

# World Journal of *Stem Cells*

*World J Stem Cells* 2021 December 26; 13(12): 1813-1946



**OPINION REVIEW**

- 1813 Stem cell-derived biofactors fight against coronavirus infection  
*Ardalan M, Chodari L, Zununi Vahed S, Hosseiniyan Khatibi SM, Eftekhari A, Davaran S, Cucchiarini M, Roshangar L, Ahmadian E*

**REVIEW**

- 1826 Application of mesenchymal stem cells derived from human pluripotent stem cells in regenerative medicine  
*Liu TM*
- 1845 Strategies to improve regenerative potential of mesenchymal stem cells  
*Choudhery MS*
- 1863 Dental mesenchymal stromal/stem cells in different microenvironments – implications in regenerative therapy  
*Okić-Đorđević I, Obradović H, Kukolj T, Petrović A, Mojsilović S, Bugarski D, Jauković A*
- 1881 Regulating the fate of stem cells for regenerating the intervertebral disc degeneration  
*Ekram S, Khalid S, Salim A, Khan I*

**ORIGINAL ARTICLE****Basic Study**

- 1905 Bone marrow mesenchymal stem cell therapy regulates gut microbiota to improve post-stroke neurological function recovery in rats  
*Zhao LN, Ma SW, Xiao J, Yang LJ, Xu SX, Zhao L*
- 1918 SmartFlare™ is a reliable method for assessing mRNA expression in single neural stem cells  
*Diana A, Setzu MD, Kokaia Z, Nat R, Maxia C, Murtas D*
- 1928 Urolithin a alleviates oxidative stress-induced senescence in nucleus pulposus-derived mesenchymal stem cells through SIRT1/PGC-1 $\alpha$  pathway  
*Shi PZ, Wang JW, Wang PC, Han B, Lu XH, Ren YX, Feng XM, Cheng XF, Zhang L*

**ABOUT COVER**

Editorial Board Member of *World Journal of Stem Cells*, Jyoti Anand Kode, MSc, PhD, Scientific Officer 'G', Kode Lab, Tumor Immunology and Immunotherapy Group; Anti-Cancer Drug Screening Facility, Advanced Centre for Treatment, Research and Education in Cancer (ACTREC), Tata Memorial Centre, Navi Mumbai 410210, Maharashtra, India. [jkode@actrec.gov.in](mailto:jkode@actrec.gov.in)

**AIMS AND SCOPE**

The primary aim of *World Journal of Stem Cells* (*WJSC*, *World J Stem Cells*) is to provide scholars and readers from various fields of stem cells with a platform to publish high-quality basic and clinical research articles and communicate their research findings online. *WJSC* publishes articles reporting research results obtained in the field of stem cell biology and regenerative medicine, related to the wide range of stem cells including embryonic stem cells, germline stem cells, tissue-specific stem cells, adult stem cells, mesenchymal stromal cells, induced pluripotent stem cells, embryonal carcinoma stem cells, hemangioblasts, lymphoid progenitor cells, etc.

**INDEXING/ABSTRACTING**

The *WJSC* is now indexed in Science Citation Index Expanded (also known as SciSearch®), Journal Citation Reports/Science Edition, Biological Abstracts, BIOSIS Previews, Scopus, PubMed, and PubMed Central. The 2021 Edition of Journal Citation Reports® cites the 2020 impact factor (IF) for *WJSC* as 5.326; IF without journal self cites: 5.035; 5-year IF: 4.956; Journal Citation Indicator: 0.55; Ranking: 14 among 29 journals in cell and tissue engineering; Quartile category: Q2; Ranking: 72 among 195 journals in cell biology; and Quartile category: Q2. The *WJSC*'s CiteScore for 2020 is 3.1 and Scopus CiteScore rank 2020: Histology is 31/60; Genetics is 205/325; Genetics (clinical) is 64/87; Molecular Biology is 285/382; Cell Biology is 208/279.

**RESPONSIBLE EDITORS FOR THIS ISSUE**

Production Editor: *Hua-Ge Yu*; Production Department Director: *Xu Guo*; Editorial Office Director: *Ze-Mao Gong*.

**NAME OF JOURNAL**

*World Journal of Stem Cells*

**ISSN**

ISSN 1948-0210 (online)

**LAUNCH DATE**

December 31, 2009

**FREQUENCY**

Monthly

**EDITORS-IN-CHIEF**

Shengwen Calvin Li, FRSM, FRSB, Carlo Ventura

**EDITORIAL BOARD MEMBERS**

<https://www.wjgnet.com/1948-0210/editorialboard.htm>

**PUBLICATION DATE**

December 26, 2021

**COPYRIGHT**

© 2021 Baishideng Publishing Group Inc

**INSTRUCTIONS TO AUTHORS**

<https://www.wjgnet.com/bpg/gerinfo/204>

**GUIDELINES FOR ETHICS DOCUMENTS**

<https://www.wjgnet.com/bpg/GerInfo/287>

**GUIDELINES FOR NON-NATIVE SPEAKERS OF ENGLISH**

<https://www.wjgnet.com/bpg/gerinfo/240>

**PUBLICATION ETHICS**

<https://www.wjgnet.com/bpg/GerInfo/288>

**PUBLICATION MISCONDUCT**

<https://www.wjgnet.com/bpg/gerinfo/208>

**ARTICLE PROCESSING CHARGE**

<https://www.wjgnet.com/bpg/gerinfo/242>

**STEPS FOR SUBMITTING MANUSCRIPTS**

<https://www.wjgnet.com/bpg/GerInfo/239>

**ONLINE SUBMISSION**

<https://www.f6publishing.com>

## Basic Study

# Bone marrow mesenchymal stem cell therapy regulates gut microbiota to improve post-stroke neurological function recovery in rats

Lin-Na Zhao, Song-Wen Ma, Jie Xiao, Li-Ji Yang, Shi-Xin Xu, Lan Zhao

**ORCID number:** Lin-Na Zhao 0000-0003-3918-6722; Song-Wen Ma 0000-0001-9934-2569; Jie Xiao 0000-0002-1579-2800; Li-Ji Yang 0000-0002-4604-408X; Shi-Xin Xu 0000-0003-2270-2911; Lan Zhao 0000-0002-7449-2947.

**Author contributions:** Zhao LN and Zhao L drafted and wrote the paper; Ma SW and Xiao J performed the experiments; Yang LJ performed the statistical analysis; Xu SX contributed to designing the experiments and revising the article.

**Institutional animal care and use committee statement:** All procedures involving animals were reviewed and approved by the Institutional Animal Care and Use Committee of the Experimental Animal Ethics Committee of Tianjin University of Traditional Chinese Medicine (IACUC protocol number: [Protocol No. TCM-LAEC2019038]).

**Conflict-of-interest statement:** The authors have no conflict of interest to declare.

**Data sharing statement:** No additional data are available.

**ARRIVE guidelines statement:** The

**Lin-Na Zhao, Song-Wen Ma, Jie Xiao, Li-Ji Yang, Shi-Xin Xu, Lan Zhao,** First Teaching Hospital of Tianjin University of Traditional Chinese Medicine, Tianjin 300381, China

**Lin-Na Zhao, Song-Wen Ma, Jie Xiao, Li-Ji Yang, Shi-Xin Xu, Lan Zhao,** National Clinical Research Center for Chinese Medicine Acupuncture and Moxibustion, Tianjin 300381, China

**Lin-Na Zhao, Shi-Xin Xu,** Tianjin Key Laboratory of Translational Research of TCM Prescription and Syndrome, Tianjin 300381, China

**Corresponding author:** Lan Zhao, PhD, Research Fellow, First Teaching Hospital of Tianjin University of Traditional Chinese Medicine, No. 88 Chang Ling Road, Xi Qing District, Tianjin 300381, China. [lanzhao69@163.com](mailto:lanzhao69@163.com)

## Abstract

### BACKGROUND

As a cellular mode of therapy, bone marrow mesenchymal stem cells (BMSCs) are used to treat stroke. However, their mechanisms in stroke treatment have not been established. Recent evidence suggests that regulation of dysregulated gut flora after stroke affects stroke outcomes.

### AIM

To investigate the effects of BMSCs on gut microbiota after ischemic stroke.

### METHODS

A total of 30 Sprague-Dawley rats were randomly divided into three groups, including sham operation control group, transient middle cerebral artery occlusion (MCAO) group, and MCAO with BMSC treatment group. The modified Neurological Severity Score (mNSS), beam walking test, and Morris water maze test were used to evaluate neurological function recovery after BMSC transplantation. Nissl staining was performed to elucidate on the pathology of nerve cells in the hippocampus. Feces from each group of rats were collected and analyzed by 16s rDNA sequencing.

### RESULTS

BMSC transplantation significantly reduced mNSS ( $P < 0.01$ ). Rats performed better in the beam walking test in the BMSC group than in the MCAO group ( $P <$

authors have read the ARRIVE Guidelines, and the manuscript was prepared and revised according to the ARRIVE Guidelines.

**Supported by** National Natural Science Foundation of China, No. 81774059 and No. 82074533; Tianjin Natural Science Foundation, No. 19JCZDJC37100.

**Country/Territory of origin:** China

**Specialty type:** Cell and tissue engineering

**Provenance and peer review:** Invited article; Externally peer reviewed.

**Peer-review model:** Single blind

**Peer-review report's scientific quality classification**

Grade A (Excellent): 0  
Grade B (Very good): 0  
Grade C (Good): 0  
Grade D (Fair): 0  
Grade E (Poor): 0

**Open-Access:** This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <https://creativecommons.org/licenses/by-nc/4.0/>

**Received:** May 5, 2021

**Peer-review started:** May 5, 2021

**First decision:** June 23, 2021

**Revised:** July 6, 2021

**Accepted:** December 11, 2021

**Article in press:** December 11, 2021

**Published online:** December 26, 2021

**P-Reviewer:** Jin W

**S-Editor:** Fan JR

**L-Editor:** Wang TQ

**P-Editor:** Fan JR

0.01). The Morris water maze test revealed that the BMSC treatment group exhibited a significant improvement in learning and memory. Nissl staining for neuronal damage assessment after stroke showed that in the BMSC group, cells were orderly arranged with significantly reduced necrosis. Moreover, BMSCs regulated microbial structure composition. In rats treated with BMSCs, the abundance of potential short-chain fatty acid producing bacteria and *Lactobacillus* was increased.

## CONCLUSION

BMSC transplantation is a potential therapeutic option for ischemic stroke, and it promotes neurological functions by regulating gut microbiota dysbiosis.

**Key Words:** Ischemic stroke; Bone marrow mesenchymal stem cells; Neurological function; Gut microbiota

©The Author(s) 2021. Published by Baishideng Publishing Group Inc. All rights reserved.

**Core Tip:** Bone marrow mesenchymal stem cell (BMSC) transplantation provides a novel approach for ischemic stroke therapy. Studies on the “gut-brain axis” indicate that gut microbiota dysbiosis affects stroke prognosis. We investigated the interactions between BMSCs and gut microbiota. Our findings indicate that the therapeutic mechanism of BMSCs on ischemic stroke treatment may involve the regulation of microbiome structure and function.

**Citation:** Zhao LN, Ma SW, Xiao J, Yang LJ, Xu SX, Zhao L. Bone marrow mesenchymal stem cell therapy regulates gut microbiota to improve post-stroke neurological function recovery in rats. *World J Stem Cells* 2021; 13(12): 1905-1917

**URL:** <https://www.wjgnet.com/1948-0210/full/v13/i12/1905.htm>

**DOI:** <https://dx.doi.org/10.4252/wjsc.v13.i12.1905>

## INTRODUCTION

Globally, stroke is a lethal disability-causing disease that affects up to 13 million people annually[1]. The latest data from the American Heart Association shows that in the United States, one person suffers a stroke after every 40 s[2]. Stroke patients exhibit recurrent attacks, which exerts a huge socio-economic burden on the society and families. Ischemic stroke is the most prevalent stroke type, accounting for 70%-80% of all stroke types[3]. Intravenous thrombolysis and endovascular thrombectomy are the primary treatment options for stroke. However, they are associated with time and technical limitations[4,5]. Therefore, it is important to develop novel therapeutic approaches for ischemic stroke.

Stem cell transplantation is considered a potential therapeutic strategy for patients after ischemic stroke[6]. Bone marrow mesenchymal stem cells (BMSCs) are a group of stem cells with various characteristics, including autologous harvesting, rapid proliferation, easy *in vitro* culture, and low immunogenicity. Moreover, they are not limited by ethical restrictions. BMSCs have the effects of neuroprotection, modulation of inflammation, immune responses, endogenous neurogenesis, and astrogenesis[7]. Specifically, their inflammatory regulatory function has been investigated in various inflammatory diseases.

An estimated 100 trillion microorganisms reside in the human gut. They are closely associated with human health and diseases[8]. The understanding of gut microbiota is only at the rudimentary stage; however, studies have confirmed the existence of bidirectional communication in the microbiota-gut-brain axis, which influences stroke treatment and prognosis[9-11]. After a stroke, the central nervous system (CNS) is injured, then, as a stress response mechanism, the hypothalamic-pituitary-adrenal axis triggers the release of adrenocorticotrophic hormone-releasing factor (CRF) and glucocorticoids[12]. Sympathetic and parasympathetic nerves directly affect gastrointestinal functions *via* communication with the enteric nervous system[10]. This induces suppressed gut motility, increased gut permeability, gut microbiota dysbiosis,



and immune cell activation. Studies have documented significant microbial diversity changes in feces of stroke patients[13,14]. Severe stroke destroys the intestinal barrier, therefore, commensal gut microbiota migrates to other organs; this is the primary cause of systemic infections after stroke[15]. A few bacterial species in gut microbiota or their metabolites regulate intestinal immunity, which regulates post-stroke immunity[16]. Animal model experiments have established that changing the gut microbiota improves the prognosis of stroke[17,18]. Despite the documented efficacy of stem cell therapy in altering the populations of gut microbiota in several inflammatory diseases, it has not determined whether it has a similar effect on ischemic stroke.

Therefore, we used a rat model of transient middle cerebral artery occlusion (MCAO) to investigate whether BMSCs can improve abnormal intestinal flora after ischemic stroke.

## MATERIALS AND METHODS

### Animals

Adult male Sprague-Dawley (SD) rats, 5-6 week old, weighing 220-250 g, were purchased from Beijing Huafukang Biotechnology Company (Beijing, China). The rats were housed in pathogen-free conditions under a 12 h-light/12 h-dark cycle at 25 °C. The Ethics Committee of Tianjin University of Traditional Chinese Medicine approved this study (approval number: TCM-LAEC2019038).

### BMSC isolation, culture, and identification

In this study, 4-wk-old SD rats were cervically dislocated. The femur and tibia were isolated and removed under sterile conditions. The Dulbecco's modified Eagle medium (DMEM) was used for flushing the bone marrow cavity, and the bone marrow flush was collected. The isolated cell suspension was sieved through a 200-mesh nylon sieve and then centrifuged (1000 r/min) for 10 min at 4 °C. The supernatant was discarded, and the cells were re-suspended with DMEM containing 10% fetal bovine serum (FBS; BI). The cell density was adjusted to  $2 \times 10^6$  cells into 25 cm<sup>2</sup> culture flasks and incubated in a cell incubator (37 °C, 5% CO<sub>2</sub>). The cells were passaged every 3-4 d, and the third-passage cells were used for further experiments. BMSCs were incubated with fluorescence antibodies, including CD90-PE, CD29-APC, CD45-PerCP, and CD31-FITC (1:100, Miltenyi, Germany), to identify the phenotype by flow cytometry (FACS Calibur, BD, San Jose, CA, United States).

### Experimental design

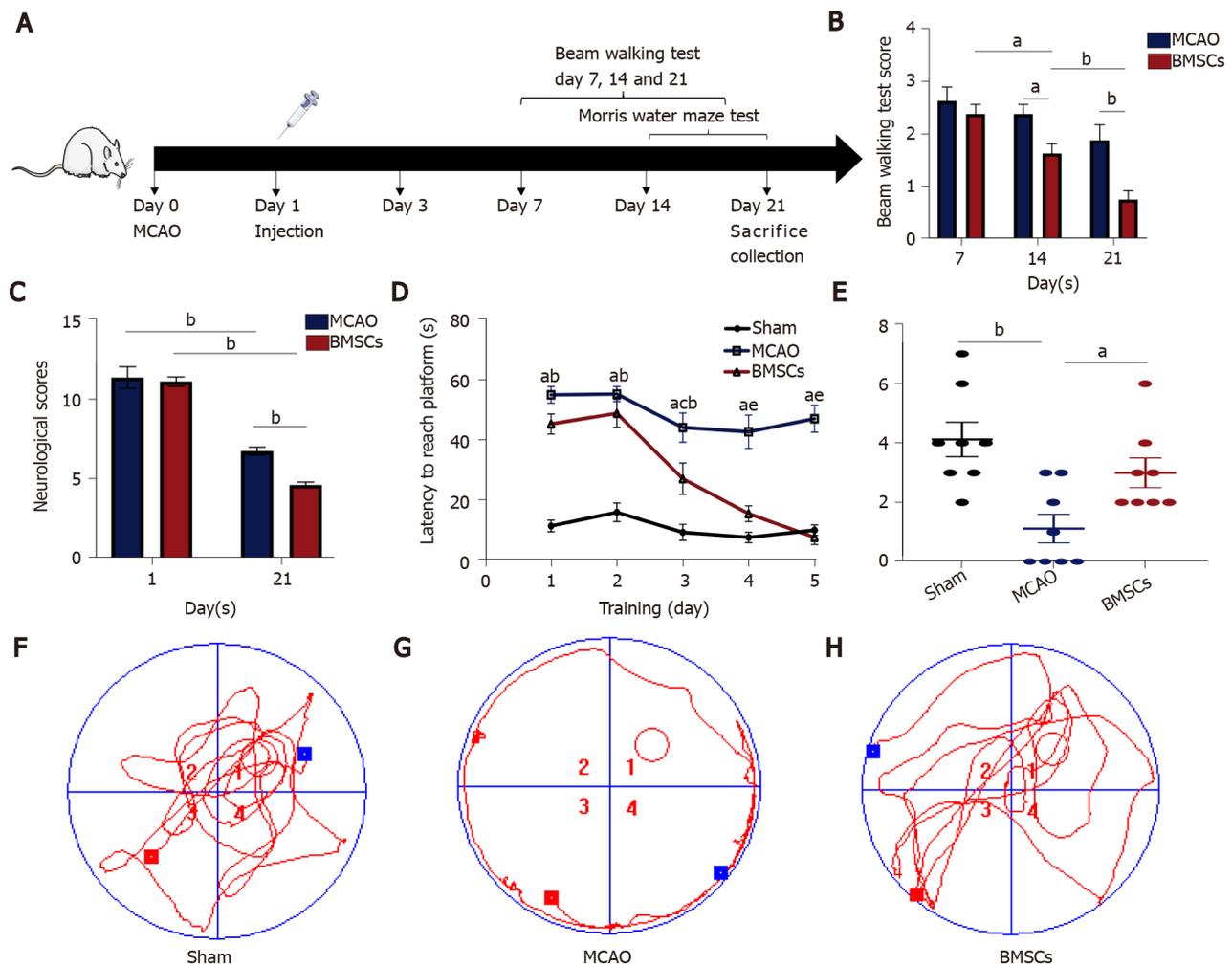
Rats were randomly divided into three groups ( $n = 10$  each): Sham operation control group (Sham), transient MCAO group, and MCAO with BMSC treatment group. The Sham and MCAO groups were injected with normal saline (PBS), and the BMSCs group was injected with  $1 \times 10^6$  BMSCs through the tail vein 24 h after reperfusion. Rats were killed after 21 d of reperfusion to collect feces and brain tissue for analysis (Figure 1A).

### MCAO

The intraluminal filament model was used to induce transient MCAO as described by Jackman *et al*[19]. Rats were anesthetized with 4% isoflurane and fixed in a supine position, and a longitudinal incision was made 0.3 cm to the right of the midline of their neck. Then, the muscles and tissues were separated to expose the right common carotid artery (CCA), external carotid artery (ECA), and internal carotid artery (ICA). Subsequently, a filament nylon suture was inserted into the right ECA and pushed until the middle cerebral artery (MCA) was obstructed. After 90 min of ischemia, the filament was removed carefully and reperfusion performed. During surgery, the rats were placed on a thermostat system to maintain body temperature.

### Neurobehavioral scores

The Longa 5-point scale was used to judge whether MCAO surgery is successful: 4, the animal died; 3, the animal could not walk in a straight line, and its body was tilted to one side; 2, the animal turned to one side during crawling; 1, the animal could not straighten its limbs and was stiff; 0, the animal was normal. If the score was 1-3, the model was considered successful, and the experiment can be carried out later; 0 and 4 were rejected. Animals with a score of 1 to 3 will be grouped for later experiments.



**Figure 1 Bone marrow mesenchymal stem cells improve neurological function after stroke.** A: Experimental design. Rats (3-5 wk) were randomly divided into three groups: Sham, middle cerebral artery occlusion (MCAO), and bone marrow mesenchymal stem cells (BMSCs). BMSCs or PBS were injected through the tail vein 1 d after MCAO. The modified Neurological Severity Score (mNSS), the beam walking test, and the Morris water maze test were evaluated before rats were killed after 21 d of reperfusion; B: mNSS was performed at days 1 and 21 after MCAO (<sup>a</sup>*P* < 0.05; <sup>b</sup>*P* < 0.01); C: Beam walking test were performed at days 7, 14, and 21 after MCAO (<sup>b</sup>*P* < 0.01); D-H: Morris water maze test. The time that rats needed to escape latency to find the hidden platform (D). <sup>a</sup>*P* < 0.01 when Sham vs MCAO; <sup>b</sup>*P* < 0.01 when Sham vs BMSCs; <sup>c</sup>*P* < 0.01 when Sham vs BMSCs; <sup>d</sup>*P* < 0.01 when MCAO vs BMSCs; <sup>e</sup>*P* < 0.01 when MCAO vs BMSCs. The number of rats crossing over the target platform on the sixth day (<sup>a</sup>*P* < 0.05; <sup>b</sup>*P* < 0.01) (E). The data are expressed as the mean ± SEM (*n* = 10). The tracks of each group on the sixth day (F-H). BMSCs: Bone marrow mesenchymal stem cells; MCAO: Middle cerebral artery occlusion.

The modified Neurological Severity Score (mNSS) was used to score the neurological function of the rats on days 1 and 21 after reperfusion, which included motor, sensory, reflex, and balance tests with a total score of 18[20]; the higher scores mean more severe injuries.

**Behavioral analysis**

Two blinded investigators observed all behavioral tests at regular times of the day. The apparatus was washed with 70% ethanol after each animal was tested to eliminate olfactory cues.

**Beam walking test:** For detecting motor coordination and balance, the beam walking test was evaluated at 7, 14, and 21 d after reperfusion. The rats were placed on a balance beam that was 1 m long, 2.5 cm wide, and 20 cm high from the ground. A soft cushion was placed under the balance beam to prevent the mouse from falling. Every mouse was scored according to the following rules: (1) If the rat crossed the balance beam smoothly without the hind limbs slipping; (2) If the rat gripped the edge of the balance beam, but the hind limbs did not dangle; (3) If the rat clutched the balance beam, and one limb dropped from the balance beam; (4) If the rat clutched the balance beam, and two limbs dropped from the balance beam or rotated on the balance beam (> 60 s); (5) If the rat tried to balance on the balance beam but fell (> 40 s); (6) If the rat tried to balance on the balance beam but failed (> 20 s); and (7) If the rats fell and did

not attempt to balance on the beam (< 20 s).

**Morris water maze test:** The Morris water maze test was performed 14 d after surgery for six consecutive days to test rats' spatial memory ability. The water maze was a circular black pool (Shanghai Xinsoft Information Technology Co., Ltd.), 150 cm in diameter, 50 cm high, and 25 cm deep, with the water temperature maintained at  $20 \pm 1$  °C. The pool was divided into four quadrants (1, 2, 3, and 4), and the circular platform was located in quadrant 1, 2 cm below the water surface. The rats were tested twice daily for 60 s for the first 5 d and were allowed to remain on the platform for 10 s after each test. On day 6, a probe trial was performed by removing the platform and allowing the rat to swim freely in the pool for the 60 s. The time and route taken by the rats to complete the task were recorded. Finally, the data were exported and analyzed using Morris water maze analysis software.

### **Histological analysis of rat brain**

The rats were fixed by perfusion in 4% paraformaldehyde (PFA). The brains were quickly removed and fixed in 4% PFA at 4 °C for 24 h. After dehydration, they were embedded in paraffin and serially sectioned into 4  $\mu$ m tissue sections for histological analysis. Nissl staining was performed to evaluate neuron damage. The histopathology of the hippocampus of brain tissues was observed with a microscope (BX43; Olympus).

### **Microbiome 16S rDNA sequencing and analysis**

The rat feces from each group were collected into 2 mL sterile freezing tubes on day 21 and stored at -80 °C until the bacterial DNA was extracted. Total bacterial DNA was extracted using DNA Extraction Kit (QIAGEN, Germany) following the manufacturer's instructions. To ensure the quality and quantity of DNA, extracted DNA was detected by agarose gel electrophoresis and stored at -20 °C until further processing. The diluted DNA was used as the template for PCR amplification of bacterial 16S rRNA genes with the barcoded primers (V3-V4 regions) and Takara Ex Taq (Takara). The PCR product was purified with AMPure XP beads (Beckman Coulter Genomics, United States) and quantified using a Qubit dsDNA assay kit (Life Technologies, United States). According to the standard protocols, equal amounts of purified amplicon were sequenced using the Illumina Miseq sequencer PE250 (Illumina, United States). The raw data were processed sequentially with the software Trimmomatic (version 0.35), Flash (version 1.2.11), QIIME (version 1.8.0), and UCHIME (version 2.4.2) to get the operational taxonomic units (OTUs). The valid tags were classified at a 97% similarity cutoff to analyze the gut microbiota diversity.

$\alpha$ -diversity is a measure of the abundance and diversity of microbial communities in a sample. In this paper, the Shannon index and Chao index were used to represent  $\alpha$ -diversity[21,22]. The Shannon index is an alpha diversity statistic for estimating the index of microbial diversity in a sample. A higher value indicates that the community is more diverse. The Chao index assesses the number of OTUs in a sample. The larger the Chao index, the higher the number of OTUs, indicating that the number of species in the sample is more numerous. The functional pathways of microbial communities for each sample were inferred using Phylogenetic Investigation of Communities by Reconstruction of Unobserved States (PICRUST) software[23]. The PICRUST software predicts the metabolic function of microorganisms by comparing the resulting 16S sequencing data with a genomic reference database of microorganisms with known metabolic functions.

### **Statistical analysis**

The results are expressed as the mean  $\pm$  SEM. The data were analyzed using one-way analysis of variance (ANOVA) and *t*-test. The difference was considered significant at  $P < 0.05$ .

## **RESULTS**

### **BMSCs improve neurological function after ischemic stroke**

The mNSS, beam walking test, and Morris water maze test were used to estimate the neurological function after ischemic stroke. The neurological deficit scores of each group of rats were evaluated at 1 and 21 d after ischemia-reperfusion (Figure 1B). Compared with the MCAO group, the BMSCs group had significantly improved

neurological function. The mNSS scores of both the MCAO and BMSCs groups were substantially lower at 21 d than on the first day ( $P < 0.01$ ). However, the BMSCs group had a more significant decrease in mNSS scores at day 21 than the MCAO group ( $P < 0.01$ ). Beam walking test showed that rats subjected to BMSCs transplantation presented a larger motor functional improvement (14 d,  $P < 0.05$ ; 21 d,  $P < 0.01$ ; **Figure 1C**)

To assess the spatial learning and memory capacity of BMSC-treated rats after stroke, the Morris water maze test was used to detect the escape latency of a random search for the hidden platform during the first 5 d. Compared to the MCAO group, the BMSCs group showed a significantly shorter duration of escaping latency ( $P < 0.05$ ; **Figure 1D**). After removing the hidden platform at 6 d, rats of the BMSCs group were easier to find the previous location of the platform site compared to those of the MCAO group, which passed over the platform site more times ( $P < 0.05$ ; **Figure 1E**). The typical swimming tracks of each group (**Figure 1F-H**) also indicated that rats treated with BMSCs had significantly improved spatial memory.

### **BMSCs alleviate neuronal loss in the hippocampus after ischemic stroke**

Nissl staining demonstrated no significant changes in neurons in the hippocampal CA1 area of the brain in the Sham group on day 21. In the MCAO group, the boundaries of the hippocampal CA1 area were irregular, the number of Nissl bodies was reduced, and a large number of neurons underwent necrosis. Compared with the MCAO group, the rat hippocampal neurons in the group treated with BMSCs were arranged in an orderly manner, and necrotic cells were significantly reduced (**Figure 2**). These results suggest that stroke causes severe neuronal damage in rats and that BMSC treatment can effectively protect neurons and prevent neuronal loss.

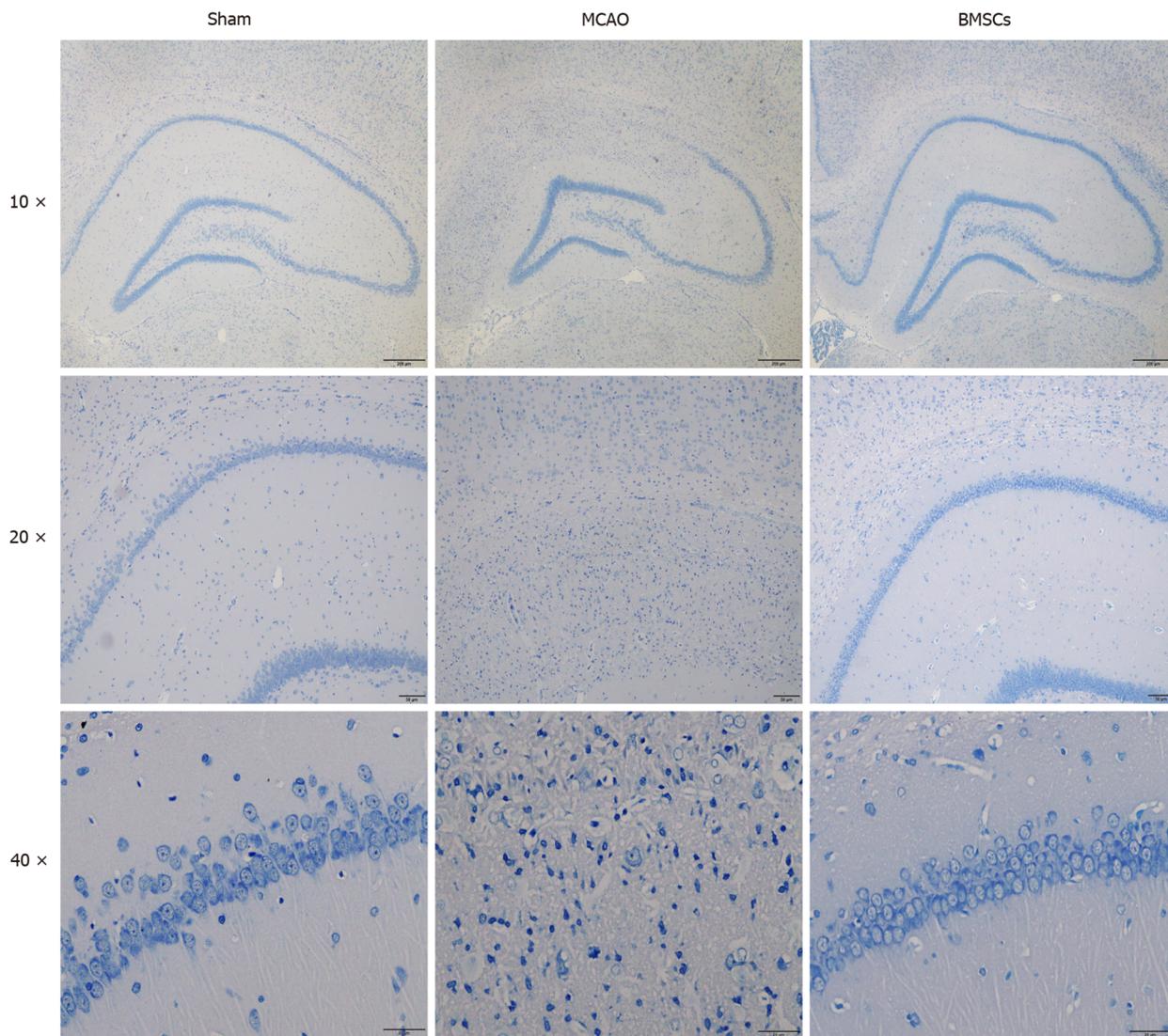
### **Effect of BMSCs on microbial $\alpha$ -diversity and structure after ischemic stroke**

To identify whether treatment with BMSCs influences the gut microbiota after ischemic stroke, we analyzed differences in species complexity and bacterial communities between populations based on OTUS and species annotation results. We obtained a total of 1494295 quality filtered 16s rRNA gene sequences from three groups of 30 samples, with an average of  $49810 \pm 1281$  reads per sample. We compared microbial  $\alpha$ -diversity between the Sham, MCAO, and BMSCs groups, and both Shannon and Chao index results showed no statistical difference between the three groups (**Figure 3A and B**).

We calculated inter-sample distances between the three groups to analyze the differences in community species composition among individual samples within each group. We demonstrate the nonmetric multi-dimensional scaling (nMDS) plot, and the principle co-ordinates analysis (PCoA) plots in **Figure 3C and D**. Different groups are presented in different colors in the figure, and samples from the same group are clustered together. The nMDS analysis and PCoA showed that MCAO and BMSCs could alter the microbiota composition significantly compared to the Sham group. However, there was no significant difference in microbiota structure between the two groups of MCAO and BMSCs. To further investigate the variability of microbial communities between the two groups, the ANOSIM test was used to test both Bray-Curtis and Unweighted Unifrac algorithms (Bray-Curtis,  $r = 0.0769$ ,  $P = 0.042$ ; Unweighted Unifrac,  $r = 0.0679$ ,  $P = 0.0415$ , respectively). The results showed significant differences in the microbial communities between the two groups.

### **BMSCs modulate gut microbiota after ischemic stroke**

We next sought to explore the effect of treatment with BMSCs on the composition of the microbial structure. **Figure 4A** shows the abundance of microorganisms in the three groups, in which Bacteroidetes, Firmicutes, Proteobacteria, and Epsilonbacteraeota were the most significant contributors at the microbial phylum level. Compared with the Sham group, MCAO and BMSC increased the relative abundance of Proteobacteria, suggesting significant differences in the gut microbiota structure after stroke. Furthermore, we analyzed the differences in the relative abundance of microorganisms between the three groups at the level of genus (**Figure 4B**). The data showed that the relative abundance of *Ruminococcaceae\_UCG-005*, *Mycoplasma*, *Ruminiclostridium\_5*, *Oceanimonas*, and *Marvinbryantia* was significantly decreased, and the relative abundance of *Escherichia-Shigella*, *Alloprevotella*, *Butyricimonas*, *ASF356*, and *Enterococcus* was increased in the MCAO group compared with the Sham group. BMSC treatment increased the relative abundance of *Ruminiclostridium\_5* and decreased *Butyricimonas* and *ASF356* at the species level. The dominant bacteria of MCAO and BMSCs are shown separately at the species level in **Figure 4C and D**. We concluded that species enriched in the BH group included *Clostridium* spp and *Lachno-*

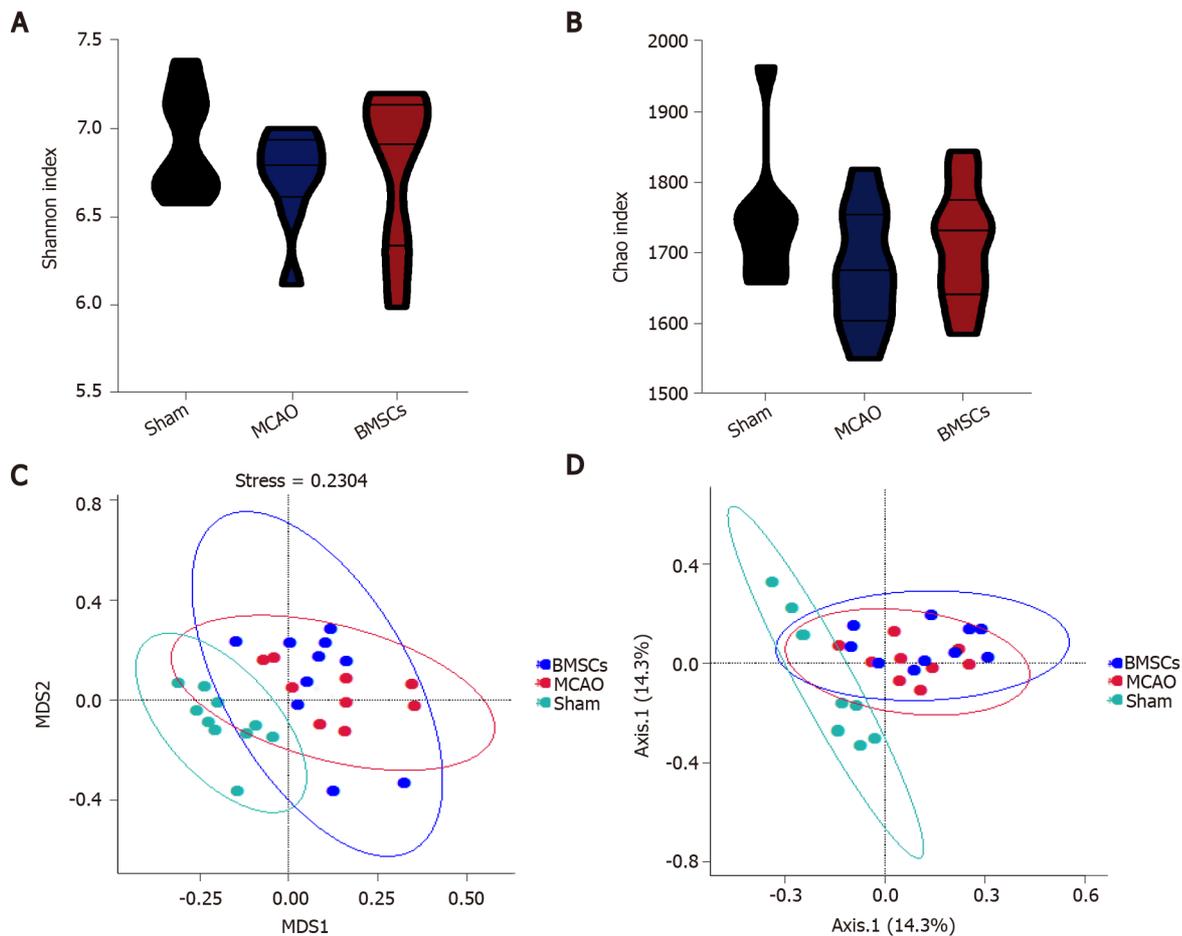


**Figure 2** Histopathological changes in brain tissue of rats. Nissl staining in the hippocampal CA1 region for the Sham, middle cerebral artery occlusion, and bone marrow mesenchymal stem cells group exhibited brain injury after 21 d post-stroke ( $n = 3$ ). Pathological observation of the hippocampus (magnification,  $\times 40$ ). CA1 region of the hippocampus (magnification,  $\times 100$ ). The morphologies of neurons in the hippocampal CA1 region (magnification,  $\times 400$ ). BMSCs: Bone marrow mesenchymal stem cells; MCAO: Middle cerebral artery occlusion.

*spiraceae* spp, which are the potential species to produce short-chain fatty acid (SCFA). A comparison of potential SCFA producing bacteria in the feces revealed that depletion occurred in the MCAO group (Figure 4E). Additionally, it was observed that the relative abundance of *Lactobacillus* was significantly increased at the genus level after BMSC treatment (Figure 4F).

#### **Predictive analysis of gut microbiota function**

PICRUSt functional prediction analysis was based on 16S sequencing data annotated in the Greengenes database. Using PICRUSt software can predict the composition of known microbial gene functions and thus statistically different functions between groups. In this study, the Kyoto Encyclopedia of Genes and Genomes (KEGG) was used to assess microbial function, and 25 differentially KEGG functional pathways were identified between MCAO and BMSCs (Figure 5). The gut microbiota of BMSCs influenced the pathways of metabolism, including "Carbohydrate Metabolism", "Biosynthesis of Other Secondary Metabolites", "Glycan Biosynthesis and Metabolism", "Lipid Metabolism", "Metabolism of Cofactors and Vitamins", "Metabolism of Other Amino Acids", and "Xenobiotics Biodegradation and Metabolism". We also found that BMSCs-enriched function pathways were associated with "Membrane Transport", "Signaling Molecules and Interaction", "Transport and Catabolism", and "Transcription".



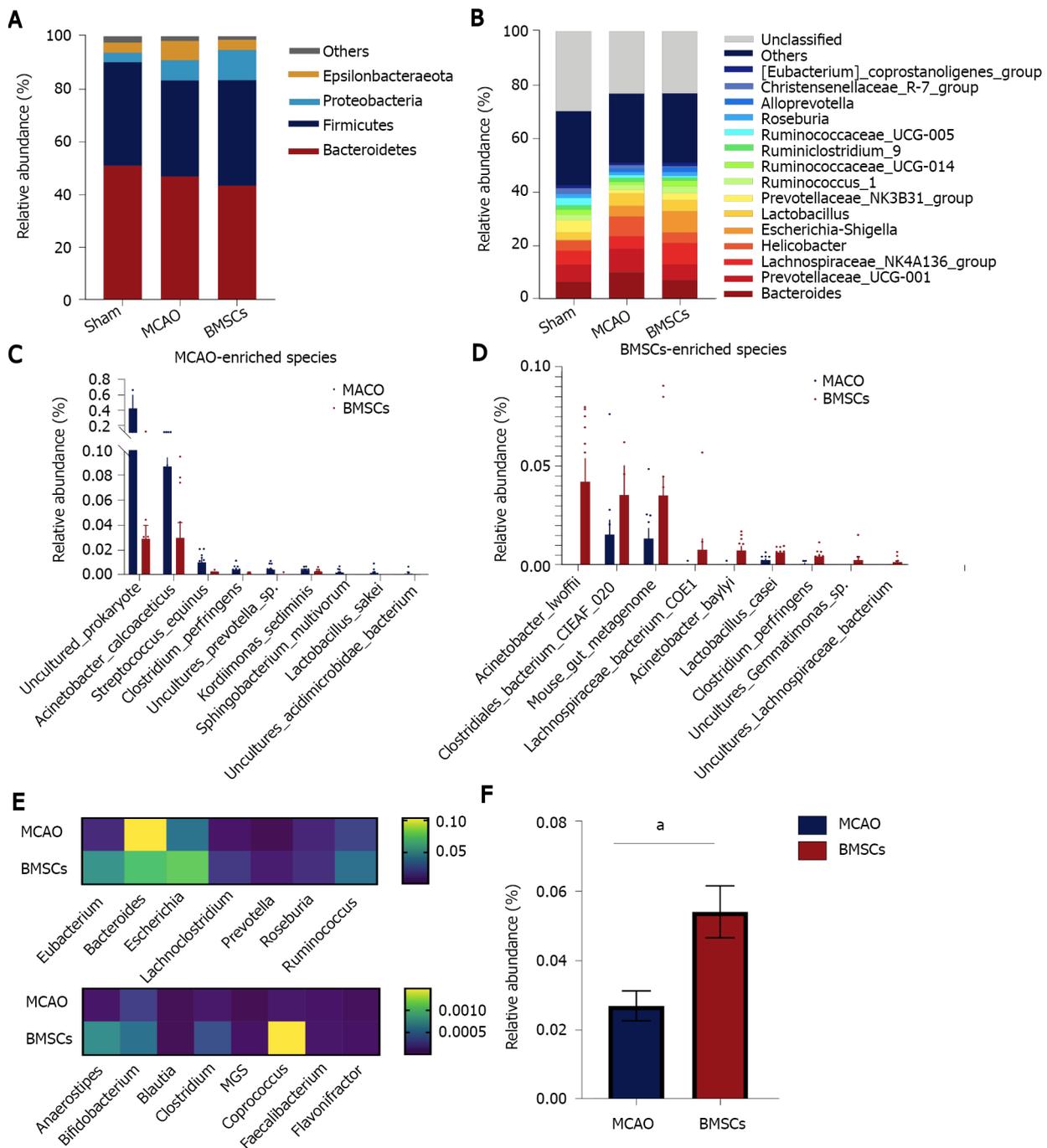
**Figure 3** Abundance and structures analysis of intestinal microecology. A and B: Shannon and Chao index to present  $\alpha$ -diversity of gut microbiota; C and D: mNDS and PCoA plot to illustrate the dissimilarities among microbiota structures. BMSCs: Bone marrow mesenchymal stem cells; MCAO: Middle cerebral artery occlusion.

## DISCUSSION

For the first time, this study showed changes in gut microbiota after ischemic stroke treatment using BMSCs. BMSCs disrupted the composition and structure of gut microbiota, thereby affecting metabolic pathways in ischemic stroke.

Evidence from basic and clinical studies show that BMSCs can effectively treat patients with ischemic stroke[24]. Transplantation of BMSCs significantly enhances neurological functions after stroke[25], consistent with our results. We established that treatment with BMSCs significantly reduced mNSS scores and enhanced balance, coordination abilities, and learning memory in rats. Notably, cerebral ischemia caused neuronal damage in the hippocampus, striatum, thalamus, and cerebellar cortices, with the CA1 region of the hippocampus being one of the most sensitive brain regions. Nissl staining revealed serious neuronal damage in rats after ischemic stroke, which explains memory impairment in the Morris water maze test. In contrast, BMSCs effectively protected the nerve cells.

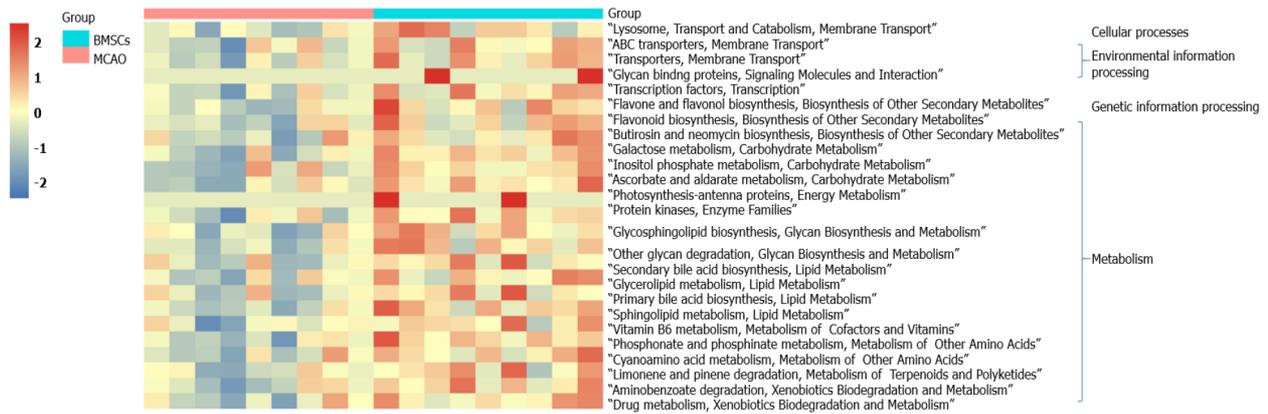
Studies have confirmed complex interactions between gut microbiota and stroke. Xia *et al*[26] reported that *Parabacteroides*, *Oscillospira*, and *Enterobacteriaceae* among others were enriched in stroke patients, whereas *Prevotella*, *Roseburia*, and *Faecalibacterium* were enriched in healthy individuals[26]. In stroke patients, dysbiosis is closely associated with metabolism and inflammation. Besides, a specific genus of gut microbiota and associated metabolites are used as potential indicators for stroke prediction and prognosis[13,27]. In stroke animal models, similar alterations in gut microbiota have been detected. Singh *et al*[9] found that the most abundant phyla of *Firmicutes*, *Bacteroidetes*, and *Actinobacteria* overgrew in MCAO mice[9]. Chen *et al*[28] reported that after stroke, rats exhibited an increase in the abundance of opportunistic pathogens, including *Alistipes*, *Bacteroides*, *Klebsiella*, *Shuttleworthia*, *Haemophilus*, *Fusobacterium*, *Faecalibacterium*, *Proteus*, and *Papillibacter*[28]. After transplantation of BMSCs, we analyzed the changes in gut microbiota to investigate the role of gut



**Figure 4 Bone marrow mesenchymal stem cells modulate the composition of gut microbiota.** A: Taxonomic composition at the phylum level; B: Taxonomic composition at the genus level; C and D: Significantly different abundances at the species level between middle cerebral artery occlusion (MCAO) and bone marrow mesenchymal stem cells groups; E: Comparison of the abundance of potential short-chain fatty acid-producing species in the MCAO and BH groups; F: Relative abundances of *Lactobacillus* at the genus level between the MCAO and BH groups ( $^*P < 0.05$ ). The data are expressed as the mean  $\pm$  SEM ( $n = 10$ ). BMSCs: Bone marrow mesenchymal stem cells; MCAO: Middle cerebral artery occlusion.

microbiota in post-stroke rats. We found that BMSCs did not alter the  $\alpha$ -diversity and structure of gut microbiota after stroke. Further assessments of the composition of microbiota structure suggested that BMSCs significantly increased the abundance of potential SCFA-producing bacteria.

*Lachnospiraceae* and *Clostridium* are the main groups of SCFA-producing bacteria [29]. For mammals, SCFA is a critical gut microbial metabolite. It can be used as a substrate for the metabolism of cholesterol, glucose, and lipids, which provide nearly 10% of daily caloric requirements[30]. Besides, it achieves its anti-inflammatory effects by activating G protein-coupled receptors (GPCR) to regulate T cells[31]. Additionally, SCFA protects and repairs the intestinal mucosal barrier by secreting mucus and stimulating tight junction protein expression[32].



**Figure 5 Alterations of microbial function.** Heatmap illustrates the difference of Kyoto Encyclopedia of Genes and Genomes (KEGG) pathways (KEGG level 3) between middle cerebral artery occlusion and bone marrow mesenchymal stem cells groups. Welch's t-test was used for statistical analysis, and the pathways is displayed when  $P < 0.05$ . BMSCs: Bone marrow mesenchymal stem cells; MCAO: Middle cerebral artery occlusion.

The abundance of *Lactobacillus* has been shown to be significantly increased in cerebral infarction patients[33]. Interestingly, we found a significantly high abundance of *Lactobacillus* in the fecal matter of the BMSCs group. Bourriaud *et al*[34] realized that butyrate-producing bacteria ferment lactic acid to produce butyrate, which reduces inflammatory responses, thereby protecting the injured brain[34]. Given that BMSCs increase the abundance of potential SCFA-producing bacteria, an increase in *Lactobacillus* leads to the production of more lactic acid to be fermented to butyrate, thereby improving neuroinflammation during stroke.

## CONCLUSION

This is the first study to elucidate on alterations in gut microbiota after BMSC treatment in an ischemic stroke condition. We found that BMSCs potentially improve neurological damage after stroke by regulating gut microbiota. This provides a basis for future research into the role of BMSCs from the perspective of the "gut-brain axis".

## ARTICLE HIGHLIGHTS

### Research background

Ischemic stroke is a highly lethal and disabling disease that has a severe impact on the quality of life of patients. Gut microbiota is closely related to the treatment and prognosis of stroke. The improvement of neurological function by bone marrow mesenchymal stem cells (BMSCs) may be related to the regulation of gut microbiota.

### Research motivation

Many studies have shown that gut microbiota plays an important role in immunity after stroke through the gut-brain axis.

### Research objectives

To observe the regulation of gut microbiota after BMSC treatment.

### Research methods

Rats were divided into three groups [Sham, middle cerebral artery occlusion (MCAO), and BMSCs]. Recovery of neurological function in rats after BMSC transplantation was observed by the modified Neurological Severity Scores (mNSS), beam walking test, and Morris water maze test. Pathological observation of hippocampal neuronal cells was conducted by Nissl staining. 16S rDNA sequencing was used to analyze the composition of gut microbiota.

### Research results

Transplantation of BMSCs significantly reduced mNSS scores ( $P < 0.01$ ), and improved balance and coordination ( $P < 0.01$ ), learning, and memory in rats. The structure of the C1 region of the hippocampus was clear and necrotic cells were significantly reduced after the intervention of BMSCs. Compared with the MCAO group, BMSCs effectively increased the relative abundance of short-chain fatty acid-producing bacteria and *Lactobacillus* in feces.

### Research conclusions

Transplantation of BMSCs can regulate gut microbiota, which provides a potential therapeutic mechanism for stroke treatment.

### Research perspectives

We demonstrated the modulatory effect of BMSCs on the gut microbiota after stroke, which provided an experimental basis for elucidating the gut-brain axis.

---

## REFERENCES

---

- 1 **GBD 2016 Neurology Collaborators.** Global, regional, and national burden of neurological disorders, 1990-2016: a systematic analysis for the Global Burden of Disease Study 2016. *Lancet Neurol* 2019; **18**: 459-480 [PMID: 30879893 DOI: 10.1016/S1474-4422(18)30499-X]
- 2 **Virani SS,** Alonso A, Benjamin EJ, Bittencourt MS, Callaway CW, Carson AP, Chamberlain AM, Chang AR, Cheng S, Delling FN, Djousse L, Elkind MSV, Ferguson JF, Fornage M, Khan SS, Kissela BM, Knutson KL, Kwan TW, Lackland DT, Lewis TT, Lichtman JH, Longenecker CT, Loop MS, Lutsey PL, Martin SS, Matsushita K, Moran AE, Mussolino ME, Perak AM, Rosamond WD, Roth GA, Sampson UKA, Satou GM, Schroeder EB, Shah SH, Shay CM, Spartano NL, Stokes A, Tirschwell DL, VanWagner LB, Tsao CW; American Heart Association Council on Epidemiology and Prevention Statistics Committee and Stroke Statistics Subcommittee. Heart Disease and Stroke Statistics-2020 Update: A Report From the American Heart Association. *Circulation* 2020; **141**: e139-e596 [PMID: 31992061 DOI: 10.1161/CIR.0000000000000757]
- 3 **Campbell BCV,** De Silva DA, Macleod MR, Coutts SB, Schwamm LH, Davis SM, Donnan GA. Ischaemic stroke. *Nat Rev Dis Primers* 2019; **5**: 70 [PMID: 31601801 DOI: 10.1038/s41572-019-0118-8]
- 4 **Emberson J,** Lees KR, Lyden P, Blackwell L, Albers G, Bluhmki E, Brott T, Cohen G, Davis S, Donnan G, Grotta J, Howard G, Kaste M, Koga M, von Kummer R, Lansberg M, Lindley RI, Murray G, Olivot JM, Parsons M, Tilley B, Toni D, Toyoda K, Wahlgren N, Wardlaw J, Whiteley W, del Zoppo GJ, Baigent C, Sandercock P, Hacke W; Stroke Thrombolysis Trialists' Collaborative Group. Effect of treatment delay, age, and stroke severity on the effects of intravenous thrombolysis with alteplase for acute ischaemic stroke: a meta-analysis of individual patient data from randomised trials. *Lancet* 2014; **384**: 1929-1935 [PMID: 25106063 DOI: 10.1016/S0140-6736(14)60584-5]
- 5 **Saver JL,** Goyal M, van der Lugt A, Menon BK, Majoie CB, Dippel DW, Campbell BC, Nogueira RG, Demchuk AM, Tomasello A, Cardona P, Devlin TG, Frei DF, du Mesnil de Rochemont R, Berkhemer OA, Jovin TG, Siddiqui AH, van Zwam WH, Davis SM, Castaño C, Sapkota BL, Franssen PS, Molina C, van Oostenbrugge RJ, Chamorro Á, Lingsma H, Silver FL, Donnan GA, Shuaib A, Brown S, Stouch B, Mitchell PJ, Davalos A, Roos YB, Hill MD; HERMES Collaborators. Time to Treatment With Endovascular Thrombectomy and Outcomes From Ischemic Stroke: A Meta-analysis. *JAMA* 2016; **316**: 1279-1288 [PMID: 27673305 DOI: 10.1001/jama.2016.13647]
- 6 **Boltze J,** Modo MM, Mays RW, Taguchi A, Jolkkonen J, Savitz SI; STEPS 4 Consortium. Stem Cells as an Emerging Paradigm in Stroke 4: Advancing and Accelerating Preclinical Research. *Stroke* 2019; **50**: 3299-3306 [PMID: 31658004 DOI: 10.1161/STROKEAHA.119.025436]
- 7 **Dabrowska S,** Andrzejewska A, Lukomska B, Janowski M. Neuroinflammation as a target for treatment of stroke using mesenchymal stem cells and extracellular vesicles. *J Neuroinflammation* 2019; **16**: 178 [PMID: 31514749 DOI: 10.1186/s12974-019-1571-8]
- 8 **Lee JY,** Tuazon JP, Ehrhart J, Sanberg PR, Borlongan CV. Gutting the brain of inflammation: A key role of gut microbiome in human umbilical cord blood plasma therapy in Parkinson's disease model. *J Cell Mol Med* 2019; **23**: 5466-5474 [PMID: 31148353 DOI: 10.1111/jcmm.14429]
- 9 **Singh V,** Roth S, Llovera G, Sadler R, Garzetti D, Stecher B, Dichgans M, Liesz A. Microbiota Dysbiosis Controls the Neuroinflammatory Response after Stroke. *J Neurosci* 2016; **36**: 7428-7440 [PMID: 27413153 DOI: 10.1523/JNEUROSCI.1114-16.2016]
- 10 **Houlden A,** Goldrick M, Brough D, Vizi ES, Lénárt N, Martinecz B, Roberts IS, Denes A. Brain injury induces specific changes in the caecal microbiota of mice *via* altered autonomic activity and mucoprotein production. *Brain Behav Immun* 2016; **57**: 10-20 [PMID: 27060191 DOI: 10.1016/j.bbi.2016.04.003]
- 11 **Spychala MS,** Venna VR, Jandzinski M, Doran SJ, Durgan DJ, Ganesh BP, Ajami NJ, Putluri N, Graf J, Bryan RM, McCullough LD. Age-related changes in the gut microbiota influence systemic inflammation and stroke outcome. *Ann Neurol* 2018; **84**: 23-36 [PMID: 29733457 DOI: 10.1002/ana.25111]

- 10.1002/ana.25250]
- 12 **Li XJ**, You XY, Wang CY, Li XL, Sheng YY, Zhuang PW, Zhang YJ. Bidirectional Brain-gut-microbiota Axis in increased intestinal permeability induced by central nervous system injury. *CNS Neurosci Ther* 2020; **26**: 783-790 [PMID: 32472633 DOI: 10.1111/cns.13401]
  - 13 **Yamashiro K**, Tanaka R, Urabe T, Ueno Y, Yamashiro Y, Nomoto K, Takahashi T, Tsuji H, Asahara T, Hattori N. Gut dysbiosis is associated with metabolism and systemic inflammation in patients with ischemic stroke. *PLoS One* 2017; **12**: e0171521 [PMID: 28166278 DOI: 10.1371/journal.pone.0171521]
  - 14 **Yin J**, Liao SX, He Y, Wang S, Xia GH, Liu FT, Zhu JJ, You C, Chen Q, Zhou L, Pan SY, Zhou HW. Dysbiosis of Gut Microbiota With Reduced Trimethylamine-N-Oxide Level in Patients With Large-Artery Atherosclerotic Stroke or Transient Ischemic Attack. *J Am Heart Assoc* 2015; **4** [PMID: 26597155 DOI: 10.1161/JAHA.115.002699]
  - 15 **Stanley D**, Mason LJ, Mackin KE, Srikhanta YN, Lyras D, Prakash MD, Nurgali K, Venegas A, Hill MD, Moore RJ, Wong CH. Translocation and dissemination of commensal bacteria in post-stroke infection. *Nat Med* 2016; **22**: 1277-1284 [PMID: 27694934 DOI: 10.1038/nm.4194]
  - 16 **Benakis C**, Brea D, Caballero S, Faraco G, Moore J, Murphy M, Sita G, Racchumi G, Ling L, Pamer EG, Iadecola C, Anrather J. Commensal microbiota affects ischemic stroke outcome by regulating intestinal  $\gamma\delta$  T cells. *Nat Med* 2016; **22**: 516-523 [PMID: 27019327 DOI: 10.1038/nm.4068]
  - 17 **Chen R**, Xu Y, Wu P, Zhou H, Lasanajak Y, Fang Y, Tang L, Ye L, Li X, Cai Z, Zhao J. Transplantation of fecal microbiota rich in short chain fatty acids and butyric acid treat cerebral ischemic stroke by regulating gut microbiota. *Pharmacol Res* 2019; **148**: 104403 [PMID: 31425750 DOI: 10.1016/j.phrs.2019.104403]
  - 18 **Sadler R**, Cramer JV, Heindl S, Kostidis S, Betz D, Zuurbier KR, Northoff BH, Heijink M, Goldberg MP, Plautz EJ, Roth S, Malik R, Dichgans M, Holdt LM, Benakis C, Giera M, Stowe AM, Liesz A. Short-Chain Fatty Acids Improve Poststroke Recovery via Immunological Mechanisms. *J Neurosci* 2020; **40**: 1162-1173 [PMID: 31889008 DOI: 10.1523/JNEUROSCI.1359-19.2019]
  - 19 **Jackman K**, Kunz A, Iadecola C. Modeling focal cerebral ischemia in vivo. *Methods Mol Biol* 2011; **793**: 195-209 [PMID: 21913102 DOI: 10.1007/978-1-61779-328-8\_13]
  - 20 **Chen J**, Sanberg PR, Li Y, Wang L, Lu M, Willing AE, Sanchez-Ramos J, Chopp M. Intravenous administration of human umbilical cord blood reduces behavioral deficits after stroke in rats. *Stroke* 2001; **32**: 2682-2688 [PMID: 11692034 DOI: 10.1161/hs1101.098367]
  - 21 **Shannon CE**. The mathematical theory of communication. 1963. *MD Comput* 1997; **14**: 306-317
  - 22 **Chao A**, Chazdon RL, Colwell RK, Shen TJ. A new statistical approach for assessing similarity of species composition with incidence and abundance data. *Ecology Letters* 2005; **8**: 148-159 [DOI: 10.1111/j.1461-0248.2004.00707.x]
  - 23 **Langille MG**, Zaneveld J, Caporaso JG, McDonald D, Knights D, Reyes JA, Clemente JC, Burkepile DE, Vega Thurber RL, Knight R, Beiko RG, Huttenhower C. Predictive functional profiling of microbial communities using 16s rRNA marker gene sequences. *Nat Biotechnol* 2013; **31**: 814-821 [PMID: 23975157 DOI: 10.1038/nbt.2676]
  - 24 **Bang OY**, Kim EH, Cha JM, Moon GJ. Adult Stem Cell Therapy for Stroke: Challenges and Progress. *J Stroke* 2016; **18**: 256-266 [PMID: 27733032 DOI: 10.5853/jos.2016.01263]
  - 25 **Cai Y**, Liu W, Lian L, Xu Y, Bai X, Xu S, Zhang J. Stroke treatment: Is exosome therapy superior to stem cell therapy? *Biochimie* 2020; **179**: 190-204 [PMID: 33010339 DOI: 10.1016/j.biochi.2020.09.025]
  - 26 **Xia GH**, You C, Gao XX, Zeng XL, Zhu JJ, Xu KY, Tan CH, Xu RT, Wu QH, Zhou HW, He Y, Yin J. Stroke Dysbiosis Index (SDI) in Gut Microbiome Are Associated With Brain Injury and Prognosis of Stroke. *Front Neurol* 2019; **10**: 397 [PMID: 31068891 DOI: 10.3389/fneur.2019.00397]
  - 27 **Li N**, Wang X, Sun C, Wu X, Lu M, Si Y, Ye X, Wang T, Yu X, Zhao X, Wei N. Change of intestinal microbiota in cerebral ischemic stroke patients. *BMC Microbiol* 2019; **19**: 191 [PMID: 31426765 DOI: 10.1186/s12866-019-1552-1]
  - 28 **Chen H**, Nwe PK, Yang Y, Rosen CE, Bielecka AA, Kuchroo M, Cline GW, Kruse AC, Ring AM, Crawford JM, Palm NW. A Forward Chemical Genetic Screen Reveals Gut Microbiota Metabolites That Modulate Host Physiology. *Cell* 2019; **177**: 1217-1231.e18 [PMID: 31006530 DOI: 10.1016/j.cell.2019.03.036]
  - 29 **Chen R**, Wu P, Cai Z, Fang Y, Zhou H, Lasanajak Y, Tang L, Ye L, Hou C, Zhao J. Puerariae Lobatae Radix with chuanxiong Rhizoma for treatment of cerebral ischemic stroke by remodeling gut microbiota to regulate the brain-gut barriers. *J Nutr Biochem* 2019; **65**: 101-114 [PMID: 30710886 DOI: 10.1016/j.jnutbio.2018.12.004]
  - 30 **LeBlanc JG**, Chain F, Martin R, Bermúdez-Humarán LG, Courau S, Langella P. Beneficial effects on host energy metabolism of short-chain fatty acids and vitamins produced by commensal and probiotic bacteria. *Microb Cell Fact* 2017; **16**: 79 [PMID: 28482838 DOI: 10.1186/s12934-017-0691-z]
  - 31 **Al Nabhani Z**, Dulauroy S, Marques R, Cousu C, Al Bounny S, Déjardin F, Sparwasser T, Bérard M, Cerf-Bensussan N, Eberl G. A Weaning Reaction to Microbiota Is Required for Resistance to Immunopathologies in the Adult. *Immunity* 2019; **50**: 1276-1288.e5 [PMID: 30902637 DOI: 10.1016/j.immuni.2019.02.014]
  - 32 **Zhao L**, Yang L, Guo Y, Xiao J, Zhang J, Xu S. New Insights into Stroke Prevention and Treatment: Gut Microbiome. *Cell Mol Neurobiol* 2021 [PMID: 33635417 DOI: 10.1007/s10571-021-01047-w]
  - 33 **Li H**, Zhang X, Pan D, Liu Y, Yan X, Tang Y, Tao M, Gong L, Zhang T, Woods CR, Du Y, Gao R, Qin H. Dysbiosis characteristics of gut microbiota in cerebral infarction patients. *Transl Neurosci*

- 2020; **11**: 124-133 [PMID: [33312718](#) DOI: [10.1515/tnsci-2020-0117](#)]
- 34 **Bourriaud C**, Robins RJ, Martin L, Kozlowski F, Tenailleau E, Cherbut C, Michel C. Lactate is mainly fermented to butyrate by human intestinal microfloras but inter-individual variation is evident. *J Appl Microbiol* 2005; **99**: 201-212 [PMID: [15960680](#) DOI: [10.1111/j.1365-2672.2005.02605.x](#)]



Published by **Baishideng Publishing Group Inc**  
7041 Koll Center Parkway, Suite 160, Pleasanton, CA 94566, USA  
**Telephone:** +1-925-3991568  
**E-mail:** [bpgoffice@wjgnet.com](mailto:bpgoffice@wjgnet.com)  
**Help Desk:** <https://www.f6publishing.com/helpdesk>  
<https://www.wjgnet.com>

