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**Occurrence of MYD88L265P and CD79B mutations in diffuse large b cell lymphoma with bone marrow infiltration: A case report**

Huang WY *et al*. MYD88L265P and CD79B in DLBCL

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**Abstract**

BACKGROUND

Over the past 20 years, we have gained a deep understanding of the biological heterogeneity of diffuse large B cell lymphoma (DLBCL) and have developed a range of new treatment programs based on the characteristics of the disease, bringing us to the era of immune-chemotherapy. However, the effectiveness and molecular mechanisms of targeted-immunotherapy remain unclear in DLBCL. Targeted-immunotherapy may be beneficial for specific subgroups of patients, thus requiring biomarker assessment.

CASE SUMMARY

Here, we report a case of MCD subtype DLBCL with MYD88L265P and CD79B mutations, considered in the initial stage as lymphoplasmic lymphoma (LPL) or Waldenstrom macroglobulinemia (WM). Flow cytometry supported this view; however, the immunohistochemical results of the lymph nodes overturned the above diagnosis, and the patient was eventually diagnosed with MCD subtype DLBCL. The presence of a monoclonal IgM component in the serum and infiltration of small lymphocytes with a phenotype compatible with WM into the bone marrow led us to propose a hypothesis that the case we report may have transformed from LPL/WM.

CONCLUSION

This highlights the possible transformation from WM to DLBCL, CD79B mutation may be a potential biomarker for predicting this conversion.

**Key Words:** Bone marrow infiltration; Case report; CD79B; Diffuse large B cell lymphoma; Ibrutinib; MYD88L265P

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**Core Tip:** This report highlights the possible transformation from Waldenstrom macroglobulinemia (WM) to diffuse large B cell lymphoma (DLBCL). Bone marrow infiltration by small lymphocytes, with an immunophenotype compatible with WM and the presence of MYD88L265P and CD79B mutations, support the hypothesis that the case may have transformed from lymphoplasmic lymphoma/WM. The CD79B mutation may be a potential biomarker for predicting the conversion of WM to DLBCL. Understanding the biology and mechanisms behind this process is important to identify susceptible patients. Frail patients could benefit from personalized low toxicity therapeutic approaches based on their mutational profile.

**INTRODUCTION**

The cell of origin (COO) is determined by the algorithm of Hans; thus, diffuse large B cell lymphoma (DLBCL) cases can be classified as germinal center B-cell-like (GCB) or non-GCB[1]. With the progression of clinical practice, COO has been found to be insufficient to explain different treatment responses. The biological heterogeneity of DLBCL may be directly due to its various genetic abnormalities related to its pathogenesis. Common genetic abnormalities in DLBCL include MYD88L265P, CD79B, BCL2, BCL6, EZH2, NOTCH2, and NOTCH1[2]. Among them, reports of CD79B and MYD88L265P are not uncommon, which are significantly related to the molecular typing of DLBCL.

CD79 is a transmembrane protein comprising two distinct peptide chains, CD79A and CD79B, which form a molecular complex with B-cell receptor (BCR). The cytoplasmic ends of the two peptide chains of CD79 contain an immunoreceptor tyrosine-based activation motif, which is involved in activating the nuclear factor kappa light chain enhancer of activated B cell (NF-κB) signaling pathway during antigen-mediated BCR activation[3]. Myeloid differentiation primary response 88 (MYD88) is a soluble adaptor protein in the cytoplasm; it belongs to the Toll/interleukin-1 receptor and death domain family members and mediates NF-κB signaling[4]. A mutation in CD79B was detectable in 30% of patients with ABC DLBCL and 3% of those with GCB DLBCL. In addition, MYD88 mutations were detected in 28% of ABC DLBCL[5]. They play an important role in DLBCL evolution. In recent years, with extensive research on multi-platform genomes, DLBCL was divided into seven genetic subtypes[2]; DLBCL with MYD88L265P and CD79B mutations were defined as the MCD genetic subtype, 42% of which had double concomitant mutations, mostly observed in the ABC subtype[2].

Here, we present a rare DLBCL case with mutations positive for MYD88L265P and CD79B, named MCD genetic subtype, characterized by discordant bone marrow (BM) infiltration, which may have transformed from indolent lymphoma.

**CASE PRESENTATION**

***Chief complaints***

fever and fatigue.

***History of present illness***

An 84-year-old bedridden man presented to the clinic with complaints of fever and fatigue and was hospitalized on January 2021.

***History of past illness***

He had been taking medication for hypertension and diabetes for more than 10 years and underwent interventional treatment for coronary atherosclerotic heart disease 4 years prior.

***Personal and family history***

He had no history of viral hepatitis B or family history of cancer.

***Physical examination***

Rales could be heard in the lungs, and edema was visible in the lower extremities.

***Laboratory examinations***

A laboratory exam revealed normocytic anemia (hemoglobin 66 g/L), elevated C reactive protein (169 mg/L), and hydrogen hexachloro platinum (III).

***Imaging examinations***

B mode ultrasonography showed several swollen cervical lymph nodes, with the largest being 1.7 cm × 1.1 cm with an abnormal structure. An abdominal computed tomography scan with contrast revealed pulmonary infection, no evidence of splenomegaly, and multiple enlarged lymph nodes in the mediastinum and abdominal region, with a maximum size of 2.5 cm × 2.5 cm (Figure 1). We completed the routine examination of the BM, and no abnormal cells were detected in the smear. However, using BM as a specimen, flow cytometry identified a group of abnormal monoclonal B cells that showed restricted expression of the intracellular kappa light chain. This group of B cells tested positive for CD19, CD20, and CD79b and were negative for CD5, CD10, CD23, and FMC7 (Figure 2). The morphology of the BM biopsy suggested that small B-cell lymphomas were scattered, indicating that the disease involved the BM (Figure 3).

In the case of monoclonal small B-lymphocyte tumors, chronic lymphocytic leukemia (CLL), lymphoplasmacytic lymphoma (LPL), and Waldenstrom macroglobulinemia (WM) should be considered. However, patients with CLL usually also express CD5 and CD23, and the peripheral blood is involved; this patient did not have these characteristics. Our case was characterized by IgM monoclonal gammopathy without plasmacytoid changes. To further confirm the diagnosis, we performed a biopsy of the patient’s cervical lymph nodes. The immunohistochemical results of the lymph nodes are as follows (Figure 4): Bcl-2(+); Bcl-6(+); CD10(-); CD20(+); CD3(-); CD5(-); CMYC(40%); CyclinD1(-); Ki67(70%); MUM-1(+); P53(60%).

**FINAL DIAGNOSIS**

The immunohistochemical results support the diagnosis of subtype activated B-cells like that of DLBCL [stage IV B, National Comprehensive Cancer Network International Prognostic Index (IPI) ≥ 6, Eastern Cooperative Oncology Group performance score = 4]. Interestingly, the patient had MYD88L265P and CD79B mutations.

**TREATMENT**

Ibrutinib is an irreversible small-molecule BTK inhibitor and a B lymphocyte signaling protein, which can effectively inhibit the proliferation and survival of malignant B lymphocytes[6]. Studies have found that compared with GCB subtypes, ibrutinib has a better effect on patients with ABC DLBCL[7]. More importantly, the combined application of ibrutinib and rituximab plus cyclophosphamide, doxorubicin, vincristine, and prednisone can significantly increase the toxicity of the drug for patients aged ≥60 years, which in turn affects the therapeutic effect[8]. The patient could not tolerate chemotherapy. After a comprehensive assessment, the patient was treated with I-R (ibrutinib 420 mg/d combined with rituxan 375 mg/m2).

**OUTCOME AND FOLLOW-UP**

Unfortunately, due to financial pressure and being long-term bedridden, this patient eventually refused to receive further treatment and died two months later.

**DISCUSSION**

Multiple studies have shown that the MYD88 mutation is present in 90% of patients with LPL/ WM; however, it is not specific to LPL/WM and is also present in DLBCL. Further research confirmed that the MYD88L265P mutation is unique to ABC and rarely occurs in GCB or primary mediastinal diffuse large B-cell lymphoma[5]. Studies have reported that continuous excessive activation of the NF-κB signaling pathway is characteristic of ABC-DLBCL. Dubois *et al*[9] further emphasized that activation of the NF-κB signaling pathway in the ABC subtype is related to the L265P mutation site, whereas non-L265P mutants harbor a mutational profile more similar to GCB-DLBCL. The impact of MYD88L265P and CD79B mutations on the prognosis of the ABC type is worth discussing, and most reports have shown that these two mutations negatively affect patient outcomes. One study showed that MYD88L265P and CD79B increase the risk of recurrence and progression. In addition, detection of the MYD88L265P mutation can effectively improve the predictive performance of the IPI scores[10]. Using next generation sequencing to detect 361 cases of DLBCL, a study showed that CD79B and MYD88L265P synergistically enhance activation of the NF-κB pathway[9]. Wilson *et al*[7] designed a phase II clinical trial involving 80 patients with relapsed or refractory DLBCL; ibrutinib had a good therapeutic effect on 80% of patients with CD79b and MYD88L265P dual mutations, whereas the seven patients with MYD88L265P mutations/CD79B wild-type cases did not show any response (0/7), suggesting that the MYD88L265P/CD79B dual mutation DLBCL may represent a unique group with stronger sensitivity to BTK inhibitors, which may be related to the CD79B-dependent BCR activation pathway.

The mechanism of the emergence of monoclonal IgM is unclear; it may be related to the differentiation of plasma cells in the BM[11]. Cox *et al*[12] analyzed 151 patients with DLBCL and found that 17 cases (11.2%) had a serum monoclonal IgM component, although none were associated with MYD88L265P mutations, which indicates there is no necessary connection between [monoclonal](C:\\Users\\Lian-Sheng Ma\\AppData\\Local\\Program Files (x86)\\Youdao\\Dict\\7.5.2.0\\resultui\\dict\\result.html?keyword=monoclonal immunoglobulin&lang=en) IgM and MYD88L265P. Cho *et al*[13] studied the relationship between the clonal status of monoclonal immunoglobulin gene rearrangement and histological B cell aggregation in the BM. The results showed that of the 394 patients with DLBCL, 32 patients had BM invasion, and only two patients with large B-cell lymphoma had no gene rearrangement detected[13]. This suggests that patients with BM invasion were more likely to have monoclonal immunoglobulin gene rearrangements. However, there is no correlation with the COO[13]. Although BM infiltration was found in our case, it showed a histological inconsistency compared with the immunohistochemistry of the peripheral lymph nodes. Regarding the biological and clinical impact, concordant BM infiltration was associated with lower progression free survival and a higher incidence of central nervous system relapse independent of the COO and IPI; therefore, the prognostic impact of discordant BM infiltration could be limited to non-CGB cases[14].

The most interesting part of this case is the possible transformation from WM to DLBCL. We support this hypothesis on two pillars: the presence of a monoclonal IgM component in the serum and infiltration of small lymphocytes with a phenotype compatible with WM into the BM. Castillo *et al*[15] retrospectively analyzed 1466 patients with WM; a total of 20 patients underwent histological transformation. Interestingly, all 20 patients had tissue transformation to DLBCL and had a high IPI score. The median survival time after transformation was not more than 3 years. Among them, 13 patients were tested for Ki67 expression, and it was found that their median value-added index was 90% (range 50%-99%), which suggests that the malignancy in these patients is higher. Two cases in this study were tested for MYD88L265P mutations before and after histological transformation, and the results were positive[15]. This also suggests that ibrutinib may be a potential treatment option for these histological conversion cases. Regarding the biological analysis of patients with diffuse large B lymphoma with BM infiltration, 24% of patients have discordant BM involvement, which manifested as infiltration by small cells forming lymphoid aggregates. This group, classified by flow cytometry (FCM), showed a wide variety of indolent B-cell lymphomas, although only one case showed DLBCL[14]. Genomic analysis technology can further determine whether this group of patients has transformed from indolent lymphoma[16]. As the transformation from WM to DLBCL is very rare, relevant literature on this was collected (Table 1).

The other interesting hypothesis concerns the role of mutations in CD79B in the transformation to DLBCL. A small sample study found that CD79B mutations may be a potential biomarker for predicting the conversion of WM to DLBCL[16]. We propose a transformation hypothesis on the basis of the antiapoptotic NF-κB pathway. The mutation of MYD88 is the most important pathogenic mechanism of LPL/WM. When LPL/WM acquires the mutation of CD79B, the reduced activity of LYN contributes to their increased surface BCR expression and constitutive BCR signaling. Chronically active BCR signaling and MYD88L265P-dependent signaling synergy promotes the conversion of WM to DLBCL (Figure 5). However, the genetic pathogenesis is a complex process that may incorporate other genetic changes to facilitate its transition to DLBCL. Schmitz *et al*[17] proposed the classification of genetic subtypes that uncovered the interrelationship between this genetic nosology and the oncogenic signaling pathway. MYD88L265P and CD79B played an important role in the evolution of WM to DLBCL, which fits the genetic characteristics of the MCD genetic subtype. It further explains why the MCD genetic subtype responds to ibrutinib[7].

In summary, with the development of genetic testing technology, research has further focused on the classification of DLBCL and its pathogenic mechanism. In the era of immune-chemotherapy, the molecular classification of DLBCL appears to offer greater prognostic interest than IPI. Furthermore, the molecular profile could help us in the choice of the optimal therapy. Ibrutinib appears a good option in patients with MCD molecular subtype DLBCL: MYD88+ CD79B+. Bone marrow infiltration by small lymphocytes, with an immunophenotype compatible with WM and the presence of MYD88L265P and CD79B mutations, supports the hypothesis that the case may have transformed from LPL/WM. Furthermore, frail patients could benefit from personalized low toxicity therapeutic approaches based on their mutational profile.

**CONCLUSION**

The CD79B mutation may be a potential biomarker for predicting the conversion of WM to DLBCL. Understanding the biology and mechanisms behind this process is important in identifying susceptible patients.

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**Footnotes**

**Informed consent statement:** Written consent was obtained from the patient’s family to participate in the study.

**Conflict-of-interest statement:** The authors declare that they have no conflicts of interest to disclose.

**CARE Checklist (2016) statement:** The authors have read, prepared and revised the manuscript according to the CARE Checklist (2016).

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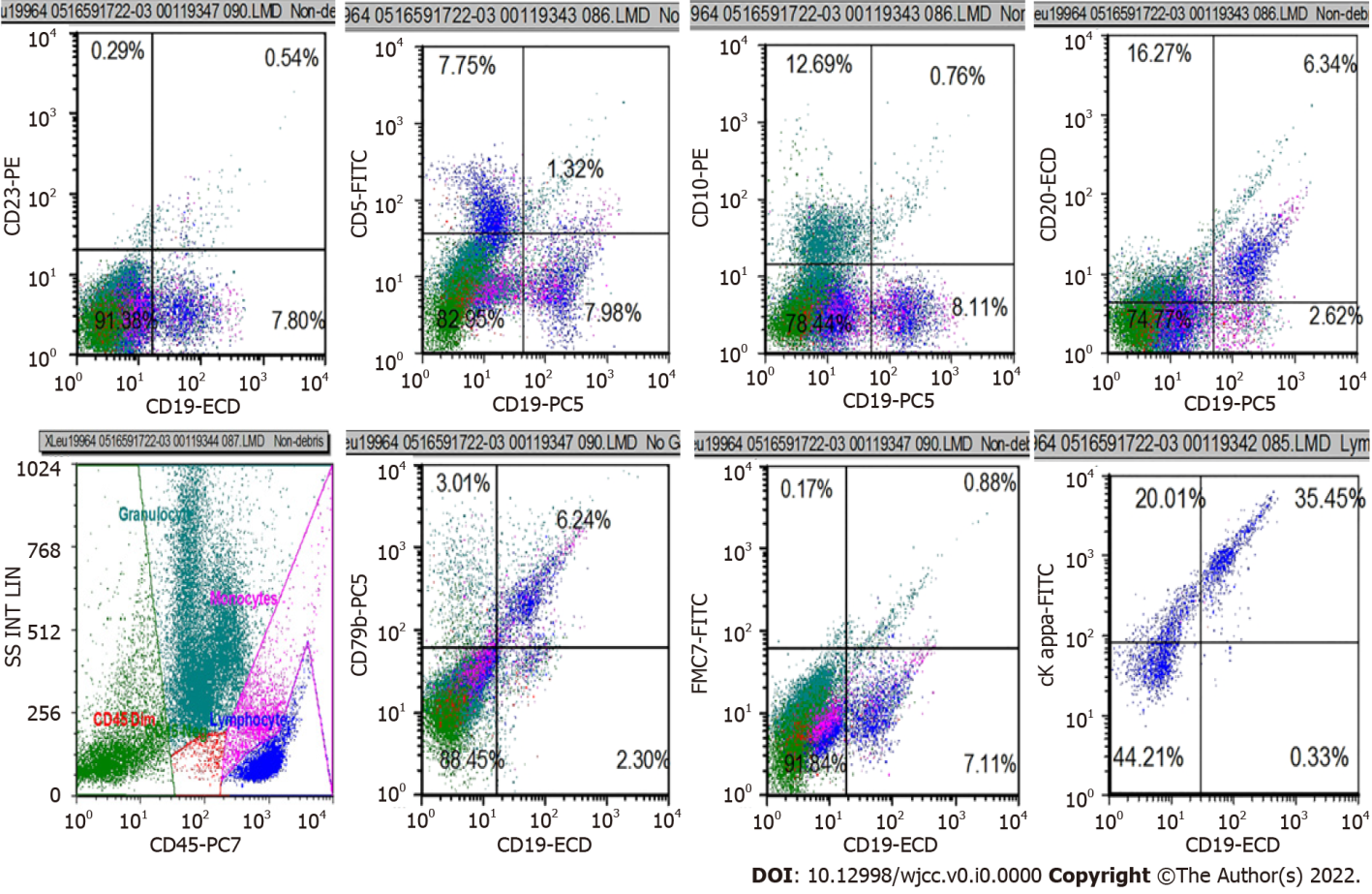
Grade D (Fair): 0

Grade E (Poor): 0

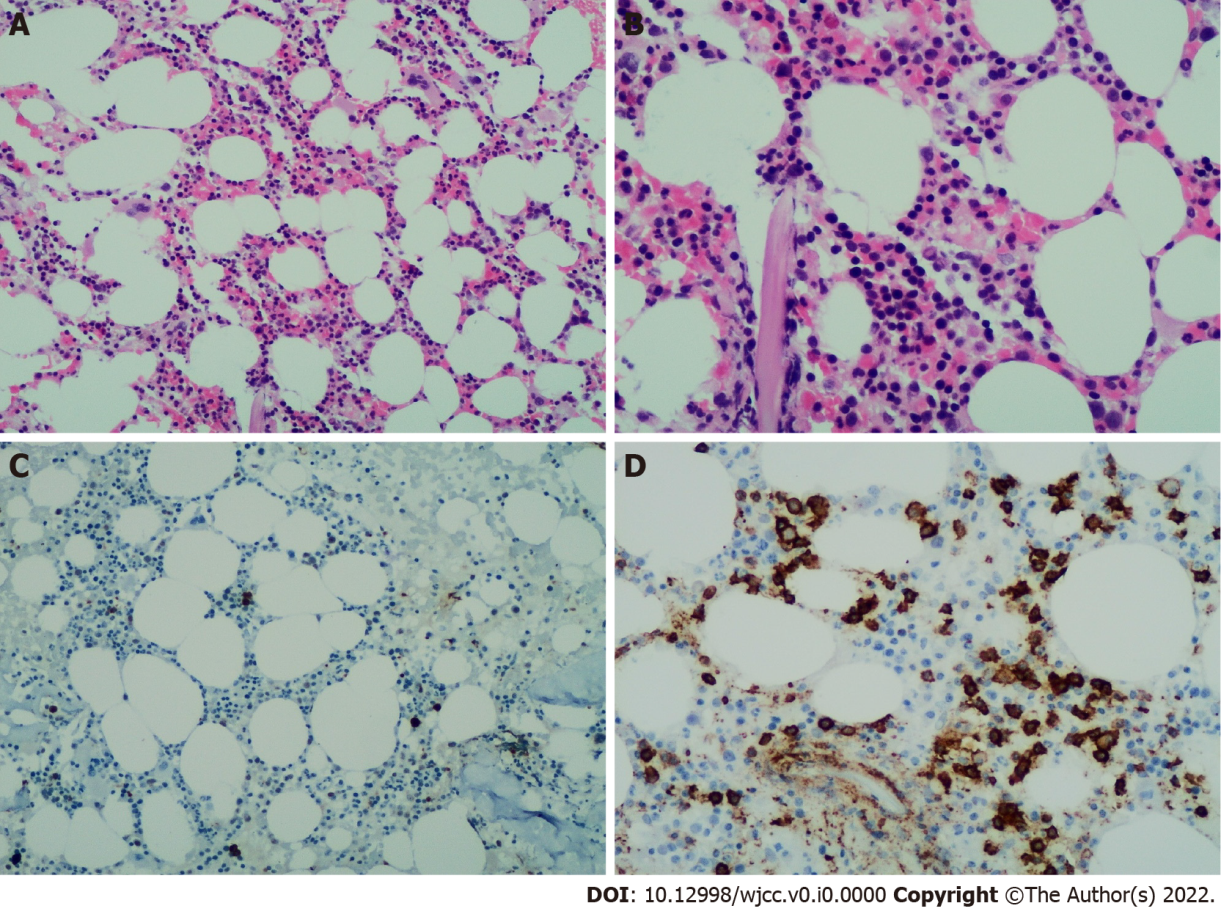
**P-Reviewer:** Baysal M, Turkey; Watanabe T, Japan **S-Editor:** Ma YJ **L-Editor:** A **P-Editor:** Ma YJ

**Figure Legends**

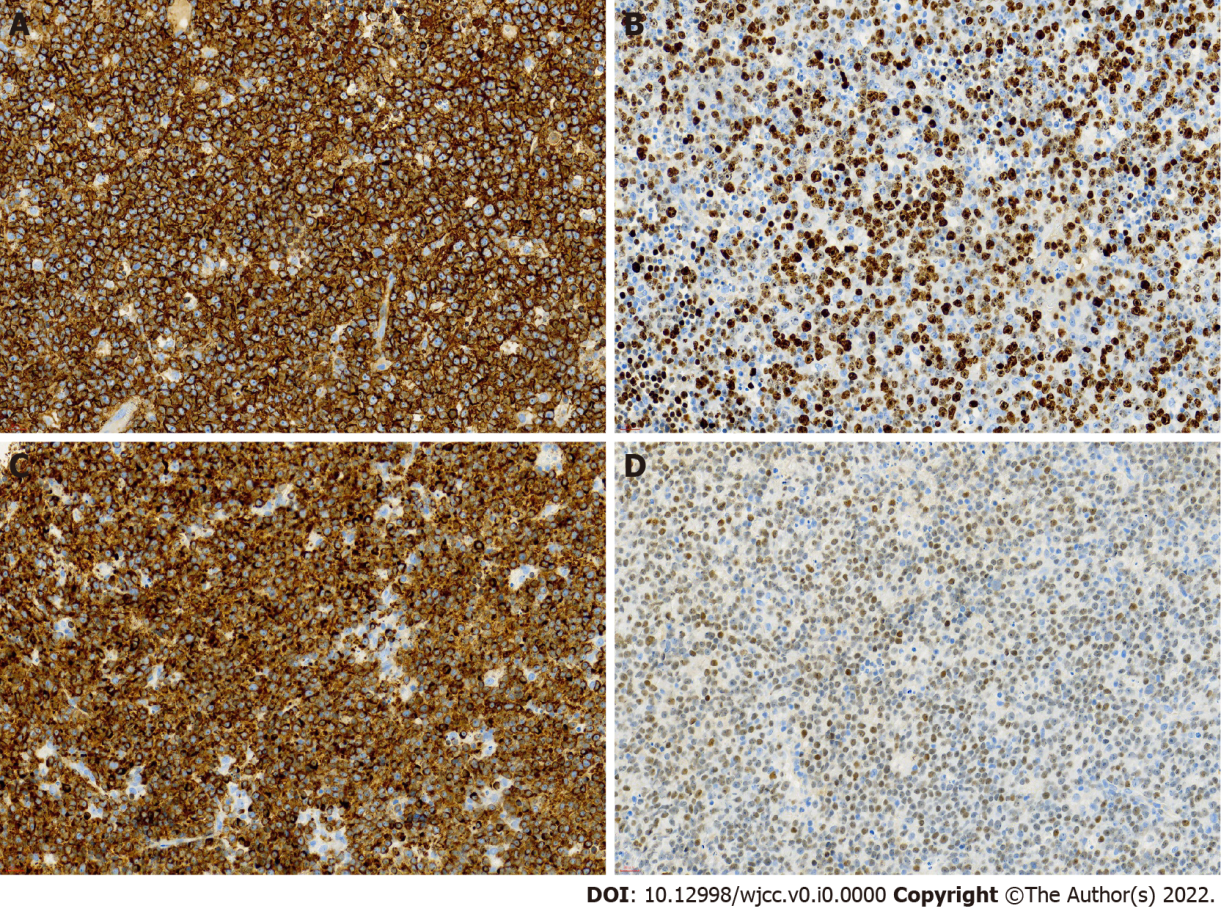


**Figure 1 Abdominal computed tomography scan with contrast revealed multiple enlarged lymph nodes in the mediastinum and abdominal, with a maximum size of 2.5 cm × 2.5 cm.** Arrows indicate multiple enlarged lymph nodes.****

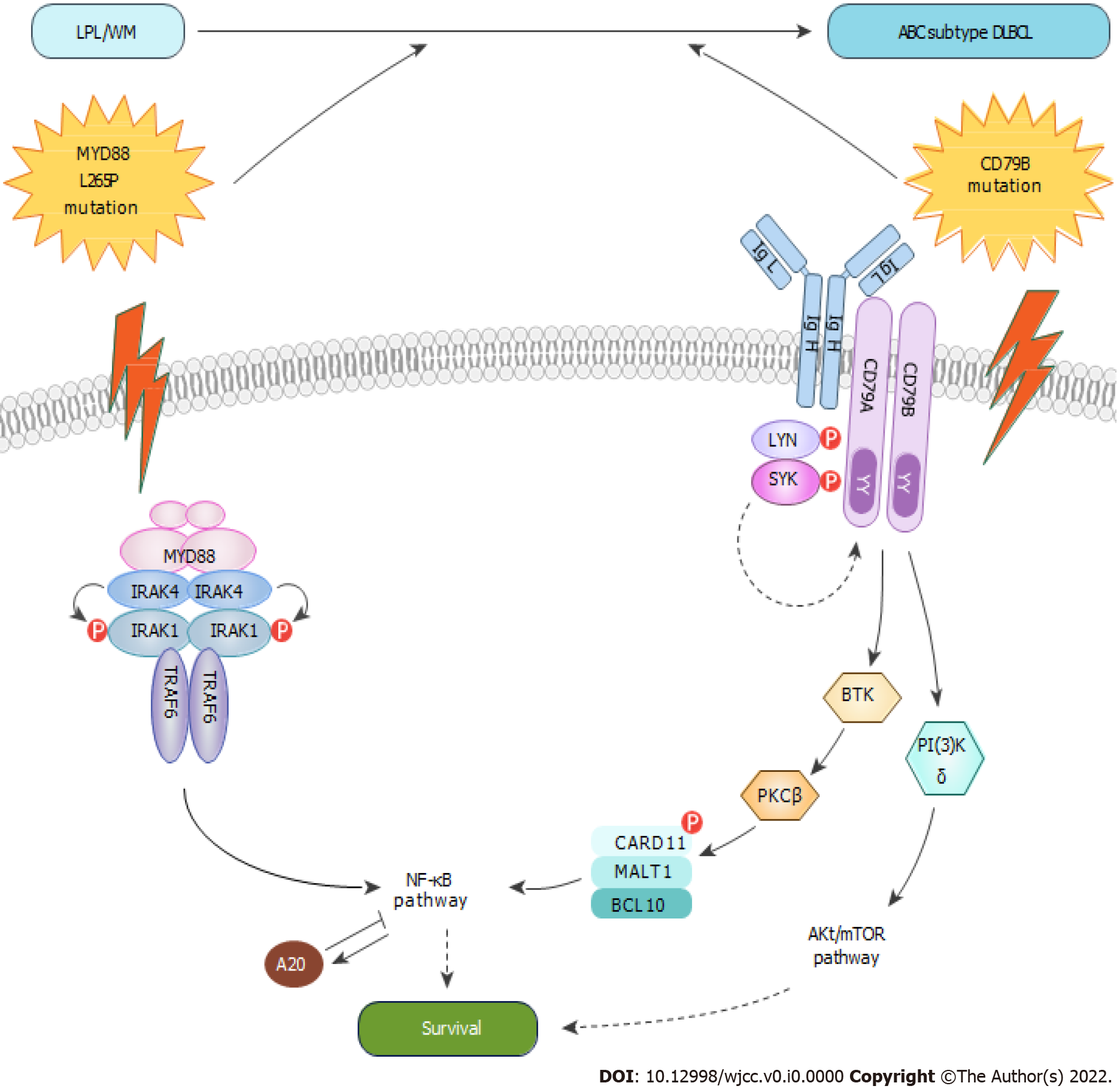
**Figure 2 Flow cytometry shows monotypic kappa light chain restricted B cells that tested positive for CD19, CD20, and CD79b, and negative for CD5, CD10, CD23, and FMC7.**

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**Figure 3 B lymphocytes had a scattered distribution (about 10%).** A, b: Histomorphology suggests that small B lymphocytes infiltrated the bone marrow (A, 10 x; B, 20 x); C, D: The expression of CD20 can be detected by immunohistochemistry (D, 20 x), and the positive rate of Ki-67 is less than 5% (C, 10 x).



**Figure 4 The expression of CD20 can be detected by immunohistochemistry (A, 40 x), Ki67 staining showed almost 70% proliferation index (B, 40 x), tumor cells were postitive for BCL-2 (C, 40 x) and BCL-6 (D, 40 x).**

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**Figure 5 nuclear factor-κB activation through the classical pathway is a hallmark of the ABC diffuse large B cell lymphoma subtype.** MYD88 is an adapter protein that couples TIR-containing receptors, regulating downstream signaling circuits, including the nuclear factor (NF)-κB. MYD88 coordinates the IRAK family kinases into a helical signaling complex through the interaction with the IRAK kinase. Phosphorylation of IRAK1 by IRAK4 will allow for the recruitment of the ubiquitin ligase TRAF6 and the activation of the downstream pathways. The mutation in MYD88L265P forms a stable, phosphorylated form of IRAK1, which upregulates gene expression signatures of NF-κB. The BCR consists of IgL and IgH chains that are noncovalently coupled to the CD79B (Ig-β) and CD79A (Ig-α) subunits, which regulate BCR surface trafficking, internalization, and expression. Upon antigen encounter, the BCR, CD79A and CD79B transmit signals to multiple downstream signaling pathways. Once BTK is recruited to the BCR signaling complex, LYN or SYK can phosphorylate and activate BTK. In turn activate PKCβ, leading to phosphorylation of CARD11 and activation of NF-κB. CBM complex, a signaling hub consisting of CARD11, BCL10, MALT1, and other proteins, which is required for the activation of classical NF-κB pathway in lymphocytes.

**Table 1 Summary of previously published series of cases**

|  |  |  |  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- |
| Ref. | **No. of case** | **A/G** | **BM** | **Extramedullary site** | **Histological type** | **Staging** | **Therapy** | **Interval (mo)1** | **Ending** | **Survival (mo)** |
| Uchino *et al*[18] | 1 | 55/M | + | Spleen | No  specified | IV B | R-CHOP | 408 | CR | > 17 |
| Owen *et al*[19] | 2 | 80/M | + | Mesenteric mass | ABC | IV | R-CVP | 36 | Dead | No specified |
| Owen *et al*[19] | 3 | 63/M | + | Lymphadenopath | GCB | IV | R-CHOP | 84 | PR | No specified |
| Shiseki *et al*[20] | 4 | 63/M | + | Lymph node | No specified | IV | THP-COP | 36 | CR | > 60 |
| Kikukawa *et al*[21] | 5 | 60/F | + | Brain | ABC | IV | R-MPV WBRT+Ara-C | 72 | PR | > 6 |
| Okolo *et al*[22] | 6 | 69/M | + | Retroperitoneal mass | No specified | IV | R-CHOP DRC | Unknown | CR | No specified |
| Kobayashi *et al*[23] | 7 | 75/M | + | Liver and ileum | GCB | IV | R-CHOP | 120 | PR | No specified |
| Elimimian *et al*[24] | 8 | 60/M | + | Inguinal lymph node | ABC | IV A | O+CHOP DHAP | 168 | Dead | 4 |

1Interval from the diagnosis of lymphoplasmic lymphoma/Waldenstrom macroglobulinemia to the diagnosis of diffuse large B cell lymphoma.

BM: Bone marrow; CR: complete response; PR: partial response; R-CVP: rituximab, cyclophosphamide, vincristine, prednisone; R-CHOP: rituximab, cyclophosphamide, doxorubicin, vincristine, prednisolone; THP: epirubicin; R-MPV: methotrexate, vincristine, procarbazine; WBRT: whole-brain radiation; Ara-C: cytarabine; DRC: dexamethasone, rituximab, cyclophosphamide; O: obinutuzumab; DHAP: cisplatin, cytarabine, dexamethasone.