**Name of Journal:** *World Journal of Hepatology*

**Manuscript NO:** 68211

**Manuscript Type:** ORIGINAL ARTICLE

***Retrospective Study***

**Correlation of hepatitis B surface antigen expression with clinicopathological and biochemical parameters in liver biopsies: A comprehensive study**

Alpsoy A *et al*. HBsAg expression in HBV hepatitis

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**Received:** May 15, 2021

**Revised:** July 10, 2021

**Accepted:** December 31, 2021

**Published online:** January 28, 2022

**Abstract**

BACKGROUND

Chronic viral B hepatitis (CHB) is a potentially life-threatening liver disease that may progress to liver failure and cirrhosis. Currently, although combinations of different laboratory methods are used in the follow-up and treatment of CHB, the failure of these procedures in some cases has led to the necessity of developing new approaches. In CHB, the intrahepatic expression pattern of viral antigens, including hepatitis B surface antigen (HBsAg), is related to different phases of inflammation. However, many studies have focused on the intracytoplasmic properties of HBsAg staining, and HBsAg positivity in liver tissue has not been evaluated by objective quantitative methods.

AIM

To investigate the relationship of image analysis-based quantitative HBsAg expression and its staining patterns with clinicopathological factors and treatment in CHB.

METHODS

A total of 140 liver biopsies from treatment-naïve cases with CHB infection were included in this study. Following diagnosis, all patients were treated with entecavir (0.5 mg) and followed up at three-month intervals. The percentage of immunohistochemical HBsAg (p-HBsAg) expression in the liver was determined in whole tissue sections of biopsies from each case by image analysis. The immunohistochemical staining pattern was also evaluated separately according to 3 different previously defined classifications.

RESULTS

A positive correlation between p-HBsAg and serum levels of hepatitis B virus (HBV) DNA and HBsAg was observed (*P* < 0.001). The p-HBsAg value was significantly higher in younger patients than in older patients. When the groups were categorized according to the hepatitis B e antigen (HBeAg) status in HBeAg-positive cases, p-HBsAg was correlated with HBV DNA, hepatitis activity index (HAI) and fibrosis scores (*P* < 0.001). In this group, p-HBsAg and HBsAg expression patterns were also correlated with the viral response (VR) and the serological response (SR) (*P* < 0.001). Multivariate analysis revealed that p-HBsAg was an independent predictor of either VR or SR (*P* < 0.001). In HBeAg-negative patients, although HBsAg expression patterns were correlated with both HAI and fibrosis, no relationship was observed among p-HBsAg, clinicopathological factors and VR.

CONCLUSION

In pretreatment liver biopsies, the immunohistochemical determination of HBsAg expression by quantitative methods, beyond its distribution within the cell,may be a good predictor of the treatment response, especially in HBeAg-positive cases.

**Key Words:** Hepatitis B; Hepatitis B surface antigens; Hepatitis B e antigens; Fibrosis; Immunohistochemistry; Image analysis

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**Citation:** Alpsoy A, Adanir H, Bayramoglu Z, Elpek GO. Correlation of hepatitis B surface antigen expression with clinicopathological and biochemical parameters in liver biopsies: A comprehensive study. *World J Hepatol* 2022; 14(1): 260-273

**URL:** https://www.wjgnet.com/1948-5182/full/v14/i1/260.htm

**DOI:** https://dx.doi.org/10.4254/wjh.v14.i1.260

**Core Tip:** This report describes a study that investigated image analysis-based quantitative hepatitis B surface antigen (HBsAg) expression and its different staining patterns in liver biopsies from patients with chronic viral B hepatitis (CHB) and correlated them with clinicopathological factors and treatment. Our findings confirmed the association of cytoplasmic HBsAg staining patterns with disease activity. Besides, the determination of immunohistochemical HBsAg expression by image analysis may be an important predictor of the response to therapy, especially in hepatitis B e antigen-positive cases. Accordingly, evaluation of the percentage of HBsAg expression by objective methods in liver tissues from treatment-naïve CHB patients might provide a useful tool in the follow-up and treatment of this disease.

**INTRODUCTION**

Despite advances in its prevention, diagnosis and treatment, chronic viral B hepatitis (CHB) continues to be an important worldwide health problem with high morbidity and mortality due to its dramatic consequences, such as cirrhosis and hepatocellular carcinoma[1]. In recent years, researchers have suggested that the combination of various laboratory methods with histopathological findings may allow the assessment of the prognosis and treatment of patients with CHB[2-4]. High liver biopsy activity scores, low viremia and high serum alanine aminotransferase (ALT) levels are associated with hepatitis B e antigen (HBeAg) seroconversion, while a decrease in serum hepatitis B surface antigen (HBsAg) levels is among the factors associated with an effective response to therapy[5-7]. Unfortunately, the presence of these factors is not always adequate to determine the progression of the disease and the efficacy of treatment. The treatment response is not sufficient in some patients with these factors, which contrasts with the adequate response in other patients without these factors, suggesting that other viral and host-related parameters may also be involved in the progression and therapy response.

Recent studies have also provided evidence that in patients with CHB, the expression of viral antigens and their expression patterns detected by immunohistochemistry may be related to clinicopathological factors and might have prognostic implications for the disease process. The expression of hepatitis B core antigen (HBcAg) and its expression patterns were found to be associated with serum HBeAg, viral replication and the hepatitis activity index (HAI)[8-11]. In addition, the results of some studies have suggested the potential use of HBcAg expression as a marker of treatment response[12-14].

Regarding HBsAg, the results of many different studies investigating the relationships between the intrahepatic expression of this antigen revealed that differences in the intracytoplasmic HBsAg staining pattern were related to viremia and active inflammation[14-18]. In these investigations, HBsAg expression was evaluated semiquantitatively, and different classifications were used for the HBsAg staining pattern. On the other hand, HBsAg, beyond being the main diagnostic marker of the disease, is a very important tool in the follow-up of the course and treatment of CHB, since a decrease in its level reflects the low replicative phase[19-21]. Therefore, more objective methods are needed to determine whether the immunohistochemical expression of HBsAg in pretreatment-performed liver biopsies can be useful in monitoring the prognosis and treatment of CHB.

Therefore, this study was undertaken to investigate the relationship between image analysis-based quantitation of HBsAg expression and corresponding staining patterns with clinicopathological factors and treatment.

**MATERIALS AND METHODS**

***Study design***

This retrospective study was conducted in accordance with the Declaration of Helsinki. Approval for this study was obtained from the Akdeniz University Faculty of Medicine Clinical Research Ethics Committee (Date: January 28, 2015; Approval number: 2015.01.211.016). All patients provided written informed consent for participation in the study.

The study group consisted of 140 patients who were followed up and treated after the diagnosis of CHB at Akdeniz University Medical School between 2015 and 2020. The inclusion criteria were the presence of positive serum HBsAg for at least 6 mo and the absence of coinfection with human immunodeficiency, hepatitis delta and hepatitis C viruses. None of these patients had end-stage liver failure, immunosuppressive therapy, malignancy, autoimmune hepatitis or alcoholic hepatitis.

Clinical and laboratory parameters were recorded from all cases on the day of liver biopsy. These included the age of the patients, sex, HBV DNA, HBsAg ALT, INR, and platelet count. Forty-nine cases with elevated ALT levels underwent antiviral therapy. Ninety-one patients with low ALT levels received this treatment based on the increase in ALT levels in the first three months of follow-up. Accordingly, all patients were treated with entecavir (0.5 mg) within three months after the liver biopsies and followed up at three-month intervals. An HBV DNA level below 20 IU/mL detected by real-time polymerase chain reaction (PCR) was defined as the viral response (VR). HBeAg seroclearance and a decrease in serum HBsAg level during follow-up > 0.5 log10 IU/mL were defined as the serological response (SR) to treatment[22].

***Laboratory analysis***

Serum HBsAg, HBeAg, antibodies against HBsAg (anti-HBs) and HBeAg were tested by using commercial kits (Abbott Laboratories, Abbott Park, IL, United States). Serum HBV-DNA was extracted from 200 μL of serum by a QIAmp DNA Blood Mini Kit (QIAGEN Inc., Valencia, CA, United States) and quantified by a real-time PCR amplification assay using a LightCycler (Roche Diagnostics, Basel, Switzerland). The detection sensitivity was 20 IU/mL[23]. Serum HBsAg was quantified with the ARCHITECT HBsAg assay (Abbott Laboratories) with a dynamic range from 0.05 IU/mL to 250 IU/mL.

***Histopathological evaluation***

Microscopic evaluation was performed on serial sections stained with hematoxylin and eosin from paraffin-embedded 10% formalin-fixed liver tissues. The HAI and fibrosis stages were determined according to the Ishak system[24]. HAI grading consists of the sum of necroinflammatory scores, including portal inflammation, periportal interface hepatitis, confluent necrosis and focal lytic necrosis, and each was given a score from 0 to 4-6 with a maximal HAI grade of 18. The fibrosis staging ranged from 0 to 6.

***Immunohistochemistry***

Immunohistochemical staining for HBsAg was performed by using a primary antibody against HBsAg (mouse monoclonal antibody, clone 3E7, Dako, Carpinteria, CA, United States). Staining was performed automatically with a BenchMark XT (Ventana Medical Systems, Tucson, AZ, United States) in the Department of Pathology, Akdeniz University Medical School.

***The quantification of HBsAg***

Image analysis was performed using a SAMBA 2005 image processor (Alcatel-TITN, Grenoble, France). This system consists of a Leitz Diaplan Microscope connected to a personal computer through a Sony color camera and a data translation frame grabber board. For detection of the percentage of HBsAg immunostaining, the system was calibrated for a 20X objective. In each area, antigen immunoreactivity was measured in the entire tissue section. The results obtained were expressed as the percentage of the ratio of the HBsAg (p-HBsAg) immunoreactive area to the total scanned area.

***The immunohistochemical staining pattern***

The immunohistochemical staining pattern was evaluated separately according to 3 different previously defined classifications[15,18].

**Staining pattern 1:** The staining was classified according to the expression type and distribution into five major patterns. Patterns A and B were characterized by diffuse cytoplasmic staining in discrete hepatocytes (Figure 1A and B). While the intensity of staining was stronger in pattern A, it was faint in pattern B. In pattern C, cytoplasmic HBsAg was similar to that in pattern B but was distributed in clusters of hepatocytes (Figure 1C). Pattern D included globular and spotty cytoplasmic staining in either discrete or clustered hepatocytes (Figure 1D). Pattern E (marginal HBsAg) was characterized by submembranous staining of HBsAg beneath the cell membranes in the hepatocyte groups (Figure 1E).

**Staining pattern 2:** The expression of HBsAg was categorized according to its intracytoplasmic expression: 1: Diffuse (Figure 1A-C); 2: Globular (Figure 1D); and 3: Submembranous (Figure 1E).

**Staining pattern 3:** In this classification,HBs staining was categorized according to the presence or absence of membranous staining (Figure 1F).

***Statistical analysis***

Statistical analysis was performed using SPSS version 25.0 (IBM Corp., Armonk, NY, United States). Nominal and ordinal data are expressed as frequencies (percentages), and continuous variables are expressed as the mean (standard error). The *χ*2test was employed to examine categorical data. Continuous variables between the groups and their relationship with the clinicopathological parameters were investigated by *t* tests. Univariate analysis, including response to therapy, was estimated with the Kaplan-Meier method. The log-rank test was employed for comparisons of response rates. A Cox proportional hazards regression model was applied for multivariate analysis. Spearman’s correlation test was used to determine relationships between p-HBsAg and the other continuous variables. The threshold *P* value accepted for statistical significance was < 0.05.

**RESULTS**

The mean age of the patients in the whole group was 42.79 ± 11.53 years, and the male to female ratio was 1.25 (Table 1). There was a positive correlation between p-HBsAg and the serum levels of HBV DNA and HBsAg (r: 0.490, r: 0.468 *P* < 0.001, respectively). When all cases were grouped according to mean age, the p-HBsAg value was significantly higher in younger patients than in older patients (48.47 ± 16.06 *vs* 31.72 ± 12.89 *P* < 0.01). According to their HBeAg status, 86 cases were HBeAg positive (61.5%), and 54 cases were HBeAg negative (38.5%). Other findings of the cases are presented in Table 1. Briefly, HBeAg-positive patients were younger than HBeAg-negative patients, and serum HBV DNA and HBsAg levels were higher in this group (*P* < 0.05). On the other hand, the INR levels were found to be lower than those in the HBeAg-negative group. The average p-HBsAg level was significantly higher in the HBeAg-negative group than in the HBeAg-positive group. There was no difference in the HBsAg staining patterns between the two groups.

The relationship between viremia and histopathological findings in cases with positive HBeAg are summarized in Table 2. The average p-HBsAg level was found to be significantly higher in patients with higher HBV DNA values than in those with low HBV DNA levels. In contrast, an inverse relationship was noted between p-HBsAg and either HAI or the fibrosis score. Regarding staining pattern 1 (SP1), the number of cases with pattern A was higher among cases with higher HBV DNA levels. Their number decreased among cases with HAI ≥ 9, and it was not observed among cases with fibrosis. When staining pattern 2 (SP2) was analyzed, it was noted that the number of cases with diffuse staining was higher in the HAI ≥ 9 group. In contrast, among the cases with high HAI and fibrosis scores, the frequency of pattern E was observed in 42% and 75% of the cases, respectively.

The relationship between viremia and histopathological findings in cases with negative HBeAg is presented in Table 3. In this group, no correlation was observed between either the p-HBsAg or the membranous staining pattern and HBV DNA, HAI, or fibrosis. In the evaluation of patients according to SP1, in cases with HAI ≥ 9, the number of patients with pattern A was lower than that of patients with HAI < 9. In the latter, pattern E was rarely observed (12% of cases). SP2 was found to be related to HBV DNA levels, and diffuse staining was more frequent in cases with a higher level of HBV DNA. However, globular staining was rarely observed in these cases.

Patients were treated for a median duration of 28 mo (range, 4 mo to 62 mo). VR was achieved in 54 of 86 HBeAg-positive patients (66.7%) and in all 54 HBeAg-negative patients (100%), with a median time to VR of 16 and 9 mo, respectively. Factors associated with VR are shown in Table 4. In HBeAg-positive patients, the p-HBsAg and HBsAg expression patterns were correlated with VR, along with age, fibrosis, HVDNA and HBsAg levels in the log-rank test (*P* < 0.05). VR was observed in 53.7% of the patients with a high p-HBsAg. Similarly, pattern A and the presence of membranous expression were frequently observed in patients with VR. In multivariate analysis, the sole independent factor associated with VR was p-HBsAg (*P* < 0.01)(Figure 2A and Table 5). Forty-seven (54.6%) of 86 HBeAg-positive patients exhibited HBeAg SR, with a median time of 22 mo (range, 4 mo to 60 mo). Univariate analysis revealed that the p-HBsAg and HBsAg expression patterns were correlated with SR together with HBsAg levels and ALT > 200 (*P* < 0.05) (Table 4). Multivariate analysis demonstrated that p-HBsAg and the HAI score were independent factors related to SR (Figure 2B and C and Table 5). On the other hand, in the HBeAg-negative group, the most valuable determinant of VR was found to be the HAI score (Figure 2D and Table 5).

**DISCUSSION**

The few previous studies that evaluated HBsAg expression by semiquantitative methods have shown that serum HBsAg levels are correlated with the number of intrahepatic HBsAg-positive cells[19,20]. This observation was interpreted as serum and hepatic HBsAg being interrelated. On the other hand, in our study, we determined the percentage of intrahepatic HBsAg expression in the whole tissue section of each case by image analysis. Our results showed that in HBeAg-positive cases, p-HBsAg was associated not only with serum HBsAg but also with serum HBV DNA levels, indicating that intrahepatic HBsAg in liver tissues is closely related to viremia. Recently, Su *et al*[25] investigated HBsAg expression in the liver and its relationship with both serum HbsAg and interferon therapy using morphometric methods in treatment-naïve HbeAg-positive patients. Parallel to the findings of the present study, they observed a strong correlation between hepatic HbsAg expression and serum HbsAg levels. In addition, the relationship between HbsAg level and HbeAg loss after interferon therapy observed in this study suggested that the HbsAg expression level may be a useful marker to determine the response to therapy.

Therefore, in our study, we also aimed to elucidate the relationship between p-HbsAg levels and the treatment response in patients receiving entecavir treatment together with other HbsAg expression patterns. In the HbeAg-positive group, VR and SR were observed more frequently in cases with pretreatment values of p-HbsAg levels above the group average. In the HbeAg-positive group, the pretreatment values of p-HbsAg were strongly related to either VR or SR, suggesting the clinical significance of p-HbsAg in the management of HbeAg-positive CHB patients with antiviral treatment. Moreover, multivariate analysis revealed that in this group, p-HbsAg was an independent factor associated with VR and SR, warranting further investigation to evaluate the potential role of p-HbsAg as a tool to predict the response to therapy.

Another interesting result of this study is the presence of an inverse relationship between p-HbsAg and HAI and fibrosis scores in the HbeAg-positive group. Parallel to this finding, in cases with high HAI and/or fibrosis scores, the serum HBV DNA and HbsAg levels were significantly lower, suggesting that in the liver, the replacement of hepatocytes by both increased inflammatory infiltration and increased connective tissue can lead to this situation.

Our study demonstrated that HbsAg expression patterns, evaluated by various immunohistochemical methods, have different relationships with clinicopathological parameters and treatment responses in patients with CHB. Many years ago, following the detection of ground-glass hepatocytes by immunohistochemical methods in the liver tissues of patients with anti-Hbe positivity in the serum and a low HBV DNA level, the relationship of HbsAg expression patterns in hepatocytes with clinicopathological factors was investigated.Some studies have shown that patterns A and B, as opposed to those of C and D, are more common in younger subjects[15,18]. However, no similar age-related difference finding in HbsAg staining patterns was observed in other studies[16]. In our study, most of the patients with patterns A and B were younger than those with patterns C and D.

When the relationship of these staining patterns with disease activity and viremia has been investigated in recent studies, most of the data indicate that patterns A and B are associated with viremia[15,18,21,26,27]. In parallel with this finding, in our study, these patterns were associated with high HBV DNA and HAI in the HbeAg-positive group and were rarely observed in cases with high fibrosis scores, supporting the hypothesis that both are associated with active disease. Interestingly, the frequent detection of pattern B in the HbeAg-negative group, especially in cases with high HAI scores, also points to a relationship with active disease in these cases[15]. While the preliminary results indicate that the marginal staining pattern (pattern E) may be a determinant of inactive virus replication, other studies concluded that pattern C may be a marker of this situation[15,28]. In our cases, pattern E was more frequent in cases with low levels of HBV DNA and more frequent in cases with high fibrosis scores in the HbeAg-positive group, supporting data about its relationship with inactive disease reported by previous studies. Similar to recent findings in HbeAg-positive cases, membranous expression was absent in 84% and 87% of patients with lower HBV DNA values and fibrosis scores, respectively[18]. These findings suggest that this pattern of staining reflects active liver disease.

In an elegant study, Yim *et al*[26] investigated the relationship between these staining patterns and antiviral treatment response in patients with CHB. Diffuse staining patterns were associated with SR in HBeAg-positive patients. However, multivariate analysis revealed that this was not an independent predictor. In our study, although HBsAg expression patterns were associated with VR and SR in univariate analysis, they were not found to be independent factors. In contrast, p-HBsAg was a good predictor of the response to therapy in HBeAg-positive cases. This finding emphasizes that the determination of HBsAg expression by quantitative methods, beyond its distribution within the cell, may be an objective marker that can be used during the course and treatment of the disease.

In recent years, there has been growing interest in developing various noninvasive methods for monitoring the course and treatment of CHB. On the other hand, although it is an expensive and invasive method, liver biopsy remains the gold standard in determining liver injury and fibrosis during the follow-up of the disease. However, beyond these parameters, our results emphasize that the HBsAg expression levels measured in pretreatment liver biopsies *via* quantitative methods may be a useful marker for determining the treatment response.

**CONCLUSION**

This study demonstrated that HBsAg expression patterns, evaluated by various immunohistochemical methods, have different relationships with clinicopathological parameters and the treatment response in patients with CHB. Although the expression pattern of HBsAg differs significantly according to the disease status, it shows lower clinical significance in predicting the response to therapy than the percentage of HBsAg expression. Therefore, the quantitative evaluation of HBsAg expression in pretreatment-naïve liver biopsies of patients with CHB may provide a prediction of the response to antiviral therapy, especially in cases with HBeAg positivity. However, our findings should be supported by additional large studies.

**ARTICLE HIGHLIGHTS**

***Research background***

The combination of various laboratory methods with histopathological findings is not always adequate to predict the progression and treatment response in patients with chronic viral B hepatitis (CHB). Therefore, efforts are being made to determine new parameters that may be useful in the follow-up of the disease.

***Research motivation***

It has been reported that differences in intracytoplasmic hepatitis B surface antigen (HBsAg) staining patterns are associated with viremia and active inflammation in CHB. However, HBsAg expression was evaluated semiquantitatively in these studies, and different classifications were used for the HBsAg staining pattern.

***Research objectives***

To investigate the relationship between image analysis-based quantitation of HBsAg expression and staining patterns with clinicopathological factors and treatment.

***Research methods***

The study group consisted of 140 patients who were followed up and treated after the diagnosis of CHB. All cases were treated with entecavir. The evaluation of immunohistochemical HBsAg staining was performed in whole tissue sections by image analysis and expressed as a percentage of HBsAg staining (p-HBsAg). The staining pattern was also evaluated separately according to 3 different previously defined classifications. The correlations among immunohistochemical and clinicopathological findings were analyzed by *χ*2 and *t*-tests. The log-rank test was employed for comparisons of response rates. A Cox proportional hazards regression model was applied for multivariate analysis. Spearman’s correlation test was used to determine relationships between p-HBsAg and the other continuous variables.

***Research results***

There was a positive correlation between p-HBsAg and the serum levels of hepatitis B virus (HBV) DNA and HBsAg. The p-HBsAg was significantly higher in younger patients than in older patients, and the mean p-HBsAg level was significantly higher in the HBeAg-negative group than in the HBeAg-positive group. In the HBeAg-positive group, the average p-HBsAg level was significantly higher in patients with higher HBV DNA values than in those with low HBV DNA levels. Furthermore, an inverse relationship was noted between p-HBsAg and either hepatitis activity index (HAI) or the fibrosis score. Regarding staining pattern, diffuse cytoplasmic staining in many discrete hepatocytes was higher among cases with higher HBV DNA levels, and their number decreased among patients with HAI ≥ 9. However, submembranous staining was more frequent in subjects with higher HAI and fibrosis scores. In the same group, the p-HBsAg and HBsAg expression patterns were correlated with the viral response (VR) and serological response (SR). Multivariate analysis revealed that p-HBsAg was an independent factor associated with VR and SR. In the HBeAg-negative group, no correlation was observed between either the p-HbsAg or the membranous staining pattern and HBV DNA, HAI, or fibrosis. Diffuse staining was more frequent in cases with a higher level of HBV DNA. On the other hand, globular staining was rarely observed in these cases. In this group, the most valuable determinant of VR was the HAI score.

***Research conclusions***

We have demonstrated that the expression pattern of HBsAg shows lower clinical significance in predicting the response to therapy than the percentage of HBsAg expression. Therefore, the quantitative evaluation of HBsAg expression in pretreatment-naïve liver biopsies of patients with CHB may predict the response to antiviral therapy, especially in cases with HBeAg positivity.

***Research perspectives***

The evaluation of HBsAg expression by image analysis in pretreatment-naïve liver biopsies may be a valuable tool to predict the response to antiviral therapy. However, our findings should be supported by additional large-scaled studies.

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**Footnotes**

**Institutional review board statement:** The study was reviewed and approved by the Akdeniz University Clinical Research Institutional Board (approval No. 16-012-211).

**Informed consent statement:** Patients were not required to give informed consent to the study because the analysis used anonymous data that were obtained after each patient agreed to treatment by written consent.

**Conflict-of-interest statement:** There is no conflict of interest.

**Data sharing statement:** No additional data are available.

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**Provenance and peer review:** Invited article; Externally peer reviewed.

**Peer-review model:** Single blind

**Peer-review started:** May 15, 2021

**First decision:** July 8, 2021

**Article in press:** December 31, 2021

**Specialty type:** Gastroenterology and hepatology

**Country/Territory of origin:** Turkey

**Peer-review report’s scientific quality classification**

Grade A (Excellent): 0

Grade B (Very good): B

Grade C (Good): 0

Grade D (Fair): 0

Grade E (Poor): 0

**P-Reviewer:** Dang SS **S-Editor:** Gao CC **L-Editor:** A **P-Editor:** Gao CC

**Figure Legends**



**Figure 1 Different staining patterns of hepatitis B surface antigen in liver tissues.**A: Dense cytoplasmic staining in many discrete hepatocytes; B: Diffuse but faint cytoplasmic staining in discrete hepatocytes; C: Diffuse faint cytoplasmic staining in a group of hepatocytes near an inflamed portal area; D: Globular and spotty staining in discrete and in a small group of hepatocytes; E: Submembranous staining in a large group of cells; F: Membranous expression of hepatitis B surface antigen. Original magnifications A and C × 200, B, D, E and F × 400, counter-stained with Mayer’s hematoxylin.

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**Figure 2 Kaplan-Meier analysis in hepatitis B surface antigen positive and hepatitis B e antigen negative cases (*P* < 0.01).** A: Cumulative incidence of viral response in hepatitis B e antigen (HBeAg) positive patients is more frequently observed in cases with higher percentage of immunohistochemical hepatitis B surface antigen (p-HBsAg) expression; B-D: In the same group, serological response is also significantly related to p-HBsAg expression (B) and HBsAg levels (C). However, in HBeAg negative in patients, VR is closely related to hepatitis activity index (D). p-HBsAg: Percentage of immunohistochemical hepatitis B surface antigen; HBsAg: Hepatitis B surface antigen; HAI: Hepatitis activity index; VR: Viral response; SR: Serological response.

**Table 1 Basic clinical, histopathological and immunohistochemical findings according to hepatitis B e antigen status**

|  |  |  |  |
| --- | --- | --- | --- |
| **Parameters** | **Total** | **HBeAg positive** | **HBeAg negative** |
| *n* | 140 | 86 | 54 |
| Age (yr) | 42.75 ± 11.53 | 40.30 ± 11.03 | 46.74 ± 11.29a |
| Gender |  |  |  |
| Male | 78 | 48 (61.5) | 30 (38.5) |
| Female | 62 | 38 (61.3) | 24 (38.7) |
| HBV DNA (log10IU/mL) | 7.27 ± 2.17 | 7.67 ± 2.39 | 6.65 ± 1.58a |
| HBsAg (log10IU/mL) | 3.85 ± 1.04 | 4.01 ± 1.07 | 3.61 ± 0.94b |
| ALT IU/L | 7.27 ± 2.17 | 155.80 ± 114.32 | 134.54 ± 98.71 |
| ALT < 200 IU/L | 91 | 52 (51.7) | 39 (42.9) |
| ALT ≥ 200 IU/L | 49 | 34 (69.4) | 15 (30.6) |
| INR | 1.08 ± 0.21 | 1.04 ± 0.14 | 1.15 ± 0.27b |
| Platelets (× 103/μL) | 226.4 ± 73.43 | 224.54 ± 70.67 | 227.59 ± 75.53 |
| HAI score ≥ 9 | 61 | 38 (62.3) | 23 (37.7) |
| HAI score < 9 | 79 | 48 (60.8) | 31 (39.2) |
| Fibrosis ≥ 4 | 53 | 33 (62.3) | 20 (37.7) |
| Fibrosis < 4 | 87 | 53 (60.9) | 34 (39.1) |
| p-HBsAg | 41.05 ± 22.78 | 47.2 ± 13.98 | 31.26 ± 13.98a |
| HBsAg SP1 |  |  |  |
| A | 33 | 17 (51.5) | 16 (48.5) |
| B | 24 | 10 (41.7) | 14 (58.3) |
| C | 21 | 16 (76.2) | 5 (23.8) |
| D | 16 | 10 (62.5) | 6 (37.5) |
| E | 46 | 33 (71.7) | 13 (28.3) |
| HBsAg SP2 |  |  |  |
| Diffuse | 59 | 29 (49.2) | 30 (50.8) |
| Globular | 34 | 24 (70.6) | 10 (29.4) |
| Submembranous | 47 | 33 (70.2) | 14 (29.8) |
| Membranous expression |  |  |  |
| Present | 53 | 33 (63.3) | 20 (37.7) |
| Absent | 87 | 53 (60.9) | 34 (39.1) |

Data are presented as mean ± SD, number (%).

a*P* < 0.05.

b*P* < 0.01.

p-HbsAg: Percentage of hepatitis B surface antigen expression; SP: Staining pattern; ALT: Alanine aminotransferase; INR: International normalized ratio, HAI: Histologic activity index; HBsAg: Hepatitis B surface antigen; HBV: Hepatitis B virus.

**Table 2 The correlation of clinicopathologic factors and hepatitis B surface antigen expression patterns with viral replication in hepatitis B e antigen positive cases**

|  |  |  |  |
| --- | --- | --- | --- |
|  | **HBV DNA** | **HAI** | **Fibrosis** |
| **Parameters** | **< 7.67** | **≥ 7.67** | **< 9** | **≥ 9** | **< 4** | **≥ 4** |
| *n* | 32 | 54 | 48 | 38 | 53 | 33 |
| Age (yr) | 48.28 ± 9.6 | 35.57 ± 8.93b | 37.79 ± 10.83 | 43.47 ± 10.59a | 36.06 ± 9.39 | 47.12 ± 10.11b |
| Gender |  |  |  |  |  |  |
| Male | 23 (71.9) | 25 (46.3)a | 23 (47.9) | 25 (65.8) | 27 (50.9) | 21 (63.6) |
| Female | 9 (28.1) | 29 (53.7) | 25 (52.1) | 13 (34.2) | 26 (49.1) | 12 (36.4) |
| HBV DNA (log10IU/mL) | - | - | 8.36 ± 2.26 | 6.79 ± 2.30a | 8.15 ± 2.32 | 6.89 ± 2.34 |
| HBsAg (log10IU/mL) | 3.02 ± 0.91 | 4.59 ± 0.64b | 4.28 ± 0.91 | 3.66 ± 1.61a | 4.19 ± 1.01 | 3.07 ± 1.10a |
| ALT IU/L | 106.97 ± 73.55 | 184.74 ± 124.85a | 148.97 ± 61.46 | 121.46 ± 50.84 | 167.74 ± 72.1 | 136.64 ± 56.2 |
| ALT< 200 IU/L | 26 (81.2) | 26 (48.1)a | 34 (70.8) | 18 (47.4)a | 32 (60.4) | 20 (60.6) |
| ALT ≥ 200 IU/L | 6 (18.8) | 28 (51.9) | 14 (29.2) | 20 (52.6) | 21 (39.6) | 13 (39.4) |
| INR | 1.03 ± 0.08 | 1.05 ± 0.16 | 1.03 ± 0.10 | 1.06 ± 0.17 | 1.04 ± 0.16 | 1.04 ± 0.09 |
| Platelets (× 103/μL) | 217.59 ± 66.85 | 233.98 ± 80.60 | 226 ± 74.74 | 228 ± 77.51 | 224.53 ± 74.72 | 232.61 ± 77.82 |
| HAI score ≥ 9 | 22 (68.8) | 16 (29.6)b | - | - | 17 (32.1) | 21 (63.6)b |
| HAI score < 9 | 10 (31.2) | 38 (70.4) | - | - | 36 (67.9) | 12 (36.4) |
| Fibrosis ≥ 4 | 19 (59.4) | 14 (25.9)a | 12 (25) | 21 (55.3)b | - | - |
| Fibrosis < 4 | 13 (40.6) | 40 (74.1) | 36 (75) | 17 (44.7) | - | - |
| p-HBsAg | 29.92 ± 10.84 | 57.44 ± 25.50b | 53.53 ± 25.12 | 39.21 ± 22.85a | 54.49 ± 26.02 | 35.49 ± 18.34b |
| HBsAg SP1 |  |  |  |  |  |  |
| A | 2 (6.2) | 15 (27.8)a | 11 (22.9 | 6 (15.8)a | 17 (32.1) | 0b |
| B | 4 (12.5) | 6 (11.1) | 1 (2.1) | 9 (23.7) | 6 (11.3) | 4 (12.1) |
| C | 6 (18.8) | 10 (18.5) | 12 (25) | 4 (10.5) | 14 (26.4) | 2 (6.1) |
| D | 4 (12.5) | 6 (11.1) | 7 (14.6) | 3 (7.9) | 8 (15.1) | 2 (6.1) |
| E | 16 (50) | 17 (31.5) | 17 (35.4) | 16 (42.1) | 8 (15.1) | 25 (75.8) |
| HBsAg SP2 |  |  |  |  |  |  |
| Diffuse | 8 (25) | 21 (38.9) | 12 (25) | 17 (44.7)a | 23 (43.4) | 6 (18.2)b |
| Globular | 8 (25) | 16 (29.6) | 19 (39.6) | 5 (13.1) | 22 (41.5) | 2 (6.1) |
| Submembranous | 16 (50) | 17 (31.5) | 17 (35.4) | 16 (42.1) | 8 (15.1) | 25 (75.8) |
| Membranous expression |  |  |  |  |  |  |
| Present | 5 (15.6) | 28 (51.9)b | 21 (43.8) | 12 (31.6) | 29 (54.7) | 4 (12.1)b |
| Absent | 27 (84.4) | 26 (48.1) | 27 (56.3) | 26 (68.4) | 24 (45.3) | 29 (87.9) |

Data are presented as mean ± SD, number (%).

a*P* < 0.05.

b*P* < 0.01.

p-HbsAg: Percentage of hepatitis B surface antigen expression; SP: Staining pattern; ALT: Alanine aminotransferase; INR: International normalized ratio; HAI: Histologic activity index; HBsAg: Hepatitis B surface antigen; HBV: Hepatitis B virus.

**Table 3 The correlation of clinicopathologic factors and hepatitis B surface antigen expression patterns with viral replication in hepatitis B e antigen negative cases**

|  |  |  |  |
| --- | --- | --- | --- |
|  | **HBV DNA** | **HAI** | **Fibrosis** |
| **Parameters** | **< 6.65** | **≥ 6.65** | **< 9** | **≥ 9** | **< 4** | **≥ 4** |
| *n* | 23 | 31 | 31 | 23 | 34 | 20 |
| Age (yr) | 48.13 ± 10.34 | 45.71 ± 12.02 | 46.52 ± 11.73 | 47.04 ± 10.92 | 45.82 ± 11.63 | 48.30 ± 10.81 |
| Gender |  |  |  |  |  |  |
| Male | 10 (43.5) | 20 (64.5) | 19 (61.3) | 11 (47.8) | 23 (67.6) | 7 (35)a |
| Female | 13 (56.5) | 11 (35.5) | 12 (38.7) | 12 (52.2) | 11 (32.4) | 13 (65) |
| HBV DNA (log10IU/mL) | - | - | 4.97 ± 1.49 | 7.56 ± 1.21a | 4.16 ± 0.97 | 3.98 ± 0.93 |
| HBsAg (log10IU/mL) | 3.19 ± 0.84 | 3.92 ± 0.90 | 3.30 ± 0.8 | 3.86 ± 0.91a | 3.46 ± 0.95 | 4.03 ± 0.98 |
| ALT IU/L | 96.09 ± 33.55 | 163.06 ± 50.85a | 102.55 ± 41.36 | 177.65 ± 80.65 | 126.18 ± 64.87 | 148.75 ± 58.6 |
| ALT < 200 IU/L | 21 (91.3) | 18 (58.1)b | 26 (83.9) | 13 (56.5)a | 25 (73.5) | 14 (70) |
| ALT ≥ 200 IU/L | 2 (8.7) | 13 (41.9) | 5 (16.1) | 10 (43.5) | 9 (26.5) | 6 (30) |
| INR | 1.13 ± 0.27 | 1.17 ± 0.28 | 1.15 ± 0.3 | 1.16 ± 0.24 | 1.22 ± 0.32 | 1.04 ± 0.10b |
| Platelets (× 103/μL) | 234.91 ± 80.6 | 216.31 ± 61.29 | 237.67 ± 73.34 | 206.64 ± 73.34 | 218.21 ± 71.85 | 236.5 ± 68.79 |
| HAI score ≥ 9 | 2 (8.7) | 21 (67.7)b | - | - | 14 (26.5) | 9 (70)b |
| HAI score < 9 | 21 (91.3) | 10 (32.3) | - | - | 20 (73.5) | 11 (30) |
| Fibrosis ≥ 4 | 4 (17.4) | 16 (51.6)a | 6 (19.4) | 14 (60.9)b | - | - |
| Fibrosis < 4 | 19 (82.6) | 15 (48.4)  | 25 (80.6) | 9 (39.1) | - | - |
| p-HBsAg | 30.39 ± 14.18 | 31.91 ± 14.04 | 28.68 ± 11.16 | 34.75 ± 16.71 | 33.21 ± 15.32 | 27.96 ± 10.93 |
| HBsAg SP1 |  |  |  |  |  |  |
| A | 6 (26.1) | 10 (32.3) | 13 (41.9) | 3 (13)a | 13 (38.2) | 3 (15) |
| B | 5 (21.7) | 9 (29) | 5 (16.1) | 9 (39.1) | 7 (20.6) | 7 (35) |
| C | 4 (17.4) | 1 (3.2) | 4 (12.9) | 1 (4.3) | 5 (14.7) | 0 |
| D | 5 (21.7) | 1 (3.2) | 5 (16.1) | 1 (4.3) | 3 (8.8) | 3 (15) |
| E | 3 (13) | 10 (32.3) | 4 (12.9) | 9 (39.1) | 6 (17.6) | 7 (35) |
| HBsAg SP2 |  |  |  |  |  |  |
| Diffuse | 11(47.8) | 19 (61.3)b | 18 (58.1) | 12 (52.2)a | 20 (58.8) | 10 (50) |
| Globular | 9 (39.1) | 2 (3.2) | 9 (29) | 1 (4.3) | 7 (20.6) | 3 (15) |
| Submembranous | 3 (13.1) | 10 (31.5) | 4 (12.9) | 10 (43.5) | 7 (20.6) | 7 (35) |
| Membranous expression |  |  |  |  |  |  |
| Present | 7 (30.4) | 13 (41.9) | 10 (32.3) | 10 (43.5) | 14 (41.2) | 6 (30) |
| Absent | 16 (69.6) | 18 (58.1) | 21 (67.7) | 13 (56.5) | 20 (58.8) | 14 (70) |

Data are presented as mean ± SD, number (%).

a*P* < 0.05.

b*P* < 0.01.

p-HbsAg: Percentage of hepatitis B surface antigen expression; SP: Staining pattern; ALT: Alanine aminotransferase; INR: International normalized ratio; HAI: Histologic activity index; HBsAg: Hepatitis B surface antigen; HBV: Hepatitis B virus.

**Table 4 The relationship between clinicopathological factors and hepatitis B surface antigen expression patterns with viral response and serological response**

|  |  |  |
| --- | --- | --- |
|  | **HBeAg positive** | **HBeAg negative** |
|  | **VR** | **SR** | **VR** |
| Parameters | Absent | Present | Absent | Present | Present |
| Age (yr) |  |  |  |  |  |
| < Mean | 11 (34.4) | 39 (72.2)b | 19 (48.7) | 31 (66)b | 24 (44.4) |
| ≥ Mean | 21 (65.6) | 15 (27.8) | 20 (51.43) | 16 (34) | 30 (55.6) |
| Gender |  |  |  |  |  |
| Male | 21 (34.4) | 27 (50) | 24 (61.5) | 24 (48.9) | 30 (55.6) |
| Female | 11 (34.4) | 27 (50) | 15 (38.5) | 23 (51.1) | 24 (44.4) |
| HBV DNA |  |  |  |  |  |
| Low | 17 (53.1) | 39 (72.2)a | 16 (41) | 16 (34) | 23 (42.6) |
| High | 15 (45.9) | 15 (27.8) | 23 (59) | 31 (66) | 31 (57.4) |
| HBsAg |  |  |  |  |  |
| Low | 21 (65.6) | 15 (27.8)a | 21 (53.8) | 15 (31.9)b | 30 (55.6) |
| High | 11 (34.4) | 39 (72.2) | 18 (46.2) | 32 (68.1) | 24 (44.4) |
| ALT |  |  |  |  |  |
| < 200 IU/L | 23 (71.9) | 29 (53.7) | 29 (74.4) | 23 (48.9)a | 39 (72.2) |
| ≥ 200 IU/L | 9 (28.1) | 25 (46.3) | 10 (25.6) | 24 (51.1) | 15 (27.8) |
| INR |  |  |  |  |  |
| Low | 15 (46.9) | 26 (48.1) | 19 (48.7) | 22 (46.8) | 39 (72.2) |
| High | 17 (53.1) | 28 (51.9) | 20 (51.3) | 25 (53.2) | 15 (27.8) |
| Platelets |  |  |  |  |  |
| Low | 15 (46.9) | 31 (57.4) | 20 (51.3) | 26 (55.3) | 31 (57.4) |
| High | 17 (53.1 | 23 (42.6) | 19 (48.7) | 21 (44.7) | 23 (42.6) |
| HAI score ≥ 9 | 13 (40.6) | 25 (46.3) | 11 (25) | 28 (59.6)a | 23 (42.6) |
| HAI score < 9 | 19 (59.4) | 29 (53.7) | 28 (75) | 19 (40.4) | 31 (57.4) |
| Fibrosis ≥ 4 | 17 (53.1) | 16 (29.6)a | 17 (43.6) | 16 (34) | 20 (37) |
| Fibrosis < 4 | 15 (46.9) | 10 (70.4) | 22 (56.4) | 31 (66) | 34 (63) |
| p-HbsAg |  |  |  |  |  |
| Low | 26 (81.3) | 25 (46.3)b | 29 (74.4) | 24 (51.1)b | 21 (38.9) |
| High | 6 (18.8) | 29 (53.7) | 10 (25.6) | 23 (48.9) | 33 (61.1) |
| HBsAg SP1 |  |  |  |  |  |
| A | 1 (3.1) | 16 (29.6)b | 2 (5.1) | 15 (31.9)b | 16 (29.6) |
| B | 0 | 10 (18.5) | 0 | 10 (21.3) | 14 (25.9) |
| C | 6 (18.8) | 10 (18.5) | 11 (28.2) | 5 (10.6) | 5 (9.3) |
| D | 6 (18.8) | 4 (7.4) | 6 (15.4) | 4 (8.5) | 6 (11.1) |
| E | 19 (59.4) | 14 (25.9) | 20 (51.3) | 13 (27.7) | 13 (24.1) |
| HBsAg SP2 |  |  |  |  |  |
| Diffuse | 1 (3.1) | 28 (58.1)b | 2 (5.1) | 2 (57.4)b | 30 (55.6) |
| Globular | 12 (37.5) | 12 (22.2) | 17 (43.6) | 7 (14.9) | 10 (18.5) |
| Submembranous | 19 (59.4) | 14 (25.9) | 20 (51.3) | 13 (27.7) | 14 (25.9) |
| Membranous expression |  |  |  |  |  |
| Absent | 27 (84.4) | 26 (48.1)b | 29 (74.4) | 24 (51.1)b | 21 (38.9) |
| Present | 5 (15.6) | 28 (51.9) | 10 (25.6) | 23 (48.9) | 33 (61.1) |

a*P* < 0.05.

b*P* < 0.01.

VR: Viral response; SR: Serological response; p-HbsAg: Percentage of hepatitis B surface antigen expression; SP: Staining pattern; ALT: Alanine aminotransferase; INR: International normalized ratio; HAI: Histologic activity index; HBsAg: Hepatitis B surface antigen; HBeAg: Hepatitis B e antigen; HBV: Hepatitis B virus.

**Table 5 The association of clinicopathological factors and hepatitis B surface antigen expression patterns with viral response and serological response**

|  |  |  |
| --- | --- | --- |
|  | **HBeAg positive (*n* = 86)** | **HBeAg negative (*n* = 54)** |
| **Parameters** | **VR** | **SR** | **VR** |
|  | **HR (95%CI)** | ***P* value1** | ***P* value2** | **HR (95%CI)** | ***P* value1** | ***P* value2** | **HR (95%CI)** | ***P* value2** |
| Age (yr) | 0.685 (0.228-2.055) | 0.001 | 0.449 | 0.892 (0.301-2.644) | 0.007 | 0.837 | 0.936 (0.460-1.904) | 0.855 |
| Gender: male *vs* female | 0.581 (0.301-1.121) | 0.72 | 0.105 | 0.516 (0.251-1.062) | 0.79 | 0.072 | 2.410 (0.986-5.985) | 0.054 |
| HBV DNA (log10IU/mL) | 0.782 (0.275-2.224) | 0.003 | 0.645 | 0.707 (0.246-2.029) | 0.40 | 0.519 | 0.620 (0.255-1.508) | 0.292 |
| HBsAg (log10IU/mL) | 1.931 (0.757-4.923) | 0.001 | 0.168 | 2.252 (0.878-5.773) | 0.001 | 0.091 | 0.700 (0.294-1.669) | 0.422 |
| ALT ≥ 200 IU/L *vs* < 200 IU/L | 0.992 (0.476-2.068) | 0.90 | 0.982 | 1.037 (0.436-2.471) | 0.018 | 0.934 | 0.854 (0.372-1.962) | 0.710 |
| INR | 0.539 (0.271-1.072) | 0.751 | 0.078 | 0.493 (0.222-1.092) | 0.901 | 0.081 | 1.815 (0.791-4.161) | 0.159 |
| Platelets (× 103/μL) | 0.619 (0.309-1.240) | 0.194 | 0.176 | 0.538 (0.243-1.189) | 0.389 | 0.125 | 0.925 (0.452-1.892) | 0.831 |
| HAI score ≥ 9 *vs* < 9 | 1.566 (0.692-3.546) | 0.282 | 0.282 | 2.881 (1.170-7.094) | 0.132 | 0.021 | 6.876 (2.219-21.309) | 0.001 |
| Fibrosis ≥ 4 *vs* < 4 | 1.245 (0.517-2.999) | 0.021 | 0.625 | 1.079 (0.427-2.730) | 0.092 | 0.872 | 0.833 (0.326-2.125) | 0.702 |
| p-HBsAg | 3.178 (1.582-6.387) | 0.001 | 0.001 | 3.990 (1.843-8.638) | 0.001 | 0.001 | 1.089 (0.575-2.063) | 0.793 |
| SP1 | 0.899 (0.495-1.634) | 0.001 | 0.727 | 3.760 (1.742-7.861) | 0.001 | 0.531 | 0.959 (0.443-2.167) | 0.990 |
| SP2 | 0.727 (0.240-2.200) | 0.001 | 0.573 | 0.721 (0.216-2.410) | 0.001 | 0.595 | 0.996 (0.552-1.798) | 0.990 |
| Membranous expression | 1.085 (0.513-2.294) | 0.001 | 0.831 | 1.082 (0.472-2.482) | 0.001 | 0.952 | 0.692 (0.246-1.944) | 0.485 |

1Univariate analysis.

2Cox regression analysis.

VR: Viral response; SR: Serological response; HR: Hazard ratio; CI: Confidence interval; p-HbsAg: Percentage of hepatitis B surface antigen expression; SP: Staining pattern; ALT: Alanine aminotransferase; INR: International normalized ratio; HAI: histologic activity index; HBsAg: Hepatitis B surface antigen; HBeAg: Hepatitis B e antigen.



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