

# World Journal of *Clinical Oncology*

*World J Clin Oncol* 2021 May 24; 12(5): 290-392



### REVIEW

- 290 Cellular based treatment modalities for unresectable hepatocellular carcinoma  
*Damiris K, Abbad H, Pyrsopoulos N*

### MINIREVIEWS

- 309 COVID-19: Where is the treatment?  
*Sabeerabi B, Vemula S, Vadde R, Nagaraju GP*
- 323 Immune response evaluation criteria in solid tumors for assessment of atypical responses after immunotherapy  
*Ippolito D, Maino C, Ragusi M, Porta M, Gandola D, Franzesi CT, Giandola TP, Sironi S*
- 335 Hepatic Langerhans cell histiocytosis: A review  
*Fu Z, Li H, Arslan ME, Ells PF, Lee H*

### ORIGINAL ARTICLE

#### Basic Study

- 342 Thymoquinone anticancer activity is enhanced when combined with royal jelly in human breast cancer  
*Moubarak MM, Chanouha N, Abou Ibrahim N, Khalife H, Gali-Muhtasib H*

#### Retrospective Study

- 355 Prognostic role of sarcopenia in metastatic colorectal cancer patients during first-line chemotherapy: A retrospective study  
*Maddalena C, Ponsiglione A, Camera L, Santarpia L, Pisanisi F, Bruzzese D, Panico C, Fiore G, Camardella S, Caramia T, Farinara A, De Placido S, Carlomagno C*

### SYSTEMATIC REVIEWS

- 367 Reduction of muscle contraction and pain in electroporation-based treatments: An overview  
*Fusco R, Di Bernardo E, D'Alessio V, Salati S, Cadossi M*
- 382 Intestinal metastasis from breast cancer: Presentation, treatment and survival from a systematic literature review  
*Bolzacchini E, Nigro O, Inversini D, Giordano M, Maconi G*

## ABOUT COVER

Editorial Board Member of *World Journal of Clinical Oncology*, Qiang Huo, MD, MSc(Med), Doctor-In-Charge, Center for Translational Medicine, Zibo Central Hospital, No. 54 West Gongqing Road, Zibo 255036, Shandong Province, China. qianghuo@mail.sdu.edu.cn

## AIMS AND SCOPE

The primary aim of *World Journal of Clinical Oncology* (WJCO, *World J Clin Oncol*) is to provide scholars and readers from various fields of oncology with a platform to publish high-quality basic and clinical research articles and communicate their research findings online.

WJCO mainly publishes articles reporting research results and findings obtained in the field of oncology and covering a wide range of topics including art of oncology, biology of neoplasia, breast cancer, cancer prevention and control, cancer-related complications, diagnosis in oncology, gastrointestinal cancer, genetic testing for cancer, gynecologic cancer, head and neck cancer, hematologic malignancy, lung cancer, melanoma, molecular oncology, neurooncology, palliative and supportive care, pediatric oncology, surgical oncology, translational oncology, and urologic oncology.

## INDEXING/ABSTRACTING

The WJCO is now abstracted and indexed in PubMed, PubMed Central, Emerging Sources Citation Index (Web of Science), China National Knowledge Infrastructure (CNKI), China Science and Technology Journal Database (CSTJ), and Superstar Journals Database.

## RESPONSIBLE EDITORS FOR THIS ISSUE

Production Editor: Ying-Yi Yuan, Production Department Director: Yun-Xiaoqian Wu, Editorial Office Director: Ze-Mao Gong.

### NAME OF JOURNAL

*World Journal of Clinical Oncology*

### ISSN

ISSN 2218-4333 (online)

### LAUNCH DATE

November 10, 2010

### FREQUENCY

Monthly

### EDITORS-IN-CHIEF

Hiten RH Patel, Stephen Safe

### EDITORIAL BOARD MEMBERS

<https://www.wjgnet.com/2218-4333/editorialboard.htm>

### PUBLICATION DATE

May 24, 2021

### COPYRIGHT

© 2021 Baishideng Publishing Group Inc

### INSTRUCTIONS TO AUTHORS

<https://www.wjgnet.com/bpg/gerinfo/204>

### GUIDELINES FOR ETHICS DOCUMENTS

<https://www.wjgnet.com/bpg/gerinfo/287>

### GUIDELINES FOR NON-NATIVE SPEAKERS OF ENGLISH

<https://www.wjgnet.com/bpg/gerinfo/240>

### PUBLICATION ETHICS

<https://www.wjgnet.com/bpg/gerinfo/288>

### PUBLICATION MISCONDUCT

<https://www.wjgnet.com/bpg/gerinfo/208>

### ARTICLE PROCESSING CHARGE

<https://www.wjgnet.com/bpg/gerinfo/242>

### STEPS FOR SUBMITTING MANUSCRIPTS

<https://www.wjgnet.com/bpg/gerinfo/239>

### ONLINE SUBMISSION

<https://www.f6publishing.com>

## Cellular based treatment modalities for unresectable hepatocellular carcinoma

Konstantinos Damiris, Hamza Abbad, Nikolaos Pyrsopoulos

**ORCID number:** Konstantinos Damiris 0000-0001-9972-740X; Hamza Abbad 0000-0002-0655-9564; Nikolaos Pyrsopoulos 0000-0002-6950-8174.

**Author contributions:** Damiris K, Abbad H, and Pyrsopoulos N equally contributed to this paper with conception and design of the study, literature review and analysis, drafting and critical revision/editing; all authors have read and approve the final manuscript.

**Conflict-of-interest statement:** The authors do not have any conflicts of interest relevant to this article.

**Open-Access:** This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

**Manuscript source:** Invited manuscript

**Konstantinos Damiris, Hamza Abbad,** Department of Medicine, Rutgers New Jersey Medical School, Newark, NJ 07103, United States

**Nikolaos Pyrsopoulos,** Department of Medicine, Division of Gastroenterology and Hepatology, Rutgers New Jersey Medical School, Newark, NJ 07103, United States

**Corresponding author:** Nikolaos Pyrsopoulos, FAASLD, AGAF, FACG, MD, PhD, Professor, Department of Medicine, Division of Gastroenterology and Hepatology, Rutgers New Jersey Medical School, 185 South Orange Ave, Newark, NJ 07103, United States.  
[pyrsopni@njms.rutgers.edu](mailto:pyrsopni@njms.rutgers.edu)

### Abstract

Hepatocellular carcinoma (HCC) is the most common primary malignancy of the liver and is unfortunately associated with an overall poor prognosis and high mortality. Early and intermediate stages of HCC allow for treatment with surgical resection, ablation and even liver transplantation, however disease progression warrants conventional systemic therapy. For years treatment options were limited to molecular-targeting medications, of which sorafenib remains the standard of care. The recent development and success of immune checkpoint inhibitors has proven to be a breakthrough in the treatment of HCC, but there is an urgent need for the development of further novel therapeutic treatments that prolong overall survival and minimize recurrence. Current investigation is focused on adoptive cell therapy including chimeric antigen receptor-T cells (CAR-T cells), T cell receptor (TCR) engineered T cells, dendritic cells, natural killer cells, and tumor infiltrating lymphocyte cells, which have shown remarkable success in the treatment of hematological and solid tumor malignancies. In this review we briefly introduce readers to the currently approved systemic treatment options and present clinical and experimental evidence of HCC immunotherapeutic treatments that will hopefully one day allow for revolutionary change in the treatment modalities used for unresectable HCC. We also provide an up-to-date compilation of ongoing clinical trials investigating CAR-T cells, TCR engineered T cells, cancer vaccines and oncolytic viruses, while discussing strategies that can help overcome commonly faced challenges when utilizing cellular based treatments.

**Key Words:** Hepatocellular carcinoma; Immunotherapy; Immune cells; Adoptive T cell therapy; Chimeric antigen receptor-T cell; Clinical trials

**Specialty type:** Oncology**Country/Territory of origin:** United States**Peer-review report's scientific quality classification**

Grade A (Excellent): A  
 Grade B (Very good): B, B, B  
 Grade C (Good): 0  
 Grade D (Fair): D  
 Grade E (Poor): 0

**Received:** March 12, 2021**Peer-review started:** March 12, 2021**First decision:** April 6, 2021**Revised:** April 19, 2021**Accepted:** April 28, 2021**Article in press:** April 28, 2021**Published online:** May 24, 2021**P-Reviewer:** Li ZM, Pongcharoen S, Zhang L**S-Editor:** Gao CC**L-Editor:** A**P-Editor:** Yuan YY

©The Author(s) 2021. Published by Baishideng Publishing Group Inc. All rights reserved.

**Core Tip:** Over the recent years investigation for safe and effective treatments for unresectable hepatocellular carcinoma (HCC) has shifted focus from various chemotherapeutic agents to immune based therapy. Although far from being finalized, immune cell-based therapy has shown efficacy in a variety of clinical trials, indicating possible future utilization alone or in combination for the prevention and treatment of HCC.

**Citation:** Damiris K, Abbad H, Pyrsopoulos N. Cellular based treatment modalities for unresectable hepatocellular carcinoma. *World J Clin Oncol* 2021; 12(5): 290-308

**URL:** <https://www.wjgnet.com/2218-4333/full/v12/i5/290.htm>

**DOI:** <https://dx.doi.org/10.5306/wjco.v12.i5.290>

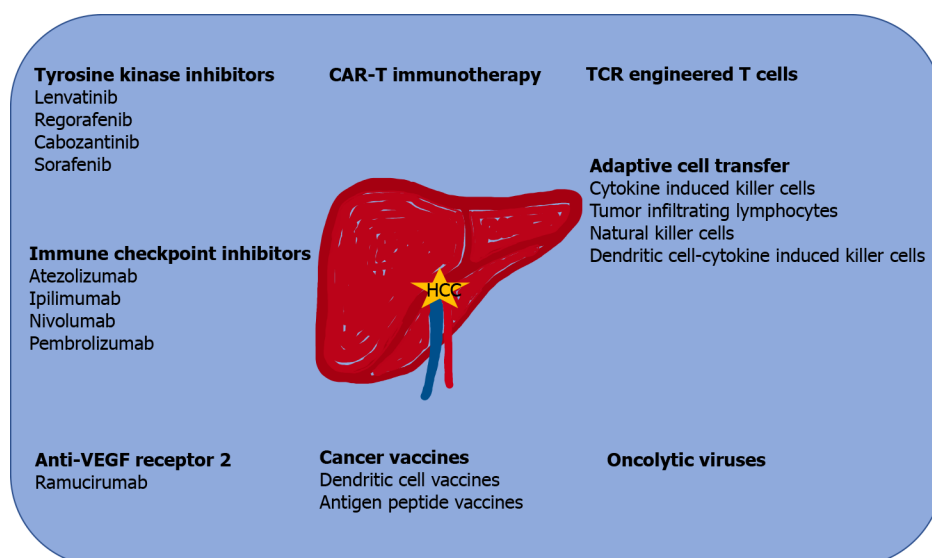
## INTRODUCTION

Hepatocellular carcinoma (HCC) is a malignant primary tumor of the liver, accounting for approximately ninety percent of total liver tumor cases worldwide[1]. HCC commonly occurs in patients with preexisting liver cirrhosis, chronic hepatitis B virus (HBV), chronic hepatitis C virus (HCV), heavy alcohol consumption, aflatoxin exposure and metabolic associated fatty liver disease associated with diabetes and metabolic syndrome[2]. According to the Global Cancer Observatory (GLOBOCAN) in 2020, HCC is considered the 6<sup>th</sup> most common cancer worldwide, with the highest incidence in eastern Asia and Africa[3]. It has been estimated that by 2025, there will be more than one million individuals diagnosed with liver cancer annually[4]. Worldwide incidence of HCC has shown a greater predominance among men in comparison to women, with most diagnoses made in patients over the age of 60 years old[5]. HCC risk factors in the United States have varied significantly over the last two decades, with a majority of cases currently related to chronic HCV infection[6].

Management of HCC varies depending on the staging status of the tumor according to the standardized Barcelona-Clinic Liver Cancer (BCLC) system endorsed by both the European Association for the Study of the Liver and the American Association for the Study of Liver Diseases[7]. Locally diagnosed tumors (BCLC stages 0-A), which are small in size ( $\leq 3$  cm), early in progression, and limited to the liver parenchyma are amendable to surgical resection, radiofrequency ablation or ideally liver transplantation[8], with a median survival rate of 60 mo[9]. Advanced HCC on the other hand is defined by a significant increase in tumor size, vascular invasion and/or metastatic disease that is not amenable to loco-regional therapy (BCLC Stage C-D)[10], warranting treatment with systemic therapy[11]. In cases necessitating liver transplantation, the Milan criteria has been adopted by The United Network for Organ Sharing in order to determine eligibility[12], however this approach is limited by organ availability[13].

For over a decade, sorafenib has been considered the first line systemic treatment for advanced HCC. More recently, other systemic treatments such as the tyrosine kinase inhibitors (lenvatinib, regorafenib, and cabozantinib), immune checkpoint inhibitors (ICIs) (atezolizumab, ipilimumab, nivolumab, pembrolizumab) and the monoclonal antibody ramucirumab have been approved as first- and second-line treatment options[14]. With only modest established improvement in overall survival (OS) and an assortment of adverse effects, there has been a recent push for the development of immunobiological treatments, which first demonstrated efficacy in the treatment of hematological cancers. Treatments include chimeric antigen receptor-T cells (CAR-T cells), T cell receptor (TCR) engineered T cells, dendritic cells (DC), natural killer (NK) cells, and tumor infiltrating lymphocyte (TIL) cellular therapies (Figure 1). In this article we will review the novel range of cellular based treatments for HCC non-amenable to loco-regional therapy and introduce readers to various preclinical and clinical trials investigating their efficacy.





**Figure 1** Treatment modalities available for unresectable (advanced) hepatocellular carcinoma, which is non-amendable to loco-regional therapies. CAR: Chimeric antigen receptor; HCC: Hepatocellular carcinoma; TCR: T cell receptor; VEGF: Vascular endothelial growth factor.

## SYSTEMIC THERAPIES: APPROVED FIRST LINE THERAPIES

### **Sorafenib**

Sorafenib is a multi-kinase inhibitor that inhibits cellular proliferation and angiogenesis through its effects on various receptor tyrosine kinases including vascular endothelial growth factor (VEGF), platelet derived growth factor (PDGF) and c-kit [15]. Its efficacy was demonstrated in two large phase III randomized controlled trials; SHARP conducted in the United States and Europe, and a similar study conducted in the Asia-Pacific focusing on patients with unresectable and advanced HCC [16]. In the SHARP trial, median OS was increased to 10.7 mo compared to 7.9 mo in placebo [17]. Similar findings were demonstrated in the Asia-Pacific trial; median OS increased in the treatment group (6.5 mo *vs* 4.2 mo) and time-to-progression (TTP) was also significantly greater (2.8 mo *vs* 1.4 mo) [18]. These findings led to sorafenib being approved by the Food and Drug Administration (FDA) in 2007, making it the standard of care and first line treatment option of advanced HCC [15,19].

### **Lenvatinib**

Lenvatinib is another inhibitor of multiple receptor tyrosine kinases that has recently gained much attention [20]. In the randomized, open-label, non-inferiority phase III trial (REFLECT), lenvatinib was compared to sorafenib in patients who were deemed to have non-resectable HCC. Findings indicated an improved median OS of 13.6 mo with lenvatinib, compared to 12.3 mo with sorafenib. Lenvatinib also demonstrated an increase in median progression-free survival (PFS) and overall response rate (ORR) [21]. This allowed for FDA approval in 2018 as a first line agent for treatment of unresectable HCC [22].

## SYSTEMIC THERAPIES: APPROVED SECOND LINE THERAPIES

### **Ramucirumab**

The phase III randomized controlled trial REACH (NCT01140347) investigated ramucirumab, a monoclonal antibody against VEGF receptor 2. While the study failed to demonstrate prolonged OS in the entire cohort, patients with elevated serum alpha-fetoprotein (AFP) (> 400 ng/mL) had significantly longer median OS when compared to placebo (7.8 mo *vs* 4.2 mo,  $P = 0.006$ ) [23]. A subsequent phase III study, REACH-2 (NCT02435433) reported a significantly prolonged median OS (8.5 mo *vs* 7.3 mo) in sorafenib-experienced patients receiving ramucirumab *vs* placebo with AFP of 400 ng/mL or greater [24]. These findings showed promise as an effective treatment option in those with elevated AFP, leading to FDA approval in May 2019.

**Regorafenib**

Regorafenib is a multikinase inhibitor that inhibits various molecules including VEGF receptor 2/3, PDGF receptor and fibroblast growth factor receptor 1[25]. In the randomized, double-blind, phase III clinical trial RESORCE (NCT01774344) patients who failed sorafenib therapy were assigned to receive regorafenib *vs* placebo. This study determined that regorafenib significantly improved OS in individuals who experienced radiological progression of HCC during sorafenib treatment (10.6 mo *vs* 7.8 mo,  $P < 0.0001$ )[26]. Based on the promising findings of the RESORCE trial and a tolerable safety profile, the FDA approved regorafenib as a second line treatment option in those who have HCC progression while on sorafenib or who are not eligible for alternative therapy[25].

**Cabozantinib**

A double-blind, phase III clinical trial CELESTIAL (NCT01908426) was conducted testing cabozantinib, a multikinase inhibitor, in patients with unresectable HCC who had progressed with sorafenib. The study demonstrated a significant prolongation of OS when compared to placebo (10.2 mo *vs* 8.0 mo,  $P = 0.005$ ) coupled with a longer median PFS when compared to placebo[27]. Based on the findings of CELESTIAL, the FDA approved cabozantinib as a second line treatment option for HCC in those who have undergone previous therapy with sorafenib.

**IMMUNOTHERAPY: ICIS**

Limited efficacy and undesirable side effects of traditional therapies (*i.e.*, sorafenib) have led to further investigation of immunotherapeutic agents over the years. A recent and emerging field of treatment includes cancer immunotherapies using ICIs that target programmed cell death protein-1 and its ligand (PD-1/PD-L1) and cytotoxic T-lymphocyte-associated protein-4 (CTLA-4), which has significantly altered treatment of various types of cancer including HCC[28].

Immune checkpoint molecules play essential roles in regulating cancer cell interactions with the host immune system; and the development of drugs that suppress these interactions used by tumor cells to evade host immunity has shown promising results. PD-1, a member of CD28, is expressed on activated immune cells including T cells, B cells, and myeloid cells; and its interaction with ligand PD-L1 negatively regulates the immune system through suppressive signals that induce peripheral tolerance[29]. In HCC tumor cells, PD-L1 is aberrantly expressed, allowing for such an escape from host immunity[30]. CTLA-4 is another member of the CD28 family, and it is induced on T cells by antigen activation leading to subsequent binding with CD80 and CD86 with more affinity compared to CD28, negatively regulating costimulatory T cell signals and allowing for peripheral tolerance[31]. Ultimately these immunotherapeutic drugs strategically suppress key checkpoints that may be used by tumor cells to evade the host immune system, hindering a hallmark for tumor progression.

**Atezolizumab**

Atezolizumab is a fully humanized monoclonal antibody targeting PD-L1 that has recently changed the treatment landscape of unresectable HCC. A phase-Ib study evaluating synergistic atezolizumab and bevacizumab (anti-VEGF antibody) in patients with previously untreated and unresectable HCC not only demonstrated an acceptable safety profile, but promising antitumor activity with ORR 36%, disease control rate (DCR) 71%, and a median PFS of 7.3 mo when compared to monotherapy with nivolumab or pembrolizumab[32]. These results led to IMbrave150 (NCT 03434379), an open-label, randomized phase III study assessing atezolizumab (anti-PDL1) in combination with bevacizumab (anti-VEGF antibody) compared to sorafenib in patients with untreated locally advanced or metastatic HCC. OS at 12 mo was significantly longer in the combination group when compared to sorafenib (67.2% *vs* 54.6%) and PFS was significantly longer in atezolizumab + bevacizumab *vs* sorafenib (6.8 mo *vs* 4.3 mo, hazard ratio for disease progression or death [0.59; 95% confidence interval (CI): 0.47-0.76,  $P < 0.001$ ], leading to FDA approval in May 2020 for the treatment of unresectable or metastatic HCC who have not received prior systemic therapy[33]. Currently a phase III, multicenter study of atezolizumab plus bevacizumab *vs* active surveillance as adjuvant therapy in HCC at high risk of recurrence after surgical resection or ablation is recruiting (IMbrave050; NCT04102098)[34]. In addition, a phase III study evaluating the safety and efficacy of atezolizumab plus

levantinib or sorafenib *vs* levantinib or sorafenib alone in locally advanced or metastatic and/or unresectable HCC following prior treatment with combination atezolizumab and bevacizumab was recently posted in February 2021 (IMbrave251; NCT04770896).

### ***Ipilimumab***

Ipilimumab is a monoclonal antibody targeting CTLA-4, which was recently granted accelerated FDA approval in May 2020, when used in combination with nivolumab (anti-PD-1) for HCC that has previously been treated with sorafenib. Approval was based on a single arm of the phase I/II clinical trial CheckMate 040 (NCT01658878), in which patients were treated with nivolumab 3 mg/kg plus ipilimumab 3 mg/kg every 3 wk for 4 doses. In addition to a substantial reduction in tumor burden, the overall ORR was 32%, with median OS 22.8 mo while having manageable safety profiles[35]. A phase III trial of ipilimumab in combination with nivolumab *vs* sorafenib or levantinib as first line treatment in patients with advanced HCC is currently recruiting participants (CheckMate 9DW; NCT04039607). A variety of clinical trials evaluating ipilimumab in combination with nivolumab during various treatment phases are ongoing and include; prior to liver resection (PRIME-HCC; NCT03682276), as neoadjuvant therapy (NCT03510871), and in combination with trans-arterial chemoembolization (CheckMate 74W; NCT04340193).

### ***Nivolumab***

Nivolumab is an anti-PD-1 monoclonal antibody that has shown promising results, leading to FDA approval for second-line treatment of advanced HCC. The phase I/II clinical trial CheckMate 040 (NCT01658878) investigated nivolumab in sorafenib-naïve and sorafenib-experienced patients with intermediate-advanced HCC and Child-Pugh Stage A. During the dose-expansion phase, ORR was 20%, while DCR was 64% with a median progression free survival of 4.1 mo[36]. The subsequent phase III study CheckMate 459 (NCT2576509) evaluating nivolumab *vs* sorafenib as first line treatments for unresectable HCC failed to demonstrate statistical significance for the primary endpoint of OS[37]. Currently a phase III trial, CheckMate-9DX (NCT 03383458), is actively evaluating recurrence free survival (RFS) in those with HCC at high risk of recurrence after curative hepatic resection or ablation.

### ***Pembrolizumab***

Pembrolizumab is a humanized anti-PD-1 monoclonal antibody that has been investigated across a variety of malignancies[38]. The phase II open-label clinical trial KEYNOTE-224 (NCT02702414) studied the safety and efficacy of pembrolizumab in patient with advanced HCC after having failed treatment with sorafenib. Favorable findings included an ORR of 17%, DCR of 61%, along with PFS of 4.8 mo, median OS of 12.9 mo and TTP of 4.9 mo[39]. This promising data led to FDA approval in November 2018 as a second line treatment after sorafenib. A subsequent phase III double-blind, randomized control trial, KEYNOTE-240 (NCT02702401) compared pembrolizumab to placebo, demonstrated longer OS and PFS; however, these findings were not statistically significant and failed to meet primary endpoints[40]. There are currently two-phase III trials that are ongoing at the time of this publication; KEYNOTE-394 (NCT03062358) , evaluating pembrolizumab in Asian patients with systemically treated advanced HCC and KEYNOTE-937 (NCT03867084) as an adjuvant therapy in HCC after curative treatment[29].

Ultimately, cancer uses a variety of unique mechanisms to evade host immune response and to develop drug resistance in HCC. This has sparked much interest in combining treatment modalities, particularly combination ICIs and checkpoint inhibitors with other chemotherapeutics. HIMALAYA (NCT03298451) is an open-label, multi-center, phase III study assessing durvalumab (anti-PDL1) plus tremelimumab (anti-CTLA4) in patients with advanced HCC. Scientists and physicians alike are eagerly awaiting results of other ongoing phase III trials such as LEAP-002 (NCT03713593) evaluating pembrolizumab and levantinib in combination and COSMIC-312 (NCT03755791) investigating atezolizumab and cabozantinib synergistically.



## BEYOND CHECKPOINT INHIBITORS

### **Cell based immunotherapy**

In recent years cancer therapy has begun shifting focus from the aforementioned conventional therapies, to the use of immune cell therapies, which utilize the host immune system to target and treat cancer. One such avenue of therapy includes gene modified T cell therapy, particularly TCR engineered T cells and CAR-T cells, which have shown promise in various malignancies and continue to be tested in clinical trials targeting HCC. Below we will discuss the concept of gene modified T cell therapies, and other modalities of adoptive cell transfer while reporting the results of recent and ongoing clinical trials where applicable.

When engineering T-cell based immune cell therapies targeting HCC, identification of tumor-associated antigens (TAAs) that can allow for appropriately mediated immune response is critical. AFP is a glycoprotein composed of 591 amino acids, identified as the first oncofetal biomarker for patients with HCC allowing for quantitative estimation of tumor burden and response to therapy[41]. Elevated levels of AFP have been found in approximately 70% of patients diagnosed with HCC, however elevations can also be found in other pathological conditions including cirrhosis, various hepatic disorders, germ cell tumors, lung cancer, gastric cancer, and pancreatic cancer[42]. Pre-clinical models have demonstrated the potential of AFP in the development of cellular immunotherapies[43,44]. Glypican-3 (GPC-3) belongs to the transmembrane heparan sulfate proteoglycan family, that regulates cellular division and growth[45]. GPC-3 expression is elevated in HCC, and recent studies have demonstrated that elevated levels correlate with an overall worse prognosis[46]. Therefore GPC-3 has been investigated in pre-clinical studies as a target for adoptive cell immunotherapy[47,48]. Melanoma antigen gene proteins (MAGE) was first identified in melanoma patients and has been found to be almost exclusively expressed in a variety of cancer tissues. Aberrant expression of MAGE has been demonstrated to significantly correlate with clinical characteristics of HCC, however functions of the multitude of MAGE proteins have yet to be thoroughly understood [49]. New York esophageal squamous cell carcinoma (NY-ESO-1) is a member of the cancer testis antigen family expressed in a variety of cancer cells including HCC[50]. NY-ESO-1 has been deemed as the most promising cancer testis antigen for the development of cancer immunotherapy with a multitude of studies demonstrating promising results across a variety of malignancies[51]. When examining HCC caused by viral infection (HBV or HCV) there is prospect in targeting viral antigens as a strategy to etiologically treat HCC. Targets include HBV S or L protein (envelope proteins), which have shown promise by eliminating HBV positive hepatocytes when targeted by antigen specific T cells[52]. Targeting of hepatitis B surface antigen has also shown efficacy in mouse models utilizing CAR-T cell therapy as well[53]. Other tumor antigens that are of particular interest include epithelial cell adhesion molecules (EpCAM)[54], mucin 1 glycoprotein (MUC1)[55] and human telomerase reverse transcriptase (hTERT)[56].

### **CAR-T and TCR engineered T cells: The basics**

Gene modified T cell therapy is a method used to deliver T cells that are genetically engineered to produce TCRs that recognize specific tumor associated antigens and their epitopes. There are currently two methods of developing gene modified T cells, CAR-T cells and TCR transgenic T-cells (engineered T-cells), which have both shown efficacy and promise in a variety of solid and hematological malignancies[57]. CAR is composed of three basic elements: (1) the extracellular antigen recognition domain, (2) the transmembrane domain and (3) the intracellular signaling domain. The extracellular portion is a single chain antigen recognition receptor composed of heavy and light chains of a monoclonal antibody specific to the tumor surface antigen, allowing for no restrictions of target antigen recognition by major histocompatibility complex (MHC) molecules. The intracellular portion is formed by combining co-stimulatory molecules to the intracellular portion of TCRs[58]. The basic process of CAR-T generation includes: (1) T cell isolation from peripheral blood mononuclear cells using immunoselective beads and activation with anti-CD3 and IL-2, (2) genetic modification of CAR-T, (3) expansion of T cells *in vitro*, (4) evaluation ensuring CAR expression and T cell viability, and (5) CAR-T infusion back into the patient[59]. TCR transgenic T cells possess a TCR on the surface consisting of two distinct protein chains; alpha and beta, which bind to the MHC of antigen presenting cells, allowing for a highly specific interaction which is not limited to membrane bound antigens[60,61]. As previously described the selection and validation of target antigens is of utmost importance when

designing both TCR-T cells and CAR-T cells that are efficacious and minimize on target/off-tumor side effects[62].

### **CAR-T immunotherapy**

CAR-T cell therapy has been tested both preclinically and clinically when targeting HCC. Below we will discuss findings of completed and ongoing clinical studies that seek to demonstrate safety and efficacy of cellular therapy. Shi *et al*[63] recently published results from two prospective phase I trials involving GPC-CAR-T cells (NCT02395250 and NCT03146234) in patients with advanced GPC3 positive relapsed or refractory HCC following chemotherapeutic induced lymphodepletion. A total of 13 patients received CAR-T cell infusion, with two partial responses and one patient with sustained stable disease after 44.2 mo. OS rates at 3 years, 1 year, and 6 mo were 10.5%, 42% and 50.3% respectively with median OS duration of 278 d according to the Kaplan-Meier method. Unfortunately, toxic effects were noted with major concern as one patient developed cytokine release syndrome (CRS) leading to death from multi-organ failure[63]. A recently completed, open-label, phase I-II study by Dai *et al*[64] demonstrated efficacy of CD133-CAR-T cell therapy in patients with biopsy proven HCC (BCLC stage C) not amenable to curative treatment. One patient demonstrated a partial response, while 66.7% of patients had stable disease after infusion with an overall PFS of 6.8 mo and median OS of 12 mo (NCT02541370). Currently an open-label, single center, phase I-II study is underway investigating CAR-T/TCR-T cell immunotherapy targeting a variety of different malignancies including hepatoma (NCT03638206). A list of currently ongoing clinical trials investigating CAR-T cells for the treatment of HCC is listed in (Table 1).

### **Improving CAR-T delivery and efficacy**

Proven clinical efficacy of CAR-T across a variety of hematological malignancies has led to the investigation of its use in solid tumors. However, treatment of solid tumors poses many challenges to clinical investigators, one of which includes access to the tumor site. Many cases of HCC occur on the background of a fibrosed and cirrhotic liver, and the presence of such fibrotic extracellular matrix (ECM) poses a barrier for CAR-T cell penetration. In order to overcome such a challenge, CAR-T cells co-expressing heparinase, which degrades ECM heparan sulfate proteoglycans were developed. These newly engineered CAR-T cells improved cellular ability to degrade the ECM and displayed antitumor activity, suggesting a feasible strategy for the development of CAR-T cells targeting stroma-rich solid tumors[65]. Similar strategies are being investigated with the use of matrix metalloproteinase 8, which can allow CAR-T cells to overcome physical barriers[66]. Local administration of immunotherapy directly into tumors has shown promising results for organ selective delivery. In a study conducted by Katz *et al*[67], not only were CAR-T cells detected in biopsies of liver metastasis, but effective CAR-T cellular response was noted within the tumor following delivery *via* hepatic artery infusion. Such results have paved light into the investigation of direct tumor inoculation *via* a variety of local administration techniques including hepatic artery infusion in a multitude of clinical trials. Novel delivery platforms including nanoparticles, scaffolds and other biomaterials are also being investigated as a safe and effective way to deliver immunotherapy to tumor cells while avoiding off-target adverse effects[68]. Recent findings from the 2020 study conducted by Ma *et al*[69], demonstrated superior targeting and anti-tumor capabilities of CAR-T cell membrane coated nanoparticles both *in vivo* and *in vitro*.

In addition to tumor location being inaccessible, the tumor microenvironment has been found to be detrimental to CAR-T cells in regard to their survival and proliferation. The consumption of glucose by malignant cells renders the surrounding environment hypoxic, acidic, and low in nutrients leading to oxidative stress; leaving glycolytic T cells unable to adequately function, ultimately hindering the immune response[70,71]. In order to help eliminate reactive oxygen species (ROS) accumulation, CAR-T cells co expressing catalase were designed, and were subsequently able to survive and function in such unfavorable environments[72]. However, oxidative stress with release of compounds such as ROS and hydrogen peroxide, not only prevents recruitment and adequate function/persistence of immune cells, it also allows for recruitment of immune suppressor cells that lead to CAR-T cell exhaustion [73]. Strategies to help overcome such exhaustion include the use of ICIs that target PD-1, PD-L1, CTLA-4 in conjunction to CAR-T cells administration[74], and by genetically deleting T-cell PD-1 protein with the use of CRISPR-CAS9 gene editing [75]. There is optimism that these findings coupled with continued advancements in biomedical engineering will allow for new delivery and tumor microenvironment altering capabilities that can help amplify immunotherapeutic response in the

**Table 1** Ongoing clinical trials investigating chimeric antigen receptor-T cell therapy in the treatment of hepatocellular carcinoma according to ClinicalTrials.gov

Product	Phase	Estimated completion	Study/identifier
GPC3- CAR-T	I	November 2021	NCT04121273
CD147- CAR-T	I	May 2022	NCT03993743
GPC3- CAR-T	I	June 2022	NCT03980288
CAR-CLD18	N/A	December 2023	NCT03302403
GPC3- CAR-T	I	May 2024	NCT03884751
GPC3/TGF-CAR-T	I	August 2024	NCT03198546
GPC3- CAR-T	I	October 2036	NCT02905188

CAR: Chimeric antigen receptor; CD: Cluster of differentiation; GPC3: Glypican-3; TGF: Transforming growth factor.

treatment of HCC.

### Improving CAR-T safety

When utilizing cellular immunotherapy with CAR-T cells, a number of important safety concerns are to be noted including CRS, central nervous system toxicity (CNS) and the “on target-off tumor” effect. CRS is characterized by flu like symptoms including fever, fatigue and headache, which can progress to hypotension and organ dysfunction secondary to uncontrolled systemic inflammatory response associated with elevated serum cytokines; particularly interleukin (IL)-6, IL-10, tumor necrosis factor  $\alpha$  (TNF- $\alpha$ ) and interferon (IFN)- $\gamma$ [76]. Symptoms of CNS toxicity can range from headache and dizziness, to memory loss and delirium, and can be found along with CRS or after the resolution of CRS. Although the exact etiology remains unclear at this time, it is speculated that various cytokines may alter the blood brain barrier[77], in conjunction with an increase in protein, white blood cells, IFN- $\gamma$ , TNF- $\alpha$ , and IL-6 within cerebrospinal fluid[78].

These adverse reactions stem from CAR-T cells recognizing TAAs, which are not only heterogenous amongst tumors, but can also be present in normal tissues, elevating the risk of off target toxicity[71]. Such toxic effects have been reported following administration of anti-ERBB2 CAR-T cells for the treatment of metastatic malignancy, which led to CRS with subsequent respiratory distress and death due to low levels of ectopic ERBB2 in lung epithelium[79]. In order to help overcome such toxicities, a variety of strategies are being investigated. One such strategy includes the controlled removal of CAR-T cells with the use of suicide genes. There has been promise with the use of a drug inducible caspase 9 system (pro-apoptotic molecule that activates downstream pro-apoptotic caspase 3 when chemically induced) leading to apoptosis of CAR-T cells on demand[80] and with a system utilizing a truncated epidermal growth factor receptor, which allows for elimination following administration of a receptor specific antibody[81]. More recently, Amatya *et al*[82] developed CAR-T cells with a caspase 9 suicide gene for the treatment of multiple myeloma. Following administration of a dimerizing agent, CAR-T cells underwent apoptosis and were no longer active.

As previously mentioned, identification of a suitable TAA is imperative in the development of efficacious CAR-T cells, however highly specific TAAs are rarely identified, leading to investigation of dual- targeted CAR-T cells. Chen *et al*[83] developed dual-target CAR-T cells (targeting GPC3 and Asialoglycoprotein receptor 1) for the treatment of HCC. Not only did these cells demonstrate superior anticancer activity coupled with higher cytokine secretion when compared to single target CAR-T cells, but their specificity toward cellular targets may reduce the risk of on target- off tumor toxicity[83]. Recent developments have shown efficacy of a novel, split anti-GPC3 CAR-T, into two components with the use of SpyCatcher (extracellular 116 amino acid sequence linked to the intracellular domain) and SpyTag (13 amino acid peptide linked to single chain variable fragment). Such dual component delivery of CAR-T cells not only suppressed tumor growth, but displayed decreased cytokine release, representing a safer alternative when compared with conventional CAR-T cells[84]. Other perspectives include formation of a split, universal and programmable CAR system that allows for enhancement of specificity, safety and programmability of

CAR delivery systems[85].

### TCR engineered T cells

TCR engineered T cells are modified T cells that are able to precisely recognize tumor surface antigen peptides and MHC, rendering a response only effective when tumor cells express both antigen epitopes and MHC molecules[57]. TCR engineered T cells specific to NY-ESO-1 have demonstrated significant antitumor effects in the treatment of multiple myeloma[86] and synovial sarcomas[87]. The presence of NY-ESO-1 overexpression in HCC has made it an investigated target when developing engineered T cells in one phase II study (NCT01967823), and two phase I studies (NCT02869217 and (NCT03159585), for which we eagerly await results. In addition, TCR engineered T cells targeting hepatitis B viral antigens in HBV related HCC have been of interest in both preclinical and clinical models[88,89]. Recently in November 2020, Chen *et al*[90] presented an abstract at the 2020 Society for Immunotherapy of Cancer, demonstrating safety and efficacy of HBV specific TCR T-cells in recurrent HBV related HCC patients post liver transplant (NCT02719782).

There are currently five ongoing trials investigating TCR engineered cells targeting various TAAs in HCC (Table 2). All trials are investigating efficacy in advanced, unresectable HCC, with the exception of (NCT03899415), which is evaluating the safety and clinical benefit of TCR engineered T cells in HBV positive HCC patients post hepatectomy or radiofrequency ablation.

### Cytokine induced killer cells

Cytokine induced killer (CIK) cell therapy is a form of adoptive cell transfer that has been investigated as adjuvant therapy in patients with HCC. CIK cells are isolated from the patient's peripheral mononuclear cells, at which point they are grown *ex vivo* with the addition of cytokines such as recombinant human IFN- $\gamma$ , anti-CD3 monoclonal antibody, and recombinant human IL-2 prior to transfusion back into the patient; allowing for recognition and subsequent death of tumor cells[91]. In a multicenter, randomized phase III trial conducted in Korea, 230 patients were assigned to receive adjuvant CIK or no adjuvant therapy (control) following treatment with surgical resection, radiofrequency ablation or percutaneous ethanol injection (NCT00699816). Median RFS was significantly prolonged in the immunotherapy group (44 mo *vs* 30 mo,  $P = 0.01$ ), coupled with lower hazard ratios for all cause death ( $P = 0.08$ ) and cancer-related death ( $P = 0.02$ )[92]. A 5-year follow-up study demonstrated continued efficacy of autologous CIK immunotherapy without any repeated injections (NCT01890291). After an average 68.5 mo follow up, RFS was 44.8% in the immunotherapy group and 33.1% in the control group, coupled with a significantly lower risk of all-cause death (hazard ratio 0.33, 95%CI: 0.15-0.76,  $P = 0.006$ )[93]. Similar results were demonstrated in a meta-analysis and systemic review conducted by Cai *et al*[94] examining adjuvant CIK after invasive treatments for HCC. Pooled analysis demonstrated a significant improvement of RFS, PFS and OS in patients receiving autologous CIK immunotherapy. Although many studies have been conducted examining the safety and efficacy of CIK as an adjuvant treatment for HCC with mixed results; research is limited when examining CIK therapy as a sole treatment for unresectable HCC.

### TILs

TIL therapy is an immunotherapeutic technique that involves isolation and subsequent cell culture followed by autologous administration of tumor specific T cells present in infiltrated tumors. Success of adoptive cell therapy using TIL is based on the diverse antigenic specificity displayed by TIL cells toward the tumor and the lytic capabilities possessed allowing for eradication of malignancy[95]. TILs play an essential role in cancer progression[96] and even as a predictive biomarker of response to neoadjuvant chemotherapy in a variety of malignancies[97]. After the recognition of tumor associated antigens[98], various types of TILs infiltrate to the tumor site; including NK cells, NK T cells, mucosal-associated invariant T cells, and gamma delta T cells[99]. CD4+ T helper cells on the other hand further differentiate in the periphery into a variety of subsets (*i.e.*, Th1, Th2, Th9, Th17, Th22, regulatory T cells, and T follicular helper cells) [100]. CD8+ and NK cells contribute to a direct cytotoxic antitumor effect induced by a cascade of activating and inactivating receptors[99].

In a study conducted by Chew *et al*[101], examining immune gene expression profiles of resected tumor, HCC patient survival was positively correlated to higher expression of inflammatory immune genes. In conjunction, there was an increased presence of NK cells and T cells promoting apoptosis and reducing proliferation in the



**Table 2 Ongoing clinical trials investigating T cell receptor engineered T cells in the treatment of hepatocellular carcinoma according to ClinicalTrials.gov**

Product/target	Phase	Estimated completion	Study/identifier
C-TCR055/AFP	I	April 2021	NCT03971747
C-TCR055/AFP	I	November 2021	NCT04368182
HBV-TCR T cell/HBV Ag	I	June 2024	NCT03899415
IMA202-101/MAGE A1	I	June 2024	NCT03441100
AFP T cells/ AFP	I	June 2026	NCT03132792

AFP: Alpha fetoprotein; HBV: Hepatitis B virus; Ag: Antigen; MAGE: Melanoma antigen gene protein.

tumors of patients with longer survival, suggesting that TILs can be used as a specific immunotherapy for treatment of HCC[101]. In a hallmark randomized trial, patients who had undergone curative resection for HCC were assigned to receive adoptive immunotherapy *vs* no adjuvant therapy. Administration of lymphocyte infusion led to a reduced frequency of recurrence by 18%, significantly longer time to first recurrence ( $P = 0.008$ ), longer RFS ( $P = 0.01$ ) and disease-specific survival ( $P = 0.04$ ) when compared to controls[102]. A phase I trial (NCT01462903) examined the use of autologous TILs in HCC patients post resection. Results demonstrated that 80% of patients remained disease free after 14-mo follow-up, while displaying a favorable side effect profile, further suggesting TIL therapy as a safe treatment method for HCC[103]. Currently an active clinical trial is investigating the safety and efficacy of autologous TILs in patients with high-risk recurrent HCC (NCT04538313). Research is limited on the use of TIL immunotherapy for unresectable HCC, and we hope that further studies can be initiated using TILs alone or in combination with other therapies such as chemotherapy or ICIs.

### NK cells

NK cells play an active and critical role in the innate and adaptive immune defense against viral infections and hepatocellular malignancy[104], forming as much as 50% of innate immunity cell infiltrate within the liver[105]. Several clinical studies have demonstrated the safety and efficacy of allogenic NK cell adoptive immunotherapy in the treatment of various malignancies[106,107] including HCC[108]. In a study conducted by Lin *et al*[108], patients were enrolled to receive cryoablation alone *vs* allogenic NK cell therapy for the treatment of unresectable HCC. Results demonstrated synergistic effects of combination therapy leading to enhanced immune function, reduction in AFP, and improved quality of life in addition to an increased median PFS (9.1 mo *vs* 7.6 mo,  $P = 0.01$ ), higher response rate (60% *vs* 46.1%,  $P < 0.05$ ) and DCR (85.7% *vs* 69.2%,  $P < 0.01$ ) when compared to cryoablation alone. Similar results were demonstrated in a study conducted by Alnaggar *et al*[109], which investigated allogenic NK cell immunotherapy in combination with irreversible electroporation for Stage IV unresectable HCC. Efficacy of treatment was synergistic, with statistically significant lower AFP levels at 1 and 3 mo after treatment ( $P < 0.01$ ), coupled with a higher median OS (10.1 mo *vs* 8.9 mo,  $P = 0.0078$ ) when comparing combo therapies *vs* electroporation alone. Currently a phase I/II study investigating allogenic NK cell therapy in patients with advanced HCC (NCT04162158) is underway, as well as a phase II/III study examining the effects of autologous NK therapy when coupled with transcatheter arterial chemoembolization (TACE) for the treatment of advanced HCC (NCT04011033). Combination therapy of “off the shelf” FT500 NK cells with ICIs is also being investigated in subjects with advanced solid tumors including HCC (NCT03841110).

The advent of genetic modification techniques has led to the use of CAR technology to develop CAR-NK cells to improve specificity and efficacy of NK cell cytotoxicity [110]. Yu *et al*[111] developed a GPC3-specific NK cell for the treatment of HCC, which demonstrated *in vitro* cytotoxicity and cytokine production in addition to potent anti-tumor activities in HCC xenografts. Similar treatment benefits were noted with the development of c-MET specific CAR-NK cells, which demonstrated specific cytotoxicity against malignant hepatic cells *in vitro*[112]. While there are many ongoing clinical trials investigating CAR-NK immunotherapy for the treatment of hematological malignancies; investigation of treatment efficacy in solid tumors remains



scarce.

### **DC-CIK cells**

DC are antigen presenting cells that bridge the gap between innate and adaptive immunity, allowing for activation of naïve T cells that promote tumor-specific T cell immune response and adaptive immunity[113]. The use of *in vitro* generated DC combined with CIK cells has shown to be an effective and promising immunotherapy technique when used alone or in combination with other treatment options for a variety of malignancies[114]. In a study conducted by Zhou *et al*[115], advanced HCC patients were assigned to receive sorafenib alone *vs* a combination of sorafenib and DC-CIK immunotherapy. Results not only demonstrated a significant reduction in AFP levels when using combination therapy, but a significant increase in clinical benefit rate (41.9% *vs* 88.6%,  $P < 0.05$ ) and prolonged median survival time (13.8 m *vs* 18.6 m,  $P < 0.05$ ) while maintaining safety. Examination of combined therapy of TACE with DC-CIK therapy *vs* TACE alone has shown statistically prolonged OS time in HCC patients with HBV[116]. A meta-analysis of 22 studies, conducted in 2019 by Cao *et al*[117], has shown a prolonged survival and reduced recurrence rate of HCC when combining conventional clinical treatment and immunotherapy with DC and/or CIKs. There are currently studies recruiting participants for treatment of various solid tumors (including HCC) with DC-CIK combination immunotherapy (NCT04214717, NCT04476641, NCT03190811).

### **HCC vaccines**

Current advancements in medicine and immunobiology have demonstrated promising treatment strategies of cancer vaccines and oncolytic immunotherapy in patients with advanced stage cancer, leading to a variety of studies under clinical investigation [118]. Modalities include DC vaccinations, vaccines that target the antigens on tumor cells which in return activate cellular and humoral immunity causing a phagocytic activity towards the tumor cells, and oncolytic viruses that can attack and destroy tumor cells. These vaccines have both prophylactic and therapeutic aspects that can aid in the treatment of advanced cases of HCC. An up-to-date list of cancer vaccines and oncolytic virus therapies under current clinical investigation is provided (Table 3).

### **Antigen peptide vaccines**

Targeting these unique antigens can contribute to a significant approach in the development of vaccines in cases of HCC. Foremost antigens include AFP, GPC3, NY-ESO-1, hTERT, and hepatocellular and melanoma antigen gene-A (*MAGE-A*) as previously mentioned[119]. GPC3 peptide, a heparan sulfate glycoprotein and member of the GPC proteoglycan group, functions through attaching to the cell membrane by a glycosphosphatidylinositol anchor, further regulating several growth factors through the Wnts, hedgehog, and the BMP signaling pathways[120]. This peptide can be an ideal target for HCC vaccine development, as GPC3 is detected in the vast majority of HCC cases[121]. In a phase I trial conducted on 33 patients to ensure the safety of the GPC3-derived peptide vaccine for advanced HCC, results indicated a well-tolerated vaccine with significant immune response, and an improvement in the median OS rate, 12.2 mo (95%CI: 6.5-18.0) in patients with high GPC3-specific cytotoxic T-lymphocytes frequencies, as compared with 8.5 mo (95%CI: 3.7-13.1) in those with low GPC3-specific cytotoxic lymphocyte frequencies ( $P = 0.033$ )[122]. However, it was not determined if these tumor-infiltrating lymphocytes detected after vaccination were GPC3 peptide-specific cytotoxic lymphocytes, prompting the initiation of a phase II trial (UMIN000005093) in patients with advanced HCC having failed sorafenib, which demonstrated induction of GPC3 peptide specific cytotoxic lymphocytes capable of infiltrating tumors[123].

### **DC vaccines**

DCs have a potent effect on the T cell responses in HCC and other malignancies by acting as antigen presenting cells towards cytotoxic T lymphocytes[124]. Prior to administration of these vaccines to subjects, allogenic DCs are loaded with tumor antigens *ex vivo*, and then stimulated with cytokines such as granulocyte-macrophage colony-stimulating factor for further expansion[125]. In a recent 2016 study, DC vaccines loaded with autologous tumor cell lysate (Hepa1-6 cells) were studied using an orthotopic murine model of HCC. Treatment using this combination not only inhibited progression of murine HCC, but results were promising with a 90% survival rate compared to survival rate lower than 5% in untreated mice, suggesting alteration of the immunosuppressive tumor microenvironment as measured by lymphocyte and

**Table 3 Ongoing clinical trials investigating cancer vaccines and oncolytic viruses in the treatment of hepatocellular carcinoma according to ClinicalTrials.gov**

Product	Phase	Estimated completion	Study/identifier
OV telomelysin (OBP-301)	I	April 2021	NCT02293850
DC vaccine	II	April 2022	NCT04317248
DC vaccine + pneumococcal 13	I	May 2022	NCT03942328
OV M1-c6v1	I	October 2022	NCT04665362
DC vaccine	I	June 2023	NCT04147078
DSP-7888 PV + nivolumab or pembrolizumab	I/II	February 2024	NCT03311334
DNAJB1-PRKACA fusion kinase vaccine	I	March 2024	NCT04248569
TAEK-VAC-HerBy vaccine	I/II	December 2024	NCT04246671

DC: Dendritic cell; OV: Oncolytic virus; PV: Peptide vaccine.

cytokine analysis[126]. Although DCs seem to be an attractive treatment modality, further research is required to establish better *in vitro* handling of DCs, better methods of cellular delivery as well as further adaptation to the in-vivo environment following cellular administration.

### Oncolytic virus therapy

The use of exogenously administered viruses has paved way as a new and promising modality of targeting cancer cells and promoting neoplastic destruction following replication and subsequent lysis of tumor cells. Such actions allow for the release of antigens after lysis, which triggers antitumor activity[29]. One such virus is Pexa-Vec (pexastimogene devacirepvec, JX-594), a modified pox virus expressing granulocyte-macrophage stimulating factor that activates both innate and adaptive immune responses, remaining as the leading oncolytic virus of interest when combating HCC [127,128]. A randomized phase II trial conducted in patients with advanced HCC, demonstrated dose-related significant improvement in OS at high doses (14.1 mo) compared to low-dose treatment (6.7 mo), coupled with tumor regression involving one complete response[129]. Current interest has sparked trials with combination of common therapies. We eagerly await the results of Pexa-Vec with nivolumab (NCT03071094), and Pexa-Vec with sorafenib (PHOCUS; NCT02562755) for treatment of patients with advanced HCC.

## CONCLUSION

Treatment of HCC has made significant strides over the past decade since the approval of sorafenib as the first line, standard of care, molecular based treatment in 2007. More recently over the past few years four novel drugs: lenvatinib, ramucirumab, regorafenib, cabozantinib, have proven to be both safe and efficacious leading to approval for clinical use as an alternative or in addition to sorafenib. Clinical trials investigating the check point inhibitors atezolizumab, ipilimumab, nivolumab, and pembrolizumab have proven efficacy and identified new treatment modalities for HCC. As discussed in the paper adoptive immunotherapy including CAR-T cells, TCR engineered T cells, CIK cells, TIL cells, NK cells, DCs, and various vaccines have proven to be a promising therapeutic strategy for the treatment of HCC, however there are still many obstacles that need to be overcome prior to clinical application. Such challenges include identification of the ideal targeted antigen, overcoming the heterogenous and immunosuppressive environment of HCC, ensuring adequate delivery and persistence of immune cells, avoiding toxicity, and continued development of safe and cost-effective measures for cellular based treatment production and subsequent delivery to patients. The differences displayed between hepatic tumors amongst individuals across various stages of liver disease can possibly explain the difference in both clinical effects and adverse outcomes of immunotherapies studied. Therefore, we believe that the future of HCC treatment lies in the prospect of combined treatment modalities to exhibit synergistic effects, which have already entered various clinical trials as described

within this paper. While most completed and ongoing studies focus on individuals with adequate underlying hepatic function (Child-Pugh Class A), it is imperative to expand studies to other patient subgroups including those with early-stage disease and those undergoing palliative treatment. We believe that continued investigation over the coming years will overcome current barriers and lead to the development of novel and multifactorial immunotherapeutic strategies allowing for great advances in the treatment of HCC.

## REFERENCES

- 1 **Kim E**, Viatour P. Hepatocellular carcinoma: old friends and new tricks. *Exp Mol Med* 2020; **52**: 1898-1907 [PMID: [33268834](#) DOI: [10.1038/s12276-020-00527-1](#)]
- 2 **Janevska D**, Chaloska-Ivanova V, Janevski V. Hepatocellular Carcinoma: Risk Factors, Diagnosis and Treatment. *Open Access Maced J Med Sci* 2015; **3**: 732-736 [PMID: [27275318](#) DOI: [10.3889/oamjms.2015.111](#)]
- 3 **Sung H**, Ferlay J, Siegel RL, Laversanne M, Soerjomataram I, Jemal A, Bray F. Global cancer statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J Clin* 2021 [PMID: [33538338](#) DOI: [10.3322/caac.21660](#)]
- 4 **Bray F**, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, Jemal A. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J Clin* 2018; **68**: 394-424 [PMID: [30207593](#) DOI: [10.3322/caac.21492](#)]
- 5 **Yang JD**, Altekruse SF, Nguyen MH, Gores GJ, Roberts LR. Impact of country of birth on age at the time of diagnosis of hepatocellular carcinoma in the United States. *Cancer* 2017; **123**: 81-89 [PMID: [27571320](#) DOI: [10.1002/cncr.30246](#)]
- 6 **Yang M**, Parikh ND, Liu H, Wu E, Rao H, Feng B, Lin A, Wei L, Lok AS. Incidence and risk factors of hepatocellular carcinoma in patients with hepatitis C in China and the United States. *Sci Rep* 2020; **10**: 20922 [PMID: [33262356](#) DOI: [10.1038/s41598-020-77515-y](#)]
- 7 **Maida M**, Orlando E, Cammà C, Cabibbo G. Staging systems of hepatocellular carcinoma: a review of literature. *World J Gastroenterol* 2014; **20**: 4141-4150 [PMID: [24764652](#) DOI: [10.3748/wjg.v20.i15.4141](#)]
- 8 **Dhir M**, Melin AA, Douaiher J, Lin C, Zhen WK, Hussain SM, Geschwind JF, Doyle MB, Abou-Alfa GK, Are C. A Review and Update of Treatment Options and Controversies in the Management of Hepatocellular Carcinoma. *Ann Surg* 2016; **263**: 1112-1125 [PMID: [26813914](#) DOI: [10.1097/SLA.0000000000001556](#)]
- 9 **Llovet JM**, Zucman-Rossi J, Pikarsky E, Sangro B, Schwartz M, Sherman M, Gores G. Hepatocellular carcinoma. *Nat Rev Dis Primers* 2016; **2**: 16018 [PMID: [27158749](#) DOI: [10.1038/nrdp.2016.18](#)]
- 10 **Kinoshita A**, Onoda H, Fushiya N, Koike K, Nishino H, Tajiri H. Staging systems for hepatocellular carcinoma: Current status and future perspectives. *World J Hepatol* 2015; **7**: 406-424 [PMID: [25848467](#) DOI: [10.4254/wjh.v7.i3.406](#)]
- 11 **Gbolahan OB**, Schacht MA, Beckley EW, LaRoche TP, O'Neil BH, Pyko M. Locoregional and systemic therapy for hepatocellular carcinoma. *J Gastrointest Oncol* 2017; **8**: 215-228 [PMID: [28480062](#) DOI: [10.21037/jgo.2017.03.13](#)]
- 12 **Václav T**. Surgical treatment of hepatocellular carcinoma. *Klin Onkol* 2020; **33**: 30-33 [PMID: [33213163](#) DOI: [10.14735/amko20203S30](#)]
- 13 **Jadlowiec CC**, Taner T. Liver transplantation: Current status and challenges. *World J Gastroenterol* 2016; **22**: 4438-4445 [PMID: [27182155](#) DOI: [10.3748/wjg.v22.i18.4438](#)]
- 14 **Rimassa L**, Pressiani T, Merle P. Systemic Treatment Options in Hepatocellular Carcinoma. *Liver Cancer* 2019; **8**: 427-446 [PMID: [31799201](#) DOI: [10.1159/000499765](#)]
- 15 **Sim HW**, Knox J. Hepatocellular carcinoma in the era of immunotherapy. *Curr Probl Cancer* 2018; **42**: 40-48 [PMID: [29150141](#) DOI: [10.1016/j.crrprobcancer.2017.10.007](#)]
- 16 **Kudo M**. Lenvatinib May Drastically Change the Treatment Landscape of Hepatocellular Carcinoma. *Liver Cancer* 2018; **7**: 1-19 [PMID: [29662829](#) DOI: [10.1159/000487148](#)]
- 17 **Llovet JM**, Ricci S, Mazzaferro V, Hilgard P, Gane E, Blanc JF, de Oliveira AC, Santoro A, Raoul JL, Forner A, Schwartz M, Porta C, Zeuzem S, Bolondi L, Greten TF, Galle PR, Seitz JF, Borbath I, Häussinger D, Giannaris T, Shan M, Moscovici M, Voliotis D, Bruix J; SHARP Investigators Study Group. Sorafenib in advanced hepatocellular carcinoma. *N Engl J Med* 2008; **359**: 378-390 [PMID: [18650514](#) DOI: [10.1056/NEJMoa0708857](#)]
- 18 **Cheng AL**, Kang YK, Chen Z, Tsao CJ, Qin S, Kim JS, Luo R, Feng J, Ye S, Yang TS, Xu J, Sun Y, Liang H, Liu J, Wang J, Tak WY, Pan H, Burock K, Zou J, Voliotis D, Guan Z. Efficacy and safety of sorafenib in patients in the Asia-Pacific region with advanced hepatocellular carcinoma: a phase III randomised, double-blind, placebo-controlled trial. *Lancet Oncol* 2009; **10**: 25-34 [PMID: [19095497](#) DOI: [10.1016/S1470-2045\(08\)70285-7](#)]
- 19 **Alqahtani A**, Khan Z, Alloghbi A, Said Ahmed TS, Ashraf M, Hammouda DM. Hepatocellular Carcinoma: Molecular Mechanisms and Targeted Therapies. *Medicina (Kaunas)* 2019; **55** [PMID: [31450841](#) DOI: [10.3390/medicina55090526](#)]
- 20 **Al-Salama ZT**, Syed YY, Scott LJ. Lenvatinib: A Review in Hepatocellular Carcinoma. *Drugs*

- 2019; **79**: 665-674 [PMID: [30993651](#) DOI: [10.1007/s40265-019-01116-x](#)]
- 21 **Kudo M**, Finn RS, Qin S, Han KH, Ikeda K, Piscaglia F, Baron A, Park JW, Han G, Jassem J, Blanc JF, Vogel A, Komov D, Evans TRJ, Lopez C, Dutcus C, Guo M, Saito K, Kraljevic S, Tamai T, Ren M, Cheng AL. Lenvatinib vs sorafenib in first-line treatment of patients with unresectable hepatocellular carcinoma: a randomised phase 3 non-inferiority trial. *Lancet* 2018; **391**: 1163-1173 [PMID: [29433850](#) DOI: [10.1016/S0140-6736\(18\)30207-1](#)]
  - 22 **Personeni N**, Pressiani T, Rimassa L. Lenvatinib for the treatment of unresectable hepatocellular carcinoma: evidence to date. *J Hepatocell Carcinoma* 2019; **6**: 31-39 [PMID: [30775342](#) DOI: [10.2147/JHC.S168953](#)]
  - 23 **Zhu AX**, Park JO, Ryoo BY, Yen CJ, Poon R, Pastorelli D, Blanc JF, Chung HC, Baron AD, Pfiffer TE, Okusaka T, Kubackova K, Trojan J, Sastre J, Chau I, Chang SC, Abada PB, Yang L, Schwartz JD, Kudo M; REACH Trial Investigators. Ramucirumab vs placebo as second-line treatment in patients with advanced hepatocellular carcinoma following first-line therapy with sorafenib (REACH): a randomised, double-blind, multicentre, phase 3 trial. *Lancet Oncol* 2015; **16**: 859-870 [PMID: [26095784](#) DOI: [10.1016/S1470-2045\(15\)00050-9](#)]
  - 24 **Zhu AX**, Kang YK, Yen CJ, Finn RS, Galle PR, Llovet JM, Assenat E, Brandi G, Lim HY, Pracht M, Rau KM, Merle P, Motomura K, Ohno I, Daniele B, Shin D, Gerken G, Abada P, Hsu Y, Kudo M. REACH-2: A randomized, double-blind, placebo-controlled phase 3 study of ramucirumab vs placebo as second-line treatment in patients with advanced hepatocellular carcinoma (HCC) and elevated baseline alpha-fetoprotein (AFP) following first-line sorafenib. *J Clin Oncol* 2018; **36**: 4003 [DOI: [10.1200/JCO.2018.36.15\\_suppl.4003](#)]
  - 25 **Cerrito L**, Ponziani FR, Garcovich M, Tortora A, Annicchiarico BE, Pompili M, Siciliano M, Gasbarrini A. Regorafenib: a promising treatment for hepatocellular carcinoma. *Expert Opin Pharmacother* 2018; **19**: 1941-1948 [PMID: [30345837](#) DOI: [10.1080/14656566.2018.1534956](#)]
  - 26 **Bruix J**, Qin S, Merle P, Granito A, Huang YH, Bodoky G, Pracht M, Yokosuka O, Rosmorduc O, Breder V, Gerolami R, Masi G, Ross PJ, Song T, Bronowicki JP, Ollivier-Hourmand I, Kudo M, Cheng AL, Llovet JM, Finn RS, LeBerre MA, Baumhauer A, Meinhardt G, Han G; RESORCE Investigators. Regorafenib for patients with hepatocellular carcinoma who progressed on sorafenib treatment (RESORCE): a randomised, double-blind, placebo-controlled, phase 3 trial. *Lancet* 2017; **389**: 56-66 [PMID: [27932229](#) DOI: [10.1016/S0140-6736\(16\)32453-9](#)]
  - 27 **Abou-Alfa GK**, Meyer T, Cheng AL, El-Khoueiry AB, Rimassa L, Ryoo BY, Cicin I, Merle P, Chen Y, Park JW, Blanc JF, Bolondi L, Klumpen HJ, Chan SL, Zagonel V, Pressiani T, Ryu MH, Venook AP, Hessel C, Borgman-Hagey AE, Schwab G, Kelley RK. Cabozantinib in Patients with Advanced and Progressing Hepatocellular Carcinoma. *N Engl J Med* 2018; **379**: 54-63 [PMID: [29972759](#) DOI: [10.1056/NEJMoa1717002](#)]
  - 28 **Hargadon KM**, Johnson CE, Williams CJ. Immune checkpoint blockade therapy for cancer: An overview of FDA-approved immune checkpoint inhibitors. *Int Immunopharmacol* 2018; **62**: 29-39 [PMID: [29990692](#) DOI: [10.1016/j.intimp.2018.06.001](#)]
  - 29 **Nakano S**, Eso Y, Okada H, Takai A, Takahashi K, Seno H. Recent Advances in Immunotherapy for Hepatocellular Carcinoma. *Cancers (Basel)* 2020; **12** [PMID: [32218257](#) DOI: [10.3390/cancers12040775](#)]
  - 30 **Gao Q**, Wang XY, Qiu SJ, Yamato I, Sho M, Nakajima Y, Zhou J, Li BZ, Shi YH, Xiao YS, Xu Y, Fan J. Overexpression of PD-L1 significantly associates with tumor aggressiveness and postoperative recurrence in human hepatocellular carcinoma. *Clin Cancer Res* 2009; **15**: 971-979 [PMID: [19188168](#) DOI: [10.1158/1078-0432.CCR-08-1608](#)]
  - 31 **Fritz JM**, Lenardo MJ. Development of immune checkpoint therapy for cancer. *J Exp Med* 2019; **216**: 1244-1254 [PMID: [31068379](#) DOI: [10.1084/jem.20182395](#)]
  - 32 **Lee MS**, Ryoo BY, Hsu CH, Numata K, Stein S, Verret W, Hack SP, Spahn J, Liu B, Abdullah H, Wang Y, He AR, Lee KH; GO30140 investigators. Atezolizumab with or without bevacizumab in unresectable hepatocellular carcinoma (GO30140): an open-label, multicentre, phase 1b study. *Lancet Oncol* 2020; **21**: 808-820 [PMID: [32502443](#) DOI: [10.1016/S1470-2045\(20\)30156-X](#)]
  - 33 **Finn RS**, Qin S, Ikeda M, Galle PR, Ducreux M, Kim TY, Kudo M, Breder V, Merle P, Kaseb AO, Li D, Verret W, Xu DZ, Hernandez S, Liu J, Huang C, Mulla S, Wang Y, Lim HY, Zhu AX, Cheng AL; IMbrave150 Investigators. Atezolizumab plus Bevacizumab in Unresectable Hepatocellular Carcinoma. *N Engl J Med* 2020; **382**: 1894-1905 [PMID: [32402160](#) DOI: [10.1056/NEJMoa1915745](#)]
  - 34 **Hack SP**, Spahn J, Chen M, Cheng AL, Kaseb A, Kudo M, Lee HC, Yopp A, Chow P, Qin S. IMbrave 050: a Phase III trial of atezolizumab plus bevacizumab in high-risk hepatocellular carcinoma after curative resection or ablation. *Future Oncol* 2020; **16**: 975-989 [PMID: [32352320](#) DOI: [10.2217/fon-2020-0162](#)]
  - 35 **Yau T**, Kang YK, Kim TY, El-Khoueiry AB, Santoro A, Sangro B, Melero I, Kudo M, Hou MM, Matilla A, Tovoli F, Knox JJ, Ruth He A, El-Rayes BF, Acosta-Rivera M, Lim HY, Neely J, Shen Y, Wisniewski T, Anderson J, Hsu C. Efficacy and Safety of Nivolumab Plus Ipilimumab in Patients With Advanced Hepatocellular Carcinoma Previously Treated With Sorafenib: The CheckMate 040 Randomized Clinical Trial. *JAMA Oncol* 2020 [PMID: [33001135](#) DOI: [10.1001/jamaoncol.2020.4564](#)]
  - 36 **El-Khoueiry AB**, Sangro B, Yau T, Crocenzi TS, Kudo M, Hsu C, Kim TY, Choo SP, Trojan J, Welling TH Rd, Meyer T, Kang YK, Yeo W, Chopra A, Anderson J, Dela Cruz C, Lang L, Neely J, Tang H, Dastani HB, Melero I. Nivolumab in patients with advanced hepatocellular carcinoma



- (CheckMate 040): an open-label, non-comparative, phase 1/2 dose escalation and expansion trial. *Lancet* 2017; **389**: 2492-2502 [PMID: [28434648](#) DOI: [10.1016/S0140-6736\(17\)31046-2](#)]
- 37 **Yau T**, Park J, Finn R, Cheng AL, Mathurin P, Edeline J, Kudo M, Han KH, Harding J, Merle P. CheckMate 459: A randomized, multi-center phase III study of nivolumab (NIVO) vs sorafenib (SOR) as first-line (1L) treatment in patients (pts) with advanced hepatocellular carcinoma (aHCC). *Ann Oncol* 2019; **30**: v874-v875 [DOI: [10.1093/annonc/mdz394.029](#)]
  - 38 **Kwok G**, Yau TC, Chiu JW, Tse E, Kwong YL. Pembrolizumab (Keytruda). *Hum Vaccin Immunother* 2016; **12**: 2777-2789 [PMID: [27398650](#) DOI: [10.1080/21645515.2016.1199310](#)]
  - 39 **Zhu AX**, Finn RS, Edeline J, Cattani S, Ogasawara S, Palmer D, Verslype C, Zagonel V, Fartoux L, Vogel A, Sarker D, Verset G, Chan SL, Knox J, Daniele B, Webber AL, Ebbinghaus SW, Ma J, Siegel AB, Cheng AL, Kudo M; KEYNOTE-224 investigators. Pembrolizumab in patients with advanced hepatocellular carcinoma previously treated with sorafenib (KEYNOTE-224): a non-randomised, open-label phase 2 trial. *Lancet Oncol* 2018; **19**: 940-952 [PMID: [29875066](#) DOI: [10.1016/S1470-2045\(18\)30351-6](#)]
  - 40 **Finn RS**, Ryoo BY, Merle P, Kudo M, Bouattour M, Lim HY, Breder V, Edeline J, Chao Y, Ogasawara S, Yau T, Garrido M, Chan SL, Knox J, Daniele B, Ebbinghaus SW, Chen E, Siegel AB, Zhu AX, Cheng AL; KEYNOTE-240 investigators. Pembrolizumab As Second-Line Therapy in Patients With Advanced Hepatocellular Carcinoma in KEYNOTE-240: A Randomized, Double-Blind, Phase III Trial. *J Clin Oncol* 2020; **38**: 193-202 [PMID: [31790344](#) DOI: [10.1200/JCO.19.01307](#)]
  - 41 **Özdemir F**, Baskiran A. The Importance of AFP in Liver Transplantation for HCC. *J Gastrointest Cancer* 2020; **51**: 1127-1132 [PMID: [32845425](#) DOI: [10.1007/s12029-020-00486-w](#)]
  - 42 **Mizejewski GJ**. Levels of alpha-fetoprotein during pregnancy and early infancy in normal and disease states. *Obstet Gynecol Surv* 2003; **58**: 804-826 [PMID: [14668662](#) DOI: [10.1097/01.OGX.0000099770.97668.18](#)]
  - 43 **Zhu W**, Peng Y, Wang L, Hong Y, Jiang X, Li Q, Liu H, Huang L, Wu J, Celis E, Merchen T, Kruse E, He Y. Identification of  $\alpha$ -fetoprotein-specific T-cell receptors for hepatocellular carcinoma immunotherapy. *Hepatology* 2018; **68**: 574-589 [PMID: [29443377](#) DOI: [10.1002/hep.29844](#)]
  - 44 **Liu H**, Xu Y, Xiang J, Long L, Green S, Yang Z, Zimdahl B, Lu J, Cheng N, Horan LH, Liu B, Yan S, Wang P, Diaz J, Jin L, Nakano Y, Morales JF, Zhang P, Liu LX, Staley BK, Priceman SJ, Brown CE, Forman SJ, Chan VW, Liu C. Targeting Alpha-Fetoprotein (AFP)-MHC Complex with CAR T-Cell Therapy for Liver Cancer. *Clin Cancer Res* 2017; **23**: 478-488 [PMID: [27535982](#) DOI: [10.1158/1078-0432.CCR-16-1203](#)]
  - 45 **Montalbano M**, Georgiadis J, Masterson AL, McGuire JT, Prajapati J, Shirafkan A, Rastellini C, Cicalese L. Biology and function of glypican-3 as a candidate for early cancerous transformation of hepatocytes in hepatocellular carcinoma (Review). *Oncol Rep* 2017; **37**: 1291-1300 [PMID: [28098909](#) DOI: [10.3892/or.2017.5387](#)]
  - 46 **Kaseb AO**, Hassan M, Lacin S, Abdel-Wahab R, Amin HM, Shalaby A, Wolff RA, Yao J, Rashid A, Vennapusa B, Feng J, Ohtomo T. Evaluating clinical and prognostic implications of Glypican-3 in hepatocellular carcinoma. *Oncotarget* 2016; **7**: 69916-69926 [PMID: [27655712](#) DOI: [10.18632/oncotarget.12066](#)]
  - 47 **Zhou F**, Shang W, Yu X, Tian J. Glypican-3: A promising biomarker for hepatocellular carcinoma diagnosis and treatment. *Med Res Rev* 2018; **38**: 741-767 [PMID: [28621802](#) DOI: [10.1002/med.21455](#)]
  - 48 **Jiang Z**, Jiang X, Chen S, Lai Y, Wei X, Li B, Lin S, Wang S, Wu Q, Liang Q, Liu Q, Peng M, Yu F, Weng J, Du X, Pei D, Liu P, Yao Y, Xue P, Li P. Anti-GPC3-CAR T Cells Suppress the Growth of Tumor Cells in Patient-Derived Xenografts of Hepatocellular Carcinoma. *Front Immunol* 2016; **7**: 690 [PMID: [28123387](#) DOI: [10.3389/fimmu.2016.00690](#)]
  - 49 **Li R**, Gong J, Xiao C, Zhu S, Hu Z, Liang J, Li X, Yan X, Zhang X, Li D, Liu W, Chong Y, Jie Y. A comprehensive analysis of the MAGE family as prognostic and diagnostic markers for hepatocellular carcinoma. *Genomics* 2020; **112**: 5101-5114 [PMID: [32941982](#) DOI: [10.1016/j.ygeno.2020.09.026](#)]
  - 50 **Gordeeva O**. Cancer-testis antigens: Unique cancer stem cell biomarkers and targets for cancer therapy. *Semin Cancer Biol* 2018; **53**: 75-89 [PMID: [30171980](#) DOI: [10.1016/j.semcancer.2018.08.006](#)]
  - 51 **Thomas R**, Al-Khadairi G, Roelands J, Hendrickx W, Dermime S, Bedognetti D, Decock J. NY-ESO-1 Based Immunotherapy of Cancer: Current Perspectives. *Front Immunol* 2018; **9**: 947 [PMID: [29770138](#) DOI: [10.3389/fimmu.2018.00947](#)]
  - 52 **Bohne F**, Chmielewski M, Ebert G, Wiegmann K, Kürschner T, Schulze A, Urban S, Krönke M, Abken H, Protzer U. T cells redirected against hepatitis B virus surface proteins eliminate infected hepatocytes. *Gastroenterology* 2008; **134**: 239-247 [PMID: [18166356](#) DOI: [10.1053/j.gastro.2007.11.002](#)]
  - 53 **Kruse RL**, Shum T, Tashiro H, Barzi M, Yi Z, Whitten-Bauer C, Legras X, Bissig-Choisat B, Garaigorta U, Gottschalk S, Bissig KD. HBsAg-redirection T cells exhibit antiviral activity in HBV-infected human liver chimeric mice. *Cytotherapy* 2018; **20**: 697-705 [PMID: [29631939](#) DOI: [10.1016/j.jcyt.2018.02.002](#)]
  - 54 **Krause J**, von Felden J, Casar C, Fründt TW, Galaski J, Schmidt C, Jung C, Itrich H, Weidemann SA, Krech T, Heumann A, Li J, Fischer L, Sauter G, Lohse AW, Wege H, Schulze K. Hepatocellular carcinoma: Intratumoral EpCAM-positive cancer stem cell heterogeneity identifies high-risk tumor



- subtype. *BMC Cancer* 2020; **20**: 1130 [PMID: [33225916](#) DOI: [10.1186/s12885-020-07580-z](#)]
- 55 **Ma Y**, Wang Z, Gong R, Li L, Wu H, Jin H. Specific cytotoxicity of MUC1 chimeric antigen receptor-engineered Jurkat T cells against hepatocellular carcinoma. *Acad J Second Mil Med Univ* 2014; **5**: 1177-1182 [DOI: [10.3724/SP.J.1008.2014.01177](#)]
  - 56 **Mizukoshi E**, Nakamoto Y, Marukawa Y, Arai K, Yamashita T, Tsuji H, Kuzushima K, Takiguchi M, Kaneko S. Cytotoxic T cell responses to human telomerase reverse transcriptase in patients with hepatocellular carcinoma. *Hepatology* 2006; **43**: 1284-1294 [PMID: [16729333](#) DOI: [10.1002/hep.21203](#)]
  - 57 **Mizukoshi E**, Kaneko S. Immune cell therapy for hepatocellular carcinoma. *J Hematol Oncol* 2019; **12**: 52 [PMID: [31142330](#) DOI: [10.1186/s13045-019-0742-5](#)]
  - 58 **Benmebarek MR**, Karches CH, Cadilha BL, Lesch S, Endres S, Kobold S. Killing Mechanisms of Chimeric Antigen Receptor (CAR) T Cells. *Int J Mol Sci* 2019; **20** [PMID: [30875739](#) DOI: [10.3390/ijms20061283](#)]
  - 59 **Chen Y**, E CY, Gong ZW, Liu S, Wang ZX, Yang YS, Zhang XW. Chimeric antigen receptor-engineered T-cell therapy for liver cancer. *Hepatobiliary Pancreat Dis Int* 2018; **17**: 301-309 [PMID: [29861325](#) DOI: [10.1016/j.hbpd.2018.05.005](#)]
  - 60 **Ping Y**, Liu C, Zhang Y. T-cell receptor-engineered T cells for cancer treatment: current status and future directions. *Protein Cell* 2018; **9**: 254-266 [PMID: [28108950](#) DOI: [10.1007/s13238-016-0367-1](#)]
  - 61 **Rochigneux P**, Chanez B, De Rauglaudre B, Mitry E, Chabannon C, Gilibert M. Adoptive Cell Therapy in Hepatocellular Carcinoma: Biological Rationale and First Results in Early Phase Clinical Trials. *Cancers (Basel)* 2021; **13** [PMID: [33450845](#) DOI: [10.3390/cancers13020271](#)]
  - 62 **Kershaw MH**, Westwood JA, Darcy PK. Gene-engineered T cells for cancer therapy. *Nat Rev Cancer* 2013; **13**: 525-541 [PMID: [23880905](#) DOI: [10.1038/nrc3565](#)]
  - 63 **Shi D**, Shi Y, Kaseb AO, Qi X, Zhang Y, Chi J, Lu Q, Gao H, Jiang H, Wang H, Yuan D, Ma H, Li Z, Zhai B. Chimeric Antigen Receptor-Glypican-3 T-Cell Therapy for Advanced Hepatocellular Carcinoma: Results of Phase I Trials. *Clin Cancer Res* 2020; **26**: 3979-3989 [PMID: [32371538](#) DOI: [10.1158/1078-0432.CCR-19-3259](#)]
  - 64 **Dai H**, Tong C, Shi D, Chen M, Guo Y, Chen D, Han X, Wang H, Wang Y, Shen P. Efficacy and biomarker analysis of CD133-directed CAR T cells in advanced hepatocellular carcinoma: a single-arm, open-label, phase II trial. *Oncoimmunology* 2020; **9**: 1846926 [PMID: [33312759](#) DOI: [10.1080/2162402X.2020.1846926](#)]
  - 65 **Caruana I**, Savoldo B, Hoyos V, Weber G, Liu H, Kim ES, Ittmann MM, Marchetti D, Dotti G. Heparanase promotes tumor infiltration and antitumor activity of CAR-redirectioned T lymphocytes. *Nat Med* 2015; **21**: 524-529 [PMID: [25849134](#) DOI: [10.1038/nm.3833](#)]
  - 66 **Mardomi A**, Abediankenari S. Matrix Metalloproteinase 8: Could it Benefit the CAR-T Cell Therapy of Solid Tumors? *Cancer Microenviron* 2018; **11**: 93-96 [PMID: [29589335](#) DOI: [10.1007/s12307-018-0208-2](#)]
  - 67 **Katz SC**, Burga RA, McCormack E, Wang LJ, Mooring W, Point GR, Khare PD, Thorn M, Ma Q, Stainken BF, Assanah EO, Davies R, Espat NJ, Junghans RP. Phase I Hepatic Immunotherapy for Metastases Study of Intra-Arterial Chimeric Antigen Receptor-Modified T-cell Therapy for CEA+ Liver Metastases. *Clin Cancer Res* 2015; **21**: 3149-3159 [PMID: [25850950](#) DOI: [10.1158/1078-0432.CCR-14-1421](#)]
  - 68 **Riley RS**, June CH, Langer R, Mitchell MJ. Delivery technologies for cancer immunotherapy. *Nat Rev Drug Discov* 2019; **18**: 175-196 [PMID: [30622344](#) DOI: [10.1038/s41573-018-0006-z](#)]
  - 69 **Ma W**, Zhu D, Li J, Chen X, Xie W, Jiang X, Wu L, Wang G, Xiao Y, Liu Z, Wang F, Li A, Shao D, Dong W, Liu W, Yuan Y. Coating biomimetic nanoparticles with chimeric antigen receptor T cell-membrane provides high specificity for hepatocellular carcinoma photothermal therapy treatment. *Theranostics* 2020; **10**: 1281-1295 [PMID: [31938065](#) DOI: [10.7150/thno.40291](#)]
  - 70 **Franchina DG**, Dostert C, Brenner D. Reactive Oxygen Species: Involvement in T Cell Signaling and Metabolism. *Trends Immunol* 2018; **39**: 489-502 [PMID: [29452982](#) DOI: [10.1016/j.it.2018.01.005](#)]
  - 71 **Dal Bo M**, De Mattia E, Baboci L, Mezzalana S, Cecchin E, Assaraf YG, Toffoli G. New insights into the pharmacological, immunological, and CAR-T-cell approaches in the treatment of hepatocellular carcinoma. *Drug Resist Updat* 2020; **51**: 100702 [PMID: [32371296](#) DOI: [10.1016/j.drug.2020.100702](#)]
  - 72 **Ligtenberg MA**, Mougiakakos D, Mukhopadhyay M, Witt K, Lladser A, Chmielewski M, Riet T, Abken H, Kiessling R. Coexpressed Catalase Protects Chimeric Antigen Receptor-Redirected T Cells as well as Bystander Cells from Oxidative Stress-Induced Loss of Antitumor Activity. *J Immunol* 2016; **196**: 759-766 [PMID: [26673145](#) DOI: [10.4049/jimmunol.1401710](#)]
  - 73 **Wu AA**, Drake V, Huang HS, Chiu S, Zheng L. Reprogramming the tumor microenvironment: tumor-induced immunosuppressive factors paralyze T cells. *Oncoimmunology* 2015; **4**: e1016700 [PMID: [26140242](#) DOI: [10.1080/2162402x.2015.1016700](#)]
  - 74 **Gargett T**, Yu W, Dotti G, Yvon ES, Christo SN, Hayball JD, Lewis ID, Brenner MK, Brown MP. GD2-specific CAR T Cells Undergo Potent Activation and Deletion Following Antigen Encounter but can be Protected From Activation-induced Cell Death by PD-1 Blockade. *Mol Ther* 2016; **24**: 1135-1149 [PMID: [27019998](#) DOI: [10.1038/mt.2016.63](#)]
  - 75 **Rupp LJ**, Schumann K, Roybal KT, Gate RE, Ye CJ, Lim WA, Marson A. CRISPR/Cas9-mediated PD-1 disruption enhances anti-tumor efficacy of human chimeric antigen receptor T cells. *Sci Rep*

- 2017; **7**: 737 [PMID: [28389661](#) DOI: [10.1038/s41598-017-00462-8](#)]
- 76 **Shimabukuro-Vornhagen A**, Gödel P, Subklewe M, Stemmler HJ, Schlößer HA, Schlaak M, Kochanek M, Böll B, von Bergwelt-Baildon MS. Cytokine release syndrome. *J Immunother Cancer* 2018; **6**: 56 [PMID: [29907163](#) DOI: [10.1186/s40425-018-0343-9](#)]
- 77 **Yáñez L**, Sánchez-Escamilla M, Perales MA. CAR T Cell Toxicity: Current Management and Future Directions. *Hemasphere* 2019; **3**: e186 [PMID: [31723825](#) DOI: [10.1097/HS9.0000000000000186](#)]
- 78 **Gust J**, Taraseviciute A, Turtle CJ. Neurotoxicity Associated with CD19-Targeted CAR-T Cell Therapies. *CNS Drugs* 2018; **32**: 1091-1101 [PMID: [30387077](#) DOI: [10.1007/s40263-018-0582-9](#)]
- 79 **Morgan RA**, Yang JC, Kitano M, Dudley ME, Laurencot CM, Rosenberg SA. Case report of a serious adverse event following the administration of T cells transduced with a chimeric antigen receptor recognizing ERBB2. *Mol Ther* 2010; **18**: 843-851 [PMID: [20179677](#) DOI: [10.1038/mt.2010.24](#)]
- 80 **Gargett T**, Brown MP. The inducible caspase-9 suicide gene system as a "safety switch" to limit on-target, off-tumor toxicities of chimeric antigen receptor T cells. *Front Pharmacol* 2014; **5**: 235 [PMID: [25389405](#) DOI: [10.3389/fphar.2014.00235](#)]
- 81 **Paszkiewicz PJ**, Fräßle SP, Srivastava S, Sommermeyer D, Hudecek M, Drexler I, Sadelain M, Liu L, Jensen MC, Riddell SR, Busch DH. Targeted antibody-mediated depletion of murine CD19 CAR T cells permanently reverses B cell aplasia. *J Clin Invest* 2016; **126**: 4262-4272 [PMID: [27760047](#) DOI: [10.1172/JCI84813](#)]
- 82 **Amatya C**, Pegues MA, Lam N, Vanasse D, Geldres C, Choi S, Hewitt SM, Feldman SA, Kochenderfer JN. Development of CAR T Cells Expressing a Suicide Gene Plus a Chimeric Antigen Receptor Targeting Signaling Lymphocytic-Activation Molecule F7. *Mol Ther* 2021; **29**: 702-717 [PMID: [33129371](#) DOI: [10.1016/j.ymthe.2020.10.008](#)]
- 83 **Chen C**, Li K, Jiang H, Song F, Gao H, Pan X, Shi B, Bi Y, Wang H, Li Z. Development of T cells carrying two complementary chimeric antigen receptors against glypican-3 and asialoglycoprotein receptor 1 for the treatment of hepatocellular carcinoma. *Cancer Immunol Immunother* 2017; **66**: 475-489 [PMID: [28035433](#) DOI: [10.1007/s00262-016-1949-8](#)]
- 84 **Liu X**, Wen J, Yi H, Hou X, Yin Y, Ye G, Wu X, Jiang X. Split chimeric antigen receptor-modified T cells targeting glypican-3 suppress hepatocellular carcinoma growth with reduced cytokine release. *Ther Adv Med Oncol* 2020; **12**: 1758835920910347 [PMID: [32215059](#) DOI: [10.1177/1758835920910347](#)]
- 85 **Cho JH**, Collins JJ, Wong WW. Universal Chimeric Antigen Receptors for Multiplexed and Logical Control of T Cell Responses. *Cell* 2018; **173**: 1426-1438.e11 [PMID: [29706540](#) DOI: [10.1016/j.cell.2018.03.038](#)]
- 86 **Rapoport AP**, Stadtmauer EA, Binder-Scholl GK, Goloubeva O, Vogl DT, Lacey SF, Badros AZ, Garfall A, Weiss B, Finklestein J, Kulikovskaya I, Sinha SK, Kronsberg S, Gupta M, Bond S, Melchiori L, Brewer JE, Bennett AD, Gerry AB, Pumphrey NJ, Williams D, Tayton-Martin HK, Ribeiro L, Holdich T, Yanovich S, Hardy N, Yared J, Kerr N, Philip S, Westphal S, Siegel DL, Levine BL, Jakobsen BK, Kalos M, June CH. NY-ESO-1-specific TCR-engineered T cells mediate sustained antigen-specific antitumor effects in myeloma. *Nat Med* 2015; **21**: 914-921 [PMID: [26193344](#) DOI: [10.1038/nm.3910](#)]
- 87 **D'Angelo SP**, Melchiori L, Merchant MS, Bernstein D, Glod J, Kaplan R, Grupp S, Tap WD, Chagin K, Binder GK, Basu S, Lowther DE, Wang R, Bath N, Tipping A, Betts G, Ramachandran I, Navenot JM, Zhang H, Wells DK, Van Winkle E, Kari G, Trivedi T, Holdich T, Pandite L, Amado R, Mackall CL. Antitumor Activity Associated with Prolonged Persistence of Adoptively Transferred NY-ESO-1<sup>e259</sup>T Cells in Synovial Sarcoma. *Cancer Discov* 2018; **8**: 944-957 [PMID: [29891538](#) DOI: [10.1158/2159-8290.CD-17-1417](#)]
- 88 **Gehring AJ**, Xue SA, Ho ZZ, Teoh D, Ruedl C, Chia A, Koh S, Lim SG, Maini MK, Stauss H, Bertoletti A. Engineering virus-specific T cells that target HBV infected hepatocytes and hepatocellular carcinoma cell lines. *J Hepatol* 2011; **55**: 103-110 [PMID: [21145860](#) DOI: [10.1016/j.jhep.2010.10.025](#)]
- 89 **Tan AT**, Yang N, Lee Krishnamoorthy T, Oei V, Chua A, Zhao X, Tan HS, Chia A, Le Bert N, Low D, Tan HK, Kumar R, Irani FG, Ho ZZ, Zhang Q, Guccione E, Wai LE, Koh S, Hwang W, Chow WC, Bertoletti A. Use of Expression Profiles of HBV-DNA Integrated into Genomes of Hepatocellular Carcinoma Cells to Select T Cells for Immunotherapy. *Gastroenterology* 2019; **156**: 1862-1876.e9 [PMID: [30711630](#) DOI: [10.1053/j.gastro.2019.01.251](#)]
- 90 **Chen W**, Cheng J, Zheng X, Yang F, Fam R, Koh S, Wai L-E, Wang T, Bertoletti A, Zhang Q. 273 Phase I study of LioCyx-M, autologous hepatitis B virus (HBV)-specific T cell receptor (TCR) T-cells, in recurrent HBV-related hepatocellular carcinoma (HCC) post-liver transplantation. *J Immunother Cancer* 2020; **8**: A167-A167 [DOI: [10.1136/jitc-2020-SITC2020.0273](#)]
- 91 **Zhang R**, Zhang Z, Liu Z, Wei D, Wu X, Bian H, Chen Z. Adoptive cell transfer therapy for hepatocellular carcinoma. *Front Med* 2019; **13**: 3-11 [PMID: [30659408](#) DOI: [10.1007/s11684-019-0684-x](#)]
- 92 **Lee JH**, Lee JH, Lim YS, Yeon JE, Song TJ, Yu SJ, Gwak GY, Kim KM, Kim YJ, Lee JW, Yoon JH. Adjuvant immunotherapy with autologous cytokine-induced killer cells for hepatocellular carcinoma. *Gastroenterology* 2015; **148**: 1383-91.e6 [PMID: [25747273](#) DOI: [10.1053/j.gastro.2015.02.055](#)]
- 93 **Lee JH**, Lee JH, Lim YS, Yeon JE, Song TJ, Yu SJ, Gwak GY, Kim KM, Kim YJ, Lee JW, Yoon

- JH. Sustained efficacy of adjuvant immunotherapy with cytokine-induced killer cells for hepatocellular carcinoma: an extended 5-year follow-up. *Cancer Immunol Immunother* 2019; **68**: 23-32 [PMID: 30232520 DOI: 10.1007/s00262-018-2247-4]
- 94 **Cai XR**, Li X, Lin JX, Wang TT, Dong M, Chen ZH, Jia CC, Hong YF, Lin Q, Wu XY. Autologous transplantation of cytokine-induced killer cells as an adjuvant therapy for hepatocellular carcinoma in Asia: an update meta-analysis and systematic review. *Oncotarget* 2017; **8**: 31318-31328 [PMID: 28412743 DOI: 10.18632/oncotarget.15454]
- 95 **Forget MA**, Tavera RJ, Haymaker C, Ramachandran R, Malu S, Zhang M, Wardell S, Fulbright OJ, Toth CL, Gonzalez AM, Thorsen ST, Flores E, Wahl A, Peng W, Amaria RN, Hwu P, Bernatchez C. A Novel Method to Generate and Expand Clinical-Grade, Genetically Modified, Tumor-Infiltrating Lymphocytes. *Front Immunol* 2017; **8**: 908 [PMID: 28824634 DOI: 10.3389/fimmu.2017.00908]
- 96 **Gomez Perdiguero E**, Geissmann F. Cancer immunology. Identifying the infiltrators. *Science* 2014; **344**: 801-802 [PMID: 24855239 DOI: 10.1126/science.1255117]
- 97 **Zeng DQ**, Yu YF, Ou QY, Li XY, Zhong RZ, Xie CM, Hu QG. Prognostic and predictive value of tumor-infiltrating lymphocytes for clinical therapeutic research in patients with non-small cell lung cancer. *Oncotarget* 2016; **7**: 13765-13781 [PMID: 26871598 DOI: 10.18632/oncotarget.7282]
- 98 **Kumar S**, Skeen MJ, Adiri Y, Yoon H, Vezys VD, Lukacher AE, Evavold BD, Ziegler HK, Boss JM. A cytokine promoter/yellow fluorescent protein reporter transgene serves as an early activation marker of lymphocyte subsets. *Cell Immunol* 2005; **237**: 131-140 [PMID: 16405934 DOI: 10.1016/j.cellimm.2005.11.003]
- 99 **Piñero Fernández J**, Luddy KA, Harmon C, O'Farrelly C. Hepatic Tumor Microenvironments and Effects on NK Cell Phenotype and Function. *Int J Mol Sci* 2019; **20** [PMID: 31450598 DOI: 10.3390/ijms20174131]
- 100 **Golubovskaya V**, Wu L. Different Subsets of T Cells, Memory, Effector Functions, and CAR-T Immunotherapy. *Cancers (Basel)* 2016; **8** [PMID: 26999211 DOI: 10.3390/cancers8030036]
- 101 **Chew V**, Tow C, Teo M, Wong HL, Chan J, Gehring A, Loh M, Bolze A, Quek R, Lee VK, Lee KH, Abastado JP, Toh HC, Nardin A. Inflammatory tumour microenvironment is associated with superior survival in hepatocellular carcinoma patients. *J Hepatol* 2010; **52**: 370-379 [PMID: 19720422 DOI: 10.1016/j.jhep.2009.07.013]
- 102 **Takayama T**, Sekine T, Makuuchi M, Yamasaki S, Kosuge T, Yamamoto J, Shimada K, Sakamoto M, Hirohashi S, Ohashi Y, Kakizoe T. Adoptive immunotherapy to lower postsurgical recurrence rates of hepatocellular carcinoma: a randomised trial. *Lancet* 2000; **356**: 802-807 [PMID: 11022927 DOI: 10.1016/s0140-6736(00)02654-4]
- 103 **Jiang SS**, Tang Y, Zhang YJ, Weng DS, Zhou ZG, Pan K, Pan QZ, Wang QJ, Liu Q, He J, Zhao JJ, Li J, Chen MS, Chang AE, Li Q, Xia JC. A phase I clinical trial utilizing autologous tumor-infiltrating lymphocytes in patients with primary hepatocellular carcinoma. *Oncotarget* 2015; **6**: 41339-41349 [PMID: 26515587 DOI: 10.18632/oncotarget.5463]
- 104 **Liu P**, Chen L, Zhang H. Natural Killer Cells in Liver Disease and Hepatocellular Carcinoma and the NK Cell-Based Immunotherapy. *J Immunol Res* 2018; **2018**: 1206737 [PMID: 30255103 DOI: 10.1155/2018/1206737]
- 105 **Gao B**, Jeong WI, Tian Z. Liver: An organ with predominant innate immunity. *Hepatology* 2008; **47**: 729-736 [PMID: 18167066 DOI: 10.1002/hep.22034]
- 106 **Lin M**, Alnaggar M, Liang S, Wang X, Liang Y, Zhang M, Chen J, Niu L, Xu K. An important discovery on combination of irreversible electroporation and allogeneic natural killer cell immunotherapy for unresectable pancreatic cancer. *Oncotarget* 2017; **8**: 101795-101807 [PMID: 29254205 DOI: 10.18632/oncotarget.21974]
- 107 **Lin M**, Liang SZ, Wang XH, Liang YQ, Zhang MJ, Niu LZ, Chen JB, Li HB, Xu KC. Clinical efficacy of percutaneous cryoablation combined with allogeneic NK cell immunotherapy for advanced non-small cell lung cancer. *Immunol Res* 2017; **65**: 880-887 [PMID: 28508945 DOI: 10.1007/s12026-017-8927-x]
- 108 **Lin M**, Liang S, Wang X, Liang Y, Zhang M, Chen J, Niu L, Xu K. Cryoablation combined with allogeneic natural killer cell immunotherapy improves the curative effect in patients with advanced hepatocellular cancer. *Oncotarget* 2017; **8**: 81967-81977 [PMID: 29137237 DOI: 10.18632/oncotarget.17804]
- 109 **Alnaggar M**, Lin M, Mesmar A, Liang S, Qaid A, Xu K, Chen J, Niu L, Yin Z. Allogeneic Natural Killer Cell Immunotherapy Combined with Irreversible Electroporation for Stage IV Hepatocellular Carcinoma: Survival Outcome. *Cell Physiol Biochem* 2018; **48**: 1882-1893 [PMID: 30092590 DOI: 10.1159/000492509]
- 110 **Nowakowska P**, Romanski A, Miller N, Odendahl M, Bonig H, Zhang C, Seifried E, Wels WS, Tonn T. Clinical grade manufacturing of genetically modified, CAR-expressing NK-92 cells for the treatment of ErbB2-positive malignancies. *Cancer Immunol Immunother* 2018; **67**: 25-38 [PMID: 28879551 DOI: 10.1007/s00262-017-2055-2]
- 111 **Yu M**, Luo H, Fan M, Wu X, Shi B, Di S, Liu Y, Pan Z, Jiang H, Li Z. Development of GPC3-Specific Chimeric Antigen Receptor-Engineered Natural Killer Cells for the Treatment of Hepatocellular Carcinoma. *Mol Ther* 2018; **26**: 366-378 [PMID: 29339014 DOI: 10.1016/j.ymthe.2017.12.012]
- 112 **Liu B**, Liu ZZ, Zhou ML, Lin JW, Chen XM, Li Z, Gao WB, Yu ZD, Liu T. Development of cMETspecific chimeric antigen receptorengineered natural killer cells with cytotoxic effects on human liver cancer HepG2 cells. *Mol Med Rep* 2019; **20**: 2823-2831 [PMID: 31524233 DOI: 10.3892/mmr.2019.9711]

- 10.3892/mmr.2019.10529]
- 113 **Gardner A**, de Mingo Pulido Á, Ruffell B. Dendritic Cells and Their Role in Immunotherapy. *Front Immunol* 2020; **11**: 924 [PMID: 32508825 DOI: 10.3389/fimmu.2020.00924]
- 114 **Wang S**, Wang X, Zhou X, Lyerly HK, Morse MA, Ren J. DC-CIK as a widely applicable cancer immunotherapy. *Expert Opin Biol Ther* 2020; **20**: 601-607 [PMID: 32033522 DOI: 10.1080/14712598.2020.1728250]
- 115 **Zhou Z**, Qin H, Weng L, Ni Y. Clinical efficacy of DC-CIK combined with sorafenib in the treatment of advanced hepatocellular carcinoma. *J BUON* 2019; **24**: 615-621 [PMID: 31128014]
- 116 **Zhang J**, Li H, Gao D, Zhang B, Zheng M, Lun M, Wei M, Duan R, Guo M, Hua J, Liu Q, Bai J, Liu H, Zheng J, Yao H. A prognosis and impact factor analysis of DC-CIK cell therapy for patients with hepatocellular carcinoma undergoing postoperative TACE. *Cancer Biol Ther* 2018; **19**: 475-483 [PMID: 29400599 DOI: 10.1080/15384047.2018.1433501]
- 117 **Cao J**, Kong FH, Liu X, Wang XB. Immunotherapy with dendritic cells and cytokine-induced killer cells for hepatocellular carcinoma: A meta-analysis. *World J Gastroenterol* 2019; **25**: 3649-3663 [PMID: 31367163 DOI: 10.3748/wjg.v25.i27.3649]
- 118 **Guo ZS**, Lu B, Guo Z, Giehl E, Feist M, Dai E, Liu W, Storkus WJ, He Y, Liu Z, Bartlett DL. Vaccinia virus-mediated cancer immunotherapy: cancer vaccines and oncolytics. *J Immunother Cancer* 2019; **7**: 6 [PMID: 30626434 DOI: 10.1186/s40425-018-0495-7]
- 119 **Park R**, Eshrat F, Al-Jumayli M, Saeed A. Immuno-Oncotherapeutic Approaches in Advanced Hepatocellular Carcinoma. *Vaccines (Basel)* 2020; **8** [PMID: 32784389 DOI: 10.3390/vaccines8030447]
- 120 **Filmus J**, Capurro M, Rast J. Glypicans. *Genome Biol* 2008; **9**: 224 [PMID: 18505598 DOI: 10.1186/gb-2008-9-5-224]
- 121 **Nobuoka D**, Yoshikawa T, Sawada Y, Fujiwara T, Nakatsura T. Peptide vaccines for hepatocellular carcinoma. *Hum Vaccin Immunother* 2013; **9**: 210-212 [PMID: 23442593 DOI: 10.4161/hv.22473]
- 122 **Sawada Y**, Yoshikawa T, Nobuoka D, Shirakawa H, Kuronuma T, Motomura Y, Mizuno S, Ishii H, Nakachi K, Konishi M, Nakagohri T, Takahashi S, Gotohda N, Takayama T, Yamao K, Uesaka K, Furuse J, Kinoshita T, Nakatsura T. Phase I trial of a glypican-3-derived peptide vaccine for advanced hepatocellular carcinoma: immunologic evidence and potential for improving overall survival. *Clin Cancer Res* 2012; **18**: 3686-3696 [PMID: 22577059 DOI: 10.1158/1078-0432.CCR-11-3044]
- 123 **Shimizu Y**, Suzuki T, Yoshikawa T, Tsuchiya N, Sawada Y, Endo I, Nakatsura T. Cancer immunotherapy-targeted glypican-3 or neoantigens. *Cancer Sci* 2018; **109**: 531-541 [PMID: 29285841 DOI: 10.1111/cas.13485]
- 124 **Fan J**, Han J, Li J, Gu A, Yin D, Song F, Wang L, Yi Y. The expression and function of immunoglobulin-like transcript 4 in dendritic cells from patients with hepatocellular carcinoma. *Hum Immunol* 2020; **81**: 714-725 [PMID: 33228921 DOI: 10.1016/j.humimm.2020.10.003]
- 125 **Gu YZ**, Zhao X, Song XR. Ex vivo pulsed dendritic cell vaccination against cancer. *Acta Pharmacol Sin* 2020; **41**: 959-969 [PMID: 32366940 DOI: 10.1038/s41401-020-0415-5]
- 126 **Wang Q**, Luan W, Warren L, Kadri H, Kim KW, Goz V, Blank S, Isabel Fiel M, Hiotis SP. Autologous Tumor Cell Lysate-Loaded Dendritic Cell Vaccine Inhibited Tumor Progression in an Orthotopic Murine Model for Hepatocellular Carcinoma. *Ann Surg Oncol* 2016; **23**: 574-582 [PMID: 26786094 DOI: 10.1245/s10434-015-5035-9]
- 127 **Kim JH**, Oh JY, Park BH, Lee DE, Kim JS, Park HE, Roh MS, Je JE, Yoon JH, Thorne SH, Kim D, Hwang TH. Systemic armed oncolytic and immunologic therapy for cancer with JX-594, a targeted poxvirus expressing GM-CSF. *Mol Ther* 2006; **14**: 361-370 [PMID: 16905462 DOI: 10.1016/j.ymthe.2006.05.008]
- 128 **Parato KA**, Breitbach CJ, Le Boeuf F, Wang J, Storbeck C, Ilkow C, Diallo JS, Falls T, Burns J, Garcia V, Kanji F, Evgin L, Hu K, Paradis F, Knowles S, Hwang TH, Vanderhyden BC, Auer R, Kim DH, Bell JC. The oncolytic poxvirus JX-594 selectively replicates in and destroys cancer cells driven by genetic pathways commonly activated in cancers. *Mol Ther* 2012; **20**: 749-758 [PMID: 22186794 DOI: 10.1038/mt.2011.276]
- 129 **Heo J**, Reid T, Ruo L, Breitbach CJ, Rose S, Bloomston M, Cho M, Lim HY, Chung HC, Kim CW, Burke J, Lencioni R, Hickman T, Moon A, Lee YS, Kim MK, Daneshmand M, Dubois K, Longpre L, Ngo M, Rooney C, Bell JC, Rhee BG, Patt R, Hwang TH, Kim DH. Randomized dose-finding clinical trial of oncolytic immunotherapeutic vaccinia JX-594 in liver cancer. *Nat Med* 2013; **19**: 329-336 [PMID: 23396206 DOI: 10.1038/nm.3089]



Published by **Baishideng Publishing Group Inc**  
7041 Koll Center Parkway, Suite 160, Pleasanton, CA 94566, USA

**Telephone:** +1-925-3991568

**E-mail:** [bpgoffice@wjgnet.com](mailto:bpgoffice@wjgnet.com)

**Help Desk:** <https://www.f6publishing.com/helpdesk>

<https://www.wjgnet.com>

