

WJG 20th Anniversary Special Issues (14): Pancreatic cancer**Pancreatic biomarkers: Could they be the answer?**

Angela Lamarca, Jaime Feliu

Angela Lamarca, Department of Medical Oncology, The Christie NHS Foundation Trust, Manchester M20 4BX, United Kingdom
Angela Lamarca, Jaime Feliu, Department of Medical Oncology, La Paz University Hospital, 28046 Madrid, Spain

Author contributions: Lamarca A and Feliu J performed manuscript writing and reviewing.

Correspondence to: Angela Lamarca, MD, PhD, Department of Medical Oncology, The Christie NHS Foundation Trust, Wilmslow Road, Manchester M20 4BX,

United Kingdom. angela.lamarca@christie.nhs.uk

Telephone: +44-16-14468106 Fax: +44-16-14463468

Received: October 29, 2013 Revised: December 11, 2013

Accepted: January 14, 2014

Published online: June 28, 2014

Core tip: Implementing the clinicopathological information with molecular characteristics for treatment individualization in pancreatic cancer seems to be one of the keys to improving survival and response to treatment. The development of new biomarkers and a better definition of the current ones are radically important. This review will summarize the most important biomarkers defined for pancreatic adenocarcinoma and their current development status.

Lamarca A, Feliu J. Pancreatic biomarkers: Could they be the answer? *World J Gastroenterol* 2014; 20(24): 7819-7829 Available from: URL: <http://www.wjgnet.com/1007-9327/full/v20/i24/7819.htm> DOI: <http://dx.doi.org/10.3748/wjg.v20.i24.7819>

Abstract

Pancreatic ductal adenocarcinoma (PDA) is known for its poor prognosis. Most of the patients are diagnosed with advanced stages, when no curative treatment is available. Currently, despite extensive clinical research on PDA, the median overall survival remains short. Diagnosis delay and primary chemo-resistance due to its intrinsic biological nature may explain the challenges to improve our results. Our knowledge about the molecular biology of PDA has exponentially increased during the last decades and its use for the development of biomarkers could help to reach better results in the clinical setting. These biomarkers could be the clue for the improvement in PDA clinical research by earlier detection strategies with diagnostic biomarkers, and by an individualization of treatment approach with prognostic and predictive biomarkers. This review summarizes the current knowledge about the molecular biology of PDA and the status of the most important prognostic and predictive biomarkers.

© 2014 Baishideng Publishing Group Inc. All rights reserved.

Key words: Pancreatic adenocarcinoma; Biomarkers; Diagnosis; Prognostic; Predictive; Treatment

INTRODUCTION

Pancreatic ductal adenocarcinoma (PDA) is known for its aggressiveness and poor prognosis: it is the fourth leading cause of cancer-related death both in men and women^[1]. Approximately 45220 patients are annually diagnosed with pancreatic adenocarcinoma; almost all are expected to die from the disease^[2]. Five-year survival rate after the diagnosis is around 5% for all the stages, reaching 20% for the localized stages and being less than 1% for those patients diagnosed with advanced disease.

The majority of pancreatic tumours (85%) are classified as adenocarcinomas (PDA), arising from the ductal epithelium. The diagnosis is mainly made in patients in their forties and the incidence is higher in men than in women (ratio 1.3:1). Some risk factors have been suggested for the development of PDA, but no standard screening has been defined yet (Table 1). Five to ten percent of the patients diagnosed with PDA have a first degree relative with the same disease, which suggests involvement of familial aggregation and/or genetic factors^[4].

Surgical resection is the only option of curative treatment. Nevertheless, because of the late presentation of

Table 1 Suggested risk factors for the development of pancreatic ductal adenocarcinoma^[3]

Hereditary syndromes	Non-hereditary risk factors
Hereditary breast/ovarian cancer (BRCA2, BRCA1, PALB2)	Nonhereditary chronic pancreatitis
Familial atypical multiple mole melanoma (FAMMM) syndrome (CDKN2A)	Diabetes mellitus, glucose metabolism, and insulin resistance
Peutz-Jeghers syndrome (STK11)	Cigarette smoking
Familial adenomatous polyposis (APC)	Obesity and physical inactivity
Hereditary nonpolyposis colon cancer (Lynch II) (DNA mismatch repair genes)	Diet (high intake of saturated fat and/or meat, particularly smoked or processed meats)
Familial pancreatic cancer (gene not identified)	Coffee and alcohol consumption
Hereditary pancreatitis (PRSS1, SPINK1)	Aspirin and nonsteroidal anti-inflammatory drug use
Ataxia telangiectasia (ATM)	History of partial gastrectomy or cholecystectomy
Li-Fraumeni syndrome (p53)	<i>Helicobacter pylori</i> infection

Table 2 Summary of the most important randomized clinical trials performed in advanced pancreatic ductal adenocarcinoma

Experimental arm treatment (number of patients included)	Median OS (mo) (Experimental arm) (95%CI)	Control arm treatment (number of patients included)	Median OS (mo) (Control arm) (95%CI)	Hazard ratio (95%CI) (P value)	Ref.
Gemcitabine (63 pts)	5.6 (data not shown)	5-FU (63 pts)	4.4 (data not shown)	Data not shown P = 0.0025	Burris <i>et al</i> ^[7] , 1997
Gemcitabine and erlotinib (285 pts)	6.24 (data not shown)	Gemcitabine (284 pts)	5.91 (data not shown)	0.82 (0.69-0.99) P = 0.038	Moore <i>et al</i> ^[8] , 2007
Gemcitabine and capecitabine (267 pts)	7.1 (6.2-7.8)	Gemcitabine (266 pts)	6.2 (5.5-7.2)	0.86 (0.72-1.02) P = 0.08	Cunningham <i>et al</i> ^[9] , 2009
FOLFIRINOX (combination of 5FU, oxaliplatin and irinotecan) (171 pts)	11.1 (9.0-13.1)	Gemcitabine (171 pts)	6.8 (5.5-7.6)	0.57 (0.45-0.73) P < 0.001	Conroy <i>et al</i> ^[10] , 2011
Gemcitabine and nab-paclitaxel (431 pts)	8.5 (7.9-9.5)	Gemcitabine (430 pts)	6.7 (6.0-7.2)	0.72 (0.62-0.83) P < 0.001	Von Hoff <i>et al</i> ^[11] , 2013

OS: Overall survival.

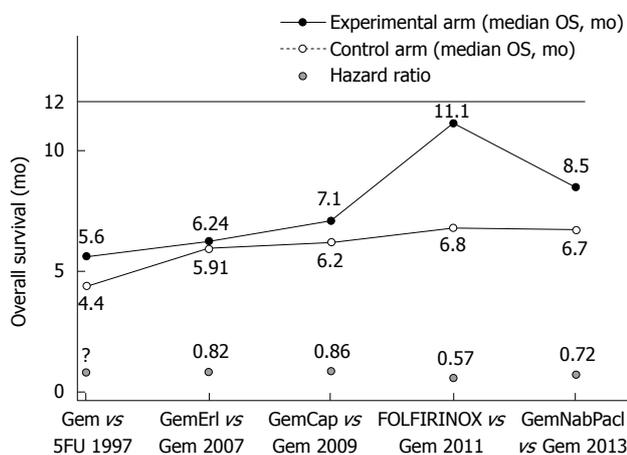


Figure 1 Multiple randomized phase III trials have been completed in the last decades; however, we have not been able to cross the barrier of 12 mo survival in advanced pancreatic cancer.

the disease, only 15%-20% of patients are diagnosed early enough to be considered for a potentially curative treatment. However, the relapse rate after surgery is high (80%-90%). Looking for a reduction in the relapse rate and an increase in the overall survival (OS), adjuvant chemotherapy is currently standard of care after resection of PDA. The most employed adjuvant chemotherapy schedules are gemcitabine or capecitabine^[5,6].

Unfortunately, most of the patients (up to 80%) are

diagnosed in advanced stages and palliative chemotherapy is the only option of treatment. The aim of this chemotherapy is to prolong OS and improve the quality of life. In 1997, gemcitabine was established as the drug of choice for the treatment of advanced PDA with OS of 5.6 mo compared to 4.4 mo in the arm with 5FU^[7]. Since then, multiple randomized studies with combination schedules have shown improvement in OS compared to single agent gemcitabine (Table 2)^[8-11]. However, as it is shown in Figure 1, the impact in the survival achieved in advanced PDA has never reached the year of median OS. This is far of being comparable to the results achieved in other malignancies such as advanced breast or colorectal cancer.

WHY THESE RESULTS? WHAT ARE THE CHALLENGES WHEN TREATING PDA?

Much effort has been employed in trying to improve the survival of our patients with PDA. The improvement in the big randomized studies with more than 1800 patients during the last decades seems to be not enough and the median OS is still less than one year after diagnosis^[12]. When we compare this data with other adenocarcinomas, for example breast or colorectal adenocarcinoma with median OS longer than 24 mo, we might wonder: are we doing the right research? What makes pancreatic cancer

Table 3 Core signalling pathways involved in pancreatic ductal adenocarcinoma

Involved pathways	PDA with pathway aberrations	Representative genes
Apoptosis	100%	<i>CASP10, VCP, CAD, HIP1</i>
DNA repair	83%	<i>ERCC4, ERCC6, EP300, RANBP2, TP53</i>
Regulation of G1/S phase	100%	<i>CDKN2A, FBXW7, CHD1, APC2</i>
Hedgehog pathway	100%	<i>TBX5, SOX3, LRP2, GLI1, GLI3, BOC, BMPR2, CREBBP</i>
Celular adhesion	79%	<i>CDH1, CDH10, CDH2, CDH7, FAT, PCDH15, PCDH17, PCDH18, PCDH9, PCDH16, PCDHB2, PCDHGA1, PCDHGA11, PCDHGC4</i>
Integrin signaling	67%	<i>ITGA4, ITGA9, ITGA11, LAMA1, LAMA4, LAMA5, FN1, ILK</i>
c-Jun N-terminal kinase signaling	96%	<i>MAP4K3, TNF, ATF2, NFATC3</i>
KRAS signaling	100%	<i>KRAS, MAP2K4, RASGRP3</i>
Regulation of invasion	92%	<i>ADAM11, ADAM12, ADAM19, ADAM5220, ADAMTS15, DPP6, MEP1A, PCSK6, APG4A, PRSS23</i>
GTP-ase dependent signaling (not κ -ras)	79%	<i>AGHGEF7, ARHGEF9, CDC42BPA, DEPDC2, PLCB3, PLCB4, RP1, PLXNB1, PRKCG</i>
TGF- β pathway	100%	<i>TGFBR2, BMPR2, SMAD4, SMAD3</i>
Wnt/Notch pathway	100%	<i>MYC, PPP2R3A, WNT9A, MAP2, TSC2, GATA6, TCF4</i>

Adapted from Jones *et al*^[19], 2008. PDA: Pancreatic ductal adenocarcinoma.

so hard to treat? Do we know enough about its molecular biology? Which is the next step?

Several reasons have been postulated for the difficulties in achieving better results in PDA^[13]: (1) delay in diagnosis due to lack of symptoms until advanced stages. Most of the patients are diagnosed with distant metastases or unresectable locally advanced disease. Moreover, due to its location in the retroperitoneum, the pancreas is difficult to access and sample with traditional endoscopic techniques. This can also raise difficulties for an early diagnosis; (2) PDA is associated with several comorbidities that could affect patients' overall health with a worse impact in the OS of those who develop the PDA (Table 1); (3) limited effect of local therapies. The relapse rate is far from being acceptable, even with adjuvant chemotherapy or a combination of adjuvant chemo-radiotherapy. One possible explanation is that "field effect" mutations may affect normal appearing cells present in the residual pancreatic tissue. This, added to the high ability of spreading, even in early stages, could explain the high chances of relapse after local radical treatment^[14]; and (4) PDA has been postulated to be primary (innate), rather than secondary (acquired), resistance to chemotherapy. Reasons for this could be both, related to the cancer cell itself and to the stroma surrounding the pancreatic cancer cells: (1) cancer cell characteristics. Different high penetrance genetic alterations have been described in PDA. One of the most frequent ones is activating mutations in κ -ras (present in > 90% of PDA), which is one of the most potent of all human oncogenes, and able to induce strong pro-growth, cell motility and invasion signals; and (2) a defining characteristic of PDA is the presence of a dense fibrotic proliferation surrounding the epithelial cells composed of various leukocytes, fibroblasts, endothelial cells and neuronal cells, as well as extracellular matrix components such as collagen and hyaluronan^[15-17]. Moreover, in contrast to many tumours that are dependent on neo-angiogenesis, PDA is poorly vascularised and therefore, poorly perfused, making the delivery of chemotherapy more difficult into the tumour

cells.

MOLECULAR BIOLOGY IN PANCREATIC CANCER: WHAT DO WE KNOW?

PDA is known to be a genetic disease, caused by inherited and acquired mutations in specific cancer-associated genes^[18]. Since the sequencing of the protein-coding exons from 20661 genes in 24 advanced ductal adenocarcinomas of the pancreas was published in 2008, a better understanding of the key pathways involved in the development and maintenance of PDA was provided^[19]. In 2012 the sequencing of 142 localized and resected PDAs was also published^[20].

The most important genes and pathways involved in PDA biology are summarized in Table 3.

According to our current knowledge, multiple combinations of all these genetic mutations are commonly found in PDA, and can be classified as follows^[21-24]: (1) mutational activation of oncogenes: predominantly K-ras; (2) inactivation of tumour suppressor genes such as *TP53*, *p16/CDKN2A*, and *SMAD4*; (3) inactivation of genome maintenance genes, such as *hMLH1* and *MSH2*, which control the repair of DNA damage. Most of these mutations are somatic aberrations. However, some germline aberrations were described (*BRCA2*, *PALB2*, *STK11*, *ATM*, *MLH1* and *MSH2*) to be involved in the development of hereditary pancreatic cancer (Tables 1 and 3)^[25].

During the last two decades, a lot of effort has been done in the definition of biological pathways involved, not only in the development/maintenance of PDA cancer cells, but also in the characterisation of the stroma surrounding the PDA cells^[15-17]. As we discussed above, the characteristics of this particular stroma are one of the explanations for the difficulties in the treatment of PDA^[26]. Some core pathways [*e.g.*, Hedgehog, Transforming growth factor (TGF)- β and Hepatocyte growth factor (HGF)-met] have shown to be involved in its development^[18,27]. Moreover, some studies are testing the effec-

Table 4 Biomarkers in pancreatic ductal adenocarcinoma

Biomarker	Prognostic biomarker	Predictive biomarker	Comments and references
MUC1	Yes		Predictive of early cancer-related death ^[37]
MSLN	Yes		Predictive of early cancer-related death ^[37]
6-gene signature	Yes		Expression of <i>FOSB</i> , <i>KLF6</i> , <i>NFKBIZ</i> , <i>ATP4A</i> , <i>GSG1</i> and <i>SIGLEC11</i> is related with metastatic spread ^[38]
VEGF	Yes		Worse survival in resected PDA ^[39]
p16	Yes		Higher expression was related to poorer prognosis ^[40]
TP53	Yes		Relation with tumour dedifferentiation and higher locoregional recurrence ^[40]
SMAD4	Yes		Higher Smad4/Dpc4 was related to bigger tumours, lymph node metastases and shorter survival ^[40] . Higher relapse rate (distant spread) ^[41] . Loss of expression correlated with resectability and better survival after surgery ^[42]
EGFR			No predictive/prognostic power ^[43,44]
K-ras	Yes		Better prognosis in Kras wild-type tumours ^[43,44]
RRM1	Yes	Yes	High expression of RRM1 showed significantly better overall survival ^[45-47] and worse response to treatment ^[47-49]
ERCC1	Yes		High ERCC1 expression showed significantly better overall survival ^[47,50,51] . No predictive power ^[49]
CTCs	Yes		More studies are awaited ^[52]
hENT1	Yes	Yes	High expression of hNENt1: worse prognosis, higher response to gemcitabine in the adjuvant setting; unclear impact in metastatic patients ^[50,53-57]
HuR	Yes	Yes	Low expression of HuR: worse prognosis ^[58] and better response to gemcitabine ^[59,60]
SPARC	Yes		Expression of SPARC in the peritumoural stroma is related with worse prognosis ^[61,62] . No predictive effect
CTGF			Preclinical data seem to suggest prognostic impact and potential predictive power for FB-3019 ^[63-66]

VEGF: Vascular endothelial growth factor; EGFR: Epidermal growth factor receptor; CTCs: Circulating tumour cells; SPARC: Secreted protein acidic and rich in cysteine; CTGF: Connective tissue growth factor.

tiveness of anti-stroma therapies in pancreatic cancer, such as Vismodegib (Hedgehog pathway inhibitor)^[28,29] and nab-paclitaxel [postulated to be a secreted protein acidic and rich in cysteine (SPARC) inhibitor]^[30].

IMPROVING OUR RESULTS THROUGH THE DEVELOPMENT OF BIOMARKERS

A biomarker has been defined as “a characteristic that is objectively measured and evaluated as an indicator of normal biological processes, pathogenic processes, or pharmacologic responses to a therapeutic intervention” by the Biomarkers Definitions Working Group^[31]. According to this Working group, three categories of biomarkers can be defined depending on the information that they provide: diagnostic, prognostic and predictive biomarkers.

As detailed above, during the last decades, our knowledge about the PDA molecular biology is increasing exponentially and lots of pathways have been implicated in this malignancy. However, apart from CA19.9 for the diagnosis of pancreatic adenocarcinoma, no other biomarkers are currently being employed in PDA for improving the clinical management of these patients^[24,32,33]. How can we apply all this new knowledge in the development of new studies looking for an improvement in OS? According to some experts, better early detection strategies with diagnostic biomarkers, treatment decisions based on prognostic biomarkers, and individualized treatment schedules based on predictive biomarkers could be the clue for the improvement in PDA research^[34,35].

A lot of work has already been done in the development of a compendium of potential PDA biomarkers worth to be included in future research^[36]. Nowadays, the three biomarker categories are being developed in PDA. The most important prognostic and predictors biomarkers for pancreatic cancer are summarized in Table 4.

Diagnostic biomarkers

The aim of the development of diagnostic biomarkers is to improve the rate of early diagnosis. CA19.9 is already employed as a diagnostic tool in combination with image techniques^[23,30]. The definition of genetic expression and proteomic patterns could improve the diagnosis of PDA, currently based on morphological pathology studies only.

Prognostic biomarkers

The potential of classifying the patients into good and bad prognostic groups could be especially useful after surgery. We could offer more aggressive chemotherapy schedule or closer follow-up to those patients with worse prognosis or more chances of relapse. Moreover, the capability of defining the relapse pattern (local *vs* distant spread) could also improve the chosen image technique or frequency for the surveillance. See more details below.

Predictive biomarkers

The definition of predictive biomarkers, both for already employed drugs and for new therapies, could enrich our prospective studies. We need to improve our ability for selecting those patients that, according to the tumour expression of predictive biomarkers in PDA, may have better response to the chosen treatment and individual-

ize the chemotherapy according to this information. See more details below.

CURRENT DEVELOPMENT OF BIOMARKERS IN PANCREATIC CANCER

The most important prognostic and predictors biomarkers for pancreatic cancer are summarized in Table 4.

PROGNOSTIC BIOMARKERS IN PDA

To look for effective biomarkers able to stratify PDA based on biologic behaviour, a survival tissue microarray of 137 resected PDAs was analysed^[37]. In a multivariate model, MUC1 (OR = 28.95, 3+ *vs* negative expression, $P = 0.004$) and MSLN (OR = 12.47, 3+ *vs* negative expression, $P = 0.01$) were highly predictive of early cancer-related death. In this study, MUC1 and MSLN were superior to pathologic features (tumour size, lymph node metastases, and nuclear grade) in predicting survival.

Stratford *et al*^[38] identified a six-gene signature (*FOSB*, *KLF6*, *NFKBIZ*, *ATP4A*, *GSG1* and *SIGLEC11*) associated with metastatic disease. The results from the training set of 34 patients were validated in an independent series of 67 patients. The six-gene signature was independently predictive of survival and superior to established clinical prognostic factors such as grade, tumour size and nodal status (HR = 4.1, 95%CI: 1.7-10.0). Patients defined to be “high-risk” had a 1-year survival rate of 55% compared to 91% in the “low-risk” group.

In 2011 a meta-analysis of immunohistochemical markers in resected pancreatic cancer was published^[39]. The aim of the study was to conduct a systematic review of the literature evaluating p53, p16, SMAD4, bcl-2, bax, vascular endothelial growth factor (VEGF) and epidermal growth factor receptor (EGFR) expression as prognostic factors in resected PDA. VEGF emerged as the most potentially informative prognostic marker (11 eligible studies, 767 patients, HR = 1.51, 95%CI: 1.18-1.92). Bcl-2, bax and p16 were also related to OS. Neither p53, SMAD4 or EGFR were found to have significant prognostic value.

The expression of hENT1, involved in the internalization of gemcitabine into the cancer cell, has been widely explored in PDA (See below, “predictive factors”). The prognostic value of the expression of hENT1 has been shown in several studies. Kim *et al*^[32] reported in 2011 a series of eighty-four resected PDAs. Total RNA was isolated from paraffin-embedded tumours and the multivariate analysis confirmed the association of low expression of hENT-1 ($P = 0.007$) with worse OS and progression free survival ($P = 0.016$).

The genes *p16*, *TP53* and *SMAD4/DPC4* were included in a study by Jones *et al*^[19] in 2008 as core pathways of PDA development and have been widely studied in PDA for its potential prognostic prediction.

A retrospective study published in 2013 aimed to clarify the implications of 3 major genes (*CDKN2A/p16*,

p53, and *SMAD4/DPC4*) with clinico-pathological findings, including survival and patterns of disease progression, in 106 patients with resected PDA^[40]. The expression of protein products of these genes was determined immunohistochemically. Genetic aberrations of these 3 genes were associated with malignant behaviour of PDA: a significant correlation was found between SMAD4/DPC4 immunolabeling and tumour size ($P = 0.006$), lymphatic invasion ($P = 0.033$) and lymph node metastasis ($P = 0.006$); loss of p16 immunolabeling ($P = 0.029$) and loss of SMAD4/DPC4 immunolabeling ($P < 0.001$) were significantly associated with shorter OS; and abnormal immunolabeling of p53 was significantly associated with tumour dedifferentiation ($P = 0.022$) and the presence of locoregional recurrence ($P = 0.020$).

Moreover, the expression of *SMAD4* has been analysed to define higher relapse rate and therefore worse prognosis, in several studies^[40-42,67]. A study including patients with resected PDA analysed the expression of cell-cycle and cell-signaling involved key proteins using immunohistochemistry in a subgroup of 129 patients^[42]. While aberrant expression of *p21*(WAF1/CIP1), cyclinD1, *p53* or *p16*(INK4A) was not associated with a difference in survival; loss of *SMAD4/DPC4* expression correlated with resectability ($P < 0.0001$) and was associated with improvement in survival after resection ($P < 0.0001$). In contrast, resection did not improve survival in patients whose tumour expressed *SMAD4/DPC4* ($P = 0.5$). The authors concluded that preoperative assessment of *SMAD4/DPC4* expression could be useful in the selection of patients that may benefit from surgical resection.

In 2011, Crane *et al*^[41] reported the results of a phase II clinical trial to assess the efficacy and safety of cetuximab, gemcitabine and oxaliplatin followed by cetuximab, capecitabine, and radiation therapy in locally advanced pancreatic cancer. Diagnostic cytology specimens were analysed for SMAD4/DPC4 protein expression (by immunohistochemistry). In this study, SMAD4/DPC4 protein expression correlated with local rather than distant disease progression ($P = 0.016$).

Finally, Iacobuzio-Donahue *et al*^[67] performed rapid autopsies on 76 patients with documented pancreatic cancer. The histological features, the status of the *KRAS2*, *p53* and *SMAD4/DPC4* genes were correlated to the stage at initial diagnosis and patterns of failure (locally destructive *vs* metastatic disease). *SMAD4/DPC4* genetic status was highly correlated with the presence of widespread metastasis but not with locally destructive tumours ($P = 0.007$).

SPARC is expressed in the cell matrix and it is involved in cell matrix interactions, wound repair, cell migration and cancer growth regulation. The high expression of *SPARC* in the peritumoural stroma was defined as a worse prognostic factor both in localized and locally advanced patients^[61,62]. The expression patterns of *SPARC* were characterized by immunohistochemistry in 299 resected pancreatic adenocarcinomas to evaluate the

Table 5 “Classic” predictive biomarkers for “classic” chemotherapies with potential interest in pancreatic cancer

Predictive biomarker	Drug	Theoretical impact ¹	Studies performed in pancreatic cancer (predictive outcome)?	Impact confirmed in pancreatic cancer?	Notes	Ref.
Thymidylate synthase	5FU	When negative, better response to 5FU	Yes	No	Predictive value in PDA not validated	[55,72-74]
DPD	5FU	When mutation DPD, more 5FU related toxicity	Yes	No	Survival benefit with S1 and DPD mutation	[73]
Topoisomerase I	Irinotecan	When positive, better response to Irinotecan	No	No	No data in pancreatic cancer	-
RRM1	Gemcitabine	When positive, better response to gemcitabine	Yes	Yes	Low expression correlates with better response	[47-49]
ERCC1	Oxaliplatin	When negative, better response to Oxaliplatin	Yes	No	No predictive effect	[49,51]
XRCC1	Oxaliplatin	When negative, better response to Oxaliplatin	No	No	No data in pancreatic cancer	-
EGFR/kras	Erlotinib	Erlotinib effective when EGFR mutation/kras wild type present	Yes	No	No predictive effect	[43,44]
PALB2	Mitomycin C	Mitomycin C effective when PALB2 mutation present	No	Yes	Case report	[75]
BRCA2	PARP inhibitors	PARP inhibitors effective when BRCA2 mutation present	Yes	Yes	Phase I trial	[76]

¹This impact is suggested in other malignancies.

prognostic significance of tumoural and peritumoural *SPARC* expression^[61]. In the multivariate analysis, *SPARC* expression in the surrounding stroma was a biomarker of worse prognosis (HR = 1.89, 95%CI: 1.31-2.74), while the expression of *SPARC* in pancreatic cancer cells remained unrelated to prognosis (HR = 1.02, 95%CI: 0.73-1.42). These data have been validated in further studies^[62]. However, in animal models this prognostic impact of *SPARC* was not verified: the prognosis was worse in those *SPARC* knock-out mice^[68].

Other prognostic biomarkers are: *Kras*^[43,44] (better prognosis in *Kras* wild type tumours), *HuR*^[59,60] (higher expression of *HuR* related to worse prognosis; See below “Predictive biomarkers”), *RRM1*^[45-47] and *ERCC1*^[47,50] (high *RRM1* and high *ERCC1* showed significantly better OS).

The analysis of circulating tumour cells is also being developed in PDA patients. However, further studies are awaited for a better understanding of its impact^[52].

Collisson *et al*^[69] reported in 2011 a study identifying three PDA molecular subtypes (classical, quasimesenchymal and exocrine-like) both in human tumours and cell lines, with different profiles of survival and response to treatment. The subtypes were defined according to the gene expression profile. These data need to be further validated but could be useful in the improvement of the individualize management of PDA^[70].

PREDICTIVE BIOMARKERS IN PDA

Use of “classic” biomarkers to predict response to “classic” chemotherapies in pancreatic cancer

Based on data from other subtypes of adenocarcinoma such as colorectal tumour or breast cancer, some studies tried to employ “classic” predictive biomarkers to improve the results achieved with standard chemotherapy in advanced PDA. Although most of these biomarkers have

not been prospectively validated in pancreatic adenocarcinoma (Table 5), the rationale for this design is the individualization of first line chemotherapy according to the molecular expression profile of each tumour.

This idea has been executed in some studies. Von Hoff *et al*^[71] published in 2012 the results of a clinical trial, where patients with PDA were treated according to molecular profiles of their tumour. The molecular analysis included immunohistochemistry, fluorescent in situ hybridization assays and immediately frozen tissue for oligonucleotide microarray gene expression assays. From the 86 patients included, there was a molecular target detected in 84 (98%) and 66 were treated according to the molecular profiling results. This was a pilot study, and the authors confirmed the feasibility of this rationale. However, prospective studies are ongoing and data are awaited for its wide use (NCT01726582, NCT01394120).

Gemcitabine response predictive biomarkers: *RRM1*, *hENT1* and *HuR*

Apart from *RRM1*^[47-49], whose lack of expression is predictive of response to gemcitabine (Tables 4 and 5), other biomarkers were suggested as predictors of response to gemcitabine in PDA: *hENT1*^[50,53-55] and *HuR*^[59,60].

Human equilibrative nucleoside transporter-1 (*hENT1*) was found to be the major gemcitabine transporter into the cell. Therefore, those cells with low expression of *hENT1* will not transport the gemcitabine into the cancer cells, avoiding its activity (inhibition of the cell growth). In contrast, increased *hENT1* abundance facilitates efficient cellular entry of gemcitabine and confers increased cytotoxicity. Nakano *et al*^[77] reported in 2007 a preclinical study with pancreatic cancer cell lines where expression of *hENT1* changed in the development of gemcitabine resistance.

However, interpreting these results in human samples is challenging. In patients receiving adjuvant treatment, the expression of hENT1 showed to be predictive biomarker for response to gemcitabine. However, this was not validated in the metastatic setting.

The multicentre ESPAC-3 trial randomized patients to adjuvant gemcitabine or 5FU after pancreatic adenocarcinoma resection^[78]. According to the safety profile, gemcitabine was chosen as the preferred agent when compared with monthly bolus (Mayo Clinic) 5-FU/LV for the adjuvant setting. The samples collected from the adjuvant ESPAC1/3 randomized trials were employed in a translational project to define the predictive value of hENT1^[54]. One-hundred and seventy-five gemcitabine treated and 176 5-FU treated patients were included in the analysis. In the gemcitabine group a significantly lower survival ($P = 0.002$) was noted with low hENT1 [median survival 17.1 (95%CI: 14.3-23.8) mo *vs* 26.2 (95%CI: 21.2-31.4) mo]. Multivariate analysis confirmed hENT1 expression as a predictive biomarker of response to gemcitabine in the adjuvant setting.

However, the findings in metastatic patients are different. During the 2013 ASCO congress, data of a new gemcitabine-like drug (CO-101) were presented^[57]. CO-101 (also known as CP-4126), a lipid-drug conjugate of gemcitabine, was rationally designed to enter cells independently of hENT1. The authors presented the results of a randomized trial comparing CO-101 and gemcitabine in the metastatic setting. The aim of the study was to determine whether CO-101 improved survival compared to gemcitabine in patients with low hENT1 tumours and to test prospectively the hypothesis that hENT1 was a predictive marker of response to gemcitabine. Unfortunately, CO-101 was not superior to gemcitabine in patients with low tumour hENT1 expression and, moreover, hENT1 expression did not predict gemcitabine treatment outcome in this study.

From these data, we conclude that while hENT1 seems to be a predictor of response in the adjuvant setting, this was not reproducible in metastatic patients. The molecular biology of the metastatic PDA may differ from the localized tumours, explaining the differences in the results.

The ubiquitous RNA-binding protein (RBP) HuR is involved in the control of gene expression, mRNA stability and translation and cellular response to internal and external signals^[79]. Through its post-transcriptional effect by targeting mRNAs, HuR can alter the cellular response to proliferative, stress, apoptotic, differentiation, senescence, inflammatory and immune signals. The high expression of HuR has already been defined as a prognostic factor in PDA and some studies postulated HuR as a predictive biomarker for response to gemcitabine in cancer cell lines^[59,60].

These results were confirmed in a series of 29 localized PDA patients in whom correlation between HuR expression levels and OS was evaluated^[58]. The results indicated an increase in risk of death in patients with low

HuR levels compared to high HuR levels among patients receiving gemcitabine. Authors concluded that HuR was regulating the key metabolic enzyme for gemcitabine activation (deoxycytidine kinase) and could be a marker for therapeutic efficacy of gemcitabine based regimens: better response in patients with high HuR expression.

SPARC

As we detailed above, SPARC has prognostic impact in PDA^[61,62]. Nab-paclitaxel is a 130-nm albumin-bound formulation of paclitaxel particles. Data from the phase I / II trial with nab-paclitaxel postulated SPARC as a predictive factor of anti-stromal therapies^[30]. SPARC status was evaluated in 36 patients and a significant increase in OS was observed in high-SPARC expression subgroup compared with patients in the low-SPARC subgroup (median OS, 17.8 mo *vs* 8.1 mo, respectively; $P = 0.0431$). Moreover, some studies in animal models postulated that the addition of nab-paclitaxel could increase the intratumoural gemcitabine delivery due to anti-stromal effect of nab-paclitaxel^[30,80]. However, the predictive impact of the expression of SPARC has not been clarified in the prospective studies with combination chemotherapy with gemcitabine and nab-paclitaxel^[11,30].

Connective tissue growth factor/CCN2

Also focused in the stroma and the importance in PDA, connective tissue growth factor (CTGF) expression was analysed in pancreatic cancer. CTGF is a cysteine-rich matricellular secreted protein, which regulates diverse cell functions including adhesion, migration, proliferation, differentiation, survival, senescence and apoptosis^[81,82].

Due to the hypoxic conditions surrounding the PDA, Eguchi *et al.*^[82] analyzed the tumour-stroma interaction signalling in cell lines of pancreatic cancer in hypoxia and normoxia using RNA interference techniques. The results showed that cell invasion was more enhanced under hypoxia than under normoxia ($P < 0.05$) and that CTGF was one of the overexpressed molecules in hypoxic conditions. Moreover, cell invasiveness was reduced by CTGF knockdown in hypoxic cancer cells ($P < 0.05$). The authors concluded that hypoxia induced CTGF expression could be a prognostic factor related to higher aggressiveness in PDA. This results match with those from other studies^[63-65].

Therefore, the data available shows that CTGF is overexpressed in PDA and facilitates local desmoplasia, tumour survival and metastasis. FG-3019 is a human monoclonal antibody to CTGF, able to control the tumour growth in cancer cell lines^[83] and tumour xenografts^[65], without damage to the healthy tissue. Neesse *et al.*^[84] reported data from animal model research to clarify the antitumour effect of FG-3019. The authors concluded that FG-3019 may have antitumour effect itself, more than improving the delivery of gemcitabine into the tumour. First data in humans were presented in ASCO-GI 2013 by Picozzi *et al.*^[66], showing that the combination with gemcitabine, erlotinib and FG-3019 was safe in ad-

vanced pancreatic cancer patients. The authors showed that baseline CTGF plasma level was related to worse survival. Further clinical data for the prognostic and predictor relevance of CTGF in humans are awaited.

FUTURE, HOW TO IMPLEMENT THE ACTUAL DATA?

There is no doubt that the knowledge in molecular biology will continue to improve in the following years. New generation techniques are being employed in PDA research and will give much more data. However, it is crucial to incorporate this knowledge in a rational way, and this could be challenging. Moreover, the huge economic cost of this research needs to be analysed. Some panels of experts have defined the most suitable way for biomarker development and its addition to the clinical research in pancreatic cancer^[12,85,86].

In conclusion, a lot of work needs to be done in the improvement of our understanding in pancreatic adenocarcinoma. Treatment individualization seems to be one of the keys, implementing the clinicopathological information with molecular characteristics. In order to achieve this, the development of new biomarkers and a better definition of the current ones are radically important. Most of the detailed biomarkers in this review are available just for research purposes; only Ca19.9 (with diagnostic and follow-up aim) is employed in the clinical practice. The results of the ongoing clinical trials with new biomarker research and the selection of the therapies according to these molecular characteristics are awaited.

REFERENCES

- Jemal A**, Bray F, Center MM, Ferlay J, Ward E, Forman D. Global cancer statistics. *CA Cancer J Clin* 2011; **61**: 69-90 [PMID: 21296855 DOI: 10.3322/caac.20107]
- Siegel R**, Naishadham D, Jemal A. Cancer statistics, 2013. *CA Cancer J Clin* 2013; **63**: 11-30 [PMID: 23335087 DOI: 10.3322/caac.21166]
- López Serrano A**. [Risk factors and early diagnosis of pancreatic cancer]. *Gastroenterol Hepatol* 2010; **33**: 382-390 [PMID: 20005016 DOI: 10.1016/j.gastrohep.2009.10.004]
- Bartsch DK**, Gress TM, Langer P. Familial pancreatic cancer-current knowledge. *Nat Rev Gastroenterol Hepatol* 2012; **9**: 445-453 [PMID: 22664588 DOI: 10.1038/nrgastro.2012.111]
- Oettle H**, Post S, Neuhaus P, Gellert K, Langrehr J, Ridwelski K, Schramm H, Fahlke J, Zuelke C, Burkart C, Gütberlet K, Kettner E, Schmalenberg H, Weigang-Koehler K, Bechstein WO, Niedergethmann M, Schmidt-Wolf I, Roll L, Doerken B, Riess H. Adjuvant chemotherapy with gemcitabine vs observation in patients undergoing curative-intent resection of pancreatic cancer: a randomized controlled trial. *JAMA* 2007; **297**: 267-277 [PMID: 17227978 DOI: 10.1001/jama.297.3.267]
- Neoptolemos JP**, Stocken DD, Friess H, Bassi C, Dunn JA, Hickey H, Beger H, Fernandez-Cruz L, Dervenis C, Lacaïne F, Falconi M, Pederzoli P, Pap A, Spooner D, Kerr DJ, Büchler MW. A randomized trial of chemoradiotherapy and chemotherapy after resection of pancreatic cancer. *N Engl J Med* 2004; **350**: 1200-1210 [PMID: 15028824 DOI: 10.1056/NEJMoa032295]
- Burris HA**, Moore MJ, Andersen J, Green MR, Rothenberg ML, Modiano MR, Cripps MC, Portenoy RK, Storniolo AM, Tarassoff P, Nelson R, Dorr FA, Stephens CD, Von Hoff DD. Improvements in survival and clinical benefit with gemcitabine as first-line therapy for patients with advanced pancreas cancer: a randomized trial. *J Clin Oncol* 1997; **15**: 2403-2413 [PMID: 9196156]
- Moore MJ**, Goldstein D, Hamm J, Figer A, Hecht JR, Gallinger S, Au HJ, Murawa P, Walde D, Wolff RA, Campos D, Lim R, Ding K, Clark G, Voskoglou-Nomikos T, Ptasynski M, Parulekar W. Erlotinib plus gemcitabine compared with gemcitabine alone in patients with advanced pancreatic cancer: a phase III trial of the National Cancer Institute of Canada Clinical Trials Group. *J Clin Oncol* 2007; **25**: 1960-1966 [PMID: 17452677 DOI: 10.1200/JCO.2006.07.9525]
- Cunningham D**, Chau I, Stocken DD, Valle JW, Smith D, Stewart W, Harper PG, Dunn J, Tudur-Smith C, West J, Falk S, Crellin A, Adab F, Thompson J, Leonard P, Ostrowski J, Eatock M, Scheithauer W, Herrmann R, Neoptolemos JP. Phase III randomized comparison of gemcitabine versus gemcitabine plus capecitabine in patients with advanced pancreatic cancer. *J Clin Oncol* 2009; **27**: 5513-5518 [PMID: 19858379 DOI: 10.1200/JCO.2009.24.2446]
- Conroy T**, Desseigne F, Ychou M, Bouché O, Guimbaud R, Bécauarn Y, Adenis A, Raoul JL, Gourgou-Bourgade S, de la Fouchardière C, Bennouna J, Bacht JB, Khemissa-Akouz F, Péré-Vergé D, Delbaldo C, Assenat E, Chauffert B, Michel P, Montoto-Grillot C, Ducreux M. FOLFIRINOX versus gemcitabine for metastatic pancreatic cancer. *N Engl J Med* 2011; **364**: 1817-1825 [PMID: 21561347 DOI: 10.1056/NEJMoa1011923]
- Von Hoff DD**, Ervin T, Arena FP, Chiorean EG, Infante J, Moore M, Seay T, Tjulandin SA, Ma WW, Saleh MN, Harris M, Reni M, Dowden S, Laheru D, Bahary N, Ramanathan RK, Tabernero J, Hidalgo M, Goldstein D, Van Cutsem E, Wei X, Iglesias J, Renschler MF. Increased survival in pancreatic cancer with nab-paclitaxel plus gemcitabine. *N Engl J Med* 2013; **369**: 1691-1703 [PMID: 24131140 DOI: 10.1056/NEJMoa1304369]
- Tabernero J**, Macarulla T. Changing the paradigm in conducting randomized clinical studies in advanced pancreatic cancer: an opportunity for better clinical development. *J Clin Oncol* 2009; **27**: 5487-5491 [PMID: 19858387 DOI: 10.1200/JCO.2009.23.3098]
- Oberstein PE**, Olive KP. Pancreatic cancer: why is it so hard to treat? *Therap Adv Gastroenterol* 2013; **6**: 321-337 [PMID: 23814611 DOI: 10.1177/1756283X13478680]
- Rhim AD**, Mirek ET, Aiello NM, Maitra A, Bailey JM, McAllister F, Reichert M, Beatty GL, Rustgi AK, Vonderheide RH, Leach SD, Stanger BZ. EMT and dissemination precede pancreatic tumor formation. *Cell* 2012; **148**: 349-361 [PMID: 22265420 DOI: 10.1016/j.cell.2011.11.025]
- Heinemann V**, Reni M, Ychou M, Richel DJ, Macarulla T, Ducreux M. Tumour-stroma interactions in pancreatic ductal adenocarcinoma: rationale and current evidence for new therapeutic strategies. *Cancer Treat Rev* 2014; **40**: 118-128 [PMID: 23849556]
- Waghray M**, Yalamanchili M, di Magliano MP, Simeone DM. Deciphering the role of stroma in pancreatic cancer. *Curr Opin Gastroenterol* 2013; **29**: 537-543 [PMID: 23892539 DOI: 10.1097/MOG.0b013e328363affe]
- Erkan M**. Understanding the stroma of pancreatic cancer: co-evolution of the microenvironment with epithelial carcinogenesis. *J Pathol* 2013; **231**: 4-7 [PMID: 23716361 DOI: 10.1002/path.4213]
- Hidalgo M**. Pancreatic cancer. *N Engl J Med* 2010; **362**: 1605-1617 [PMID: 20427809 DOI: 10.1056/NEJMra0901557]
- Jones S**, Zhang X, Parsons DW, Lin JC, Leary RJ, Angenendt P, Mankoo P, Carter H, Kamiyama H, Jimeno A, Hong SM, Fu B, Lin MT, Calhoun ES, Kamiyama M, Walter K, Nikolskaya T, Nikolsky Y, Hartigan J, Smith DR, Hidalgo M, Leach

- SD, Klein AP, Jaffee EM, Goggins M, Maitra A, Iacobuzio-Donahue C, Eshleman JR, Kern SE, Hruban RH, Karchin R, Papadopoulos N, Parmigiani G, Vogelstein B, Velculescu VE, Kinzler KW. Core signaling pathways in human pancreatic cancers revealed by global genomic analyses. *Science* 2008; **321**: 1801-1806 [PMID: 18772397 DOI: 10.1126/science.1164368]
- 20 **Biankin AV**, Waddell N, Kassahn KS, Gingras MC, Muthuswamy LB, Johns AL, Miller DK, Wilson PJ, Patch AM, Wu J, Chang DK, Cowley MJ, Gardiner BB, Song S, Harliwong I, Idrisoglu S, Nourse C, Nourbakhsh E, Manning S, Wani S, Gongora M, Pajic M, Scarlett CJ, Gill AJ, Pinho AV, Rooman I, Anderson M, Holmes O, Leonard C, Taylor D, Wood S, Xu Q, Nones K, Fink JL, Christ A, Bruxner T, Cloonan N, Kolle G, Newell F, Pinese M, Mead RS, Humphris JL, Kaplan W, Jones MD, Colvin EK, Nagrial AM, Humphrey ES, Chou A, Chin VT, Chantrill LA, Mawson A, Samra JS, Kench JG, Lovell JA, Daly RJ, Merrett ND, Toon C, Epari K, Nguyen NQ, Barbour A, Zeps N, Kakkar N, Zhao F, Wu YQ, Wang M, Muzny DM, Fisher WE, Brunnicardi FC, Hodges SE, Reid JG, Drummond J, Chang K, Han Y, Lewis LR, Dinh H, Buhay CJ, Beck T, Timms L, Sam M, Begley K, Brown A, Pai D, Panchal A, Buchner N, De Borja R, Denroche RE, Yung CK, Serra S, Onetto N, Mukhopadhyay D, Tsao MS, Shaw PA, Petersen GM, Gallinger S, Hruban RH, Maitra A, Iacobuzio-Donahue CA, Schlick RD, Wolfgang CL, Morgan RA, Lawlor RT, Capelli P, Corbo V, Scardoni M, Tortora G, Tempero MA, Mann KM, Jenkins NA, Perez-Mancera PA, Adams DJ, Largaespada DA, Wessels LF, Rust AG, Stein LD, Tuveson DA, Copeland NG, Musgrove EA, Scarpa A, Eshleman JR, Hudson TJ, Sutherland RL, Wheeler DA, Pearson JV, McPherson JD, Gibbs RA, Grimmond SM. Pancreatic cancer genomes reveal aberrations in axon guidance pathway genes. *Nature* 2012; **491**: 399-405 [PMID: 23103869 DOI: 10.1038/nature11547]
- 21 **Hilgers W**, Kern SE. Molecular genetic basis of pancreatic adenocarcinoma. *Genes Chromosomes Cancer* 1999; **26**: 1-12 [PMID: 10440999 DOI: 10.1002/(SICI)1098-2264(199909)26:1<1::AID-GCC1>3.0.CO;2-X]
- 22 **Sakorafas GH**, Tsiotos GG. Molecular biology of pancreatic cancer: potential clinical implications. *BioDrugs* 2001; **15**: 439-452 [PMID: 11520255 DOI: 10.2165/00063030-200115070-00003]
- 23 **Schmid RM**. Genetic basis of pancreatic cancer. *Best Pract Res Clin Gastroenterol* 2002; **16**: 421-433 [PMID: 12079267 DOI: 10.1053/bega.2002.0316]
- 24 **Baumgart M**, Heinmüller E, Horstmann O, Becker H, Ghadimi BM. The genetic basis of sporadic pancreatic cancer. *Cell Oncol* 2005; **27**: 3-13 [PMID: 15750203]
- 25 **Cowgill SM**, Muscarella P. The genetics of pancreatic cancer. *Am J Surg* 2003; **186**: 279-286 [PMID: 12946833 DOI: 10.1016/S0002-9610(03)00226-5]
- 26 **Whatcott CJ**, Posner RG, Von Hoff DD, Han H. Desmoplasia and chemoresistance in pancreatic cancer. In: Grippo PJ, Munshi HG, editors. *Pancreatic Cancer and Tumor Microenvironment*. Trivandrum (India): Transworld Research Network, 2012: Chapter 8 [PMID: 22876390]
- 27 **Hidalgo M**. New insights into pancreatic cancer biology. *Ann Oncol* 2012; **23** Suppl 10: x135-x138 [PMID: 22987949 DOI: 10.1093/annonc/mds313]
- 28 **Hao K**, Tian XD, Qin CF, Xie XH, Yang YM. Hedgehog signaling pathway regulates human pancreatic cancer cell proliferation and metastasis. *Oncol Rep* 2013; **29**: 1124-1132 [PMID: 23292285]
- 29 **Catenacci DVT**, Bahary N, Nattam SR, Marsh RW, Wallace JA, Rajdev L, Cohen DJ, Sleckman SG, Lenz HJ, Stiff PJ, Thomas SP, Xu P, Henderson L, Horiba MN, Vannier M, Karrison T, Stadler WM, Kindler HL. A phase IB/randomized phase II study of gemcitabine (G) plus placebo (P) or vismodegib (V), a hedgehog (Hh) pathway inhibitor, in patients (pts) with metastatic pancreatic cancer (PC): Interim analysis of a University of Chicago phase II consortium study. ASCO 2013, Abstract No: 4012. Available from: URL: <http://meetinglibrary.asco.org/content/117069-132>
- 30 **Von Hoff DD**, Ramanathan RK, Borad MJ, Laheru DA, Smith LS, Wood TE, Korn RL, Desai N, Trieu V, Iglesias JL, Zhang H, Soon-Shiong P, Shi T, Rajeshkumar NV, Maitra A, Hidalgo M. Gemcitabine plus nab-paclitaxel is an active regimen in patients with advanced pancreatic cancer: a phase I/II trial. *J Clin Oncol* 2011; **29**: 4548-4554 [PMID: 21969517 DOI: 10.1200/JCO.2011.36.5742]
- 31 **Biomarkers Definitions Working Group**. Biomarkers and surrogate endpoints: preferred definitions and conceptual framework. *Clin Pharmacol Ther* 2001; **69**: 89-95 [PMID: 11240971 DOI: 10.1067/mcp.2001.113989]
- 32 **Kim BJ**, Kim YH, Sinn DH, Kang KJ, Kim JY, Chang DK, Son HJ, Rhee PL, Kim JJ, Rhee JC. Clinical usefulness of glycosylated hemoglobin as a predictor of adenomatous polyps in the colorectum of middle-aged males. *Cancer Causes Control* 2010; **21**: 939-944 [PMID: 20373014 DOI: 10.1007/s10552-010-9543-4]
- 33 **Fong ZV**, Winter JM. Biomarkers in pancreatic cancer: diagnostic, prognostic, and predictive. *Cancer J* 2012; **18**: 530-538 [PMID: 23187839 DOI: 10.1097/PPO.0b013e31827654ea]
- 34 **Winter JM**, Yeo CJ, Brody JR. Diagnostic, prognostic, and predictive biomarkers in pancreatic cancer. *J Surg Oncol* 2013; **107**: 15-22 [PMID: 22729569 DOI: 10.1002/jso.23192]
- 35 **Costello E**, Greenhalf W, Neoptolemos JP. New biomarkers and targets in pancreatic cancer and their application to treatment. *Nat Rev Gastroenterol Hepatol* 2012; **9**: 435-444 [PMID: 22733351 DOI: 10.1038/nrgastro.2012.119]
- 36 **Harsha HC**, Kandasamy K, Ranganathan P, Rani S, Ramabadran S, Gollapudi S, Balakrishnan L, Dwivedi SB, Telikicherla D, Selvan LD, Goel R, Mathivanan S, Marimuthu A, Kashyap M, Vizza RF, Mayer RJ, Decaprio JA, Srivastava S, Hanash SM, Hruban RH, Pandey A. A compendium of potential biomarkers of pancreatic cancer. *PLoS Med* 2009; **6**: e1000046 [PMID: 19360088 DOI: 10.1371/journal.pmed.1000046]
- 37 **Winter JM**, Tang LH, Klimstra DS, Brennan MF, Brody JR, Rocha FG, Jia X, Qin LX, D'Angelica MI, DeMatteo RP, Fong Y, Jarnagin WR, O'Reilly EM, Allen PJ. A novel survival-based tissue microarray of pancreatic cancer validates MUC1 and mesothelin as biomarkers. *PLoS One* 2012; **7**: e40157 [PMID: 22792233 DOI: 10.1371/journal.pone.0040157]
- 38 **Stratford JK**, Bentrem DJ, Anderson JM, Fan C, Volmar KA, Marron JS, Routh ED, Caskey LS, Samuel JC, Der CJ, Thorne LB, Calvo BF, Kim HJ, Talamonti MS, Iacobuzio-Donahue CA, Hollingsworth MA, Perou CM, Yeh JJ. A six-gene signature predicts survival of patients with localized pancreatic ductal adenocarcinoma. *PLoS Med* 2010; **7**: e1000307 [PMID: 20644708 DOI: 10.1371/journal.pmed.1000307]
- 39 **Smith RA**, Tang J, Tudur-Smith C, Neoptolemos JP, Ghaneh P. Meta-analysis of immunohistochemical prognostic markers in resected pancreatic cancer. *Br J Cancer* 2011; **104**: 1440-1451 [PMID: 21448172 DOI: 10.1038/bjc.2011.110]
- 40 **Oshima M**, Okano K, Muraki S, Haba R, Maeba T, Suzuki Y, Yachida S. Immunohistochemically detected expression of 3 major genes (CDKN2A/p16, TP53, and SMAD4/DPC4) strongly predicts survival in patients with resectable pancreatic cancer. *Ann Surg* 2013; **258**: 336-346 [PMID: 23470568 DOI: 10.1097/SLA.0b013e3182827a65]
- 41 **Crane CH**, Varadhachary GR, Yordy JS, Staerke GA, Javle MM, Safran H, Haque W, Hobbs BD, Krishnan S, Fleming JB, Das P, Lee JE, Abbruzzese JL, Wolff RA. Phase II trial of cetuximab, gemcitabine, and oxaliplatin followed by chemoradiation with cetuximab for locally advanced (T4) pancreatic adenocarcinoma: correlation of Smad4(Dpc4) immunostaining with pattern of disease progression. *J Clin Oncol* 2011; **29**: 3037-3043 [PMID: 21709185 DOI: 10.1200/JCO.2010.33.8038]
- 42 **Biankin AV**, Morey AL, Lee CS, Kench JG, Biankin SA,

- Hook HC, Head DR, Hugh TB, Sutherland RL, Henshall SM. DPC4/Smad4 expression and outcome in pancreatic ductal adenocarcinoma. *J Clin Oncol* 2002; **20**: 4531-4542 [PMID: 12454109 DOI: 10.1200/JCO.2002.12.063]
- 43 **Boeck S**, Jung A, Laubender RP, Neumann J, Egg R, Goritschan C, Ormanns S, Haas M, Modest DP, Kirchner T, Heinemann V. KRAS mutation status is not predictive for objective response to anti-EGFR treatment with erlotinib in patients with advanced pancreatic cancer. *J Gastroenterol* 2013; **48**: 544-548 [PMID: 23435671 DOI: 10.1007/s00535-013-0767-4]
- 44 **Boeck S**, Jung A, Laubender RP, Neumann J, Egg R, Goritschan C, Vehling-Kaiser U, Winkelmann C, Fischer von Weikersthal L, Clemens MR, Gauler TC, Märten A, Klein S, Kojouharoff G, Barner M, Geissler M, Greten TF, Mansmann U, Kirchner T, Heinemann V. EGFR pathway biomarkers in erlotinib-treated patients with advanced pancreatic cancer: translational results from the randomised, crossover phase 3 trial AIO-PK0104. *Br J Cancer* 2013; **108**: 469-476 [PMID: 23169292 DOI: 10.1038/bjc.2012.495]
- 45 **Xie H**, Jiang W, Jiang J, Wang Y, Kim R, Liu X, Liu X. Predictive and prognostic roles of ribonucleotide reductase M1 in resectable pancreatic adenocarcinoma. *Cancer* 2013; **119**: 173-181 [PMID: 22736490 DOI: 10.1002/cncr.27715]
- 46 **Tanaka M**, Javle M, Dong X, Eng C, Abbruzzese JL, Li D. Gemcitabine metabolic and transporter gene polymorphisms are associated with drug toxicity and efficacy in patients with locally advanced pancreatic cancer. *Cancer* 2010; **116**: 5325-5335 [PMID: 20665488 DOI: 10.1002/cncr.25282]
- 47 **Akita H**, Zheng Z, Takeda Y, Kim C, Kittaka N, Kobayashi S, Marubashi S, Takemasa I, Nagano H, Dono K, Nakamori S, Monden M, Mori M, Doki Y, Bepler G. Significance of RRM1 and ERCC1 expression in resectable pancreatic adenocarcinoma. *Oncogene* 2009; **28**: 2903-2909 [PMID: 19543324 DOI: 10.1038/onc.2009.158]
- 48 **Jordheim LP**, Seve P, Trédan O, Dumontet C. The ribonucleotide reductase large subunit (RRM1) as a predictive factor in patients with cancer. *Lancet Oncol* 2011; **12**: 693-702 [PMID: 21163702 DOI: 10.1016/S1470-2045(10)70244-8]
- 49 **Valsecchi ME**, Holdbrook T, Leiby BE, Pequignot E, Littman SJ, Yeo CJ, Brody JR, Witkiewicz AK. Is there a role for the quantification of RRM1 and ERCC1 expression in pancreatic ductal adenocarcinoma? *BMC Cancer* 2012; **12**: 104 [PMID: 22436573 DOI: 10.1186/1471-2407-12-104]
- 50 **Pérez-Torras S**, García-Manteiga J, Mercadé E, Casado FJ, Carbó N, Pastor-Anglada M, Mazo A. Adenoviral-mediated overexpression of human equilibrative nucleoside transporter 1 (hENT1) enhances gemcitabine response in human pancreatic cancer. *Biochem Pharmacol* 2008; **76**: 322-329 [PMID: 18589402 DOI: 10.1016/j.bcp.2008.05.011]
- 51 **Fisher SB**, Patel SH, Bagci P, Kooby DA, El-Rayes BF, Staley CA, Adsay NV, Maithel SK. An analysis of human equilibrative nucleoside transporter-1, ribonucleoside reductase subunit M1, ribonucleoside reductase subunit M2, and excision repair cross-complementing gene-1 expression in patients with resected pancreatic adenocarcinoma: implications for adjuvant treatment. *Cancer* 2013; **119**: 445-453 [PMID: 22569992 DOI: 10.1002/cncr.27619]
- 52 **Khoja L**, Backen A, Sloane R, Menasce L, Ryder D, Krebs M, Board R, Clack G, Hughes A, Blackhall F, Valle JW, Dive C. A pilot study to explore circulating tumour cells in pancreatic cancer as a novel biomarker. *Br J Cancer* 2012; **106**: 508-516 [PMID: 22187035 DOI: 10.1038/bjc.2011.545]
- 53 **Maréchal R**, Bachet JB, Mackey JR, Dalban C, Demetter P, Graham K, Couvelard A, Svrcek M, Bardier-Dupas A, Hammel P, Sauvanet A, Louvet C, Paye F, Rougier P, Penna C, André T, Dumontet C, Cass CE, Jordheim LP, Matera EL, Closset J, Salmon I, Devière J, Emile JF, Van Laethem JL. Levels of gemcitabine transport and metabolism proteins predict survival times of patients treated with gemcitabine for pancreatic adenocarcinoma. *Gastroenterology* 2012; **143**: 664-74. e1-6 [PMID: 22705007 DOI: 10.1053/j.gastro.2012.06.006]
- 54 **Neoptolemos JP**, Greenhalf W, Ghaneh P, Palmer DH, Cox TF, Garner E, Campbell F, Mackey JR, Moore MJ, Valle JW, McDonald A, Tebbutt NC, Dervenis C, Glimelius B, Charnley BM, Lacaine F, Mayerle J, Rawcliffe CL, Bassi C, Buchler MW. HENT1 tumor levels to predict survival of pancreatic ductal adenocarcinoma patients who received adjuvant gemcitabine and adjuvant 5FU on the ESPAC trials. ASCO 2013, J Clin Oncol 31, 2013 (suppl; abstr 4006). Available from: URL: <http://meetinglibrary.asco.org/content/111340-132>
- 55 **Tsujie M**, Nakamori S, Nakahira S, Takahashi Y, Hayashi N, Okami J, Nagano H, Dono K, Umeshita K, Sakon M, Monden M. Human equilibrative nucleoside transporter 1, as a predictor of 5-fluorouracil resistance in human pancreatic cancer. *Anticancer Res* 2007; **27**: 2241-2249 [PMID: 17695509]
- 56 **Kim R**, Tan A, Lai KK, Jiang J, Wang Y, Rybicki LA, Liu X. Prognostic roles of human equilibrative transporter 1 (hENT-1) and ribonucleoside reductase subunit M1 (RRM1) in resected pancreatic cancer. *Cancer* 2011; **117**: 3126-3134 [PMID: 21264835 DOI: 10.1002/cncr.25883]
- 57 **Poplin E**, Wasan H, Rolfe L, Raponi M, Ikdahl T, Bondarenko I, Davidenko I, Bondar V, Garin A, Boeck SH, Heinemann V, Bassi C, Evans TRJ, Voong C, Kaur P, Isaacson JD, Allen AR. Randomized multicenter, phase II study of CO-101 versus gemcitabine in patients with metastatic pancreatic ductal adenocarcinoma (mPDAC) and a prospective evaluation of the of the association between tumor hENT1 expression and clinical outcome with gemcitabine treatment; J Clin Oncol 31, 2013 (suppl; abstr 4007). Available from: URL: <http://meetinglibrary.asco.org/content/113230-132>
- 58 **Brody J**, Dasgupta A, Costantino CL, Kennedy E, Yeo CJ, Witkiewicz AK. Correlation of HuR cytoplasmic expression in pancreatic cancer and overall patient survival when treated with gemcitabine in the adjuvant setting. ASCO 2009, J Clin Oncol 27: 15s, 2009 (suppl; abstr 11097). Available from: URL: <http://meetinglibrary.asco.org/content/35081-65>
- 59 **Costantino CL**, Witkiewicz AK, Kuwano Y, Cozzitorto JA, Kennedy EP, Dasgupta A, Keen JC, Yeo CJ, Gorospe M, Brody JR. The role of HuR in gemcitabine efficacy in pancreatic cancer: HuR Up-regulates the expression of the gemcitabine metabolizing enzyme deoxycytidine kinase. *Cancer Res* 2009; **69**: 4567-4572 [PMID: 19487279 DOI: 10.1158/0008-5472.CAN-09-0371]
- 60 **Richards NG**, Rittenhouse DW, Freydin B, Cozzitorto JA, Grenda D, Rui H, Gonye G, Kennedy EP, Yeo CJ, Brody JR, Witkiewicz AK. HuR status is a powerful marker for prognosis and response to gemcitabine-based chemotherapy for resected pancreatic ductal adenocarcinoma patients. *Ann Surg* 2010; **252**: 499-505; discussion 505-6 [PMID: 20739850]
- 61 **Infante JR**, Matsubayashi H, Sato N, Tonascia J, Klein AP, Riall TA, Yeo C, Iacobuzio-Donahue C, Goggins M. Peritumoral fibroblast SPARC expression and patient outcome with resectable pancreatic adenocarcinoma. *J Clin Oncol* 2007; **25**: 319-325 [PMID: 17235047 DOI: 10.1200/JCO.2006.07.8824]
- 62 **Mantoni TS**, Schendel RR, Rödel F, Niedobitek G, Al-Assar O, Masamune A, Brunner TB. Stromal SPARC expression and patient survival after chemoradiation for non-resectable pancreatic adenocarcinoma. *Cancer Biol Ther* 2008; **7**: 1806-1815 [PMID: 18787407 DOI: 10.4161/cbt.7.11.6846]
- 63 **Jacobson A**, Cunningham JL. Connective tissue growth factor in tumor pathogenesis. *Fibrogenesis Tissue Repair* 2012; **5**: S8 [PMID: 23259759 DOI: 10.1186/1755-1536-5-S1-S8]
- 64 **Ijichi H**, Chytil A, Gorska AE, Aakre ME, Bieri B, Tada M, Mohri D, Miyabayashi K, Asaoka Y, Maeda S, Ikenoue T, Tateishi K, Wright CV, Koike K, Omata M, Moses HL. Inhibiting Cxcr2 disrupts tumor-stromal interactions and improves survival in a mouse model of pancreatic ductal adenocarcinoma. *J Clin Invest* 2011; **121**: 4106-4117 [PMID: 21926469 DOI: 10.1172/JCI42754]
- 65 **Aikawa T**, Gunn J, Spong SM, Klaus SJ, Korc M. Connec-

- tive tissue growth factor-specific antibody attenuates tumor growth, metastasis, and angiogenesis in an orthotopic mouse model of pancreatic cancer. *Mol Cancer Ther* 2006; **5**: 1108-1116 [PMID: 16731742 DOI: 10.1158/1535-7163.MCT-05-0516]
- 66 **Picozzi VJ**, Pipas JM, Koong A, Giaccia A, Bahary N, Krishnamurthi SS, Lopez CD, O'Dwyer PJ, Modelska K, Poolman V, Chou J, Zhong M, Porter S, Neff T, Valone F; FG-3019, a human monoclonal antibody to CTGF, with gemcitabine/erlotinib in patients with locally advanced or metastatic pancreatic ductal adenocarcinoma. ASCO GI 2013, *J Clin Oncol* 30: 2012 (suppl 34; abstr 213). Available from: URL: <http://meetinglibrary.asco.org/content/106279-133>
- 67 **Iacobuzio-Donahue CA**, Fu B, Yachida S, Luo M, Abe H, Henderson CM, Vilardeell F, Wang Z, Keller JW, Banerjee P, Herman JM, Cameron JL, Yeo CJ, Halushka MK, Eshleman JR, Raben M, Klein AP, Hruban RH, Hidalgo M, Laheru D. DPC4 gene status of the primary carcinoma correlates with patterns of failure in patients with pancreatic cancer. *J Clin Oncol* 2009; **27**: 1806-1813 [PMID: 19273710 DOI: 10.1200/JCO.2008.17.7188]
- 68 **Arnold SA**, Rivera LB, Miller AF, Carbon JG, Dineen SP, Xie Y, Castrillon DH, Sage EH, Puolakkainen P, Bradshaw AD, Brekken RA. Lack of host SPARC enhances vascular function and tumor spread in an orthotopic murine model of pancreatic carcinoma. *Dis Model Mech* 2010; **3**: 57-72 [PMID: 20007485 DOI: 10.1242/dmm.003228]
- 69 **Collisson EA**, Sadanandam A, Olson P, Gibb WJ, Truitt M, Gu S, Cooc J, Weinkle J, Kim GE, Jakkula L, Feiler HS, Ko AH, Olshen AB, Danenberg KL, Tempero MA, Spellman PT, Hanahan D, Gray JW. Subtypes of pancreatic ductal adenocarcinoma and their differing responses to therapy. *Nat Med* 2011; **17**: 500-503 [PMID: 21460848 DOI: 10.1038/nm.2344]
- 70 **Dancey JE**, Bedard PL, Onetto N, Hudson TJ. The genetic basis for cancer treatment decisions. *Cell* 2012; **148**: 409-420 [PMID: 22304912 DOI: 10.1016/j.cell.2012.01.014]
- 71 **Von Hoff DD**, Stephenson JJ, Rosen P, Loesch DM, Borad MJ, Anthony S, Jameson G, Brown S, Cantafio N, Richards DA, Fitch TR, Wasserman E, Fernandez C, Green S, Sutherland W, Bittner M, Alarcon A, Mallery D, Penny R. Pilot study using molecular profiling of patients' tumors to find potential targets and select treatments for their refractory cancers. *J Clin Oncol* 2010; **28**: 4877-4883 [PMID: 20921468 DOI: 10.1200/JCO.2009.26.5983]
- 72 **Formentini A**, Sander S, Denzer S, Straeter J, Henne-Bruns D, Kornmann M. Thymidylate synthase expression in resectable and unresectable pancreatic cancer: role as predictive or prognostic marker? *Int J Colorectal Dis* 2007; **22**: 49-55 [PMID: 16538493 DOI: 10.1007/s00384-006-0111-z]
- 73 **Kondo N**, Murakami Y, Uemura K, Sudo T, Hashimoto Y, Nakashima A, Ohge H, Sueda T. Prognostic impact of dihydropyrimidine dehydrogenase expression on pancreatic adenocarcinoma patients treated with S-1-based adjuvant chemotherapy after surgical resection. *J Surg Oncol* 2011; **104**: 146-154 [PMID: 21538357 DOI: 10.1002/jso.21955]
- 74 **Brody JR**, Hucl T, Costantino CL, Eshleman JR, Gallmeier E, Zhu H, van der Heijden MS, Winter JM, Wikiewicz AK, Yeo CJ, Kern SE. Limits to thymidylate synthase and TP53 genes as predictive determinants for fluoropyrimidine sensitivity and further evidence for RNA-based toxicity as a major influence. *Cancer Res* 2009; **69**: 984-991 [PMID: 19155291 DOI: 10.1158/0008-5472.CAN-08-3610]
- 75 **Villarreal MC**, Rajeshkumar NV, Garrido-Laguna I, De Jesus-Acosta A, Jones S, Maitra A, Hruban RH, Eshleman JR, Klein A, Laheru D, Donehower R, Hidalgo M. Personalizing cancer treatment in the age of global genomic analyses: PALB2 gene mutations and the response to DNA damaging agents in pancreatic cancer. *Mol Cancer Ther* 2011; **10**: 3-8 [PMID: 21135251 DOI: 10.1158/1535-7163.MCT-10-0893]
- 76 **Fogelman DR**, Wolff RA, Kopetz S, Javle M, Bradley C, Mok I, Cabanillas F, Abbruzzese JL. Evidence for the efficacy of Iniparib, a PARP-1 inhibitor, in BRCA2-associated pancreatic cancer. *Anticancer Res* 2011; **31**: 1417-1420 [PMID: 21508395]
- 77 **Nakano Y**, Tanno S, Koizumi K, Nishikawa T, Nakamura K, Minoguchi M, Izawa T, Mizukami Y, Okumura T, Kohgo Y. Gemcitabine chemoresistance and molecular markers associated with gemcitabine transport and metabolism in human pancreatic cancer cells. *Br J Cancer* 2007; **96**: 457-463 [PMID: 17224927 DOI: 10.1038/sj.bjc.6603559]
- 78 **Neoptolemos JP**, Stocken DD, Bassi C, Ghaneh P, Cunningham D, Goldstein D, Padbury R, Moore MJ, Gallinger S, Mariette C, Wente MN, Izbicki JR, Friess H, Lerch MM, Dervenis C, Oláh A, Butturini G, Doi R, Lind PA, Smith D, Valle JW, Palmer DH, Buckels JA, Thompson J, McKay CJ, Rawcliffe CL, Büchler MW. Adjuvant chemotherapy with fluorouracil plus folinic acid vs gemcitabine following pancreatic cancer resection: a randomized controlled trial. *JAMA* 2010; **304**: 1073-1081 [PMID: 20823433 DOI: 10.1001/jama.2010.1275]
- 79 **Srikantan S**, Gorospe M. HuR function in disease. *Front Biosci (Landmark Ed)* 2012; **17**: 189-205 [PMID: 22201738 DOI: 10.2741/3921]
- 80 **Frese KK**, Neesse A, Cook N, Bapiro TE, Lolkema MP, Jodrell DI, Tuveson DA. nab-Paclitaxel potentiates gemcitabine activity by reducing cytidine deaminase levels in a mouse model of pancreatic cancer. *Cancer Discov* 2012; **2**: 260-269 [PMID: 22585996 DOI: 10.1158/2159-8290.CD-11-0242]
- 81 **Charrier A**, Brigstock DR. Regulation of pancreatic function by connective tissue growth factor (CTGF, CCN2). *Cytokine Growth Factor Rev* 2013; **24**: 59-68 [PMID: 22884427 DOI: 10.1016/j.cytogfr.2012.07.001]
- 82 **Eguchi D**, Ikenaga N, Ohuchida K, Kozono S, Cui L, Fujiwara K, Fujino M, Ohtsuka T, Mizumoto K, Tanaka M. Hypoxia enhances the interaction between pancreatic stellate cells and cancer cells via increased secretion of connective tissue growth factor. *J Surg Res* 2013; **181**: 225-233 [PMID: 22795353 DOI: 10.1016/j.jss.2012.06.051]
- 83 **Dornhöfer N**, Spong S, Bennewith K, Salim A, Klaus S, Kambham N, Wong C, Kaper F, Sutphin P, Nacamuli R, Höckel M, Le Q, Longaker M, Yang G, Koong A, Giaccia A. Connective tissue growth factor-specific monoclonal antibody therapy inhibits pancreatic tumor growth and metastasis. *Cancer Res* 2006; **66**: 5816-5827 [PMID: 16740721 DOI: 10.1158/0008-5472.CAN-06-0081]
- 84 **Neesse A**, Frese KK, Bapiro TE, Nakagawa T, Sternlicht MD, Seeley TW, Pilarsky C, Jodrell DI, Spong SM, Tuveson DA. CTGF antagonism with mAb FG-3019 enhances chemotherapy response without increasing drug delivery in murine ductal pancreas cancer. *Proc Natl Acad Sci USA* 2013; **110**: 12325-12330 [PMID: 23836645 DOI: 10.1073/pnas.1300415110]
- 85 **Tempero MA**, Klimstra D, Berlin J, Hollingsworth T, Kim P, Merchant N, Moore M, Pleskow D, Wang-Gillam A, Lowy AM. Changing the way we do business: recommendations to accelerate biomarker development in pancreatic cancer. *Clin Cancer Res* 2013; **19**: 538-540 [PMID: 23344262 DOI: 10.1158/1078-0432.CCR-12-2745]
- 86 **McShane LM**, Altman DG, Sauerbrei W, Taube SE, Gion M, Clark GM. REporting recommendations for tumor MARKer prognostic studies (REMARK). *Breast Cancer Res Treat* 2006; **100**: 229-235 [PMID: 16932852 DOI: 10.1007/s10549-006-9242-8]

P- Reviewers: Liu JY, Kozarek R S- Editor: Qi Y
L- Editor: Wang TQ E- Editor: Wang CH





Published by **Baishideng Publishing Group Inc**

8226 Regency Drive, Pleasanton, CA 94588, USA

Telephone: +1-925-223-8242

Fax: +1-925-223-8243

E-mail: bpgoffice@wjgnet.com

Help Desk: <http://www.wjgnet.com/esps/helpdesk.aspx>

<http://www.wjgnet.com>



ISSN 1007-9327

