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**Gut microbiome in non-alcoholic fatty liver disease associated hepatocellular carcinoma: Current knowledge and potential for therapeutics**

Said I *et al.* Gut microbiome in NAFLD HCC

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**Abstract**

Metabolic diseases such as nonalcoholic fatty liver disease (NAFLD) are rising in incidence and are an increasingly common cause of cirrhosis and hepatocellular carcinoma (HCC). The gut microbiome is closely connected to the liver *via* the portal vein, and has recently been identified as a predictor of liver disease state. Studies in NAFLD, cirrhosis and HCC have identified certain microbial signatures associated with these diseases, with the disease-associated microbiome changes collectively referred to as dysbiosis. The pathophysiologic underpinnings of these observations are an area of ongoing investigation, with current evidence demonstrating that the gut microbiome can influence liver disease and carcinogenesis *via* effects on intestinal permeability (leaky gut) and activation of the innate immune system. In the innate immune system, pathogen recognition receptors (Toll like receptors) on resident liver cells and macrophages cause liver inflammation, fibrosis, hepatocyte proliferation and reduced antitumor immunity, leading to chronic liver disease and carcinogenesis. Dysbiosis-associated changes include increase in secondary bile acids and reduced expression of FXR (nuclear receptor), which have also been associated with deleterious effects on lipid and carbohydrate metabolism associated with progressive liver disease. Longitudinal experimental and clinical studies are needed in different populations to examine these questions further. The role of therapeutics that modulate the microbiome is an emerging field with experimental studies showing the potential of diet, probiotics, fecal microbiota transplantation and prebiotics in improving liver disease in experimental models. Clinical studies are ongoing with preliminary evidence showing improvement in liver enzymes and steatosis. The microbial profile is different in responders to cancer immunotherapy including liver cancer, but whether or not manipulation of the microbiome can be utilized to affect response is being investigated.

**Key Words:** Microbiome; Gut microbiome; Hepatocellular carcinoma; Nonalcoholic fatty liver disease; Pathophysiology; Treatment

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**Core Tip:** The gut microbiome is intimately linked to nonalcoholic fatty liver disease, cirrhosis and hepatocellular carcinoma. The breakdown of the intestinal barrier in liver disease, innate immune system stimulation and bile acid profile changes are increasingly found in association with these diseases. Manipulation of the microbiome by diet, probiotics, prebiotics and other agents is a promising area of investigation.

**INTRODUCTION**

Liver cancer (hepatocellular carcinoma, HCC) is the seventh commonest cancer worldwide and the third commonest cause of cancer related mortality accounting for over 800000 deaths in 2020[1]. Over the past 4 decades in the US there has been a 4 fold increase in HCC incidence in the US[2].

Liver cancer is most commonly seen in association with cirrhosis of the liver as well as chronic hepatitis B (HBV) infection without cirrhosis[3]. Common causes of cirrhosis include non-alcoholic and alcoholic fatty liver disease, hepatitis C and hepatitis B infection as well as autoimmune and biliary diseases. Well known risk factors for liver cancer in cirrhosis and chronic HBV include male sex, smoking, alcohol excess, aflatoxin (rare), viral load in HBV, and metabolic factors such as diabetes and obesity[4].

Although the prevalence of hepatitis B and C are decreasing globally, liver cancer rates have increased due to the rise in cases of obesity, nonalcoholic fatty liver disease (NAFLD) and type 2 diabetes, largely fueled by a poor Western diet[5]. NAFLD related liver cancer is the fastest growing cause of liver cancer and related mortality in the US[6].

In NAFLD related HCC, factors such as age, genetic predisposition, diabetes and obesity have been found in association with the development of NAFLD-related HCC[6]. In NAFLD a significant minority of liver cancers (10%-15%) can occur even in the absence of cirrhosis and has been linked to the underlying liver inflammation, fibrosis with increased risk in diabetics[6]. The gut microbiome has been proposed as a leading risk factor associated with liver cancer. In obesity related metabolic diseases the microbial profile of the intestine has been linked to progressive liver disease and carcinogenesis both in experimental models and in human studies[7].

**The Gut Microbiome and NAFLD-HCC**

***The gut microbiome***

The gut microbiome refers to a multispecies community of resident microbes that includes a wide variety of bacteria, fungi, viruses as well as archaea, residing in the gut[8]. Nearly 100 trillion microbium occupy the intestinal tract particularly in the large intestine. Although small intestinal microbiota also exist this is a less well studied area compared to the large intestine. Most of the research in the human microbiome has been done on bacterial stool microorganisms which are a reasonable approximation of the intestinal microbiome. Studies of the microbiome in intestinal biopsies have been done to a lesser extent and there may be qualitative and quantitative differences in measuring the microbiome adherent to the mucosa *vs* present in stool.

The gut microbiome exhibits many benefits of commensalism for the host and plays an important role in regulating host immunity beginning in utero, maintaining a mucosal defense against pathogens, facilitating nutrient metabolismincluding assistance in digestion and as a prominent source of key vitamins and energy harvest[9]. The microbiome also plays a critical role in the pathogenesis of metabolic diseases, inflammatory and autoimmune conditions both within the gastrointestinal tract and in remote sites[10].

***The gut microbiome and liver disease***

The liver receives the majority of its blood supply through the portal vein and is exposed to the microbiome either directly through microbial translocation or *via* microbial metabolites and products[11]. The homeostasis between gut microbes and host is mediated by an intact barrier function (tight junction) of colonic epithelial cells, thick mucus layer as well as IgA and antimicrobial surface peptides, achieved by interaction of the microbes and pathogen recognition receptors that promote a healthy tolerogenic immune response allowing symbiosis[12-14]. This exposure to the microbiome has a critical role in development of a normal immune response through priming and modulation of the immune response in the gut mucosa and the liver. This is exemplified in experiments in knockout mice lacking aspects of the innate immune system (*e.g.,* TLR5) in which dysbiosis has been reported[14].

Emerging evidence shows that in liver disease including metabolic fatty liver disease (NAFLD), cirrhosis and liver cancer, the microbiome varies significantly from the microbiome in healthy individuals both compositionally as well as functionally[11]. These diseases associated changes in the microbiome are referred to as dysbiosis and have been associated with metabolic liver disease and liver cancer in both animal experiments and in human studies. Dysbiosis, is strongly linked to fatty liver disease, type 2 diabetes and other metabolic disease[14]. The presence/absence of certain microorganisms can allow for identification of the severity of liver disease (serving as a diagnostic signature) and potentially guide emerging therapies. *Escherichia coli* (E. coli) is enriched in the gut of NAFLD patients with more advanced fibrosis and HCC[15] and Bacteroides bacteria were found in higher concentrations in cirrhotic patients with HCC patients as compared to cirrhotic patients without HCC[16].

In experimental models, antibiotics and gut sterilization can reduce the prevalence of HCCs in obese mice suggesting that microbiota dysbiosis plays a crucial role in the pathogenesis of HCC[17].

**PATHOPHYSIOLOGY OF GUT DYSBIOSIS IN LIVER DISEASE AND LIVER CANCER**

***Leaky gut, endotoxemia, innate immune system and the inflammatory response***

The intestinal microbiome in liver disease is potentially influenced by the liver disease itself, and in turn the intestinal microbiome can also influence the progression of liver disease.

In cirrhosis the underlying changes of portal hypertension influence intestine transit and permeability resulting in the so called “leaky gut” seen in cirrhosis[7]. This increased intestinal permeability allows increased passage of bacterial products, metabolites and bacteria *via* the portal vein to the liver resulting in endotoxinemia. Bacterial cell wall components such as lipopolysaccharide (LPS) from *Gram negative bacteria* and lipoteichoic acid (LTA) from *Gram positive bacteria* (also referred to as PAMPs or pathogen associated molecular pattern) are increased in the circulation in patients with increasing degree of advanced liver disease and in animal models of liver disease[7,18]. Measurements of LPS in portal vein, mean portal vein LPS levels increased in chronic liver injury from < 3 pg/mL in healthy volunteers to 4.9 pg/mL, 7.9 pg/mL and 10.2 pg/mL in patients with Child–Turcotte-Pugh cirrhosis stage A, B and C respectively[19].

In dysbiosis the host-microbiota balance is lost and the delivery of PAMPs like LPS and LTA *via* the portal vein, outside of the intestine where they exist in a symbiotic relationship with the host, is associated with activation of the innate immune systems *via* Pattern recognition receptors (such as Toll like receptors, TLR 4, 5 and TLR 9) found on the liver resident cells (hepatocytes, stellate cells) and liver resident macrophages (Kupffer cells)[11]. Activation of the immune response results in cytokine and chemokine expression and the recruitment of inflammatory cells in the liver, hepatocyte proliferation as well as hepatic stellate activation which result in progressive liver inflammation, fibrosis and liver cancer[7,19,20] (Figure 1).

Endotoxemia directly promotes chronic inflammation in the intestine as well as systemically in the liver, adipose tissue and vasculature though activation of cytokine and cell mediated pathways. This increases the risk of metabolic complications such as atherosclerosis, diabetes and nonalcoholic fatty liver disease which are common concurrent conditions[8,14]. The presence of NAFLD and liver diseases further impairs the ability of the liver to deal with gut derived endotoxins arriving *via* the portal vein. Chronic liver conditions such as NAFLD, particularly in the presence of diabetes are an increasing cause of liver cancer[6]. NAFLD patients with cirrhosis with HCC have an enhanced intestinal inflammatory status compared to those without HCC and healthy subjects, as demonstrated by the increased fecal calprotectin concentration[16]. Increased intestinal permeability, intestinal bacterial overgrowth and elevated serum endotoxin, all have been reported in NAFLD and NAFLD-HCC[7].

In experimental (mouse) models of liver cancer, TLR4 and the intestinal microbiota were not required for HCC initiation but are critical for HCC promotion. Activation of this innate immune system promotes liver cell proliferation *via* increased levels of the hepatomitogen epiregulin, and prevention of apoptosis (Figure 1). Gut sterilization restricted to late stages of hepatocarcinogenesis reduced HCC in this model. Germ-free status or TLR4 inactivation also reduce HCC by 80%-90% further attesting to the importance of the microbiome in carcinogenesis[17].

Intestinal permeability is increased in patients with compensated liver cirrhosis, regardless of the presence of HCC. A comparison of the patterns of cytokine and chemokine plasma levels between NAFLD related HCC in cirrhosis and NAFLD cirrhosis without HCC observed a specific inflammatory milieu in the HCC group. IL8, IL13, CCL3, CCL4, and CCL5 were significantly increased in the presence of HCC and, their plasma levels correlated with circulating activated monocytes and monocytic myeloid derived suppressor cells (mMDSCs). Activation of TLR4 by LPS is one of the most important inflammatory stimulations that can enhance the expression of CCL3, CCL4, and CCL5 by hepatic stellate cells (HSC) and monocytes[16].

LPS–TLR4 interaction also plays a role in hepatocarcinogenesis. IL8 has been associated with HCC development, tumor burden, and prognosis, similar to what has been reported for CCL2, CCL3, and CCL5. in mouse models the activation of HSC through TLR4 Leads to the secretion of CXCL1, the homologue of human IL8, causing neutrophil recruitment to the liver. Similar mechanisms have been postulated to promote mMDSC recruitment to the liver, favoring HCC progression[21].

LTA from Gram positive bacteria enhances the production of a Senescence associated secretory phenotype (SASP) of HSC in conjunction with an obesity induced gut microbial metabolite, deoxycholic acid (DCA) (secondary bile acid increased in the presence of dysbiosis in obesity). Cellular senescence a relatively recently described phenomenon is a complex process whereby senescent cells can induce cell cycle arrest as well as involve the secretion of factors that can result in tissue inflammation, repair and regeneration and affect the behavior of neighboring cells[22].

The phenotype of senescent cells involves secretion of a series of inflammatory cytokines, chemokines, matrix-remodeling factors, and growth factors. Whereas in early life and development the SASP may have anticancer effects (through cell cycle arrest) it has also been associated with the biology of aging, chronic inflammation and carcinogenesis in chronic conditions such as NAFLD. In one study of Steatohepatitis associated HCC increased production of senescence associated secretory factors like IL-6 were expressed by Fibroblasts in steatohepatitis associated cancers and non-tumoral stellate cells compared to convention HCCs)[23]. HCC development is promoted by this induction of cellular senescence and SASPin HSC in the tumor microenvironment. Corroborating these hypotheses, animal models of HCC demonstrate that vancomycin treatment significantly reduced obesity-associated liver cancer development. However, vancomycin treatment with DCA plus LTA significantly promoted liver cancer development, accompanied by increased levels of SASP factors[18].

Endotoxemia induced TLR 2 induction leads to COX2 mediated PGE production which suppresses antitumor immunity by inhibiting antitumor cytokine production from liver immune cells leading to HCC progression in a mouse model[7,18]. In human HCCs with noncirrhotic, NAFLD, COX2 overexpression and excess PGE production are detected[7,18]. Although these studies suggest that hepatocellular inflammation may be secondary to altered intestinal permeability and translocation of either intact bacteria or microbial cell components into circulation the causal link between them is not completely clarified[24].

The liver immune resident macrophages (Kupffer cells) and vascular system act to clear microbes that have penetrated the intestinal wall. Liver disease itself can cause dysfunction of this barrier and may promote increased intestinal permeability and translocation of bacteria or their components[25]. Liver disease in this model may directly contribute to the alteration of intestinal permeability and microbiome dysbiosis through portal hypertension.

***Bile acids and interaction with gut microbiome, role in liver disease***

**Changes in bile acid metabolism are present in advanced liver disease:** Bile acids have antimicrobial effects mediated directly or *via* induction of FXR nuclear receptors found in the intestine and liver that are closely linked with bile acid metabolism[26,27].Bile antimicrobial properties are observed as a result of bile’s capability of carrying out membrane-damaging effects. Electron microscopy and enzyme assay-based evidence has indicated in previous studies that, upon exposure to bile, cells shrink and lose intracellular material, thereby compromising membrane integrity[26]. Furthermore, bile has numerous other effects on bacterial stability such as changing the structure of RNA, inducing DNA damage, and alteration in protein structure, causing misfolding or denaturation, *via* detergent action[24]. The demonstration of the potential role of FXR against overgrowth is a possible area of potential research with utilization of synthetic FXR agonists in patients with reduced or obstructed bile flow who are at risk for bacterial overgrowth[27]. In cirrhosis there is decrease in total fecal bile acids and change in the bile acid profile which can result in intestinal bacterial overgrowth as bile acids have direct bacteriostatic effects[28-30]. Dysbiosis in liver disease has been associated with reduced Gram positives like *Ruminnocococcacea* and increased *Enterobacteracieas* associated with decreased bile acid levels and with increased inflammation and LPS.

Bile acids are secreted by the liver and undergo extensive enterohepatic circulation in the small intestine and are influenced by the intestine microbiome. In obesity associated microbiomes there is increased conversion of primary bile acids like chenodeoxycholic acid to DCA which are toxic to the liver[31]. Unlike mice the human liver cannot convert DCA back to primary bile acids as it lacks the enzyme (7 hydroxylase) and secondary bile acids can accumulate to very high levels in the liver. After undergoing enterohepatic re-circulation increased DCA accumulation in the liver can cause oxidative injury to mitochondria and cell walls and increased reactive oxygen species development. Secondary BAs promote HCC development by activating SASP in hepatic stellate cells and hepatocyte proliferation *via* the hepatic mTOR pathway[32]. DCA has been associated with dysbiosis (Clostridium clusters) and development of HCC development in a obesity associated mouse model[33]. Since high fat diet can result in high DCA levels in healthy male volunteers, the DCA induced changes in stellate cells (SASP) may contribute to obesity associated HCC[33]. Increased DCA was associated with not only the increased relative abundance of specific bacterial groups, including *Bacteroidaceae* and *Lachnospiraceae spp.*, but also advanced fibrosis in NAFLD[34].

The production of bile acids in the liver and reabsorption in the ileum is regulated by a feedback mechanism with both bile acid induced activation of nuclear receptors like *FXR* in the intestine downregulating the apical bile salt pumps that transport bile back to the liver and activation of FXR nuclear receptors in the liver by bile acids downregulating their production. *TGR 5* is another universal bile acid sensing receptor that interacts with bile acids. Activation of these bile acid sensing receptors has been shown to have important anti-inflammatory and metabolic effects resulting in decreased liver lipogenesis, stellate cell activation, reduced gluconeogenesis resulting in improved liver metabolic profile[35].

Normal bile acids profiles are also involved in maintaining healthy intestinal epithelial barrier function *via* FXR and EGFR dependent pathways. The FXR dependent pathway is linked to the gut microbiota and liver disease *via* its regulation of bile acid concentration and composition. The presence of FXRs and their related target genes help maintain homeostasis of bile acid, glucose and fat levels in the liver and intestine. Inhibition of FXR disrupts this homeostasis and can lead to cholestasis, an excess buildup of bile acids in the liver. A lack of bile acids entering the intestine contributes to dysbiosis, which is commonly seen in conjunction with liver disease[31,36]. EGFR is important for the regeneration of liver cells post-injury, and the expression of this receptor is controlled by the bile acid profile in the gut-liver axis. An excess buildup of bile acids in the liver (caused by FXR inhibition, for example) results in overexpression of EGFR and its ligands, which is a trend commonly seen in patients with HCC[36].

Reduced levels of FXR expression have been found in mouse models that are associated with reduced expression of tumor suppressors (like SHP) and to liver cancer development.

In a mouse model of liver cancer, altering gut bacteria had an anti-tumor effect mediated *via* bile acid signaling that increased antitumor NKT cells in the liver. Primary bile acids were associated with antitumor effects *via* bile acid signaling through chemokines (CXCL16) whereas secondary bile acids were associated with reduced antitumor immunity. The use of vancomycin that reduced bacteria that convert primary to secondary bile acids were associated with increased NKT cell accumulation and reduced liver tumors[37].

***Gut microbiome, metabolites and liver disease***

The gut microbiome also impacts host metabolic processes such as energy extraction from food and is a major environmental factor contributing to NAFLD. The gut microbiota have the potential to increase intrahepatic fat through mechanisms such as altered appetite signaling, increased energy extraction from diet and altered expression of genes involved in de novo lipogenesis or oxidation[38,39]. There is evidence suggesting that microbiota may play a significant role in diurnal/circadian rhythm regulation a key process in mammalian metabolism that synchronizes metabolism to night and day light cycles[40]. In an animal model diurnal metabolic rhythms in metabolism were influenced by the intestinal microbiome via expression of intestinal epithelial histone deacetylase 3 (HDAC3) which takes inputs from the microbiota and circadian cycles and relays the signals from the inputs towards the host genes responsible for metabolism specifically in lipid transport promoting diet induced obesity[41]. Therefore, the possible disruption of the human microbiome can lead to worsening obesity and disruption of metabolic homeostasis leading to metabolic diseases like NAFLD and associate complications like cirrhosis and HCC.

***Gut metabolites***

Short chain fatty acids (SCFAs) are important for colonic epithelial integrity and serve as a valuable nutritional source in the colon. SCFA including formate, acetate, propionate and butyrate can enter the liver through portal vein and cause lipid accumulation and glucogenesis[38,42]. In NAFLD there are reduced levels of short chain anti-inflammatory fatty acids further affecting host energy absorption[43]*.* Other human NAFLD studies have shown increased SCFA levels (acetate and propionate) associated with reduced tREG cells and other markers of reduced immunologic progression of liver disease[44]. Circulating levels of butyrate are inversely related to portal hypertension, endotoxemia, and systemic inflammation in patients with cirrhosis**.** The effects of the microbiome on SCFA are still being investigated and discrepant results in studies may be due to variations in patient diet, age and other environmental factors. Based on the preponderance of preclinical data there is interest in investigation of the antisteatotic effects of SCFA supplementation in NAFLD.

Branched chain amino acids (BCAA) *vs* aromatic amino acids (AAA) balance can induce insulin resistance and steatosis and is associated with certain bacterial species (*Prevotella* and *Bacteroides*). BCAA have been positively associated with simple steatosis to NASH, NASH-cirrhosis and HCC, while Glutathione was inversely associated, although reverse effects are found in human and animal studies. Metabolites derived from aromatic amino acids can have anti-inflammatory effects in host cells[45].

***Microbiota profiles in liver disease***

Thus far there are no prospective longitudinal clinical studies showing correlation of microbial profile with HCC risk. Most studies are cross-sectional correlating microbial profiles with HCC risk while trying to control for other predictors. Functional studies that show a microbial risk profile development of HCC are animal based (mice based). In various models, germ free mice or sterilized mice have lower HCC risk. Administration of MAMPs can increase this risk (Table 1).

Identification of microbial signatures associated with HCC has the potential to be used for disease diagnostics in patients at risk of HCC. The microbiome also has the potential to be used for therapy (*e.g.*, in conjunction with cancer immunotherapy, see therapy section below).

The microbiome in obese individuals has reduced bacterial diversity and a higher potential for inflammation in a study of obesity, bimodal gene distribution was observed. Reduced bacterial diversity in obese individuals can lead to dysbiosis, which is associated with the development of liver diseases such as NAFLD and HCC - and so obesity is considered a risk factor for metabolic liver disease. A higher risk group was characterized by a higher prevalence of anti-inflammatory species such as *F. prausnitzii*, and an increased production potential of organic acids (including butyrate). In contrast, lower risk groups showed higher relative abundance of potentially proinflammatory *Bacteroides spp.* and genes involved in oxidative stress response. These groups were associated with insulin resistance but not BMI[46].

*E. coli*, E*nterobacteriaceae spp.*, and *Klebsiella pneumonia* have been identified as ethanol-producing bacteria and were found to be relatively abundant in NAFLD patients[47,48]. In NAFLD advanced fibrosis was associated with an increased abundance of *Proteobacteria* and *E. coli* and a decrease in Firmicutes[49]. This gut microbiota profile promotes absorption of monosaccharides from the gut lumen, resulting in the induction of *de novo* hepatic lipogenesis[50].

Increased DCA was associated with not only the increased relative abundance of specific bacterial groups, including *Bacteroidaceae* and *Lachnospiraceae spp.*, but also advanced fibrosis in NAFLD[48,51].

A study from China showed a microbial signature profile (distinct bacterial species) that were present in early stage HCC with cirrhosis compared to cirrhosis without HCC and healthy controls). While diversity of species was decreased in cirrhosis compared to controls it was increased in early-stage HCC compared to cirrhosis without HCC. Bacteria producing butyrate (potentially beneficial SCFA associated with improved gut barrier and liver immunity) were decreased in HCC while those producing LPS were increased[52].

An Italian study showed *E. coli* overgrowth in the intestines of HCC and cirrhosis patients as well as increased levels of *Bacteroides* and *Ruminococcaceae spp.* and decreased levels of *Akkermensia* and *Bifidobacterium spp*. *Akkermansia* and *Bifidobacterium* were inversely correlated with calprotectin concentration, which in turn was associated with humoral and cellular inflammatory markers. A similar pattern was also observed for Bacteroides. This study suggests that gut microbiota profile and systemic inflammation are significantly correlated in NAFLD-HCC[18]. Another study also demonstrated *Bacteroides* and *Ruminococcaceae* increased but *Bifidobacterium* decreased in NAFLD-HCC[48].

The biggest differences in microbial profiles so far have been found between patients with cirrhosis and healthy patients and less so than those between cirrhosis with or without HCC. Thus the impact of the microbiome may be more in development of cirrhosis (the biggest risk factor for HCC) rather than HCC development itself. Differences in microbial profiles in studies from different countries suggest there are important regional differences, influenced by dietary, genetic or underlying cause of cirrhosis (*e.g.,* HBV HCC in Asia *vs* non HBV HCC in other part of the world) that should be accounted for in future studies.

***Therapeutics, gut microbiome, role in liver cancer***

Manipulation of the gut microbiome to alleviate disease including liver disease is a burgeoning area of research. Studies have looked at diet and impact on microbiome. The impact of using prebiotics, prebiotics and antibiotics to modulate the microbiome to impact liver disease is an area of active research as is the impact of the microbiome in influencing efficacy of liver cancer chemotherapeutics (Table 2).

There is epidemiologic evidence that dietary patterns and nutrients are associated with liver cancer (red meat, added sugar, particularly high fructose, processed food have been shown to pose a higher risk whereas fruit, vegetables, omega-3 oil (fish), coffee potentially lower risk). Animal (and some human) studies in liver disease also show high fat diet promoting worsened intestinal barrier function, decreased bacterial diversity and endotoxemia reaching the liver and adipose tissue promoting chronic inflammation and metabolic disease. Potential mechanisms whereby diet can influence cancer risk is through influencing inflammatory pathways, and potentially through modulation of the gut microbiome.A high fat diet results in a higher proportion of gram-negative bacteria - such as *Bacteroides* - in the gut microbiome, resulting in a higher concentration of the LPS contained within the cell membrane of these bacteria. LPS activates toll-like receptors 4 and 9, which contributes to the fibrosis seen in NAFLD, NASH and HCC development[5,11,64].

Besides diet, other factors also regulate the microbiome and endotoxemia including genetics, and exogenous factors like exercise and alcohol.

***Potential therapeutic strategies***

Probiotics, prebiotics, fecal microbiota transplant (FMT), antibiotics for dysbiosis. No clinical studies have shown that intervening with the microbiome can influence the risk of HCC development in human trials thus far. In animal studies prebiotics (which are dietary fermentable substrates that can modulate microbiome growth) have been shown to reduce hepatic triglyceride accumulation *via* inhibition of lipogenesis and reduced expression of genes such as FAS[53].

In humans, supplementation with prebiotics such as oligofructose has been associated with decreased hepatic inflammatory markers. In a clinical trial with 14 participants, changes in body weight, glucose tolerance, and inflammatory markers among others were followed and observed over the course of 9 mo. The individuals treated with the prebiotics had markedly better markers than the individuals treated with the placebo and the overall rates of steatosis in patients treated with the prebiotic that were affected by non-alcoholic steatohepatitis did decrease[54].

Probiotics are live bacteria which are beneficial to the host. Probiotics in mice have been shown to enhance bile acid fecal excretion, reduce bile acid reabsorption in the intestine and reverse abnormal bile acid metabolism seen in dysbiosis. *Lactobacillus* and *Bifidobacterium spp.* have been reported to reduce gut inflammation and improve gut barrier function by remodeling the gut microbiota[53]. *Lactobacilli*, was administered orally to mice in a model of liver disease and reduced the frequency of NASH considerably while also reducing inflammation and fibrosis in the liver. In human studies, administration of *Lactobacillus acidophilus* reduced AST and ALT levels in NAFLD patients[55].

The modulation of gut microbiome *via* FMT has demonstrated decreased hepatic lipid accumulation and decreased pro-inflammatory cytokine levels after FMT in a mouse model of liver disease fed a high fat diet[56]. Additionally, FMT increased the relative abundance of beneficial bacterial species of *Christensenellaceae* and *Lactobacillus*, improved gut barrier function, and increased butyrate production and reduced endotoxemia[58]. In a human trial, FMT from lean donors to individuals with metabolic syndrome temporarily increased insulin sensitivity whereas autologous FMT from the donors with metabolic syndrome did not show this change[57].

Antibiotics have been studied in experimental models of liver disease. In a mouse model of liver cancer, vancomycin treatment significantly reduced obesity-associated liver cancer development[18]. The use of antibiotics in a preclinical mouse model indicated that chronic oral administration of antibiotics decreased secondary bile acid levels, hepatic lipid accumulation, and attenuated hepatic inflammation and fibrosis *via* modulating the composition of gut microbiota[58,59].

In one study conducted on liver cancer model of mice, it was shown that various species of *Clostridium bacteria* can accumulate and potentially result in suppression of natural antitumor mechanisms against tumors found in the liver. Mice that were affected with the tumors were given antibiotic treatment that affected the state of the gut microbiome and resulted in a reduction in the growth of tumors in the liver as well as the metastasis of tumors originating in the liver. Antibiotics however may have divergent effects and Mahana *et al*[60] showed that mice treated with antibiotics exhibited severe insulin resistance and NAFLD associated with a change in composition of the gut microbiota from *Firmicutes* to *Bifidobacterium* and *Prevotella*.

The microbiome also has the potential to be used in conjunction with cancer immunotherapy. *Bacteroides* and *Bifidobacteria* species can assist T-cell-based immunotherapies in combating against cancer in the liver[61].

In a study of liver cancer undergoing immunotherapy (checkpoint inhibitor, anti-PD-1), gut microbiota profiles were different in patients responding to chemotherapy. These profiles varied during therapy and may enable early identification (within 6 wk changes in bacteria) of responders *vs* non responder to immunotherapy. Changes in the microbiome that correlated with response or non-response were seen as early as 6 wk. after start of immunotherapy[34]. Pathways by which the microbiome influenced cancer therapy efficacy may include the effect of certain bacterial species in improving host immunity, decreased intestinal permeability, decreased oxidative stress and decreased growth of pathogenic bacteria.

Probiotics and FMT are currently being investigated in cancer treatment as an adjuvant strategy to increase the efficacy of chemotherapy and immunotherapy[62].

FXR agonists that are in clinical use for patients with cholestatic liver diseases like PBC and being investigated for NAFLD have the potential to restore healthy gut microbiota and ameliorate metabolic diseases though effect on carbohydrate and lipid metabolism. In animal models, use of these agents have led to decreased endogenous bile acid levels and improved bacterial profile with increase in the proportion of Firmicutes[63].

**CONCLUSION**

***Future directions***

The role of the microbiome in chronic liver disease, particularly NAFLD and associated liver cancer is being elucidated through experimental and clinical studies. With the increasing epidemic of NAFLD and liver cancer this is an exciting and critical area of investigation.

Active areas of investigation include searching for effective HCC treatment and prevention in patients with chronic liver disease utilizing the knowledge gained from studies on the microbiome in liver disease. Utilizing the presence of distinct gut microbial profiles in earlier stage chronic liver disease, such as NAFLD and NASH, is emerging as an area for potential diagnostics as well as for therapeutics. Early identification of the signs of progressive liver disease, such as decreased microbiome diversity, increase in cytokine expression, and leaky gut may be important in preventing HCC development.

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**Footnotes**

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**Figure Legends**



**Figure 1 Activation of the immune response results in cytokine and chemokine expression and the recruitment of inflammatory cells in the liver, hepatocyte proliferation as well as hepatic stellate activation which result in progressive liver inflammation, fibrosis and liver cancer.**

**Table 1** **Microbiota profiles in liver disease**

|  |  |  |  |  |
| --- | --- | --- | --- | --- |
| **Study** | **Bacterial species increased** | **Bacterial species decreased** | **Putative impact of microbial change in group** | **Comparison group** |
| Advanced fibrosis/cirrhosis |
| Loomba *et al*[49], 2017, Cell Metab  | Proteobacteria Include *E. coli*  | Firmicutes  | Higher incidence of enzymes for butyrate, and lower for lactate and acetate  | NAFLD advanced fibrosis/cirrhosis compared to NAFLD mild/moderate fibrosis |
| Ponziani *et al*[18], 2019, Hepatology | **Phylum:** *Proteobacteria,* *Bacteroidetes,* *Cyanobacteria;* **Family:** *Lactobacillaceae,* *Enterobacteriaceae,* *Prevotellaceae,* *Bacteroidaceae,* *Streptococcaceae,* *Enterococcaceae,* *Veillonellaceae* | **Phylum:** *Verrucomicrobia;* **Family:** *Verrucomicrobiaceae;* *Methanobacteriaceae* | Intestinal inflammation,Increase in Intestinal permeability, increased systemic inflammation | NAFLD cirrhosis compared to healthy controls |
| HCC |  |  |  |  |
| Ren *et al*[52], 2019, Gut | **Phylum:** *Acinetobacter*; **Genus:** *Gemmiger,* *Parabacteroides,* *Paraprevotella,* others | **Phylum:** *Verrucomicrobia* | Increased LPS producer with liver inflammation and oxidative damage; Decreased butyrate production resulting in intestinal mucosal disruption | Early HCC compared to cirrhosis  |
| Ren *et al*[52], 2019, Gut | **Genus:** *Klebsiella,* *Haemophilus* | *Verrucomicrobia (Akkermansia);* **Genus:** *Alistipes,* *Phascolarctobacterium,* *Ruminococus* | Increased LPS producer with liver inflammation and oxidative damage; Decreased butyrate production resulting in intestinal mucosal disruption | Early HCC compared to healthy controls |
| Ponziani *et al*[16], 2019, Hepatology | **Phylum:** *Bacteroidetes*; **Family:** *Bacteroidaceae,* *Streptococcaceae,* *Enterococcaceae* | **Family:** *Verrucomicrobiaceae,* *Bifidobacteriaceae* | Intestinal inflammation, Increase in Intestinal permeability, increased systemic inflammation | HCC in NAFLD Compared to NAFLD cirrhosis without HCC |
| Grat M *et al*[15], 2016, Transplant Proc | *E. coli* |  | LPS and inflammation within liver | HCC cirrhosis Compared to non-HCC cirrhosis |

Selected studies (human) showing different microbiota profiles in cirrhosis and hepatocellular carcinoma (HCC) with emphasis on non-alcoholic fatty liver disease associated HCC. HCC: Hepatocellular carcinoma; NAFLD: Non-alcoholic fatty liver disease; LPS: Lipopolysaccharide.

**Table 2** **Therapeutics, gut microbiome, role in liver cancer**

|  |  |  |  |
| --- | --- | --- | --- |
| **Study** | **Agents** | **Population** | **Outcomes** |
| Animal models |  |  |  |
| Borges Haubert *et al*[64], 2015, Nutr Metab Insights  | *Prebiotics* fructoligosaccharides | NAFLD rat model | Decreased liver fat *via* decreased lipogenesis |
| Liu *et al*[65], 2020, J Nutr Biochem | *Probiotic* Lactobacillus rhamnosus | Mouse model of liver disease (HFD) | Reduced NASH frequency, reduced steatosis inflammation and apoptosis in liver |
| Zhou *et al*[56], 2017, Sci Rep | *FMT* | Mouse model of liver disease (HFD) | Decreased hepatic lipid and proinflammatory cytokines, increased lactobacillus, improved gut barrier function, reduced endotoxemia, increase butyrate  |
| Yoshimoto *et al*[33], 2013, Nature  | *Vancomycin* | Mouse model of liver disease (HFD) | Reduced liver cancer |
| Janssen *et al*[58], 2017, Lipid Res  | Antibiotics (ampicillin, neomycin, vancomycin and metronidazole) | Mouse model of NAFLD | Decreased secondary bile acids, decreased liver inflammation and fibrosis |
| Friedman *et al*[63], 2018, Gastroenterology | FXR agonist obeticholic acid | Mouse model of NAFLD | Decreased endogenous bile acid; Increased bacterial profile with Gram + including Firmicutes |
| Humans studies |  |  |  |
| Yang *et al*[5], 2020, Br J Nutrition | Diet and incident cancer risk- summary of studies  | Worldwide epidemiologic studies of diet and liver cancer risk | *Higher risk for liver cancer:* Red Meat, Added Sugar, Processed food; *Lowered risk for liver cancer:* Vegetables, Fruits, Omega 3 oil, Coffee |
| Monem *et al*[55], 2017, Euroasian J hepatogastro | *Probiotics* Lactobacillus | NAFLD patients | Decreased AST and ALT |
| Bomhof *et al*[54], 2019, Eur J Nutrition | *Prebiotic* Oligofructose | NASH patients | Decreased hepatic inflammatory markers, deceased weight, improved glucose tolerance, decreased steatosis, decreased clostridium cluster XA and I and enhanced Bifidobacterium |
| Vrieze *et al*[57], 2012, Gastroenterology | FMT trial | FMT from lean donor to individuals with metabolic syndrome | Improved insulin sensitivity, Increase in butyrate producing bacteria |

Selected studies (animal and human) of microbiome modulating therapies tested in non alcoholic steatohepatitis (NASH) and hepatocellular carcinoma (HCC) with emphasis on NASH HCC. NASH: Non alcoholic steatohepatitis; NAFLD: Non alcoholic fatty liver disease; HCC: Hepatocellular carcinoma; FMT: Fecal microbiota transplant; PD-1: Programmed death-1; FXR: Farnesoid X receptor.