World J Diabetes 2023 July 15; 14(7): 939-1145





Published by Baishideng Publishing Group Inc

Contents

Monthly Volume 14 Number 7 July 15, 2023

OPINION REVIEW

939	Access to novel anti-diabetic agents in resource limited settings: A brief commentary
	Naidoo P, Naidoo K, Karamchand S, Leisegang RF

REVIEW

- 942 Detection, management, and prevention of diabetes-related foot disease in the Australian context McNeil S, Waller K, Poy Lorenzo YS, Mateevici OC, Telianidis S, Qi S, Churilov I, MacIsaac RJ, Galligan A
- 958 Novel insights regarding the role of noncoding RNAs in diabetes Macvanin MT, Gluvic Z, Bajic V, Isenovic ER
- 977 Implications of receptor for advanced glycation end products for progression from obesity to diabetes and from diabetes to cancer

Garza-Campos A, Prieto-Correa JR, Domínguez-Rosales JA, Hernández-Nazará ZH

- Advanced glycation end product signaling and metabolic complications: Dietary approach 995 Khan MI, Ashfaq F, Alsayegh AA, Hamouda A, Khatoon F, Altamimi TN, Alhodieb FS, Beg MMA
- 1013 Tight junction disruption and the pathogenesis of the chronic complications of diabetes mellitus: A narrative review

Robles-Osorio ML, Sabath E

MINIREVIEWS

- 1027 Klotho: A new therapeutic target in diabetic retinopathy? Puddu A, Maggi DC
- Type 2 diabetes and thyroid cancer: Synergized risk with rising air pollution 1037 Kruger EM, Shehata SA, Toraih EA, Abdelghany AA, Fawzy MS
- 1049 Liver or kidney: Who has the oar in the gluconeogenesis boat and when? Sahoo B, Srivastava M, Katiyar A, Ecelbarger C, Tiwari S

ORIGINAL ARTICLE

Basic Study

1057 Network-pharmacology-based research on protective effects and underlying mechanism of Shuxin decoction against myocardial ischemia/reperfusion injury with diabetes

Yang L, Jian Y, Zhang ZY, Qi BW, Li YB, Long P, Yang Y, Wang X, Huang S, Huang J, Zhou LF, Ma J, Jiang CQ, Hu YH, Xiao WJ



Contents

Monthly Volume 14 Number 7 July 15, 2023

1077 Analysis of N6-methyladenosine-modified mRNAs in diabetic cataract

Cai L, Han XY, Li D, Ma DM, Shi YM, Lu Y, Yang J

Retrospective Cohort Study

1091 Long-term quality-of-care score for predicting the occurrence of acute myocardial infarction in patients with type 2 diabetes mellitus

Li PI, Guo HR

Retrospective Study

1103 Correlation between glycated hemoglobin A1c, urinary microalbumin, urinary creatinine, β2 microglobulin, retinol binding protein and diabetic retinopathy

Song JJ, Han XF, Chen JF, Liu KM

Observational Study

1112 Glucose metabolism profile recorded by flash glucose monitoring system in patients with hypopituitarism during prednisone replacement

Han MM, Zhang JX, Liu ZA, Xu LX, Bai T, Xiang CY, Zhang J, Lv DQ, Liu YF, Wei YH, Wu BF, Zhang Y, Liu YF

1126 Association between cardiorespiratory fitness level and insulin resistance in adolescents with various obesity categories

La Grasta Sabolic L, Pozgaj Sepec M, Valent Moric B, Cigrovski Berkovic M

CASE REPORT

1137 Maturity-onset diabetes of the young type 9 or latent autoimmune diabetes in adults: A case report and review of literature

Zhou GH, Tao M, Wang Q, Chen XY, Liu J, Zhang LL



Contents

Monthly Volume 14 Number 7 July 15, 2023

ABOUT COVER

Editorial Board Member of World Journal of Diabetes, Sonia Eiras, BSc, PhD, Senior Researcher, Traslational Cardiology, Health Research Institute, University Hospital of Santiago de Compostela, Santiago de Compostela 15706, Spain. sonia.eiras.penas@sergas.es

AIMS AND SCOPE

The primary aim of World Journal of Diabetes (WJD, World J Diabetes) is to provide scholars and readers from various fields of diabetes with a platform to publish high-quality basic and clinical research articles and communicate their research findings online.

WID mainly publishes articles reporting research results and findings obtained in the field of diabetes and covering a wide range of topics including risk factors for diabetes, diabetes complications, experimental diabetes mellitus, type 1 diabetes mellitus, type 2 diabetes mellitus, gestational diabetes, diabetic angiopathies, diabetic cardiomyopathies, diabetic coma, diabetic ketoacidosis, diabetic nephropathies, diabetic neuropathies, Donohue syndrome, fetal macrosomia, and prediabetic state.

INDEXING/ABSTRACTING

The WID is now abstracted and indexed in Science Citation Index Expanded (SCIE, also known as SciSearch®), Current Contents/Clinical Medicine, Journal Citation Reports/Science Edition, PubMed, PubMed Central, Reference Citation Analysis, China National Knowledge Infrastructure, China Science and Technology Journal Database, and Superstar Journals Database. The 2023 Edition of Journal Citation Reports® cites the 2022 impact factor (IF) for WJD as 4.2; IF without journal self cites: 4.1; 5-year IF: 4.5; Journal Citation Indicator: 0.69; Ranking: 51 among 145 journals in endocrinology and metabolism; and Quartile category: Q2.

RESPONSIBLE EDITORS FOR THIS ISSUE

Production Editor: Yu-Xi Chen; Production Department Director: Xu Guo; Editorial Office Director: Jia-Ru Fan.

NAME OF JOURNAL	INSTRUCTIONS TO AUTHORS		
World Journal of Diabetes	https://www.wjgnet.com/bpg/gerinfo/204		
ISSN	GUIDELINES FOR ETHICS DOCUMENTS		
ISSN 1948-9358 (online)	https://www.wjgnet.com/bpg/GerInfo/287		
LAUNCH DATE	GUIDELINES FOR NON-NATIVE SPEAKERS OF ENGLISH		
June 15, 2010	https://www.wjgnet.com/bpg/gerinfo/240		
FREQUENCY	PUBLICATION ETHICS		
Monthly	https://www.wjgnet.com/bpg/GerInfo/288		
EDITORS-IN-CHIEF	PUBLICATION MISCONDUCT		
Lu Cai, Md. Shahidul Islam, Michael Horowitz	https://www.wjgnet.com/bpg/gerinfo/208		
EDITORIAL BOARD MEMBERS	ARTICLE PROCESSING CHARGE		
https://www.wjgnet.com/1948-9358/editorialboard.htm	https://www.wjgnet.com/bpg/gerinfo/242		
PUBLICATION DATE	STEPS FOR SUBMITTING MANUSCRIPTS		
July 15, 2023	https://www.wjgnet.com/bpg/GerInfo/239		
COPYRIGHT	ONLINE SUBMISSION		
© 2023 Baishideng Publishing Group Inc	https://www.f6publishing.com		

© 2023 Baishideng Publishing Group Inc. All rights reserved. 7041 Koll Center Parkway, Suite 160, Pleasanton, CA 94566, USA E-mail: bpgoffice@wjgnet.com https://www.wjgnet.com



WJD

Submit a Manuscript: https://www.f6publishing.com

World J Diabetes 2023 July 15; 14(7): 977-994

DOI: 10.4239/wjd.v14.i7.977

ISSN 1948-9358 (online)

REVIEW

Implications of receptor for advanced glycation end products for progression from obesity to diabetes and from diabetes to cancer

Andrea Garza-Campos, José Roberto Prieto-Correa, José Alfredo Domínguez-Rosales, Zamira Helena Hernández-Nazará

Specialty type: Endocrinology and metabolism

Provenance and peer review: Invited article; Externally peer reviewed.

Peer-review model: Single blind

Peer-review report's scientific quality classification

Grade A (Excellent): 0 Grade B (Very good): B, B, B Grade C (Good): C Grade D (Fair): 0 Grade E (Poor): 0

P-Reviewer: Pahlavani HA, Iran; Preziosi F, Italy; Srinivasu PN, India; Yang JS, China

Received: January 9, 2023 Peer-review started: January 9, 2023 First decision: January 17, 2023 Revised: January 31, 2023 Accepted: April 17, 2023 Article in press: April 17, 2023 Published online: July 15, 2023



Andrea Garza-Campos, José Roberto Prieto-Correa, Programa de Doctorado en Ciencias en Biología Molecular en Medicina, Universidad de Guadalajara, Guadalajara 44340, Jalisco, Mexico

Andrea Garza-Campos, José Roberto Prieto-Correa, José Alfredo Domínguez-Rosales, Zamira Helena Hernández-Nazará, Departamento de Biología Molecular y Genómica, Instituto de Investigación en Enfermedades Crónico-Degenerativas, Universidad de Guadalajara, Guadalajara 44340, Jalisco, Mexico

Corresponding author: Zamira Helena Hernández-Nazará, MD, PhD, Departamento de Biología Molecular y Genómica, Instituto de Investigación en Enfermedades Crónico-Degenerativas, Universidad de Guadalajara, Sierra Mojada 950, Col. Independencia C.P. 44350, Guadalajara 44340, Jalisco, Mexico. zamirahelena@yahoo.com.mx

Abstract

Obesity and type 2 diabetes mellitus (T2DM) are chronic pathologies with a high incidence worldwide. They share some pathological mechanisms, including hyperinsulinemia, the production and release of hormones, and hyperglycemia. The above, over time, affects other systems of the human body by causing tissue hypoxia, low-grade inflammation, and oxidative stress, which lay the pathophysiological groundwork for cancer. The leading causes of death globally are T2DM and cancer. Other main alterations of this pathological triad include the accumulation of advanced glycation end products and the release of endogenous alarmins due to cell death (i.e., damage-associated molecular patterns) such as the intracellular proteins high-mobility group box protein 1 and protein S100 that bind to the receptor for advanced glycation products (RAGE) - a multiligand receptor involved in inflammatory and metabolic and neoplastic processes. This review analyzes the latest advanced reports on the role of RAGE in the development of obesity, T2DM, and cancer, with an aim to understand the intracellular signaling mechanisms linked with cancer initiation. This review also explores inflammation, oxidative stress, hypoxia, cellular senescence, RAGE ligands, tumor microenvironment changes, and the "cancer hallmarks" of the leading tumors associated with T2DM. The assimilation of this information could aid in the development of diagnostic and therapeutic approaches to lower the morbidity and mortality associated with these diseases.

Raisbideng® WJD | https://www.wjgnet.com

Key Words: Type 2 diabetes; Cancer; Obesity; Advanced glycation end product receptor; Receptor for advanced glycation end products; Glycation end products, advanced

©The Author(s) 2023. Published by Baishideng Publishing Group Inc. All rights reserved.

Core Tip: The receptor for advanced glycation products (RAGE) is involved in every stage of the pathophysiological pathways that lead to the progression of obesity, type 2 diabetes, and cancer. This article provides a focused discussion on the stages of obesity leading to the development of metabolic diseases and provides a broad overview of the contribution of RAGE to the development of diabetes and cancer.

Citation: Garza-Campos A, Prieto-Correa JR, Domínguez-Rosales JA, Hernández-Nazará ZH. Implications of receptor for advanced glycation end products for progression from obesity to diabetes and from diabetes to cancer. World J Diabetes 2023; 14(7): 977-994 URL: https://www.wjgnet.com/1948-9358/full/v14/i7/977.htm DOI: https://dx.doi.org/10.4239/wjd.v14.i7.977

INTRODUCTION

Obesity, diabetes, and cancer are chronic diseases, the prevalences of which have all increased in parallel, and are leading causes of death worldwide[1]. However, the forecasts for these health problems are not encouraging. For example, the prevalence of diabetes is estimated to increase by 2045, specifically in middle-income countries to 21.1%, in high-income countries to 12.2%, and in low-income countries to 11.9%. Meanwhile, the incidence of malignant neoplasms in people under 50 years of age is also rising[2,3].

Although esophageal adenocarcinoma has a direct link to obesity, and pancreatic cancer can debut with type 2 diabetes mellitus (T2DM), there is an evident connection between the three disorders. Moreover, there is confusion about their shared lifestyle risk factors, including sedentariness and consumption of highly processed foods[4-6]. Regarding the common pathological mechanisms of obesity, T2DM, and cancer, expansion of adipose tissue (AT) results in the production of excess estrogen, adipokines, and inflammatory molecules that can lead to systemic or localized low-grade inflammation. In addition, omental and visceral adiposity is related to hyperinsulinemia and increased levels of insulinlike growth factor-1 (IGF-1)[7]. The metabolic abnormalities and lipo-glucotoxicity associated with insulin resistance and T2DM also cause an increase in inflammatory cytokines and oxidative stress. As a result, neoplastic processes can be triggered by T2DM and, likewise, obesity[8].

The pathogenic mechanisms that link obesity, T2DM, and cancer are complex and multifactorial. Because there is a notion of progression from obesity to T2DM towards cancer, our motivation for this review was to provide a detailed and up-to-date discussion on these mechanisms in the context of a single molecule known as the receptor for advanced glycation end products (RAGE). As such, this narrative review incorporates the conceptual framework and reports on findings extracted from two literature databases, the Reference Citation Analysis (https://www.referencecitationanalysis.com/) and PubMed, to provide a reflective discussion of RAGE's implications for the progression of obesity to T2DM and from T2DM to cancer.

RAGE is an immunoglobulin superfamily member and a type I pattern-recognition receptor. It is also a sensitive environmental sensor with several endogenous and external ligands. Furthermore, it is a widely expressed modulator of inflammatory and oxidative stress pathways with vast metabolic implications[9]. RAGE isoforms include soluble forms (sRAGE) that act as decoy receptors, sequester circulating ligands, and attenuate membrane RAGE signaling[10]. Soluble forms derived from membrane-localized RAGE are released into the circulation by proteolytic cleavage (cRAGE), and endogenously secreted RAGE (esRAGE) is formed by alternative splicing. In addition to the sRAGE isoforms and the fulllength membrane receptor (flRAGE) - the only isoform that participates in signal transduction, there are also the dominant-negative isoforms lacking the cytoplasmic tail and the truncated isoform lacking the V-type immunoglobulin domain[11] (Figure 1A).

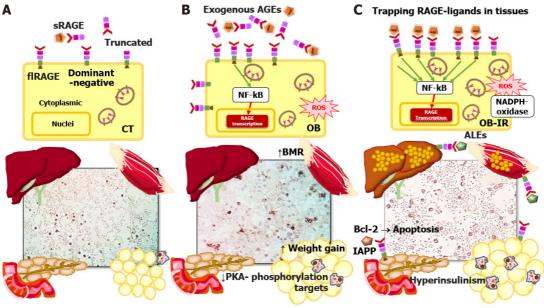
OBESITY AND T2DM

Initially, the function of RAGE was established in the context of chronic disease, specifically T2DM and its complications, in which persistent hyperglycemia triggers inflammation, oxidative stress, and endothelial damage[12,13]. However, there is more evidence that an increase in RAGE ligands is present in the early stages of metabolic dysfunction in obesity [14,15].

RAGE ligands

The most recognized ligands of RAGE are the advanced glycosylation end products (AGEs) and lipid oxidation adducts (ALEs). These are taken in from diet or produced by endogenous metabolism through non-enzymatic and spontaneous





DOI: 10.4239/wjd.v14.i7.977 Copyright ©The Author(s) 2023.

Figure 1 Receptor for advanced glycation products signaling and molecular mechanisms involved in progression from obesity to type 2 diabetes mellitus. Receptor for advanced glycation products (RAGE)-ligand signaling in healthy control subjects, obese individuals (OB), and OB with insulin resistance is illustrated. A: Full-length, total soluble, dominant-negative (intracytoplasmic, lacking domain), and truncated (lacking a V-terminal) RAGE isoforms; B: Basal metabolic rate increase in muscle, decreased phosphorylation targets of protein kinase A, and weight gain (adipose tissue) are findings in obesity related to increased RAGE isoforms and ligands; C: The mechanism trapping RAGE-ligand in tissues involves translocation of cytoplasmic RAGE to the membrane, inflammation (nuclear factor-kappa B), and oxidative stress (NADPH-oxidase) in peripheral mononuclear blood cells, liver, muscle, pancreas, and adipose tissue. The B cell lymphoma-2 proto-oncogene mediates RAGE apoptosis signaling in pancreatic beta cells and leads to type 2 diabetes mellitus. Advanced glycosylation end products; advanced lipoperoxidation end products, and islet amyloid polypeptide (also known as amyloid) are RAGE ligands. RAGE: Receptor for advanced glycation products; SRAGE: Soluble receptor for advanced glycation products; BMR: Basal metabolic rate; PKA: Protein kinase A; NF-kB: Nuclear factor-kappa B; PBMCs: Peripheral mononuclear blood cells; Bcl-2: B cell lymphoma-2; AGEs: Advanced glycosylation end products; IAPP: Islet amyloid polypeptide.

Maillard-type reactions in which proteins and nucleic acids react with carbohydrates, lipids, or their intermediate metabolites[16,17].

Foods cooked by roasting, grilling, frying, drying, heating, or adding artificial colorants, salt, oil, or sugar are often present in ultra-processed foods to make them suitable to store[6]. In addition to those above, an increase in the diet's caloric, fat, and glycemic indices leads to a significant rise in the levels of circulating AGEs. Some exogenous-derived food AGEs are Nδ-(5-hydro-5-methil-4-imidazolon-2-il)-ornithina (MG-H1), Nε-carboxyethyl lysine (CEL), and Nε-carboxy-methyl lysine (CML), in addition to the precursor methylglyoxal[18-20].

The problem gets worse when an individual also consumes other substances like alcohol and tobacco. Cigarettes are a source of AGEs, and smoking them causes RAGE expression to rise, which is linked to airway inflammation in chronic obstructive pulmonary disease and causes sRAGE to decrease in smoke-induced cardiovascular disease[21,22]. The increase in mitochondrial-derived reactive oxygen species (ROS) caused by the RAGE pathway in smoke-exposed skeletal muscle is one of the hypothesized mechanisms in this regard[23]. *In vitro*, oral squamous cell carcinoma treated with cigarette smoke extract showed an increase in RAGE with a link to a rise in invasive ability[24]. Additionally, RAGE is elevated in alcoholic liver disease, affecting blood triglycerides, low-density lipoprotein cholesterol, and alanine transaminase levels. RAGE also contributes to the accumulation of lipid droplets in the liver and modifies the expression of SREBP1, a transcription factor involved in lipid homeostasis[25].

Serum AGE accumulation from the diet can lead to cross-link formation that irreversibly changes endogenous proteins independent of glycemic control. Birukov *et al*[26] found that in people with prediabetes and T2DM, there were significant variations in the levels of AGEs in the skin. Additionally, AGE measurements in that study were related to factors such as waist circumference, glycated hemoglobin (commonly known as hemoglobin A1c) levels, C-reactive protein levels, and vascular stiffness. Further research is required to determine the sensitivity and accuracy of testing AGE accumulation and its relationship to disease status.

In addition to the above, other natural substances such as catechols, myeloperoxidase systems, and the polyol pathway are implicated in producing endogenous AGEs in obesity and states of insulin resistance[27,28]. Likewise, the link between AGEs in obesity and T2DM is the accumulation of lipids and their oxidized products. Thus, the accumulation of free fatty acids and subsequent ALE production aids in the progression of obesity to T2DM[29,30]. Oxidative stress promotes the lipoperoxidation of membranes and the production of metabolites such as 4-hydroxyl-trans2-nonenal, acrolein, aldehydes such as malondialdehyde (MDA), and ketoaldehydes such as 4-oxo-trans-2-nonenal. These may start with obesity and insulin resistance and can result in the creation of endogenous ALEs like MDA-Lys[17,31,32]. Further

Zaishidene® WJD | https://www.wjgnet.com

studies are required on the mechanism by which the progression from obesity to T2DM is affected by the ALEs-RAGE interaction and their aldehyde precursors produced by lipid peroxidation.

In this regard, in obese subjects, RAGE induces migration of macrophages because of the rise in lipid peroxidation and the accumulation of ALEs in renal tissue that leads to kidney injury[33]. Patients with T2DM have high levels of ALE (MDA-Lys), which induces the activation and adherence of monocytes to endothelial cells by increasing the expression of monocyte chemotactic protein-1 (MCP-1) and activating the nuclear factor-kappa B (NF-kB) pathway causing inflammation[34]. Recent comprehensive reviews have addressed endogenous and exogenous AGE and ALE formation in obesity[17], T2DM, and cancer[28,35,36].

There is consistent evidence regarding how ultra-processed foods, ALEs, and AGEs disrupt the microbiota causing dysbiosis, the subsequent translocation of lipopolysaccharide (LPS), and endotoxemia[37,38]. Likewise, dysbiosis is related to obesity, low-grade inflammation, and the progression of insulin resistance and T2DM[39]. However, few publications implicate RAGE as an LPS ligand to mediate inflammatory processes in obesity [40]. This issue needs further investigation, and an exciting future research opportunity may focus on T2DM prevention with respect to the relationship between AGEs/ALEs, RAGE, and dysbiosis.

According to the most recent definitions, chronic low-grade inflammation begins when molecules and metabolites, resulting from altered cell function and structure and foods, stimulate receptors and activate their signaling cascades with dysregulated energy homeostasis. To this end, RAGE mediates danger signals to the body and metabolic stress characteristic of innate immune systems, since RAGE detects ligands from microbes via exogenous pathogen-associated molecular patterns such as LPS. Furthermore, damage-associated molecular pattern (DAMP) ligands are derived from endogenous sources such as high-mobility group box protein 1 (HMGB1), S100/calgranulins, amyloid deposits like βamyloid peptide, and macrophage-1 antigen[41].

AGE and ALE metabolites can be considered DAMPs that are not derived from exogenous sources such as the diet, and the term "metabolism-associated molecular pattern" is proposed for these specific ligands. It is essential to differentiate between them and demonstrate that both endogenous and external components are involved in these responses [42]. An opportunity for experts in the field is to reach a consensus with respect to the classification of all exogenous and endogenous ligands for pattern-recognition receptors.

RAGE-trapping ligands

Several investigations in human subjects have found an association between obesity and low circulating AGE levels^[43]. Complex detoxification and clearance kinetics of AGEs could lead to inconsistent study results. The concept of entrapment of AGE in tissues proposes that AGEs are no longer circulating because they are trapped in tissues as metabolic risks increase in individuals[44-46] (Figure 1C).

For instance, high RAGE expression in AT is implicated in its dysfunction and is evidence of a link between RAGE signaling and the progression of obesity to associated metabolic disorder. A high level of RAGE expression in human epicardial AT is related to its thickening, low glucose transporter type 4 expression, and high HMGB1 expression[47]. In this context, visceral omental AT and fetal membrane samples from women with gestational diabetes revealed higher levels of RAGE and the HMGB1 ligand, respectively^[48]. RAGE signaling pathway proteins were also found to be expressed differently in omental and subcutaneous biopsies from obese people with healthy phenotypes. Subcutaneous AT showed a higher correlation between the RAGE signaling axis, inflammatory markers, and the homeostatic model assessment of insulin resistance (HOMA-IR)[49]. A study with a murine RAGE (-/-) model demonstrated protection against inflammation and oxidative stress and protection against insulin resistance. Interestingly, this model showed that the most beneficial characteristics of RAGE knockout were found in female mice^[50]. Additionally, RAGE is related to the adaptive thermogenesis function of brown AT through the decline in energy expenditure caused by a high-fat diet, possibly mediated *via* the accumulation of AGEs[51,52] (Figure 1B).

In addition to dysregulation in AT discussed above, chronic inflammation also plays a pivotal role in obesity-related insulin resistance that leads to metabolic dysfunction in the liver and muscle. Insulin resistance is characterized by alterations in insulin signaling in sensitive tissues, hyperinsulinemia with defects in glucose uptake in muscle and AT, impaired suppression of hepatic glucose production, and ectopic accumulation of fat in the muscle and liver through reesterification of fatty acids from AT[53,54] (Figure 1C). To this end, an increase in AGE accumulation in liver biopsies has been linked to RAGE expression, lipid accumulation, and the degree of liver damage without association with the measurements of sRAGE and circulating serum AGEs[55,56]. These studies demonstrate how RAGE affects hepatic conditions caused by the accumulation of AGEs in tissue in non-alcoholic liver disease.

RAGE expression and the accumulation of AGEs are linked to weight gain, inflammation, and oxidative stress markers in human muscle tissue [57]. For instance, one study demonstrated that RAGE expression and the accumulation of AGEs in skeletal muscle in a fructose-supplemented murine model were related to alterations in the oral glucose tolerance test curve, increased triglycerides, inflammatory response, increased basal metabolic rate, and resting metabolic rate[58]. Moreover, chronic AGE exposure is linked to sarcopenia[59]. However, the implications of obesity- and T2DM-induced RAGE expression in muscle tissue are less well explored in humans[60].

Along with the mechanism of trapping excess RAGE ligands in tissues, it is known that the sRAGE form eliminates dangerous circulating ligands and functions as a competitive inhibitor of ligands that might bind to cellular RAGE, supported by studies in which sRAGE levels were found to be low[61-64]. The role of sRAGE in metabolic diseases is debatable because it depends on the degree of disease development and the levels of cell and tissue damage[65]. The cRAGE levels are initially high in acute conditions, triggered by cleavage of flRAGE, which increases its AGE-binding activity. The main variations of sRAGE are attributed to the production of cRAGE shedding by metalloproteinases[66] to compensate for the increase in AGEs in the early stages of low-grade inflammation[67-69]. As the concentration of sRAGE decreases, sequestration and competitive inhibition of ligands decrease and as such they can reach cellular fIRAGE,

leading to an inflammatory response and subsequent tissue damage[68-70] (Figure 1C).

In prediabetes, plasma levels of sRAGE and esRAGE are all negatively correlated with the HOMA-IR index of insulin resistance and MDA. This correlation matches their reduction as insulin resistance develops in an oxidative environment [67]. Another study with similar results comparing healthy people to those with prediabetes and T2DM found low levels of esRAGE and an inverse linkage with S100A12[71]. Miranda et al[62] showed that all RAGE isoforms were lower when grouped by pancreatic dysfunction (*i.e.*, healthy controls, individuals with glucose intolerance, and those with T2DM). Thus, according to the above, the negative correlation of sRAGE with RAGE ligands or increase of the AGE/esRAGE index seems to be more related to individuals with obesity-related insulin resistance and early T2DM[72], and low cRAGE concentrations are a marker of aging[72,73]. Even the elevated AGE/esRAGE index could distinguish between those with non-alcoholic fatty liver disease without T2DM and healthy individuals[74]. Further studies are needed to determine the precise interactions between sRAGE, esRAGE, cRAGE, and their ligands in these disease states.

Since sRAGE and resting energy expenditure are related, one of the most recent discoveries regarding the expression of soluble variants is sRAGE's contribution to adaptive negative energy balance. In an investigation of the influence of sRAGE on the change in energy expenditure that occurs during weight loss, it was found that, under caloric restriction, adaptive changes arise that slow down energy expenditure. Specifically, after a 3-mo intervention for weight loss due to caloric restriction, energy expenditure increased by 52.6 kcal/d for each 100 pg/mL increase in basal sRAGE levels. Increases in esRAGE and cRAGE similarly translated to concomitant rises in energy expenditure, by 181.6 kcal/d and 56.1 kcal, respectively. This finding illustrates the potential impact of a RAGE feedback mechanism, in which a reduction in sRAGE could slow energy expenditure during weight loss[75]. Furthermore, one mechanism by which RAGE controls energy expenditure is through the suppression of adaptative thermogenesis in white and brown AT via the decline of β adrenergic signaling in adipocytes blocking protein kinase A (PKA) phosphorylation targets[76].

Still more, the subcellular localization of RAGE can change, a process related to oligomerization in the membrane after RAGE interaction with ligands[77]. A previous study demonstrated increased localization of RAGE in the cell membrane, rather than the cytoplasm, in peripheral blood mononuclear cells of obese individuals with insulin resistance compared with healthy individuals. As such, sRAGE correlates negatively with the HOMA-IR index and tissue damage markers[78] (Figure 1A-C). Peripheral blood mononuclear cells may provide an accessible platform to study the relationship between ligands and cellular RAGE, detect systemic inflammation, and relate these to tissue damage. The preceding argument needs to be tested by additional research.

In T2DM, the pancreas loses its ability to secrete enough insulin in response to meals. One of the mechanisms of pancreas failure is low-grade systemic inflammation. The activating signaling of RAGE in response to ligand binding results in RAGE autoregulation through the increase of its synthesis, which is mediated by NF-kB[79]. In vivo and in vitro models have shown that oxidative stress and inflammation are induced by AGE stimuli through NF-kB activation and the formation of ROS, respectively^[80]. These events are evidenced by the increase in the inflammatory serum marker Creactive protein, particularly in obesity[81]. Some antioxidants and drugs can modulate the AGEs-RAGE axis and the activation of NF-kB, leading to the reduction of lipid peroxidation products in obesity models[82-84].

RAGE expression in the pancreas may be an essential mechanism for the development of T2DM in humans, based on evidence from both in vitro and in vivo glycolipotoxicity studies[85-87]. In a rodent model of diet-induced hyperglycemia, endogenous AGE products are produced, and RAGE expression is observed in pancreatic islets[87]. RAGE inhibition prevented the increase of its expression, and decreased B cell lymphoma-2 (Bcl-2) expression and apoptosis of beta cells treated with glycation serum. However, RAGE inhibition did not restore the ability of the beta cells to secrete insulin in response to glucose[85]. RAGE endocytosis regulated by Rab31 ligand can inhibit apoptosis mediated by the pAkt/Bcl-2 pathway in beta cells treated with glycation serum [88]. In another study, the pancreas of db/db transgenic mice that lack the leptin receptor but express RAGE (+/+) have less beta cell mass and less apoptosis, is glucose intolerant, and has decreased insulin secretion. Likewise, when the MIN6 pancreatic beta cell line was treated with palmitate or oleate and leptin antagonists to induce RAGE expression, pancreatic damage occurred[86]. Another mouse model of diabetes induced by streptozotocin and a high-cholesterol diet treated with the water-soluble carotenoid crocin showed attenuated atrophic effects in pancreatic tissue and decreased blood glucose levels through decreases in the expression of RAGE and LOX-1[89].

DAMP/RAGE reports such as the activation of S100b/RAGE and the subsequent loss of beta cells by apoptosis via NADPH oxidase and the protection of sRAGE against amyloid deposition, beta cell loss, and glucose intolerance demonstrate that they interact[90,91]. All of these findings suggest that RAGE can lead to pancreatic failure and the progression of T2DM.

T2DM AND CANCER

Several studies have shown that the incidence of various malignancies increases in patients with T2DM. However, more rigorous statistical analyses of observational studies demonstrate a more significant association of T2DM with colorectal, pancreatic, hepatocellular, breast, and endometrial carcinomas. Even so, there are biases in these studies that make it challenging to study the confounding variables of T2DM leading to cancer[92]. A more recent study included statistical analysis of the "Mendelian randomization" studies to analyze genetic data from large-scale international consortia. Ultimately, it allowed to link a possible causal relationship between genetically predicted T2DM and endometrial and pancreatic cancer risks, and between the variable fasting insulin levels and breast cancer risk. In addition, numerous studies have demonstrated the impact of glycemic traits on the emergence of different malignancies, establishing a relationship between T2DM and cancer[93].



Metabolic and hormonal factors found in patients with obesity, insulin resistance, and T2DM, such as hyperinsulinism, hyperglycemia, IGF-1, adipokines, and estrogens, all of which are closely related to inflammation and oxidative stress, function in the long-term as risk factors that support transformation to neoplastic cells in diabetic patients[94].

Estrogens

The increase in estrogen levels in obese patients is due to the positive regulation of the aromatase enzyme, encoded by CYP19A1 and secreted by cells of the tumor stromal microenvironment. The activation mechanisms are triggered in response to hypoxia, with activation of hypoxia-inducible factor-1 alpha (HIF-1 α), fat tissue hormones (e.g., adipokine leptin, which increases aromatase expression by phosphorylating serine at position 485 of AMPK and inhibiting the aromatase suppressor), and inflammation processes[95]. Estrogen receptors are transcriptional factors of DNA reprogramming that transduce extranuclear signals, resulting in the regulation of ion channels or kinase cascades such as PKC/PKA/PI3-K/MAPK. The metabolic effects of estrogen on both tumor and normal cells are survival, cell proliferation, and immunomodulation[96]. Estrogens are the most relevant risk factor for endometrial and breast cancers, especially in postmenopausal women. Recently, studies have shown that the microbiota is a source of estrogen-like compounds or estrogen mimics that could be involved in cancer progression[97].

Hyperinsulinism and IGF-1

The insulin receptor (IR) and insulin receptor substrate (IRS) are phosphorylated at Ser/Thr residues by inflammatory cytokines and oxidative stress, resulting in insulin resistance and compensatory hyperinsulinemia[98]. Insulin induces proliferation in tissues not involved in metabolism. Binding to its receptor (i.e., IR) activates the RAS/RAF/MAPK kinase-dependent/ERK signaling pathways and increases cell survival and migration[99]. Another mechanism is mediated by IGF-1, a hormone structurally and functionally similar to insulin that binds to IR and its receptor (*i.e.*, IGFR). This receptor, like IR, activates pathways that increase cell proliferation, and insulin enhances the liver's production of IGF-1, elevating the mitogenic activity of cancer cells expressing the IGFR[7,100]. The nuclear protein HMGA1 contributes to the potentiation of insulin action. In addition, the HMGA1 protein overexpressed in triple-negative breast cancer cells functions in chromatin remodeling and gene expression regulation, indirectly promoting enhanced IR expression through the inhibitory effect on p53 expression, which usually keeps IR expression turned off.

Hyperglycemia

Although hyperglycemia is the primary cause of T2DM pathophysiological abnormalities, it also contributes to the development of cancer through several processes that either directly or indirectly harm DNA, RNA, lipids, and proteins. The production of ROS, accumulation of mutations and inhibition of their repair, alteration of the immune system, alteration of metabolism, and activation of oncogenes and inactivation tumor suppressor genes are some of the carcinogenic effects that result from the formation of AGEs through non-enzymatic reactions and the subsequent activation of RAGE[101]. Endogenous AGEs are categorized according to their precursor as follows: Glyoxal (GO)derived compounds including glyoxal lysine dimer, N7-(carboxymethyl)arginine, and CML; methylglyoxal-derived, including MG-H1, methylglyoxal lysine, argpyrimidine, and CEL; 3-deoxyglucosone-derived, including pyrraline, pentosidine, and deoxyglucosone lysine dimer; and derivatives of glucose, fructose, and glyceraldehyde that form DNA adducts or cross-link with lysine or arginine altering protein structure and function[102]. These non-enzymatic protein modifications elevate oxidative stress and inflammation by binding with cell surface receptors such as RAGE. Exogenous AGEs play a role in the progression of cancer in addition to endogenous AGEs[29,103,104]. The metabolism and pathogenic effects of endogenous and exogenous AGEs have recently been the subject of extensive reviews[105].

RAGE AND CANCER

RAGE, inflammation, and oxidative stress

Interactions between RAGE and its ligands in T2DM result in various cellular responses, including activation of signaling pathways that cause oxidative stress and inflammation, which in turn cause various pathophysiological effects such as apoptosis, autophagy[106], senescence, and osteogenic differentiation[107], remodeling processes of the extracellular matrix, and activation of fibroblasts significant in vascular, neuronal [108], and musculoskeletal processes [109]. AGEs in T2DM accumulate in the extracellular matrix, forming cross-links with type I collagen and allowing long-lasting activation of RAGE. This also initiates a complex signaling network that allows the formation of ROS, activates the signaling pathway through ERK1/2 which then phosphorylates and activates NF-kB, and directly induces inflammation. Another alternative signaling pathway to the AGE-RAGE/ERK1-2/PKC pathway involves Rap-1, which induces inflammation, remodeling of the extracellular matrix, and oxidative stress[110].

RAGE and hypoxia

Hypoxia is frequent in solid malignant neoplasms due to the high proliferation of neoplastic cells, which does not allow rapid vascularization of neoplastic tissue so that the oxygen demand exceeds the supply. Another factor is the formation of new blood vessels that do not have the integrity of their vascular wall; a continuous outflow of blood results in tissue oxygenation deficiency [11]. Under these conditions, a series of genes regulated by HIF-1 α are activated, allowing survival through the expression of genes that promote angiogenesis, metabolic reprogramming, lipid accumulation[112], inhibition of apoptosis, invasion, and metastasis. HIF-1 α also promotes inflammation via NF-kB signaling in hypoxic

environments. In this tumor niche with inflammation, hypoxia, and cell death, DAMPs activate the NF-kB pathway mediated by RAGE, thereby amplifying HIF-1α activity[113]. In this hypoxic setting, stromal cells are also affected by RAGE; this is the case in adipocytes, where the AGE/RAGE/NF-kB pathway is activated and prolongs the inflammatory and hypoxic processes. Other effects of hypoxia include the stimulation of cell adhesion mediated by MCP-1, chemotaxis, and the polarization of macrophages towards a proinflammatory phenotype, specifically through the RAGE/NF-kB pathway in tryptophan hydroxylase 1 monocytes[114].

RAGE, survival, and programmed cell death

Cell death is a physiological process that keeps tissues healthy by systematically removing damaged cells to prevent an immune response. Although necrosis is a kind of cell death, it is pathological and only happens when there has been a significant tissue injury coupled with an immune response. Non-pathological cell death can take many forms, including apoptosis, necroptosis, and autophagy[115]. RAGE is involved in all three death pathways and can be activated by AGEs, HMGB1, and S100. In normal tissues, both the intrinsic and extrinsic apoptosis pathways are activated[116], and ROS, NF-kB, and MAPK mediate the stimulation. High levels of ROS induce the apoptosis pathway, but if they are low, autophagy is activated. Reduced HMGB1 activates Beclin-1-mediated autophagy pathways, but if oxidized, it activates apoptosis[117].

RAGE promotes cancer cell autophagy, which eventually permits survival by utilizing nutrients through the catabolism of their cellular components in a blood-free environment with no access to external nutrients and hypoxia. RAGE-dependent signaling pathways that promote autophagy involve PI3K, NF-kB/Beclin-1, PKC, and/or RAF/p38-MAPK/ERK[118]. Likewise, in cancer cells, apoptosis is inhibited, which indirectly allows cell perpetuation and survival. The pathways that inhibit apoptosis start with the binding of HMBG1/RAGE, which induces the formation of ROS and activation of NF-kB; another pathway involves Akt and matrix metalloproteinase-9[119].

RAGE and senescent cells

Cell senescence is present in T2DM and cancer. Frequently occurring in tissues undergoing metabolic shock, chronic inflammation, or oxidative stress, cell senescence is a physiological response that aims to prevent genomic instability and the consequent DNA damage that leads to metabolic reprogramming. In addition, senescence relates to decreasing immune surveillance, thus facilitating cancer initiation and progression[109,119-121]. The same markers found in the carcinogenesis process discussed above, such as IGF, HIF-1 α , AGEs, and RAGE, were discovered in a proteomics study looking for plasma proteins that indicate a senescence-related decline in health[122]. In a model of endothelial senescence induced by protein products of advanced oxidation, the presence of modified p53 at amino acid K386 by SUMOylation was associated with evasion of apoptosis[123].

RAGE ligands

RAGE aids in the removal of endotoxins and debris from apoptotic bodies during the processes of oxidative stress, hypoxia, and inflammation. Cellular damage occurs that causes the release of intracellular molecules that, outside the cell, behave as alarmins, specifically the S100 and HMBG1 proteins, also known as DAMPs, which act as endogenous RAGE ligands[124]. These proteins are also known as "moonlighting proteins" since they have various functions depending on their location. For example, when the HMGB1 protein locates inside the nucleus, it organizes chromatin[125]. In contrast, S100 is a protein that functions as a Ca²⁺ sensor[126], and like HMGB1, when located extracellularly, it functions as an alarmin. Tumor initiation and progression, as well as tissue damage, are significantly influenced by endogenous DAMP/ RAGE ligand signaling. Numerous malignancies, including colorectal[127], hepatocarcinoma[128], pancreatic[129], breast, and endometrial cancers, overexpress HMGB1 and S100[35].

The primary ligands that bind to RAGE in cancer cells, such as AGEs, HMGB1, and S100, activate several signaling pathways such as PI3K/Akt, ERK 1/2, JAK/STAT, Ras/MAPK, Rac/cdc42, p14/p42, p38, and SAP/JNK/MAPK, and transcription factors such as NF-kB, STAT3, HIF-1α, AP-1, and CREB[118,130], and thus activate a series of genes whose functions are essential in the initiation, promotion, and extension of various malignant neoplasms. These functions are known as "cancer hallmarks" and include cell proliferation, inhibition of apoptosis, inhibition of tumor suppressor genes, evasion of immunity, increased survival, invasion, metastasis, angiogenesis, genomic instability due to failure to repair mutations, and metabolic dysregulation[35,124] (Table 1).

RAGE and tumor microenvironment

Tumorigenesis is the process by which healthy cells develop the capacity to become cancerous cells, which implies, in addition to genetic and epigenetic alterations in DNA, the formation of the tumor microenvironment. The tumor microenvironment is determined by the interaction between resident immune cells, mesenchymal stromal cells, and tumor cells, the paracrine signaling between them, and the anatomical niche built-up by the extracellular matrix and blood vessels. In addition to cancer-affected fibroblasts, the tumor microenvironment contains infiltrating tumor-associated macrophages that promote tumor survival[131,132]. The tumor microenvironment includes the extracellular matrix, blood vascular structures, and paracrine signaling between stromal cells and tumor cells (Figure 2).

Recent studies have revealed that the involved cells and specialized three-dimensional structures are unique to each tumor by tissue[133-135]. Table 1 outlines the traits of the tumor microenvironment in hepatocarcinoma, colorectal, breast, and pancreatic cancers with RAGE implications. These findings demonstrate that RAGE promotes different adaptive phenomena for the survival, initiation, and progression of malignant tumors. Nevertheless, it is necessary to mention that RAGE overexpression varies in cancer related to T2DM because of the cellular heterogeneity of the neoplastic process. The Human Protein Atlas database shows RAGE detection rates in malignant cells by immunohisto-

Table 1 Studies published between 2018 and 2022 on receptor for advanced glycation products-ligands, related activated pathways, and cancer hallmarks in the most frequent neoplasms found in diabetic patients Ligands and signaling Molecule TS/AM/CL Neoplasia Cancer hallmarks Ref. pathway expressed AGE/RAGE, ERK1/2; Akt, c-IL-8/CXCR1/2 CL; CAFs, TNBC (MDA-Santolla et al[137] Breast cancer Migration and invasion MB-231 cells) fos HMGA1 CL; TNBC (MDA-MB-231 Cell proliferation, Shah et al[138] metastasis, and EMT and Hs578) HMGB1/RAGE Motility, migration, TS; human breast cancer, Chen et al[139] invasion, and dysregu-AM; NOD/SCID mice, lation of metabolism CL; human breast cancer cells (MCF-7, T-470, BT474, MDA-MB-231, ZR-75-30, BT549) and human fibroblast cells HFL1 HMGB1/PI3K/Akt PD-L1 Cell proliferation, CL; human breast cancer Amornsupak et al cells (MDA-MB-231 P, migration, invasion, and [140]T-cell apoptosis MDA-MB-231 BM) HMGB1, PI3K/Akt, mTOR HIF-1α, VEGF Migration and TS: human breast cancer He *et al*[141] angiogenesis CL; human breast cancer cells MCF-7 HMGB1/RAGE Downregulation of Cell growth, invasion, and TS; human breast cancer Wang et al[142] miR-205 EMT CL; TNBC (MDA-MB-231, MDA-MB-453, MDA-MB-468) and NTNBC (MCF-7, MCF-10F) HMGB1/RAGE, ERK 1/2, Bone metastasis and AM; 4T1 mice CL; mouse Okui et al[143] breast cancer 4T1, primary CREB neurite outgrowth of nervous system cells rat nervous system cells DRG, rat DRG/mouse neuroblastoma hybrid cells F11, immortalized rat DRG neuronal cells 50B11 S100A14/RAGE, NF-kB CCL2/CXCL5 Migration, invasion, and TS; human breast cancer Li et al[144] lung metastasis and paired adjacent breast normal, metastatic lymph node, and non-metastatic lymph node AM; BALB/c, BALB/c, SCID beige, C57BL/6J, CMV-CreC57BL/6J, S200-/- and S100A14-/- PyMT mice CL: human breast cancer cells MCF7, MCF10A, T47D, SKBR3, BT549, MDA-MB-231, MCF10AT, MCFCA1h, MCFCA1a and mouse breast cancer cells 4T1 S100A7/RAGE, PI3K/Akt, IGF-1 Angiogenesis CL; human breast cancer Muoio et al[145] ERK1/2, STAT3 cells MCF-7, T47D, and HUVECs cells S100A7/RAGE, cPLA PGE2, CD163+ AM; NOD SCID gamma Immunosuppression, M2-Mishra et al[146] macrophages, CD4+, mice CL; human breast CD8+, and T cells cancer cells MDA-MB-231, MDA-MB-468 and mouse mammary cancer cells MVT-1 S100A8/A9-RAGE, FAK, Akt, FLNA, CTGF, Cyr61 Cell proliferation and CL; HEK293T and TNBC Rigiracciolo et al Hippo-YAK migration (MDA-MB-23 and BT-549) [147]

LPS/S100A7/TLR4/RAGE

WJD | https://www.wjgnet.com

984

Migration and invasion

Wilkie et al[148]

AM; orthotopic breast

cancer C57BL/6 mice model CL; murine mammary cancer cells EO771, MTV-1, murine metastatic mammary cells EO.2, human breast carcinoma cells SUM 159,

				MDA-MB-231 and MDA- MB-468	
	acHMGB1/RAGE, S100A4/RAGE, Gas6/AXL	CXCR4, CXCL12, CCL2, CD151 and α3 β1-integrin	Cell proliferation, invasion, intravasation, and EMT	AM; murine orthotopic mammary cancer CL; human MSCs, geminin overexpressing breast tumors Gem197, Gem240, Gem256, Gem257 and Gem270 cells, CAFs, and M0- TAMs and M2-TAMs	Ryan et al <mark>[149]</mark>
Colorectal cancer	S100A16		Cancer prognostic marker	TS; human colorectal cancer	Sun <i>et al</i> [150]
	HMGB1/RAGE	PD-1	Cancer prognostic marker	TS; human colorectal cancer CL; human colorectal cancer cells SW480, and SW620	Huang et al[<mark>151</mark>]
	S100B/RAGE, NF-kB	VEGF-A	Proliferation, migration, and angiogenesis	CL; human colon cancer cells HCT116	Zheng et al[152]
	IGF1R-Ras/RAGE-HMGB1,		Oncogenesis	TS; Human colorectal from diabetic patients	Niu <i>et al</i> [<mark>153</mark>]
	AGEs/RAGE, KLF5	MDM upregulation and RB and p53 downregulation	Cancer initiation and development	AM; diabetic mouse model and CL; human colon cancer cells HCT116	Wang <i>et al</i> [<mark>154</mark>]
	TCTP, HMGB1/RAGE, NF-kB		Invasion and metastasis	TS; human colorectal AM; tumor xenografts BALB/c nude mice CL; human colon adenocarcinoma cells LoVo	Huang et al[155]
	S100A9/RAGE/TLR4	Arg-1, iNOS, IL-10 and ROS	Immune suppression and MDSC chemotaxis	TS; human colorectal cancer and normal colon CL; Human colorectal cells LoVo, and MDSCs	Huang et al[156]
	HMGB1/RAGE, Kras/Yap1		Cell proliferation	CL; human colorectal cancer cells HCT116 and SW480	Qian et al[<mark>157</mark>]
	S100B/RAGE, p38/pAkt/mTOR	VEGF-R2, iNOS, VEGF	Cell proliferation, migration, invasion, and angiogenesis	CL; human colon adenocarcinoma cells CaCo	Seguella et al[158]
	HMGB1/RAGE, pERK1/2, pDRP1		Cell viability, autophagy, and chemoresistance	TS; human colorectal AM; athymic nude BALBC/c mice CL; human colorectal cells SW480, SW620, and LoVo	Huang et al[159]
Hepatocellular carcinoma	S100A9-TLR4/RAGE-ROS,	NET	Cell proliferation, invasion, and metastasis	TS; HBV+ and HBV- hepatocellular carcinoma AM; BALB/c mice and C57BL/6 mice CL; human liver cells QSG-7701, human hepatocellular carcinoma cells HepG2.2.15, mouse hepatocellular carcinoma cells H22 and HUVEC cells	Zhan et al[<mark>160</mark>]
	HMGB1/RAGE		Cell proliferation and tumor differentiation	TS; primary hepatocellular carcinoma	Ando et al[<mark>161</mark>]
	HMGB1/RAGE, ATG7		Cell proliferation, fibrosis, and autophagy	TS; mouse hepatocellular carcinoma AM; Atg7, RAGE, HMGB1 transgenic C57BL/6Jmouse	Khambu <i>et al</i> [128]
	HMGB1/RAGE, JNK, OCT4/TGFb1	miR-21, CD44	Migration and invasion	TS; human hepatocellular carcinoma AM; BALB/c nu/nu mice CL; human hepatocellular carcinoma cells HepG2, HCCLM3, Huh7, SMMC7721 and MHCC97H	Li et al <mark>[162]</mark>

Gaisbideng® WJD | https://www.wjgnet.com

	S100A4/RAGE, b-catenin	OCT4, SOX2, CD44 and Nanog (stem cell-associated genes)	Fibrosis and carcino- genesis	TS; human hepatocellular carcinoma AM; S100a4- EGFP, S100A4 ^{+/+GFP} , S100A4 ^{-/-} transgenic mice. CL; human hepatocellular carcinoma cells Huh7 and murine liver cancer cells Hep1-6	Li et al[163]
	HMGB1/RAGE, ERK1/2	CXCL2, IL-8, TNF, IL-6, IL-10, IL-23-p19	Macrophage activation and inflammation	AM; primary murine hepatocytes from male C57BI/6J mice, and primary murine splenocytes from male C57BI/CJ CC; murine hepatoma cells Hepa1-6 and Hep-56.1D, human hepatoma cells HepG2, RAW 264.7 macrophages and monocytic cells THP1	Bachmann <i>et al</i> [164]
	HMGB1/RAGE, NF-kB	circRNA 101368, miR-200a	Cell migration	TS; human hepatocellular carcinoma CL; human hepatocellular carcinoma cells HCCLM3, MHCC97L, SMMC7721, Hep3B, HepG2 cells, and normal hepatocyte cells THLE-3	Li et al <mark>[165]</mark>
Pancreatic cancer	RAGE	NET	Neutrophil autophagy	TS; human pancreatic carcinoma AM; Wild type C57BL6 mice and RAGE ^{-/-} C57BL6 mice, orthotopic pancreatic cancer model, CL; murine pancreatic cancer cells Panc02, MDSCs cells	Boone <i>et al</i> [166]
	RAGE, PI3K/AKT/mTOR		Cell viability	CL; human pancreatic cancer cells MIA Paca-2, BxPC-3, AsPC-1, HPAC, PANC-1, MIA Paca ^{GEMR}	Lan et al <mark>[167]</mark>
	RAGE, ERK1/2/Akt	Alpha 2 and alpha 1 integrin downregu- lation	Cell proliferation, invasion, and migration	CL; human pancreatic cancer cells Panc-1	Swami et al[129]
	AGE/RAGE, TGFb1	a-SMA, collagen 1, IL-6	Fibrosis and EMT	TS; human pancreatic ductal adenocarcinoma AM; WT-C57BL/6 and RG-C57BL/6 nice CL; primary PSC, human pancreatic ductal adenocarcinoma cells BxPC-3 and AsPC-1	Uchida et al[<mark>168</mark>]
	HMGB1/RAGE, PI3K/Akt	Atg5, Beclin-1, LC3-II	Autophagia and apoptosis inhibition	CL; human pancreatic cancer cells MIA Paca-2 and MIA Paca ^{GEMR}	Chen <i>et al</i> [169]

AGE: Advanced glycation end products; Akt: Protein kinase B; AM: Animal model; Arg-1: Arginase-1; ATG7: Autophagy related 7; CAFs: Cancerassociated fibroblasts; CCL2: CC-chemokine ligand 2; circRNA: Circular RNA; CL: Cell line; cPLA: Cytosolic phospholipase A2; CREB: cAMP response element-binding protein; CTGF: Connective tissue growth factor; CXCL: CXC motif chemokine ligand; CXCR: C-X-C Chemokine receptor; Cyr61: Cysteinerich angiogenic 61; DRG: Dorsal rat ganglion; pDRP1: Phosphorylated dynamin-related protein 1; EMT: Epithelial-mesenchymal transition; ERK 1/2: Extracellular signal regulated kinase 1/2; FAK: Focal adhesion kinase; FLNA: Filamin A alpha; HIF-1a: Hypoxia-inducible factor-1 alpha; HBV: Hepatitis B virus; Gas6: Growth arrest-specific gene 6; HMG: High mobility group; acHMGB1: Acetylated high mobility group B1; HUVECs: Human umbilical vein endothelial cells; IGF-1: Insulin-like growth factor-1; JNK: Jun N-terminal kinase; KLF5: Kruppel-like factor 5; LPS: Lipopolysaccharide; MDA-MB-231 P: MDA-MB-231 Parental cells; MDA-MB-231 BM: MDA-MB-231 bone marrow; MDM: Mouse double minute 2 homolog; MDSCs: Myeloid-derived suppressor cells; MIA Paca^{GEMR}: MIA Paca gemcitabine resistant; MSCs: Mesenchymal stem cells; mTOR: Mammalian target of rapamycin; NET: Neutrophil extracellular traps; NF-kB: Nuclear factor-kappa B; iNOS: Inducible nitric oxide synthase; NTNBC: Non-triple-negative breast cancer; OCT-4: Octamer-binding transcription factor 4; PI3K: Phosphoinositide 3-kinase; PD-L1: Programmed death ligand 1; PGE2: Prostaglandin E2; PSC: Pancreatic stellate cell; Ras: Rat sarcoma virus; RB: Retinoblastoma; RAGE: Receptor for advanced glycation end products; S100: Soluble 100% protein; a-SMA: Alphasmooth muscle actin; SOX2: SRY (sex determining region Y)-box 2; STAT3: Signal transducer and activator of transcription 3; TAMs: Tumor-associated macrophages; TCTP: Translationally controlled tumor protein; TGFb1: Tumor growth factor beta 1; THP1: Human monocytic cell line derived from acute monocytic leukemia; TLR: Toll-like receptor; TNBC: Triple-negative breast cancer; TNF: Tumor necrosis factor; TS: Tissue sample; VEGF: Vascular endothelial growth factor; Yap1: Yes associated protein 1.

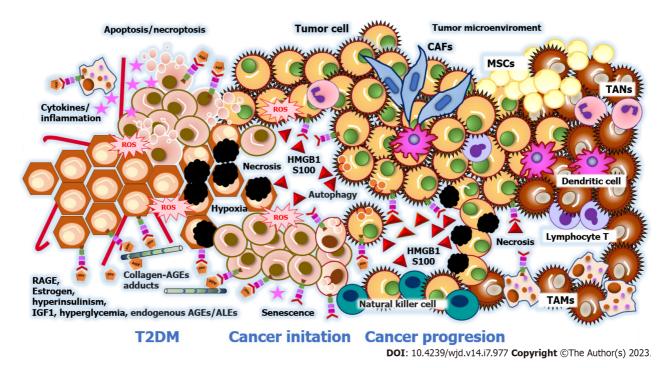


Figure 2 Tumor microenvironment in type 2 diabetes mellitus. In type 2 diabetes mellitus patients, elevated estrogen levels, hyperinsulinemia, insulin-like growth factor-1 levels, hyperglycemia, endogenous advanced glycosylation end products (AGEs), and advanced lipoperoxidation end products (ALEs) promote cancer initiation and progression in the tumor microenvironment (TME). Receptor for advanced glycation products (RAGE) plays an essential role in the TME by promoting inflammation, oxidative stress, endotoxin clearance, senescence, and programmed cell death by binding to endogenous AGE/ALE ligands and damageassociated molecular patterns, primarily the high mobility group box 1 proteins and S100 proteins. To overcome a hypoxic and acidic microenvironment, tumor cells coordinate a metabolic program (Warburg effect), cell survival (senescence and cell death program), angiogenesis, extracellular matrix remodeling, proliferation, invasion, and metastasis. Tumor cells interact with resident immune cells and recruit mesenchymal stromal cells, cancer-associated fibroblasts, tumor-associated macrophages, tumor-associated neutrophils. RAGE: Receptor for advanced glycation products; ROS: Reactive oxygen species; T2DM: Type 2 diabetes mellitus patients; IGF-1: Insulin-like growth factor-1; AGEs: Advanced glycosylation end products; ALEs: Advanced lipoperoxidation end products; HMGB1: High mobility group box 1 proteins; MSCs: Mesenchymal stromal cells; CAFs: Cancer-associated fibroblasts; TAMs: Tumor-associated macrophages; TANs: Tumor-associated neutrophils.

chemistry as follows: Hepatocarcinoma at 50%; pancreatic cancer at 33.3%; breast cancer at 25%; endometrial cancer at 16.6%; and colorectal cancer at 8.3% [136].

CONCLUSION

RAGE is an environmental sensor with complex and multiple functions involved in every stage along the pathophysiological pathways that lead to the progression of obesity, T2DM, and cancer. Therefore, it is crucial to analyze each of the processes that RAGE is involved in, as the assimilation of this information could help in developing more accurate diagnostic and treatment approaches. For instance, this review has highlighted how RAGE acts from the earliest stages of the initiation and development of obesity, T2DM, and cancer. Recognizing all participating RAGE isoforms in their tissue and cellular locations could predict the progression points and provide diagnostic markers. In this manner, we would also be able to distinguish between a patient who is obese, has a low grade of inflammation, and is on the frontline of developing T2DM or most likely to respond to nutritional intervention.

On the other hand, RAGE participates in the initiation of neoplastic processes. Since its presence indicates cellular senescence and the presence of cancer cells with more aggressive activity, it is not surprising related to a poor prognosis and has potential as a cancer biomarker to help predict patient outcomes. Since RAGE participates even in the first stages, it has potential as a preventive and immunomodulator for therapeutic purposes to reduce morbidity and mortality associated with the development of obesity, T2DM, and cancer. Inhibitors of RAGE may be helpful in the treatment of obesity and diabetes mellitus. Studies have shown that RAGE is overexpressed in AT. Obesity is well known to contribute to inflammation and insulin resistance, which are hallmarks of obesity and diabetes. RAGE inhibitors could reduce inflammation and improve insulin sensitivity in obesity and T2DM; however, the majority of RAGE inhibitor studies have focused on cancer treatment. Some RAGE inhibitors under study are cromolyn, RAP, RAGE peptide antagonist, and gefitinib. While there are currently no RAGE-specific therapies approved for use in humans, there are pre-clinical studies investigating the potential of RAGE inhibitors as a treatment for various diseases. We review herein the topically relevant literature, delimiting by process, organ, and tissue to provide a progressive and systemic overview. It should be read and generalized with caution, as there are still many gaps in the knowledge about RAGE since most studies are experimental-based (in mice) and cross-sectional studies (in humans).

ACKNOWLEDGEMENTS

The authors acknowledge Ruelas-Cinco EC for providing some photographic images of RAGE immunocytochemistry in peripheral blood mononuclear cells from her thesis.

FOOTNOTES

Author contributions: Garza-Campos A and Prieto-Correa JR contributed to the writing, reviewing, and editing of the manuscript; Prieto-Correa JR and Domínguez-Rosales JA prepared the table; Garza-Campos A and Hernández-Nazará ZH prepared the figures; Domí nguez-Rosales JA contributed to the writing and performed the majority of the reviewing and editing of the manuscript; Hernández-Nazará ZH and Domínguez-Rosales JA conceptualized the study and designed the outline for the paper; Hernández-Nazará ZH wrote the first draft; and all authors read and approved the final manuscript.

Supported by the Founding Proyectos de Impulso a la Investigación to Hernandez-Nazara ZH from Universidad de Guadalajara, Mexico, No. PIN 2020-I.

Conflict-of-interest statement: All the authors report no relevant conflicts of interest for this article.

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: https://creativecommons.org/Licenses/by-nc/4.0/

Country/Territory of origin: Mexico

ORCID number: Andrea Garza-Campos 0000-0003-0413-7090; José Roberto Prieto-Correa 0000-0003-4580-0231; José Alfredo Domínguez-Rosales 0000-0002-8560-5855; Zamira Helena Hernández-Nazará 0000-0003-2319-8470.

S-Editor: Wang JJ L-Editor: Wang TQ P-Editor: Zhao S

REFERENCES

- 1 World Health Organization. Noncommunicable diseases. [cited 5 December 2022]. Available from: https://www.who.int/health-topics/ noncommunicable-diseases#tab=tab 1
- Sun H, Saeedi P, Karuranga S, Pinkepank M, Ogurtsova K, Duncan BB, Stein C, Basit A, Chan JCN, Mbanya JC, Pavkov ME, 2 Ramachandaran A, Wild SH, James S, Herman WH, Zhang P, Bommer C, Kuo S, Boyko EJ, Magliano DJ. IDF Diabetes Atlas: Global, regional and country-level diabetes prevalence estimates for 2021 and projections for 2045. Diabetes Res Clin Pract 2022; 183: 109119 [PMID: 34879977 DOI: 10.1016/j.diabres.2021.109119]
- Ugai T, Sasamoto N, Lee HY, Ando M, Song M, Tamimi RM, Kawachi I, Campbell PT, Giovannucci EL, Weiderpass E, Rebbeck TR, Ogino 3 S. Is early-onset cancer an emerging global epidemic? Current evidence and future implications. Nat Rev Clin Oncol 2022; 19: 656-673 [PMID: 36068272 DOI: 10.1038/s41571-022-00672-8]
- Renchan AG, Tyson M, Egger M, Heller RF, Zwahlen M. Body-mass index and incidence of cancer: a systematic review and meta-analysis of 4 prospective observational studies. Lancet 2008; 371: 569-578 [PMID: 18280327 DOI: 10.1016/S0140-6736(08)60269-X]
- Huxley R, Ansary-Moghaddam A, Berrington de González A, Barzi F, Woodward M. Type-II diabetes and pancreatic cancer: a meta-analysis 5 of 36 studies. Br J Cancer 2005; 92: 2076-2083 [PMID: 15886696 DOI: 10.1038/sj.bjc.6602619]
- Lane MM, Davis JA, Beattie S, Gómez-Donoso C, Loughman A, O'Neil A, Jacka F, Berk M, Page R, Marx W, Rocks T. Ultraprocessed food 6 and chronic noncommunicable diseases: A systematic review and meta-analysis of 43 observational studies. Obes Rev 2021; 22: e13146 [PMID: 33167080 DOI: 10.1111/obr.13146]
- 7 Roberts DL, Dive C, Renehan AG. Biological mechanisms linking obesity and cancer risk: new perspectives. Annu Rev Med 2010; 61: 301-316 [PMID: 19824817 DOI: 10.1146/annurev.med.080708.082713]
- van Greevenbroek MM, Schalkwijk CG, Stehouwer CD. Obesity-associated low-grade inflammation in type 2 diabetes mellitus: causes and 8 consequences. Neth J Med 2013; 71: 174-187 [PMID: 23723111]
- Chuah YK, Basir R, Talib H, Tie TH, Nordin N. Receptor for advanced glycation end products and its involvement in inflammatory diseases. 9 Int J Inflam 2013; 2013: 403460 [PMID: 24102034 DOI: 10.1155/2013/403460]
- Bierhaus A, Nawroth PP. Multiple levels of regulation determine the role of the receptor for AGE (RAGE) as common soil in inflammation, 10 immune responses and diabetes mellitus and its complications. Diabetologia 2009; 52: 2251-2263 [PMID: 19636529 DOI: 10.1007/s00125-009-1458-9]
- Hudson BI, Carter AM, Harja E, Kalea AZ, Arriero M, Yang H, Grant PJ, Schmidt AM. Identification, classification, and expression of RAGE 11 gene splice variants. FASEB J 2008; 22: 1572-1580 [PMID: 18089847 DOI: 10.1096/fj.07-9909com]
- Schmidt AM, Yan SD, Yan SF, Stern DM. The biology of the receptor for advanced glycation end products and its ligands. Biochim Biophys 12 Acta 2000; 1498: 99-111 [PMID: 11108954 DOI: 10.1016/s0167-4889(00)00087-2]
- Clynes R, Moser B, Yan SF, Ramasamy R, Herold K, Schmidt AM. Receptor for AGE (RAGE): weaving tangled webs within the 13 inflammatory response. Curr Mol Med 2007; 7: 743-751 [PMID: 18331232 DOI: 10.2174/156652407783220714]



- Feng Z, Zhu L, Wu J. RAGE signalling in obesity and diabetes: focus on the adipose tissue macrophage. Adipocyte 2020; 9: 563-566 [PMID: 14 32892690 DOI: 10.1080/21623945.2020.1817278]
- Arivazhagan L, Popp CJ, Ruiz HH, Wilson RA, Manigrasso MB, Shekhtman A, Ramasamy R, Sevick MA, Schmidt AM. The RAGE/ 15 DIAPH1 axis: mediator of obesity and proposed biomarker of human cardiometabolic disease. Cardiovasc Res 2022 [PMID: 36448548 DOI: 10.1093/cvr/cvac175]
- Bettiga A, Fiorio F, Di Marco F, Trevisani F, Romani A, Porrini E, Salonia A, Montorsi F, Vago R. The Modern Western Diet Rich in 16 Advanced Glycation End-Products (AGEs): An Overview of Its Impact on Obesity and Early Progression of Renal Pathology. Nutrients 2019; 11 [PMID: 31366015 DOI: 10.3390/nu11081748]
- Arivazhagan L, López-Díez R, Shekhtman A, Ramasamy R, Schmidt AM. Glycation and a Spark of ALEs (Advanced Lipoxidation End 17 Products) - Igniting RAGE/Diaphanous-1 and Cardiometabolic Disease. Front Cardiovasc Med 2022; 9: 937071 [PMID: 35811725 DOI: 10.3389/fcvm.2022.937071
- Nemet I, Varga-Defterdarović L, Turk Z. Methylglyoxal in food and living organisms. Mol Nutr Food Res 2006; 50: 1105-1117 [PMID: 18 17103372 DOI: 10.1002/mnfr.200600065]
- Poulsen MW, Hedegaard RV, Andersen JM, de Courten B, Bügel S, Nielsen J, Skibsted LH, Dragsted LO. Advanced glycation endproducts in 19 food and their effects on health. Food Chem Toxicol 2013; 60: 10-37 [PMID: 23867544 DOI: 10.1016/j.fet.2013.06.052]
- Scheijen JLJM, Clevers E, Engelen L, Dagnelie PC, Brouns F, Stehouwer CDA, Schalkwijk CG. Analysis of advanced glycation endproducts 20 in selected food items by ultra-performance liquid chromatography tandem mass spectrometry: Presentation of a dietary AGE database. Food Chem 2016; 190: 1145-1150 [PMID: 26213088 DOI: 10.1016/j.foodchem.2015.06.049]
- Robinson AB, Stogsdill JA, Lewis JB, Wood TT, Reynolds PR. RAGE and tobacco smoke: insights into modeling chronic obstructive 21 pulmonary disease. Front Physiol 2012; 3: 301 [PMID: 22934052 DOI: 10.3389/fphys.2012.00301]
- Prasad K, Dhar I, Caspar-Bell G. Role of Advanced Glycation End Products and Its Receptors in the Pathogenesis of Cigarette Smoke-22 Induced Cardiovascular Disease. Int J Angiol 2015; 24: 75-80 [PMID: 26060376 DOI: 10.1055/s-0034-1396413]
- Kwon OS, Decker ST, Zhao J, Hoidal JR, Heuckstadt T, Sanders KA, Richardson RS, Layec G. The receptor for advanced glycation end 23 products (RAGE) is involved in mitochondrial function and cigarette smoke-induced oxidative stress. Free Radic Biol Med 2023; 195: 261-269 [PMID: 36586455 DOI: 10.1016/j.freeradbiomed.2022.12.089]
- Chapman S, Mick M, Hall P, Mejia C, Sue S, Abdul Wase B, Nguyen MA, Whisenant EC, Wilcox SH, Winden D, Reynolds PR, Arroyo JA. 24 Cigarette smoke extract induces oral squamous cell carcinoma cell invasion in a receptor for advanced glycation end-products-dependent manner. Eur J Oral Sci 2018; 126: 33-40 [PMID: 29226456 DOI: 10.1111/eos.12395]
- Li Y, Qin M, Zhong W, Liu C, Deng G, Yang M, Li J, Ye H, Shi H, Wu C, Lin H, Chen Y, Huang S, Zhou C, Lv Z, Gao L. RAGE promotes 25 dysregulation of iron and lipid metabolism in alcoholic liver disease. Redox Biol 2023; 59: 102559 [PMID: 36502724 DOI: 10.1016/j.redox.2022.102559]
- 26 Birukov A, Cuadrat R, Polemiti E, Eichelmann F, Schulze MB. Advanced glycation end-products, measured as skin autofluorescence, associate with vascular stiffness in diabetic, pre-diabetic and normoglycemic individuals: a cross-sectional study. Cardiovasc Diabetol 2021; **20**: 110 [PMID: 34176469 DOI: 10.1186/s12933-021-01296-5]
- Fujiwara Y, Kiyota N, Tsurushima K, Yoshitomi M, Mera K, Sakashita N, Takeya M, Ikeda T, Araki T, Nohara T, Nagai R. Natural 27 compounds containing a catechol group enhance the formation of NE-(carboxymethyl)lysine of the Maillard reaction. Free Radic Biol Med 2011; 50: 883-891 [PMID: 21195168 DOI: 10.1016/j.freeradbiomed.2010.12.033]
- Anderson MM, Requena JR, Crowley JR, Thorpe SR, Heinecke JW. The myeloperoxidase system of human phagocytes generates Nepsilon-28 (carboxymethyl)lysine on proteins: a mechanism for producing advanced glycation end products at sites of inflammation. J Clin Invest 1999; **104**: 103-113 [PMID: 10393704 DOI: 10.1172/JCI3042]
- 29 Twarda-Clapa A, Olczak A, Białkowska AM, Koziołkiewicz M. Advanced Glycation End-Products (AGEs): Formation, Chemistry, Classification, Receptors, and Diseases Related to AGEs. Cells 2022; 11 [PMID: 35455991 DOI: 10.3390/cells11081312]
- Ryder E, Pedreañez A, Vargas R, Peña C, Fernandez E, Diez-Ewald M, Mosquera J. Increased proinflammatory markers and lipoperoxidation 30 in obese individuals: Inicial inflammatory events? Diabetes Metab Syndr 2015; 9: 280-286 [PMID: 25470639 DOI: 10.1016/j.dsx.2014.04.022]
- Mishra S, Mishra BB. Study of Lipid Peroxidation, Nitric Oxide End Product, and Trace Element Status in Type 2 Diabetes Mellitus with and 31 without Complications. Int J Appl Basic Med Res 2017; 7: 88-93 [PMID: 28584737 DOI: 10.4103/2229-516X.205813]
- Jaganjac M, Tirosh O, Cohen G, Sasson S, Zarkovic N. Reactive aldehydes--second messengers of free radicals in diabetes mellitus. Free 32 Radic Res 2013; 47 Suppl 1: 39-48 [PMID: 23521622 DOI: 10.3109/10715762.2013.789136]
- Iacobini C, Menini S, Ricci C, Scipioni A, Sansoni V, Mazzitelli G, Cordone S, Pesce C, Pugliese F, Pricci F, Pugliese G. Advanced 33 lipoxidation end-products mediate lipid-induced glomerular injury: role of receptor-mediated mechanisms. J Pathol 2009; 218: 360-369 [PMID: 19334049 DOI: 10.1002/path.2536]
- Shanmugam N, Figarola JL, Li Y, Swiderski PM, Rahbar S, Natarajan R. Proinflammatory effects of advanced lipoxidation end products in 34 monocytes. Diabetes 2008; 57: 879-888 [PMID: 18003754 DOI: 10.2337/db07-1204]
- Palanissami G, Paul SFD. RAGE and Its Ligands: Molecular Interplay Between Glycation, Inflammation, and Hallmarks of Cancer-a Review. 35 Horm Cancer 2018; 9: 295-325 [PMID: 29987748 DOI: 10.1007/s12672-018-0342-9]
- van Dongen KCW, Kappetein L, Miro Estruch I, Belzer C, Beekmann K, Rietjens IMCM. Differences in kinetics and dynamics of 36 endogenous versus exogenous advanced glycation end products (AGEs) and their precursors. Food Chem Toxicol 2022; 164: 112987 [PMID: 35398182 DOI: 10.1016/j.fct.2022.112987]
- 37 Lyte JM, Gabler NK, Hollis JH. Postprandial serum endotoxin in healthy humans is modulated by dietary fat in a randomized, controlled, cross-over study. Lipids Health Dis 2016; 15: 186 [PMID: 27816052 DOI: 10.1186/s12944-016-0357-6]
- Li Y, Peng Y, Shen Y, Zhang Y, Liu L, Yang X. Dietary polyphenols: regulate the advanced glycation end products-RAGE axis and the 38 microbiota-gut-brain axis to prevent neurodegenerative diseases. Crit Rev Food Sci Nutr 2022; 1-27 [PMID: 35587161 DOI: 10.1080/10408398.2022.2076064]
- Cani PD, Amar J, Iglesias MA, Poggi M, Knauf C, Bastelica D, Neyrinck AM, Fava F, Tuohy KM, Chabo C, Waget A, Delmée E, Cousin B, 39 Sulpice T, Chamontin B, Ferrières J, Tanti JF, Gibson GR, Casteilla L, Delzenne NM, Alessi MC, Burcelin R. Metabolic endotoxemia initiates obesity and insulin resistance. Diabetes 2007; 56: 1761-1772 [PMID: 17456850 DOI: 10.2337/db06-1491]
- Wang L, Wu J, Guo X, Huang X, Huang Q. RAGE Plays a Role in LPS-Induced NF-KB Activation and Endothelial Hyperpermeability. 40 Sensors (Basel) 2017; 17 [PMID: 28358333 DOI: 10.3390/s17040722]



- Fritz G. RAGE: a single receptor fits multiple ligands. Trends Biochem Sci 2011; 36: 625-632 [PMID: 2201901] DOI: 41 10.1016/j.tibs.2011.08.008]
- Wang X, Wang Y, Antony V, Sun H, Liang G. Metabolism-Associated Molecular Patterns (MAMPs). Trends Endocrinol Metab 2020; 31: 42 712-724 [PMID: 32807598 DOI: 10.1016/j.tem.2020.07.001]
- Turki Jalil A, Alameri AA, Iqbal Doewes R, El-Schrawy AA, Ahmad I, Ramaiah P, Kadhim MM, Kzar HH, Sivaraman R, Romero-Parra RM, 43 Ansari MJ, Fakri Mustafa Y. Circulating and dietary advanced glycation end products and obesity in an adult population: A paradox of their detrimental effects in obesity. Front Endocrinol (Lausanne) 2022; 13: 966590 [PMID: 36531466 DOI: 10.3389/fendo.2022.966590]
- Ruiz HH, Ramasamy R, Schmidt AM. Advanced Glycation End Products: Building on the Concept of the "Common Soil" in Metabolic 44 Disease. Endocrinology 2020; 161 [PMID: 31638645 DOI: 10.1210/endocr/bqz006]
- Gaens KH, Goossens GH, Niessen PM, van Greevenbroek MM, van der Kallen CJ, Niessen HW, Rensen SS, Buurman WA, Greve JW, Blaak 45 EE, van Zandvoort MA, Bierhaus A, Stehouwer CD, Schalkwijk CG. Nɛ-(carboxymethyl)lysine-receptor for advanced glycation end product axis is a key modulator of obesity-induced dysregulation of adipokine expression and insulin resistance. Arterioscler Thromb Vasc Biol 2014; **34**: 1199-1208 [PMID: 24723555 DOI: 10.1161/ATVBAHA.113.302281]
- Sebeková K, Krivošíková Z, Gajdoš M. Total plasma Nɛ-(carboxymethyl)lysine and sRAGE levels are inversely associated with a number of 46 metabolic syndrome risk factors in non-diabetic young-to-middle-aged medication-free subjects. Clin Chem Lab Med 2014; 52: 139-149 [PMID: 23509221 DOI: 10.1515/cclm-2012-0879]
- Dozio E, Vianello E, Briganti S, Lamont J, Tacchini L, Schmitz G, Corsi Romanelli MM. Expression of the Receptor for Advanced Glycation 47 End Products in Epicardial Fat: Link with Tissue Thickness and Local Insulin Resistance in Coronary Artery Disease. J Diabetes Res 2016; 2016: 2327341 [PMID: 26788516 DOI: 10.1155/2016/2327341]
- Santangelo C, Filardi T, Perrone G, Mariani M, Mari E, Scazzocchio B, Masella R, Brunelli R, Lenzi A, Zicari A, Morano S. Cross-talk 48 between fetal membranes and visceral adipose tissue involves HMGB1-RAGE and VIP-VPAC2 pathways in human gestational diabetes mellitus. Acta Diabetol 2019; 56: 681-689 [PMID: 30820673 DOI: 10.1007/s00592-019-01304-x]
- Ruiz HH, Nguyen A, Wang C, He L, Li H, Hallowell P, McNamara C, Schmidt AM. AGE/RAGE/DIAPH1 axis is associated with 49 immunometabolic markers and risk of insulin resistance in subcutaneous but not omental adipose tissue in human obesity. Int J Obes (Lond) 2021; 45: 2083-2094 [PMID: 34103691 DOI: 10.1038/s41366-021-00878-3]
- Du Z, Wu J, Feng Z, Ma X, Zhang T, Shu X, Xu J, Wang L, Luo M. RAGE displays sex-specific differences in obesity-induced adipose tissue 50 insulin resistance. Biol Sex Differ 2022; 13: 65 [PMID: 36348465 DOI: 10.1186/s13293-022-00476-6]
- Song F, Hurtado del Pozo C, Rosario R, Zou YS, Ananthakrishnan R, Xu X, Patel PR, Benoit VM, Yan SF, Li H, Friedman RA, Kim JK, 51 Ramasamy R, Ferrante AW Jr, Schmidt AM. RAGE regulates the metabolic and inflammatory response to high-fat feeding in mice. Diabetes 2014; 63: 1948-1965 [PMID: 24520121 DOI: 10.2337/db13-1636]
- Ding YS, Malik N, Mendoza S, Tuchman D, Del Pozo CH, Diez RL, Schmidt AM. PET imaging study of brown adipose tissue (BAT) activity 52 in mice devoid of receptor for advanced glycation end products (RAGE). J Biosci 2019; 44 [PMID: 31502571]
- da Silva Rosa SC, Nayak N, Caymo AM, Gordon JW. Mechanisms of muscle insulin resistance and the cross-talk with liver and adipose 53 tissue. Physiol Rep 2020; 8: e14607 [PMID: 33038072 DOI: 10.14814/phy2.14607]
- Li M, Chi X, Wang Y, Setrerrahmane S, Xie W, Xu H. Trends in insulin resistance: insights into mechanisms and therapeutic strategy. Signal 54 Transduct Target Ther 2022; 7: 216 [PMID: 35794109 DOI: 10.1038/s41392-022-01073-0]
- 55 Priken K, Tapia G, Cadagan C, Quezada N, Torres J, D'Espessailles A, Pettinelli P. Higher hepatic advanced glycation end products and liver damage markers are associated with nonalcoholic steatohepatitis. Nutr Res 2022; 104: 71-81 [PMID: 35635899 DOI: 10.1016/i.nutres.2022.04.005]
- Gaens KH, Niessen PM, Rensen SS, Buurman WA, Greve JW, Driessen A, Wolfs MG, Hofker MH, Bloemen JG, Dejong CH, Stehouwer CD, 56 Schalkwijk CG. Endogenous formation of Ne-(carboxymethyl)lysine is increased in fatty livers and induces inflammatory markers in an in vitro model of hepatic steatosis. J Hepatol 2012; 56: 647-655 [PMID: 21907687 DOI: 10.1016/j.jhep.2011.07.028]
- de la Maza MP, Uribarri J, Olivares D, Hirsch S, Leiva L, Barrera G, Bunout D. Weight increase is associated with skeletal muscle 57 immunostaining for advanced glycation end products, receptor for advanced glycation end products, and oxidation injury. Rejuvenation Res 2008; 11: 1041-1048 [PMID: 19086911 DOI: 10.1089/rej.2008.0786]
- Rai AK, Jaiswal N, Maurya CK, Sharma A, Ahmad I, Ahmad S, Gupta AP, Gayen JR, Tamrakar AK. Fructose-induced AGEs-RAGE 58 signaling in skeletal muscle contributes to impairment of glucose homeostasis. J Nutr Biochem 2019; 71: 35-44 [PMID: 31272030 DOI: 10.1016/j.jnutbio.2019.05.016]
- 59 Dozio E, Vettoretti S, Lungarella G, Messa P, Corsi Romanelli MM. Sarcopenia in Chronic Kidney Disease: Focus on Advanced Glycation End Products as Mediators and Markers of Oxidative Stress. Biomedicines 2021; 9 [PMID: 33918767 DOI: 10.3390/biomedicines9040405]
- Riuzzi F, Sorci G, Sagheddu R, Chiappalupi S, Salvadori L, Donato R. RAGE in the pathophysiology of skeletal muscle. J Cachexia 60 Sarcopenia Muscle 2018; 9: 1213-1234 [PMID: 30334619 DOI: 10.1002/jcsm.12350]
- Basta G, Sironi AM, Lazzerini G, Del Turco S, Buzzigoli E, Casolaro A, Natali A, Ferrannini E, Gastaldelli A. Circulating soluble receptor for 61 advanced glycation end products is inversely associated with glycemic control and S100A12 protein. J Clin Endocrinol Metab 2006; 91: 4628-4634 [PMID: 16926247 DOI: 10.1210/jc.2005-2559]
- Miranda ER, Somal VS, Mey JT, Blackburn BK, Wang E, Farabi S, Karstoft K, Fealy CE, Kashyap S, Kirwan JP, Quinn L, Solomon TPJ, 62 Haus JM. Circulating soluble RAGE isoforms are attenuated in obese, impaired-glucose-tolerant individuals and are associated with the development of type 2 diabetes. Am J Physiol Endocrinol Metab 2017; 313: E631-E640 [PMID: 28811295 DOI: 10.1152/ajpendo.00146.2017]
- Momma H, Niu K, Kobayashi Y, Huang C, Chujo M, Otomo A, Tadaura H, Miyata T, Nagatomi R. Higher serum soluble receptor for 63 advanced glycation end product levels and lower prevalence of metabolic syndrome among Japanese adult men: a cross-sectional study. Diabetol Metab Syndr 2014; 6: 33 [PMID: 24602408 DOI: 10.1186/1758-5996-6-33]
- Zaki M, Kamal S, Kholousi S, El-Bassyouni HT, Yousef W, Reyad H, Mohamed R, Basha WA. Serum soluble receptor of advanced glycation 64 end products and risk of metabolic syndrome in Egyptian obese women. EXCLI J 2017; 16: 973-980 [PMID: 28900377 DOI: 10.17179/excli2017-275
- 65 Biswas SK, Mohtarin S, Mudi SR, Anwar T, Banu LA, Alam SM, Fariduddin M, Arslan MI. Relationship of Soluble RAGE with Insulin Resistance and Beta Cell Function during Development of Type 2 Diabetes Mellitus. J Diabetes Res 2015; 2015: 150325 [PMID: 26078977 DOI: 10.1155/2015/1503251
- Hudson BI, Dong C, Gardener H, Elkind MS, Wright CB, Goldberg R, Sacco RL, Rundek T. Serum levels of soluble receptor for advanced 66 glycation end-products and metabolic syndrome: the Northern Manhattan Study. Metabolism 2014; 63: 1125-1130 [PMID: 25012910 DOI:



10.1016/j.metabol.2014.05.011]

- 67 Huang M, Que Y, Shen X. Correlation of the plasma levels of soluble RAGE and endogenous secretory RAGE with oxidative stress in prediabetic patients. J Diabetes Complications 2015; 29: 422-426 [PMID: 25659638 DOI: 10.1016/j.jdiacomp.2014.12.007]
- Prasad K. Is there any evidence that AGE/sRAGE is a universal biomarker/risk marker for diseases? Mol Cell Biochem 2019; 451: 139-144 68 [PMID: 29961210 DOI: 10.1007/s11010-018-3400-2]
- Erusalimsky JD. The use of the soluble receptor for advanced glycation-end products (sRAGE) as a potential biomarker of disease risk and 69 adverse outcomes. Redox Biol 2021; 42: 101958 [PMID: 33839083 DOI: 10.1016/j.redox.2021.101958]
- Prasad K, Khan AS, Bhanumathy KK. Does AGE-RAGE Stress Play a Role in the Development of Coronary Artery Disease in Obesity? Int J 70 Angiol 2022; 31: 1-9 [PMID: 35221846 DOI: 10.1055/s-0042-1742587]
- Di Pino A, Urbano F, Zagami RM, Filippello A, Di Mauro S, Piro S, Purrello F, Rabuazzo AM. Low Endogenous Secretory Receptor for 71 Advanced Glycation End-Products Levels Are Associated With Inflammation and Carotid Atherosclerosis in Prediabetes. J Clin Endocrinol Metab 2016; 101: 1701-1709 [PMID: 26885882 DOI: 10.1210/jc.2015-4069]
- 72 Sabbatinelli J, Castiglione S, Macri F, Giuliani A, Ramini D, Vinci MC, Tortato E, Bonfigli AR, Olivieri F, Raucci A. Circulating levels of AGEs and soluble RAGE isoforms are associated with all-cause mortality and development of cardiovascular complications in type 2 diabetes: a retrospective cohort study. Cardiovasc Diabetol 2022; 21: 95 [PMID: 35668468 DOI: 10.1186/s12933-022-01535-3]
- 73 Scavello F, Tedesco CC, Castiglione S, Maciag A, Sangalli E, Veglia F, Spinetti G, Puca AA, Raucci A. Modulation of soluble receptor for advanced glycation end products isoforms and advanced glycation end products in long-living individuals. Biomark Med 2021; 15: 785-796 [PMID: 34236256 DOI: 10.2217/bmm-2020-0856]
- 74 Palma-Duran SA, Kontogianni MD, Vlassopoulos A, Zhao S, Margariti A, Georgoulis M, Papatheodoridis G, Combet E. Serum levels of advanced glycation end-products (AGEs) and the decoy soluble receptor for AGEs (sRAGE) can identify non-alcoholic fatty liver disease in age-, sex- and BMI-matched normo-glycemic adults. Metabolism 2018; 83: 120-127 [PMID: 29409822 DOI: 10.1016/j.metabol.2018.01.023]
- 75 Popp CJ, Zhou B, Manigrasso MB, Li H, Curran M, Hu L, St-Jules DE, Alemán JO, Vanegas SM, Jay M, Bergman M, Segal E, Sevick MA, Schmidt AM. Soluble Receptor for Advanced Glycation End Products (sRAGE) Isoforms Predict Changes in Resting Energy Expenditure in Adults with Obesity during Weight Loss. Curr Dev Nutr 2022; 6: nzac046 [PMID: 35542387 DOI: 10.1093/cdn/nzac046]
- Hurtado Del Pozo C, Ruiz HH, Arivazhagan L, Aranda JF, Shim C, Daya P, Derk J, MacLean M, He M, Frye L, Friedline RH, Noh HL, Kim 76 JK, Friedman RA, Ramasamy R, Schmidt AM. A Receptor of the Immunoglobulin Superfamily Regulates Adaptive Thermogenesis. Cell Rep 2019; **28**: 773-791.e7 [PMID: 31315054 DOI: 10.1016/j.celrep.2019.06.061]
- Popa I, Ganea E, Petrescu SM. Expression and subcellular localization of RAGE in melanoma cells. Biochem Cell Biol 2014; 92: 127-136 77 [PMID: 24697697 DOI: 10.1139/bcb-2013-0064]
- 78 Ruelas Cinco EDC, Ruíz Madrigal B, Domínguez Rosales JA, Maldonado González M, De la Cruz Color L, Ramírez Meza SM, Torres Baranda JR, Martínez López E, Hernández Nazará ZH. Expression of the receptor of advanced glycation end-products (RAGE) and membranal location in peripheral blood mononuclear cells (PBMC) in obesity and insulin resistance. Iran J Basic Med Sci 2019; 22: 623-630 [PMID: 31231489 DOI: 10.22038/ijbms.2019.34571.8206]
- Li J, Schmidt AM. Characterization and functional analysis of the promoter of RAGE, the receptor for advanced glycation end products. J Biol 79 Chem 1997; 272: 16498-16506 [PMID: 9195959 DOI: 10.1074/jbc.272.26.16498]
- Yan SD, Schmidt AM, Anderson GM, Zhang J, Brett J, Zou YS, Pinsky D, Stern D. Enhanced cellular oxidant stress by the interaction of 80 advanced glycation end products with their receptors/binding proteins. J Biol Chem 1994; 269: 9889-9897 [PMID: 8144582]
- 81 Corica D, Aversa T, Ruggeri RM, Cristani M, Alibrandi A, Pepe G, De Luca F, Wasniewska M. Could AGE/RAGE-Related Oxidative Homeostasis Dysregulation Enhance Susceptibility to Pathogenesis of Cardio-Metabolic Complications in Childhood Obesity? Front Endocrinol (Lausanne) 2019; 10: 426 [PMID: 31316471 DOI: 10.3389/fendo.2019.00426]
- Xia B, Zhu R, Zhang H, Chen B, Liu Y, Dai X, Ye Z, Zhao D, Mo F, Gao S, Wang XD, Bromme D, Wang L, Wang X, Zhang D. Lycopene 82 Improves Bone Quality and Regulates AGE/RAGE/NF-KB Signaling Pathway in High-Fat Diet-Induced Obese Mice. Oxid Med Cell Longev 2022; 2022: 3697067 [PMID: 35222796 DOI: 10.1155/2022/3697067]
- Pereira ENGDS, Araujo BP, Rodrigues KL, Silvares RR, Martins CSM, Flores EEI, Fernandes-Santos C, Daliry A. Simvastatin Improves 83 Microcirculatory Function in Nonalcoholic Fatty Liver Disease and Downregulates Oxidative and ALE-RAGE Stress. Nutrients 2022; 14 [PMID: 35277075 DOI: 10.3390/nu14030716]
- 84 Ji J, Feng M, Huang Y, Niu X. Liraglutide inhibits receptor for advanced glycation end products (RAGE)/reduced form of nicotinamideadenine dinucleotide phosphate (NAPDH) signaling to ameliorate non-alcoholic fatty liver disease (NAFLD) in vivo and vitro. Bioengineered 2022; 13: 5091-5102 [PMID: 35164657 DOI: 10.1080/21655979.2022.2036902]
- Zhu Y, Shu T, Lin Y, Wang H, Yang J, Shi Y, Han X. Inhibition of the receptor for advanced glycation endproducts (RAGE) protects 85 pancreatic β-cells. Biochem Biophys Res Commun 2011; 404: 159-165 [PMID: 21111711 DOI: 10.1016/j.bbrc.2010.11.085]
- Han D, Yamamoto Y, Munesue S, Motoyoshi S, Saito H, Win MT, Watanabe T, Tsuneyama K, Yamamoto H. Induction of receptor for 86 advanced glycation end products by insufficient leptin action triggers pancreatic β -cell failure in type 2 diabetes. Genes Cells 2013; 18: 302-314 [PMID: 23410183 DOI: 10.1111/gtc.12036]
- Kehm R, Rückriemen J, Weber D, Deubel S, Grune T, Höhn A. Endogenous advanced glycation end products in pancreatic islets after short-87 term carbohydrate intervention in obese, diabetes-prone mice. Nutr Diabetes 2019; 9: 9 [PMID: 30858378 DOI: 10.1038/s41387-019-0077-x]
- Bai R, Zhang T, Gao Y, Shu T, Zhou Y, Wang F, Chang X, Tang W, Zhu Y, Han X. Rab31, a receptor of advanced glycation end products 88 (RAGE) interacting protein, inhibits AGE induced pancreatic β-cell apoptosis through the pAKT/BCL2 pathway. Endocr J 2022; 69: 1015-1026 [PMID: 35314532 DOI: 10.1507/endocrj.EJ21-0594]
- 89 Bayatpoor ME, Mirzaee S, Karami Abd M, Mohammadi MT, Shahyad S, Bahari Z, Raouf Sarshoori J. Crocin treatment decreased pancreatic atrophy, LOX-1 and RAGE mRNA expression of pancreas tissue in cholesterol-fed and streptozotocin-induced diabetic rats. J Complement Integr Med 2019; 17 [PMID: 31532754 DOI: 10.1515/jcim-2019-0117]
- Lee BW, Chae HY, Kwon SJ, Park SY, Ihm J, Ihm SH. RAGE ligands induce apoptotic cell death of pancreatic β -cells via oxidative stress. Int 90 J Mol Med 2010; 26: 813-818 [PMID: 21042774]
- Abedini A, Cao P, Plesner A, Zhang J, He M, Derk J, Patil SA, Rosario R, Lonier J, Song F, Koh H, Li H, Raleigh DP, Schmidt AM. RAGE 91 binds preamyloid IAPP intermediates and mediates pancreatic β cell proteotoxicity. J Clin Invest 2018; **128**: 682-698 [PMID: 29337308 DOI: 10.1172/JCI85210]
- 92 Tsilidis KK, Kasimis JC, Lopez DS, Ntzani EE, Ioannidis JP. Type 2 diabetes and cancer: umbrella review of meta-analyses of observational



studies. BMJ 2015; 350: g7607 [PMID: 25555821 DOI: 10.1136/bmj.g7607]

- Pearson-Stuttard J, Papadimitriou N, Markozannes G, Cividini S, Kakourou A, Gill D, Rizos EC, Monori G, Ward HA, Kyrgiou M, Gunter 93 MJ, Tsilidis KK. Type 2 Diabetes and Cancer: An Umbrella Review of Observational and Mendelian Randomization Studies. Cancer Epidemiol Biomarkers Prev 2021; 30: 1218-1228 [PMID: 33737302 DOI: 10.1158/1055-9965.EPI-20-1245]
- 94 Dashti SG, Simpson JA, Viallon V, Karahalios A, Moreno-Betancur M, Brasky T, Pan K, Rohan TE, Shadyab AH, Thomson CA, Wild RA, Wassertheil-Smoller S, Ho GYF, Strickler HD, English DR, Gunter MJ. Adiposity and breast, endometrial, and colorectal cancer risk in postmenopausal women: Quantification of the mediating effects of leptin, C-reactive protein, fasting insulin, and estradiol. Cancer Med 2022; 11: 1145-1159 [PMID: 35048536 DOI: 10.1002/cam4.4434]
- Brown KA. Metabolic pathways in obesity-related breast cancer. Nat Rev Endocrinol 2021; 17: 350-363 [PMID: 33927368 DOI: 95 10.1038/s41574-021-00487-0]
- 96 Mahboobifard F, Pourgholami MH, Jorjani M, Dargahi L, Amiri M, Sadeghi S, Tehrani FR. Estrogen as a key regulator of energy homeostasis and metabolic health. Biomed Pharmacother 2022; 156: 113808 [PMID: 36252357 DOI: 10.1016/j.biopha.2022.113808]
- Parida S, Sharma D. The Microbiome-Estrogen Connection and Breast Cancer Risk. Cells 2019; 8 [PMID: 31847455 DOI: 97 10.3390/cells8121642]
- Scully T, Ettela A, LeRoith D, Gallagher EJ. Obesity, Type 2 Diabetes, and Cancer Risk. Front Oncol 2020; 10: 615375 [PMID: 33604295 98 DOI: 10.3389/fonc.2020.615375]
- Kang C, LeRoith D, Gallagher EJ. Diabetes, Obesity, and Breast Cancer. Endocrinology 2018; 159: 3801-3812 [PMID: 30215698 DOI: 99 10.1210/en.2018-00574]
- Hopkins BD, Goncalves MD, Cantley LC. Insulin-PI3K signalling: an evolutionarily insulated metabolic driver of cancer. Nat Rev Endocrinol 100 2020; 16: 276-283 [PMID: 32127696 DOI: 10.1038/s41574-020-0329-9]
- Ramteke P, Deb A, Shepal V, Bhat MK. Hyperglycemia Associated Metabolic and Molecular Alterations in Cancer Risk, Progression, 101 Treatment, and Mortality. Cancers (Basel) 2019; 11 [PMID: 31546918 DOI: 10.3390/cancers11091402]
- Lai SWT, Lopez Gonzalez EJ, Zoukari T, Ki P, Shuck SC. Methylglyoxal and Its Adducts: Induction, Repair, and Association with Disease. Chem Res Toxicol 2022; 35: 1720-1746 [PMID: 36197742 DOI: 10.1021/acs.chemrestox.2c00160]
- Adeshara KA, Bangar N, Diwan AG, Tupe RS. Plasma glycation adducts and various RAGE isoforms are intricately associated with oxidative stress and inflammatory markers in type 2 diabetes patients with vascular complications. Diabetes Metab Syndr 2022; 16: 102441 [PMID: 35247657 DOI: 10.1016/j.dsx.2022.102441]
- Eva TA, Barua N, Chowdhury MM, Yeasmin S, Rakib A, Islam MR, Emran TB, Simal-Gandara J. Perspectives on signaling for biological-104 and processed food-related advanced glycation end-products and its role in cancer progression. Crit Rev Food Sci Nutr 2022; 62: 2655-2672 [PMID: 33307763 DOI: 10.1080/10408398.2020.1856771]
- Rao NL, Kotian GB, Shetty JK, Shelley BP, Dmello MK, Lobo EC, Shankar SP, Almeida SD, Shah SR. Receptor for Advanced Glycation End 105 Product, Organ Crosstalk, and Pathomechanism Targets for Comprehensive Molecular Therapeutics in Diabetic Ischemic Stroke. Biomolecules 2022; 12 [PMID: 36421725 DOI: 10.3390/biom12111712]
- 106 Li S, Yang D, Gao X, Yao S, Wang S, Zhu J, Shu J. Argpyrimidine bonded to RAGE regulates autophagy and cell cycle to cause periodontal destruction. J Cell Physiol 2022; 237: 4460-4476 [PMID: 36166691 DOI: 10.1002/jcp.30886]
- Wang Y, Jiang C, Shang Z, Qiu G, Yuan G, Xu K, Hou Q, He Y, Liu Y. AGEs/RAGE Promote Osteogenic Differentiation in Rat Bone 107 Marrow-Derived Endothelial Progenitor Cells via MAPK Signaling. J Diabetes Res 2022; 2022: 4067812 [PMID: 35155684 DOI: 10.1155/2022/4067812]
- Gottschalk G, Peterson D, Knox K, Maynard M, Whelan RJ, Roy A. Elevated ATG13 in serum of patients with ME/CFS stimulates oxidative 108 stress response in microglial cells via activation of receptor for advanced glycation end products (RAGE). Mol Cell Neurosci 2022; 120: 103731 [PMID: 35487443 DOI: 10.1016/j.mcn.2022.103731]
- Teissier T, Temkin V, Pollak RD, Cox LS. Crosstalk Between Senescent Bone Cells and the Bone Tissue Microenvironment Influences Bone 109 Fragility During Chronological Age and in Diabetes. Front Physiol 2022; 13: 812157 [PMID: 35388291 DOI: 10.3389/fphys.2022.812157]
- Burr SD, Dorroh CC, Stewart JA Jr. Rap1a Activity Elevated the Impact of Endogenous AGEs in Diabetic Collagen to Stimulate Increased 110 Myofibroblast Transition and Oxidative Stress. Int J Mol Sci 2022; 23 [PMID: 35562872 DOI: 10.3390/ijms23094480]
- Taneja S, Vetter SW, Leclerc E. Hypoxia and the Receptor for Advanced Glycation End Products (RAGE) Signaling in Cancer. Int J Mol Sci 111 2021; 22 [PMID: 34360919 DOI: 10.3390/ijms22158153]
- Seo J, Yun JE, Kim SJ, Chun YS. Lipid metabolic reprogramming by hypoxia-inducible factor-1 in the hypoxic tumour microenvironment. 112 *Pflugers Arch* 2022; **474**: 591-601 [PMID: 35348849 DOI: 10.1007/s00424-022-02683-x]
- Nie Y, Yang D, Oppenheim JJ. Alarmins and Antitumor Immunity. Clin Ther 2016; 38: 1042-1053 [PMID: 27101817 DOI: 113 10.1016/j.clinthera.2016.03.021
- Zhou J, Bai W, Liu Q, Cui J, Zhang W. Intermittent Hypoxia Enhances THP-1 Monocyte Adhesion and Chemotaxis and Promotes M1 114 Macrophage Polarization via RAGE. Biomed Res Int 2018; 2018: 1650456 [PMID: 30402462 DOI: 10.1155/2018/1650456]
- D'Arcy MS. Cell death: a review of the major forms of apoptosis, necrosis and autophagy. Cell Biol Int 2019; 43: 582-592 [PMID: 30958602 115 DOI: 10.1002/cbin.11137]
- Tang D, Kang R, Coyne CB, Zeh HJ, Lotze MT. PAMPs and DAMPs: signal 0s that spur autophagy and immunity. Immunol Rev 2012; 249: 116 158-175 [PMID: 22889221 DOI: 10.1111/j.1600-065X.2012.01146.x]
- Tang D, Kang R, Cheh CW, Livesey KM, Liang X, Schapiro NE, Benschop R, Sparvero LJ, Amoscato AA, Tracey KJ, Zeh HJ, Lotze MT. 117 HMGB1 release and redox regulates autophagy and apoptosis in cancer cells. Oncogene 2010; 29: 5299-5310 [PMID: 20622903 DOI: 10.1038/onc.2010.261]
- Waghela BN, Vaidya FU, Ranjan K, Chhipa AS, Tiwari BS, Pathak C. AGE-RAGE synergy influences programmed cell death signaling to 118 promote cancer. Mol Cell Biochem 2021; 476: 585-598 [PMID: 33025314 DOI: 10.1007/s11010-020-03928-y]
- 119 Kang R, Tang D, Schapiro NE, Livesey KM, Farkas A, Loughran P, Bierhaus A, Lotze MT, Zeh HJ. The receptor for advanced glycation end products (RAGE) sustains autophagy and limits apoptosis, promoting pancreatic tumor cell survival. Cell Death Differ 2010; 17: 666-676 [PMID: 19834494 DOI: 10.1038/cdd.2009.149]
- Kumar V, Agrawal R, Pandey A, Kopf S, Hoeffgen M, Kaymak S, Bandapalli OR, Gorbunova V, Seluanov A, Mall MA, Herzig S, Nawroth 120 PP. Compromised DNA repair is responsible for diabetes-associated fibrosis. EMBO J 2020; 39: e103477 [PMID: 32338774 DOI: 10.15252/embj.2019103477
- 121 Melia F, Udomjarumanee P, Zinovkin D, Arghiani N, Pranjol MZI. Pro-tumorigenic role of type 2 diabetes-induced cellular senescence in



colorectal cancer. Front Oncol 2022; 12: 975644 [PMID: 36059680 DOI: 10.3389/fonc.2022.975644]

- Moaddel R, Ubaida-Mohien C, Tanaka T, Lyashkov A, Basisty N, Schilling B, Semba RD, Franceschi C, Gorospe M, Ferrucci L. Proteomics 122 in aging research: A roadmap to clinical, translational research. Aging Cell 2021; 20: e13325 [PMID: 33730416 DOI: 10.1111/acel.13325]
- Chen Y, Liu Z, Chen H, Huang X, Lei Y, Liang Q, Wei J, Zhang Q, Guo X, Huang Q. p53 SUMOylation Mediates AOPP-Induced Endothelial 123 Senescence and Apoptosis Evasion. Front Cardiovasc Med 2021; 8: 795747 [PMID: 35187108 DOI: 10.3389/fcvm.2021.795747]
- Garay-Sevilla ME, Gomez-Ojeda A, González I, Luévano-Contreras C, Rojas A. Contribution of RAGE axis activation to the association 124 between metabolic syndrome and cancer. Mol Cell Biochem 2021; 476: 1555-1573 [PMID: 33398664 DOI: 10.1007/s11010-020-04022-z]
- Pujals M, Resar L, Villanueva J. HMGA1, Moonlighting Protein Function, and Cellular Real Estate: Location, Location! 125 Biomolecules 2021; 11 [PMID: 34572547 DOI: 10.3390/biom11091334]
- Bresnick AR, Weber DJ, Zimmer DB. S100 proteins in cancer. Nat Rev Cancer 2015; 15: 96-109 [PMID: 25614008 DOI: 10.1038/nrc3893] 126
- Yu GH, Li SF, Wei R, Jiang Z. Diabetes and Colorectal Cancer Risk: Clinical and Therapeutic Implications. J Diabetes Res 2022; 2022: 127 1747326 [PMID: 35296101 DOI: 10.1155/2022/1747326]
- Khambu B, Hong H, Liu S, Liu G, Chen X, Dong Z, Wan J, Yin XM. The HMGB1-RAGE axis modulates the growth of autophagy-deficient 128 hepatic tumors. Cell Death Dis 2020; 11: 333 [PMID: 32382012 DOI: 10.1038/s41419-020-2536-7]
- Swami P, Thiyagarajan S, Vidger A, Indurthi VSK, Vetter SW, Leclerc E. RAGE Up-Regulation Differently Affects Cell Proliferation and 129 Migration in Pancreatic Cancer Cells. Int J Mol Sci 2020; 21 [PMID: 33086527 DOI: 10.3390/ijms21207723]
- El-Far AH, Sroga G, Jaouni SKA, Mousa SA. Role and Mechanisms of RAGE-Ligand Complexes and RAGE-Inhibitors in Cancer 130 Progression. Int J Mol Sci 2020; 21 [PMID: 32443845 DOI: 10.3390/ijms21103613]
- 131 Rojas A, Schneider I, Lindner C, Gonzalez I, Morales MA. The RAGE/multiligand axis: a new actor in tumor biology. Biosci Rep 2022; 42 [PMID: 35727208 DOI: 10.1042/BSR20220395]
- Muthyalaiah YS, Jonnalagadda B, John CM, Arockiasamy S. Impact of Advanced Glycation End products (AGEs) and its receptor (RAGE) 132 on cancer metabolic signaling pathways and its progression. *Glycoconj J* 2021; **38**: 717-734 [PMID: 35064413 DOI: 10.1007/s10719-021-10031-x]
- Ennis CS, Llevenes P, Qiu Y, Dries R, Denis GV. The crosstalk within the breast tumor microenvironment in type II diabetes: Implications for 133 cancer disparities. Front Endocrinol (Lausanne) 2022; 13: 1044670 [PMID: 36531496 DOI: 10.3389/fendo.2022.1044670]
- Azizian-Farsani F, Abedpoor N, Hasan Sheikhha M, Gure AO, Nasr-Esfahani MH, Ghaedi K. Receptor for Advanced Glycation End Products 134 Acts as a Fuel to Colorectal Cancer Development. Front Oncol 2020; 10: 552283 [PMID: 33117687 DOI: 10.3389/fonc.2020.552283]
- Mollace A, Coluccio ML, Donato G, Mollace V, Malara N. Cross-talks in colon cancer between RAGE/AGEs axis and inflammation/ 135 immunotherapy. Oncotarget 2021; 12: 1281-1295 [PMID: 34194625 DOI: 10.18632/oncotarget.27990]
- Uhlen M, Zhang C, Lee S, Sjöstedt E, Fagerberg L, Bidkhori G, Benfeitas R, Arif M, Liu Z, Edfors F, Sanli K, von Feilitzen K, Oksvold P, 136 Lundberg E, Hober S, Nilsson P, Mattsson J, Schwenk JM, Brunnström H, Glimelius B, Sjöblom T, Edqvist PH, Djureinovic D, Micke P, Lindskog C, Mardinoglu A, Ponten F. A pathology atlas of the human cancer transcriptome. Science 2017; 357 [PMID: 28818916 DOI: 10.1126/science.aan2507]
- Santolla MF, Talia M, Cirillo F, Scordamaglia D, De Rosis S, Spinelli A, Miglietta AM, Nardo B, Filippelli G, De Francesco EM, Belfiore A, 137 Lappano R, Maggiolini M. The AGEs/RAGE Transduction Signaling Prompts IL-8/CXCR1/2-Mediated Interaction between Cancer-Associated Fibroblasts (CAFs) and Breast Cancer Cells. Cells 2022; 11 [PMID: 35954247 DOI: 10.3390/cells11152402]
- Shah SN, Cope L, Poh W, Belton A, Roy S, Talbot CC Jr, Sukumar S, Huso DL, Resar LM. HMGA1: a master regulator of tumor progression 138 in triple-negative breast cancer cells. PLoS One 2013; 8: e63419 [PMID: 23658826 DOI: 10.1371/journal.pone.0063419]
- Chen Y, Cai L, Guo X, Li Z, Liao X, Zhang X, Huang L, He J. HMGB1-activated fibroblasts promote breast cancer cells metastasis via 139 RAGE/aerobic glycolysis. Neoplasma 2021; 68: 71-78 [PMID: 33030958 DOI: 10.4149/neo_2020_200610N620]
- Amornsupak K, Thongchot S, Thinyakul C, Box C, Hedayat S, Thuwajit P, Eccles SA, Thuwajit C. HMGB1 mediates invasion and PD-L1 140 expression through RAGE-PI3K/AKT signaling pathway in MDA-MB-231 breast cancer cells. BMC Cancer 2022; 22: 578 [PMID: 35610613 DOI: 10.1186/s12885-022-09675-1]
- He H, Wang X, Chen J, Sun L, Sun H, Xie K. High-Mobility Group Box 1 (HMGB1) Promotes Angiogenesis and Tumor Migration by 141 Regulating Hypoxia-Inducible Factor 1 (HIF-1α) Expression via the Phosphatidylinositol 3-Kinase (PI3K)/AKT Signaling Pathway in Breast Cancer Cells. Med Sci Monit 2019; 25: 2352-2360 [PMID: 30930461 DOI: 10.12659/MSM.915690]
- Wang L, Kang FB, Wang J, Yang C, He DW. Downregulation of miR-205 contributes to epithelial-mesenchymal transition and invasion in 142 triple-negative breast cancer by targeting HMGB1-RAGE signaling pathway. Anticancer Drugs 2019; 30: 225-232 [PMID: 30334817 DOI: 10.1097/CAD.000000000000705]
- Okui T, Hiasa M, Ryumon S, Ono K, Kunisada Y, Ibaragi S, Sasaki A, Roodman GD, White FA, Yoneda T. The HMGB1/RAGE axis induces 143 bone pain associated with colonization of 4T1 mouse breast cancer in bone. J Bone Oncol 2021; 26: 100330 [PMID: 33204606 DOI: 10.1016/j.jbo.2020.100330]
- Li X, Wang M, Gong T, Lei X, Hu T, Tian M, Ding F, Ma F, Chen H, Liu Z. A S100A14-CCL2/CXCL5 signaling axis drives breast cancer 144 metastasis. Theranostics 2020; 10: 5687-5703 [PMID: 32483412 DOI: 10.7150/thno.42087]
- Muoio MG, Talia M, Lappano R, Sims AH, Vella V, Cirillo F, Manzella L, Giuliano M, Maggiolini M, Belfiore A, De Francesco EM. 145 Activation of the S100A7/RAGE Pathway by IGF-1 Contributes to Angiogenesis in Breast Cancer. Cancers (Basel) 2021; 13 [PMID: 33557316 DOI: 10.3390/cancers13040621]
- Mishra S, Charan M, Shukla RK, Agarwal P, Misri S, Verma AK, Ahirwar DK, Siddiqui J, Kaul K, Sahu N, Vyas K, Garg AA, Khan A, Miles 146 WO, Song JW, Bhutani N, Ganju RK. cPLA2 blockade attenuates S100A7-mediated breast tumorigenicity by inhibiting the immunosuppressive tumor microenvironment. J Exp Clin Cancer Res 2022; 41: 54 [PMID: 35135586 DOI: 10.1186/s13046-021-02221-0]
- Rigiracciolo DC, Nohata N, Lappano R, Cirillo F, Talia M, Adame-Garcia SR, Arang N, Lubrano S, De Francesco EM, Belfiore A, Gutkind 147 JS, Maggiolini M. Focal Adhesion Kinase (FAK)-Hippo/YAP transduction signaling mediates the stimulatory effects exerted by S100A8/A9-RAGE system in triple-negative breast cancer (TNBC). J Exp Clin Cancer Res 2022; 41: 193 [PMID: 35655319 DOI: 10.1186/s13046-022-02396-0]
- Wilkie T, Verma AK, Zhao H, Charan M, Ahirwar DK, Kant S, Pancholi V, Mishra S, Ganju RK. Lipopolysaccharide from the commensal 148 microbiota of the breast enhances cancer growth: role of S100A7 and TLR4. Mol Oncol 2022; 16: 1508-1522 [PMID: 33969603 DOI: 10.1002/1878-0261.12975]
- Ryan D, Koziol J, ElShamy WM. Targeting AXL and RAGE to prevent geminin overexpression-induced triple-negative breast cancer 149 metastasis. Sci Rep 2019; 9: 19150 [PMID: 31844158 DOI: 10.1038/s41598-019-55702-w]



- Sun X, Wang T, Zhang C, Ning K, Guan ZR, Chen SX, Hong TT, Hua D. S100A16 is a prognostic marker for colorectal cancer. J Surg Oncol 150 2018; 117: 275-283 [PMID: 28876468 DOI: 10.1002/jso.24822]
- Huang CY, Chiang SF, Ke TW, Chen TW, Lan YC, You YS, Shiau AC, Chen WT, Chao KSC. Cytosolic high-mobility group box protein 1 151 (HMGB1) and/or PD-1+ TILs in the tumor microenvironment may be contributing prognostic biomarkers for patients with locally advanced rectal cancer who have undergone neoadjuvant chemoradiotherapy. Cancer Immunol Immunother 2018; 67: 551-562 [PMID: 29270668 DOI: 10.1007/s00262-017-2109-5
- Zheng J, Zhu W, He F, Li Z, Cai N, Wang HH. An Aptamer-Based Antagonist against the Receptor for Advanced Glycation End-Products 152 (RAGE) Blocks Development of Colorectal Cancer. Mediators Inflamm 2021; 2021: 9958051 [PMID: 34035661 DOI: 10.1155/2021/9958051]
- Niu S, Zhao ZG, Lyu XM, Zhao M, Wang XZ, Liu WN, Zhao W, Zhang XH, Wang Y. [The expression and significance of IGF1R-Ras/ 153 RAGE-HMGB1 pathway in colorectal cancer patients with type 2 diabetes mellitus]. Zhonghua Zhong Liu Za Zhi 2020; 42: 391-395 [PMID: 32482028 DOI: 10.3760/cma.j.cn112152-112152-20190906-00580]
- 154 Wang P, Lu YC, Li YF, Wang L, Lee SC. Advanced Glycation End Products Increase MDM2 Expression via Transcription Factor KLF5. J Diabetes Res 2018; 2018: 3274084 [PMID: 30271790 DOI: 10.1155/2018/3274084]
- 155 Huang M, Geng Y, Deng Q, Li R, Shao X, Zhang Z, Xu W, Wu Y, Ma Q. Translationally controlled tumor protein affects colorectal cancer metastasis through the high mobility group box 1-dependent pathway. Int J Oncol 2018; 53: 1481-1492 [PMID: 30066846 DOI: 10.3892/ijo.2018.4502]
- Huang M, Wu R, Chen L, Peng Q, Li S, Zhang Y, Zhou L, Duan L. S100A9 Regulates MDSCs-Mediated Immune Suppression via the RAGE 156 and TLR4 Signaling Pathways in Colorectal Carcinoma. Front Immunol 2019; 10: 2243 [PMID: 31620141 DOI: 10.3389/fimmu.2019.02243]
- Qian F, Xiao J, Gai L, Zhu J. HMGB1-RAGE signaling facilitates Ras-dependent Yap1 expression to drive colorectal cancer stemness and 157 development. Mol Carcinog 2019; 58: 500-510 [PMID: 30456802 DOI: 10.1002/mc.22944]
- Seguella L, Capuano R, Pesce M, Annunziata G, de Conno B, Sarnelli G, Aurino L, Esposito G. S100B Protein Stimulates Proliferation and 158 Angiogenic Mediators Release through RAGE/pAkt/mTOR Pathway in Human Colon Adenocarcinoma Caco-2 Cells. Int J Mol Sci 2019; 20 [PMID: 31266264 DOI: 10.3390/ijms20133240]
- Huang CY, Chiang SF, Chen WT, Ke TW, Chen TW, You YS, Lin CY, Chao KSC, Huang CY. HMGB1 promotes ERK-mediated 159 mitochondrial Drp1 phosphorylation for chemoresistance through RAGE in colorectal cancer. Cell Death Dis 2018; 9: 1004 [PMID: 30258050 DOI: 10.1038/s41419-018-1019-6]
- 160 Zhan X, Wu R, Kong XH, You Y, He K, Sun XY, Huang Y, Chen WX, Duan L. Elevated neutrophil extracellular traps by HBV-mediated S100A9-TLR4/RAGE-ROS cascade facilitate the growth and metastasis of hepatocellular carcinoma. Cancer Commun (Lond) 2023; 43: 225-245 [PMID: 36346061 DOI: 10.1002/cac2.12388]
- Ando K, Sakoda M, Ueno S, Hiwatashi K, Iino S, Minami K, Kawasaki Y, Hashiguchi M, Tanoue K, Mataki Y, Kurahara H, Maemura K, 161 Shinchi H, Natsugoe S. Clinical Implication of the Relationship Between High Mobility Group Box-1 and Tumor Differentiation in Hepatocellular Carcinoma. Anticancer Res 2018; 38: 3411-3418 [PMID: 29848691 DOI: 10.21873/anticanres.12609]
- Li J, Ren H, Wang J, Zhang P, Shi X. Extracellular HMGB1 promotes CD44 expression in hepatocellular carcinoma via regulating miR-21. 162 Aging (Albany NY) 2021; 13: 8380-8395 [PMID: 33661757 DOI: 10.18632/aging.202649]
- 163 Li Y, Wang J, Song K, Liu S, Zhang H, Wang F, Ni C, Zhai W, Liang J, Qin Z, Zhang J. S100A4 promotes hepatocellular carcinogenesis by intensifying fibrosis-associated cancer cell stemness. Oncoimmunology 2020; 9: 1725355 [PMID: 32117590 DOI: 10.1080/2162402X.2020.1725355
- Bachmann M, Lamprecht L, Gonther S, Pfeilschifter J, Mühl H. A murine cellular model of necroinflammation displays RAGE-dependent 164 cytokine induction that connects to hepatoma cell injury. J Cell Mol Med 2020; 24: 10356-10366 [PMID: 32697038 DOI: 10.1111/jcmm.15649
- Li S, Gu H, Huang Y, Peng Q, Zhou R, Yi P, Chen R, Huang Z, Hu X, Tang D. Circular RNA 101368/miR-200a axis modulates the migration 165 of hepatocellular carcinoma through HMGB1/RAGE signaling. Cell Cycle 2018; 17: 2349-2359 [PMID: 30265210 DOI: 10.1080/15384101.2018.1526599]
- Boone BA, Orlichenko L, Schapiro NE, Loughran P, Gianfrate GC, Ellis JT, Singhi AD, Kang R, Tang D, Lotze MT, Zeh HJ. The receptor for 166 advanced glycation end products (RAGE) enhances autophagy and neutrophil extracellular traps in pancreatic cancer. Cancer Gene Ther 2015; 22: 326-334 [PMID: 25908451 DOI: 10.1038/cgt.2015.21]
- Lan CY, Chen SY, Kuo CW, Lu CC, Yen GC. Quercetin facilitates cell death and chemosensitivity through RAGE/PI3K/AKT/mTOR axis in 167 human pancreatic cancer cells. J Food Drug Anal 2019; 27: 887-896 [PMID: 31590760 DOI: 10.1016/j.jfda.2019.07.001]
- Uchida C, Mizukami H, Hara Y, Saito T, Umetsu S, Igawa A, Osonoi S, Kudoh K, Yamamoto Y, Yamamoto H, Yagihashi S, Hakamada K. 168 Diabetes in Humans Activates Pancreatic Stellate Cells via RAGE in Pancreatic Ductal Adenocarcinoma. Int J Mol Sci 2021; 22 [PMID: 34769147 DOI: 10.3390/ijms222111716]
- 169 Chen SY, Hsu YH, Wang SY, Chen YY, Hong CJ, Yen GC. Lucidone inhibits autophagy and MDR1 via HMGB1/RAGE/PI3K/Akt signaling pathway in pancreatic cancer cells. Phytother Res 2022; 36: 1664-1677 [PMID: 35224793 DOI: 10.1002/ptr.7385]





Published by Baishideng Publishing Group Inc 7041 Koll Center Parkway, Suite 160, Pleasanton, CA 94566, USA Telephone: +1-925-3991568 E-mail: bpgoffice@wjgnet.com Help Desk: https://www.f6publishing.com/helpdesk https://www.wjgnet.com

