**Name of Journal:** *World Journal of Clinical Cases*

**Manuscript NO:** 64590

**Manuscript Type:** ORIGINAL ARTICLE

***Retrospective Study***

**Association of serum pepsinogen with degree of gastric mucosal atrophy in an asymptomatic population**

Cai HL *et al*. Pepsinogen for gastric mucosal atrophy evaluation

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**Author contributions:** Cai HL performed the research and wrote the paper; Tong YL designed the research, contributed to the analysis, and supervised the report.

**Supported by** National Natural Science Foundation of China, No. 71804161 and No. 72074188.

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**Received:** March 26, 2021

**Revised:** May 6, 2021

**Accepted:** September 27, 2021

**Published online:** November 6, 2021

**Abstract**

BACKGROUND

Atrophic gastritis is a precancerous lesion of the stomach. It has been reported that pepsinogen (PG) can reflect the morphology and function of the gastric mucosa, and it is therefore used as a marker for the early diagnosis of atrophic gastritis.

AIM

To evaluate the diagnostic value of serum PG for degree of gastric mucosal atrophy in asymptomatic Chinese upon physical examination.

METHODS

Medical data were collected from subjects who underwent transnasal gastroscopy between October 2016 and October 2018. For each study subject, serum PG levels and presence of *Helicobacter pylori* (*H. pylori*) infection were investigated. Pathology was evaluated using the Operative Link for Gastritis Assessment (OLGA) classification and Operative Link on Gastric Intestinal Metaplasia Assessment (OLGIM) systems. All statistical analyses were carried out using SPSS statistical software.

RESULTS

A total of 2256 subjects were enrolled and 1922 cases were finally included in the study. Based on the OLGA grading system, the levels of PGI were slightly decreased, while those of PGII were slightly increased. The PGI/PGII ratio (PGR) was reduced with increasing atrophy. The association between PG and OLGA grading was higher compared with that between PG and the OLGIM grading system. Compared with the OLGA-0 group, a statistically significant difference was observed in the mean age of OLGA-I, III, and IV groups (*P* < 0.05). In the *H. pylori*-positive subjects, the PGR levels were notably lower in the OLGA-I, II, and III groups compared with the OLGA-0 group (*P* <0.05). *H. pylori*-positive subjects exhibited significantly higher PGI and PGII serum levels and a significantly lower PGR compared with *H. pylori*-negative patients in different OLGA groups (*P* < 0.05).

CONCLUSION

Serum PG levels may represent a non-invasive screening marker for gastric mucosal atrophy in asymptomatic subjects.

**Key Words:** Pepsinogen; *Helicobacter pylori*; Operative Link for Gastritis Assessment; Atrophic gastritis; Gastric mucosal atrophy; Biomarker

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**Citation:** Cai HL, Tong YL. Association of serum pepsinogen with degree of gastric mucosal atrophy in an asymptomatic population. *World J Clin Cases* 2021; 9(31): 9431-9439

URL: <https://www.wjgnet.com/2307-8960/full/v9/i31/9431.htm>

DOI: https://dx.doi.org/10.12998/wjcc.v9.i31.9431

**Core Tip:** The current study evaluated the diagnostic value of serum pepsinogen (PG) as a screening marker for atrophic gastritis in asymptomatic healthy check-up populations in different regions of China. Serum PG levels were closely associated with Operative Link for Gastritis Assessment grading and could be used as an effective non-invasive screening tool for atrophic gastritis in asymptomatic subjects. In addition, better results could be obtained in *Helicobacter pylori*-positive individuals. Screening is more necessary in the elderly, and the application of the aforementioned screening tool may be beneficial for this population.

**INTRODUCTION**

In 2014, approximately 410400 new cases of gastric cancer (GC) were diagnosed in China, accounting for 10.79% of all cancer cases[1]. The prognosis of GC is closely associated with the times of diagnosis and treatment. The 5-year survival rate of advanced GC is less than 30%, while early GC, with a 5-year survival rate of over 90%, can be treated by endoscopy[2]. Therefore, improving the diagnostic rate of early GC is an efficient and feasible method to improve the survival rate of patients with GC.

GC follows the previously described Correa cascade of active gastritis-atrophic gastritis-intestinalization-intraepithelial neoplasia-GC, where atrophic gastritis is considered as the turning point and is referred to as a gastric precancerous lesion. Therefore, effective screening of subjects with gastric precancerous lesions and appropriate intervention can reduce the incidence of GC and increase the diagnostic rate of early GC.

In 2005, the International Atrophy Study Group proposed the Operative Link for Gastritis Assessment (OLGA) grading and staging system for chronic gastritis[3]. This staging system represents the extent and degree of gastric mucosal atrophy to link the histopathology of chronic gastritis to the risk of GC. In terms of pathological diagnosis, the consistency of gastric mucosal atrophy diagnosis among different physicians is low, while that of intestinal metaplasia is high. Therefore, in 2010, the standard Operative Link on Gastric Intestinal Metaplasia Assessment (OLGIM) staging system put forth the proposal to replace the term “atrophy” with “intestinal metaplasia”[4]. Several studies have now confirmed that the OLGA/OLGIM staging system, reflecting the severity of atrophic gastritis and the risk of GC, facilitates the identification of patients with a high risk of developing GC (OLGA/OLGIM stages III and IV), thus promoting the early diagnosis and prevention of the disease[5,6].

Endoscopy is an invasive diagnostic approach that requires a large number of samples, and its diagnostic accuracy is affected by the practicing pathologists themselves. Therefore, endoscopy is not considered as an ideal method in clinical practice, particularly for the physical examination of healthy individuals. Nevertheless, screening for atrophy and risk of GC in healthy individuals using non-invasive approaches is of great importance.

Serum levels of pepsinogen (PG) can reflect the morphology and function of the gastric mucosa. PGI reflects the function of gastric acid secretion of the gastric gland. Therefore, it has been reported that the levels of PGI are reduced when gastric mucosal glands are atrophied. Additionally, the PGII serum levels are mainly associated with the extent of lesions in the gastric mucosa of the gastric fundus. Increased PGII serum levels are associated with gastric fundus atrophy, intestinal metaplasia, or pseudopyloric metaplasia and dysplasia. Several studies have demonstrated that the PGI/PGII ratio (PGR) was associated with progressive gastric mucosal atrophy and GC[7,8].

The current study aimed to evaluate the diagnostic value of the serum PG levels and the presence of *Helicobacter pylori* (*H. pylori*) infection as a screening tool for atrophic gastritis in asymptomatic Chinese who attend standard health check-up in China.

**MATERIALS AND METHODS**

***Study population***

A total of nine health management centers from different areas of China participated in the study, which was conducted between October 2016 and October 2018. The centers were the No. 924 Hospital of the People's Liberation Army of China (southern China), the Second Affiliated Hospital of Zhejiang University College of Medicine, the Zhongshan Affiliated Hospital of Xiamen University, the Kunshan Hospital of Traditional Chinese Medicine (all in eastern China), the Sichuan Provincial People’s Hospital, the Southwest Affiliated Hospital of the Third Military Medical University (both in southwest China), the Jilin City People’s Hospital (northeast China), the PLA General Hospital, and the Jingzhou Hospital of Traditional Chinese Medicine (both in central-northern China). More specifically, subjects between 20-80 years of age, without gastrointestinal symptoms, and who presented for asymptomatic health check-up that included transnasal endoscopy were enrolled in the present study.

Participants with one of the following symptoms or clinical findings were excluded from the present study: (1) Subjects with contraindications to transnasal gastroscopy; (2) Previous history of definitive benign or malignant diseases of the upper gastrointestinal tract, including peptic ulcers, gastric polyps, esophageal cancer, and GC, or history of surgery; (3) Treatment with acid suppressants within the past month; (4) Serious organic diseases, including heart, liver, and kidney diseases; (5) Severe mental illness precluding an ability to cooperate; or (6) Current pregnancy or lactation.

***Determination of serum PG levels, H. pylori test, and endoscopy***

All participants received serum PG testing, *H. pylori* testing, and transnasal endoscopy in a single-day hospital visit. Approximately 5 mL of fasting blood was collected from each subject. Serum PG levels were measured by particle-based chemiluminescence immunoassay using the PGI and PGII kits (Abbott Laboratories Inc., Chicago, IL, United States). *H. pylori* infection was evaluated using a 13C-urea breath test (Shenzhen Zhonghe Headway Bio-Sci & Tech Co., Ltd., Shenzhen, China) or serum *H. pylori* antibodies (MP Biomedicals, Santa Ana, CA, United States) in combination with pathological *H. pylori* findings. Individuals with a positive result for any of the aforementioned three tests were categorized into a *H. pylori*-positive group, while those with a negative result for all tests were categorized into a *H. pylori*-negative group. Endoscopy was performed by experienced endoscopists who were blinded to the *H. pylori* and PG test results. Pathological samples were obtained from the gastric body, angulus, and antrum according to the Chinese Consensus on Screening and Endoscopic Diagnosis and Treatment of Early GC (2014 version)[9]. Additional biopsies were performed at the sites where lesions were detected.

***Statistical analysis***

The data are expressed as the mean ± standard deviation for normally distributed data and as the median ± interquartile range for non-normally distributed data. Statistical analyses were performed using SPSS statistical software (version 20; IBM Corp., Armonk, NY, United States). Age, PGI and PGII levels, and PGR were tested for normal distribution by the Kolmogorov-Smirnov test. After confirming a normal distribution, ANOVA with *post hoc* Scheffe’s test was used for further assessment. Bonferroni’s correction and Pearson’s chi-square test were used to evaluate the differences between *H. pylori*-negative and *H. pylori*-positive patients. Receiver operating characteristic (ROC) curve analysis was used to estimate the cutoff values for PGI and PGR. *P*-values less than 0.05 were considered statistically significant.

**RESULTS**

Among the total 2256 subjects included in the current study, 14 were diagnosed with GC (0.6%), including six cases of early GC. In addition, one subject was diagnosed with pharyngeal cancer. There were 326 cases (15.9%) of atrophic gastritis and 391 cases (19.0%) of intestinal metaplasia, while the total *H. pylori* infection rate was 41.7%. A total of 1922 subjects, with an average age of 52.3 ± 9.8 years and male to female ratio of 1.2:1 (1065/857), underwent serological, gastroscopic, and pathological examinations (Table 1).

Based on OLGA grading, the PGI serum levels were slightly decreased, while those of PGII were slightly increased. In addition, the PGR was reduced in conjunction with increasing degrees of atrophy. Compared with the OLGA-0 group, the PGII levels were slightly increased and PGR was slightly decreased in the OLGA-I to IV groups (*P* < 0.05). Additionally, compared with the OLGA-I and II groups, PGR levels in the OLGA-III and IV groups were significantly reduced (*P* < 0.05). A statistically significant difference was also observed in the PGR between the OLGA-II and OLGA-III/IV groups (*P* < 0.05). A notable difference in PGR was also identified between the GC group and both OLGA-0 and I groups (*P* < 0.05; Table 2).

Furthermore, as shown in Table 2, compared with the OLGIM-0 group, the PGI levels were significantly lower in the OLGIM-II, III, and IV groups (*P* < 0.05), and PGR was reduced in the OLGIM-II and III groups (*P* <0.05). The PGI serum levels were significantly lower in the OLGIM-III/IV groups, and PGR was also markedly decreased in the OLGIM- II/ III groups compared with the OLGIM-I group (*P* < 0.05). Finally, compared with the OLGIM-II group, the OLGIM-III group exhibited significantly reduced PGR (*P* <0.05; Table 3).

The aforementioned results indicated that the PG serum levels were more relevant to OLGA grade compared with the OLGIM grade. Therefore, the OLGA grading system was applied for the subsequent analyses.

Rugge *et al*’s[10] prospective study confirmed that the high-risk stage (defined as stage III or IV by the OLGA classification) is closely associated with high risk of GC. Accordingly, OLGA I and II groups were then combined in a low-risk group and compared with the OLGA-0 group. The best cutoff for low risk was estimated at PGI ≤ 73.14 ng/mL [area under the curve (AUC) = 0.585, sensitivity = 62.1%, specificity = 53.8%] and PGR ≤ 11.54 (AUC = 0.611, sensitivity = 43.2%, specificity = 77.7%) (Figure 1). The OLGA III and IV groups were combined in a high-risk group and compared with the remaining three groups; the best cutoff for high risk was estimated at PGI ≤ 64.00 ng/mL (AUC = 0.631, sensitivity = 67.2%, specificity = 61.2%) and PGR ≤ 9.11 (AUC = 0.740, sensitivity = 53.0%, specificity = 91.8%) (Figure 2).

Compared with the OLGA-0 group, the mean ages in the OLGA-I, III, and IV groups were significantly higher (*P* <0.05) and increased with an increasing OLGA grade. In addition, a statistically significant difference was observed in the *H. pylori* positivity rate between the OLGA-I/II/III/IV groups and the OLGA-0 group (*P* <0.05; Table 4).

Further analyses were performed with the *H. pylori*-negative or positive groups. In the OLGA-0 group, *H. pylori* positive subjects exhibited significantly higher PGI and PGII serum levels and a significantly lower PGR compared with *H. pylori*-negative patients (*P* < 0.05). The same trend was observed among *H. pylori*-positive/*H. pylori*-negative subjects in the OLGA-I group (*P* < 0.05). Additionally, *H. pylori*-positive subjects in the OLGA-II group had significantly elevated PGI and PGII levels compared with the *H. pylori*-negative subjects (*P* < 0.05). In the OLGA-III group, significantly increased serum levels of PGII were observed in the *H. pylori*-positive group compared with the *H. pylori*-negative group, while the ratio PGI/PGII was notably reduced. By contrast, no statistically significant differences were obtained in the PG levels between the *H. pylori*-positive/*H. pylori*-negative subjects in the OLGA-IV and GC groups (*P* > 0.05).

In the *H. pylori*-positive population, compared with the OLGA-0 group, both PGI levels and PGR were significantly lower in the OLGA-I and III groups (*P* < 0.05), and PGR was also notably reduced lower in the OLGA-II group (*P* < 0.05). In the *H. pylori*-negative population, the serum levels of PGI and PGR were markedly decreased in the OLGA-I/II (*P* < 0.05) and GC groups (*P* < 0.05; Table 5).

**DISCUSSION**

Atrophic gastritis is considered as one of the main precursor lesions in the Correa cascade, which may eventually result in GC. A study reported that the odds of developing GC 5 years after the diagnosis of atrophic gastritis, enterocolitis, mild to moderate atypical hyperplasia, and severe atypical hyperplasia were 0.1%, 0.25%, 0.6%, and 6%, respectively[1].

The current study found that approximately 15.9% of the asymptomatic individuals undergoing standard physical examination exhibited atrophic gastritis and 19.0% exhibited enterocolitis. Among these patients, 2.5% suffered from severe atrophic gastritis (OLGA grade III or IV). These patients have a higher risk of progression to GC after 5 years. Therefore, effective screening of such individuals in the asymptomatic population *via* endoscopic surveillance interventions are important for reducing the incidence of GC and improving the diagnostic rate of early GC.

Serum PG levels reflect the morphological and functional status of the gastric mucosa. It has been reported that the gradual decrease of PGR is associated with the progression of gastric mucosal atrophy and GC[2]. A meta-analysis of 31 studies, including 1520 patients with GC and 2265 with atrophic gastritis, suggested that serum PG levels could be considered as a potent, non-invasive, population-based screening tool for the diagnosis of GC and atrophic gastritis[11]. Additionally, a study by Zoalfaghari *et al*[12] indicated that the serum PGI levels and PGR could be potential serological markers for the diagnosis of atrophic gastritis, with a high sensitivity and specificity. The present study also demonstrated that PGI levels and PGR were significantly decreased with increasing atrophy, with PGR showing more significant diagnostic value. The association between PG levels and OLGA grade was more pronounced compared with the OLGIM grading system, suggesting that PG could be more closely associated with atrophy compared with intestinal chemosis. Furthermore, ROC curve analysis showed that serum PGI levels and PGR could be used to predict atrophy and intestinal metaplasia of the gastric mucosa in asymptomatic health check-up subjects in China.

Several regional studies have demonstrated that the *H. pylori* infection rate is approximately 40%-55% in China.The overall *H. pylori* infection rate in the study population was 39.39%. The results showed that the PGI and PGII serum levels were significantly increased, and PGR was markedly decreased in the *H. pylori*-infected group. *H. pylori* infection also affected the levels of PG in different pathological subgroups[13]. Al-Ezzy *et al*[14] reported that *H. pylori* infection can affect the expression of the *Fas* gene through inflammatory factors, thus regulating PG concentration.

Furthermore, the results of the current study also demonstrated that patients with high-grade lesions were older and had a higher rate of *H. pylori* infection. A positive association has been reported between the severity of gastric atrophy or intestinal metaplasia and the age of the subjects[15,16]. Additionally, a study showed that the prevalence of high-risk OLGA stage was increased with increasing age in *H. pylori*-negative subjects[17]. A Korean study demonstrated that advanced age, long-term smoking, and *H. pylori* infection were independent risk factors associated with advanced stages determined by the OLGA staging system[18]. The transition from *H. pylori* infection into GC is a multistep process, including the progression of chronic gastritis into precancerous lesions and ultimately to GC[19]. The presence or absence of *H. pylori* infection allows a preliminary prediction of whether a patient is at high risk by OLGA staging. Therefore, it has been suggested that routine endoscopy may not be necessary for all patients by OLGA staging or those at a low-risk by OLGIM stage, since *H. pylori* eradication can effectively prevent the development of GC. Effectively eradicating *H. pylori* before the age of 40 years could reduce the number of routine endoscopies in individuals requiring GC surveillance and substantially reduce the related healthcare burden.

The current study has some limitations in. First, the overall sample size in the study was not large enough; in particular, the number of patients with OLGA stage III/IV and GC was insufficient, which could cause statistical bias. Therefore, further studies with increased sample size are needed to verify the results of the present study. Second, there could be a selection bias in this study, since all enrolled subjects voluntarily underwent physical examination and gastroscopy, suggesting that their economic status was moderate.

**CONCLUSION**

The present study demonstrated that serum PG levels were closely associated with OLGA classification. The results suggested that PG levels could be used as an effective non-invasive screening tool for atrophic gastritis in the asymptomatic population undergoing standard physical examination, while the diagnostic value of PG levels could be more potent in patients with *H. pylori* infection. This screening tool could be valuable and represent an important screening method for the elderly population.

**ARTICLE HIGHLIGHTS**

***Research background***

Atrophic gastritis is a precancerous lesion of the stomach. Pepsinogen (PG) has been reported to reflect the morphology and function of the gastric mucosa.

***Research motivation***

PG can be used for non-invasive screening of atrophic gastritis and even gastric cancer (GC). Effective screening of subjects with gastric precancerous lesions and appropriate intervention can reduce the incidence of GC and increase the diagnostic rate of early GC.

***Research objectives***

The main objective of this research was to evaluate the diagnostic value of serum PG in the degree of gastric mucosal atrophy in an asymptomatic Chinese population undergoing standard physical examination.

***Research methods***

The study subjects underwent transnasal gastroscopy, and serum PG levels and the presence of *Helicobacter pylori* (*H. pylori*) infection were investigated to assess the diagnostic accuracy of PG for evaluating the degree of gastric mucosal atrophy. Pathology was evaluated using the Operative Link for Gastritis Assessment (OLGA) classification and Operative Link on Gastric Intestinal Metaplasia Assessment (OLGIM) systems. ANOVA with *post hoc* Scheffe’s test was used for further assessment of the differences in age, PGI and PGII levels, and PGI/PGII ratio (PGR). Bonferroni’s correction and Pearson’s chi-square test were used to evaluate the differences between *H. pylori*-negative and *H. pylori*-positive patients. Receiver operating characteristic curve analysis was used to estimate the cutoff values for PGI, PGII, and PGR.

***Research results***

The association between PG and OLGA grade was higher compared with that between PG and the OLGIM grading system. Based on the OLGA grading system, the levels of PGI were slightly decreased, while those of PGII were slightly increased. PGR was reduced with increasing atrophy (*P* < 0.05). A slightly increasing trend was observed in the mean age of different OLGA groups. *H. pylori*-positive subjects exhibited significantly higher PGI and PGII serum levels and a significantly lower PGR compared with *H. pylori*-negative patients in the different OLGA groups (*P* < 0.05).

***Research conclusions***

Serum levels of PG are closely associated with OLGA stage and could be used as an effective non-invasive screening tool for evaluating the degree of gastric mucosal atrophy in asymptomatic subjects.

***Research perspectives***

Future studies could focus on the cutoff values of PG for the diagnosis of precancerous lesions or early gastric cancer in different regions of China

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**Footnotes**

**Institutional review board statement:** This study was reviewed and approved by the Human Research Ethics Committee of the Second Hospital Affiliated to Zhejiang University School of Medicine (No. 2015-079).

**Informed consent statement:** All subjects provided written informed consent prior to enrollment in this study.

**Conflict-of-interest statement:** The authors declare that they have no related financial relationships to disclose.

**Data sharing statement:** No additional data are available.

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**Manuscript source:** Unsolicited manuscript

**Peer-review started:** March 26, 2021

**First decision:** April 28, 2021

**Article in press:** September 27, 2021

**Specialty type:** Gastroenterology and hepatology

**Country/Territory of origin:** China

**Peer-review report’s scientific quality classification**

Grade A (Excellent): 0

Grade B (Very good): B

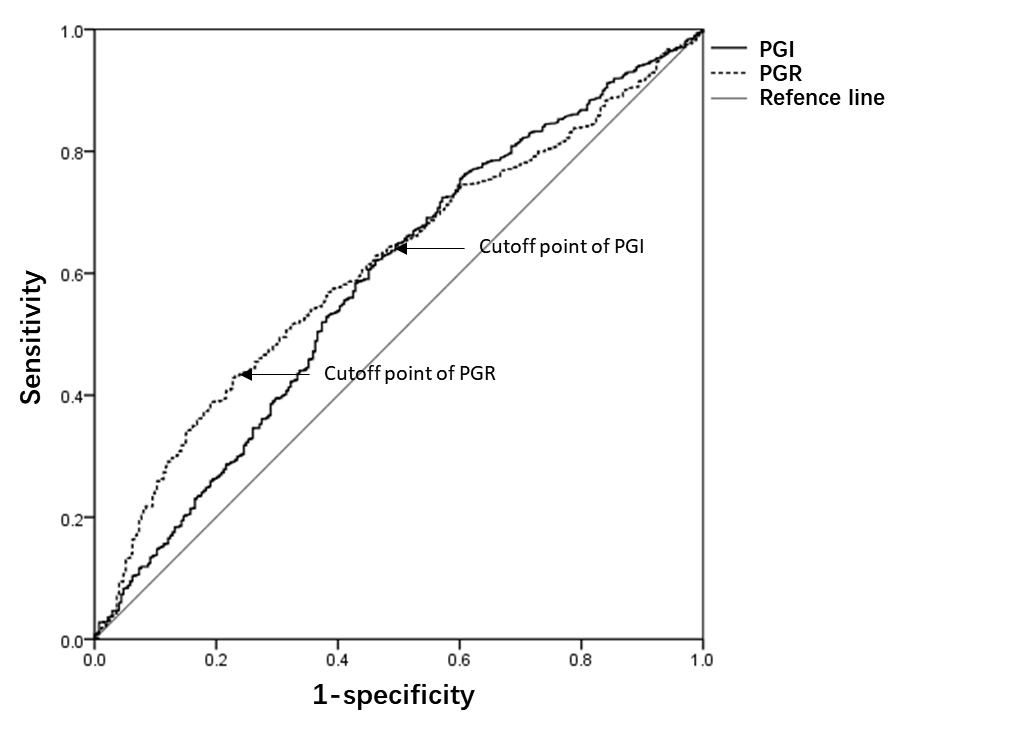
Grade C (Good): C, C

Grade D (Fair): D

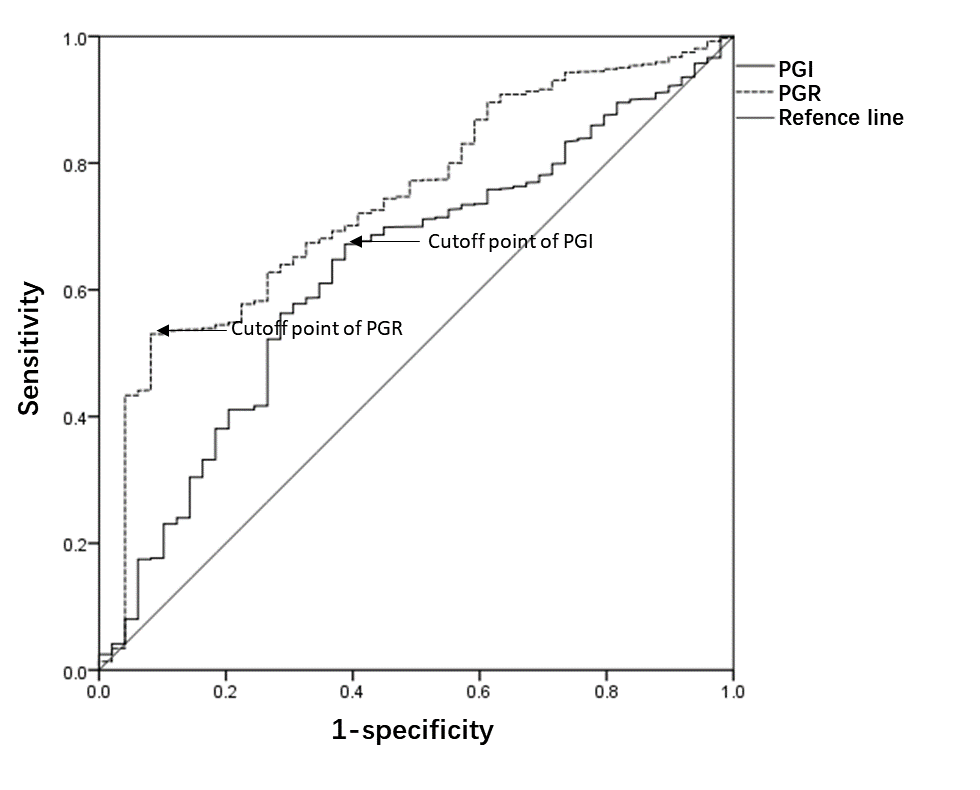
Grade E (Poor): 0

**P-Reviewer:** Cho JH, Mohammadi M, Yücel O **S-Editor:** Liu M **L-Editor:** Wang TQ **P-Editor:** Liu M

**Figure Legends**



**Figure 1 Operating characteristics curve analysis of pepsinogen I and pepsinogen I/pepsinogen II ratio for the low-risk group.** TheOperative Link for Gastritis Assessment (OLGA) I and II groups were combined as the low-risk group and compared with the OLGA-0 group. PG: Pepsinogen; PGR: Pepsinogen I/pepsinogen II ratio; OLGA: Operative Link for Gastritis Assessment.



**Figure 2 Operating characteristics curve analysis of pepsinogen I and pepsinogen I/pepsinogen II ratio for the high-risk group.** TheOperative Link for Gastritis Assessment (OLGA) III and IV groups were combined as the high-risk group and compared with OLGA 0, I, and II groups. PG: Pepsinogen; PGR: Pepsinogen I/pepsinogen II ratio; OLGA: Operative Link for Gastritis Assessment.

**Table 1 Baseline characteristics of the 1922 subjects involved in the study, *n* (%)**

|  |  |
| --- | --- |
| **Age** | **52.3 ± 9.8** |
| Gender (male) | 1065 (55.41) |
| *Helicobacter pylori* | 757(39.39) |
| Gastric cancer | 10 (0.52) |

**Table 2 Serum pepsinogen levels in different OLGA groups**

|  |  |  |  |  |
| --- | --- | --- | --- | --- |
| **OLGA grade** | ***n*** | **PGI (ng/mL)** | **PGII (ng/mL)** | **PGR** |
| 0 | 1590 | 90.4 ± 82.2 | 8.3 ± 8.6 | 10.1 ± 9.3 |
| I | 185 | 68.7 ± 71.21 | 7.8 ± 8.5 | 8.1 ± 5.61 |
| II | 88 | 73.7 ± 80.71 | 9.8 ± 8.8 | 7.5 ± 4.61 |
| III | 43 | 60.8 ± 51.11 | 11.7 ± 12.81,2 | 6.7 ± 4.41,2,3 |
| IV | 6 | 52.1 ± 37.31 | 15.5 ± 9.9 | 4.1 ± 3.71,2,3 |

1*P* < 0.05, compared with the OLGA-0 group.

2*P* < 0.05, compared with the OLGA-I group.

3*P* < 0.05, compared with the OLGA-II group.

OLGA: Operative Link for Gastritis Assessment; PG: Pepsinogen; PGR: Pepsinogen I/pepsinogen II ratio.

**Table 3** **Serum pepsinogen levels in different OLGIM groups**

|  |  |  |  |  |
| --- | --- | --- | --- | --- |
| **OLGIM grade** | ***n*** | **PGI (ng/mL)** | **PGII (ng/mL)** | **PGR** |
| 0 | 1522 | 91.1 ± 83.8 | 8.5 ± 8.9 | 9.9 ± 8.8 |
| I | 252 | 79.2 ± 80.71 | 8.1 ± 8.4 | 9.0 ± 8.3 |
| II | 89 | 66.4 ± 76.51 | 8.8 ± 7.6 | 7.3 ± 6.31,2,3 |
| III | 41 | 56.3 ± 45.11,2 | 9.5 ± 6.72 | 6.3 ± 4.11,2,3 |
| IV | 8 | 60.8 ± 26.41,2 | 8.6 ± 11.7 | 6.7 ± 18.8 |

1*P* < 0.05, compared with the OLGIM-0 group.

2*P* < 0.05, compared with the OLGIM-I group.

3*P* < 0.05, compared with the OLGIM-II group.

OLGA: Operative Link for Gastritis Assessment; PG: Pepsinogen; PGR: Pepsinogen I/pepsinogen II ratio.

**Table 4 Age, sex, and presence of *Helicobacter pylori* infection in different Operative Link for Gastritis Assessment groups, *n* (%)**

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
| **OLGA grade** | **0** | **I** | **II** | **III** | **IV** |
| Sex (male) | 866 (54.5) | 109 (58.9) | 57 (64.8) | 25 (58.1) | 3 (50.0) |
| Age | 51.9 ± 9.9 | 54.99 ± 9.11 | 53.49 ± 8.8 | 55.42 ± 9.91 | 56.33 ± 8.21 |
| *Helicobacter pylori*-positive | 583 (36.7) | 86 (46.5)1 | 51 (60.0)1 | 27 (62.8)1 | 5 (83.3)1 |

1*P* < 0.05, compared with the OLGA-0 group.

OLGA: Operative Link for Gastritis Assessment.

**Table 5** **Serum pepsinogen levels in the OLGA groups after stratification according to the presence of *Helicobacter pylori* infection**

|  |  |  |  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- |
|  | **OLGA-0** | | **I** | | **II** | | **III** | | **IV** | |
|  | *Helicobacter pylori*-(*n* = 1007） | *Helicobacter pylori* + (583) | *Helicobacter pylori* - (*n* = 99 | *Helicobacter pylori*+ (*n* = 86) | *Helicobacter pylori*- (*n* = 37) | *Helicobacter pylori*+ (*n* = 51) | *Helicobacter pylori*- (*n* = 16 | *Helicobacter pylori*+ (*n* = 27) | *Helicobacter pylori*-(*n* = 1) | *Helicobacter pylori*+ (*n* = 5) |
| PGI (ng/mL) | 82.18 ± 78.64 | 101.70 ± 89.011 | 56.71 ± 66.102 | 73.05 ± 78.201,2 | 55.40 ± 54.182 | 92.12 ± 73.511 | 60.00 ± 47.55 | 60.8 ± 54.302 | 34.3 | 63.50 ± 43.882 |
| PGII (ng/mL) | 7.35 ± 6.61 | 10.80 ± 11.361 | 6.60 ± 6.31 | 10.05 ± 10.231 | 7.10 ± 4.88 | 13.00 ± 10.491 | 7.00 ± 6.13 | 13.5 ± 10.641 | 3.8 | 16.40 ± 8.70 |
| PGR | 10.68 ± 8.76 | 8.82 ± 9.891 | 8.77 ± 5.072 | 7.24 ± 6.281,2 | 7.94 ± 3.542 | 7.18 ± 5.512 | 8.32 ± 3.25 | 4.50 ± 3.451,2 | 9.026316 | 3.90 ± 2.652 |

1*P* < 0.05, within the same pathological subgroup, compared with the *Helicobacter pylori*- group.

2*P* < 0.05, within the same *Helicobacter pylori* subgroup, compared with the OLGA-0 group.

OLGA: Operative Link for Gastritis Assessment; PG: Pepsinogen; PGR: Pepsinogen I/pepsinogen II ratio.



Published by **Baishideng Publishing Group Inc**

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