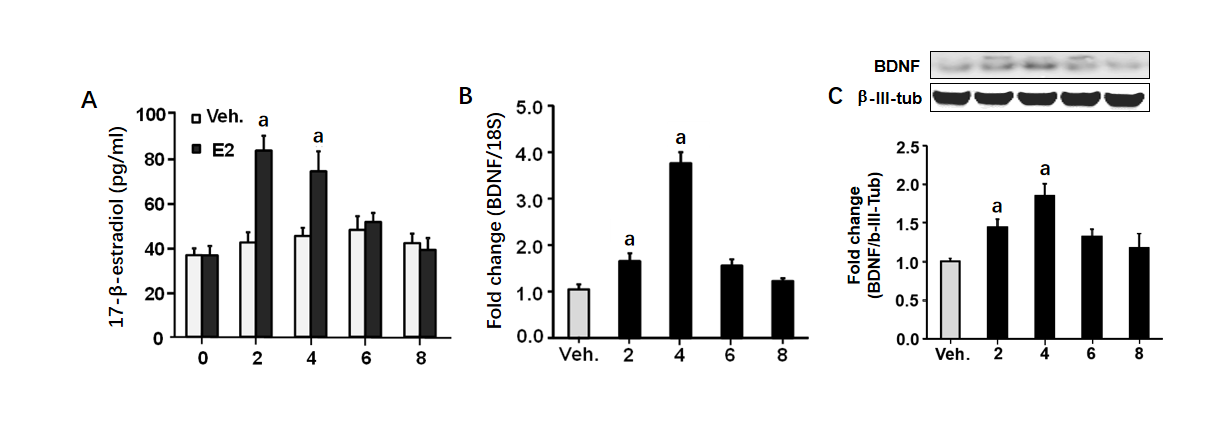
**Figure 2 Patch clamp recording in colonic dorsal root ganglion neurons from female rats.** A: Patch clamp process of cell labeling. Under isoflurane anesthesia, the lipid soluble fluorescent dye 9-DiI was injected into muscularis externa of the exposed distal colon (left figure). Lumbosacral (L6–S2) dorsal root ganglions (upper photograph) were isolated and DiI-labeled neurons were identified by fluorescence microscopy (lower photograph). Electrophysiological properties of each neuron were measured using whole-cell current and voltage clamp protocols (right figure); B: Rheobase from all four experimental groups (*n* = 5 rats, 45 cells in each group, one-way ANOVA, a*P* < 0.05 *vs* control or b*P* < 0.05 *vs* CAS); C: Representative action potentials (APs) elicited by current injection at 2 × the rheobase in neurons from control, chronic adult stress (CAS), chronic prenatal stress (CPS) and CPS + CAS female rats; D: Membrane input resistance from all four groups (*n* = 5 rats, 45 cells in each group, one-way ANOVA, a*P* < 0.05 *vs* control; b*P* < 0.05 *vs* CPS); E: Number of APs elicited by current injection at either 2 × and 3 × the rheobase in all four experimental groups (two-way ANOVA, a*P* < 0.05 *vs* control; b*P* < 0.05 *vs* CPS); F: AP overshoot recorded from all four experimental groups (a*P* < 0.05 *vs* control); G: The proportion of neurons from each experimental group exhibiting spontaneous APs. Red numbers represent spontaneous AP firing cells; black numbers represent total cells; H: Representative total, IK and IA current tracings and average values of potassium currents: Itotal, IK and IA are shown in female CPS + CAS, CAS, CPS (*n* = 15 neurons, from 5 rats in each group), and control groups (*n* = 12 neurons from 5 rats); two-way ANOVA, a*P* < 0.05 *vs* each control group.



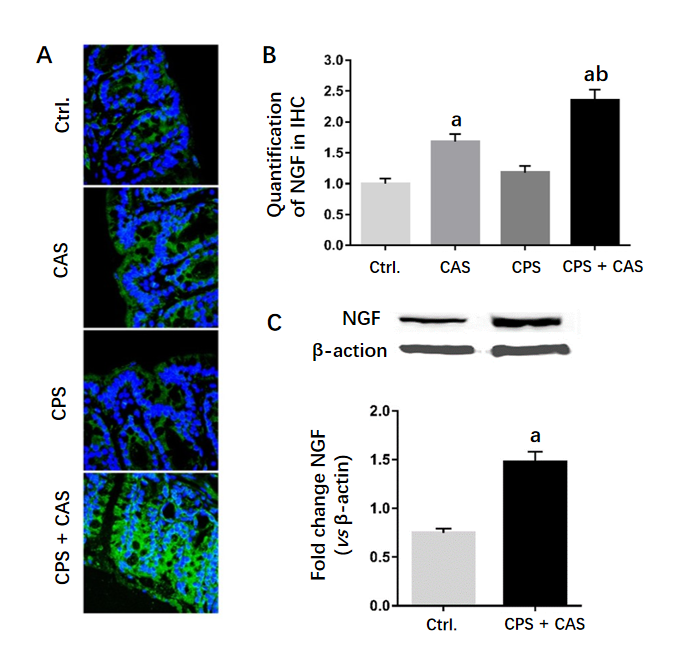
**Figure 3 Effects of chronic prenatal stress, chronic adult stress, ovariectomy, and letrozole treatment on plasma estrogen levels in female rats.** A: Plasma estrogen level in control and chronic prenatal stress (CPS) rats by estrus cycle phase (*n* = 8 rats, one-way ANOVA, a*P* < 0.05 *vs* control proestrus/estrus (P-E) phase; b*P* < 0.05 *vs* CPS diestrus (D) phase); B: Plasma estrogen levels increased in CPS rats and following chronic adult stress (CAS) 24 h after the last adult stressor (*n* = 8 rats, one-way ANOVA, a*P* < 0.05 *vs* control; b*P* < 0.05 *vs* CPS); C: Ovariectomy (OVX) significantly reduced CPS female rat plasma estrogen levels before and after CAS (*n* = 5 rats, one-way ANOVA, a*P* < 0.05 *vs* sham group); D: Letrozole treatment significantly reduced CPS female rat plasma estrogen levels before or after CAS (*n* = 5 rats, one-way ANOVA, a*P* < 0.05 *vs* vehicle group; c*P* < 0.0001); E: Plasma norepinephrine levels from control, CAS, CPS and CPS + CAS group female rats (*n* = 5 rats, one-way ANOVA, a*P* < 0.05 *vs* control; b*P* < 0.05 *vs* CPS); F: Plasma adrenocorticotropic hormone (ACTH) levels from control and CPS + CAS group female rats (*n* = 5 rats, *t*-test, a*P* < 0.05 *vs* control).



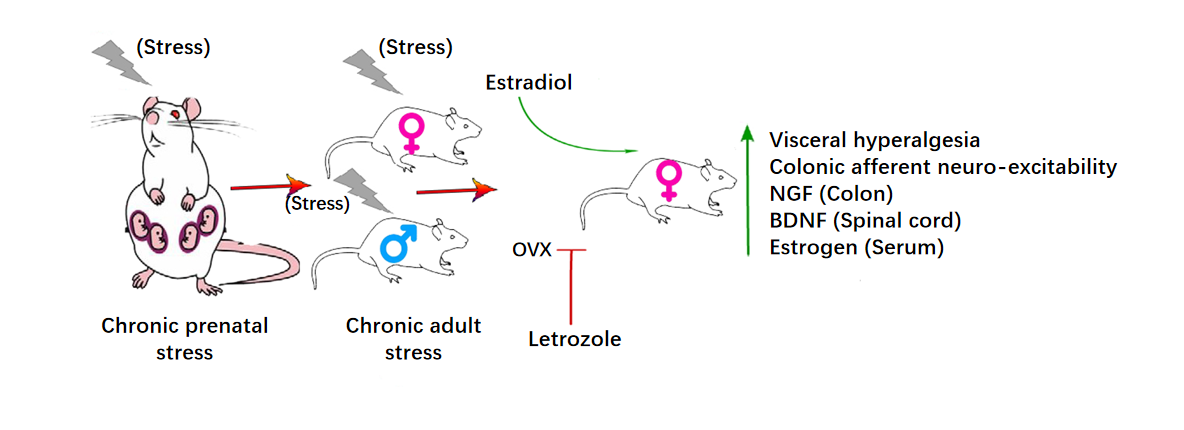
**Figure 4 Effects of Letrozole treatment on colon dorsal root ganglion neuron excitability.** A: Rheobase (*n* = 45 cells in 6 rats in each group, *t*-test, c*P* < 0.001 *vs* Veh. + chronic adult stress [CAS] + chronic prenatal stress [CPS]); B: Membrane input resistance (RIn) (*t*-test, a*P* < 0.05); C: Action potential (AP) overshoot (*t*-test, a*P* < 0.05); D: Number of APs elicited by current injection at 2 × and 3 × rheobase (two-way ANOVA, a*P* < 0.05; b*P* < 0.01).



**Figure 5 Brain-derived neurotrophic factor expression in lumbar-sacral spinal cord is regulated by estrogen.** A: Plasma estrogen levels in cycling females that received a bolus estradiol (E2) infusion on day 1; B: Lumbar-sacral spinal cord brain-derived neurotrophic factor (BDNF) mRNA following bolus estrogen infusion; C: Lumbar-sacral spinal cord BDNF protein following bolus estrogen infusion. (*n* = 8 rats in each group, two-way ANOVA, a*P* < 0.05 *vs* vehicle group).



**Figure 6 Nerve growth factor expression level in the colon wall.** A: Immunohistochemical staining of nerve growth factor (NGF; green) was detected with nuclear counterstaining staining (blue) in controls, chronic adult stress (CAS), chronic prenatal stress (CPS) and CPS + CAS group female rat colon walls. × 400 magnification representative pictures were shown; B: Quantification of NGF levels from colon wall in immunohistochemistry (IHC) (*n* = 4 rats in each group, one-way ANOVA, a*P* < 0.05 *vs* control group; b*P* < 0.05 *vs* CPS group); C: Western blots of NGF protein from control and CPS + CAS female rats colon wall tissue (*n* = 6 rats in each group, *t*-test, a*P* < 0.05 *vs* control group).



**Figure 7 Summary diagram of estrogen re-enhanced visceral hyperalgesia investigated in chronic prenatal stress plus chronic adult stress models.** BDNF: Brain-derived neurotrophic factor; NGF: Nerve growth factor.

**Table 1 Electrophysiological characteristics of dorsal horn neurons**

|  |  |  |  |  |
| --- | --- | --- | --- | --- |
| **Classification** | **Ctr., *n* = 36** | **CAS, *n* = 34** | **CPS, *n* = 29** | **CAS + CPS, *n* = 45** |
| Spontaneous spike number | 7.4 ± 4.0 | 16.9 ± 6.6 | 11.7 ± 4.8 | 44.8 ± 14.6c |
| Membrane capacitance (pF) | 72.1 ± 4.9 | 95.4 ± 5.8 | 85.1 ± 6.2 | 93.8 ± 8.9a |
| Action potential threshold (mV) | -27.1 ± 2.5 | -29.2 ± 1.8 | -34.6 ± 1.6 | -38.6 ± 1.4c |
| Resting membrane potential (mV) | -60.1 ± 1.7 | -50.1 ± 1.3 | -53.8 ± 1.5 | -56.4 ± 1.3 |
| Cell diameter (μm) | 32 ± 0.9 | 29 ± 0.6 | 31 ± 0.8 | 31 ± 0.6 |
| Time constant (μm) | 545.5 ± 51.1 | 737.7 ± 70.4 | 595.9 ± 54.1 | 535.3 ± 42.5 |
| Action potential amplitude (mV) | 79.0 ± 4.7 | 80.2 ± 3.9 | 77.4 ± 4.2 | 85.6 ± 3.7 |
| Duration (ms) | 8.38 ± 0.97 | 11.2 ± 0.95 | 12.3 ± 1.83 | 8.89 ± 0.60 |

Values means ± standard error. a*P* < 0.05; c*P* < 0.001 *vs* control group. CAS: Chronic adult stress; CPS: Chronic prenatal stress.

**Table 2 Electrophysiological characteristics of dorsal horn neurons after letrozole treatment**

|  |  |  |
| --- | --- | --- |
| **Classification** | **Veh. + CAS + CPS, *n* = 60** | **Let + CAS + CPS, *n* = 27** |
| Spontaneous spike number | 34.6 ± 11.3 | 8.04 ± 4.62c |
| Membrane capacitance (pF) | 86.8 ± 7.1 | 83.7 ± 6.7 |
| Action potential threshold (mV) | -34.6 ± 1.6 | -27.6 ± 2.1a |
| Resting membrane potential (mV) | -54.9 ± 1.3 | -54.6± 2.2 |
| Cell diameter (μm) | 31 ± 0.5 | 27 ± 1.6 |
| Time constant (um) | 502.7 ± 34.5 | 454.0 ± 36.6 |
| Action potential amplitude (mV) | 90.9 ± 3.7 | 89.8 ± 7.2 |
| Duration (ms) | 8.67 ± 0.63 | 10.70 ± 2.22 |

Values are means ± standard error. a*P* < 0.05; c*P* < 0.001 *vs* control group.CAS: Chronic adult stress; CPS: Chronic prenatal stress.