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**Gastrointestinal tumors in transplantation: Two case reports and review of literature**

Stammler R *et al*. GISTs and transplantation

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**Abstract**

BACKGROUND

Gastrointestinal stromal tumors (GISTs) are the most common mesenchymal tumors of the gastrointestinal tract. As most of them harbor a*KIT* mutation (75%), selective kinase inhibitors are the therapeutic option and show a sustained objective response among patients with metastatic or unresectable GISTs. A well-known higher risk of neoplasm has been described among renal transplant recipients (RTRs). Nevertheless, only few cases of GIST onset among transplant patients have been reported in the literature.

CASE SUMMARY

Here, we describe 2 cases of gastric GIST occurring during the follow-up of RTRs. We also review the existing literature concerning GIST occurrence in transplant patients. In total and in association with our 2 cases, 16 patients have been reported. The median age was 59.5 years and 69% were male. With a median tumor size of 45 mm, no patient displayed metastatic dissemination at diagnosis. Time from transplantation to diagnosis was highly variable between 5 mo and 21 years. Histopathological data mostly revealed high risk of progression (43%). Death increased to 29% during follow-up. Surgical treatment was systematically performed when the tumor was operable (94%). The use of adjuvant therapy was uncommon (19%).

CONCLUSION

GISTs represent rare but potentially severe malignant complication among transplant patients.

**Key Words:** Gastrointestinal stromal tumors; Imatinib mesylate; Transplantation; Kidney transplantation; Proto-oncogene protein c-KIT; Case report

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**Core Tip:** Although a well-known higher risk of neoplasm has been described among renal transplant recipients (RTRs), few cases of gastrointestinal stromal tumors (GISTs) have been reported. We describe 2 cases of gastric GIST among RTRs and provide a review of the literature. We report 16 patients with a median age of 59.5 years, and 69% were male. No patient displayed metastasis at diagnosis. Time from transplantation to diagnosis varied between 5 mo and 21 years. Histopathology revealed high risk of progression (43%). Death increased to 29%. Surgical treatment was commonly performed (94%). The use of adjuvant therapy was uncommon (19%).

**INTRODUCTION**

Gastrointestinal stromal tumors (GISTs) are the most common mesenchymal tumors of the gastrointestinal (GI) tract[1]. GISTs arise from interstitial cells of Cajal (ICC), which are specialized mesenchymal cells located within the muscle of the GI tract. ICC play a critical role in regulating smooth muscle function and GI tract motility[2]. GISTs are mainly located in the stomach (55%) or the small bowel (30%). About 10% to 47% of patients have metastatic disease at diagnosis[3-5]. About 95% of GISTs display positive staining for the receptor tyrosine kinase KIT (or CD117), 75% of these tumors harbor a *KIT* gene mutation and 10% a platelet-derived growth factor receptor A (*PDGFRA*) gene mutation[6]. Among KIT-negative GISTs, immunohistochemical expression of discovered on GIST-1 (DOG-1) was found in 76% of the cases[7]. Consequently, selective tyrosine kinase inhibitors targeting KIT receptor have been used. The first one, imatinib mesylate (Gleevec®; Novartis, Basel, Switzerland), has shown a sustained objective response in a phase III trial among patients with metastatic or unresectable GISTs in immunocompetent patients[8].

In renal transplant recipients (RTRs), an increased risk of cancer has been reported especially for non-melanoma skin cancer, virus-associated cancer and lymphoproliferative disorders[9]. Currently, malignancy represents a major cause of mortality among RTRs[10]. Nonetheless, only few cases of GIST have been reported among transplant patients. Overall, 8 cases of GIST[11-17] and 2 cases of extra GIST (EGIST)[14,18] have previously been reported in RTRs and respectively 3 cases[19-21] and 1 case[22] in liver transplant recipients.

We report 2 cases of GIST occurring in RTRs and provide a review of the existing literature concerning GIST occurrence in transplant patients.

**CASE PRESENTATION**

***Chief complaints***

**Case 1:** A 60-year-old Caucasian man without any symptoms.

**Case 2:** A 56-year-old Caucasian man presented with upper GI hemorrhage.

***History of present illness***

**Case 1:** Hepatic magnetic resonance imaging (MRI) was performed to explore abnormal hepatic tests. MRI revealed a 32 mm spherical tumor of the lesser curvature of the stomach.

**Case 2:** The upper GI hemorrhage led to gastric endoscopy, which revealed a spherical gastric tumor in the fundus.

***History of past illness***

**Case 1:** He had end-stage renal disease with a kidney biopsy compatible with nephronophthisis despite negative screening for mutation in hepatocyte nuclear factor 1 beta (*HNF1B*) gene. Hemodialysis was initiated in 2016. In October 2019, he received a kidney transplant from a deceased donor. The initial immunosuppressive therapy combined basiliximab, steroids, tacrolimus, and everolimus. Renal function at hospital discharge was 94 µmol/L, (normal range 53 µmol/L to 97 µmol/L). Initial maintenance immunosuppressive therapy associated steroids, tacrolimus, and everolimus. Due to relapsing lymphocele, everolimus was switched to mycophenolate mofetil (MMF). Moreover, a pre-existing mild cytolysis and cholestasis worsened after transplantation leading to the discontinuation of cotrimoxazole and MMF, which were replaced by atovaquone and belatacept (NULOJIX®; Bristol-Myers Squibb, New York, NY, United States), respectively.

**Case 2:** The patient developed end-stage renal disease of unknown origin. He received a kidney transplantation from a deceased donor. Due to preformed donor specific antibodies (anti-Cw15, mean fluorescence intensity of 6130) on the day of transplantation, induction immunosuppressive therapy combined basiliximab, steroids, MMF, cyclosporine, and intravenous immunoglobulins. At 10 d after surgery, a kidney biopsy was performed due to delayed graft function. It revealed acute tubular necrosis associated with possible acute humoral rejection (g1 cpt0 v0 i0 t0 according to Banff’s classification[23,24], C4d immunostaining was negative). A treatment with high dose steroids, five plasma exchanges and rituximab[25] was initiated allowing improvement of renal function with a nadir in serum creatinine level of 170 µmol/L. Maintenance immunosuppressive therapy included steroids, cyclosporine, and MMF.

***Personal and family history***

**Case 1:** His other past medical history consisted in nonalcoholic steatohepatitis, Hashimoto’s thyroiditis, and hypertension.

**Case 2:** The patient had no significant personal or family history.

***Physical examination***

**Case 1:** On admission, physical examination was unremarkable.

**Case 2:** Physical examination was unremarkable except for hematemesis.

***Laboratory examinations***

**Case 1:** The patient had mild cytolysis and cholestasis without any other biological abnormality.

**Case 2:** No abnormal blood test was noticed on admission.

***Imaging examinations***

**Case 1:** Body computed tomography (CT) scan confirmed the absence of metastatic dissemination.

**Case 2:** Body CT scan was consistent with local tumor without metastatic localizations.

***Initial diagnosis***

**Case 1:** Upper GI endoscopy found a 3 cm submucosal tumor of the lesser curvature of the stomach. Tumor biopsies were performed using endoscopic ultrasound guidance. Cytological examination revealed spindle-shaped cells that showed positive staining for c-KIT and DOG-1 in immunohistochemistry, confirming the diagnosis of GIST (Figure 1).

**Case 2:** The gastric endoscopy revealed a spherical gastric tumor in the fundus with a typical macroscopic aspect of GIST.

***Initial treatment***

**Case 1:**Partial gastrectomy was performed without complication.

**Case 2:** Partial gastrectomy was performed.

***Course of illness in the hospital***

**Case 1:** No complication associated with the GIST of its treatment was noticed.

**Case 2:** The patient was rapidly discharged after partial gastrectomy without complication.

**FINAL DIAGNOSIS**

***Case 1***

Histopathology revealed a 27 mm stromal tumor strongly positive for KIT and moderately positive for DOG-1 with a mitotic count of 2 mitosis for 5 mm2. The tumor harbored an exon 11 (p. Val559Ala c.1676T>C) *KIT* mutation[23].

***Case 2***

Histopathology report described a 51 mm GIST strongly positive for KIT harboring a mitotic count of 10 mitosis for 5 mm2. Of note, an exon 18 D842V *PDGFRA* mutation was identified.

**TREATMENT**

***Case 1***

Regarding the very low risk of progression, no adjuvant therapy was initiated.

***Case 2***

No adjuvant treatment was initiated at the time of diagnosis.

**OUTCOME AND FOLLOW-UP**

***Case 1***

The patient remains in remission at the 1-year follow-up.

***Case 2***

Two years later, a follow-up MRI revealed hepatic vascular nodules compatible with metastatic lesions. Treatment with imatinib mesylate was initiated. In the absence of a tumor response, imatinib was discontinued 4 mo later and sunitinib (SUTENT©; Bayer, Germany), an anti-angiogenic multikinase inhibitor (anti vascular endothelial growth factor-1, -2, -3, PDGFR-α,-β, c-KIT, fms-like tyrosine kinase 3, and RET) was introduced. Five months later, the onset of thrombopenia, neutropenia, and hepatic cytolysis led to replacement of sunitinib with regorafenib (STIVARGA©; Bayer Pharma AG, Germany), another multikinase inhibitor. Due to side effects and tumor progression, regorafenib was discontinued and dasatinib (SPRYCEL©; Bristol-Myers Squibb, New York, NY, United States) was introduced. Disease progression finally led to stopping all therapies in April 2019. Selective transarterial embolization was performed complicated with artery dissection of the kidney transplant requiring stent implantation. The patient was finally admitted with a clinical presentation of hydrops concomitant with acute renal injury and peritoneal carcinosis. The patient eventually died due to disease progression.

**DISCUSSION**

GISTs represent an uncommon malignant complication of immunosuppression state in solid organ transplantation. We describe 2 cases of typical GIST occurring early in the course of kidney transplantation. The first patient developed an isolated gastric GIST 5 mo after transplantation and the second 4 years after. Both were nonmetastatic at diagnosis although the second patient developed multiple hepatic metastasis 2 years after complete tumor resection. Of note, the mutation of *PDGFRA* D842V in the second case was associated with resistance to imatinib mesylate.

We looked for previously reported cases of GIST in the literature during the course of transplantation. We searched PubMed and Web of Science databases using the following Medical Subject Headings words: “Gastrointestinal stromal tumors” AND “Kidney transplantation” or “Gastrointestinal stromal tumors” AND “Transplantation.” Using these terms, we found 8 and 31 articles, respectively. Only 12 articles were analyzed. From 2007 to 2020, 14 cases of GIST have been reported in transplant recipients[11-22]. We excluded reports of GIST occurring among nontransplant or bone marrow transplant patients. We also excluded article types different than case reports or case series.

Table 1 summarizes the main features of these patients including the 2 cases described in the present manuscript. Tables 2 and 3 give details on the 14 cases reported. In our literature review, the typical patient profile was a male patient with a median age of 59.5-years-old, who developed large nonmetastatic gastric tumors (median size, 45 mm). The delay between transplantation and diagnosis was highly variable, ranging between 5 mo and 21 years. Histopathological data mostly revealed high risk of progression (42.8%) and death occurred in 29% of the cases during follow-up. Surgical treatment was systematically performed if the tumor features were suitable (94%). The use of adjuvant therapy was uncommon (19%).

Several prognostic classifications have been used to evaluate the risk of recurrence of GIST after surgery. In 2002, Fletcher *et al*[26] claimed size of the tumor and mitotic count, Miettinen and Lasota[27] in 2006 added tumor location and Joensuu *et al*[28] in 2012 adjoined rupture of the tumoral capsule and male gender. Heinrich *et al*[29] demonstrated that *PDGFRA* and *c-KIT* were mutually exclusive proto-oncogenic mutations with similar biologicals consequences, even if associated with different prognostics. Molecular predictors of response to imatinib have been widely studied. Underlying *KIT* or *PDGFRA* mutations are the strongest predictors of imatinib sensitivity[30]. Mutations directly located in the *PDGFRA* binding site of imatinib or inducing variations in tridimensional conformation of the tyrosine kinase receptor and subsequently hiding the binding site, may explain inefficacy of therapy. For instance, *KIT* exon 9 mutation is less sensitive to imatinib and *PDFGRA* exon 18 D842V mutations is associated with imatinib resistance. Nevertheless, these mutations have been correlated with opposite courses of the disease, indolent for *PDFGRA* exon 18 D842V mutation but aggressive for *KIT* exon 9 mutation[31]. These data should highlight the importance of molecular biomarkers to evaluate prognosis of GIST or EGIST at diagnosis.

Guidelines for the diagnosis, treatment, and follow-up of GIST have recently been published[32]. Management of local or locoregional disease should always aim for complete resection whenever possible. Otherwise, neoadjuvant treatment with imatinib for 6 to 12 mo should be used in case of sensitive mutation with an overall response rate of 50%[30]. Moreover, high-risk patients, as previously described, should receive adjuvant imatinib for a duration of 3 years[33]. Imatinib remains the first-line therapy for metastatic GIST. Several other targeted therapies such as sunitinib or regorafenib have emerged as second- or third-line treatment, and more recently avapritinib and ripretinib. Several biomarkers, such as *KIT* or *PDGFRA* mutations, are used as predictive factors for tumoral response to refine therapeutic strategies[32]. Data are missing concerning the level of tyrosine kinase inhibitors’ efficacy in transplanted patients.

Data about the management of immunosuppressive therapy after the diagnosis of GIST are scarce. As both imatinib mesylate and cyclosporin are extensively metabolized by cytochrome P450 3A4, interaction occurrence has been documented[12]. Reduction in the dosage of cyclosporin should be performed if this treatment is maintained. Mammalian target of rapamycin inhibitors (mTORis) have shown antiproliferative properties among transplant patients. Schöffski *et al*[34] highlighted the potential efficacy of association of everolimus and imatinib in imatinib-resistant GIST in a phase II trial. Cheung *et al*[14] reported a case of complete tumoral response with sirolimus in a transplant patient with imatinib-resistant GIST. Among the patients described in Tables 1 and 2, mTORis have been initiated or switched in 4 of them. Three of them were alive and relapse-free at last follow-up and the last patient died from pneumonia 2 years after GIST diagnosis.

This study had several limitations. First, the retrospective analysis of GIST cases impairs the reliability of the data. Very few cases of GIST occurring after solid organ transplantation have been described in the last 15 years reducing the significance of this literature review. Moreover, it was unclear if GIST was a *de novo* feature in our first patient because of the short delay (5 mo) between transplantation and tumor discovery. Unfortunately, the latest available CT scan was performed 7 years before the transplantation. However, some previous cases reported GIST onset within the 1st year following transplantation[14,18-22].

**CONCLUSION**

To conclude, GISTs represent rare but potentially severe malignant complication among transplant patients. Further analysis of prognosis value of new biomarkers should improve therapeutic strategies.

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**Footnotes**

**Informed consent statement:** Written informed consent was obtained from the patient for publication of this case report and any accompanying images.

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**Figure Legends**



**Figure 1 Histopathological features of case 1 gastrointestinal stromal tumor.** A: Hematoxylin and eosin staining showing exophytic development of a tumor from the mucosa musculus (magnification, 2.5 ×); B: Hematoxylin and eosin staining showing spindle cell morphology composed of relatively uniform cells arranged in short fascicles (magnification, 20 ×); C: Immunohistochemistry showing strong positive staining for the protooncogene c-KIT (white asterisk; magnification, 5 ×); D: Immunohistochemistry showing strong positive staining for discovered on gastrointestinal stromal tumor protein 1 (white asterisk; magnification, 5 ×). M: Mucosa; MM: Mucosa musculus; T: Tumor.

**Table 1 Characteristics of 16 transplanted patients with gastrointestinal stromal tumors**

|  |  |  |
| --- | --- | --- |
|  | **Overall number** |  |
| Male sex, *n* (%) | 16 | 11 (69) |
| Age (yr), median (min; max) | 16 | 59.5 (23; 74) |
| Type of organ transplantation, *n* (%) | 16 |  |
| Kidney | 12 (75) |
| Liver | 4 (25) |
| Location of primitive tumor, *n* (%)  | 16 |  |
| Stomach | 9 (56) |
| Small bowel | 3 (19) |
| Colon | 1 (6) |
| Other: pelvis, perineum, mesentery | 3 (19) |
| Time from transplantation to diagnosis (mo), median (min; max) | 16 | 32 (5; 252) |
| Metastatic dissemination at diagnosis, *n* (%) | 16 | 0 (0) |
| Tumor size (mm), median (min; max) | 15 | 45 (10; 230) |
| Risk of progression according to Joensuu’s criteria, *n* (%) | 14 |  |
| Very low | 2 (14) |
| Low | 4 (29) |
| Intermediate | 2 (14) |
| High | 6 (43) |
| Surgical treatment, *n* (%)  | 16 | 15 (94) |
| Adjuvant treatment, *n* (%) | 16 | 3 (19) |
| Modification of immunosuppression, *n* (%) | 11 | 9 (82) |
| Death during follow-up, *n* (%)  | 14 | 4 (29) |

**Table 2 Clinical features and immunosuppression regimen of 16 transplant patients with gastrointestinal stromal tumor**

|  |  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- | --- |
| **Ref.** | **Age (yr)/sex** | **Transplanted organ** | **Time from transplantation to diagnosis** | **Location of primitive GIST** | **Metastasis at diagnosis** | **Evolution/delay** | **Immunosuppression before diagnosis** | **Immunosuppression after diagnosis** |
| Agaimy and Wünsch[11] | 59/F | Kidney | 40 mo | Stomach | No | Relapse 68 mo | Not described | Not described |
| Agaimy and Wünsch[11] | 58/F | Kidney | 96 mo | Small bowel | No | Not described | Not described | Not described |
| Saidi *et al*[19] | 54/M | Liver | 11 mo | Colon | No | Not described | Tac, Aza | Not described |
| Camargo *et al*[22] | 64/M | Liver | 7 mo | Perineum | No | Not described | Tac, mycophenolate sodium | Not described |
| Tu *et al*[18] | 57/F | Kidney | 6 mo | Pelvis | No | Not described | Steroids, CsA, MMF | CsA and MMF at half dosage; rapamycin-containing regimensteroids withdrawn |
| Mulder *et al*[12] | 72/M | Kidney | 21 yr | Stomach | No | Peritoneal metastasis/24 mo | Steroids, CsA | Steroids, CsA (60% reduction in dosage) |
| Mrzljak *et al*[20] | 53/M | Liver | 12 mo | Jejunum | No | No | Tac, MMF | Same |
| Cimen *et al*[13] | 46/F | Kidney | 18 yr | Stomach | No | Not described | Steroids, CsA, Aza | Same with reduced dosage of CsA |
| Cheung *et al*[14] | 64/M | Kidney | 2 yr | Stomach | No | Yes/2 yr | Steroids, Tac, MMF | Steroids, Tac (reduced dosage), everolimus |
| Cheung *et al*[14] | 48/M | Kidney | 1 yr | Mesentery | Multiple tumors | No | CsA, MMF | CsA withdrawal, sirolimus introduction |
| Patiño *et al*[15] | 23/F | Kidney | 13 yr | Stomach | No | Local relapse/3 yr | Steroids, Tac, MMF | Not described |
| Xie *et al*[21] | 60/M | Liver | 11 mo | Stomach | No | No | Tac, sirolimus, MMF | Same |
| Elkabets *et al*[17] | 74/M | Kidney | 7 yr | Stomach | No | No | Steroids, CsA, MMF | Switch CsA to mTOR inhibitor |
| Takahashi *et al*[16] | 64/M | Kidney | 72 mo | Small bowel | No | No | Steroids, CsA, MMF | Stop CsA |
| Stammler *et al* | 60/M | Kidney | 5 mo | Stomach | No | No | Steroids, Tac, MMF | Switch MMF to belatacept |
| Stammler *et al* | 64/M | Kidney | 51 mo | Stomach | No | Yes/23 mo | Steroids, CsA, MMF | Switch CsA to Tac |

Aza: Azathioprine; CsA: Cyclosporine; F: Female; GIST: Gastrointestinal stromal tumor; M: Male; MMF: Mycophenolate mofetil; mTOR: Mammalian target of rapamycin; Tac: Tacrolimus.

**Table 3 Histopathological features, treatments, and outcome of 16 transplant patients with gastrointestinal stromal tumor**

|  |  |  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- | --- | --- |
| **Ref.** | **Size (mm)** | **Mitotic count** | **Fletcher’s criteria** | **Joensuu’s criteria** | **Mutation identified** | **Resection** | **Initial adjuvant treatment** | **Second line treatment** | **Outcome** |
| Agaimy and Wünsch[11] | 35 | < 5/50 | Low | Low | Not described | Yes | No | Not described | Alive and relapse free at 68 mo |
| Agaimy and Wünsch[11] | 230 | 14/50 | High | High | Not described | Yes | No | Not described | Not described |
| Saidi *et al*[19] | 25 | 1/50 | Low | High | Not described | Yes | No | Not described | Alive and relapse free at 18 mo |
| Camargo *et al*[22] | 50 | 5/50 | Intermediate | High | Not described | Yes | No | Not described | Alive and relapse free at 20 mo |
| Tu *et al*[18] | 45 | 2-3/50 | Low | Low | *PDGFRA* exon 18 V824V | Yes | No | Not described | Alive and relapse free 24 mo |
| Mulder *et al*[12] | 50 | > 10/50 | High | High | Not described | Yes | No | Imatinib 400 mg/d then 200 mg/d | Died 44 mo |
| Mrzljak *et al*[20] | 10 | 1/50 | Very low | Very low | Not described | Yes | No | No | Died 3 yr after from acute leukemia |
| Cimen *et al*[13] | 150 | 14/50 | High | High | *KIT* T574del | Yes | Imatinib 400 mg/d | Not described | Alive and relapse free at 12 mo |
| Cheung *et al*[14] | 30 | 9/50 | Intermediate | Intermediate | Not described | Yes | No | No | Died from pneumonia at 2 yr |
| Cheung *et al*[14] | Not described | Not described | Not described | Not described | Not described | No | Imatinib 400 mg/d for 1 yr | Switch CsA to sirolimus | Alive and relapse free at 10 yr |
| Patiño *et al*[15] | 58 | Not described | Intermediate or high | Intermediate of high | Not described | Yes | No | Imatinib 400 mg/d | Alive and relapse free/5 yr after imatinib initiation |
| Xie *et al*[21] | 10 | < 5/50 | Very low | Very low | *KIT* exon 11  | Yes | No | No | Not described |
| Elkabets *et al*[17] | 31 | Not described | Not described | Not described | Not described | Yes | No | No | Alive and relapse free at 40 mo |
| Takahashi *et al*[16] | 110 | 20/50 | High | High | *KIT* exon 11 | Yes | Imatinib 400 mg/d reduced to 3000 mg/d | No | Alive and relapse free at 18 mo |
| Stammler *et al* | 27 | 2/5 | Low | Low | *KIT* exon 11 | Yes | No | No | Alive and relapse free at 2 mo |
| Stammler *et al* | 51 | 10/50 | High | High | *PDGFRA* exon 18 | Yes | No | Sunitinib then regorafenib then dasatinib | Died 56 mo later |

CsA: Cyclosporine.



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