**Name of Journal:** *World Journal of Gastroenterology*

**Manuscript NO:** 79724

**Manuscript Type:** REVIEW

**Microbiota revolution: How gut microbes regulate our lives**

Colella M *et al*. Gut microbiota and life

Marica Colella, Ioannis Alexandros Charitos, Andrea Ballini, Concetta Cafiero, Skender Topi, Raffaele Palmirotta, Luigi Santacroce

**Marica Colella, Raffaele Palmirotta, Luigi Santacroce,** Interdisciplinary Department of Medicine, Section of Microbiology and Virology, University of Bari “Aldo Moro”, Bari 70124, Italy

**Ioannis Alexandros Charitos,** Maugeri Clinical Scientific Research Institutes (IRCCS) of Pavia - Division of Pneumology and Respiratory Rehabilitation, Scientific Institute of Bari, Bari 70124, Italy

**Andrea Ballini,** Department of Clinical and Experimental Medicine, University of Foggia, Foggia 71122, Italy

**Andrea Ballini,** Department of Precision Medicine, University of Campania “Luigi Vanvitelli”, Naples 80138, Italy

**Concetta Cafiero,** Area of Molecular Pathology, Anatomic Pathology Unit, Fabrizio Spaziani Hospital, Frosinone 03100, Italy

**Skender Topi,** Department of Clinical Disciplines, School of Technical Medical Sciences, University of Elbasan “A. Xhuvani”, Elbasan 3001, Albania

**Author contributions:** Colella M contributed to the conceptualization and data collection; Charitos IA involved in the original manuscript writing and drafting, and resources; Ballini A take part in the manuscript revision and formal analysis; Cafiero C and Palmirotta R contributed to the investigation; Palmirotta R and Santacroce L involved in the validation of this manuscript; Santacroce L contributed to the conceptualization, project administration, and funding of this manuscript; and all the authors have read, discussed, and approved the original manuscript and the final version of it.

**Corresponding author: Luigi Santacroce, MD, Professor,** Interdisciplinary Department of Medicine, Section of Microbiology and Virology, University of Bari “Aldo Moro”, Pzza Giulio Cesare, 11, Bari 70124, Italy. luigi.santacroce@uniba.it

**Received:** March 28, 2023

**Revised:** May 16, 2023

**Accepted:** July 10, 2023

**Published online:**

**Abstract**

The human intestine is a natural environment ecosystem of a complex of diversified and dynamic microorganisms, determined through a process of competition and natural selection during life. Those intestinal microorganisms called microbiota and are involved in a variety of mechanisms of the organism, they interact with the host and therefore are in contact with the organs of the various systems. However, they play a crucial role in maintaining host homeostasis, also influencing its behaviour. Thus, microorganisms perform a series of biological functions important for human well-being. The host provides the microorganisms with the environment and nutrients, simultaneously drawing many benefits such as their contribution to metabolic, trophic, immunological, and other functions. For these reasons it has been reported that its quantitative and qualitative composition can play a protective or harmful role on the host health. Therefore, a dysbiosis can lead to an association of unfavourable factors which lead to a dysregulation of the physiological processes of homeostasis. Thus, it has previously noted that the gut microbiota can participate in the pathogenesis of autoimmune diseases, chronic intestinal inflammation, diabetes mellitus, obesity and atherosclerosis, psychic disorders (*e.g.,* neurological diseases, autism, *etc.*) colorectal cancer, and more.

**Key Words:** Microbiology;Human microbiota; Intestinal microbiota; Immune system; Metabolites; Dysbiosis; Probiotics; Diseases; Cancer

Colella M, Charitos IA, Ballini A, Cafiero C, Topi S, Palmirotta R, Santacroce L. Microbiota revolution: How gut microbes regulate our lives. *World J Gastroenterol* 2023; In press

**Core Tip:** The microbial populations that colonize the human body constitute a complex ecosystem; several cells much higher than the total number of cells in the human body. It is generally accepted that the human gut microbiota is a focus of research interest due to its complexity, involvement with health, and involvement in various pathological conditions. The need to extensively elucidate and document hitherto unknown aspects of the gastrointestinal microbiota, its associations with health fuels the need for further study. Furthermore, on the subject in question it has constituted a trigger for carrying out the present review of the research sources received so far.

**INTRODUCTION**

The human organism is colonized by trillions of microbes. The microbiome refers to all the genes of microbes found in various locations in an individual’s body. Instead for microbiota we mean the total of microorganisms quantitatively and qualitatively present. The human host and the microbiota have co-evolved for the benefit of both parties especially the intestinal one[1,2]. In fact, on the one hand, the host provides space, suitable conditions, and food for the growth of the intestinal microbiota and this in turn generally participates in obtaining useful substances and induces resistance to various infections. The relationship between the human gut microbiota and the host is symbiotic, in which both the host and the microorganisms are mutually beneficial. For its part, the host offers a place of growth and nourishment to the symbiotic intestinal bacteria, which in turn favors the function of the host on the one hand by inducing resistance to infections and on the other hand by facilitating the absorption of digested food[3]. It appears, therefore, that eukaryotic hosts and symbiotic bacteria have “co-evolved” with mutual interactions based on nutritional benefits enjoyed by both parties. When this balance is disturbed (dysbiosis) for various reasons, such as repeated and inappropriate use of antibiotics or alcohol abuse, pathological conditions may arise, such as mild chronic intestinal inflammation or metabolic disorders[4]. The interactions of (symbiotic) microbes with each other is of particular interest. A negative correlation has been observed between members of the phylum *Bacteroidota*, (such as that of the *Prevotellaceae* spp*.*) in the intestine, which may reflect alternative “metabolic specializations”[2,5]. Thus, the gut microbiota is of particular importance for the maintenance of human health. In fact, various biological mechanisms are microbiota-dependent because they cannot be performed autonomously and are useful for health homeostasis. In general, the concept of a “superorganism” refers to the bidirectional and therefore beneficial activity between the host organism and the gut microbiota. The microbiota, in addition, has both a protective and trophic role influencing, as we have mentioned, several homeostatic processes of the host organism, *e.g.*, tissue trophism, immune balance, metabolic activity, neuro-endocrine function, *etc.*[6,7]. Intestinal microbiota represents the most populous community of the entire organism. In fact, the stomach has about 103-104 bacteria, the duodenum 105-106 and the terminal ileum 108-109 bacteria (per gram of tissue). However, the most populated is the large intestinal one representing around 1012-14 bacteria per gram of tissue[2,6]. The colon is the part of the gastrointestinal tract where a complex set of microorganisms develops from the moment of birth. The microbiota of the large intestine is denser and more diverse than the human microbiota of the small intestine. Intestine are enriched in the phyla of *Bacillota* and *Actinomycetota*, while *Bacteroidota* and *Lachnospiracae* are more abundant in colonic samples[8]. It is estimated that about 400-500 different genera of microorganisms constitute the intestinal microbiota, while the 90% of them species are predominantly anaerobic. Most of them belong to two genera, *Bacteroidota* and *Bacillota*. The remaining bacterial populations belong to *Pseudomonadota*, *Actinomycetota*, *Fusobacteria* and V*errucomicrobia*[9]. An estimated 70% of these microbial communities are bacteria that cannot be cultured with conventional microbiological techniques. It has been observed that the microbial populations of the intestinal mucosa differ from those found in the intestinal lumen. The large intestine hosts the most numerous microbial cells which vary in species from individual to individual and are organized in localized microbial communities along its path. These microorganisms appear not to follow stable topological patterns along the intestine as they are influenced by the local oxygen concentration. Thus, facultative aerobic communities reside near the intestinal walls, which are sites of high oxygen concentration, and anaerobes prefer the intestinal lumen, where oxygen concentration is lower[10]. Although there is great diversity in the microbes that colonize the gut, it has been found that the gut microbiota of most individuals can be classified into three main microbial groups or “enterotypes” depending on the predominance of genera: *Bacteroides* (enterotype 1); *Prevotella* (enterotype 2) or *Ruminococcus* (enterotype 3). More recent data, however, show that the division into three enterotypes is a simplification and that there are many intermediate states in the gut. The prevalence of each enterotype is mainly determined by dietary factors[2]. However, it appears that everyone has a fixed bacterial strain, even though the composition may vary. One of the main functions of the intestinal microbiota is to protect the intestine from pathogenic microorganisms, it also contributes to the development of a healthy immune system, regulates intestinal motility and participates in metabolism. More specifically, the gut microbiota promotes the regulation of the immune system through: (1) Stimulation of the immune response against potential pathogens; and (2) Suppression of the immune response against food and symbiotic antigens[11]. It is estimated that 80% of antibody production occurs locally in the gut, mainly supplying immunoglobulin A. In addition, they participate in host metabolism, drugs metabolism, maintenance of mucosal structural integrity, bile salts metabolism, plant fibers, mucus, and fatty acid catabolism[12]. In addition to bacteria that have already been studied quite a lot, recently it has been hypothesized that yeasts are involved in starch metabolism. Understanding the relationship between yeasts and the immune system is characterized as difficult and there are few data on their action. In some special cases, they can also act therapeutically[13]. Thus, there is an amphidromic relationship between microbes and hosts (Figure 1).

Finally, the probiotics bacteria are the “good” bacteria that are found in gut microbiota but also are added in specific dietary or ferment foods. Probiotics aid the organism by strengthening our immunity and help with gastrointestinal health, especially in conditions such as the irritable bowel syndrome (IBS)[14]. In the 1989, Fuller listed some beneficial effects and therapeutic applications of probiotic bacteria. We find probiotics in dairy products, such as yogurt and aged cheeses, but also in other foods such as pickled vegetables, *etc.* The consumption of probiotics promotes the growth of desirable microorganisms, overcoming potentially harmful bacteria and strengthening the body’s defences. Several scientific articles commenting on these results, refer to studies using cultures such as of *Lactobacillus acidophilus* and *Bifidobacterium*, *Streptococcus thermophilus* and *Lactobacillus delbrueckii* subsp. *bulgaricus*[14,15]. Consuming probiotic products can cause a plethora of positives effects on the human body. Probiotics, thanks to their antimicrobial properties action can fight intestinal and other diseases, aid in various infections (such as those from coronavirus disease 2019 and other), reduce levels serum cholesterol, stimulate the immune system, reduce allergy symptoms, aid with lactose intolerance, can prevent hypercholesterolemia and osteoporosis and other[15].

**MAIN FACTORS AFFECTING THE INTESTINAL MICROBIOTA**

***Inheritance***

The mechanisms of host-bacteria interactions have not been fully described. Little is known about the relationship between the host genotype and its gut bacteria. However, there is strong evidence that it affects intestinal populations. Simple genetic mutations can lead to changes in the composition of microorganisms in the gut[16]. Further investigations are needed to delineate the mechanisms by which this occurs. There are few studies comparing the gut microbiota between family members. It has been observed that monozygotic and dizygotic twins had a greater similarity in gut microbiota between monozygotic than dizygotic siblings, indicating the importance of genetic background. Furthermore, it was observed in an animal model that the similarities in the gut microbiota of the same mouse strain were greater than in mice of a different species even with a common environmental effect. The role of parental genetic influence on the colonization of microorganisms in the intestine has not been elucidated.

***Geographical location***

The diversity of the gut microbiota in children living in rural areas is greater than that of children in developed countries. It was noted in a study among Caucasians and Asians (Chinese) in the United States and Hong Kong that there were qualitative differences and quantitative in the intestinal microbiota[17-19]. It was found that children from a country in West Africa (Burkina Faso) that had a high presence of *Bacteroidota* with a greater presence of the genus *Xylanibacter* and *Prevotella* (which allow the hydrolysis of xylan and cellulose) with a reduced presence of *Bacillota*. Indeed, the higher presence of short-chain fatty acids (SCFAs) was noted. Therefore, the intestinal microbiota can help the host to optimize its energy intake from dietary fibers according to nutrition and needs, thus also protecting against infective and inflammatory processes[20].

***Intrauterine period and delivery***

There is a growing body of data suggesting that the human gut microbiota begins before infant birth. Meconium (the baby’s first stool) contains bacteria, where *Bacillota* phyla predominate, the bacteria through the placenta and circulatory system enter the intestinal lumen of the foetal intestine. After human birth, the gut is colonized by many microbial strains, and everyone has their own gut microbiota which changes further throughout their life[2,6]. How the baby is born is very important as it affects the variety of microorganisms which will be installed. During normal delivery the new-born receives microorganisms from the mother’s vagina or intestinal tract, whereas in caesarean section the new-born is exposed to those from the hospital environment. The gut microbiota in these newborns can be disturbed for months to years. Newborns born with normal delivery develop a microbiota that reflects the vaginal microbiota, while those born with caesarean section develop a skin microbiota resulting in delayed microbial colonization by *Bacteroides,* and *Bifidobacterium* spp*.*[16,21].

***Diet***

Many are observed in the first months of a person’s life changes in the gut microbiota, while the stability and diversity of microbiota genes increase after the first three years of life[21]. Breastfeeding or consuming breastmilk substitutes in infancy has significant effects on intestinal microbiota of the new-born and in the development of the immune system[22]. In animal models, the two different ways of feeding babies develop different populations of microorganisms in the gut which lead to differences in the immune system. Immunity is affected up to the first five years of life[23]. In humans there are few studies on the different administration of milk to newborns. Undoubtedly, however, the intestinal microbiota of the child influences the development of the immune system and its metabolism, just as in adult life[24]. Breastfed infants are exposed to the gut microbiota offered by milk, which contains more than 700 species of bacteria. Breast milk contains many protective factors that breast milk does not contain. In addition, changes in the infant’s gut microbiota appear to predispose to diseases and his later life[2,16]. Subsequently adult dietary habits are one of these most important environmental factors shaping the growth of microorganisms in the human gut. Short or long-term consumption of animal or vegetable products changes the structure of the intestinal microbiota. The short-term diet based on animal foods increases the microorganisms of genera such as *Bacteroides*, *Alistipes*, *Bilophila* and decreases the levels of *Roseburia*, *Eubacterium rectale*, *Ruminococusbromii* belonging to the *Bacillota* phyla (populations that mainly metabolize polysaccharides of plant origin)[25]. Instead, it has been noted that long-term consumption of fruit and vegetables by the elderly is associated with an increase in populations of the genus *Prevotella*. It was noticed that a change of nutrition style (*i.e.,* sugar-rich foods, a shift from a low-fat vegetable polysaccharide to a high-fat vegetable polysaccharide, and so) modifies the microbiota, qualitatively and quantitatively, through a change of the microhabitat in 24 h and so on changes in metabolic pathways also occur. Although the gut microbiota responds to short-term changes in diet, long-term nutrient changes appear to determine the type of qualitative and quantitative microorganism population also for other such as the oral ones[26,27]. There are differences in the presence of gut bacteria in the two diets, reflecting differences in metabolism. of carbohydrates and proteins. In both cases, the colonization of fungi, viruses and yeasts is observed[25]. It has been observed that the consumption of foods of plant origin is also associated with a beneficial set in the intestinal microbiota. A high correlation has been noted between a plant-based diet and increased levels of SCFAs and some dietary fibres that degrade *Bacillota* phyla. Indeed, the characteristics of the Mediterranean-type diet, *i.e.,* increased intake of cereals, fruit, vegetables, and legumes offer many benefits for the human intestine[28]. An observational study found that dietary fiber from beans, fruits, and vegetables was associated with an abundance of and *Actinomycetota* phyla and *Clostridium* spp*.* But also, the consumption of sour milk, many dairy products, or the consumption of fermented foods (such as Greek yogurt, kefir, and others) affect the intestinal microbiota and change its structure. The use of substances of abuse (such as alcohol, cocaine and other) or chemical xenobiotics can cause in human health is associated with quantitative and qualitative changes in the intestinal microbiota (dysbiosis)[29,30].

***Lifestyle***

Modern lifestyles can influence the intestinal microbiota, sedentary life contributes to obesity and combined with a positive energy balance and the consumption of foods of animal origin rich in saturated fat leads to the rearrangement of populations at the intestinal level. Although studies show conflicting results, an increase in *Bacillota* and a decrease in *Bacteroidota* have been observed in individuals consuming foods of animal origin[6]. Frequent and moderate exercise influences the intestinal defence as well as on the genes of intestinal microorganisms. In a study on the effect of exercise on the intestinal microbiota of 22 athletes, the beneficial influence of exercise on the diversity of intestinal microorganisms is underlined and the combination with diet is claimed to affect the gut microbiota structure[31,32]. Smoking also affects the composition of the gut microbiota, increasing populations of the *Bacteroides-Prevotella* genera. Stress affects gut motility, which can change the structure of microbial populations, which has even been blamed for the development of inflammatory bowel disease (IBD)[33].

***Use of antibiotics***

Antibiotics target pathogenic microorganisms, but they also have a negative effect on intestinal symbionts. Unfortunately, they affect the intestinal microbiota especially if they are broad-spectrum antibiotics that are used for all diseases[34]. Gut microbiota diversity is reduced, many strains are lost, and their re-emergence is gradual and long-term, which is a major concern of experts in the field of intestinal health. It was observed that most of the bacteria affecting antibiotics response mainly belong to the *Bacillota* and *Pseudomonadota* phyla[35,36].

**THE ROLE OF THE HOST/INTESTINAL MICROBIOTA CROSS-TALKING AXIS**

***Nutritional and gut-influenced energy conservation by metabolic processes (metabolome)***

Several studies have shown that the intestinal mucosa needs to be colonized by microorganisms to take on its integral structure. For example, mice raised in a sterile environment developed fewer blood vessels in their intestinal villi. Sterile growth also showed that there is defective growth in gut-associated lymphoid tissue and antibody production. Also, in the context of the sterile environment, fewer Peyer’s patches develop, there are fewer cells in the dermis, and fewer plasma cells in the germinal centres of the mesenteric lymph nodes than in the development data in a nonsterile environment[37]. The intestinal microbiota could be considered a “metabolic organ” performing multiple functions needed for maintaining the health status of the host organism orchestrating an amphidromous communication with it. The catabolism of a number of both endogenous and exogenous indigestible molecules (*i.e.,* cholesterol, fibers, bile acids, excreted mucus, *etc.*), is one of the most important activities of the intestinal microbiota accounting for 10% of the host’s energy requirement every day[38,39]. So, the gut microorganisms (especially bacteria and commensal fungi) produce SCFAs to gain energy but also give trophic substances and energy to the host organism. Finally, certain bacterial species can synthesize vitamins such as B12, folic acid, thiamin, biotin, vitamin K, amino acids and more[40]. As proof, *Bacteroides* *thetaiotaomicron* can metabolize polysaccharides that reach the large intestine as is in Figure 2. Indeed, it has many enzymes such as glycoside hydrolases and polysaccharide lyases that break down pectins, arabinose, *etc.*

Archaea, such as *Methanobrevibacter smithii*, establish beneficial relationships with other bacteria to eliminate the H2 by-products, thus facilitating the yield of ATP[41,42]. The gut microbiota benefits the host in many ways contributing to the conservation of energy from several common non-digestible polysaccharides through enzymes such as glycoside hydrolases and other non-encoded enzymes in the human genome[43]. Studies of mice with a germ-free gut microbiota revealed that the gut microbiota improves the regulation of fat accumulation and obesity mainly due to an increase in energy production from food. These (germ-free) mice are protected against obesity and metabolic syndrome[44]. It was noted that restoring an eubiotic intestinal microbiota in these animals leads to an increase in insulin resistance and fasting blood glucose, but also of the level of liver triglycerides and body fat amount.

The intestinal microbiota improves the absorption of monosaccharides, which in turn leads to an increased lipogenesis with consequent accumulation of triglycerides both in the liver and in the fat tissue. With food utilization, obese mice have been reported to conserve energy from food more efficiently than lean-wild-type mice[45]. Distal gut microbiota’s composition of obese mice revealed that the relative increase of *Bacteroidota* and *Bacillota* resulted in a modified metabolic potential of the gut microbiota, giving it a greater ability to use energy from the diet. Interestingly, this obesity trait of his was transmissible through faecal transplants from obese mice as opposed to lean germ-free mice. Obese mice also possess multiple methanogenic archaea which can increase the efficiency of bacterial fermentation through H2 removal[46]. It was observed that, after gut co-colonization, both *Bacteroides thetaiotaomicron* and *Methanobrevibacter smithii* boost up the efficiency and specificity of bacterial fermentation by removing bacterial polysaccharides, increasing adiposity compared to mice colonized with only one of the two organisms[47,48]. Further studies have highlighted further correlations regarding the possibility of increased energy saving through induced microbiota nutrition or genetically induced obesity microbiota[49]. For that, age and diet are important factors not only for the composition of the gut microbiota, but also for energy saving possibilities. In addition, the link between gut microbiota and liver diseases has been studied mainly in obesity and non-alcoholic steatohepatitis/nonalcoholic fatty liver disease patients, and in liver failure conditions (*e.g.,* cirrhosis, hepatic encephalopathy, infections, hepatocellular carcinoma), total parenteral nutrition-associated liver disease, cholangitis primary sclerosing, primary biliary cirrhosis[50]. It has been noted that dysbiosis of the intestinal microbiota leads to intestinal motility disorders (stasis, with bacterial proliferation). These conditions lead to increased intestinal permeability, impaired immune response with an excessive increase in tolerance in bacterial translocation by the intestinal microbiota with subsequent liver injury[50,51].

***Immunological actions and resistance to colonization by pathogens***

The host’s organism meets both the pathogenic microbes of the environment and the microbes of the intestinal microbiota. Previous studies of the immune system have focused on the mechanisms by which this system can defend itself against pathogenic microbes. In addition, the microorganisms of the intestinal microbiota produce antimicrobial substances such as bacteriocins and hydrogen peroxide which inhibit the growth of others with pathogenic behaviour[51]. The immune system has evolved in such a way that it can accommodate symbiotic bacterial communities of increasing complexity while retaining the ability to fight pathogenic bacteria. The microbiota regulates the development and function of the innate and acquired immune systems[11]. Even in healthy conditions of the host, there is a continuous stimulation of the immune system by the intestinal microbiota. This condition leads to a basal state of “low physiological inflammation” representing an effective first line of defence against pathogenic microbes. Furthermore, both resident and pathogenic microbes compete for available sites and nutrients, so the microbiota exerts a protective role metabolizing those nutrients that are necessary for the pathogens survival and producing molecules that inhibit their growth[2,52]. In fact, it has been shown that the introduction of certain molecules produced by *Bacteroides thetaiotamicron* and *Eubacterium rectale* can induce the production of specific mucosal glycans. These can be metabolized by these 2 bacterial species only but not by pathogens, thus preventing their proliferation. Hence, diet appears to have a pivotal role in microbial composition modifications[53]. The immune system works by learning, *i.e.,* at the beginning of life it has the necessary components (cells, internal cellular mediators, *etc.*) but does not have data available from the environment, which it acquires in the first years of life through contact with other people and the natural environment. If these early childhood data are inappropriate, the regulatory mechanisms of the immune system could fail. As a result, the immune system attacks not only pathogenic microorganisms but also harmless targets such as pollen, house dust and food antigens, leading to the onset of allergic diseases[54]. The microorganisms together with digestive enzymes, the mucus layer, intestinal peristalsis, and the epithelial barrier with “tight” connections (tight junctions) constitutes the non-immune component of the body’s immune response. The functions of the intestinal microbiota in terms of defence of the organism are on the one hand to influence in a decisive way the arise of the gut immunity (for which reference was made to the trophic role) and on the other to prevent possible invasion of pathogens through a direct effect on them and/or through the “activation” of the host’s immune mechanism[55,56]. As far as natural immunity is concerned, it has the ability, by recognizing characteristic pattern molecules [pathogen associated molecular patterns (PAMPs)] on microorganisms, to separate potentially pathogenic microbes from “unwanted” antigens. More specifically, the cells of the natural immunity using proline rich proteins (PRPs) receptors (pattern recognition receptors) detect PAMPs[57]. PRPs also participate in the release of cytokines and the activation of acquired immunity. However, there are many types of PRP, and of the most important are the Toll-like receptors (TLRs) found in dendritic cells, neutrophils, macrophages, and intestinal mucosal epithelial cells. The most known PAMPs recognized by PRP receptors are bacterial carbohydrates (*e.g.,* mannose, glucides of lipopolysaccharide), bacterial peptides (*e.g.,* flagellin), peptidoglycans and lipoteichoic acid of Gram+, fungal lipoproteins, glycans, viral genomes. Since these molecules are also found in symbiotic microorganisms, they are called microbe-associated molecular patterns (MAMPs)[58]. Thus, MAMPs seem to be able to modify the expression of TLRs in natural immunity cells. Thus, the recognition of MAMPs triggers the activation of the nuclear factor kappa-light-chain-enhancer of activated B cells (NF-κB) pathway, which results in the production of cytokines, the activation of other auxiliary and necessary molecules on the antigen-presenting cells, which ultimately results in the activation of T-cells, *i.e.,* acquired immunity[59]. However, microorganisms can modulate the natural immunity changing the quality and the amount of mucus from the intestinal mucoid cells. Mucus forms a natural barrier to stop an infection directly adhering to them, collects the bacterial by-products, and protect epithelial cells from secretions thanks to its low pH rich and richness in lytic enzymes. These events activate body’s defences[60]. As far as the acquired immunity in the gastrointestinal system is concerned, it is “based” in the gut associated lymphoid tissue which is made up of the Peyer’s Patches and the mesenteric lymph nodes. The spleens of germ-free mice were found to contain increasingly smaller germinal centres in the lymph nodes and reduced numbers of memory CD T4 cells in the intestinal epithelium, that the production of cytokines belongs to a Th2-type of immune response, and that these animals have a reduced ability to secrete germicidal agents[61]. Recolonization of these muscles with muscle-specific bacteria can reverse some of these perturbations, as experimentally demonstrated by the restoration of systemic T-cell deficiency and Th1/Th2 imbalance of sterile muscle microbes after single colonization of the muscle gut with the bacterium *Bacteroides fragilis*. Such studies collectively highlight the importance of the gut microbiota for the normal development of the peripheral immune system in immunocompetent hosts[62].

***Influence of the host’s health condition***

According to clinical studies that have been done in recent years, they show a correlation of the change in the composition of the gut microbiota with several serious pathological conditions such as: autoimmune diseases, type 2 diabetes (T2D), weight gain and obesity, IBD, asthma and chronic sinusitis, mental health disorders, dermatological problems, Alzheimer’s disease, poor immune system health, gastroesophageal reflux disease, constipation or diarrhea, cancerous conditions and other[63].

**INTESTINAL MICROBIOTA ROLE IN DISEASES**

***IBS and******IBD***

Functional bowel disorders such as IBS are defined solely by symptom-based diagnostic criteria. IBS is characterized by abdominal pain or discomfort and changes in bowel habits. Although the aetiology is multifactorial, recent studies to understand the pathophysiology of IBS have revealed that changes in the normal gut microbiota may play a role in IBS-associated low-grade intestinal inflammation[2,40]. Indeed, intestinal microbial dysbiosis is involved in the pathogenesis of IBS by facilitating the adhesion of the pathogenic microorganism to the intestinal wall and several studies revealed that. Indeed, there is a clear separation between the gastrointestinal microbiota of patients with IBS and that of healthy controls[64]. That is, IBS is characterized by an increase in the genera *Dorea*, *Clostridium* (as *Bacillota* phylum) and *Ruminococcus* with an important loss in the population of the genera *Bifidobacterium* and *Faecalibacterium*. According to a trial on IBS in young patients who were divided into subgroups with different bacterial fingerprints, an increase of *Bacillota* phylum compared to *Bacteroidota* phylum exists, differing from healthy patients[65,66]. Finally, another study on a paediatric population with IBS showed a variation of *Bacillota* and *Pseudomonadota* phyla amount, with many of the genera *Dorea* and *Ruminococcus*, and *Haemophilus parainfluenzae* species. The *Bacteroides* genus was found to be smaller in paediatric patients with IBS than in healthy controls. Typically, these studies could ultimately lead to the design of targeted therapies[67].

Idiopathic IBDs are immune-driven chronic diseases and represented by ulcerative colitis and Crohn’s disease. Today they are an adverse immune response to endogenous symbiotic gut microorganisms with or without the involvement of the autoimmunity process. Studies have shown a change in the composition of the gut microbiota in IBD[68]. Furthermore, there is a qualitative reduction in it, *i.e.,* on the bio-diversity of the population with a typical reduction of *Bacillota* phylum strains (such as *Bifidobacteria*, spp*., Faecalibacterium prausnitzii*, and *Lactobacillaceae* families) while an increase in the microorganisms that are attached to the mucus is observed. *Bacillota* strains are the main producers of SCFAs, such as butyric acid, which has immunomodulatory properties[69]. However, it has not been clarified whether the disturbance in the microbiota is the cause of the disease or is the result of it. The normal, non-inflamed gut contains many immune cells which are in such a state of activation that there is no complete immune response to the microbes of the normal microbiota and food antigens. This is due to the activation of potent mechanisms of immune regulation, such as the stimulation of regulatory T cells expressing the transcription factor FoxP3 to suppress inflammation[66]. But, under environmental stimuli (*e.g.,* certain infections) the activation of the intestinal immune system starts on a large scale but is subsequently suppressed. In IBD patients this suppression of this immune response may not be adequately regulated. The eubiosis of the intestinal microbiota is under the control of the host through immune and epithelial responses, diet, drugs use (especially antibiotics), genetics, and more[2]. In turn, the microbiota, has significant effects on host epithelial and immune function thanks to its structural components and metabolism, that can become permanent by epigenetic effects. From childbirth, when the human microbiota is established, these host effects may influence the risk of developing IBD later in life[63]. It is worth noting here that in most studies the incidence of IBD increases particularly in the second to fourth decades of life, while some studies even report a second peak in the sixth and seventh decades. Specifically, therefore, components of the microbes can promote or protect against disease. The community microbes in patients with ulcerative colitis and Crohn’s disease have been shown to be different from unaffected individuals, a state of dysbiosis: The presence of disease-causing microorganisms (such as those form *Pseudomonadota* phyla and adherent *Escherichia coli*) and to which directed immune response and/or loss of microorganisms that inhibit inflammation (*e.g.,* such as *Faecalibacterium prausnitzii*). But many changes and inflammation results in changes in the microbial community. Also, antibiotics (such as nitroimidazoles, quinolones) and certain diets change the gut microbiota and may improve Crohn’s disease symptoms. In fact, the use of antibiotics really has a place in the treatment of Crohn’s disease[2,16,70].

***Immune dysregulation (allergies and autoimmunity)***

Effectively stimulated immunity of both local and systemic mucosa is required for a mature gut microbiota. If this does not happen, a dysregulation can occur which can lead to allergic manifestations or an asthmatic phenotype from early life. It has been noted that developing countries have a lower incidence of allergic diseases such as asthma than countries developed. Thus the “hygiene hypothesis” was developed, which according to this lack of exposure to pathogenic bacteria or products of non-pathogenic bacteria can cause this condition by a negative effect on the development of the immune system[71]. Subsequently, the “microbiota hypothesis” was formulated, in which it is proposed that changes in diet, as well as increased use of antibiotics in developed societies, lead to a less diverse gut microbiota in its microbial components. This “immature” gut microbiota, as it has been defined, alters the development of the immune system interrupting the proper evolution of events that allow the development of immune tolerance and thus increasing the risk of developing allergic hypersensitivity[72]. More specifically, levels of *Bifidobacterium* and *Enterococci* appeared to be related to allergic symptoms in the first months of life. An increased *Bacteroidota*/*Bifidobacterium* ratio was reported during the second year of life in children who developed symptoms of atopy children who eventually developed allergies were less frequently colonized with *Lactobacillaceae* phyla, *Bifidobacterium* and *Clostridiodies difficile* strains during the 2nd mo of life[73]. Autoimmune diseases (such as rheumatoid arthritis and other) can be sharing a common pathogenesis, an immune-mediated attack on the body’s own organs. Autoimmune diseases usually show a variety of characteristics and symptoms. Concerning these characteristics, the most common may include, for example, an increase in epithelial (such as intestinal) and vessel permeability, mitochondrial dysfunction, progressive inflammation and chronic infections, imbalance of the hypothalamic-pituitary-adrenal (HPA) axis, and microbiota dysbiosis. Symptoms, on the other hand, can manifest as chronic fatigue, allergic phenomena, poor cognitive function, mood and mental disorders and pain, skin rashes, gastrointestinal disorders, and so[74,75]. An example of the genetically predisposed autoimmune diseases of the small intestine is celiac disease, an age-independent condition. The symptoms occur after the ingestion of the toxic epitopes of gluten (*i.e.,* the proteins present in wheat, rye, barley and, less so, in oats). In celiac disease there is an increase in intraepithelial lymphocytes and an atrophy of the intestinal villi. The autoimmune mechanism is due to the presence of various autoantigens, where the most important is tissue transglutaminase that is also an important diagnostic marker. Furthermore, it is often associated with other autoimmune diseases, such as insulin dependent diabetes mellitus (IDDM), or type 1 diabetes (T1D), thyroiditis, *etc.* This suggests that these diseases share common pathogenetic pathways[76-79]. Finally, the intestinal microbiota seems to play a fundamental role in the development and course of celiac disease. There is an unfavourable qualitative and quantitative composition of the intestinal microbiota characterized by a greater presence of the genus *Bacteroides* and *Escherichia coli* and a minor presence of *Bifidobacterium* spp. (*e.g.,* *B. longum* compared to healthy controls). Furthermore, this condition does not seem to change even after a gluten-free diet[80,81]. Finally, it has been noted that children born by caesarean section have a higher risk to develop the disease[82,83].

***Diabetes mellitus***

The exact role of the gut microbiota in the pathogenesis of T1D remains unknown. Data from experimental models support the idea that some bacterial families may act protectively against divalent metal transporter 1 (DMT1)*.* Studies have shown that the use of antibiotics in experimental animals can prevent the occurrence of type 1[84]. But also, the administration of probiotics with bacterial strains in experimental animals prevented the onset or delayed the progression of the disease[85]. In another study, it was found that accidentally infecting mice with a spore forming bacterium resulted in a reduction in the incidence of DMT1[86]. Similarly, incubation of mycobacterium and streptococci in laboratory animals protected them from developing diabetes. Studies of the gut microbiota in mice that developed DMT1 and mice that did not develop diabetes reported that, at the onset of the disease, the two groups differed in the concentrations and type of microbes[87]. Stool samples from animals (mice) that developed diabetes contained higher concentrations of so-called probiotic bacteria, such as from *Lactobacillaceae* phyla and *Bifidobacterium* spp*.*, in contrast to mice that did not develop diabetes which showed higher concentrations of *Bacteroides*, *Eubacterium* and *Ruminococcus*. It is worth mentioning that the *Lactobacillus johnsonii* strain prevents the development of diabetes when administered to mice[88,89]. Because of the possible relationship between the microbiota and IBD, most of the research around the gut microbiota has been done in patients with IBD, whereas studies in patients with type 1 diabetes mellitus (T1DM) are limited. In addition to increased intestinal permeability, patients with T1DM show an increase in inflammatory cells in the gut and decreased numbers of CD4, CD25, and T-cells, which are the master regulator of the immune system. Another study showed that individuals who developed T1DM had higher concentrations of *Bacteroidota* and lower concentrations of *Bacillota* compared to healthy controls. In addition, individuals who developed DMT1 were colonized with a lower number of bacteria compared to healthy controls[90,91].

The manifestation of T2DM is due to the combination of reduced insulin secretion from the β-cells of the pancreas and increased insulin resistance, as well as the disruption of incretin secretion from the gastrointestinal system. Several studies associated the microbiota with the development of T2DM, which is particularly characterized by a decrease in the concentrations of the phylum *Bacillota* (such as the genus *Roseburia* from the *Lachnospiraceae* family and the *Enterococcus faecalis* from genus *Enterococcus*, and other). Changes in the number of *Bifidobacteria*, *Lactobacillaceae* phylum, *Clostridioides* genus, as well as the *Bacillota*/*Bacteroidota* ratio have also been observed in the intestinal microbiota of children with T1D[91-93]. Obesity and T2D share the presence of low-grade inflammation that occurs in tissues involved in the regulation of metabolism, such as the liver, adipose tissue, and muscle. Inflammation is characterized by an increase in cytokines, interleukin (IL)-6, IL-1 and tumor necrosis factor-alpha with the result being insulin resistance. There are several studies in which the gut microbiota has been associated with the presence of obesity, insulin resistance and T2D, such as, and that probiotic treatment affects the metabolic control of patients with T2D. The gut microbiota contributes to the development of T2D[94]. Both studies showed that people with T2D had reduced concentrations of *Clostridiales* bacteria (*Roseburia* spp*.* and *Enterococcus faecalis*)[95]. T2D, independent of obesity, may also affect the structural composition of the microbiota. T2D may have a high presence of Gram-negative intestinal bacteria, such as *Bacteroidota*. In fact, in diabetic mice a reduction in *Bacteroides/Prevotella* spp*.* has been linked to an improvement in metabolic endotoxemia and lowering of laboratory tests of inflammation[96]. Changes in the number of *Bifidobacterium*, *Lactobacillaceae* phylum, *Clostridium* as well as the ratio *Bacillota*/*Bacteroidota* have also been observed in the intestinal microbiota of children with T1D[97]. Similar changes in the composition of the intestinal microbiota have been reported in patients with T2D. A lot of different theories have been proposed about the biomechanisms to explain the effect of the intestinal microbiota on insulin resistance and T2D, the main ones being metabolic inflammation, modification of incretin secretion and the production of hydroxybutyric acid. Lipopolysaccharides are endotoxins that are usually found in the outer membrane of Gram-negative bacteria and cause so-called metabolic inflammation, which is characterized by the release of pro-inflammatory factors[98]. The role of lipopolysaccharides in the pathogenesis of metabolic diseases was demonstrated by a study in mice fed a normal diet, in which the infusion of lipopolysaccharides caused insulin resistance in the liver, glucose intolerance, as well as an increase in adipose tissue. In addition, lipopolysaccharides may can lead to the expression of NF-κB and stimulate the activity of mitogen-activated protein kinase metabolic pathways in adipocytes. A study in fed leptin-deficient mice physiologically showed that increased intestinal permeability to lipopolysaccharide resulted in a change in the proportion of Gram-negative bacteria in the intestinal lumen, which was associated with the presence of insulin resistance. The modification of the intestinal microbiota by administering probiotic treatment to obese mice acted favourably on the intestinal barrier, reducing lipopolysaccharide-induced metabolic inflammation[99]. It has been found that increasing concentrations of *Bifidobacterium* modifies the inflammatory response in obese mice by increasing the production of glucagon like peptides (GLPs), while reducing intestinal permeability. The increase in *Bifidobacterium* concentrations induced by probiotic treatment is probably associated with an increase in the levels of gut-secreted peptides GLP-1 and YY, which exert a favourable effect, reducing insulin resistance and improving β-cell function. In addition, probiotic treatment caused an increase in GLP-2 levels in the colon, improved intestinal barrier function, and ultimately reduced plasma lipopolysaccharide levels[100]. It was found that people with T2D had an increase in the number of various opportunistic gut pathogens and a decrease in the concentrations of hydroxybutyric acid-producing bacteria. Hydroxybutyric acid is the main source of energy for maintaining the function of the cells of the digestive system. In the large intestine, hydroxybutyric acid is mainly produced by the bacteria *Clostridium coccoides* and *Eubacterium rectale*. However, changes in gut bacteria were found in colon cancer patients and in elderly subjects, suggesting that hydroxybutyric acid-producing bacteria could potentially have a protective role in the functioning of the gut microbiota[101,102].

***Obesity and atherosclerosis***

In obese individuals the accumulation and thus the increase of energy is related to the transfer of hydrogen between certain bacterial taxa. In fact, this effect has been noted both from the hydrogen-producing *Prevotellaceae* and from the methanogenic archaea that use hydrogen[103]. In obese individuals this increase and accumulation of energy due to the relative abundance of *Gammaproteobacteria* and the less presence of *Clostridium* genus. Additionally, obese individuals harbour clusters of H2-producing bacteria, primarily members of the family *Prevotellaceae* and some groups of *Bacillota*. These H2-producing bacteria coexist in the gastrointestinal tracts of obese individuals with relatively large numbers of H2-oxidizing methanogens belonging to archaea. Methanogens account up to 10% of all anaerobes in the colon[104,105]. Plant polysaccharides and dietary fibers are fermented by intestinal bacteria to produce SCFAs. An increase in methanogenic oxidation of H2 facilitates fermentation, which produces more SCFAs[106]. It can also be utilized directly by hydrotrophic methanogenic agents, while propionate, butyrate and lactate can be fermented with acetate and H2, where the latter is utilized by hydrotrophic methanogenic bacteria. As a consequence, the increase in methane oxidation should increase the conversion of plant polysaccharides into SCFAs, especially the acetate. SCFAs produced by fermentative bacteria are absorbed through the human intestinal epithelium, whereas H2 is an energy exchange factor within bacterial communities[107].

Atherosclerotic vascular disease is caused by environmental and genetic factors such as food and associated microorganisms. We currently know three circulating phospholipid-related molecules considered promoters of atherosclerosis: Trimethylamine N-oxide (TMAO), choline and betaine that could be used as biomarkers to predict cardiovascular disease risk[108]. These three phospholipid-related molecules were identified by analysis of plasma metabolites from 50 patients with atherosclerotic disease using liquid chromatography/mass spectrometry compared to 50 healthy samples. It has been noted that used apoE-deficient mice as a model of atherosclerosis and showed that plasma TMAO levels in apoE-deficient mice correlated positively with the area of aortic damage. The activity level of hepatic flavin monooxygenases, which convert trimethylamine (TMA) to TMAO, correlated positively with plasma TMAO levels in both mice and humans[109,110]. In the apoE-deficient mice an antibiotic treatment modify plasma TMAO level and atherosclerosis size, suggesting that gut microbiota significantly influence the development of atherosclerosis in apoE-deficient mice reducing it. This was demonstrated by adding 1% choline to the diets of apoE-deficient mice, as it increased the formation of foam cells as well as the expression of the scavenger receptors CD36 and SRA1 on macrophages, which is normally prevented by the administration of broad-spectrum antibiotics. A new pathway linking dietary lipid intake, gut microbiota and atherosclerosis has been found. Dietary L-carnitine is metabolized to TMA by gut microbiota and further converted to TMAO in the liver, which accelerates atherosclerosis in mice[111]. 16S rRNA sequencing of cecum bacteria in mice fed a diet supplemented with L-carnitine showed that the family *Prevotellaceae* were increased and positively correlated with plasma TMA level[112]. Furthermore, circulating TMAO in individuals with non-selective diet seems to be much higher than in those with a vegetables-based diet. Faecal microbiota analysis and plasma TMAO levels have been correlated in individuals with *Prevotella* enterotype in respect of those with *Bacteroides* enterotype, showing that these are higher in the first. L-carnitine use in individuals with different diet habits showed that a varied diet led to the production of more TMAO than in vegans or vegetarians. This confirm that both diet and lifestyle can change both the gut microbiota composition and its ability to metabolize TMA and TMAO from dietary L-carnitine[113]. Based on metagenomics, the genus *Collinsella* in faecal microbiota seems to be increased in patients with symptomatic atherosclerosis, while the genera *Eubacterium* and *Roseburia* is abundant in healthy subjects. Furthermore, in patients with atherosclerosis, metabolomics of faecal microbiota suggests an increase of the expression of genes coding for peptidoglycan synthesis, and a fall of hydrogenase levels. In addition, serum carotenoids, especially β-carotene, were decreased suggesting that symptomatic patients with atherosclerosis could have changes in the gut microbiota and a basic inflammatory state[113,114].

***Neurological and psychiatric disturbance***

As for the brain, everyone knows that it sends messages throughout the body. The intestine seems to respond. One of the most interesting effects of probiotics on the body focuses on the presence of the gut-brain axis (GBA). The GBA is the connection and the two-way communication between the gut [enteric nervous system (ENS)] and the central nervous system (CNS). This axis is a complex two-way pathway essential for metabolism homeostasis, the influence it has on emotions, mood and in general higher cognitive functions. It is a complex system in which it participates the CNS, the autonomic nervous system, the brain, the spinal cord, and the HPA axis[2,115]. The GBA axis indicates the bidirectional relationship and interdependence between CNS and ENS. It is important to understand this interaction and how a healthy intestinal microbiota can affect the body’s nervous system and vice versa. Through a multitude of mechanisms, hundreds of substances and neurotransmitters, the relationship between the two complex systems is in constant change which can have both positive and negative effects. Stress is a quite common factor that can affect mood, psychology and, apparently, the body’s intestinal microbiota[116,117]. When the organism exposed to a stressful social situation even for a period of only two hours, the microbiota undergoes significant changes, a significant change in its profile and change in the ratio of the main races of bacteria. Additionally, the brain, under the right conditions, can influence composition and functionality of the intestinal microbiota. It may alter intestinal permeability thereby allowing bacterial antigens to enter the epithelium and stimulate a mucosal immune response. Acute stress can increase colonic permeability, leading to overproduction of interferon-γ[116]. Finally, dysbiosis, *i.e.,* alterations caused in the gut due to stress, facilitate the expression of infectious bacteria. An example is the secretion of norepinephrine during surgery which causes expression of *Pseudomonas aeruginosa*, which can lead to intestinal sepsis. In addition, norepinephrine can stimulate the proliferation of enteric pathogens and increase the infectious properties of *Campylobacter jejuni*[117-119]. Finally, it can favour the overgrowth of non-pathogenic *Escherichia coli* isolates, as well as pathogenic *Escherichia coli* type 0157:H7[3,116]. In continuation of the above, it seems that probiotics have the mechanisms to deal with the complications of stress. Cortisol is a substance produced when the organism is in a state of stress and experiences situations related to anxiety and depression. It was found that probiotics led to a decrease in its release of this substance. In addition, the metabolic products of probiotics, SCFAs, offer beneficial actions. The ENS becomes a recipient of bacterial metabolites. SCFAs such as butyric, acetic, and propionic acids are the main metabolic products of bacterial metabolism[120]. In addition to their essential presence for the multitude of beneficial actions they offer, they can stimulate the sympathetic nervous system, release serotonin in the mucosa and positively affect memory and the learning process[116]. The above-mentioned negative effects of unpleasant psychological states indicate this strong dependence and relationship between CNS and ENS. It is worth focusing on the reverse course, that is, how the health of the body’s microbiota can, through probiotic bacteria, give the appropriate signals to the brain to influence possible diseases. Studies show that balance in the microbiota is related to our emotions, as well as how our brains process information from our senses, such as sights, sounds, tastes. Scientists suspect that gut microbiota disorders may play a role in autism spectrum disorders, depression, anxiety, and chronic pain[121]. The GBA is a communication system between the gut and the brain *via* neural, hormonal, and immune circuits, offering the gut microbiota and its metabolites a potential pathway to access the brain. This communication system is bidirectional and allows the brain to regulate gastrointestinal functions such as peristaltic movements and mucus production as well as immune functions[122]. Considerable progress has been made in the past decade in understanding the ways in which the gut microbiota is linked to the brain. Stress conditions affects the gut microbiota composition and that two-way communication between gut microbiota and the CNS influences the host’s response to stress. Stress has been shown to affect the integrity of the intestinal epithelium and alter peristalsis, secretions, and mucus production, thereby altering the gut microbiota environment and causing changes in microbial composition and/or metabolism[2,123]. Finally, regarding autism, it has been found that children with autism typically have a higher abundance of *Pseudomonadota*, *Bacteroidota* and a lower abundance of *Bacillota* and *Bifidobacteria* than healthy children. In fact, many classes of bacteria that make up *Bacillota* and *Clostridia* spp*.*, have been found to appear in higher percentages in autistic children with a history of gastrointestinal problems, while at the same time and despite the overall higher abundance of *Bacteroidota*, and lower *Prevotella* were observed. Therefore, in addition to quantifying the relative increase or decrease of the populations of some phyla, it has been deemed necessary to determine the populations of specific intestinal symbiotic organisms to understand the significance of some physiological changes in the gut and/or brain[124].

***Colorectal cancer***

Colonic metabolism may be protective against carcinogenesis under eubiotic conditions[125] (Figure 3). Several environmental and individual factors associated with the development of colorectal cancer, and the interactions between them. It is estimated that 20% of human cancers are related to conditions of chronic inflammation and/or persistent infections. As for example *Helicobacter pylori* infection is often associated with ulcer and gastric cancer, hepatitis B virus and hepatitis C virus can facilitate hepatocellular carcinoma, IBDs can develop colorectal cancer[126]. The composition of the gut microbiota according to studies may contribute to the mechanism of carcinogenesis either through diet or through its anti-inflammatory effect on the gut mucosa. A number of research has showed a link between *Fusobacterium* genus and the development of colorectal cancer. Most recently an association between *Fusobacterium nucleatum* and colorectal cancer was established by a molecular cytogenetic technique[127,128].

The gut microbiota in patients with colorectal cancer is characterized by increased diversity in *Clostridiaceae* family (such as those from *Clostridium* genus), and an increase in *Bacteroides* and *Bifidobacterium* spp. In contrast, the gut microbiota of individuals with a reduced risk of developing cancer has a high population of lactate-producing bacteria, such as *Eubacterium aerofaciens* and *Lactobacillaceae* phyla[129]. Studies on animals led to propose a model of carcinogenesis based on the consideration that certain mutations in the intestinal epithelial cells lead to a loosening of the intercellular junctions and a reduced production of mucus, compromising the integrity of the intestinal mucosa. So, there is a translocation of bacteria from the lumen to the epithelium where microbial products bind to tumor-associated macrophage receptors inducing the release of inflammatory factors (such as IL-1, IL-6, IL-23) which in turn leads to the production of IL-17 stimulating T-helper lymphocytes. IL-17 activates the transcription factor signal transducer and activator of transcription 3 in epithelial cells which will lead to increased survival and proliferation of epithelial cells resulting in further mutations facilitating carcinogenesis[130,131]. These changes in the epithelium burden the already compromised epithelial integrity, exacerbating bacterial allostasis and contributing to the vicious cycle: Bacterial allostasis - inflammation - cancer. The IL-23/IL-17 axis in tumour-induced inflammation is caused by barrier loss. Thus, otherwise “good” bacteria in the gut microbiota can be transformed into a carcinogen due to the altered host defence mechanism, indicating the uniqueness of the mucosal environment in preventing the development of cancer and may can lead to a cancer progression in other sites such as make the gut/breast microbiota’s axis[132-135].

**CONCLUSION**

A large population of microorganisms of various composition gradually colonizes the human organism, externally and internally, and forms the so-called microbiota. The most important and then populous is the intestinal one. The organism represents a stable and nutrient-rich habitat for the gut microbiota to thrive, and therefore the health of the host is of primary importance to them. In turn, the host benefits because they not only prevent colonization by pathogens, but also provide a continuous and dynamic effect on their host’s homeostasis. Thus, this symbiosis creates the host/gut microbiota axis that influences its health throughout an individual’s life. In fact, a gut microbiota disorder has been related to many diseases (such as asthma, diabetes, obesity, neurological disorders, *etc.*) but also psychic and mental disorders. However, not all correlation mechanisms are yet understood, which is why more research is needed into the composition of the microbiota that is the cause or result of a disease. The aim of this study is on the human microbiota and its composition for the correlation of conditions during the life course that can be physiological or pathological. In fact, the scientific community is still trying to understand and relate its development and change to various pathological conditions that occur in humans.

**REFERENCES**

1 **Cresci GA**, Bawden E. Gut Microbiome: What We Do and Don't Know. *Nutr Clin Pract* 2015; **30**: 734-746 [PMID: 26449893 DOI: 10.1177/0884533615609899]

2 **Santacroce L**, Man A, Charitos IA, Haxhirexha K, Topi S. Current knowledge about the connection between health status and gut microbiota from birth to elderly. A narrative review. *Front Biosci (Landmark Ed)* 2021; **26**: 135-148 [PMID: 34162042 DOI: 10.52586/4930]

3 **Duncan SH**, Lobley GE, Holtrop G, Ince J, Johnstone AM, Louis P, Flint HJ. Human colonic microbiota associated with diet, obesity and weight loss. *Int J Obes (Lond)* 2008; **32**: 1720-1724 [PMID: 18779823 DOI: 10.1038/ijo.2008.155]

4 **Conlon MA**, Bird AR. The impact of diet and lifestyle on gut microbiota and human health. *Nutrients* 2014; **7**: 17-44 [PMID: 25545101 DOI: 10.3390/nu7010017]

5 **Ferraris RP**, Vinnakota RR. Intestinal nutrient transport in genetically obese mice. *Am J Clin Nutr* 1995; **62**: 540-546 [PMID: 7661115 DOI: 10.1093/ajcn/62.3.540]

6 **Di Domenico M**, Ballini A, Boccellino M, Scacco S, Lovero R, Charitos IA, Santacroce L. The Intestinal Microbiota May Be a Potential Theranostic Tool for Personalized Medicine. *J Pers Med* 2022; **12** [PMID: 35455639 DOI: 10.3390/jpm12040523]

7 **Lazar V**, Ditu LM, Pircalabioru GG, Picu A, Petcu L, Cucu N, Chifiriuc MC. Gut Microbiota, Host Organism, and Diet Trialogue in Diabetes and Obesity. *Front Nutr* 2019; **6**: 21 [PMID: 30931309 DOI: 10.3389/fnut.2019.00021]

8 **Thursby E**, Juge N. Introduction to the human gut microbiota. *Biochem J* 2017; **474**: 1823-1836 [PMID: 28512250 DOI: 10.1042/BCJ20160510]

9 **Jeffery IB**, O'Toole PW. Diet-microbiota interactions and their implications for healthy living. *Nutrients* 2013; **5**: 234-252 [PMID: 23344252 DOI: 10.3390/nu5010234]

10 **Ravcheev DA**, Thiele I. Systematic genomic analysis reveals the complementary aerobic and anaerobic respiration capacities of the human gut microbiota. *Front Microbiol* 2014; **5**: 674 [PMID: 25538694 DOI: 10.3389/fmicb.2014.00674]

11 **Yang T**, Owen JL, Lightfoot YL, Kladde MP, Mohamadzadeh M. Microbiota impact on the epigenetic regulation of colorectal cancer. *Trends Mol Med* 2013; **19**: 714-725 [PMID: 24051204 DOI: 10.1016/j.molmed.2013.08.005]

12 **Robles Alonso V**, Guarner F. Linking the gut microbiota to human health. *Br J Nutr* 2013; **109** Suppl 2: S21-S26 [PMID: 23360877 DOI: 10.1017/S0007114512005235]

13 **Ianiro G**, Bruno G, Lopetuso L, Beghella FB, Laterza L, D'Aversa F, Gigante G, Cammarota G, Gasbarrini A. Role of yeasts in healthy and impaired gut microbiota: the gut mycome. *Curr Pharm Des* 2014; **20**: 4565-4569 [PMID: 24180411 DOI: 10.2174/13816128113196660723]

14 **Inchingolo F**, Santacroce L, Cantore S, Ballini A, Del Prete R, Topi S, Saini R, Dipalma G, Arrigoni R. Probiotics and EpiCor® in human health. *J Biol Regul Homeost Agents* 2019; **33**: 1973-1979 [PMID: 31858774 DOI: 10.23812/19-543-L]

15 **Isacco CG**, Ballini A, De Vito D, Nguyen KCD, Cantore S, Bottalico L, Quagliuolo L, Boccellino M, Di Domenico M, Santacroce L, Arrigoni R, Dipalma G, Inchingolo F. Rebalancing the Oral Microbiota as an Efficient Tool in Endocrine, Metabolic and Immune Disorders. *Endocr Metab Immune Disord Drug Targets* 2021; **21**: 777-784 [PMID: 32727337 DOI: 10.2174/1871530320666200729142504]

16 **Rodríguez JM**, Murphy K, Stanton C, Ross RP, Kober OI, Juge N, Avershina E, Rudi K, Narbad A, Jenmalm MC, Marchesi JR, Collado MC. The composition of the gut microbiota throughout life, with an emphasis on early life. *Microb Ecol Health Dis* 2015; **26**: 26050 [PMID: 25651996 DOI: 10.3402/mehd.v26.26050]

17 **Vaahtovuo J**, Toivanen P, Eerola E. Study of murine faecal microflora by cellular fatty acid analysis; effect of age and mouse strain. *Antonie Van Leeuwenhoek* 2001; **80**: 35-42 [PMID: 11761365 DOI: 10.1023/a:1012058107731]

18 **Clarke SF**, Murphy EF, O'Sullivan O, Lucey AJ, Humphreys M, Hogan A, Hayes P, O'Reilly M, Jeffery IB, Wood-Martin R, Kerins DM, Quigley E, Ross RP, O'Toole PW, Molloy MG, Falvey E, Shanahan F, Cotter PD. Exercise and associated dietary extremes impact on gut microbial diversity. *Gut* 2014; **63**: 1913-1920 [PMID: 25021423 DOI: 10.1136/gutjnl-2013-306541]

19 **Avershina E**, Storrø O, Øien T, Johnsen R, Pope P, Rudi K. Major faecal microbiota shifts in composition and diversity with age in a geographically restricted cohort of mothers and their children. *FEMS Microbiol Ecol* 2014; **87**: 280-290 [PMID: 24112053 DOI: 10.1111/1574-6941.12223]

20 **De Filippo C**, Di Paola M, Ramazzotti M, Albanese D, Pieraccini G, Banci E, Miglietta F, Cavalieri D, Lionetti P. Diet, Environments, and Gut Microbiota. A Preliminary Investigation in Children Living in Rural and Urban Burkina Faso and Italy. *Front Microbiol* 2017; **8**: 1979 [PMID: 29081768 DOI: 10.3389/fmicb.2017.01979]

21 **Santacroce L**, Charitos IA, Ballini A, Inchingolo F, Luperto P, De Nitto E, Topi S. The Human Respiratory System and its Microbiome at a Glimpse. *Biology (Basel)* 2020; **9** [PMID: 33019595 DOI: 10.3390/biology9100318]

22 **Jandhyala SM**, Talukdar R, Subramanyam C, Vuyyuru H, Sasikala M, Nageshwar Reddy D. Role of the normal gut microbiota. *World J Gastroenterol* 2015; **21**: 8787-8803 [PMID: 26269668 DOI: 10.3748/wjg.v21.i29.8787]

23 **David LA**, Maurice CF, Carmody RN, Gootenberg DB, Button JE, Wolfe BE, Ling AV, Devlin AS, Varma Y, Fischbach MA, Biddinger SB, Dutton RJ, Turnbaugh PJ. Diet rapidly and reproducibly alters the human gut microbiome. *Nature* 2014; **505**: 559-563 [PMID: 24336217 DOI: 10.1038/nature12820]

24 **Santacroce L**, Imbimbo C, Ballini A, Crocetto F, Scacco S, Cantore S, Di Zazzo E, Colella M, Jirillo E. Testicular Immunity and Its Connection with the Microbiota. Physiological and Clinical Implications in the Light of Personalized Medicine. *J Pers Med* 2022; **12** [PMID: 36013286 DOI: 10.3390/jpm12081335]

25 **De Filippis F**, Pellegrini N, Vannini L, Jeffery IB, La Storia A, Laghi L, Serrazanetti DI, Di Cagno R, Ferrocino I, Lazzi C, Turroni S, Cocolin L, Brigidi P, Neviani E, Gobbetti M, O'Toole PW, Ercolini D. High-level adherence to a Mediterranean diet beneficially impacts the gut microbiota and associated metabolome. *Gut* 2016; **65**: 1812-1821 [PMID: 26416813 DOI: 10.1136/gutjnl-2015-309957]

26 **Lovreglio P**, Bukvic N, Fustinoni S, Ballini A, Drago I, Foà V, Guanti G, Soleo L. Lack of genotoxic effect in workers exposed to very low doses of 1,3-butadiene. *Arch Toxicol* 2006; **80**: 378-381 [PMID: 16307232 DOI: 10.1007/s00204-005-0046-0]

27 **Santacroce L**, Muzio EL, Bottalico L, Spirito F, Charitos IA, Passarelli PC, Jirillo E. Subversion of the Oral Microbiota and Induction of Immune-Mediated Systemic Inflammation with Special Reference to Periodontitis: Current Knowledge and Perspectives. *Endocr Metab Immune Disord Drug Targets* 2023; **23**: 470-484 [PMID: 35770408 DOI: 10.2174/1871530322666220629101357]

28 **Dominianni C**, Sinha R, Goedert JJ, Pei Z, Yang L, Hayes RB, Ahn J. Sex, body mass index, and dietary fiber intake influence the human gut microbiome. *PLoS One* 2015; **10**: e0124599 [PMID: 25874569 DOI: 10.1371/journal.pone.0124599]

29 **Baranowski RW**, Skelly LE, Josse AR, Fajardo VA. Exploring the Effects of Greek Yogurt Supplementation and Exercise Training on Serum Lithium and Its Relationship With Musculoskeletal Outcomes in Men. *Front Nutr* 2021; **8**: 798036 [PMID: 35004824 DOI: 10.3389/fnut.2021.798036]

30 **Contuzzi N**, Casalino G, Boccaccio A, Ballini A, Charitos IA, Bottalico L, Santacroce L. Metals Biotribology and Oral Microbiota Biocorrosion Mechanisms. *J Funct Biomater* 2022; **14** [PMID: 36662061 DOI: 10.3390/jfb14010014]

31 **Arslan N**. Obesity, fatty liver disease and intestinal microbiota. *World J Gastroenterol* 2014; **20**: 16452-16463 [PMID: 25469013 DOI: 10.3748/wjg.v20.i44.16452]

32 **Wegierska AE**, Charitos IA, Topi S, Potenza MA, Montagnani M, Santacroce L. The Connection Between Physical Exercise and Gut Microbiota: Implications for Competitive Sports Athletes. *Sports Med* 2022; **52**: 2355-2369 [PMID: 35596883 DOI: 10.1007/s40279-022-01696-x]

33 **Savin Z**, Kivity S, Yonath H, Yehuda S. Smoking and the intestinal microbiome. *Arch Microbiol* 2018; **200**: 677-684 [PMID: 29626219 DOI: 10.1007/s00203-018-1506-2]

34 **Santacroce L**, Di Domenico M, Montagnani M, Jirillo E. Antibiotic Resistance and Microbiota Response. *Curr Pharm Des* 2023; **29**: 356-364 [PMID: 36537602 DOI: 10.2174/1381612829666221219093450]

35 **Ramirez J**, Guarner F, Bustos Fernandez L, Maruy A, Sdepanian VL, Cohen H. Antibiotics as Major Disruptors of Gut Microbiota. *Front Cell Infect Microbiol* 2020; **10**: 572912 [PMID: 33330122 DOI: 10.3389/fcimb.2020.572912]

36 **Bottalico L**, Charitos IA, Potenza MA, Montagnani M, Santacroce L. The war against bacteria, from the past to present and beyond. *Expert Rev Anti Infect Ther* 2022; **20**: 681-706 [PMID: 34874223 DOI: 10.1080/14787210.2022.2013809]

37 **Xu Z**, Knight R. Dietary effects on human gut microbiome diversity. *Br J Nutr* 2015; **113** Suppl: S1-S5 [PMID: 25498959 DOI: 10.1017/S0007114514004127]

38 **Vernocchi P**, Del Chierico F, Putignani L. Gut Microbiota Metabolism and Interaction with Food Components. *Int J Mol Sci* 2020; **21** [PMID: 32456257 DOI: 10.3390/ijms21103688]

39 **Rowland I**, Gibson G, Heinken A, Scott K, Swann J, Thiele I, Tuohy K. Gut microbiota functions: metabolism of nutrients and other food components. *Eur J Nutr* 2018; **57**: 1-24 [PMID: 28393285 DOI: 10.1007/s00394-017-1445-8]

40 **Bottalico L**, Castellaneta F, Charitos IA. From hydrotherapy to the discovery of the gut microbiota: the historical gastrointestinal health concept. *Pharmacophore* 2020; **11**: 82-90

41 **Ndeh D**, Munoz Munoz J, Cartmell A, Bulmer D, Wills C, Henrissat B, Gray J. The human gut microbe Bacteroides thetaiotaomicron encodes the founding member of a novel glycosaminoglycan-degrading polysaccharide lyase family PL29. *J Biol Chem* 2018; **293**: 17906-17916 [PMID: 30262663 DOI: 10.1074/jbc.RA118.004510]

42 **Turnbaugh PJ**, Ley RE, Mahowald MA, Magrini V, Mardis ER, Gordon JI. An obesity-associated gut microbiome with increased capacity for energy harvest. *Nature* 2006; **444**: 1027-1031 [PMID: 17183312 DOI: 10.1038/nature05414]

43 **Murphy EF**, Cotter PD, Healy S, Marques TM, O'Sullivan O, Fouhy F, Clarke SF, O'Toole PW, Quigley EM, Stanton C, Ross PR, O'Doherty RM, Shanahan F. Composition and energy harvesting capacity of the gut microbiota: relationship to diet, obesity and time in mouse models. *Gut* 2010; **59**: 1635-1642 [PMID: 20926643 DOI: 10.1136/gut.2010.215665]

44 **Arrigoni R**, Cammarota F, Porro R, Cantore S, Dioguardi M, Cazzolla AP, De Leonardis F, Polimeno L, Zerman N, Di Cosola M, Mastrangelo F, Santacroce L, Ballini A. Natural Bioactive Compounds against Oxidative Stress: Dietary Polyphenols Strike Back. *Endocr Metab Immune Disord Drug Targets* 2023; **23**: 764-776 [PMID: 36345247 DOI: 10.2174/1871530323666221107092553]

45 **Dao MC**, Everard A, Aron-Wisnewsky J, Sokolovska N, Prifti E, Verger EO, Kayser BD, Levenez F, Chilloux J, Hoyles L; MICRO-Obes Consortium, Dumas ME, Rizkalla SW, Doré J, Cani PD, Clément K. Akkermansia muciniphila and improved metabolic health during a dietary intervention in obesity: relationship with gut microbiome richness and ecology. *Gut* 2016; **65**: 426-436 [PMID: 26100928 DOI: 10.1136/gutjnl-2014-308778]

46 **Samuel BS**, Hansen EE, Manchester JK, Coutinho PM, Henrissat B, Fulton R, Latreille P, Kim K, Wilson RK, Gordon JI. Genomic and metabolic adaptations of Methanobrevibacter smithii to the human gut. *Proc Natl Acad Sci U S A* 2007; **104**: 10643-10648 [PMID: 17563350 DOI: 10.1073/pnas.0704189104]

47 **Schneeberger M**, Everard A, Gómez-Valadés AG, Matamoros S, Ramírez S, Delzenne NM, Gomis R, Claret M, Cani PD. Akkermansia muciniphila inversely correlates with the onset of inflammation, altered adipose tissue metabolism and metabolic disorders during obesity in mice. *Sci Rep* 2015; **5**: 16643 [PMID: 26563823 DOI: 10.1038/srep16643]

48 **Goodrich JK**, Waters JL, Poole AC, Sutter JL, Koren O, Blekhman R, Beaumont M, Van Treuren W, Knight R, Bell JT, Spector TD, Clark AG, Ley RE. Human genetics shape the gut microbiome. *Cell* 2014; **159**: 789-799 [PMID: 25417156 DOI: 10.1016/j.cell.2014.09.053]

49 **Hrncir T**, Hrncirova L, Kverka M, Hromadka R, Machova V, Trckova E, Kostovcikova K, Kralickova P, Krejsek J, Tlaskalova-Hogenova H. Gut Microbiota and NAFLD: Pathogenetic Mechanisms, Microbiota Signatures, and Therapeutic Interventions. *Microorganisms* 2021; **9** [PMID: 33946843 DOI: 10.3390/microorganisms9050957]

50 **Santacroce L**, Sardaro N, Topi S, Pettini F, Bottalico L, Cantore S, Cascella G, Del Prete R, Dipalma G, Inchingolo F. The pivotal role of oral microbiota in health and disease. *J Biol Regul Homeost Agents* 2020; **34**: 733-737 [PMID: 32492992 DOI: 10.23812/20-127-L-45]

51 **Belkaid Y**, Hand TW. Role of the microbiota in immunity and inflammation. *Cell* 2014; **157**: 121-141 [PMID: 24679531 DOI: 10.1016/j.cell.2014.03.011]

52 **Mahowald MA**, Rey FE, Seedorf H, Turnbaugh PJ, Fulton RS, Wollam A, Shah N, Wang C, Magrini V, Wilson RK, Cantarel BL, Coutinho PM, Henrissat B, Crock LW, Russell A, Verberkmoes NC, Hettich RL, Gordon JI. Characterizing a model human gut microbiota composed of members of its two dominant bacterial phyla. *Proc Natl Acad Sci U S A* 2009; **106**: 5859-5864 [PMID: 19321416 DOI: 10.1073/pnas.0901529106]

53 **Rook GA**. Regulation of the immune system by biodiversity from the natural environment: an ecosystem service essential to health. *Proc Natl Acad Sci U S A* 2013; **110**: 18360-18367 [PMID: 24154724 DOI: 10.1073/pnas.1313731110]

54 **Gieryńska M**, Szulc-Dąbrowska L, Struzik J, Mielcarska MB, Gregorczyk-Zboroch KP. Integrity of the Intestinal Barrier: The Involvement of Epithelial Cells and Microbiota-A Mutual Relationship. *Animals (Basel)* 2022; **12** [PMID: 35049768 DOI: 10.3390/ani12020145]

55 **Ma J**, Piao X, Mahfuz S, Long S, Wang J. The interaction among gut microbes, the intestinal barrier and short chain fatty acids. *Anim Nutr* 2022; **9**: 159-174 [PMID: 35573092 DOI: 10.1016/j.aninu.2021.09.012]

56 **Amarante-Mendes GP**, Adjemian S, Branco LM, Zanetti LC, Weinlich R, Bortoluci KR. Pattern Recognition Receptors and the Host Cell Death Molecular Machinery. *Front Immunol* 2018; **9**: 2379 [PMID: 30459758 DOI: 10.3389/fimmu.2018.02379]

57 **Cochet F**, Peri F. The Role of Carbohydrates in the Lipopolysaccharide (LPS)/Toll-Like Receptor 4 (TLR4) Signalling. *Int J Mol Sci* 2017; **18** [PMID: 29099761 DOI: 10.3390/ijms18112318]

58 **Liu T**, Zhang L, Joo D, Sun SC. NF-κB signaling in inflammation. *Signal Transduct Target Ther* 2017; **2**: 17023-17023 [PMID: 29158945 DOI: 10.1038/sigtrans.2017.23]

59 **Paone P**, Cani PD. Mucus barrier, mucins and gut microbiota: the expected slimy partners? *Gut* 2020; **69**: 2232-2243 [PMID: 32917747 DOI: 10.1136/gutjnl-2020-322260]

60 **Kennedy EA**, King KY, Baldridge MT. Mouse Microbiota Models: Comparing Germ-Free Mice and Antibiotics Treatment as Tools for Modifying Gut Bacteria. *Front Physiol* 2018; **9**: 1534 [PMID: 30429801 DOI: 10.3389/fphys.2018.01534]

61 **Qi R**, Sun J, Qiu X, Zhang Y, Wang J, Wang Q, Huang J, Ge L, Liu Z. The intestinal microbiota contributes to the growth and physiological state of muscle tissue in piglets. *Sci Rep* 2021; **11**: 11237 [PMID: 34045661 DOI: 10.1038/s41598-021-90881-5]

62 **Hou K**, Wu ZX, Chen XY, Wang JQ, Zhang D, Xiao C, Zhu D, Koya JB, Wei L, Li J, Chen ZS. Microbiota in health and diseases. *Signal Transduct Target Ther* 2022; **7**: 135 [PMID: 35461318 DOI: 10.1038/s41392-022-00974-4]

63 **Tavakoli P**, Vollmer-Conna U, Hadzi-Pavlovic D, Grimm MC. A Review of Inflammatory Bowel Disease: A Model of Microbial, Immune and Neuropsychological Integration. *Public Health Rev* 2021; **42**: 1603990 [PMID: 34692176 DOI: 10.3389/phrs.2021.1603990]

64 **Ghoshal UC**, Shukla R, Ghoshal U, Gwee KA, Ng SC, Quigley EM. The gut microbiota and irritable bowel syndrome: friend or foe? *Int J Inflam* 2012; **2012**: 151085 [PMID: 22577594 DOI: 10.1155/2012/151085]

65 **Rajilić-Stojanović M**, Biagi E, Heilig HG, Kajander K, Kekkonen RA, Tims S, de Vos WM. Global and deep molecular analysis of microbiota signatures in fecal samples from patients with irritable bowel syndrome. *Gastroenterology* 2011; **141**: 1792-1801 [PMID: 21820992 DOI: 10.1053/j.gastro.2011.07.043]

66 **Jeffery IB**, O'Toole PW, Öhman L, Claesson MJ, Deane J, Quigley EM, Simrén M. An irritable bowel syndrome subtype defined by species-specific alterations in faecal microbiota. *Gut* 2012; **61**: 997-1006 [PMID: 22180058 DOI: 10.1136/gutjnl-2011-301501]

67 **Saulnier DM**, Riehle K, Mistretta TA, Diaz MA, Mandal D, Raza S, Weidler EM, Qin X, Coarfa C, Milosavljevic A, Petrosino JF, Highlander S, Gibbs R, Lynch SV, Shulman RJ, Versalovic J. Gastrointestinal microbiome signatures of pediatric patients with irritable bowel syndrome. *Gastroenterology* 2011; **141**: 1782-1791 [PMID: 21741921 DOI: 10.1053/j.gastro.2011.06.072]

68 **Vijay A**, Valdes AM. Role of the gut microbiome in chronic diseases: a narrative review. *Eur J Clin Nutr* 2022; **76**: 489-501 [PMID: 34584224 DOI: 10.1038/s41430-021-00991-6]

69 **Khan I**, Ullah N, Zha L, Bai Y, Khan A, Zhao T, Che T, Zhang C. Alteration of Gut Microbiota in Inflammatory Bowel Disease (IBD): Cause or Consequence? IBD Treatment Targeting the Gut Microbiome. *Pathogens* 2019; **8** [PMID: 31412603 DOI: 10.3390/pathogens8030126]

70 **Caparrós E**, Wiest R, Scharl M, Rogler G, Gutiérrez Casbas A, Yilmaz B, Wawrzyniak M, Francés R. Dysbiotic microbiota interactions in Crohn's disease. *Gut Microbes* 2021; **13**: 1949096 [PMID: 34313550 DOI: 10.1080/19490976.2021.1949096]

71 **Ferreira C**, Veldhoen M. Host and microbes date exclusively. *Cell* 2012; **149**: 1428-1430 [PMID: 22726431 DOI: 10.1016/j.cell.2012.06.005]

72 **Russell SL**, Gold MJ, Hartmann M, Willing BP, Thorson L, Wlodarska M, Gill N, Blanchet MR, Mohn WW, McNagny KM, Finlay BB. Early life antibiotic-driven changes in microbiota enhance susceptibility to allergic asthma. *EMBO Rep* 2012; **13**: 440-447 [PMID: 22422004 DOI: 10.1038/embor.2012.32]

73 **Sjögren YM**, Jenmalm MC, Böttcher MF, Björkstén B, Sverremark-Ekström E. Altered early infant gut microbiota in children developing allergy up to 5 years of age. *Clin Exp Allergy* 2009; **39**: 518-526 [PMID: 19220322 DOI: 10.1111/j.1365-2222.2008.03156.x]

74 **Rose NR**. Prediction and Prevention of Autoimmune Disease in the 21st Century: A Review and Preview. *Am J Epidemiol* 2016; **183**: 403-406 [PMID: 26888748 DOI: 10.1093/aje/kwv292]

75 **Topi S**, Bottalico L, Charitos IA, Colella M, Di Domenico M, Palmirotta R, Santacroce L. Biomolecular Mechanisms of Autoimmune Diseases and Their Relationship with the Resident Microbiota: Friend or Foe? *Pathophysiology* 2022; **29**: 507-536 [PMID: 36136068 DOI: 10.3390/pathophysiology29030041]

76 **Kucera P**, Nováková D, Behanová M, Novak J, Tlaskalová-Hogenová H, Andel M. Gliadin, endomysial and thyroid antibodies in patients with latent autoimmune diabetes of adults (LADA). *Clin Exp Immunol* 2003; **133**: 139-143 [PMID: 12823288 DOI: 10.1046/j.1365-2249.2003.02205.x]

77 **Tlaskalová-Hogenová H**, Stěpánková R, Kozáková H, Hudcovic T, Vannucci L, Tučková L, Rossmann P, Hrnčíř T, Kverka M, Zákostelská Z, Klimešová K, Přibylová J, Bártová J, Sanchez D, Fundová P, Borovská D, Srůtková D, Zídek Z, Schwarzer M, Drastich P, Funda DP. The role of gut microbiota (commensal bacteria) and the mucosal barrier in the pathogenesis of inflammatory and autoimmune diseases and cancer: contribution of germ-free and gnotobiotic animal models of human diseases. *Cell Mol Immunol* 2011; **8**: 110-120 [PMID: 21278760 DOI: 10.1038/cmi.2010.67]

78 **Tannock GW**, Savage DC. Influences of dietary and environmental stress on microbial populations in the murine gastrointestinal tract. *Infect Immun* 1974; **9**: 591-598 [PMID: 4593471 DOI: 10.1128/iai.9.3.591-598.1974]

79 **Nadal I**, Donant E, Ribes-Koninckx C, Calabuig M, Sanz Y. Imbalance in the composition of the duodenal microbiota of children with coeliac disease. *J Med Microbiol* 2007; **56**: 1669-1674 [PMID: 18033837 DOI: 10.1099/jmm.0.47410-0]

80 **Collado MC**, Donat E, Ribes-Koninckx C, Calabuig M, Sanz Y. Imbalances in faecal and duodenal Bifidobacterium species composition in active and non-active coeliac disease. *BMC Microbiol* 2008; **8**: 232 [PMID: 19102766 DOI: 10.1186/1471-2180-8-232]

81 **Mårild K**, Stephansson O, Montgomery S, Murray JA, Ludvigsson JF. Pregnancy outcome and risk of celiac disease in offspring: a nationwide case-control study. *Gastroenterology* 2012; **142**: 39-45.e3 [PMID: 21995948 DOI: 10.1053/j.gastro.2011.09.047]

82 **Decker E**, Engelmann G, Findeisen A, Gerner P, Laass M, Ney D, Posovszky C, Hoy L, Hornef MW. Cesarean delivery is associated with celiac disease but not inflammatory bowel disease in children. *Pediatrics* 2010; **125**: e1433-e1440 [PMID: 20478942 DOI: 10.1542/peds.2009-2260]

83 **Durazzo M**, Ferro A, Gruden G. Gastrointestinal Microbiota and Type 1 Diabetes Mellitus: The State of Art. *J Clin Med* 2019; **8** [PMID: 31684011 DOI: 10.3390/jcm8111843]

84 **Yadav H**, Jain S, Sinha PR. Antidiabetic effect of probiotic dahi containing Lactobacillus acidophilus and Lactobacillus casei in high fructose fed rats. *Nutrition* 2007; **23**: 62-68 [PMID: 17084593 DOI: 10.1016/j.nut.2006.09.002]

85 **Elding Larsson H**, Vehik K, Gesualdo P, Akolkar B, Hagopian W, Krischer J, Lernmark Å, Rewers M, Simell O, She JX, Ziegler A, Haller MJ; TEDDY Study Group. Children followed in the TEDDY study are diagnosed with type 1 diabetes at an early stage of disease. *Pediatr Diabetes* 2014; **15**: 118-126 [PMID: 24034790 DOI: 10.1111/pedi.12066]

86 **Membrez M**, Blancher F, Jaquet M, Bibiloni R, Cani PD, Burcelin RG, Corthesy I, Macé K, Chou CJ. Gut microbiota modulation with norfloxacin and ampicillin enhances glucose tolerance in mice. *FASEB J* 2008; **22**: 2416-2426 [PMID: 18326786 DOI: 10.1096/fj.07-102723]

87 **Rebello CJ**, Burton J, Heiman M, Greenway FL. Gastrointestinal microbiome modulator improves glucose tolerance in overweight and obese subjects: A randomized controlled pilot trial. *J Diabetes Complications* 2015; **29**: 1272-1276 [PMID: 26424589 DOI: 10.1016/j.jdiacomp.2015.08.023]

88 **Cani PD**, Bibiloni R, Knauf C, Waget A, Neyrinck AM, Delzenne NM, Burcelin R. Changes in gut microbiota control metabolic endotoxemia-induced inflammation in high-fat diet-induced obesity and diabetes in mice. *Diabetes* 2008; **57**: 1470-1481 [PMID: 18305141 DOI: 10.2337/db07-1403]

89 **Mokhtari P**, Metos J, Anandh Babu PV. Impact of type 1 diabetes on the composition and functional potential of gut microbiome in children and adolescents: possible mechanisms, current knowledge, and challenges. *Gut Microbes* 2021; **13**: 1-18 [PMID: 34101547 DOI: 10.1080/19490976.2021.1926841]

90 **Whang A**, Nagpal R, Yadav H. Bi-directional drug-microbiome interactions of anti-diabetics. *EBioMedicine* 2019; **39**: 591-602 [PMID: 30553752 DOI: 10.1016/j.ebiom.2018.11.046]

91 **Moreno-Indias I**, Cardona F, Tinahones FJ, Queipo-Ortuño MI. Impact of the gut microbiota on the development of obesity and type 2 diabetes mellitus. *Front Microbiol* 2014; **5**: 190 [PMID: 24808896 DOI: 10.3389/fmicb.2014.00190]

92 **Ballini A**, Scacco S, Boccellino M, Santacroce L, Arrigoni R. Microbiota and Obesity: Where Are We Now? *Biology (Basel)* 2020; **9** [PMID: 33255588 DOI: 10.3390/biology9120415]

93 **Iatcu CO**, Steen A, Covasa M. Gut Microbiota and Complications of Type-2 Diabetes. *Nutrients* 2021; **14** [PMID: 35011044 DOI: 10.3390/nu14010166]

94 **Zhang L**, Chu J, Hao W, Zhang J, Li H, Yang C, Yang J, Chen X, Wang H. Gut Microbiota and Type 2 Diabetes Mellitus: Association, Mechanism, and Translational Applications. *Mediators Inflamm* 2021; **2021**: 5110276 [PMID: 34447287 DOI: 10.1155/2021/5110276]

95 **Craciun CI**, Neag MA, Catinean A, Mitre AO, Rusu A, Bala C, Roman G, Buzoianu AD, Muntean DM, Craciun AE. The Relationships between Gut Microbiota and Diabetes Mellitus, and Treatments for Diabetes Mellitus. *Biomedicines* 2022; **10** [PMID: 35203519 DOI: 10.3390/biomedicines10020308]

96 **Murri M**, Leiva I, Gomez-Zumaquero JM, Tinahones FJ, Cardona F, Soriguer F, Queipo-Ortuño MI. Gut microbiota in children with type 1 diabetes differs from that in healthy children: a case-control study. *BMC Med* 2013; **11**: 46 [PMID: 23433344 DOI: 10.1186/1741-7015-11-46]

97 **Sikalidis AK**, Maykish A. The Gut Microbiome and Type 2 Diabetes Mellitus: Discussing a Complex Relationship. *Biomedicines* 2020; **8** [PMID: 31936158 DOI: 10.3390/biomedicines8010008]

98 **Caesar R**, Reigstad CS, Bäckhed HK, Reinhardt C, Ketonen M, Lundén GÖ, Cani PD, Bäckhed F. Gut-derived lipopolysaccharide augments adipose macrophage accumulation but is not essential for impaired glucose or insulin tolerance in mice. *Gut* 2012; **61**: 1701-1707 [PMID: 22535377 DOI: 10.1136/gutjnl-2011-301689]

99 **Cani PD**, Possemiers S, Van de Wiele T, Guiot Y, Everard A, Rottier O, Geurts L, Naslain D, Neyrinck A, Lambert DM, Muccioli GG, Delzenne NM. Changes in gut microbiota control inflammation in obese mice through a mechanism involving GLP-2-driven improvement of gut permeability. *Gut* 2009; **58**: 1091-1103 [PMID: 19240062 DOI: 10.1136/gut.2008.165886]

100 **Dioguardi M**, Cantore S, Quarta C, Sovereto D, Zerman N, Pettini F, Muzio LL, Cosola MD, Santacroce L, Ballini A. Correlation between Diabetes Mellitus and Peri-implantitis: A Systematic Review. *Endocr Metab Immune Disord Drug Targets* 2023; **23**: 596-608 [PMID: 36281861 DOI: 10.2174/1871530323666221021100427]

101 **Markowiak-Kopeć P**, Śliżewska K. The Effect of Probiotics on the Production of Short-Chain Fatty Acids by Human Intestinal Microbiome. *Nutrients* 2020; **12** [PMID: 32316181 DOI: 10.3390/nu12041107]

102 **Zhang C**, Yin A, Li H, Wang R, Wu G, Shen J, Zhang M, Wang L, Hou Y, Ouyang H, Zhang Y, Zheng Y, Wang J, Lv X, Wang Y, Zhang F, Zeng B, Li W, Yan F, Zhao Y, Pang X, Zhang X, Fu H, Chen F, Zhao N, Hamaker BR, Bridgewater LC, Weinkove D, Clement K, Dore J, Holmes E, Xiao H, Zhao G, Yang S, Bork P, Nicholson JK, Wei H, Tang H, Zhang X, Zhao L. Dietary Modulation of Gut Microbiota Contributes to Alleviation of Both Genetic and Simple Obesity in Children. *EBioMedicine* 2015; **2**: 968-984 [PMID: 26425705 DOI: 10.1016/j.ebiom.2015.07.007]

103 **Marzullo P**, Di Renzo L, Pugliese G, De Siena M, Barrea L, Muscogiuri G, Colao A, Savastano S; Obesity Programs of nutrition, Education, Research and Assessment (OPERA) Group. From obesity through gut microbiota to cardiovascular diseases: a dangerous journey. *Int J Obes Suppl* 2020; **10**: 35-49 [PMID: 32714511 DOI: 10.1038/s41367-020-0017-1]

104 **Sarmiento F**, Leigh JA, Whitman WB. Genetic systems for hydrogenotrophic methanogens. *Methods Enzymol* 2011; **494**: 43-73 [PMID: 21402209 DOI: 10.1016/B978-0-12-385112-3.00003-2]

105 **Ballini A**, Cantore S, Fatone L, Montenegro V, De Vito D, Pettini F, Crincoli V, Antelmi A, Romita P, Rapone B, Miniello G, Perillo L, Grassi FR, Foti C. Transmission of nonviral sexually transmitted infections and oral sex. *J Sex Med* 2012; **9**: 372-384 [PMID: 22023797 DOI: 10.1111/j.1743-6109.2011.02515.x]

106 **Ungerfeld EM**. Corrigendum: Shifts in metabolic hydrogen sinks in the methanogenesis-inhibited ruminal fermentation: a meta-analysis. *Front Microbiol* 2015; **6**: 538 [PMID: 26082760 DOI: 10.3389/fmicb.2015.00538]

107 **Ilyas A**, Wijayasinghe YS, Khan I, El Samaloty NM, Adnan M, Dar TA, Poddar NK, Singh LR, Sharma H, Khan S. Implications of trimethylamine N-oxide (TMAO) and Betaine in Human Health: Beyond Being Osmoprotective Compounds. *Front Mol Biosci* 2022; **9**: 964624 [PMID: 36310589 DOI: 10.3389/fmolb.2022.964624]

108 **Xia X**, Li X, Xie F, Yuan G, Cheng D, Peng C. Non-targeted metabonomic analysis of plasma in patients with atherosclerosis by liquid chromatography-mass spectrometry. *Ann Transl Med* 2022; **10**: 133 [PMID: 35284547 DOI: 10.21037/atm-22-118]

109 **Wang X**, Ishimori N, Korstanje R, Rollins J, Paigen B. Identifying novel genes for atherosclerosis through mouse-human comparative genetics. *Am J Hum Genet* 2005; **77**: 1-15 [PMID: 15931593 DOI: 10.1086/431656]

110 **Getz GS**, Reardon CA. Diet, Microbes, and Murine Atherosclerosis. *Arterioscler Thromb Vasc Biol* 2018; **38**: 2269-2271 [PMID: 30354225 DOI: 10.1161/ATVBAHA.118.311513]

111 **Koeth RA**, Wang Z, Levison BS, Buffa JA, Org E, Sheehy BT, Britt EB, Fu X, Wu Y, Li L, Smith JD, DiDonato JA, Chen J, Li H, Wu GD, Lewis JD, Warrier M, Brown JM, Krauss RM, Tang WH, Bushman FD, Lusis AJ, Hazen SL. Intestinal microbiota metabolism of L-carnitine, a nutrient in red meat, promotes atherosclerosis. *Nat Med* 2013; **19**: 576-585 [PMID: 23563705 DOI: 10.1038/nm.3145]

112 **Karlsson FH**, Fåk F, Nookaew I, Tremaroli V, Fagerberg B, Petranovic D, Bäckhed F, Nielsen J. Symptomatic atherosclerosis is associated with an altered gut metagenome. *Nat Commun* 2012; **3**: 1245 [PMID: 23212374 DOI: 10.1038/ncomms2266]

113 **Ballini A**, Charitos IA, Cantore S, Topi S, Bottalico L, Santacroce L. About Functional Foods: The Probiotics and Prebiotics State of Art. *Antibiotics (Basel)* 2023; **12** [PMID: 37106999 DOI: 10.3390/antibiotics12040635]

114 **Carabotti M**, Scirocco A, Maselli MA, Severi C. The gut-brain axis: interactions between enteric microbiota, central and enteric nervous systems. *Ann Gastroenterol* 2015; **28**: 203-209 [PMID: 25830558]

115 **Bashir Y**, Khan AU. The interplay between the gut-brain axis and the microbiome: A perspective on psychiatric and neurodegenerative disorders. *Front Neurosci* 2022; **16**: 1030694 [PMID: 36389228 DOI: 10.3389/fnins.2022.1030694]

116 **Jacobson A**, Yang D, Vella M, Chiu IM. The intestinal neuro-immune axis: crosstalk between neurons, immune cells, and microbes. *Mucosal Immunol* 2021; **14**: 555-565 [PMID: 33542493 DOI: 10.1038/s41385-020-00368-1]

117 **Charitos IA**, Topi S, Gagliano-Candela R, De Nitto E, Polimeno L, Montagnani M, Santacroce L. The Toxic Effects of Endocrine Disrupting Chemicals (EDCs) on Gut Microbiota: Bisphenol A (BPA) A Review. *Endocr Metab Immune Disord Drug Targets* 2022; **22**: 716-727 [PMID: 35339192 DOI: 10.2174/1871530322666220325114045]

118 **Gao F**, Guo R, Ma Q, Li Y, Wang W, Fan Y, Ju Y, Zhao B, Gao Y, Qian L, Yang Z, He X, Jin X, Liu Y, Peng Y, Chen C, Chen Y, Gao C, Zhu F, Ma X. Stressful events induce long-term gut microbiota dysbiosis and associated post-traumatic stress symptoms in healthcare workers fighting against COVID-19. *J Affect Disord* 2022; **303**: 187-195 [PMID: 35157946 DOI: 10.1016/j.jad.2022.02.024]

119 **Montagnani M,** Bottalico L, Potenza MA, Charitos IA, Topi S, Colella M, Santacroce L. The Crosstalk between Gut Microbiota and Nervous System: A Bidirectional Interaction between Microorganisms and Metabolome. *Int J Mol Sci* 2023; **24** [PMID: 37373470 DOI: 10.3390/ijms241210322]

120 **Ganci M**, Suleyman E, Butt H, Ball M. The role of the brain-gut-microbiota axis in psychology: The importance of considering gut microbiota in the development, perpetuation, and treatment of psychological disorders. *Brain Behav* 2019; **9**: e01408 [PMID: 31568686 DOI: 10.1002/brb3.1408]

121 **O'Mahony SM**, Clarke G, Borre YE, Dinan TG, Cryan JF. Serotonin, tryptophan metabolism and the brain-gut-microbiome axis. *Behav Brain Res* 2015; **277**: 32-48 [PMID: 25078296 DOI: 10.1016/j.bbr.2014.07.027]

122 **Foster JA**, McVey Neufeld KA. Gut-brain axis: how the microbiome influences anxiety and depression. *Trends Neurosci* 2013; **36**: 305-312 [PMID: 23384445 DOI: 10.1016/j.tins.2013.01.005]

123 **Ghaisas S**, Maher J, Kanthasamy A. Gut microbiome in health and disease: Linking the microbiome-gut-brain axis and environmental factors in the pathogenesis of systemic and neurodegenerative diseases. *Pharmacol Ther* 2016; **158**: 52-62 [PMID: 26627987 DOI: 10.1016/j.pharmthera.2015.11.012]

124 **O'Keefe SJ**. Diet, microorganisms and their metabolites, and colon cancer. *Nat Rev Gastroenterol Hepatol* 2016; **13**: 691-706 [PMID: 27848961 DOI: 10.1038/nrgastro.2016.165]

125 **Hibino S**, Kawazoe T, Kasahara H, Itoh S, Ishimoto T, Sakata-Yanagimoto M, Taniguchi K. Inflammation-Induced Tumorigenesis and Metastasis. *Int J Mol Sci* 2021; **22** [PMID: 34063828 DOI: 10.3390/ijms22115421]

126 **McCoy AN**, Araújo-Pérez F, Azcárate-Peril A, Yeh JJ, Sandler RS, Keku TO. Fusobacterium is associated with colorectal adenomas. *PLoS One* 2013; **8**: e53653 [PMID: 23335968 DOI: 10.1371/journal.pone.0053653]

127 **Inamura K**, Hamada T, Bullman S, Ugai T, Yachida S, Ogino S. Cancer as microenvironmental, systemic and environmental diseases: opportunity for transdisciplinary microbiomics science. *Gut* 2022 [PMID: 35820782 DOI: 10.1136/gutjnl-2022-327209]

128 **Rebersek M**. Gut microbiome and its role in colorectal cancer. *BMC Cancer* 2021; **21**: 1325 [PMID: 34895176 DOI: 10.1186/s12885-021-09054-2]

129 **Zhang X,** Yu D, Wu D, Gao X, Shao F, Zhao M, Wang J, Ma J, Wang W, Qin X, Chen Y, Xia P, Wang S. Tissue-resident Lachnospiraceae family bacteria protect against colorectal carcinogenesis by promoting tumor immune surveillance.*Cell Host Microbe*2023; **31**: 418-432.e8 [PMID: 36893736 DOI: 10.1016/j.chom.2023.01.013]

130 **Polimeno L**, Barone M, Mosca A, Viggiani MT, Joukar F, Mansour-Ghanaei F, Mavaddati S, Daniele A, Debellis L, Bilancia M, Santacroce L, Di Leo A. Soy Metabolism by Gut Microbiota from Patients with Precancerous Intestinal Lesions. *Microorganisms* 2020; **8** [PMID: 32218321 DOI: 10.3390/microorganisms8040469]

131 **Murata M**. Inflammation and cancer. *Environ Health Prev Med* 2018; **23**: 50 [PMID: 30340457 DOI: 10.1186/s12199-018-0740-1]

132 **Polimeno L**, Barone M, Mosca A, Viggiani MT, Di Leo A, Debellis L, Troisi M, Daniele A, Santacroce L. Gut Microbiota Imbalance is Related to Sporadic Colorectal Neoplasms. A Pilot Study. *Appl Sci* 2019; **9**: 5491 [DOI: 10.3390/app9245491]

133 **Niekamp P,** Kim CH. Microbial Metabolite Dysbiosis and Colorectal Cancer. *Gut Liver* 2023; **17**: 190-203[PMID: 36632785 DOI: 10.5009/gnl220260]

134 **Marino MM**, Nastri BM, D'Agostino M, Risolo R, De Angelis A, Settembre G, Rienzo M, D'Esposito V, Abbondanza C, Formisano P, Ballini A, Santacroce L, Boccellino M, Di Domenico M. Does Gut-breast Microbiota Axis Orchestrates Cancer Progression? *Endocr Metab Immune Disord Drug Targets* 2022; **22**: 1111-1122 [PMID: 35362389 DOI: 10.2174/1871530322666220331145816]

135 **Charles A**, Thomas RM. The Influence of the microbiome on the innate immune microenvironment of solid tumors. *Neoplasia* 2023; **37**: 100878 [PMID: 36696837 DOI: 10.1016/j.neo.2023.100878]

**Footnotes**

**Conflict-of-interest statement:** All the authors report no relevant conflicts of interest for this article.

**Open-Access:** This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: https://creativecommons.org/Licenses/by-nc/4.0/

**Provenance and peer review:** Invited article; Externally peer reviewed.

**Peer-review model:** Single blind

**Peer-review started:** March 28, 2023

**First decision:** April 26, 2023

**Article in press:**

**Specialty type:** Gastroenterology and hepatology

**Country/Territory of origin:** Italy

**Peer-review report’s scientific quality classification**

Grade A (Excellent): A, A

Grade B (Very good): B

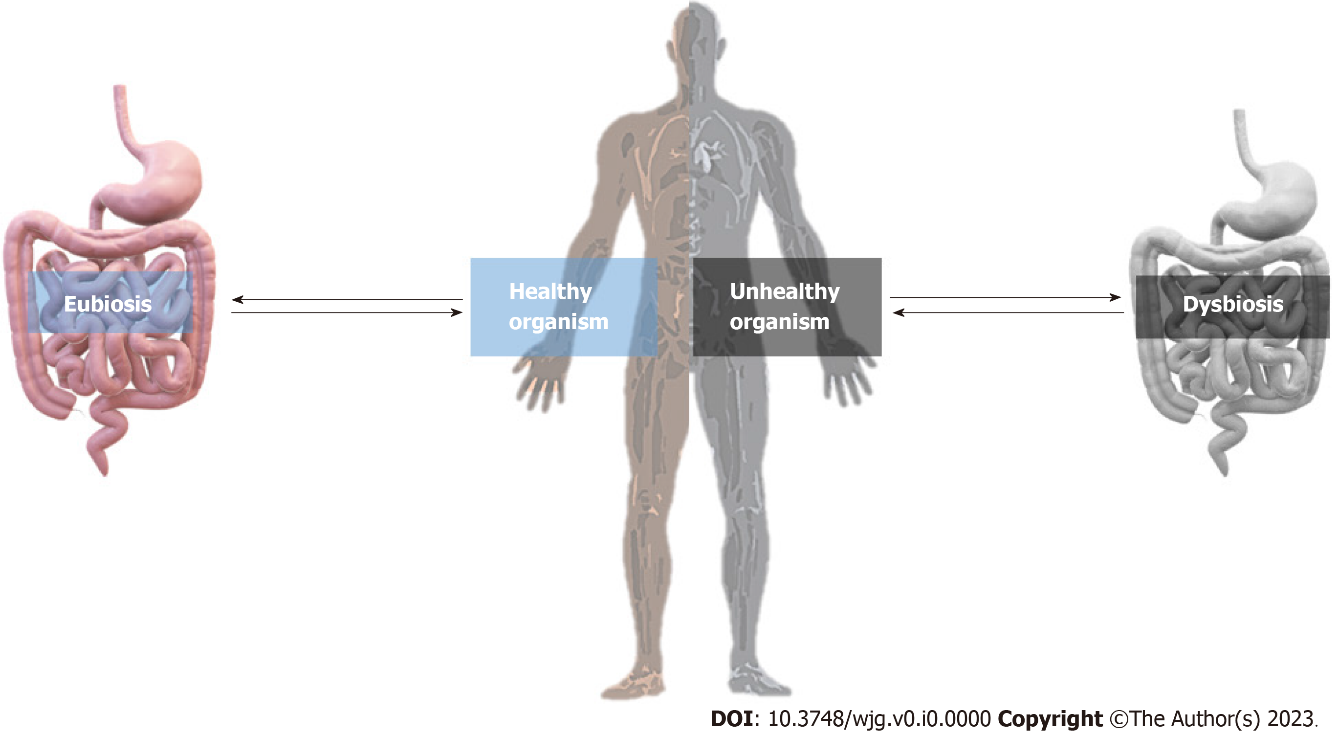
Grade C (Good): 0

Grade D (Fair): 0

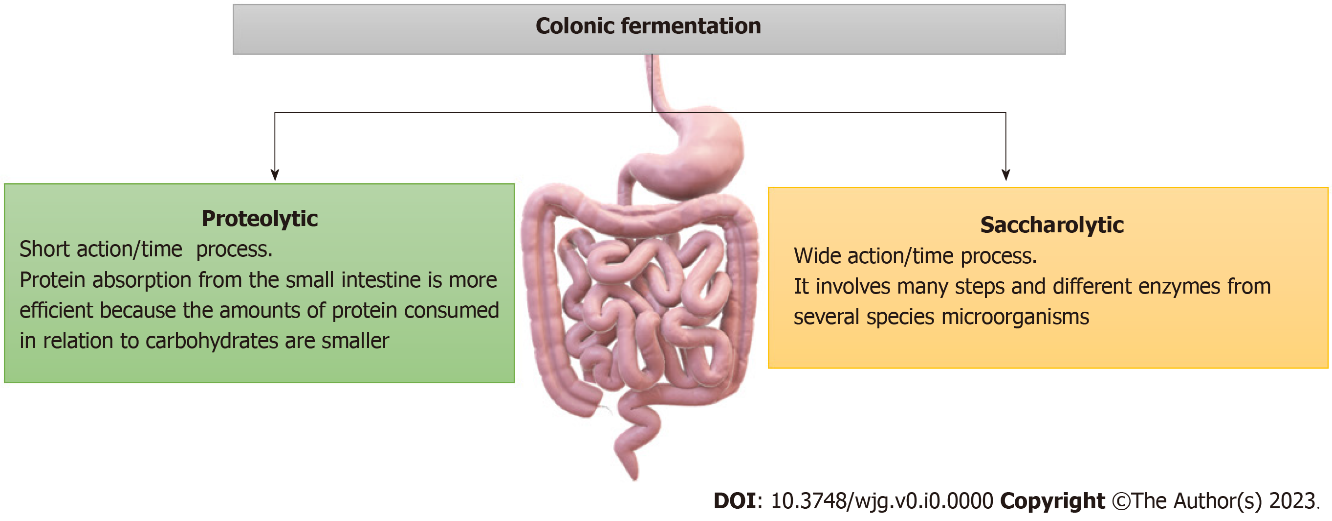
Grade E (Poor): 0

**P-Reviewer:** Ogino S, United States; Qin Y, China **S-Editor:** Wang JJ **L-Editor:** A **P-Editor:** Wang JJ

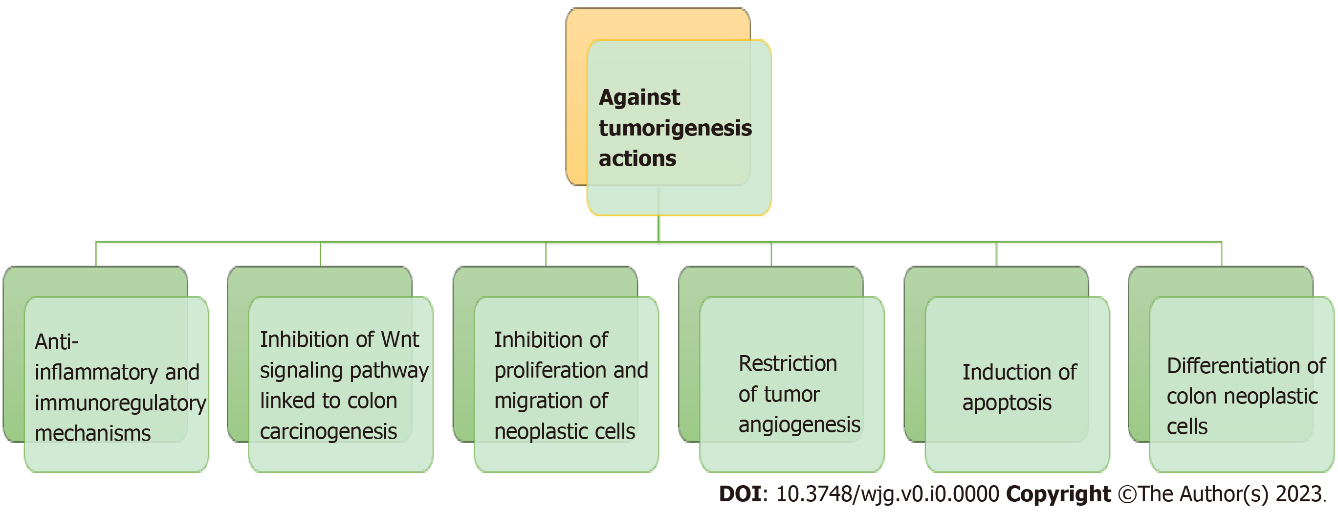
**Figure Legends**



**Figure 1 The cross-talking axis host/gut microbiota: The gastrointestinal microbiota plays an important role in host physiology, metabolism, and nutrition.** An organism with a regular maintenance of the physiological homeostasis leads to eubiosis of the gut microbiota and vice versa. Conversely, an altered physiological homeostasis leads to gut microbiota dysbiosis and vice versa. An alteration in the gut microbial community is linked to several disturbances gut conditions, including cancer, obesity, and a variety of gut disorders. The contribution of beneficial components of the gut microbiota to host physiology, metabolism, and immune function has become the focus of scientific research and will undoubtedly lead to new therapeutic approaches.



**Figure 2** **The metabolic activity by the colonic microbiota.** The intestinal microbiota finds an environment rich in polysaccharides which are not digested by stomach enzymes. Fermentation of polysaccharides by intestinal bacteria leads to the production of acetate, butyrate, and propionate, which are used as a carbon source by intestinal mucosal cells. The initial fermentation of the carbohydrate that escaped digestion in the small intestine is followed by the utilization and cross-distribution of metabolites by various members of the microbiota, and then the synthesis of short-chain fatty acids (butyrate, propionate, acetate). Proteolytic fermentation differs from saccharolytic fermentation because it releases many potentially toxic nitrogen and sulfur metabolites, such as ammonia, amines, nitrates, nitrites, and hydrogen sulfide[32,40].



**Figure 3 The inhibitory properties of butyrate on tumorigenesis through various mechanisms by the colonic microbiota.** Many of the metabolites of protein fermentation can be taken up by other microorganisms and synthesized into active carcinogens. For example, amines and nitrates can be used by facultatively anaerobic and anaerobic colonic bacteria and catalysed the formation of N-nitrosamines, which are among the strongest procarcinogens. Reduced levels of butyrate in the body are not only an indication of the possibility of cancer, but also indicate the severity of the cancer and its course in the body.