

World Journal of *Gastroenterology*

World J Gastroenterol 2022 April 14; 28(14): 1384-1502



Contents

Weekly Volume 28 Number 14 April 14, 2022

EDITORIAL

- 1384 Hepatocellular adenoma: Where are we now?
Wang X, Zhang X

OPINION REVIEW

- 1394 Endoluminal vacuum-assisted therapy to treat rectal anastomotic leakage: A critical analysis
Vignali A, De Nardi P

REVIEW

- 1405 Viral hepatitis: Past, present, and future
Odenwald MA, Paul S
- 1430 Osteosarcopenia in autoimmune cholestatic liver diseases: Causes, management, and challenges
Pugliese N, Arcari I, Aghemo A, Lania AG, Lleo A, Mazzotti G

ORIGINAL ARTICLE

Basic Study

- 1444 Syngeneic implantation of mouse hepatic progenitor cell-derived three-dimensional liver tissue with dense collagen fibrils
Tamai M, Adachi E, Kawase M, Tagawa YI

Retrospective Cohort Study

- 1455 Clinical classification of symptomatic heterotopic pancreas of the stomach and duodenum: A case series and systematic literature review
LeCompte MT, Mason B, Robbins KJ, Yano M, Chatterjee D, Fields RC, Strasberg SM, Hawkins WG

Retrospective Study

- 1479 Radiomics signature: A potential biomarker for β -arrestin1 phosphorylation prediction in hepatocellular carcinoma
Che F, Xu Q, Li Q, Huang ZX, Yang CW, Wang LY, Wei Y, Shi YJ, Song B

LETTER TO THE EDITOR

- 1494 Comment on review article: Chronic hepatitis C virus infection cascade of care in pediatric patients
Bouare N, Keita M, Delwaide J
- 1499 Comments on "Effect of type 2 diabetes mellitus in the prognosis of acute-on-chronic liver failure patients in China"
Wang W, Pan CC, Zhao WY, Sheng JY, Wu QQ, Chen SS

ABOUT COVER

Editorial Board Member of *World Journal of Gastroenterology*, Sung-Chul Lim, MD, PhD, Professor, Head, Department of Pathology, Chosun University Hospital, 365 Pilmun-daero, Dong-gu, Gwangju 61453, South Korea. sclim@chosun.ac.kr

AIMS AND SCOPE

The primary aim of *World Journal of Gastroenterology* (WJG, *World J Gastroenterol*) is to provide scholars and readers from various fields of gastroenterology and hepatology with a platform to publish high-quality basic and clinical research articles and communicate their research findings online. WJG mainly publishes articles reporting research results and findings obtained in the field of gastroenterology and hepatology and covering a wide range of topics including gastroenterology, hepatology, gastrointestinal endoscopy, gastrointestinal surgery, gastrointestinal oncology, and pediatric gastroenterology.

INDEXING/ABSTRACTING

The WJG is now indexed in Current Contents®/Clinical Medicine, Science Citation Index Expanded (also known as SciSearch®), Journal Citation Reports®, Index Medicus, MEDLINE, PubMed, PubMed Central, and Scopus. The 2021 edition of Journal Citation Report® cites the 2020 impact factor (IF) for WJG as 5.742; Journal Citation Indicator: 0.79; IF without journal self cites: 5.590; 5-year IF: 5.044; Ranking: 28 among 92 journals in gastroenterology and hepatology; and Quartile category: Q2. The WJG's CiteScore for 2020 is 6.9 and Scopus CiteScore rank 2020: Gastroenterology is 19/136.

RESPONSIBLE EDITORS FOR THIS ISSUE

Production Editor: Hua-Ge Yin, Production Department Director: Xu Guo, Editorial Office Director: Ze-Mao Gong.

NAME OF JOURNAL

World Journal of Gastroenterology

ISSN

ISSN 1007-9327 (print) ISSN 2219-2840 (online)

LAUNCH DATE

October 1, 1995

FREQUENCY

Weekly

EDITORS-IN-CHIEF

Andrzej S Tarnawski

EDITORIAL BOARD MEMBERS

<http://www.wjgnet.com/1007-9327/editorialboard.htm>

PUBLICATION DATE

April 14, 2022

COPYRIGHT

© 2022 Baishideng Publishing Group Inc

INSTRUCTIONS TO AUTHORS

<https://www.wjgnet.com/bpg/gerinfo/204>

GUIDELINES FOR ETHICS DOCUMENTS

<https://www.wjgnet.com/bpg/GerInfo/287>

GUIDELINES FOR NON-NATIVE SPEAKERS OF ENGLISH

<https://www.wjgnet.com/bpg/gerinfo/240>

PUBLICATION ETHICS

<https://www.wjgnet.com/bpg/GerInfo/288>

PUBLICATION MISCONDUCT

<https://www.wjgnet.com/bpg/gerinfo/208>

ARTICLE PROCESSING CHARGE

<https://www.wjgnet.com/bpg/gerinfo/242>

STEPS FOR SUBMITTING MANUSCRIPTS

<https://www.wjgnet.com/bpg/GerInfo/239>

ONLINE SUBMISSION

<https://www.f6publishing.com>



Hepatocellular adenoma: Where are we now?

Xi Wang, Xuchen Zhang

Specialty type: Gastroenterology and hepatology

Provenance and peer review: Invited article; Externally peer reviewed.

Peer-review model: Single blind

Peer-review report's scientific quality classification

Grade A (Excellent): A
Grade B (Very good): B
Grade C (Good): C
Grade D (Fair): 0
Grade E (Poor): 0

P-Reviewer: Kotelevets SM, Russia; Mrzljak A, Croatia; Sugimura H, Japan

Received: December 5, 2021

Peer-review started: December 5, 2021

First decision: January 8, 2022

Revised: January 9, 2022

Accepted: March 6, 2022

Article in press: March 6, 2022

Published online: April 14, 2022



Xi Wang, Xuchen Zhang, Department of Pathology, Yale University School of Medicine, New Haven, CT 06520, United States

Corresponding author: Xuchen Zhang, MD, PhD, Associate Professor, Department of Pathology, Yale University School of Medicine, 310 Cedar Street, PO Box 208023, New Haven, CT 06520, United States. xuchen.zhang@yale.edu

Abstract

Hepatocellular adenoma (HCA) is a benign hepatocellular neoplasm, commonly occurs in young women with a history of oral contraceptive use. Complications including hemorrhage and malignant transformation necessitate the need for a thorough understanding of the underlying molecular signatures in this entity. Recent molecular studies have significantly expanded our knowledge of HCAs. The well-developed phenotype-genotype classification system improves clinical management through identifying "high risk" subtype of HCAs. In this article, we attempt to provide updated information on clinical, pathologic and molecular features of each subtype of HCAs.

Key Words: Hepatocellular adenoma; Subtype; Pathology; Classification; Hepatocellular carcinoma

©The Author(s) 2022. Published by Baishideng Publishing Group Inc. All rights reserved.

Core Tip: Hepatocellular adenoma (HCA) has been well recognized as a benign liver neoplasm with risks of hemorrhage and malignant transformation. Studies revealed that different HCA subtypes with specific genetic mutation and pathologic findings are associated with different clinical features. Currently HCAs are classified into at least 5 major subtypes, involving 4 different pathways driving HCA pathogenesis: Hepatocyte nuclear factor 1A, interleukin-6/the Janus kinase/signal transducer and activator of transcription, β -catenin, and Sonic hedgehog pathway.

Citation: Wang X, Zhang X. Hepatocellular adenoma: Where are we now? *World J Gastroenterol* 2022; 28(14): 1384-1393

URL: <https://www.wjgnet.com/1007-9327/full/v28/i14/1384.htm>

DOI: <https://dx.doi.org/10.3748/wjg.v28.i14.1384>

INTRODUCTION

Hepatocellular adenoma (HCA) is an uncommon benign liver neoplasm with two major complications: Hemorrhage and malignant transformation. Epidemiological data from the United States and European countries have revealed HCAs occur mainly in young to middle-aged women (median ages: 36-38 years, female/male ratio of 8:1), often with a history of long-term use of oral contraceptives (OCPs)[1,2]. Besides the well-known risk factor of exogenous estrogen exposure, other risk factors, such as androgen use, obesity, fatty liver disease, glycogen storage disease (GSD), especially GSD type 1, hepatic vascular disorders and other genetic disorders are also associated with the occurrence of HCA[2-6]. However, epidemiological data from Asia, where the use of OCPs and the incidence of obesity are lower, are unclear. Limited studies from Taiwan, China and Japan showed a male predominance in patients with HCA[7-11]. Further exploration of these epidemiological differences is needed to better understand the pathogenesis of HCA.

Historically, HCA was thought to be a single group of tumor. Recent advances in HCA clinicopathologic features and molecular biology have not only enhanced our understanding of the disease pathogenesis, but also significantly transitioned into the daily practice of pathology-morpho-molecular correlation. It is known now that HCA is a heterogeneous group of liver tumors with heterogeneous etiology, clinical presentation, risk of malignant transformation or hemorrhage, radiologic findings, histopathologic features, clinical management strategies and underlying molecular changes. In the current review, we will provide an overview of the current knowledge of how HCA histomorphology correlates to its underlying molecular changes, as well as discuss the controversies in some HCA variants.

HCA: MORPHO-MOLECULAR CLASSIFICATION

Currently, HCAs are classified into hepatocyte nuclear factor 1A (*HNF-1A*) inactivated HCA (H-HCA), inflammatory HCA (I-HCA), β -catenin activated HCA (b-HCA), as well as unclassified HCA (U-HCA) based on underlying molecular changes in 3 different pathways driving benign hepatocytic proliferation: *HNF-1A*, interleukin-6/the Janus kinase/signal transducer and activator of transcription (IL-6/JAK/STAT), and β -catenin signaling. A new subtype of Sonic hedgehog HCA (shHCA) has been recently described[2], which still needs further characterization. The different mutations in HCA identified to date are summarized in Table 1.

H-HCA

As the first identified subtype, *HNF-1A*-inactivated HCA was proposed by a French group in their pioneer work published in 2002. In an attempt to search for tumor suppressor gene in HCA, Bluteau *et al*[12] genotyped DNA from HCAs and confirmed the bi-allelic inactivation of *HNF-1A* gene in a subgroup of tumors. *HNF-1A* is an important transcription factor regulating hepatocytes differentiation. It is mainly expressed in pancreatic beta cells, intestine, and liver, that plays an important role in the regulation of glycolipid metabolism[13,14]. Specifically, *HNF-1A* positively regulates *FABP1* gene, which encodes liver fatty acid binding protein (L-FABP). As a result of *HNF-1A* inactivated mutation, expression of L-FABP is downregulated, along with diffuse steatosis due to dysregulated lipogenesis.

As expected, *HNF-1A*-inactivated HCA histomorphologically shows diffuse marked steatosis (Figure 1A). Immunohistochemical stain for L-FABP is negative (Figure 1B), indicating the functional loss of *HNF-1A* gene. Of note, some H-HCAs may not show steatosis and some other subtype HCAs such as I-HCAs can be seen with marked steatosis. Therefore, steatosis alone should not be used as a sole feature to diagnose H-HCAs. This subtype of HCA approximately composes of 30%-35% of overall HCAs and is mainly due to somatic mutation. Typical H-HCA can also be detected in magnetic resonance imaging (MRI), with diffuse and homogenous signal dropout on T1-weighted images, due to massive fat component[15]. In the clinical aspect, cases of familial liver adenomatosis (greater than 10 adenomas in the liver) have been consistently linked to the germline mutation of *HNF-1A*[16], which is also the genotype in "Maturity onset diabetes of the young, type 3 (MODY3)". Thus, detection of liver adenomatosis with H-HCA is suggested to start family screen for familial adenomatosis, MODY3 diabetes, and the *HNF-1A* germline mutation[17]. Although H-HCA is not usually associated with malignant transformation, hepatocellular carcinoma (HCC) has been reported to arise in the settings of sporadic HCAs[18] as well as those in patients with hepatic vascular disorders or MODY3[19], especially in female patients with multiple lesions without significant steatosis and presence of myxoid change, peliosis and sinusoidal dilatation[3].

I-HCA

Accumulating evidence suggests a critical role of inflammatory response in tumorigenesis, including STAT signaling pathway in the development of breast and lung cancer[20]. Similarly, a subtype of inflammatory epithelial tumors has been described in the family of HCA, involving the IL-6/JAK/STAT3 signaling pathway[21]. IL-6 belongs to the IL-6 cytokine family, which binds to the ligand-

Table 1 Morpho-molecular features of the subtypes of hepatocellular adenoma

	Key pathogenesis	Histology	Immunohistochemical stains	Clinical features
H-HCA	<i>HNF1A</i> inactivating mutation: Negative regulation of glycolipid metabolism and L-FABP	Marked steatosis	L-FABP: Negative	Associated with maturity onset diabetes of the young (MODY3) and familial hepatic adenomatosis
I-HCA	IL-6/JAK/STAT3 pathway mutations: (1) Constitutive activation of inflammatory pathway; and (2) Upregulation of acute reactants and hepatocellular proliferation	(1) Inflammatory infiltration; (2) Pseudoportal tracts; (3) Ductular reaction; and (4) Sinusoidal dilatation/peliosis	SSA and CRP: Diffuse positive	Obesity, metabolic syndrome, glycogen storage disease, high alcohol consumption, inflammatory syndrome
b-HCA	(1) <i>CTNNB1</i> gene mutation: Activation of signaling pathway and upregulation of targeted genes including GS; and (2) Level of activation depends on mutation loci (Exon 3 Non-S45: Strong activation; S45: Weak activation, and T41: Moderate activation; Exon 7/8: Weak activation)	More atypical features: Pseudoacini and mild cytologic atypia	(1) β -catenin: Aberrant nuclear expression; and (2) GS: Positive	More in men, anabolic steroids use, glycogen storage disease, high risk of malignant transformation, and high risk of bleeding (exon 7/8)
b-IHCA	Share the features of both b-HCA and I-HCA			
shHCA	<i>INHBE-GLI1</i> gene fusion: Constitutive activation of Sonic hedgehog pathway	Hemorrhage	PTGSD and ASS1	High risk of bleeding and obesity
Unclassified	Not other specified			
Uncommon subtypes	Myxoid HCA, pigmented HCA, atypical HCA, I-HCA in cirrhotic liver			

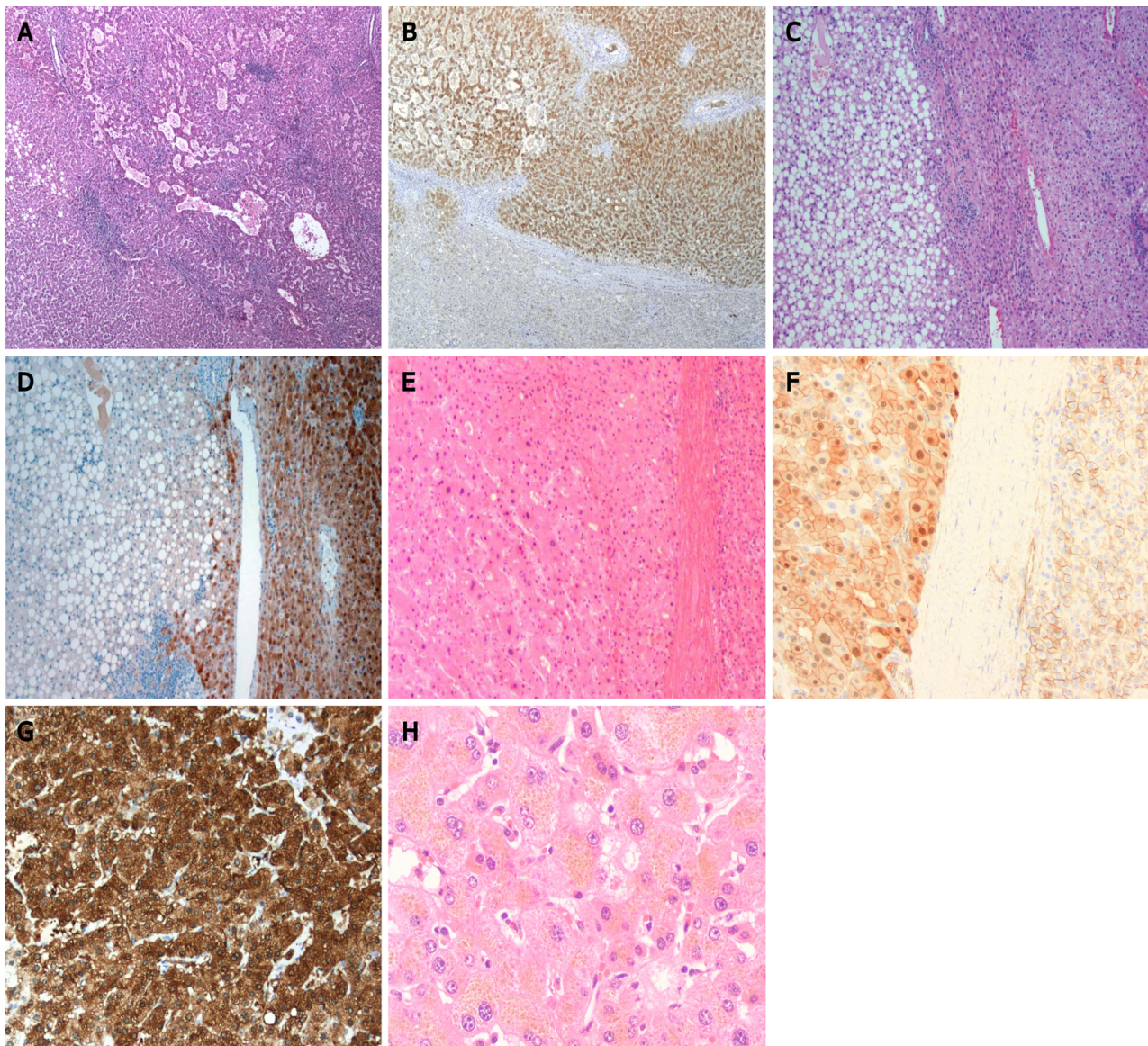
HNF1A: Hepatocyte nuclear factor 1A; L-FABP: Liver fatty acid binding protein; SSA: Serum amyloid A; CRP: C-reactive protein; CTNNB1: Catenin beta 1; GS: Glutamine synthetase; INHBE-GLI1: Inhibin beta E chain/ glioma-associated oncogene 1; PTGDS: Prostaglandin D synthase; ASS1: Argininosuccinate Synthase 1; HCA: Hepatocellular adenoma; I-HCA: Inflammatory HCA; H-HCA: *HNF1A*-inactivated HCA; shHCA: Sonic hedgehog HCA; b-HCA: β -catenin mutated HCA.

specific receptor gp130 to initiate the downstream signaling. This activates downstream signaling pathways such as the Src-homology 2 domain-containing tyrosine phosphatase 2 (SHP2)-Ras-ERK, JAK1/2-STAT3, mosaic G-protein alpha-subunit (GNAS), and the phosphatidylinositol-3-kinase/ Akt and the mechanistic target of rapamycin (PI3K-AKT-mTOR), and further the expression of target genes that regulate cell survival, proliferation and angiogenesis[2,22]. Rebouissou *et al*[21] demonstrated that 60% of I-HCA harbored small in-frame deletions of *IL6ST* gene (encodes the signaling co-receptor gp130), resulting in the constitutive activation of IL-6 signaling and hepatocellular proliferation in the absence of ligand binding. The remaining I-HCAs are linked to other mutations that belong to IL-6/JAK/STAT3 family. The inflammatory response is well-observed histologically[23], characterized by inflammatory infiltration of lymphocytes, plasma cells and neutrophils, ductular reaction, and sinusoidal dilatation/peliosis (Figure 1C). Immunohistochemical stain is remarkable for diffuse overexpression of acute-phase inflammatory protein such as serum amyloid A (SAA) and C-reactive protein (CRP) (Figure 1D).

Clinically, I-HCA is frequently associated with obesity, metabolic syndrome and high alcohol consumption. Furthermore, the risk of developing HCA in patients with GSD is high. The HCAs in patients with GSD are mainly I-HCAs (52%), followed by b-HCAs (28%) and U-HCAs (20%), but never H-HCAs[5]. A few I-HCA cases had also been reported to induce systemic AA amyloidosis[24]. Typical I-HCA also has unique MRI features, with hyperintensity on T2-weighted images due to sinusoidal dilatation[15]. This subtype approximately composes of 35%-45% of overall HCAs, and about 10% also have β -catenin activation. While the mutation alone is not associated with malignant transformation, it is important to remember that I-HCA can co-exist with β -catenin mutation, the latter of which can transform into HCC[2].

b-HCA

Aberrant Wnt/ β -catenin signaling has been identified underlying pathogenesis of many diseases including the well-known familial adenomatous polyposis[25]. In the liver, it also plays an essential role in regulating various cellular events including differentiation, proliferation, survival and others. Not surprisingly, β -catenin (cadherin-associated protein) beta 1 (*CTNNB1*) gene mutations have been reported in around 20%-40% of HCC cases. Chen *et al*[26] detected interstitial deletions in the *CTNNB1* gene in HCAs, indicating the dysregulation of Wnt/ β -catenin as a possible preneoplastic pathway for hepatocellular proliferation. Exome sequencing study[27] further subclassified into two types: Exon 3



DOI: 10.3748/wjg.v28.i14.1384 Copyright © The Author(s) 2022.

Figure 1 Histologic features of hepatocellular adenoma. A: Hepatocyte nuclear factor 1A (HNF-1A) inactivated hepatocellular adenoma showing marked steatosis (Hematoxylin-eosin stain, original magnification 200 ×); B: HNF-1A inactivated hepatocellular adenoma showing loss expression of liver fatty acid binding protein in tumor component (Immunohistochemical stain, original magnification 200 ×); C: Inflammatory hepatocellular adenoma showing marked sinusoidal dilatation and pseudoportal tract with inflammatory infiltrate and ductular reaction (Hematoxylin-eosin stain, original magnification 200 ×); D: Inflammatory hepatocellular adenoma showing strong and diffuse expression of C-reactive protein in tumor component (Immunohistochemical stain, original magnification 200 ×); E: β -catenin activated hepatocellular adenoma showing mild cytologic atypia and pseudoacini (Hematoxylin-eosin stain, original magnification 200 ×); F: β -catenin activated hepatocellular adenoma showing nuclear expression of β -catenin in tumor component (Immunohistochemical stain, original magnification 400 ×); G: β -catenin activated hepatocellular adenoma showing strong and diffuse expression of glutamine synthetase in tumor component (Immunohistochemical stain, original magnification 200 ×); H: Pigmented hepatocellular adenoma showing marked cytoplasmic lipofuscin (Hematoxylin-eosin stain, original magnification 400 ×).

and exon 7/8 with distinct canonical and non-canonical mutations respectively. In the canonical pathway, inactive β -catenin is attached with E-cadherin at cell membrane to maintain cell-cell adhesion. Cytoplasmic β -catenin is sequestered by a “destruction protein complex”, then phosphorylated, and further recognized by E3 ubiquitin, eventually degraded in the cytoplasm by proteasomes. The overall net effect is low β -catenin levels within the cells. In the activate states or upon WNT binding, the phosphorylation activity is inhibited and the disheveled protein complex becomes disintegrated. As a result, β -catenin accumulates in the cytoplasm and translocates into the nucleus to regulate downstream target gene expression, such as a glutamate-ammonia ligase, which codes for glutamine synthase (GS) and leucine-rich-repeat containing G protein-coupled receptor-5 (LGR5). Thus, the overall net effect is abnormally high expression level of β -catenin within the nucleus and cytoplasm[28]. HCAs with β -catenin mutation are divided into two groups based on the mutation loci in the *CTNNB1* coding for β -catenin: β -catenin exon 3 mutated HCA and β -catenin exon 7/8 mutated HCA[29].

b-HCA exon 3: Approximately 10%-15% of HCAs harbored canonical mutations in *CTNNB1* gene exon 3. The level of β -catenin pathway activation further depends on the specific mutations: Large in-frame deletions, D32-S37 deletions and T41 exon 3 mutations are associated with high level, whereas S45 exon 3 mutations are associated with low level of activation[29]. Comparing with other HCA subtypes, this subtype HCA has more atypical features including mild cytologic atypia, cholestasis, and/or pseudoacinar formation (Figure 1E), which sometimes can be difficult to distinguish from well differentiated HCC. Of note, no specific histologic features can be reliably used to predict a diagnosis of b-HCA. Immunohistochemical stains for β -catenin shows aberrant nuclear and cytoplasmic expression (Figure 1F). While the interpretation of β -catenin expression can be challenging, a better surrogate marker, GS, is also widely used. A recent study has revealed that diffuse homogenous GS staining pattern was strongly associated with exon 3 non-S45 mutation (Figure 1G). Whereas, a diffuse heterogeneous GS staining pattern with strong positivity at the tumor periphery indicated exon 3 S45 mutation [30].

Clinically, β -catenin exon 3 mutated HCA is more frequent in men than other subtypes and is more often associated with androgen exposure. In addition, this subtype especially with large in-frame deletions is associated with a high risk of malignant transformation. It is hypothesized that *CTNNB1* exon 3 mutation is the earliest genetic alteration, while additional mutations such as telomere reverse transcriptase (TERT) promoter mutation is involved in the final step of transition from HCA to HCC [27].

b-HCA exon 7/8: A smaller subset of HCAs (5%-10%) harbor non-canonical mutations of *CTNNB1* gene in exon 7 and 8, which is exclusive to mutations in exon 3. In comparison with exon 3 mutations, mutations in exon 7 and 8 result in a weak activation of β -catenin. A focal patchy GS staining pattern in the tumor with strong positivity at the tumor periphery by immunohistochemistry can indicate exon 7/8 mutation[30]. This subtype is associated with a low risk of malignant transformation. A recent study demonstrated that b-HCA exon 7/8 is significantly associated with tumor hemorrhage[31].

β -catenin activated I-HCA

While β -catenin mutations are nearly exclusive to *HNF-1A* mutations, they can be associated with altered JAK/STAT pathway and demonstrate inflammatory features. If the I-HCA shows beta-catenin activation, then a diagnosis of I-HCA with beta-catenin activation can be rendered [β -catenin activated I-HCA (b-IHCA)], which shares the histopathologic features of both b-HCA and I-HCA, and carries malignant transformation potential similarly. It is thus important to continue workup for β -catenin inactivation when a diagnosis of I-HCA is made.

shHCA

ShHCA is a recently recognized subtype of HCA with sonic hedgehog pathway activation[2]. This subtype represents approximately 4% of overall HCAs and was previously classified as unclassified HCA due to the lack of mutations in typical HCA genes. Genetic studies demonstrated that this type of HCA is caused by inhibin beta E subunit (*INHBE*) gene and *GLI1* gene fusion (INHBE-GLI1). The Hedgehog pathway is a complex signal transduction pathway including 4 main components: The ligand Hedgehog, the receptor Patched, the signal transducer Smoothened and the effector transcription factor, Gli[32]. It is not only crucial for embryogenesis of liver, but also plays a role in liver regeneration. Evidence has shown that the Hedgehog pathway is dormant in healthy adult liver, while significantly activated after liver injury. INHBE is a growth factor belonging to the transforming growth factor-beta family. It is highly expressed in the liver and regulates hepatocellular growth and differentiation. As a result, the INHBE-GLI1 fusion leads to uncontrolled activation of sonic hedgehog pathway due to overexpression of transcription factor GLI1.

Clinically, shHCAs are more frequently seen in women and are associated with higher body mass index and/or OCP use. While there are no specific histopathologic features in this HCA subtype, it seems that prostaglandin D synthase and argininosuccinate synthase 1 (ASS1), particularly ASS1 positivity by immunohistochemistry is a hallmark of shHCA[2,33]. Although ASS1 may be expressed in other HCA subtypes with hemorrhage and thought to be a marker of hemorrhage[34], a recent study showed ASS1 expression did not correlate with HCA hemorrhagic complications[35]. Interestingly, higher risk of hemorrhage has been consistently observed both histologically and clinically in shHCA[2, 31]. This serious complication warrants a thorough understanding of pathogenesis of shHCA for better diagnosis and clinical management.

Unclassified HCA

Even though extensive studies have been conducted to explore molecular features of HCAs, approximately 5%-10% remain unclassified. These HCAs lack distinct histopathologic features and any specific molecular abnormality[2].

UNUSUAL SUBTYPES AND CONTROVERSIES

In addition, uncommon HCA subtypes have been well documented, which do not fit well into the current classification.

Pigmented HCA

Pigmented HCAs are a heterogeneous group of HCAs with different genetic mutations, which contain pigment deposition of lipofuscin (Figure 1H) as confirmed by electron microscopy[36]. Although H-HCA is the commonest subtype, other subtypes such as b-HCA, I-HCA, b-IHCA, and unclassified HCA also can be seen in pigmented HCAs[36-38]. Pigmented HCAs often show histologic atypia with higher risk malignant transformation. Besides the heavy deposition of lipofuscin in tumor cells, lipofuscin pigment deposition is commonly seen in hepatocytes of the background livers[36]. Although unclear, the underlying biology may be related to dysregulation of autophagy resulting in lipofuscin accumulation, which could contribute to carcinogenesis including the liver[39,40].

Myxoid HCA

Myxoid HCAs are characterized by the deposition of myxoid materials between the hepatic cords within the tumor[41-44]. In addition to loss of L-FABP expression and/or *HNF-1A* mutation as documented in all myxoid HCAs, a recent study identified recurrent mutations in genes within the protein kinase A (PKA) pathway or in genes that regulate the PKA pathway, such as *GNAS*, *CDKN1B* (p27) and *RNF123*, in myxoid HCAs[44]. Myxoid HCAs are often seen in individuals with older age and carry a high risk of malignant transformation[3,44]. It is still controversial whether this is a rare variant of H-HCA with additional mutations or a distinct subtype of HCAs.

Atypical/borderline HCA/hepatocellular neoplasm of uncertain malignant potential

HCA, as described above, has potential of malignant transformation that sometimes can be challenging to distinguish from well-differentiated HCC, especially in biopsy specimens. Various terms, such as atypical HCA, borderline HCA, atypical hepatocellular neoplasm, well-differentiated hepatocellular neoplasm with atypical or borderline features, and hepatocellular neoplasm of uncertain malignant potential (HUMP) have been used for hepatocellular neoplasms that demonstrate features atypical for HCA but insufficient for an unequivocal diagnosis of HCC. So far there are no widely accepted criteria in diagnosing this entity, however, these clinical (male, females > 50 years or < 15 years) and pathologic (focal cytological atypia, small cell change, pseudoacini, focal reticulin network loss, presence of β -catenin activation or *CTNNB1* mutations) features have been consistently used when an atypical/borderline HCA (A-HCA)/HUMP is diagnosed[45-47]. A recent study showed that greater than 60% HCAs would be re-classified as A-HCAs/HUMPs using the above suggested criteria. Furthermore, in this study A-HCA/HUMP does not seem to correlate in patients with or without synchronous or metachronous HCC[45]. The high rate of HCAs placed in the category of A-HCA/HUMP, particularly in resected tumors may cause confusion to clinicians in managing those tumors. Molecular study of *TERT* promoter mutations, a marker of HCC, may be useful to distinguish A-HCA/HUMP from well-differentiated HCC and predict the risk of malignant transformation. Studies have demonstrated that *TERT* promoter mutations have been identified in 17% of A-HCAs/HUMPs compared with 50%-60% of HCC[48]. Further study to refine the widely accepted criteria of diagnosing A-HCA/HUMP and to predict its malignant behavior by combining the clinical, pathological and molecular features is warranted.

I-HCA in cirrhotic livers

HCAs typically arise in the livers without significant fibrosis. The background liver can be histologically normal or have steatosis or steatohepatitis or other genetic and vascular disorders that have been recognized as risk factors for developing HCAs. Thus, a solid mass arising in a cirrhotic liver is generally not considered as HCA. However, rare HCAs with SAA positivity or harboring activating mutations in *IL6ST* or *STAT3* same as I-HCAs have been described in metabolic syndrome and/or alcoholic cirrhosis[49,50]. Of note, some cirrhotic nodules and HCCs can be positive for CRP and SAA, the two important I-HCA markers. Furthermore, activation of IL-6/gp130/STAT3-dependent pathway is involved in the development of liver fibrosis[51]. Therefore, classifying those SAA-positive cirrhotic nodules with *IL6ST* or *STAT3* mutations as I-HCAs is thought to be premature[52]. Additional data are warranted to confirm whether HCA can arise in a background liver with cirrhosis.

MANAGEMENT

Guidelines from both the American College of Gastroenterology[53] and the European Association for Study of the Liver[54] recommend a surgical resection when HCAs are > 5 cm, since they have a higher potential for hemorrhage and malignant transformation. The adoption of a genotype-phenotype classi-

fication is increasingly relevant to clinical decision-making given growing evidence that the risk of complications is likely dependent on HCA subtype and gender. Currently, the treatment of HCA is based on HCA subtype and gender[17,54,55]. Briefly, surgery is recommended for all men with HCAs since they carry a high risk of malignant transformation. For women, after the initial 6 mo lifestyle management including the discontinuation of OCPs and control of body weight, the management of HCAs is based on the size and HCA subtypes. For tumors persistently > 5 cm, or increasing in size after lifestyle change, irrespective of their subtypes, resection or curative treatment is indicated. For tumors < 5 cm of the H-HCA subtype, or those that are either inflammatory or β -catenin negative on biopsy, conservative management is recommended. Of note, surgical resection is recommended for b-HCAs, b-IHCAs and A-HCAs/HUMPs, irrespective of size. Of note, HCAs arising in patients with underlying liver diseases, such as GSD and hepatic vascular disorders seem to have a higher risk of malignant transformation, that should also be considered when manage these HCAs.

CONCLUSION

Recent molecular studies have significantly expanded our knowledge of HCAs. The newly developed phenotype-genotype classification system not only fulfils our curiosity academically, but more importantly, helps improving clinical management through identifying the “high risk” subtype of HCAs. However, further studies are needed, including how to incorporate the uncommon HCA subtypes into the classification system, how to effectively guide the HCA management by using the classification system and identifying the underlying molecular changes of the unclassified HCAs.

FOOTNOTES

Author contributions: Wang X reviewed the literature and drafted the manuscript; Zhang X provided overall intellectual input, reviewed the literature, acquired the histological images, and edited the final version of the manuscript; all authors approved the final version to be published.

Conflict-of-interest statement: The authors have no conflicts of interest to disclose.

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <https://creativecommons.org/licenses/by-nc/4.0/>

Country/Territory of origin: United States

ORCID number: Xi Wang 0000-0001-8916-5132; Xuchen Zhang 0000-0002-1484-4672.

S-Editor: Fan JR

L-Editor: A

P-Editor: Fan JR

REFERENCES

- 1 **Margolskee E**, Bao F, de Gonzalez AK, Moreira RK, Lagana S, Sireci AN, Sepulveda AR, Remotti H, Lefkowitz JH, Salomao M. Hepatocellular adenoma classification: a comparative evaluation of immunohistochemistry and targeted mutational analysis. *Diagn Pathol* 2016; **11**: 27 [PMID: 26961851 DOI: 10.1186/s13000-016-0475-5]
- 2 **Nault JC**, Couchy G, Balabaud C, Morcrette G, Caruso S, Blanc JF, Bacq Y, Calderaro J, Paradis V, Ramos J, Scoazec JY, Gnemmi V, Sturm N, Guettier C, Fabre M, Savier E, Chiche L, Labrune P, Selves J, Wendum D, Pilati C, Laurent A, De Muret A, Le Bail B, Rebouissou S, Imbeaud S; GENTHEP Investigators, Bioulac-Sage P, Letouzé E, Zucman-Rossi J. Molecular Classification of Hepatocellular Adenoma Associates With Risk Factors, Bleeding, and Malignant Transformation. *Gastroenterology* 2017; **152**: 880-894.e6 [PMID: 27939373 DOI: 10.1053/j.gastro.2016.11.042]
- 3 **Putra J**, Ferrell LD, Gouw ASH, Paradis V, Rishi A, Sempoux C, Balabaud C, Thung SN, Bioulac-Sage P. Malignant transformation of liver fatty acid binding protein-deficient hepatocellular adenomas: histopathologic spectrum of a rare phenomenon. *Mod Pathol* 2020; **33**: 665-675 [PMID: 31570768 DOI: 10.1038/s41379-019-0374-x]
- 4 **Cheng L**, Jain D, Kakar S, Torbenson MS, Wu TT, Yeh MM. Hepatocellular neoplasms arising in genetic metabolic disorders: steatosis is common in both the tumor and background liver. *Hum Pathol* 2021; **108**: 93-99 [PMID: 33245984 DOI: 10.1016/j.humpath.2020.11.012]
- 5 **Calderaro J**, Labrune P, Morcrette G, Rebouissou S, Franco D, Prévot S, Quaglia A, Bedossa P, Libbrecht L, Terracciano L, Smit GP, Bioulac-Sage P, Zucman-Rossi J. Molecular characterization of hepatocellular adenomas developed in patients

- with glycogen storage disease type I. *J Hepatol* 2013; **58**: 350-357 [PMID: 23046672 DOI: 10.1016/j.jhep.2012.09.030]
- 6 **Sakellariou S**, Al-Hussaini H, Scalori A, Samyn M, Heaton N, Portmann B, Tobal K, Quaglia A. Hepatocellular adenoma in glycogen storage disorder type I: a clinicopathological and molecular study. *Histopathology* 2012; **60**: E58-E65 [PMID: 22372484 DOI: 10.1111/j.1365-2559.2011.04153.x]
 - 7 **Huang WC**, Liao JY, Jeng YM, Liu KL, Lin CN, Song HL, Tsai JH. Hepatocellular adenoma in Taiwan: Distinct ensemble of male predominance, overweight/obesity, and inflammatory subtype. *J Gastroenterol Hepatol* 2020; **35**: 680-688 [PMID: 31698521 DOI: 10.1111/jgh.14903]
 - 8 **Wang H**, Yang C, Rao S, Ji Y, Han J, Sheng R, Zeng M. MR imaging of hepatocellular adenomas on genotype-phenotype classification: A report from China. *Eur J Radiol* 2018; **100**: 135-141 [PMID: 29496071 DOI: 10.1016/j.ejrad.2018.01.023]
 - 9 **Sasaki M**, Yoneda N, Kitamura S, Sato Y, Nakanuma Y. Characterization of hepatocellular adenoma based on the phenotypic classification: The Kanazawa experience. *Hepatol Res* 2011; **41**: 982-988 [PMID: 21883740 DOI: 10.1111/j.1872-034X.2011.00851.x]
 - 10 **Fukusato T**, Soejima Y, Kondo F, Inoue M, Watanabe M, Takahashi Y, Aso T, Uozaki H, Sano K, Sanada Y, Niki T. Preserved or enhanced OATP1B3 expression in hepatocellular adenoma subtypes with nuclear accumulation of β -catenin. *Hepatol Res* 2015; **45**: E32-E42 [PMID: 25418671 DOI: 10.1111/hepr.12453]
 - 11 **Izu A**, Sugitani M, Kinukawa N, Matsumura H, Ogawa M, Moriyama M, Yamazaki S, Takayama T, Hano H, Yao T, Kanda H, Suzuki K, Hayashi S, Ariizumi S, Yamamoto M, Morishita Y, Matsumoto K, Nakamura N, Nakano M. Hepatocellular adenoma, approximately half and predominantly inflammatory subtype, in 38 Japanese patients with several differences in age, gender, and clinical background factors from Western populations. *Hepatol Res* 2021; **51**: 336-342 [PMID: 33381872 DOI: 10.1111/hepr.13613]
 - 12 **Bluteau O**, Jeannot E, Bioulac-Sage P, Marqués JM, Blanc JF, Bui H, Beaudoin JC, Franco D, Balabaud C, Laurent-Puig P, Zucman-Rossi J. Bi-allelic inactivation of TCF1 in hepatic adenomas. *Nat Genet* 2002; **32**: 312-315 [PMID: 12355088 DOI: 10.1038/ng1001]
 - 13 **Cereghini S**, Yaniv M, Cortese R. Hepatocyte dedifferentiation and extinction is accompanied by a block in the synthesis of mRNA coding for the transcription factor HNF1/LFB1. *EMBO J* 1990; **9**: 2257-2263 [PMID: 2357969]
 - 14 **Yamagata K**, Oda N, Kaisaki PJ, Menzel S, Furuta H, Vaxillaire M, Southam L, Cox RD, Lathrop GM, Boriraj VV, Chen X, Cox NJ, Oda Y, Yano H, Le Beau MM, Yamada S, Nishigori H, Takeda J, Fajans SS, Hattersley AT, Iwasaki N, Hansen T, Pedersen O, Polonsky KS, Bell GI. Mutations in the hepatocyte nuclear factor-1alpha gene in maturity-onset diabetes of the young (MODY3). *Nature* 1996; **384**: 455-458 [PMID: 8945470 DOI: 10.1038/384455a0]
 - 15 **Shanbhogue A**, Shah SN, Zaheer A, Prasad SR, Takahashi N, Vikram R. Hepatocellular adenomas: current update on genetics, taxonomy, and management. *J Comput Assist Tomogr* 2011; **35**: 159-166 [PMID: 21412084 DOI: 10.1097/RCT.0b013e31820bad61]
 - 16 **Bacq Y**, Jacquemin E, Balabaud C, Jeannot E, Scotto B, Branchereau S, Laurent C, Bourlier P, Pariente D, de Muret A, Fabre M, Bioulac-Sage P, Zucman-Rossi J. Familial liver adenomatosis associated with hepatocyte nuclear factor 1alpha inactivation. *Gastroenterology* 2003; **125**: 1470-1475 [PMID: 14598263 DOI: 10.1016/j.gastro.2003.07.012]
 - 17 **Védie AL**, Sutter O, Ziol M, Nault JC. Molecular classification of hepatocellular adenomas: impact on clinical practice. *Hepat Oncol* 2018; **5**: HEP04 [PMID: 30302195 DOI: 10.2217/hep-2017-0023]
 - 18 **Hechtman JF**, Abou-Alfa GK, Stadler ZK, Mandelker DL, Roehrl MHA, Zehir A, Vakiani E, Middha S, Klimstra DS, Shia J. Somatic HNF1A mutations in the malignant transformation of hepatocellular adenomas: a retrospective analysis of data from MSK-IMPACT and TCGA. *Hum Pathol* 2019; **83**: 1-6 [PMID: 30121369 DOI: 10.1016/j.humpath.2018.08.004]
 - 19 **Stueck AE**, Qu Z, Huang MA, Campreciós G, Ferrell LD, Thung SN. Hepatocellular Carcinoma Arising in an HNF-1 α -Mutated Adenoma in a 23-Year-Old Woman with Maturity-Onset Diabetes of the Young: A Case Report. *Semin Liver Dis* 2015; **35**: 444-449 [PMID: 26676820 DOI: 10.1055/s-0035-1567827]
 - 20 **Grivennikov S**, Karin M. Autocrine IL-6 signaling: a key event in tumorigenesis? *Cancer Cell* 2008; **13**: 7-9 [PMID: 18167335 DOI: 10.1016/j.ccr.2007.12.020]
 - 21 **Rebouissou S**, Amessou M, Couchy G, Poussin K, Imbeaud S, Pilati C, Izard T, Balabaud C, Bioulac-Sage P, Zucman-Rossi J. Frequent in-frame somatic deletions activate gp130 in inflammatory hepatocellular tumours. *Nature* 2009; **457**: 200-204 [PMID: 19020503 DOI: 10.1038/nature07475]
 - 22 **Rosell R**, Bertran-Alamillo J, Molina MA, Taron M. IL-6/gp130/STAT3 signaling axis in cancer and the presence of in-frame gp130 somatic deletions in inflammatory hepatocellular tumors. *Future Oncol* 2009; **5**: 305-308 [PMID: 19374537 DOI: 10.2217/fon.09.3]
 - 23 **Fievet P**, Sevestre H, Boudjelal M, Noel LH, Kemeny F, Franco D, Delamarre J, Capron JP. Systemic AA amyloidosis induced by liver cell adenoma. *Gut* 1990; **31**: 361-363 [PMID: 2157638 DOI: 10.1136/gut.31.3.361]
 - 24 **Calderaro J**, Letouze E, Bayard Q, Boulai A, Renault V, Deleuze JF, Bestard O, Franco D, Zafrani ES, Nault JC, Moutschen M, Zucman-Rossi J. Systemic AA Amyloidosis Caused by Inflammatory Hepatocellular Adenoma. *N Engl J Med* 2018; **379**: 1178-1180 [PMID: 30231230 DOI: 10.1056/NEJMc1805673]
 - 25 **Clevers H**, Nusse R. Wnt/ β -catenin signaling and disease. *Cell* 2012; **149**: 1192-1205 [PMID: 22682243 DOI: 10.1016/j.cell.2012.05.012]
 - 26 **Chen YW**, Jeng YM, Yeh SH, Chen PJ. P53 gene and Wnt signaling in benign neoplasms: beta-catenin mutations in hepatic adenoma but not in focal nodular hyperplasia. *Hepatology* 2002; **36**: 927-935 [PMID: 12297840 DOI: 10.1053/jhep.2002.36126]
 - 27 **Pilati C**, Letouze E, Nault JC, Imbeaud S, Boulai A, Calderaro J, Poussin K, Franconi A, Couchy G, Morcrette G, Mallet M, Taouji S, Balabaud C, Terris B, Canal F, Paradis V, Scoazec JY, de Muret A, Guettier C, Bioulac-Sage P, Chevet E, Calvo F, Zucman-Rossi J. Genomic profiling of hepatocellular adenomas reveals recurrent FRK-activating mutations and the mechanisms of malignant transformation. *Cancer Cell* 2014; **25**: 428-441 [PMID: 24735922 DOI: 10.1016/j.ccr.2014.03.005]
 - 28 **Jiang Y**, Han QJ, Zhang J. Hepatocellular carcinoma: Mechanisms of progression and immunotherapy. *World J Gastroenterol* 2019; **25**: 3151-3167 [PMID: 31333308 DOI: 10.3748/wjg.v25.i25.3151]
 - 29 **Rebouissou S**, Franconi A, Calderaro J, Letouze E, Imbeaud S, Pilati C, Nault JC, Couchy G, Laurent A, Balabaud C,

- Bioulac-Sage P, Zucman-Rossi J. Genotype-phenotype correlation of CTNNB1 mutations reveals different β -catenin activity associated with liver tumor progression. *Hepatology* 2016; **64**: 2047-2061 [PMID: [27177928](#) DOI: [10.1002/hep.28638](#)]
- 30 **Sempoux C**, Gouw ASH, Dunet V, Paradis V, Balabaud C, Bioulac-Sage P. Predictive Patterns of Glutamine Synthetase Immunohistochemical Staining in CTNNB1-mutated Hepatocellular Adenomas. *Am J Surg Pathol* 2021; **45**: 477-487 [PMID: [33560657](#) DOI: [10.1097/PAS.0000000000001675](#)]
 - 31 **Julien C**, Le-Bail B, Ouazzani Touhami K, Frulio N, Blanc JF, Adam JP, Laurent C, Balabaud C, Bioulac-Sage P, Chiche L. Hepatocellular Adenoma Risk Factors of Hemorrhage: Size Is Not the Only Concern! *Ann Surg* 2021; **274**: 843-850 [PMID: [34334644](#) DOI: [10.1097/SLA.0000000000005108](#)]
 - 32 **Machado MV**, Diehl AM. Hedgehog signalling in liver pathophysiology. *J Hepatol* 2018; **68**: 550-562 [PMID: [29107151](#) DOI: [10.1016/j.jhep.2017.10.017](#)]
 - 33 **Sala M**, Gonzales D, Leste-Lasserre T, Dugot-Senat N, Paradis V, Di Tommaso S, Dupuy JW, Pitard V, Dourthe C, Sciarra A, Sempoux C, Ferrell LD, Clouston AD, Miller G, Yeh MM, Thung S, Gouw ASH, Quaglia A, Han J, Huan J, Fan C, Crawford J, Nakanuma Y, Harada K, le Bail B, Castain C, Frulio N, Trillaud H, Possenti L, Blanc JF, Chiche L, Laurent C, Balabaud C, Bioulac-Sage P, Raymond AA, Saltel F. ASS1 Overexpression: A Hallmark of Sonic Hedgehog Hepatocellular Adenomas; Recommendations for Clinical Practice. *Hepatol Commun* 2020; **4**: 809-824 [PMID: [32490318](#) DOI: [10.1002/hep4.1514](#)]
 - 34 **Henriet E**, Abou Hammoud A, Dupuy JW, Dartigues B, Ezzoukry Z, Dugot-Senat N, Leste-Lasserre T, Pallares-Lupon N, Nikolski M, Le Bail B, Blanc JF, Balabaud C, Bioulac-Sage P, Raymond AA, Saltel F. Argininosuccinate synthase 1 (ASS1): A marker of unclassified hepatocellular adenoma and high bleeding risk. *Hepatology* 2017; **66**: 2016-2028 [PMID: [28646562](#) DOI: [10.1002/hep.29336](#)]
 - 35 **Lehrke HD**, Van Treeck BJ, Allende D, Denham LJ, Gonzalez RS, Moreira RK, Mounajjed T, Naini BV, Smoot RL, Zreik RT, Jenkins S, Graham RP. Does Argininosuccinate Synthase 1 (ASS1) Immunohistochemistry Predict an Increased Risk of Hemorrhage for Hepatocellular Adenomas? *Appl Immunohistochem Mol Morphol* 2020; **28**: 464-470 [PMID: [31135443](#) DOI: [10.1097/PAL.0000000000000774](#)]
 - 36 **Mounajjed T**, Yasir S, Aleff PA, Torbenson MS. Pigmented hepatocellular adenomas have a high risk of atypia and malignancy. *Mod Pathol* 2015; **28**: 1265-1274 [PMID: [26205181](#) DOI: [10.1038/modpathol.2015.83](#)]
 - 37 **Souza LN**, de Martino RB, Thompson R, Strautnieks S, Heaton ND, Quaglia A. Pigmented well-differentiated hepatocellular neoplasm with beta-catenin mutation. *Hepatobiliary Pancreat Dis Int* 2015; **14**: 660-664 [PMID: [26663015](#) DOI: [10.1016/s1499-3872\(15\)60381-2](#)]
 - 38 **Hoshino K**, Harimoto N, Muranushi R, Hagiwara K, Yamanaka T, Ishii N, Tsukagoshi M, Igarashi T, Watanabe A, Kubo N, Araki K, Tomomasa R, Nobusawa S, Aishima S, Nakashima O, Shirabe K. Unclassified hepatocellular adenoma with histological brown pigment deposition and serum PIVKA-II level elevation: a case report. *Surg Case Rep* 2020; **6**: 94 [PMID: [32382834](#) DOI: [10.1186/s40792-020-00853-6](#)]
 - 39 **Liu L**, Liao JZ, He XX, Li PY. The role of autophagy in hepatocellular carcinoma: friend or foe. *Oncotarget* 2017; **8**: 57707-57722 [PMID: [28915706](#) DOI: [10.18632/oncotarget.17202](#)]
 - 40 **Marsman DS**, Goldsworthy TL, Popp JA. Contrasting hepatocytic peroxisome proliferation, lipofuscin accumulation and cell turnover for the hepatocarcinogens Wy-14,643 and clofibrate. *Carcinogenesis* 1992; **13**: 1011-1017 [PMID: [1600604](#) DOI: [10.1093/carcin/13.6.1011](#)]
 - 41 **Salaria SN**, Graham RP, Aishima S, Mounajjed T, Yeh MM, Torbenson MS. Primary hepatic tumors with myxoid change: morphologically unique hepatic adenomas and hepatocellular carcinomas. *Am J Surg Pathol* 2015; **39**: 318-324 [PMID: [25602798](#) DOI: [10.1097/PAS.0000000000000382](#)]
 - 42 **Young JT**, Kurup AN, Graham RP, Torbenson MS, Venkatesh SK. Myxoid hepatocellular neoplasms: imaging appearance of a unique mucinous tumor variant. *Abdom Radiol (NY)* 2016; **41**: 2115-2122 [PMID: [27334021](#) DOI: [10.1007/s00261-016-0812-x](#)]
 - 43 **De Vos N**, Van der Meulen J, Van Der Linden M, Claes K, Candaele AS, Vanlander A, Troisi RI, Van Vlierberghe H, Smeets P, Van Dorpe J, Hoorens A. Myxoid hepatocellular adenoma, a rare variant of hepatocellular adenoma with distinct imaging features: A case report with immunohistochemical and molecular analysis and literature review. *Clin Res Hepatol Gastroenterol* 2021; **45**: 101478 [PMID: [32620388](#) DOI: [10.1016/j.clinre.2020.06.004](#)]
 - 44 **Rowan DJ**, Yasir S, Chen ZE, Mounajjed T, Erdogan Damgard S, Cummins L, Zhang L, Whitcomb E, Falck V, Simon SM, Singhi AD, Torbenson MS. Morphologic and Molecular Findings in Myxoid Hepatic Adenomas. *Am J Surg Pathol* 2021; **45**: 1098-1107 [PMID: [34232602](#) DOI: [10.1097/PAS.0000000000001711](#)]
 - 45 **Larson BK**, Guindi M. Applying criteria for hepatocellular neoplasm of uncertain malignant potential reclassifies more than half of hepatocellular adenomas. *Ann Diagn Pathol* 2021; **55**: 151833 [PMID: [34597957](#) DOI: [10.1016/j.anndiagpath.2021.151833](#)]
 - 46 **Choi WT**, Kakar S. Atypical Hepatocellular Neoplasms: Review of Clinical, Morphologic, Immunohistochemical, Molecular, and Cytogenetic Features. *Adv Anat Pathol* 2018; **25**: 254-262 [PMID: [29649004](#) DOI: [10.1097/PAP.0000000000000189](#)]
 - 47 **Bedossa P**, Burt AD, Brunt EM, Callea F, Clouston AD, Dienes HP, Goodman ZD, Gouw AS, Hubscher SG, Roberts EA, Roskams T, Terracciano L, Tiniakos DG, Torbenson MS, Wanless IR. Well-differentiated hepatocellular neoplasm of uncertain malignant potential: proposal for a new diagnostic category. *Hum Pathol* 2014; **45**: 658-660 [PMID: [24529331](#) DOI: [10.1016/j.humpath.2013.09.020](#)]
 - 48 **Cancer Genome Atlas Research Network**. Comprehensive and Integrative Genomic Characterization of Hepatocellular Carcinoma. *Cell* 2017; **169**: 1327-1341.e23 [PMID: [28622513](#) DOI: [10.1016/j.cell.2017.05.046](#)]
 - 49 **Calderaro J**, Nault JC, Balabaud C, Couchy G, Saint-Paul MC, Azoulay D, Mehdaoui D, Luciani A, Zafrani ES, Bioulac-Sage P, Zucman-Rossi J. Inflammatory hepatocellular adenomas developed in the setting of chronic liver disease and cirrhosis. *Mod Pathol* 2016; **29**: 43-50 [PMID: [26516697](#) DOI: [10.1038/modpathol.2015.119](#)]
 - 50 **Sasaki M**, Yoneda N, Sawai Y, Imai Y, Kondo F, Fukusato T, Yoshikawa S, Kobayashi S, Sato Y, Matsui O, Nakanuma Y. Clinicopathological characteristics of serum amyloid A-positive hepatocellular neoplasms/nodules arising in alcoholic

- cirrhosis. *Histopathology* 2015; **66**: 836-845 [PMID: [25318388](#) DOI: [10.1111/his.12588](#)]
- 51 **Xiang DM**, Sun W, Ning BF, Zhou TF, Li XF, Zhong W, Cheng Z, Xia MY, Wang X, Deng X, Wang W, Li HY, Cui XL, Li SC, Wu B, Xie WF, Wang HY, Ding J. The HLF/IL-6/STAT3 feedforward circuit drives hepatic stellate cell activation to promote liver fibrosis. *Gut* 2018; **67**: 1704-1715 [PMID: [28754776](#) DOI: [10.1136/gutjnl-2016-313392](#)]
 - 52 **Torbenson M**. Hepatic Adenomas: Classification, Controversies, and Consensus. *Surg Pathol Clin* 2018; **11**: 351-366 [PMID: [29751879](#) DOI: [10.1016/j.path.2018.02.007](#)]
 - 53 **Marrero JA**, Ahn J, Rajender Reddy K; American College of Gastroenterology. ACG clinical guideline: the diagnosis and management of focal liver lesions. *Am J Gastroenterol* 2014; **109**: 1328-47; quiz 1348 [PMID: [25135008](#) DOI: [10.1038/ajg.2014.213](#)]
 - 54 **European Association for the Study of the Liver (EASL)**. EASL Clinical Practice Guidelines on the management of benign liver tumours. *J Hepatol* 2016; **65**: 386-398 [PMID: [27085809](#) DOI: [10.1016/j.jhep.2016.04.001](#)]
 - 55 **Klomphehouwer AJ**, de Man RA, Dioguardi Burgio M, Vilgrain V, Zucman-Rossi J, Ijzermans JNM. New insights in the management of Hepatocellular Adenoma. *Liver Int* 2020; **40**: 1529-1537 [PMID: [32464711](#) DOI: [10.1111/liv.14547](#)]



Published by **Baishideng Publishing Group Inc**
7041 Koll Center Parkway, Suite 160, Pleasanton, CA 94566, USA

Telephone: +1-925-3991568

E-mail: bpgoffice@wjgnet.com

Help Desk: <https://www.f6publishing.com/helpdesk>

<https://www.wjgnet.com>

