

World Journal of *Clinical Cases*

World J Clin Cases 2022 April 6; 10(10): 2976-3320



Contents

Thrice Monthly Volume 10 Number 10 April 6, 2022

REVIEW

- 2976** Gut microbiota in gastrointestinal diseases during pregnancy
Liu ZZ, Sun JH, Wang WJ
- 2990** Targeting metabolism: A potential strategy for hematological cancer therapy
Tang X, Chen F, Xie LC, Liu SX, Mai HR

MINIREVIEWS

- 3005** Elevated intra-abdominal pressure: A review of current knowledge
Łagosz P, Sokolski M, Biegus J, Tycinska A, Zymlinski R

ORIGINAL ARTICLE

Case Control Study

- 3014** Changes in corneal nerve morphology and function in patients with dry eyes having type 2 diabetes
Fang W, Lin ZX, Yang HQ, Zhao L, Liu DC, Pan ZQ
- 3027** Combined sevoflurane-dexmedetomidine and nerve blockade on post-surgical serum oxidative stress biomarker levels in thyroid cancer patients
Du D, Qiao Q, Guan Z, Gao YF, Wang Q

Retrospective Cohort Study

- 3035** Early warning prevention and control strategies to reduce perioperative venous thromboembolism in patients with gastrointestinal cancer
Lu Y, Chen FY, Cai L, Huang CX, Shen XF, Cai LQ, Li XT, Fu YY, Wei J
- 3047** Dose-response relationship between risk factors and incidence of COVID-19 in 325 hospitalized patients: A multicenter retrospective cohort study
Zhao SC, Yu XQ, Lai XF, Duan R, Guo DL, Zhu Q

Retrospective Study

- 3060** Preventive online and offline health management intervention in polycystic ovary syndrome
Liu R, Li M, Wang P, Yu M, Wang Z, Zhang GZ
- 3069** Evidence-based intervention on postoperative fear, compliance, and self-efficacy in elderly patients with hip fracture
Fu Y, Zhu LJ, Li DC, Yan JL, Zhang HT, Xuan YH, Meng CL, Sun YH
- 3078** Significance of dysplasia in bile duct resection margin in patients with extrahepatic cholangiocarcinoma: A retrospective analysis
Choe JW, Kim HJ, Kim JS

- 3088** Diagnostic value and safety of medical thoracoscopy for pleural effusion of different causes

Liu XT, Dong XL, Zhang Y, Fang P, Shi HY, Ming ZJ

Observational Study

- 3101** Oxaliplatin-induced neuropathy and colo-rectal cancer patient's quality of life: Practical lessons from a prospective cross-sectional, real-world study

Prutianu I, Alexa-Stratulat T, Cristea EO, Nicolau A, Moisuc DC, Covrig AA, Ivanov K, Croitoru AE, Miron MI, Dinu MI, Ivanov AV, Marinca MV, Radu I, Gafton B

- 3113** Breast-conserving surgery and sentinel lymph node biopsy for breast cancer and their correlation with the expression of polyligand proteoglycan-1

Li FM, Xu DY, Xu Q, Yuan Y

SYSTEMATIC REVIEWS

- 3121** Clinical significance of aberrant left hepatic artery during gastrectomy: A systematic review

Tao W, Peng D, Cheng YX, Zhang W

META-ANALYSIS

- 3131** Betel quid chewing and oral potential malignant disorders and the impact of smoking and drinking: A meta-analysis

Lin HJ, Wang XL, Tian MY, Li XL, Tan HZ

- 3143** Effects of physical exercise on the quality-of-life of patients with haematological malignancies and thrombocytopenia: A systematic review and meta-analysis

Yang YP, Pan SJ, Qiu SL, Tung TH

CASE REPORT

- 3156** Primary malignant peritoneal mesothelioma mimicking tuberculous peritonitis: A case report

Lin LC, Kuan WY, Shiu BH, Wang YT, Chao WR, Wang CC

- 3164** Endoscopic submucosal dissection combined with adjuvant chemotherapy for early-stage neuroendocrine carcinoma of the esophagus: A case report

Tang N, Feng Z

- 3170** Lymph-node-first presentation of Kawasaki disease in a 12-year-old girl with cervical lymphadenitis caused by *Mycoplasma pneumoniae*: A case report

Kim N, Choi YJ, Na JY, Oh JW

- 3178** Tuberculosis-associated hemophagocytic lymphohistiocytosis misdiagnosed as systemic lupus erythematosus: A case report

Chen WT, Liu ZC, Li MS, Zhou Y, Liang SJ, Yang Y

- 3188** Migration of a Hem-o-Lok clip to the renal pelvis after laparoscopic partial nephrectomy: A case report

Sun J, Zhao LW, Wang XL, Huang JG, Fan Y

- 3194** Ectopic intrauterine device in the bladder causing cystolithiasis: A case report
Yu HT, Chen Y, Xie YP, Gan TB, Gou X
- 3200** Giant tumor resection under ultrasound-guided nerve block in a patient with severe asthma: A case report
Liu Q, Zhong Q, Zhou NN, Ye L
- 3206** Myomatous erythrocytosis syndrome: A case report
Shu XY, Chen N, Chen BY, Yang HX, Bi H
- 3213** Middle thyroid vein tumor thrombus in metastatic papillary thyroid microcarcinoma: A case report and review of literature
Gui Y, Wang JY, Wei XD
- 3222** Severe pneumonia and acute myocardial infarction complicated with pericarditis after percutaneous coronary intervention: A case report
Liu WC, Li SB, Zhang CF, Cui XH
- 3232** IgA nephropathy treatment with traditional Chinese medicine: A case report
Zhang YY, Chen YL, Yi L, Gao K
- 3241** Appendico-vesicocolonic fistula: A case report and review of literature
Yan H, Wu YC, Wang X, Liu YC, Zuo S, Wang PY
- 3251** *Scedosporium apiospermum* infection of the lumbar vertebrae: A case report
Shi XW, Li ST, Lou JP, Xu B, Wang J, Wang X, Liu H, Li SK, Zhen P, Zhang T
- 3261** Woman diagnosed with obsessive-compulsive disorder became delusional after childbirth: A case report
Lin SS, Gao JF
- 3268** Emphysematous pyelonephritis: Six case reports and review of literature
Ma LP, Zhou N, Fu Y, Liu Y, Wang C, Zhao B
- 3278** Atypical infantile-onset Pompe disease with good prognosis from mainland China: A case report
Zhang Y, Zhang C, Shu JB, Zhang F
- 3284** *Mycobacterium tuberculosis* bacteremia in a human immunodeficiency virus-negative patient with liver cirrhosis: A case report
Lin ZZ, Chen D, Liu S, Yu JH, Liu SR, Zhu ML
- 3291** Cervical aortic arch with aneurysm formation and an anomalous right subclavian artery and left vertebral artery: A case report
Wu YK, Mao Q, Zhou MT, Liu N, Yu X, Peng JC, Tao YY, Gong XQ, Yang L, Zhang XM
- 3297** Dedifferentiated chondrosarcoma of the middle finger arising from a solitary enchondroma: A case report
Yonezawa H, Yamamoto N, Hayashi K, Takeuchi A, Miwa S, Igarashi K, Morinaga S, Asano Y, Saito S, Tome Y, Ikeda H, Nojima T, Tsuchiya H

- 3306** Endoscopic-catheter-directed infusion of diluted (-)-noradrenaline for atypical hemobilia caused by liver abscess: A case report
Zou H, Wen Y, Pang Y, Zhang H, Zhang L, Tang LJ, Wu H
- 3313** *Pneumocystis jiroveci* pneumonia after total hip arthroplasty in a dermatomyositis patient: A case report
Hong M, Zhang ZY, Sun XW, Wang WG, Zhang QD, Guo WS

ABOUT COVER

Editorial Board Member of *World Journal of Clinical Cases*, Hui-Jeong Hwang, MD, PhD, Associate Professor, Department of Cardiology, Kyung Hee University Hospital at Gangdong, Kyung Hee University College of Medicine, Seoul 05278, South Korea. neonic7749@hanmail.net

AIMS AND SCOPE

The primary aim of *World Journal of Clinical Cases* (WJCC, *World J Clin Cases*) is to provide scholars and readers from various fields of clinical medicine with a platform to publish high-quality clinical research articles and communicate their research findings online.

WJCC mainly publishes articles reporting research results and findings obtained in the field of clinical medicine and covering a wide range of topics, including case control studies, retrospective cohort studies, retrospective studies, clinical trials studies, observational studies, prospective studies, randomized controlled trials, randomized clinical trials, systematic reviews, meta-analysis, and case reports.

INDEXING/ABSTRACTING

The WJCC is now indexed in Science Citation Index Expanded (also known as SciSearch®), Journal Citation Reports/Science Edition, Scopus, PubMed, and PubMed Central. The 2021 Edition of Journal Citation Reports® cites the 2020 impact factor (IF) for WJCC as 1.337; IF without journal self cites: 1.301; 5-year IF: 1.742; Journal Citation Indicator: 0.33; Ranking: 119 among 169 journals in medicine, general and internal; and Quartile category: Q3. The WJCC's CiteScore for 2020 is 0.8 and Scopus CiteScore rank 2020: General Medicine is 493/793.

RESPONSIBLE EDITORS FOR THIS ISSUE

Production Editor: Xu Guo; Production Department Director: Xiang Li; Editorial Office Director: Jin-Lai Wang.

NAME OF JOURNAL

World Journal of Clinical Cases

ISSN

ISSN 2307-8960 (online)

LAUNCH DATE

April 16, 2013

FREQUENCY

Thrice Monthly

EDITORS-IN-CHIEF

Bao-Gan Peng, Jerzy Tadeusz Chudek, George Kontogeorgos, Maurizio Serati, Ja Hyeon Ku

EDITORIAL BOARD MEMBERS

<https://www.wjgnet.com/2307-8960/editorialboard.htm>

PUBLICATION DATE

April 6, 2022

COPYRIGHT

© 2022 Baishideng Publishing Group Inc

INSTRUCTIONS TO AUTHORS

<https://www.wjgnet.com/bpg/gerinfo/204>

GUIDELINES FOR ETHICS DOCUMENTS

<https://www.wjgnet.com/bpg/GerInfo/287>

GUIDELINES FOR NON-NATIVE SPEAKERS OF ENGLISH

<https://www.wjgnet.com/bpg/gerinfo/240>

PUBLICATION ETHICS

<https://www.wjgnet.com/bpg/GerInfo/288>

PUBLICATION MISCONDUCT

<https://www.wjgnet.com/bpg/gerinfo/208>

ARTICLE PROCESSING CHARGE

<https://www.wjgnet.com/bpg/gerinfo/242>

STEPS FOR SUBMITTING MANUSCRIPTS

<https://www.wjgnet.com/bpg/GerInfo/239>

ONLINE SUBMISSION

<https://www.f6publishing.com>



Targeting metabolism: A potential strategy for hematological cancer therapy

Xue Tang, Fen Chen, Li-Chun Xie, Si-Xi Liu, Hui-Rong Mai

Specialty type: Medicine, research and experimental

Provenance and peer review: Unsolicited article; Externally peer reviewed.

Peer-review model: Single blind

Peer-review report's scientific quality classification

Grade A (Excellent): 0
Grade B (Very good): B
Grade C (Good): 0
Grade D (Fair): 0
Grade E (Poor): 0

P-Reviewer: Azmi AS, United States

Received: July 15, 2021

Peer-review started: July 15, 2021

First decision: October 18, 2021

Revised: November 1, 2021

Accepted: February 27, 2022

Article in press: February 27, 2022

Published online: April 6, 2022



Xue Tang, Fen Chen, Li-Chun Xie, Si-Xi Liu, Hui-Rong Mai, Department of Hematology and Oncology, Shenzhen Children's Hospital, Shenzhen 518038, Guangdong Province, China

Corresponding author: Hui-Rong Mai, MD, Chief Doctor, Department of Hematology and Oncology, Shenzhen Children's Hospital, No. 7019 Yitian Road, Futian, Shenzhen 518038, Guangdong Province, China. maihuirong@163.com

Abstract

Most hematological cancer-related relapses and deaths are caused by metastasis; thus, the importance of this process as a target of therapy should be considered. Hematological cancer is a type of cancer in which metabolism plays an essential role in progression. Therefore, we are required to block fundamental metastatic processes and develop specific preclinical and clinical strategies against those biomarkers involved in the metabolic regulation of hematological cancer cells, which do not rely on primary tumor responses. To understand progress in this field, we provide a summary of recent developments in the understanding of metabolism in hematological cancer and a general understanding of biomarkers currently used and under investigation for clinical and preclinical applications involving drug development. The signaling pathways involved in cancer cell metabolism are highlighted and shed light on how we could identify novel biomarkers involved in cancer development and treatment. This review provides new insights into biomolecular carriers that could be targeted as anticancer biomarkers.

Key Words: Metabolism; Metastasis; Hematological cancer; Biomarker; Cancer; Anticancer

©The Author(s) 2022. Published by Baishideng Publishing Group Inc. All rights reserved.

Core Tip: Hematological cancer is a type of cancer in which metabolism plays an essential role in progression. We provide a summary of recent developments in the understanding of metabolism in hematological cancer and provide a general understanding of biomarkers currently used and under investigation for clinical and preclinical applications involving drug development. This review provides new insights into biomolecular carriers that could be targeted as anticancer biomarkers.

Citation: Tang X, Chen F, Xie LC, Liu SX, Mai HR. Targeting metabolism: A potential strategy for hematological cancer therapy. *World J Clin Cases* 2022; 10(10): 2990-3004

URL: <https://www.wjgnet.com/2307-8960/full/v10/i10/2990.htm>

DOI: <https://dx.doi.org/10.12998/wjcc.v10.i10.2990>

INTRODUCTION

Resistance to cancer therapy is diverse and multifactorial, and is the most difficult challenge in oncology. Although the early phase of treatment is often successful, resistance 75% characterized by relapse, anti-tumoral drug desensitization, or even aggressive spread 75% can develop. Many studies have investigated the basis of tumorigenesis, progression, and drug resistance, and over the last decade, the diverse roles of host cells in promoting cancer development and progression have been comprehensively studied. Different biomarkers serve as cancer drug targets and as diagnostic markers, and distinct cancer-related signaling pathways involved in tumor progression have been characterized. An increasing number of data indicate that cancer therapy can induce host-mediated local and systemic responses, many of which modify the equilibrium of the tumor microenvironment, facilitating or supporting tumor progression, as well as resistance to antitumor treatment. Most hematological cancer-related relapses and deaths are caused by metastasis; thus, the importance of this process as a therapeutic target should be considered. Metastatic tumor cells that arrive in distant tissues, surrounded by unfamiliar cells and a foreign microenvironment, are likely to die; however, those that survive can generate metastatic tumors with markedly different biology from that of the primary tumor.

This review describes recent advances in understanding how tumor progression is precisely tuned by signaling pathways, how these regulators control therapeutic outcome, the mechanisms that regulate the metabolism in hematological cancer, and the biomarkers currently used and under investigation for clinical and preclinical applications involving drug development. Evaluating the host response and the biological targets for cancer therapy is key to the development of precision medicine in oncology.

CANCER SIGNALING PATHWAYS

Hypoxia signaling in cancer and approaches to enforce tumor regression

Cancer cells progress as a consequence of genetic alterations of signaling pathways that promote cell growth and survival, whereas their expansion relies on nutrient storage. Oxygen limitation is central in control of neovascularization, glucose metabolism, survival, and tumor spread. This pleiotropic action is orchestrated by hypoxia-inducible factor (HIF), which is a master transcriptional factor in nutrient stress signaling[1]. HIF-1 α is recognized as an important cancer drug target, despite its location and expression pattern. Many recent studies have shown promising evidence to correlate HIF-1 α and tumor metastasis, angiogenesis, poor prognosis, and drug resistance. Low oxygen supply (or hypoxia) is a common characteristic in solid tumors. Since an adaptive response to hypoxic stress is triggered, tumor cells undergo several survival activation pathways to initiate their essential biological processes in ways that are different compared with normal cells. Recent findings at both cellular and molecular levels rule out a role for the HIF-1 α pathway and its downstream signalosome as a crucial cancer cell survival pathway. Targeting the HIF-1 α pathway has been challenging due to its expression level, location, and drug accessibility, but promising progress has been made in the past 20 years. This section attempts to summarize the role and regulatory mechanism of HIF-1 α during cancer development, and recent therapeutic approaches specifically targeting this important pathway[1].

HIF-1 α is the most studied of the HIF proteins, and the roles of other HIF-1 subunits remain elusive in cancer cells related to intracellular pH regulation, metabolism, activation, invasion, autophagy, apoptosis, and necrosis. Targeting HIF is a focus of efforts to develop novel anticancer therapies. There are many new therapeutic approaches to accelerate necrotic cell death (not apoptosis, which is considered to inhibit cancer cell growth) and tumor regression by targeting metabolism within the microenvironment and downstream signaling[2] (see Figure 1A).

Recent research suggests that induction of HIF proteins in normoxic conditions (oxygen tensions between 10%-21%) is likely to have serious consequences, such as chronic inflammation or pathological symptoms[3]. Chronic inflammation is self-perpetuating, distorting the microenvironment as a result of aberrantly active transcription factors[4]. As a consequence, alterations in growth factors, chemokines, cytokines, and reactive oxygen species balance occur within the cellular milieu that enable the growth and survival needed for *de novo* development of cancer and metastasis[5]. It is thought that understanding the crosstalk between two key transcription factors, nuclear factor-kappaB (NF- κ B) and HIF, will greatly enhance the process of drug development[6].

HIF activity is also involved in angiogenesis that is required for tumor growth, so HIF inhibitors, such as phenethyl isothiocyanate and acriflavine, have been under investigation for anticancer effects[7,8].

HIF signaling pathways in metabolic regulation

Different metabolic regulatory mechanisms affect the functions of HIF, highlighting the possibility that HIF activation alters the metabolic signatures in cancer, and disordered metabolism triggers HIF-1 α activation of HIF. HIF hydroxylation was assumed to be regulated by the Krebs cycle product 2-oxoglutarate (2-OG), which is a substrate in reductive amidation/oxidative deamidation. The latter is further catalyzed by glutamate dehydrogenase[9]. It is likely that substrates like 2-OG become metabolic signal receivers by modulating HIF hydroxylase function. In parallel, 2-OG decreases in the cytosol in the absence of amino acid complementation related to the activation of prolyl hydroxylase domain protein 2. This signaling pathway is dependent upon mammalian target of rapamycin complex 1 (mTORC1), although the mechanism by which the low concentration of 2-OG could alter the HIF hydroxylase function in the context of cancer remains elusive[10].

Besides activation by multifactorial stimuli, including stress and danger signals, the HIF signalosome complex is activated by different antitumor molecules and signaling pathway oncogenes. The most striking observation concerns mutation of the tumor suppressor von Hippel-Lindau protein[11]. This ubiquitin E3 Ligase subunit targets HIF- α and is responsible for regulating ubiquitin-mediated protein degradation *via* the proteasome pathway[12].

The Ras/Raf/MAPK signaling pathway is affected by the activity of HIF-1 α , and regulated by transactivation by several coactivators, such as cyclic AMP response-element-binding protein (see Figures 1B and 1C). In addition, different kinases, such as p42/p44 MAPK or p38, all contribute to the activation of HIF when transcription factor p300 is also transactivated in the nuclei for p300-induced gene expressions[13]. Diverse interactions between HIF-1 α and p53 tumor suppressor pathways have been recently reported, but the results are controversial; p53 induction and activation have also been shown to suppress HIF activity in some aspects of cancer. In particular, different modes of interactions between p53 and HIF-1 α show differential outcomes by adopting different molecular complexes and different signaling pathways[14].

Despite HIF-1 α activation in cancer cells targeting a group of metabolic enzymes, signaling energy regulators and transporters suggest that HIF contributes to the Warburg effect (described in detail below). Several studies (mostly using murine models) have shown that upregulated glycolysis continues independently of HIF-1 α expression[15]. Experiments on the mechanisms of oncogenes and tumor suppressors have revealed a large spectrum of crosstalk comprising glycolysis regulation and oncogenic glycolytic activation, and these could result in HIF-1 activity *in vitro* and *in vivo*[16]. Therefore, the enzymes that target HIF-1 α in the glycolytic pathway suggest that HIF is involved in different canonical and noncanonical pathways leading to glycolysis upregulation in cancer, mimicking the Warburg effect, without being restricted by mitochondrial involvement (see Figure 1A).

Triple pathways involving PI3K–Akt/Ras–ERK/mTORC1 as examples of oncogenic signaling pathways for tumor progression

Many of the genes commonly mutated in cancer, especially during proliferative growth under aerobic glycolysis (or under the Warburg effect), generate subproducts by regulating the PI3K-Akt and Ras-ERK pathways in the early stages of tumor development from benign tissue. These pathways are differentially activated in response to hormone or cytokine signaling exclusively elicited from cell surface receptors, cytosolic receptors, and integrin molecules, which can bind to adhesion receptors (see Figures 1C and 1D). However, genetic alterations from chromosomal defects, genomic errors, transcriptomic dysfunction, and even epigenetic disorders can lead to constitutive signaling, like an oncogene signaling pathway without on-off control, in the absence of stimuli[17].

The PI3K-Akt pathway is activated through amplification or activating mutations acting on PI3K-Akt-pathway molecules, such as type I PI3K isoform PIK3CA (p110a), Akt, and the Akt signaling pathway regulator and adaptor protein PIK3R1, among others, or through splicing deletion or signaling-induced inactivation of the phosphatases hydrolyzing PI3K products acting on the phosphatase and tensin homolog and INPP4B tumor suppressors. Therefore, diverse mutations in the tumor suppressors tuberous sclerosis complex (TSC)1 and TSC2 are activated *via* signaling controlled by mTORC1[18] (see Figure 1D). The latter is indeed one important target molecule in PI3K-Akt signaling.

The Ras-ERK signaling pathway is induced through Ras mutations, or the homologous molecule Raf, which constitutively activate these proteins. Alternatively, inactivation of GTPase-activating proteins, including RASAL2 and NF1, could also lead to Ras activation (see Figure 1B)[19]. An important downstream target of Ras-ERK signaling is the transcription factor Myc. Notably, Myc (*i.e.*, c-Myc) as a transcription factor promotes transcriptionally important enzymes for survival and proliferation, and therefore, mutations that alter the Ras-ERK pathway can, when activated, amplify gene expression to promote cancer cell proliferation[20].

In addition to the specific mutations described above, other mutations can act *via* the Ras-ERK and PI3K-Akt pathways, including those that mutate the structure of oncogenes, trigger gene over-amplification, and cause fusion protein products in these cancers or proliferation-related signaling

pathways. For example, mutations of kinases, such as epidermal growth factor receptor (EGFR) and ErbB2 [examples of receptor tyrosine kinases (RTKs)], fibroblast growth factor receptor, and platelet-derived growth factor receptor (PDGFR), can be frequently detected in cancer patients. Mutations of oncogenes for G protein-coupled receptors provide another pathway to activate them[21].

Deregulated synthesis of growth factors also plays an important role in almost all cancers. Incorrect mRNA splicing-based growth factors in cancer cells expressing the appropriate receptor leads to an autocrine loop, driving persistent signaling that lacks a stop signal. Alternatively, these signaling molecules involved in carcinogenesis can be produced by proximal cells (*via* paracrine stimulation). Both possibilities involve the Ras-ERK or PI3K-Akt signaling pathways. AKT signaling is also multipotent, as it can interact with glycogen synthase kinase 3 to be recruited to mitochondria-related cell death *via* the FAS/FASL axis. AKT signaling also negatively controls the cancer suppressor gene for p53[22].

NF- κ B in cancer development and progression

The earliest evidence implicating pathogen-induced NF- κ B activation in the development of cancer was the existence of the reticuloendotheliosis virus T viral oncogene that causes avian reticuloendothelial lymphomatosis[23], and v-Rel, which shares a Rel transactivation domain with the mammalian homologs NF- κ B1, NF- κ B2, RelA (p65), cRel, and RelB to compose the NF- κ B complex[24] (see Figure 1E).

Different oncogenes from the oncogenic viruses are key activators that are responsible for NF- κ B activation in cancer T lymphocytes[25]. The cells, under such restricted response, promote cell proliferation, survival, and inflammation, contributing to the pathogenesis of lymphomas and adult T-cell lymphoblastic leukemias.

Human papillomavirus viral proteins E6 and E7 that inactivate p53 and Rb tumor suppressor genes have also been involved in NF- κ B activation, and are associated with carcinogenesis of the larynx, oropharynx, and cervix. Hepatitis B and C viruses lead to hepatocellular carcinoma, and *Helicobacter pylori* generates ulcerative colitis and gastrointestinal carcinoma, where NF- κ B activation is triggered [26].

Major chemical and physical carcinogens implicated in the initiation and/or promotion of human cancer can also activate NF- κ B. Specifically, nicotine and carcinogens in tobacco and betel nut (*Areca catechu*), have been demonstrated in the pathogenesis of head and neck and lung cancer, inducing AKT and NF- κ B and promoting cell proliferation and survival and inflammation[27]. Nicotine has been reported to directly activate these pathways *via* nicotinic receptors and AKT, whereas chemotherapy and radiation-induced DNA damage have been reported to induce NF- κ B activation *via* nuclear to cytoplasmic signaling mechanisms involving SUMOylation of the I κ B kinase complex. tumor necrosis factor- α /interferon- γ (IFN- γ), radiation, and certain chemotherapeutic drugs also induce NF- κ B activation and several target antiapoptotic genes (*TRAF*, *IAP*, *BCL-2* and *Bcl-XL*) that protect cells from therapeutic injury by these efficient chemical antitumor agents.

Warburg effect, a metabolically regulated mechanism of cancer cell proliferation

Cancer cells can grow with a high rate of glycolysis followed by lactic acid fermentation, even in the presence of abundant oxygen. They can even proliferate in a hypoxic microenvironment. This condition requires cancer cell mitochondria and related apparatus to shut down all the processes of cellular respiration[28]. The energy that supports this growth is provided by aerobic glycolysis. This phenomenon is opposite to the Pasteur effect in that the energy of aerobic glycolysis is dependent on mitochondrial oxidation. The Warburg effect shows that cancer cell energy under such circumstances is supported *via* aerobic glycolysis after mitochondrial dysfunction[29]. Cellular respiration occurs by metabolism of glucose to pyruvate and is independent of the tricarboxylic acid cycle, which is replaced by lactate dehydrogenase, which converts glucose into lactic acid that is excreted into the extracellular environment. This leads to the increased generation of additional metabolites that may particularly benefit proliferating cells.

Although the Warburg effect has been studied extensively since its discovery in 1924, its precise nature remains unclear, which hampers research of its therapeutic potential. The Warburg effect forms the basis of positron emission tomography, in which a radioactive glucose analog is injected and can be detected at higher concentrations in malignant tumors compared with healthy tissues[30].

The Warburg effect may simply be considered a consequence of mitochondrial damage in cancer, *via* an adaptation mode for low-oxygen environments within tumors. Cancer genes working as a network, help to shut down the mitochondrial functions that could be actively involved in cancer cell apoptosis [31]. In some cancers, this effect could be due to the presence of mutations in the tumor suppressor genes involved in regulation of glycolytic enzymes within mitochondria, including the M2 splice isoform of pyruvate kinase. For example, mutations in TP53 affect energy metabolism and increase glycolysis in breast cancer and colorectal carcinoma (CRC)[32]. The Warburg effect is associated with tightly regulated glucose uptake and utilization, which indicates how mitochondrial activity is regulated. Tumor cells present with increased rates of glycolysis, which can be manifested as mitochondrial damage[33].

MAJOR HEMATOLOGY CANCER TYPES

Leukemia

Leukemia is characterized by a large increase in the numbers of leukocytes in the circulation or bone marrow. Leukemia is defined as acute or chronic, and as myelogenous, lymphocytic, or mixed phenotype. Acute leukemia affects immature cells; the disease develops rapidly, with symptoms including anemia, fever, bleeding, and swelling of the lymph nodes. In chronic leukemia, the cells develop and are transported to the tissues, but the cells do not function normally.

Acute myeloid leukemia (AML) is a disorder characterized by a clonal proliferation derived from primitive hematopoietic stem cells or progenitor cells. Acute lymphoblastic leukemia (ALL) is the most common cancer found in children. When ALL develops from T cells, it is called T-ALL. T-ALL represents 15% of pediatric ALL and 25% of adult ALL[34].

Chronic myeloid leukemia is a clonal myeloproliferative neoplasm characterized by a genetic change called the Philadelphia chromosome. More than 95% of cases with chronic lymphocytic leukemia involve B lymphocytes, with the expression of CD19, CD23, CD21, CD24, and CD40, as well as CD5[35].

Growing evidence shows that deregulation of PI3K/AKT/mTORC1 signaling contributes to the pathogenesis of leukemia. Upregulated mTORC1 and mTORC2 activity has been reported to play a critical role in leukemia initiation, propagation and relapse[36-41]. mTOR constitutive activation is usually found in leukemia patients, which contributes to chemoresistance, disease progression, and unfavorable prognosis. Constitutive NF- κ B activation protects tumor cells from apoptosis and plays a crucial role in the acquisition of resistance to chemotherapy[42,43]. Constitutive NF- κ B activation frequently occurs in patients with leukemia[44].

Lymphoma

Lymphoma is a type of blood cancer that affects the lymphatic system. Abnormal lymphocytes become lymphoma cells, which are found in the lymph nodes, spleen, thymus, bone marrow, and other parts of the body[45]. The two main types of lymphoma are Hodgkin's lymphoma (HL) and non-HL (NHL). The differences in these two types are certain unique characteristics of the different lymphoma cells. In NHL, PI3K/AKT/mTOR upregulation is found frequently[46]. HL cells also have unchecked PI3K pathway activation[47]. HL cells in classical HL patients show the constitutive activity of both the canonical and noncanonical NF- κ B signaling pathways[48].

Multiple myeloma

Multiple myeloma is a cancer of the plasma cells. Plasma cells are white blood cells that normally produce antibodies. The activated mTOR signaling pathway is regarded as an essential pathway associated with disease progression[49]. The deregulated activity of the NF- κ B family of transcription factors has also been implicated in the pathogenesis of multiple myeloma, with multiple signals through the canonical and noncanonical arms to activate the NF- κ B system in myeloma cells. In fact, NF- κ B signaling promotes proliferation, survival and drug resistance of myeloma cells[50].

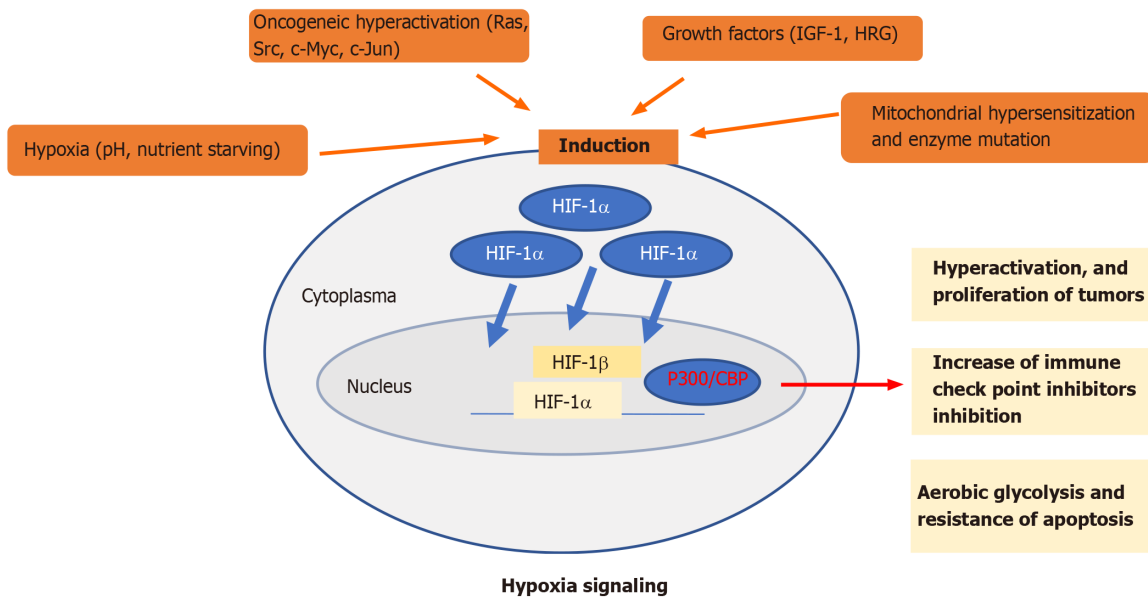
TARGETS FOR THE METABOLIC SIGNALING PATHWAY FOR HEMATOLOGICAL CANCERS

Target for the PI3K/AKT/mTOR pathway

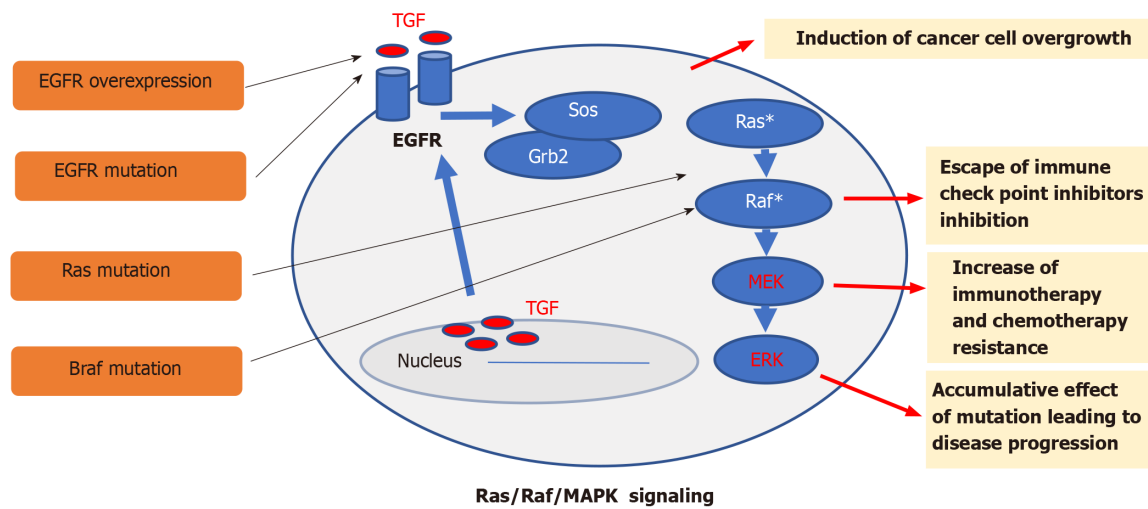
Since mTOR functions as a point of convergence between a nutrient-sensing pathway (*via* mTORC1) and as a regulator of AKT itself (*via* mTORC2), mTOR plays an important role in controlling cellular metabolism and energy homeostasis in normal and cancer cells, which is fundamental in developing effective therapies for leukemia. Several molecules that target the PI3K/AKT/mTOR signaling pathway have been investigated, showing potential therapeutic efficacy in hematological cancers, alone or in combination with chemotherapeutic drugs.

Rapamycin, an immunosuppressant and antiproliferative agent, strongly inhibits mTORC1 activity. Rapamycin forms a complex with a FK506 binding protein 12 (FKBP12), and this complex directly interacts and inhibits mTORC1, leading to cell cycle arrest and apoptosis. Rapamycin demonstrates antileukemic activity in AML blast cells, and in combination with etoposide (a topoisomerase inhibitor) shows a synergistic effect in an AML mouse model[51]. An mTORC1/2-specific inhibitor blocks AKT phosphorylation in AML cell lines and blast cells, suppresses activation of two regulator proteins S6K (S6 Kinase 1) and eukaryotic translation initiation factor 4E-binding protein 1 (4EBP1) and elicits potent antileukemic effects[52]. NVPBEZ235, a strong inhibitor of PI3K and mTORC1/2 complexes, shows strong inhibitory effects on leukemia cell proliferation and survival[53]. The emergence of compensatory mechanisms induced by long-term treatment of primary AML blast cells with PI3K/AKT/mTOR inhibitors has been discovered[54]. These mechanisms involve the upregulation of RTKs (insulin-like growth factor receptor 1, PDGFR, and EGFR). Therefore, the combined treatment with RTK inhibitors, such as sunitinib, linsitinib, or quizartinib, together with PI3K/AKT/mTOR inhibitors, is

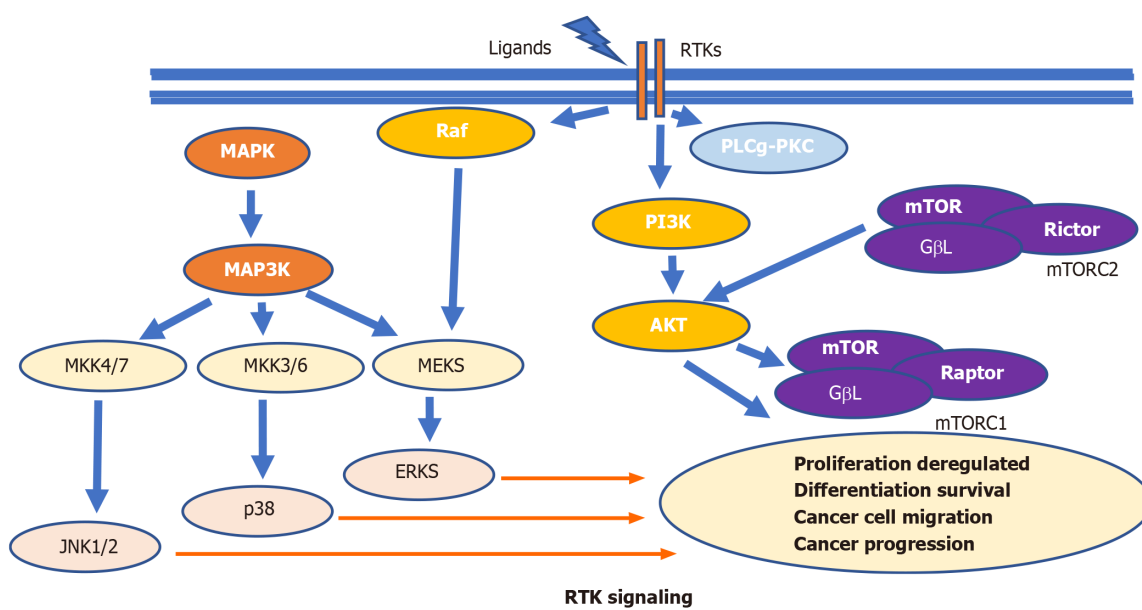
A



B



C



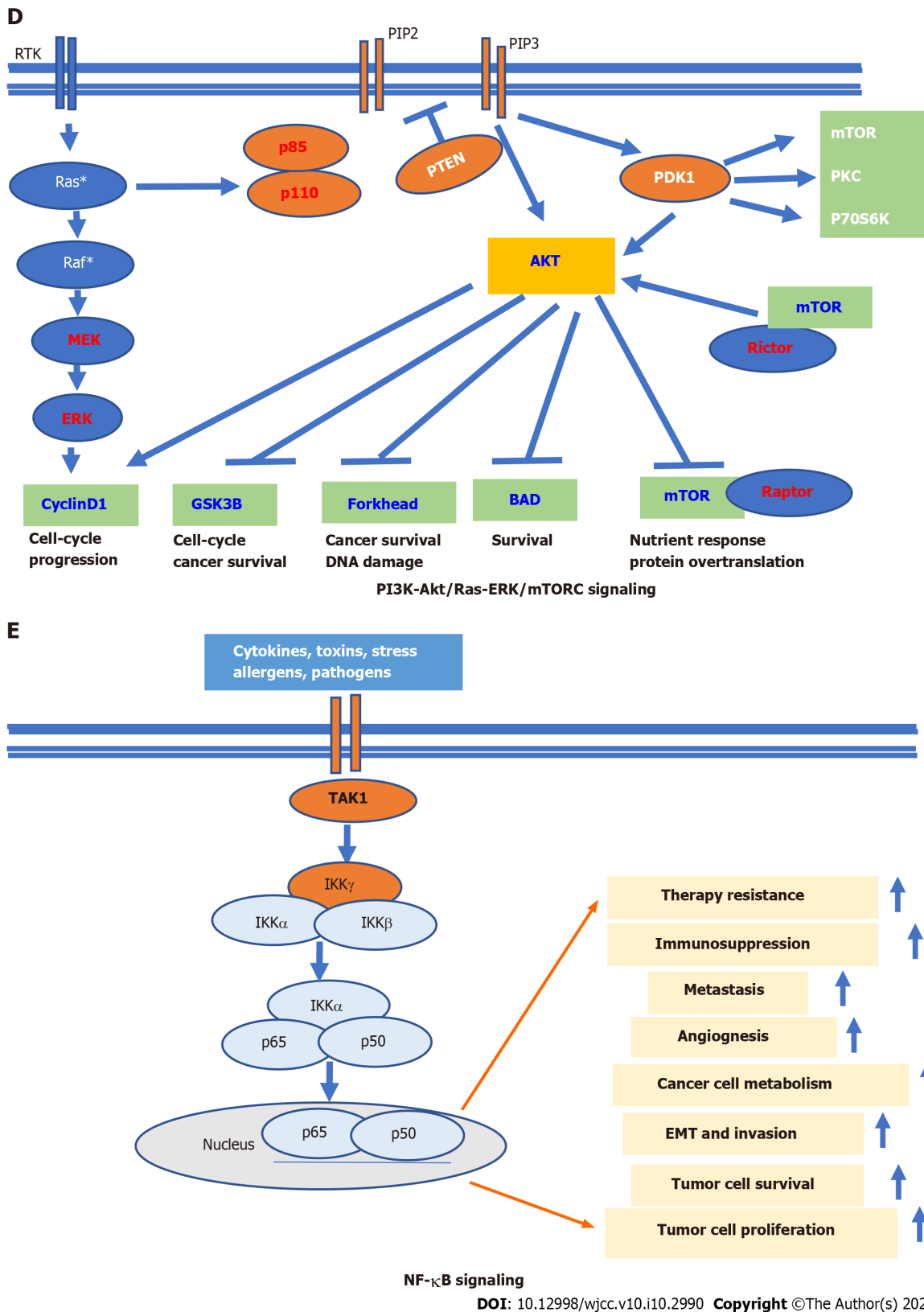


Figure 1 Summary of various signaling pathways of hematological cancers that induce cancer development and therapy resistance. A: Hypoxia signaling; B: Ras/Raf/MAPK signaling; C: RTK signaling; D: PI3K-AKT/Ras-ERK/mTOR signaling; E: NF-κB signaling. HIF: Hypoxia-inducible factor; IGF: Insulin-like growth factor; HRG: Heregulin; CBP: Cyclic AMP response-element-binding protein; TGF: Transforming growth factor; EGFR: Epidermal growth factor receptor; GSK: Glycogen synthase kinase; PDK: Phosphoinositide-dependent kinases; PKC: Protein kinase C; mTORC: Mammalian target of rapamycin complex; mTOR: Mammalian target of rapamycin; RTK: Receptor tyrosine kinase; NF-κB: Nuclear factor-kappaB.

recommended to induce a potent cytotoxic effect on AML blast cells in a clinical setting. Despite these promising findings, the combination of rapamycin analogs and chemotherapy failed to display the expected synergistic effect in clinical studies. One of the main reasons could be the presence of drug

resistance in several cell lines, caused by mutations in mTOR, FKBP12, or one of mTOR's substrates, such as 4EBP1, S6K, and cyclin-dependent kinase inhibitor[55]. Moreover, mTORC1 inhibition can produce feedback mechanisms mediated by S6K/IRS-1 that in turn increases PI3K/AKT activity, reducing the anticancer activity of mTORC1 inhibitors[54]. Some other inhibitors of mTORC, AKT, and PI3K have also been investigated for other hematological cancers; some of which also showed beneficial effects, see Table 1.

Taken together, these results suggest the potential importance of PI3K/AKT/mTOR pathway inhibitors alone or in combination with other chemotherapeutics for the treatment of leukemia.

Target for NF- κ B

Constitutive NF- κ B activation protects tumor cells from apoptotic stimuli and plays a crucial role in the acquisition of resistance to chemotherapy[81]. Several antitumor agents enhance NF- κ B activation, promoting development of these mechanisms of resistance. In this context, inhibition of the NF- κ B signaling pathway has emerged as an attractive therapeutic strategy for cancer. Bortezomib, a proteasome inhibitor, has been approved by the United States' Food and Drug Administration to treat multiple myeloma and now are in clinical trials for AML treatment[44]. Although the mechanism of proteasome inhibition is not fully understood, one of the important activities associated with the anti-myeloma functions of bortezomib is its ability to suppress the NF- κ B signaling pathway[82]. In general, I κ B, a cellular inhibitory protein of NF- κ B, is targeted by the ubiquitin-proteasome pathway for degradation after its phosphorylation at serine residues 32 and 36. Inhibition of the proteasome pathway by bortezomib has been shown to impede the degradation of I κ B α , thus blocking NF- κ B in the cytoplasm and preventing NF- κ B nuclear translocation and activation of NF- κ B target genes[83]. Some of the different kinases or proteases that participate in NF- κ B activation, such as mucosa-associated lymphoid tissue lymphoma translocation protein 1, are also being evaluated as potential targets[59,60].

Application of biomarkers involved in cancer metabolism

Cancer biomarkers are often identified from the observed phenotype alteration, genetic modification, and epigenetic switching. These molecules could be useful to determine the type of cancer, stage of cancer progression, or the spatiotemporal switch from benign status to disease progression of cancer [84]. A great number of genes including EGFR, c-myc, and Ras, have been used for cancer screening; for example, mutations in BRCA molecules are used to screen for breast cancer in the female[62-64]. Therefore, it has become more necessary to determine the origins of primary or metastatic tumors in different sites. Interestingly, chromosome modifications in tumor cells was shown to be potential indicators (markers) of tumor progression and metastasis. If these epigenetic signatures could be quantified and classified, we could use this information to determine subtypes of different cancers, as well as stage of their disease progression[85].

Prognosis and treatment predictions

Prognosis is one important step linking risk assessment and treatment. As mentioned above, characterized and clinically approved cancer biomarkers can be essential to determine the severity of each cancer by providing a proof of concept of an efficient treatment based on the *in vitro* or pharmacological analyses. Such prognostic biomarkers include: Tissue inhibitor of metalloproteinase, a marker characterizing progressive stage of myeloma; estrogen receptor/progesterone receptor, and overexpressed receptor tyrosine-protein kinase erbB-2 (HER2), which could be associated with breast cancer; and c-KIT, a proto-oncogene which could help to identify stromal tumors in the intestine. These molecules are shown not only to identify cancer types but also to specify resistance level to anticancer drugs, which could help to determine efficient treatment to reduce the burden of patients and increase their chance of survival[86].

Follow-up of anticancer treatment

Cancer biomarkers can be used to continuously monitor the efficacy of cancer treatment. As discussed above, these biomarkers can significantly decrease treatment cost and disease burden. The S100 calcium-binding protein B (S100-B) has the potential to follow up treatment response in melanoma. Melanocytes produce pigment in the epidermis, which is associated with high expression of S100-B protein in cancer cells but not in benign cells. Therefore, the anticancer response can be monitored by the decrease in S100-B in the circulation.

In addition, tumor cells undergo apoptosis *via* the release of several intracellular complexes, such as cytochrome c, cytokeratin-18 (cleaved form), nucleosomes, and other molecules. A number of analyses have shown the roles of these molecules in monitoring the cancer progression, metastasis, and eradication, which assist in providing the essential information of treatment[67,68].

Cancer biomarkers used in cancer research

There is an increased focus on biomarkers for their applications in developing cancer drugs. For example, 60 years ago, it was discovered that a major population of chronic myelogenous leukemia patients had specific defects not only in gene expression but also at the level of chromosomes, which

Table 1 Summary of signaling pathways of hematological cancers

Signaling pathway	Biomarkers	Clinical roles as biomarkers	Ref.
Hypoxia signaling	HIF-1 α	Hypoxia strongly correlated clinically to B cell lymphoma and AML proliferation, progression and drug resistance by avoiding oxidative glycolysis but not aerobic glycolysis	[56,57]
Ras/Raf/MAPK	P42/P44 MAPK, P38, P300	Gain-of-function mutation of Kras or Braf (like other kinases) in the tumor cells raise significant resistance to cancer therapy	[58,59]
PI3K-AKT/Ras-ERK/mTOR	Ras-ERK	Mutations affecting kinase activity residues of BRAF (Ras family) and MAPK (ERK family) are correlated to cancer severity, associated to upregulation of these genes in cancer patients as well as increased resistance to conventional chemotherapy; increase of immune checkpoint inhibitors on cancer cells also decreases efficiency of targeted immunotherapy	[60,61]
	mTOR	mTOR constitutive activation is usually found in leukemia patients, which contributes to chemoresistance, disease progression, and unfavorable prognosis	[36-41]
	PI3K-AKT	PTEN, a negative regulator in the PI3K-AKT signaling, becomes inactivated during tumor progression, which deviates the normal signaling and leads to over-reactivation in cancer cells	[62-66]
RTK	Meks/MKKs/ERKs	Mutations that affect RTK signaling often lead to cellular cancerous transformation, and exhibit very limited access to anti-cancer therapy targets	[67,68]
	VEGFR and EGFR	VEGF/VEGFR expression is upregulated in several types of hematolymphoid tumors. It is likely that patients with AML may benefit from EGFR inhibition therapy	[69-76]
NF- κ B	NF- κ B	NF- κ B activity not only promotes tumor cell proliferation, suppresses apoptosis, and attracts angiogenesis but also induces epithelial mesenchymal transition, which facilitates distant metastasis; additionally, it is hardly druggable for specific targeted therapy	[77-80]

Different cellular signaling pathways triggering hematological tumorigenesis, and clinical implications as biomarkers or druggable targets is presented. AML: Acute myeloid leukemia; EGFR: Epidermal growth factor receptor; HIF: Hypoxia-inducible factor; mTOR: Mammalian target of rapamycin; RTK: Receptor tyrosine kinase; NF- κ B: Nuclear factor-kappaB; VEGF: Vascular endothelial growth factor; VEGFR: Vascular endothelial growth factor receptor.

were named Philadelphia chromosomes. If both chromosomes are present in the same patient, a fusion protein called BCR-ABL is expressed, which represents a critical cancer-inducing gene and a key gene that is monitored for the physiological manifestations at the early stage of leukemia. BCR-ABL can still be simply measured to classify the type of leukemia. Owing to these discovered molecules, several targeted inhibitors were developed. For example, imatinib targets and inhibits the function of BCR-ABL protein and drastically diminishes the number of cells with Philadelphia chromosomes[87].

Investigations on surrogate endpoints of the disease are ongoing. Biomarkers may predict the side effects of anti-cancer drugs, and thus, increase survival rate. These markers can prevent patients from undergoing tumor biopsies and shorten the duration of clinical trials' period. The decrease in cancer progression and duration of post-treatment survival could be quantifiable for determining the effect of anticancer drugs. Once biomarker surrogates have been identified, anticancer drug design and validation will become less time-consuming and more cost-effective before entering clinical trials.

New signals from the invasive front

MicroRNA: Studies have been carried out extensively on microRNA (miRNA) and cancer. Profiling of the miRNome (global miRNA expression levels in a certain organism) has become prevalent, and abundant miRNome data are currently available for various cancers[88]. The pattern of miRNA expression can be correlated with a distinct cancer type, stage, and other clinical variables, so miRNA profiling can be used as a biomarker for cancer diagnosis and prognosis. Advanced analyses indicate that miRNAs also play roles in almost all fields of cancer biology, such as angiogenesis, invasion/metastasis, proliferation, and apoptosis. Therefore, an ever-increasing number of studies have identified miRNAs as potential biomarkers for cancer diagnosis, prognosis, and also as therapeutic targets or tools using specific individual miRNAs or clustered groups of miRNAs, which need further investigation and validation in different clinical and research studies[89].

Circulating tumor cells: Surrogate markers, including circulating tumor cells (CTCs) and circulating miRNAs, are getting more attention in recent years[90]. These markers are correlated with the number of tumor cells present in the blood. However, because of the low numbers, CTCs are still very difficult to detect and isolate at a satisfactorily high purity and efficiency. New techniques and research are required for their application into clinical practice[91]. However, the clinical regulation and corresponding medical criteria to define the sensitivity and positivity of interpretation of results remain to be optimized, and this protocol requires equipment dedicated to detecting CTCs and validating the results.

Long noncoding RNA: Long noncoding RNAs (lncRNAs) act as crucial biomarkers in tumors. With the development of molecular biology techniques, lncRNAs have gradually become a research hotspot in the field of tumor research[92]. lncRNAs comprise RNA molecules with sizes greater than 200

nucleotides and do not encode any protein. lncRNAs have been demonstrated to regulate various biological activities and processes, such as epigenetics, cell cycle, and regulation of cell differentiation [93]. Different lncRNAs profiles have been observed in various types of tumors compared with normal tissues, and lncRNAs with dysregulated expression can be tumor-promoting or tumor-suppressing factors. lncRNAs have multidimensional regulatory mechanisms, such as activating/repressing the expression of neighboring genes, encoding the upstream promoter of a protein-coding gene, mediating chromatin modifying, binding to transcription factors and specific proteins, regulating post-transcriptional mRNA decay, and acting as sponges of miRNAs. Different lncRNAs have been detected and shown to be significant for measuring development of non-small cell lung cancer, CRC, gastric cancer, lung squamous cell carcinoma, and breast cancer[94].

Exosomes: Exosomes are extracellular vesicles that have pleiotropic functions in living organisms. These 50-nm to 140-nm nanoparticles transport various materials, including DNA, RNA, proteins, and lipids. Exosomes were first identified as recycled fractions of intravesicular membranes released by reticulocytes following endocytosis of the transferrin receptor. Reticulocytes undergo drastic alterations in cell size, shape, and deformability during maturation. Toward the last stage of this process, transferrin receptors are exocytosed with the help of multivesicular bodies (MVB) that carry 50-nm small vesicles or exosomes[95]. These bilipid-layered vesicles carry a certain number of molecules from the plasma membrane and the interior of the cytoplasm. The release of exosomes occurs when MVBs fuse with the plasma membrane. MVBs are now well known as intracellular endosomal organelles dispersed in the cytoplasm. Exosomes in cancer biology have attracted a lot of attention for their role in the development of the tumor microenvironment. Exosomes play a role in creating a premetastatic niche conducive to metastasis at distant sites. These exosome-delivered tolerogenic signals to cancer-specific immune cells could therefore interrupt immune cell proliferation and induce apoptosis of activated CD8⁺ T lymphocytes, interfering with monocyte differentiation and negatively favoring the expansion of regulatory T cells, leading to immunosuppression (peripheral tolerance) through a paracrine effect. Chen *et al*[96] recently showed that programmed death ligand 1 (PD-L1) expression is enhanced when melanoma cells are exposed to IFN- γ , resulting in increased PD-L1 expression on circulating human-derived melanoma exosomes[97].

Also, EGFR and other metabolic reprogramming using miRNA regulation (*e.g.*, miR-155 and miR-210)[98] show a reverse Warburg effect that contributes to tumor-specific CD8⁺ T-cell inhibition. Therefore, the exosome-derived tumor microenvironment not only creates a favorable immediate layer but a macrostructure to facilitate the metastatic process[99].

Tumor exosome secretion is suggested to participate in promoting cancer cell invasiveness. For example, exosomes derived from pancreatic ductal adenocarcinoma (PDAC) can induce niche formation of premetastatic hepatocytes. This pathological niche formation in a murine model increases the burden of hepatocyte metastasis. Uptake of PDAC-derived exosomes by Kupffer cells could induce a high level of transforming growth factor- β secretion and greater production of fibronectin by stromal cells in the liver[100]. Apparently, this fiber-like microenvironment enhances the recruitment of type 1 macrophages derived from bone marrow. The extracellular matrix is clearly greater in exosomes derived from patients with early-stage PDAC compared with later stages. When MIF expression in the PDAC-derived exosomes was blocked, the formation of the pre-metastatic niche in the liver and subsequent tumor metastasis is also prevented. These observations suggest that the molecules expressed in the tumor exosomes help to prime or target the tumor tissue to become metastatic and resistant to chemotherapy. Therefore, these exosomes could act as a prognostic biomarker to monitor PDAC progression and liver metastasis development and progression[101]; however, precision therapy is still far from being designed, due to the unstable expression level and isoforms of materials presented within the exosomes[102].

CONCLUSION

A more profound understanding of the cellular and molecular mechanisms of cancer and malignant disorders has translated into a biologically adapted classification providing evidence for therapeutic development. Imatinib, a BCR-BRL inhibitor, has been shown to dramatically improve B cell lymphoma patients' prognosis, and it has been established that a significant number of patients defined by older classification systems exhibited poor prognosis, probably due to the biomarkers selected at that time. Cancer therapies, including corticosteroids, IFN- α , chemotherapies, targeted monoclonal antibodies, and small molecule drugs, can elicit significant clinical effects by targeting these biomarkers, with variable durability of response, followed by short- and long-term adverse effects lasting for an undetermined period. Therefore, in the future, a new generation of targeted drugs with better resolution and precision is needed.

For example, Pitson *et al*[103] have found that SK1 is activated by site-directed phosphorylation of ERK-1/2 of S225. Future studies can use anti-phosphor S225 SK1 monoclonal antibodies to determine the impact of phosphorylated SK1 on clinical prognosis[104-106].

It is likely that more biomarkers are continuously being discovered and identified, and new technologies are necessary to measure biomarkers at the time of diagnosis. Ideally, these methods should be high-throughput as well as tissue-sensitive, cost-effective, and rapid[107,108]. However, an additional challenge for molecular and cell biologists, geneticists, and clinical investigators lies in bridging the gap between their worlds and that of biotechnology. Notably, some overlapping technologies have already created unexpected successes in the last decade and are continuously being developed. Essentially, anti-cancer drug discovery frameworks targeted at developing anti-cancer drugs, specifically for metastasis, should be taken into consideration globally. Extensive advanced development of a more sensitive detection method is required.

Limitations of these detection assays include the logistical challenges associated with high-quality results from fresh biopsy specimens in the hospital setting. The ongoing development of these promising techniques of high resolution and detection sensitivity, with the translation to a widespread clinical application from basic science, will also be vitally reviewed and appreciated by all aspects. There are still some examples regarding the technologies under investigation, including high-contrast fluorescence detection, multispectral optoacoustic tomography, shortwave infrared emitting nanoprobes, and novel magnetic resonance imaging that is non-toxic and applied with highly permissive contrast agents. In addition, it is important to detect micro-metastasis in the initial stage of development in cancers of different types using all the possible approaches, which will guarantee adapted and appropriate treatment regimens in both ongoing and to-be-started clinical trials.

FOOTNOTES

Author contributions: Liu SX and Mai HR contributed equally to this work; Liu SX and Mai HR designed the research study; Tang X, Chen F and Xie LC performed the research; Tang X, Mai HR and Liu SX wrote the manuscript; and all authors have read and approved the final manuscript.

Supported by Sanming Project of Medicine in Shenzhen, No. SZSM201512033; Shenzhen Fund for Guangdong Provincial High-level Clinical Key Specialties, No. SZGSP012; Shenzhen Key Medical Discipline Construction Fund, No. SZXK034; and Shenzhen Healthcare Research Project, No. SZLY2018015.

Conflict-of-interest statement: The authors declare no conflicts of interest.

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <https://creativecommons.org/licenses/by-nc/4.0/>

Country/Territory of origin: China

ORCID number: Xue Tang 0000-0001-7164-7555; Fen Chen 0000-0002-2187-1775; Li-Chun Xie 0000-0002-6183-5904; Si-Xi Liu 0000-0003-1674-2685; Hui-Rong Mai 0000-0002-8970-9221.

S-Editor: Wang JJ

L-Editor: A

P-Editor: Wang JJ

REFERENCES

- Masoud GN, Li W. HIF-1 α pathway: role, regulation and intervention for cancer therapy. *Acta Pharm Sin B* 2015; **5**: 378-389 [PMID: 26579469 DOI: 10.1016/j.apsb.2015.05.007]
- Persi E, Duran-Frigola M, Damaghi M, Roush WR, Aloy P, Cleveland JL, Gillies RJ, Ruppin E. Systems analysis of intracellular pH vulnerabilities for cancer therapy. *Nat Commun* 2018; **9**: 2997 [PMID: 30065243 DOI: 10.1038/s41467-018-05261-x]
- Iommarini L, Porcelli AM, Gasparre G, Kurelac I. Non-Canonical Mechanisms Regulating Hypoxia-Inducible Factor 1 Alpha in Cancer. *Front Oncol* 2017; **7**: 286 [PMID: 29230384 DOI: 10.3389/fonc.2017.00286]
- Palazon A, Goldrath AW, Nizet V, Johnson RS. HIF transcription factors, inflammation, and immunity. *Immunity* 2014; **41**: 518-528 [PMID: 25367569 DOI: 10.1016/j.immuni.2014.09.008]
- Bhoo-Pathy N, Verkooijen HM, Tan EY, Miao H, Taib NA, Brand JS, Dent RA, See MH, Subramaniam S, Chan P, Lee SC, Hartman M, Yip CH. Trends in presentation, management and survival of patients with de novo metastatic breast cancer in a Southeast Asian setting. *Sci Rep* 2015; **5**: 16252 [PMID: 26536962 DOI: 10.1038/srep16252]
- Balamurugan K. HIF-1 at the crossroads of hypoxia, inflammation, and cancer. *Int J Cancer* 2016; **138**: 1058-1066 [PMID: 25784597 DOI: 10.1002/ijc.29519]
- Wu X, Zhou QH, Xu K. Are isothiocyanates potential anti-cancer drugs? *Acta Pharmacol Sin* 2009; **30**: 501-512 [PMID: 19111111 DOI: 10.1007/s12248-009-9111-1]

- 19417730 DOI: [10.1038/aps.2009.50](https://doi.org/10.1038/aps.2009.50)]
- 8 **Zargar P**, Ghani E, Mashayekhi FJ, Ramezani A, Eftekhari E. Acridine enhances the antitumor activity of the chemotherapeutic drug 5-fluorouracil in colorectal cancer cells. *Oncol Lett* 2018; **15**: 10084-10090 [PMID: [29928378](https://pubmed.ncbi.nlm.nih.gov/29928378/) DOI: [10.3892/ol.2018.8569](https://doi.org/10.3892/ol.2018.8569)]
 - 9 **Ardö Y**. Flavour formation by amino acid catabolism. *Biotechnol Adv* 2006; **24**: 238-242 [PMID: [16406465](https://pubmed.ncbi.nlm.nih.gov/16406465/) DOI: [10.1016/j.biotechadv.2005.11.005](https://doi.org/10.1016/j.biotechadv.2005.11.005)]
 - 10 **Strowitzki MJ**, Cummins EP, Taylor CT. Protein Hydroxylation by Hypoxia-Inducible Factor (HIF) Hydroxylases: Unique or Ubiquitous? *Cells* 2019; **8** [PMID: [31035491](https://pubmed.ncbi.nlm.nih.gov/31035491/) DOI: [10.3390/cells8050384](https://doi.org/10.3390/cells8050384)]
 - 11 **Kibel A**, Iliopoulos O, DeCaprio JA, Kaelin WG Jr. Binding of the von Hippel-Lindau tumor suppressor protein to Elongin B and C. *Science* 1995; **269**: 1444-1446 [PMID: [7660130](https://pubmed.ncbi.nlm.nih.gov/7660130/) DOI: [10.1126/science.7660130](https://doi.org/10.1126/science.7660130)]
 - 12 **Kong X**, Alvarez-Castelao B, Lin Z, Castaño JG, Caro J. Constitutive/hypoxic degradation of HIF- α proteins by the proteasome is independent of von Hippel Lindau protein ubiquitylation and the transactivation activity of the protein. *J Biol Chem* 2007; **282**: 15498-15505 [PMID: [17403672](https://pubmed.ncbi.nlm.nih.gov/17403672/) DOI: [10.1074/jbc.M700704200](https://doi.org/10.1074/jbc.M700704200)]
 - 13 **Freedman SJ**, Sun ZY, Poy F, Kung AL, Livingston DM, Wagner G, Eck MJ. Structural basis for recruitment of CBP/p300 by hypoxia-inducible factor-1 α . *Proc Natl Acad Sci U S A* 2002; **99**: 5367-5372 [PMID: [11959990](https://pubmed.ncbi.nlm.nih.gov/11959990/) DOI: [10.1073/pnas.082117899](https://doi.org/10.1073/pnas.082117899)]
 - 14 **Nieminen AL**, Qanungo S, Schneider EA, Jiang BH, Agani FH. Mdm2 and HIF-1 α interaction in tumor cells during hypoxia. *J Cell Physiol* 2005; **204**: 364-369 [PMID: [15880652](https://pubmed.ncbi.nlm.nih.gov/15880652/) DOI: [10.1002/jcp.20406](https://doi.org/10.1002/jcp.20406)]
 - 15 **Ramanathan M**, Luo W, Csóka B, Haskó G, Lukashev D, Sitkovsky MV, Leibovich SJ. Differential regulation of HIF-1 α isoforms in murine macrophages by TLR4 and adenosine A(2A) receptor agonists. *J Leukoc Biol* 2009; **86**: 681-689 [PMID: [19477908](https://pubmed.ncbi.nlm.nih.gov/19477908/) DOI: [10.1189/jlb.0109021](https://doi.org/10.1189/jlb.0109021)]
 - 16 **Masson N**, Ratcliffe PJ. Hypoxia signaling pathways in cancer metabolism: the importance of co-selecting interconnected physiological pathways. *Cancer Metab* 2014; **2**: 3 [PMID: [24491179](https://pubmed.ncbi.nlm.nih.gov/24491179/) DOI: [10.1186/2049-3002-2-3](https://doi.org/10.1186/2049-3002-2-3)]
 - 17 **Mendoza MC**, Er EE, Blenis J. The Ras-ERK and PI3K-mTOR pathways: cross-talk and compensation. *Trends Biochem Sci* 2011; **36**: 320-328 [PMID: [21531565](https://pubmed.ncbi.nlm.nih.gov/21531565/) DOI: [10.1016/j.tibs.2011.03.006](https://doi.org/10.1016/j.tibs.2011.03.006)]
 - 18 **Laplanche M**, Sabatini DM. mTOR signaling in growth control and disease. *Cell* 2012; **149**: 274-293 [PMID: [22500797](https://pubmed.ncbi.nlm.nih.gov/22500797/) DOI: [10.1016/j.cell.2012.03.017](https://doi.org/10.1016/j.cell.2012.03.017)]
 - 19 **Sears R**, Gray JW. Epigenomic Inactivation of RasGAPs Activates RAS Signaling in a Subset of Luminal B Breast Cancers. *Cancer Discov* 2017; **7**: 131-133 [PMID: [28167613](https://pubmed.ncbi.nlm.nih.gov/28167613/) DOI: [10.1158/2159-8290.CD-16-1423](https://doi.org/10.1158/2159-8290.CD-16-1423)]
 - 20 **Chen Y**, Zhou C, Ji W, Mei Z, Hu B, Zhang W, Zhang D, Wang J, Liu X, Ouyang G, Zhou J, Xiao W. ELL targets c-Myc for proteasomal degradation and suppresses tumour growth. *Nat Commun* 2016; **7**: 11057 [PMID: [27009366](https://pubmed.ncbi.nlm.nih.gov/27009366/) DOI: [10.1038/ncomms11057](https://doi.org/10.1038/ncomms11057)]
 - 21 **De Luca A**, Maiello MR, D'Alessio A, Pergameno M, Normanno N. The RAS/RAF/MEK/ERK and the PI3K/AKT signalling pathways: role in cancer pathogenesis and implications for therapeutic approaches. *Expert Opin Ther Targets* 2012; **16** Suppl 2: S17-S27 [PMID: [22443084](https://pubmed.ncbi.nlm.nih.gov/22443084/) DOI: [10.1517/14728222.2011.639361](https://doi.org/10.1517/14728222.2011.639361)]
 - 22 **McCubrey JA**, Steelman LS, Chappell WH, Abrams SL, Montalto G, Cervello M, Nicoletti F, Fagone P, Malaponte G, Mazzerino MC, Candido S, Libra M, Bäscke J, Mijatovic S, Maksimovic-Ivanic D, Milella M, Tafuri A, Cocco L, Evangelisti C, Chiarini F, Martelli AM. Mutations and deregulation of Ras/Raf/MEK/ERK and PI3K/Pten/Akt/mTOR cascades which alter therapy response. *Oncotarget* 2012; **3**: 954-987 [PMID: [23006971](https://pubmed.ncbi.nlm.nih.gov/23006971/) DOI: [10.18632/oncotarget.652](https://doi.org/10.18632/oncotarget.652)]
 - 23 **Boehmelt M**, Madruga J, Dörfler P, Briegel K, Schwarz H, Enrietto PJ, Zenke M. Dendritic cell progenitor is transformed by a conditional v-Rel estrogen receptor fusion protein v-RelER. *Cell* 1995; **80**: 341-352 [PMID: [7834754](https://pubmed.ncbi.nlm.nih.gov/7834754/) DOI: [10.1016/0092-8674\(95\)90417-4](https://doi.org/10.1016/0092-8674(95)90417-4)]
 - 24 **Oeckinghaus A**, Ghosh S. The NF- κ B family of transcription factors and its regulation. *Cold Spring Harb Perspect Biol* 2009; **1**: a000034 [PMID: [20066092](https://pubmed.ncbi.nlm.nih.gov/20066092/) DOI: [10.1101/cshperspect.a000034](https://doi.org/10.1101/cshperspect.a000034)]
 - 25 **Sun SC**, Cesarman E. NF- κ B as a target for oncogenic viruses. *Curr Top Microbiol Immunol* 2011; **349**: 197-244 [PMID: [20845110](https://pubmed.ncbi.nlm.nih.gov/20845110/) DOI: [10.1007/82_2010_108](https://doi.org/10.1007/82_2010_108)]
 - 26 **Yim EK**, Park JS. The role of HPV E6 and E7 oncoproteins in HPV-associated cervical carcinogenesis. *Cancer Res Treat* 2005; **37**: 319-324 [PMID: [19956366](https://pubmed.ncbi.nlm.nih.gov/19956366/) DOI: [10.4143/crt.2005.37.6.319](https://doi.org/10.4143/crt.2005.37.6.319)]
 - 27 **Park MH**, Hong JT. Roles of NF- κ B in Cancer and Inflammatory Diseases and Their Therapeutic Approaches. *Cells* 2016; **5** [PMID: [27043634](https://pubmed.ncbi.nlm.nih.gov/27043634/) DOI: [10.3390/cells5020015](https://doi.org/10.3390/cells5020015)]
 - 28 **Osellame LD**, Blacker TS, Duchon MR. Cellular and molecular mechanisms of mitochondrial function. *Best Pract Res Clin Endocrinol Metab* 2012; **26**: 711-723 [PMID: [23168274](https://pubmed.ncbi.nlm.nih.gov/23168274/) DOI: [10.1016/j.beem.2012.05.003](https://doi.org/10.1016/j.beem.2012.05.003)]
 - 29 **Araujo L**, Khim P, Mkhikian H, Mortales CL, Demetriou M. Glycolysis and glutaminolysis cooperatively control T cell function by limiting metabolite supply to N-glycosylation. *Elife* 2017; **6** [PMID: [28059703](https://pubmed.ncbi.nlm.nih.gov/28059703/) DOI: [10.7554/eLife.21330](https://doi.org/10.7554/eLife.21330)]
 - 30 **Zhu A**, Lee D, Shim H. Metabolic positron emission tomography imaging in cancer detection and therapy response. *Semin Oncol* 2011; **38**: 55-69 [PMID: [21362516](https://pubmed.ncbi.nlm.nih.gov/21362516/) DOI: [10.1053/j.seminoncol.2010.11.012](https://doi.org/10.1053/j.seminoncol.2010.11.012)]
 - 31 **Bhat TA**, Kumar S, Chaudhary AK, Yadav N, Chandra D. Restoration of mitochondria function as a target for cancer therapy. *Drug Discov Today* 2015; **20**: 635-643 [PMID: [25766095](https://pubmed.ncbi.nlm.nih.gov/25766095/) DOI: [10.1016/j.drudis.2015.03.001](https://doi.org/10.1016/j.drudis.2015.03.001)]
 - 32 **Harami-Papp H**, Pongor LS, Munkácsy G, Horváth G, Nagy ÁM, Ambrus A, Hauser P, Szabó A, Tretter L, Györfy B. TP53 mutation hits energy metabolism and increases glycolysis in breast cancer. *Oncotarget* 2016; **7**: 67183-67195 [PMID: [27582538](https://pubmed.ncbi.nlm.nih.gov/27582538/) DOI: [10.18632/oncotarget.11594](https://doi.org/10.18632/oncotarget.11594)]
 - 33 **Senyilmaz D**, Teleman AA. Chicken or the egg: Warburg effect and mitochondrial dysfunction. *F1000Prime Rep* 2015; **7**: 41 [PMID: [26097714](https://pubmed.ncbi.nlm.nih.gov/26097714/) DOI: [10.12703/P7-41](https://doi.org/10.12703/P7-41)]
 - 34 **Pui CH**, Robison LL, Look AT. Acute lymphoblastic leukaemia. *Lancet* 2008; **371**: 1030-1043 [PMID: [18358930](https://pubmed.ncbi.nlm.nih.gov/18358930/) DOI: [10.1016/S0140-6736\(08\)60457-2](https://doi.org/10.1016/S0140-6736(08)60457-2)]
 - 35 **Janku F**. Phosphoinositide 3-kinase (PI3K) pathway inhibitors in solid tumors: From laboratory to patients. *Cancer Treat Rev* 2017; **59**: 93-101 [PMID: [28779636](https://pubmed.ncbi.nlm.nih.gov/28779636/) DOI: [10.1016/j.ctrv.2017.07.005](https://doi.org/10.1016/j.ctrv.2017.07.005)]
 - 36 **Bertacchini J**, Heidari N, Mediani L, Capitani S, Shahjehani M, Ahmadzadeh A, Saki N. Targeting PI3K/AKT/mTOR

- network for treatment of leukemia. *Cell Mol Life Sci* 2015; **72**: 2337-2347 [PMID: [25712020](#) DOI: [10.1007/s00018-015-1867-5](#)]
- 37 **Fang Y**, Yang Y, Hua C, Xu S, Zhou M, Guo H, Wang N, Zhao X, Huang L, Yu F, Cheng H, Wang ML, Meng L, Cheng T, Yuan W, Ma D, Zhou J. Rictor has a pivotal role in maintaining quiescence as well as stemness of leukemia stem cells in MLL-driven leukemia. *Leukemia* 2017; **31**: 414-422 [PMID: [27499138](#) DOI: [10.1038/leu.2016.223](#)]
 - 38 **Evangelisti C**, Chiarini F, McCubrey JA, Martelli AM. Therapeutic Targeting of mTOR in T-Cell Acute Lymphoblastic Leukemia: An Update. *Int J Mol Sci* 2018; **19** [PMID: [29949919](#) DOI: [10.3390/ijms19071878](#)]
 - 39 **Janku F**, Yap TA, Meric-Bernstam F. Targeting the PI3K pathway in cancer: are we making headway? *Nat Rev Clin Oncol* 2018; **15**: 273-291 [PMID: [29508857](#) DOI: [10.1038/nrclinonc.2018.28](#)]
 - 40 **Hoshii T**, Tadokoro Y, Naka K, Ooshio T, Muraguchi T, Sugiyama N, Soga T, Araki K, Yamamura K, Hirao A. mTORC1 is essential for leukemia propagation but not stem cell self-renewal. *J Clin Invest* 2012; **122**: 2114-2129 [PMID: [22622041](#) DOI: [10.1172/JCI62279](#)]
 - 41 **Willems L**, Tamburini J, Chapuis N, Lacombe C, Mayeux P, Bouscary D. PI3K and mTOR signaling pathways in cancer: new data on targeted therapies. *Curr Oncol Rep* 2012; **14**: 129-138 [PMID: [22350330](#) DOI: [10.1007/s11912-012-0227-y](#)]
 - 42 **Lopez-Guerra M**, Colomer D. NF-kappaB as a therapeutic target in chronic lymphocytic leukemia. *Expert Opin Ther Targets* 2010; **14**: 275-288 [PMID: [20148715](#) DOI: [10.1517/14728221003598930](#)]
 - 43 **Vilimas T**, Mascarenhas J, Palomero T, Mandal M, Buonamici S, Meng F, Thompson B, Spaulding C, Macaroun S, Alegre ML, Kee BL, Ferrando A, Miele L, Aifantis I. Targeting the NF-kappaB signaling pathway in Notch1-induced T-cell leukemia. *Nat Med* 2007; **13**: 70-77 [PMID: [17173050](#) DOI: [10.1038/nm1524](#)]
 - 44 **Zhou J**, Ching YQ, Chng WJ. Aberrant nuclear factor-kappa B activity in acute myeloid leukemia: from molecular pathogenesis to therapeutic target. *Oncotarget* 2015; **6**: 5490-5500 [PMID: [25823927](#) DOI: [10.18632/oncotarget.3545](#)]
 - 45 **Shanbhag S**, Ambinder RF. Hodgkin lymphoma: A review and update on recent progress. *CA Cancer J Clin* 2018; **68**: 116-132 [PMID: [29194581](#) DOI: [10.3322/caac.21438](#)]
 - 46 **Schatz JH**. Targeting the PI3K/AKT/mTOR pathway in non-Hodgkin's lymphoma: results, biology, and development strategies. *Curr Oncol Rep* 2011; **13**: 398-406 [PMID: [21755275](#) DOI: [10.1007/s11912-011-0187-7](#)]
 - 47 **Johnston PB**, Inwards DJ, Colgan JP, Laplant BR, Kabat BF, Habermann TM, Micallef IN, Porrata LF, Ansell SM, Reeder CB, Roy V, Witzig TE. A Phase II trial of the oral mTOR inhibitor everolimus in relapsed Hodgkin lymphoma. *Am J Hematol* 2010; **85**: 320-324 [PMID: [20229590](#) DOI: [10.1002/ajh.21664](#)]
 - 48 **Weniger MA**, Küppers R. NF-κB deregulation in Hodgkin lymphoma. *Semin Cancer Biol* 2016; **39**: 32-39 [PMID: [27221964](#) DOI: [10.1016/j.semcancer.2016.05.001](#)]
 - 49 **Tong J**, Yu Q, Xu W, Yu W, Wu C, Wu Y, Yan H. Montelukast enhances cytotoxic effects of carfilzomib in multiple myeloma by inhibiting mTOR pathway. *Cancer Biol Ther* 2019; **20**: 381-390 [PMID: [30359543](#) DOI: [10.1080/15384047.2018.1529112](#)]
 - 50 **Roy P**, Sarkar UA, Basak S. The NF-κB Activating Pathways in Multiple Myeloma. *Biomedicines* 2018; **6** [PMID: [29772694](#) DOI: [10.3390/biomedicines6020059](#)]
 - 51 **Xu Q**, Thompson JE, Carroll M. mTOR regulates cell survival after etoposide treatment in primary AML cells. *Blood* 2005; **106**: 4261-4268 [PMID: [16150937](#) DOI: [10.1182/blood-2004-11-4468](#)]
 - 52 **Altman JK**, Sassano A, Kaur S, Glaser H, Kroczyńska B, Redig AJ, Russo S, Barr S, Platanias LC. Dual mTORC2/mTORC1 targeting results in potent suppressive effects on acute myeloid leukemia (AML) progenitors. *Clin Cancer Res* 2011; **17**: 4378-4388 [PMID: [21415215](#) DOI: [10.1158/1078-0432.CCR-10-2285](#)]
 - 53 **Chapuis N**, Tamburini J, Green AS, Vignon C, Bardet Y, Neyret A, Pannetier M, Willems L, Park S, Maccone A, Maira SM, Ifrah N, Dreyfus F, Herauld O, Lacombe C, Mayeux P, Bouscary D. Dual inhibition of PI3K and mTORC1/2 signaling by NVP-BEZ235 as a new therapeutic strategy for acute myeloid leukemia. *Clin Cancer Res* 2010; **16**: 5424-5435 [PMID: [20884625](#) DOI: [10.1158/1078-0432.CCR-10-1102](#)]
 - 54 **Bertacchini J**, Guida M, Accordi B, Mediani L, Martelli AM, Barozzi P, Petricoin E 3rd, Liotta L, Milani G, Giordan M, Luppi M, Forghieri F, De Pol A, Cocco L, Basso G, Marmiroli S. Feedbacks and adaptive capabilities of the PI3K/Akt/mTOR axis in acute myeloid leukemia revealed by pathway selective inhibition and phosphoproteome analysis. *Leukemia* 2014; **28**: 2197-2205 [PMID: [24699302](#) DOI: [10.1038/leu.2014.123](#)]
 - 55 **Grupp PA**, Boylan JM, Sanders JA. The physiology and pathophysiology of rapamycin resistance: implications for cancer. *Cell Cycle* 2011; **10**: 1050-1058 [PMID: [21389767](#) DOI: [10.4161/cc.10.7.15230](#)]
 - 56 **Wang Y**, Liu Y, Malek SN, Zheng P. Targeting HIF1α eliminates cancer stem cells in hematological malignancies. *Cell Stem Cell* 2011; **8**: 399-411 [PMID: [21474104](#) DOI: [10.1016/j.stem.2011.02.006](#)]
 - 57 **Matolay O**, Méhes G. Sustain, Adapt, and Overcome-Hypoxia Associated Changes in the Progression of Lymphatic Neoplasia. *Front Oncol* 2019; **9**: 1277 [PMID: [31824854](#) DOI: [10.3389/fonc.2019.01277](#)]
 - 58 **Wang AX**, Qi XY. Targeting RAS/RAF/MEK/ERK signaling in metastatic melanoma. *IUBMB Life* 2013; **65**: 748-758 [PMID: [23893853](#) DOI: [10.1002/iub.1193](#)]
 - 59 **Santarpia L**, Lippman SM, El-Naggar AK. Targeting the MAPK-RAS-RAF signaling pathway in cancer therapy. *Expert Opin Ther Targets* 2012; **16**: 103-119 [PMID: [22239440](#) DOI: [10.1517/14728222.2011.645805](#)]
 - 60 **Steelman LS**, Franklin RA, Abrams SL, Chappell W, Kempf CR, Bäsecke J, Stivala F, Donia M, Fagone P, Nicoletti F, Libra M, Ruvolo P, Ruvolo V, Evangelisti C, Martelli AM, McCubrey JA. Roles of the Ras/Raf/MEK/ERK pathway in leukemia therapy. *Leukemia* 2011; **25**: 1080-1094 [PMID: [21494257](#) DOI: [10.1038/leu.2011.66](#)]
 - 61 **Giménez N**, Martínez-Trillos A, Montraveta A, Lopez-Guerra M, Rosich L, Nadeu F, Valero JG, Aymerich M, Magnano L, Rozman M, Matutes E, Delgado J, Baumann T, Gine E, González M, Alcoceba M, Terol MJ, Navarro B, Colado E, Payer AR, Puente XS, López-Otín C, Lopez-Guillermo A, Campo E, Colomer D, Villamor N. Mutations in the RAS-BRAF-MAPK-ERK pathway define a specific subgroup of patients with adverse clinical features and provide new therapeutic options in chronic lymphocytic leukemia. *Haematologica* 2019; **104**: 576-586 [PMID: [30262568](#) DOI: [10.3324/haematol.2018.196931](#)]
 - 62 **Roszak J**, Smok-Pieniążek A, Stępnik M. Transcriptomic analysis of the PI3K/Akt signaling pathway reveals the dual

- role of the c-Jun oncogene in cytotoxicity and the development of resistance in HL-60 leukemia cells in response to arsenic trioxide. *Adv Clin Exp Med* 2017; **26**: 1335-1342 [PMID: [29442453](#) DOI: [10.17219/acem/65475](#)]
- 63 **Wu Y**, Hu Y, Yu X, Zhang Y, Huang X, Chen S, Li Y, Zeng C. TAL1 mediates imatinib-induced CML cell apoptosis via the PTEN/PI3K/AKT pathway. *Biochem Biophys Res Commun* 2019; **519**: 234-239 [PMID: [31493871](#) DOI: [10.1016/j.bbrc.2019.08.164](#)]
 - 64 **Marshall JDS**, Whitecross DE, Mellor P, Anderson DH. Impact of p85α Alterations in Cancer. *Biomolecules* 2019; **9** [PMID: [30650664](#) DOI: [10.3390/biom9010029](#)]
 - 65 **Fayard E**, Moncayo G, Hemmings BA, Holländer GA. Phosphatidylinositol 3-kinase signaling in thymocytes: the need for stringent control. *Sci Signal* 2010; **3**: re5 [PMID: [20716765](#) DOI: [10.1126/scisignal.3135re5](#)]
 - 66 **Jiang N**, Dai Q, Su X, Fu J, Feng X, Peng J. Role of PI3K/AKT pathway in cancer: the framework of malignant behavior. *Mol Biol Rep* 2020; **47**: 4587-4629 [PMID: [32333246](#) DOI: [10.1007/s11033-020-05435-1](#)]
 - 67 **Regad T**. Targeting RTK Signaling Pathways in Cancer. *Cancers (Basel)* 2015; **7**: 1758-1784 [PMID: [26404379](#) DOI: [10.3390/cancers7030860](#)]
 - 68 **Han B**, Wei W, Hua F, Cao T, Dong H, Yang T, Yang Y, Pan H, Xu C. Requirement for ERK activity in sodium selenite-induced apoptosis of acute promyelocytic leukemia-derived NB4 cells. *J Biochem Mol Biol* 2007; **40**: 196-204 [PMID: [17394769](#) DOI: [10.5483/BMBRep.2007.40.2.196](#)]
 - 69 **Iqbal N**, Iqbal N. Human Epidermal Growth Factor Receptor 2 (HER2) in Cancers: Overexpression and Therapeutic Implications. *Mol Biol Int* 2014; **2014**: 852748 [PMID: [25276427](#) DOI: [10.1155/2014/852748](#)]
 - 70 **Gazdar AF**. Activating and resistance mutations of EGFR in non-small-cell lung cancer: role in clinical response to EGFR tyrosine kinase inhibitors. *Oncogene* 2009; **28** Suppl 1: S24-S31 [PMID: [19680293](#) DOI: [10.1038/onc.2009.198](#)]
 - 71 **Koch S**, Claesson-Welsh L. Signal transduction by vascular endothelial growth factor receptors. *Cold Spring Harb Perspect Med* 2012; **2**: a006502 [PMID: [22762016](#) DOI: [10.1101/cshperspect.a006502](#)]
 - 72 **Ding W**, Knox TR, Tschumper RC, Wu W, Schwager SM, Boysen JC, Jelinek DF, Kay NE. Platelet-derived growth factor (PDGF)-PDGF receptor interaction activates bone marrow-derived mesenchymal stromal cells derived from chronic lymphocytic leukemia: implications for an angiogenic switch. *Blood* 2010; **116**: 2984-2993 [PMID: [20606160](#) DOI: [10.1182/blood-2010-02-269894](#)]
 - 73 **Song G**, Li Y, Jiang G. Role of VEGF/VEGFR in the pathogenesis of leukemias and as treatment targets (Review). *Oncol Rep* 2012; **28**: 1935-1944 [PMID: [22993103](#) DOI: [10.3892/or.2012.2045](#)]
 - 74 **Dias S**, Hattori K, Zhu Z, Heissig B, Choy M, Lane W, Wu Y, Chadburn A, Hyjek E, Gill M, Hicklin DJ, Witte L, Moore MA, Rafii S. Autocrine stimulation of VEGFR-2 activates human leukemic cell growth and migration. *J Clin Invest* 2000; **106**: 511-521 [PMID: [10953026](#) DOI: [10.1172/JCI18978](#)]
 - 75 **Mahmud H**, Kornblau SM, Ter Elst A, Scherpen FJ, Qiu YH, Coombes KR, de Bont ES. Epidermal growth factor receptor is expressed and active in a subset of acute myeloid leukemia. *J Hematol Oncol* 2016; **9**: 64 [PMID: [27488458](#) DOI: [10.1186/s13045-016-0294-x](#)]
 - 76 **Lainey E**, Wolfromm A, Sukkurwala AQ, Micol JB, Fenaux P, Galluzzi L, Kepp O, Kroemer G. EGFR inhibitors exacerbate differentiation and cell cycle arrest induced by retinoic acid and vitamin D3 in acute myeloid leukemia cells. *Cell Cycle* 2013; **12**: 2978-2991 [PMID: [23974111](#) DOI: [10.4161/cc.26016](#)]
 - 77 **Xiu Y**, Dong Q, Li Q, Li F, Borchering N, Zhang W, Boyce B, Xue HH, Zhao C. Stabilization of NF-κB-Inducing Kinase Suppresses MLL-AF9-Induced Acute Myeloid Leukemia. *Cell Rep* 2018; **22**: 350-358 [PMID: [29320732](#) DOI: [10.1016/j.celrep.2017.12.055](#)]
 - 78 **Braun T**, Carvalho G, Fabre C, Grosjean J, Fenaux P, Kroemer G. Targeting NF-kappaB in hematologic malignancies. *Cell Death Differ* 2006; **13**: 748-758 [PMID: [16498458](#) DOI: [10.1038/sj.cdd.4401874](#)]
 - 79 **Mansouri L**, Papakonstantinou N, Ntoufa S, Stamatopoulos K, Rosenquist R. NF-κB activation in chronic lymphocytic leukemia: A point of convergence of external triggers and intrinsic lesions. *Semin Cancer Biol* 2016; **39**: 40-48 [PMID: [27491692](#) DOI: [10.1016/j.semcancer.2016.07.005](#)]
 - 80 **Xia Y**, Shen S, Verma IM. NF-κB, an active player in human cancers. *Cancer Immunol Res* 2014; **2**: 823-830 [PMID: [25187272](#) DOI: [10.1158/2326-6066.CIR-14-0112](#)]
 - 81 **Godwin P**, Baird AM, Heavey S, Barr MP, O'Byrne KJ, Gately K. Targeting nuclear factor-kappa B to overcome resistance to chemotherapy. *Front Oncol* 2013; **3**: 120 [PMID: [23720710](#) DOI: [10.3389/fonc.2013.00120](#)]
 - 82 **Richardson PG**, Mitsiades C, Schlossman R, Munshi N, Anderson K. New drugs for myeloma. *Oncologist* 2007; **12**: 664-689 [PMID: [17602058](#) DOI: [10.1634/theoncologist.12-6-664](#)]
 - 83 **Wu ZH**, Shi Y. When ubiquitin meets NF-κB: a trove for anti-cancer drug development. *Curr Pharm Des* 2013; **19**: 3263-3275 [PMID: [23151140](#) DOI: [10.2174/1381612811319180010](#)]
 - 84 **Herceg Z**, Hainaut P. Genetic and epigenetic alterations as biomarkers for cancer detection, diagnosis and prognosis. *Mol Oncol* 2007; **1**: 26-41 [PMID: [19383285](#) DOI: [10.1016/j.molonc.2007.01.004](#)]
 - 85 **Grade M**, Difilippantonio MJ, Camps J. Patterns of Chromosomal Aberrations in Solid Tumors. *Recent Results Cancer Res* 2015; **200**: 115-142 [PMID: [26376875](#) DOI: [10.1007/978-3-319-20291-4_6](#)]
 - 86 **Lasota J**, Jasinski M, Sarlomo-Rikala M, Miettinen M. Mutations in exon 11 of c-Kit occur preferentially in malignant versus benign gastrointestinal stromal tumors and do not occur in leiomyomas or leiomyosarcomas. *Am J Pathol* 1999; **154**: 53-60 [PMID: [9916918](#) DOI: [10.1016/S0002-9440\(10\)65250-9](#)]
 - 87 **O'Brien SG**, Guilhot F, Larson RA, Gathmann I, Baccarani M, Cervantes F, Cornelissen JJ, Fischer T, Hochhaus A, Hughes T, Lechner K, Nielsen JL, Rousselot P, Reiffers J, Saglio G, Shepherd J, Simonsson B, Gratwohl A, Goldman JM, Kantarjian H, Taylor K, Verhoef G, Bolton AE, Capdeville R, Druker BJ, IRIS Investigators. Imatinib compared with interferon and low-dose cytarabine for newly diagnosed chronic-phase chronic myeloid leukemia. *N Engl J Med* 2003; **348**: 994-1004 [PMID: [12637609](#) DOI: [10.1056/NEJMoa022457](#)]
 - 88 **Lee YS**, Dutta A. MicroRNAs in cancer. *Annu Rev Pathol* 2009; **4**: 199-227 [PMID: [18817506](#) DOI: [10.1146/annurev.pathol.4.110807.092222](#)]
 - 89 **Peng Y**, Croce CM. The role of MicroRNAs in human cancer. *Signal Transduct Target Ther* 2016; **1**: 15004 [PMID: [29263891](#) DOI: [10.1038/sigtrans.2015.4](#)]

- 90 **Lianidou ES**, Markou A, Strati A. The Role of CTCs as Tumor Biomarkers. *Adv Exp Med Biol* 2015; **867**: 341-367 [PMID: [26530376](#) DOI: [10.1007/978-94-017-7215-0_21](#)]
- 91 **Chinen LTD**, Abdallah EA, Braun AC, Flores BCTCP, Corassa M, Sanches SM, Fanelli MF. Circulating Tumor Cells as Cancer Biomarkers in the Clinic. *Adv Exp Med Biol* 2017; **994**: 1-41 [PMID: [28560666](#) DOI: [10.1007/978-3-319-55947-6_1](#)]
- 92 **Renganathan A**, Felley-Bosco E. Long Noncoding RNAs in Cancer and Therapeutic Potential. *Adv Exp Med Biol* 2017; **1008**: 199-222 [PMID: [28815541](#) DOI: [10.1007/978-981-10-5203-3_7](#)]
- 93 **Li Y**, Yang X, Kang X, Liu S. The regulatory roles of long noncoding RNAs in the biological behavior of pancreatic cancer. *Saudi J Gastroenterol* 2019; **25**: 145-151 [PMID: [30720003](#) DOI: [10.4103/sjg.SJG_465_18](#)]
- 94 **Zhang X**, Wang W, Zhu W, Dong J, Cheng Y, Yin Z, Shen F. Mechanisms and Functions of Long Non-Coding RNAs at Multiple Regulatory Levels. *Int J Mol Sci* 2019; **20** [PMID: [31717266](#) DOI: [10.3390/ijms20225573](#)]
- 95 **Rout ED**, Webb TL, Laurence HM, Long L, Olver CS. Transferrin receptor expression in serum exosomes as a marker of regenerative anaemia in the horse. *Equine Vet J* 2015; **47**: 101-106 [PMID: [24708277](#) DOI: [10.1111/evj.12235](#)]
- 96 **Chen S**, Crabill GA, Pritchard TS, McMiller TL, Wei P, Pardoll DM, Pan F, Topalian SL. Mechanisms regulating PD-L1 expression on tumor and immune cells. *J Immunother Cancer* 2019; **7**: 305 [PMID: [31730010](#) DOI: [10.1186/s40425-019-0770-2](#)]
- 97 **Tai YL**, Chen KC, Hsieh JT, Shen TL. Exosomes in cancer development and clinical applications. *Cancer Sci* 2018; **109**: 2364-2374 [PMID: [29908100](#) DOI: [10.1111/cas.13697](#)]
- 98 **Qiu S**, Lin S, Hu D, Feng Y, Tan Y, Peng Y. Interactions of miR-323/miR-326/miR-329 and miR-130a/miR-155/miR-210 as prognostic indicators for clinical outcome of glioblastoma patients. *J Transl Med* 2013; **11**: 10 [PMID: [23302469](#) DOI: [10.1186/1479-5876-11-10](#)]
- 99 **Huang T**, Deng CX. Current Progresses of Exosomes as Cancer Diagnostic and Prognostic Biomarkers. *Int J Biol Sci* 2019; **15**: 1-11 [PMID: [30662342](#) DOI: [10.7150/ijbs.27796](#)]
- 100 **Costa-Silva B**, Aiello NM, Ocean AJ, Singh S, Zhang H, Thakur BK, Becker A, Hoshino A, Mark MT, Molina H, Xiang J, Zhang T, Theilen TM, Garcia-Santos G, Williams C, Ararso Y, Huang Y, Rodrigues G, Shen TL, Labori KJ, Lothe IM, Kure EH, Hernandez J, Doussot A, Ebbesen SH, Grandgenett PM, Hollingsworth MA, Jain M, Mallya K, Batra SK, Jarnagin WR, Schwartz RE, Matei I, Peinado H, Stanger BZ, Bromberg J, Lyden D. Pancreatic cancer exosomes initiate pre-metastatic niche formation in the liver. *Nat Cell Biol* 2015; **17**: 816-826 [PMID: [25985394](#) DOI: [10.1038/ncb3169](#)]
- 101 **Sun W**, Luo JD, Jiang H, Duan DD. Tumor exosomes: a double-edged sword in cancer therapy. *Acta Pharmacol Sin* 2018; **39**: 534-541 [PMID: [29542685](#) DOI: [10.1038/aps.2018.17](#)]
- 102 **Whiteside TL**. The effect of tumor-derived exosomes on immune regulation and cancer immunotherapy. *Future Oncol* 2017; **13**: 2583-2592 [PMID: [29198150](#) DOI: [10.2217/fon-2017-0343](#)]
- 103 **Pitson SM**, Xia P, Leclercq TM, Moretti PA, Zebol JR, Lynn HE, Wattenberg BW, Vadas MA. Phosphorylation-dependent translocation of sphingosine kinase to the plasma membrane drives its oncogenic signalling. *J Exp Med* 2005; **201**: 49-54 [PMID: [15623571](#) DOI: [10.1084/jem.20040559](#)]
- 104 **Gao Y**, Gao F, Chen K, Tian ML, Zhao DL. Sphingosine kinase 1 as an anticancer therapeutic target. *Drug Des Devel Ther* 2015; **9**: 3239-3245 [PMID: [26150697](#) DOI: [10.2147/DDDT.S83288](#)]
- 105 **Watson C**, Long JS, Orange C, Tannahill CL, Mallon E, McGlynn LM, Pyne S, Pyne NJ, Edwards J. High expression of sphingosine 1-phosphate receptors, S1P1 and S1P3, sphingosine kinase 1, and extracellular signal-regulated kinase-1/2 is associated with development of tamoxifen resistance in estrogen receptor-positive breast cancer patients. *Am J Pathol* 2010; **177**: 2205-2215 [PMID: [20889557](#) DOI: [10.2353/ajpath.2010.100220](#)]
- 106 **Heffernan-Stroud LA**, Obeid LM. Sphingosine kinase 1 in cancer. *Adv Cancer Res* 2013; **117**: 201-235 [PMID: [23290781](#) DOI: [10.1016/B978-0-12-394274-6.00007-8](#)]
- 107 **Janvilisri T**, Suzuki H, Scaria J, Chen JW, Charoensawan V. High-Throughput Screening for Biomarker Discovery. *Dis Markers* 2015; **2015**: 108064 [PMID: [26060333](#) DOI: [10.1155/2015/108064](#)]
- 108 **Long NP**, Park S, Anh NH, Nghi TD, Yoon SJ, Park JH, Lim J, Kwon SW. High-Throughput Omics and Statistical Learning Integration for the Discovery and Validation of Novel Diagnostic Signatures in Colorectal Cancer. *Int J Mol Sci* 2019; **20** [PMID: [30642095](#) DOI: [10.3390/ijms20020296](#)]



Published by **Baishideng Publishing Group Inc**
7041 Koll Center Parkway, Suite 160, Pleasanton, CA 94566, USA

Telephone: +1-925-3991568

E-mail: bpgoffice@wjgnet.com

Help Desk: <https://www.f6publishing.com/helpdesk>

<https://www.wjgnet.com>

