

Effect of *GP73* silencing on proliferation and apoptosis in hepatocellular cancer

Yu-Long Zhang, You-Cheng Zhang, Wei Han, Yu-Min Li, Geng-Nian Wang, Shao Yuan, Feng-Xian Wei, Jia-Feng Wang, Jian-Jun Jiang, Ya-Wu Zhang

Yu-Long Zhang, You-Cheng Zhang, Wei Han, Yu-Min Li, Geng-Nian Wang, Shao Yuan, Feng-Xian Wei, Jia-Feng Wang, Jian-Jun Jiang, Ya-Wu Zhang, Department of General Surgery, Lanzhou University Second Hospital, Lanzhou 730030, Gansu Province, China

Yu-Long Zhang, You-Cheng Zhang, Wei Han, Yu-Min Li, Ya-Wu Zhang, Hepato-Biliary-Pancreatic Institute, Lanzhou University Second Hospital, Lanzhou 730030, Gansu Province, China

Yu-Long Zhang, You-Cheng Zhang, Wei Han, Yu-Min Li, Ya-Wu Zhang, Gansu Provincial-Level Key Laboratory of Digestive System Tumors, Lanzhou 730030, Gansu Province, China

Author contributions: Zhang YL, Zhang YC, Han W and Li YM designed the research; Zhang YL, Wang GN, Wei FX, Wang JF and Yuan S performed the research; Jiang JJ and Zhang YW contributed new reagents or analytic tools; Zhang YL and Zhang YC analyzed the data; Zhang YL and Zhang YC wrote the paper. **Supported by** The Grant from Gansu Provincial-level Key Laboratory of Digestive System Tumors Open projects Research Fund, No. lzujbky-2011-t03

Correspondence to: You-Cheng Zhang, MD, PhD, Department of General Surgery, Lanzhou University Second Hospital, Cuiyingmen 82, Chengguan District, Lanzhou 730030, Gansu Province, China. zhangyouchengphd@163.com

Telephone: +86-931-8942287 Fax: +86-931-8942287

Received: December 25, 2013 Revised: February 28, 2014

Accepted: April 21, 2014

Published online: August 28, 2014

Abstract

AIM: To investigate the roles of Golgi protein (GP) 73 in the regulation of cell proliferation and apoptosis.

METHODS: Stealth RNAi targeting *GP73* gene sequence was used to silence its expression in Hep G2 cells and Bel7402 cells. Stealth RNAi effects were assessed by reverse transcriptase polymerase chain reaction and ELISA. Cell proliferation assay and cell cycle analysis were assessed by MTT assay and flow cytometry. Apoptosis was assessed by flow cytometry and

transmission electron microscopy. Apoptosis-related proteins were assessed by western immunoblot analysis.

RESULTS: Stealth RNAi targeting *GP73* gene sequence markedly reduced the expression of *GP73* gene. The reduction of GP73 in Hep G2 cells and Bel7402 cells inhibited cell proliferation and induced apoptosis, however, terminal apoptosis occurred in Hep G2 cells, but early apoptosis occurred in Bel7402 cells. Reduced expression of *GP73* gene might lead to a reduction in Bcl-2/Bax ratio, an increase in cytochrome c, but a reduction in capase-3.

CONCLUSION: GP73 might play an important role in proliferation and apoptosis in hepatocellular carcinoma cells.

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Key words: Hepatocellular carcinoma; Golgi protein-73; Cell proliferation; Apoptosis

Core tip: Stealth RNAi targeting Golgi protein (GP)73 gene sequence markedly reduced the expression of *GP73* gene. Reduction of GP73 in Hep G2 cells and Bel7402 cells inhibited cell proliferation and induced apoptosis, however, terminal apoptosis occurred in Hep G2 cells, but early apoptosis occurred in Bel7402 cells. Reduced expression of *GP73* gene might lead to a reduction in Bcl-2/ Bax ratio, an increase in cytochrome c, but a reduction in capase-3. GP73 might play an important role in proliferation and apoptosis in Hep G2 and Bel7402 cells.

Zhang YL, Zhang YC, Han W, Li YM, Wang GN, Yuan S, Wei FX, Wang JF, Jiang JJ, Zhang YW. Effect of *GP73* silencing on proliferation and apoptosis in hepatocellular cancer. *World J Gastroenterol* 2014; 20(32): 11287-11296 Available from: URL: <http://www.wjgnet.com/1007-9327/full/v20/i32/11287.htm> DOI:

INTRODUCTION

Hepatocellular carcinoma (HCC) is the fifth most common malignancy worldwide, and > 80% of HCC occurs in developing countries^[1]. HCC remains a significant concern of cancer research because of its poor survival rate and high rates of recurrence^[2,3]. Prognosis of surgical or loco-regional therapies for patients with intermediate- and advanced-stage disease remains poor^[4]. Several effective gene-targeting agents are currently being tested in preclinical studies^[5-9]. Unfortunately, no chemotherapy is effective for HCC patients.

Kladney *et al.*^[10] first isolated Golgi protein (GP)73 in a genetic screen. Expression of GP73 is increased markedly in HCC cells and its serum levels appear to be predictive of HCC^[11-15], and several studies have reported the use of GP73 as a serum marker for HCC^[16-22]. GP73 may be elevated even when small undetectable tumors are present^[23]. The physiological and pathological roles of GP73 have attracted considerable attention in recent years^[24-30]. However the function of GP73 in hepatic carcinoma cells remains obscure. The expression of GP73 was silenced in the HCC cell line Bel7402 and Hep G2 by stealth RNAi, which serves as a powerful technology to block specifically the expression of target genes in the present study^[31-35]. The effects of GP73 on cell proliferation and apoptosis were also evaluated in this study.

MATERIALS AND METHODS

Stealth RNAi

According to the siRNA design guidelines^[27,28], one RNAi target sequence was selected corresponding to the nucleotides of RNAi-Stealth RNAi of the human GP73 mRNA (GenBank Accession No. NM177937.2). The sequence of the synthesized oligonucleotide was: HSS181966: sense 5'-GGAAACGGGCGUCGAG-CAUGAAGU-3', anti-sense 5'-ACUUCAUGC UGCU-ACGCCCGUUUCC-3'.

Transfection

Lipofectamine RNAi Max transfection agent (Invitrogen, Carlsbad, CA, United States) was used to transfect synthesized Stealth RNAi against GP73 into Hep G2 and Bel7402 cells. BLOCK-iT Alexa Fluor Red Fluorescent (Invitrogen) was used to confirm the transfection efficiency of each duplex siRNA. Stealth RNAi, fluorescent logo or negative control duplexes were delivered into Hep G2 and Bel7402 cells through reverse transfection.

Reverse transcriptase polymerase chain reaction

To analyze quantitatively the effects of Stealth RNAi on GP73 mRNA, cells were transfected with Stealth RNAi or negative control in culture flasks. After 24 and 48 h, cells were harvested by trypsinization and rinsed

twice with cold PBS. TRIzol reagent (Invitrogen) was used to extract total RNA. Two-step real-time reverse transcriptase polymerase chain reaction (RT-PCR) kits (TakaRa, Japan) was used to perform first-strand cDNA synthesis and amplification. The 7000 Sequence Detection System (Applied Biosystems, Foster City, CA, United States) was used to perform quantitative PCR amplifications. A 20- μ L reaction volume containing 10 μ L 2 \times SYBR Premix Ex TaqTM (TakaRa) was used to carry out this reaction. β -Actin was used as an internal standard. The primer sequences were: GP73 sense 5'-GTGCT-GGTGCCAGCCTGTTA-3' and anti-sense 5'-AGT-GCTCTAGGCCA TTGATTGATTG-3', β -actin sense 5'-GCAAGCAGGAGTATGACGAGT-3' and anti-sense 5'-GCAAGCAGGAGTATGACGAGT-3'. Thermal cycle conditions: 95 °C for 30 s, followed by 40 cycles of 94 °C for 5 s, and 61 °C for 30 s. The Δ Ct of each group was calculated by the formula: Δ Ct = Ct_{GP73} - Ct _{β -actin}. $\Delta\Delta$ Ct was calculated by Δ Ct_{treated} - Δ Ct_{control}. The fold change for GP73 expression levels of the treated groups were calculated using 2^{- $\Delta\Delta$ Ct}. The primers described above and the comparative threshold (Ct) method were used to calculate the relative amount of mRNA in the treated samples compared to the control samples. The real-time PCR assays were performed in triplicate.

GP73 proteins detection

To analyze quantitatively the effects of Stealth RNAi on GP73 protein levels in supernatant of Hep G2 and Bel7402 cells, cells were transfected with Stealth RNAi or negative control in culture flasks. After 24 and 48 h, the supernatant was collected, and GP73 protein levels in supernatant of Hep G2 and Bel7402 cells were detected using a commercially available human GP73 ELISA kit. The ELISAs were performed in triplicate.

Cell proliferation assay and cell cycle analysis

Cell proliferation was measured by MTT assay, according to the manufacturer's instructions. Propidium iodide (PI) staining of the nuclei was used to monitor cell cycle assay. Seventy-five percent cold alcohol was used overnight to fix the cells, and then the cells were resuspended in 300 μ L PBS and stained with 500 μ L PI (250 μ g/mL) for 30 min in the dark. Flow cytometry was used to analyze cells. Cell cycle assays were performed in triplicate.

Apoptosis assessment with flow cytometry and transmission electron microscopy

To analyze quantitatively the effects of Stealth RNAi on apoptosis, cells were transfected with Stealth RNAi or negative control in culture flasks. After 48 h, cells were harvested by trypsinization and rinsed twice with cold PBS. Cells were resuspended in 200 μ L binding buffer and then treated with 10 μ L Annexin V-FITC and 5 μ L PI (Sigma, St Louis, MO, United States) for 15 min. Flow cytometric analysis of cells was performed with an EpicsXL Coulter flow cytometer (Beckman-Coulter, United States). All assays were repeated three times.

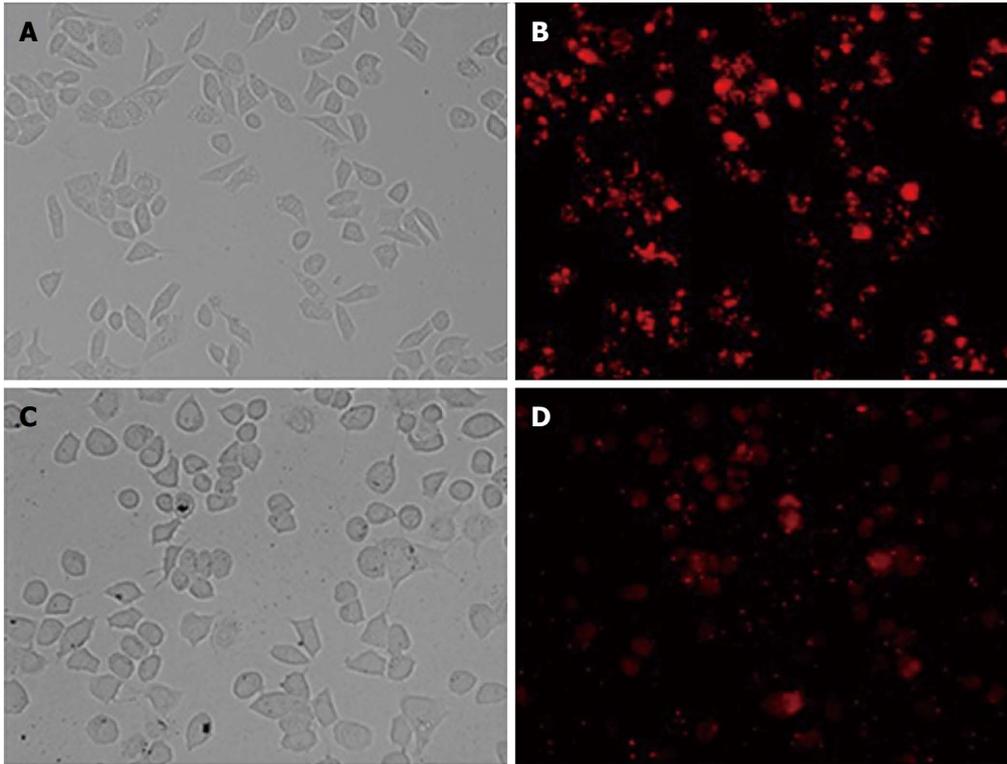


Figure 1 BLOCK-iT Alexa Fluor red fluorescent control. The BLOCK-iT Alexa Fluor red fluorescent control (30 nmol/L) was transfected into Hep G2 and Bel7402 cells using Lipofectamine RNAi MAX Transfection Reagent. Twenty-four hours after transfection, growth medium was removed and replaced with PBS. Hep G2 and Bel7402 cell localization of the Alexa Fluor Red Fluorescent control oligo is seen with fluorescent microscopy (B and D, $\times 200$). Over 80% of the cells took up the control oligo and retained a normal morphology, as seen in bright field (A and C, $\times 200$).

Transmission electron microscopy (TEM) was used to detect apoptosis. Cells were harvested by trypsinization and rinsed twice with cold PBS. Glutaraldehyde (0.3%) was used to prefix the cells overnight, and 10 mL/L osmic acid was used to post-fix the cells. The cells were observed under a JEM-1230 transmission electron microscope (Jeol, Japan).

Protein extraction and western immunoblot analysis

To analyze quantitatively the mechanism of Stealth RNAi on apoptosis, Hep G2 cells were transfected with Stealth RNAi or negative control in culture flasks. After 48 h, cells were harvested by trypsinization and rinsed twice with cold PBS. Hep G2 cells (5×10^5) were lysed by lysis buffer (phenylmethylsulfonyl fluoride), then drawing the protein standard curve to calculate the density of total protein. Ten percent SDS-PAGE was used to separate proteins, and proteins were transferred to nitrocellulose membranes. Anti-GP73, Bax, Bcl-2, cytochrome *c* and procaspase-3, β -actin primary antibody (1:1000; Abcam, Cambridge, MA, United States) were used to incubate membranes, and then, anti-rabbit secondary antibody conjugated with horseradish peroxidase was used to incubate membranes (1:5000) again. Western Blotting Substrate (Bio-Rad) was used to visualize immunoreactive proteins. All assays were repeated three times.

Statistical analysis

Mean \pm SD was used to express the data. The data

among the three groups were compared by one-way analysis of variance followed by Bonferroni correction. Some data were also analyzed by Student's *t* test. SPSS for Windows version 19.0 was used for statistical analysis. $P < 0.01$ was considered significant.

RESULTS

Transfection efficiency

Twenty-four hours after transfection, Bel7402 and Hep G2 cells were observed by fluorescent microscopy. The efficiency of transfection was $> 80\%$ when using 30 nmol/L final concentrations of oligo duplex and 1×10^5 /mL cells (Figure 1). Therefore, 30 nmol/L siRNA and 1×10^5 /mL Bel7402 and Hep G2 cells were used to perform subsequent experiments.

Expression of GP73 after transfection

After Stealth siRNA targeting GP73 was transfected into Hep G2 and Bel7402 cells, mRNA levels were measured by RT-PCR. There was an obvious reduction of GP73 mRNA levels in the Stealth RNAi group. Quantification analysis revealed that GP73 mRNA was reduced by 68.7% and 90.3% of the blank control for Hep G2 cells, and 56.7% and 88.7% of the blank control for Bel7402 cells at 24 and 48 h after transfection, respectively. A significant difference was found between the blank control and Stealth RNAi groups (Figure 2A and B, $P < 0.01$). ELISA of supernatant demonstrated that GP73 protein

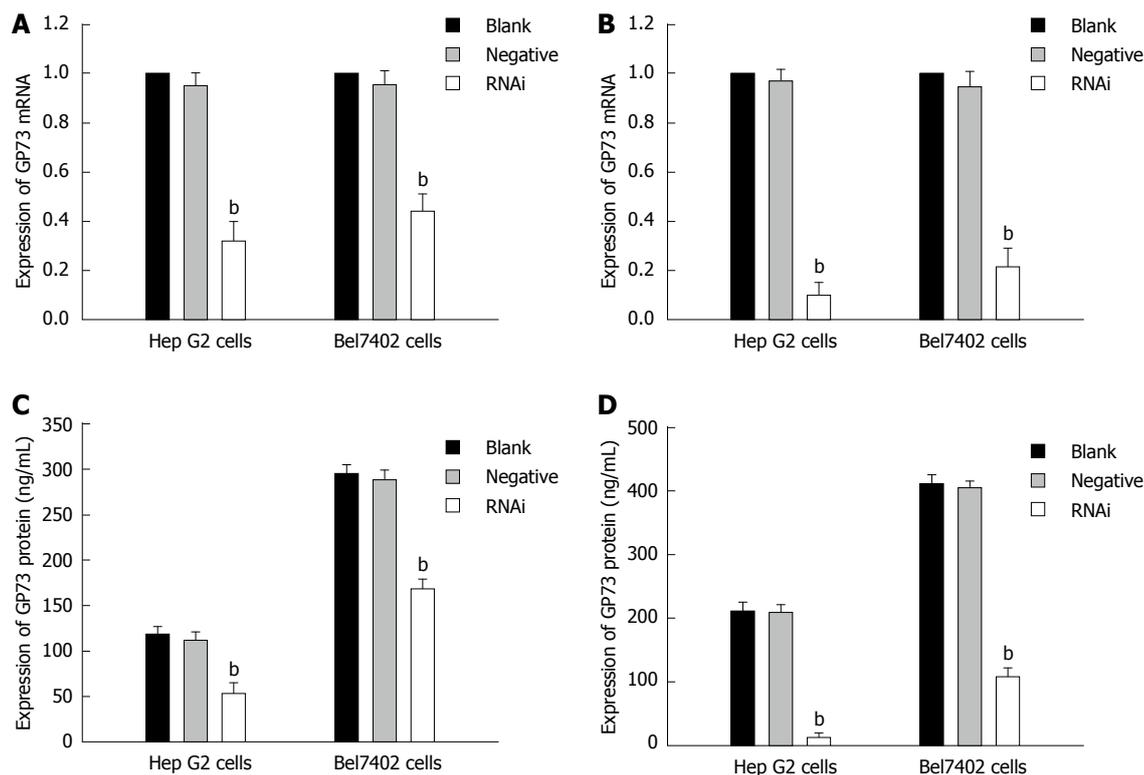


Figure 2 Expression of GP73 mRNA and protein for Hep G2 and Bel7402 cells after transfection with Stealth RNAi against GP73, negative control RNAi or blank control. A: Relative expression of GP73 mRNA level at 24 h after transfection analyzed by quantitative real-time reverse transcription polymerase chain reaction (RT-PCR); B: Relative expression of GP73 mRNA level at 48 h after transfection analyzed by quantitative RT-PCR; C: Expression of GP73 proteins in supernatant of Hep G2 and Bel7402 cells were detected by enzyme-linked immunosorbent assay (ELISA) at 24 and 48 h; D: Expression of GP73 proteins in supernatant of Hep G2 and Bel7402 cells was detected by ELISA at 48 h. The corresponding values of GP73 protein in the three groups at 24 and 48 h after transfection. ^b $P < 0.01$ vs blank group.

levels were also significantly reduced compared with the blank control. GP73 protein levels for blank control, negative control and RNAi groups at 24 and 48 h after transfection are shown in Figure 2C and D ($P < 0.01$). There was a significant difference between the blank control and RNAi groups (Figure 2C and D, $P < 0.01$). Expression of GP73 in Hep G2 cells was also significantly lower than in Bel7402 cells after transfection.

Silencing of GP73 gene decreased viability and proliferation in Bel7402 and Hep G2 cells

As illustrated in Figure 3A, the viability of Hep G2 and Bel7402 cells was reduced significantly after transfection. The inhibition rate in the RNAi group was 27.09% and 50.53% at 24 and 48 h after transfection for Hep G2 cells, respectively, and 21.3% and 46.4% at 24 and 48 h after transfection for Bel7402 cells (Figure 3A, $P < 0.01$). As illustrated in Figure 3B and C, an apoptotic peak was seen in the RNAi group of Hep G2 cells, and cell cycle analysis showed that proliferation of Hep G2 cells yielded 78.22% \pm 0.35% cells in the G₀/G₁ phase, 20.72% \pm 1.19% cells in S phase, and 1.07% \pm 1.09% cells in G₂/M phase 48 h after transfection. The percentage of cells in G₀/G₁ phase increased significantly, however, the percentage of cells in G₂/M phase decreased significantly ($P < 0.01$). Proliferation control Bel7402 cells yielded

88.81% \pm 1.13% cells in G₀/G₁ phase, 12.1% \pm 1.12% cells in S phase, and 0.98% \pm 1.08% cells in G₂/M phase 48 h after transfection in the RNAi group. The percentage of cells in G₀/G₁ phase increased significantly, however, the percentage of cells in G₂/M phase decreased significantly in both cells.

Silencing of GP73 gene induces apoptosis

After transfection for 48 h, the apoptotic cells were markedly increased in the RNAi group compared to those in the negative and blank control groups in Hep G2 and Bel7402 cells. Their values are shown in Figure 4 ($P < 0.01$). The results showed that terminal- and early-stage apoptotic cells were markedly increased in the RNAi group compared to the blank control group in both cells, and terminal-stage apoptosis mainly occurred in Hep G2 cells after transfection, however, early-stage apoptosis mainly occurred in Bel7402 cells.

Changes in ultrastructural morphology of cells

Normal control Hep G2 cells were observed by TEM (Figure 5A). Some typical manifestations of apoptosis such as vacuoles, nuclear fragmentation and apoptotic bodies were observed in the RNAi group of Hep G2 cells (Figure 5B). Normal control Bel7402 cells were observed (Figure 5C). Dense chromatin and some vacuoles could

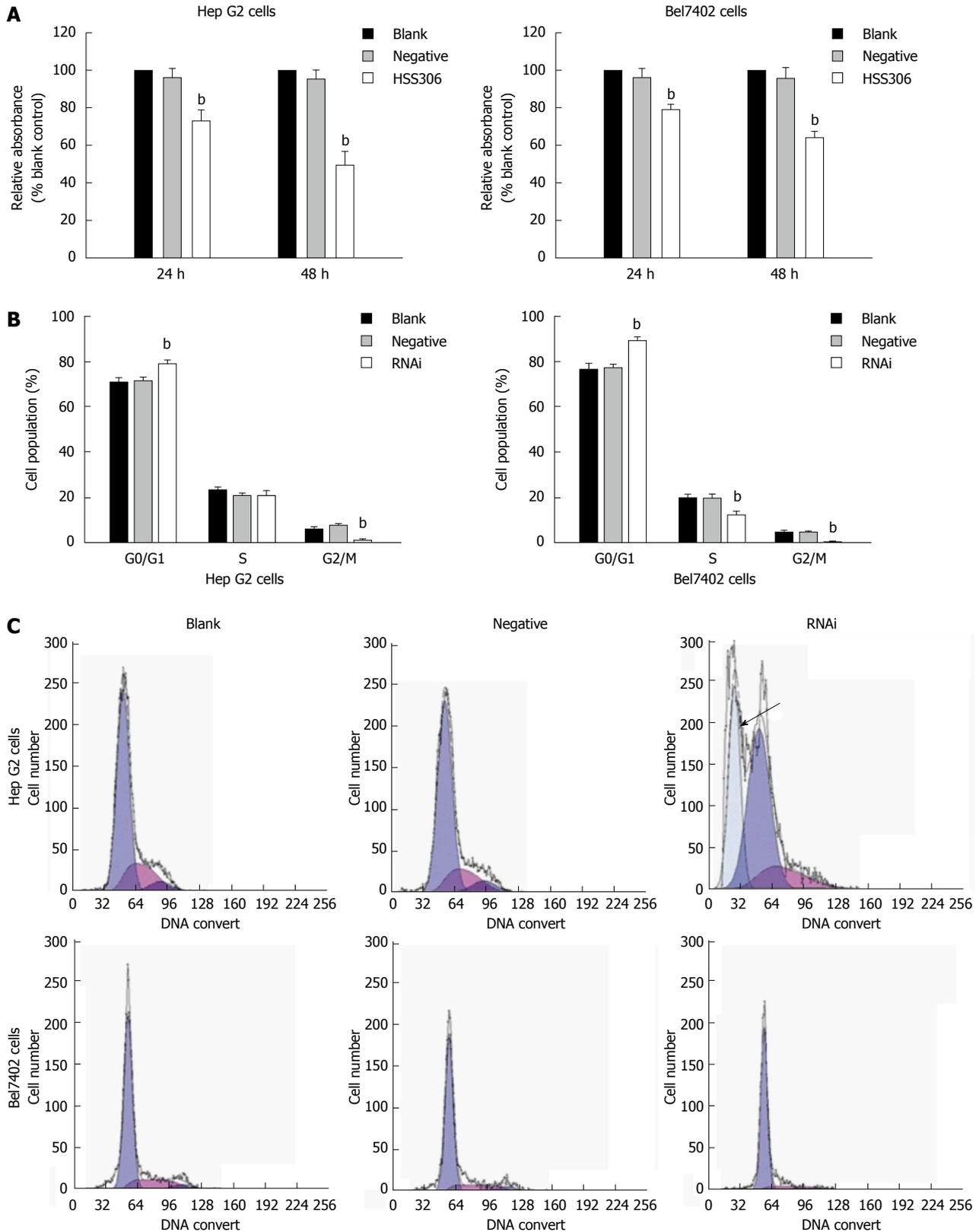


Figure 3 Silencing of *GP73* gene decreased cell proliferation. A: MTT analysis at 24 and 48 h (% blank control) for Hep G2 and Bel7402 cells; B: Cell cycle assay was analyzed by flow cytometry; C: Flow cytometric analysis of the cell cycle distribution of Hep G2 and Bel7402 cells at 24 h after transfection. Cells were washed, fixed, and stained with propidium iodide, and analyzed using Beckman-Coulter Epics flow cytometer. ^b*P* < 0.01 vs blank group.

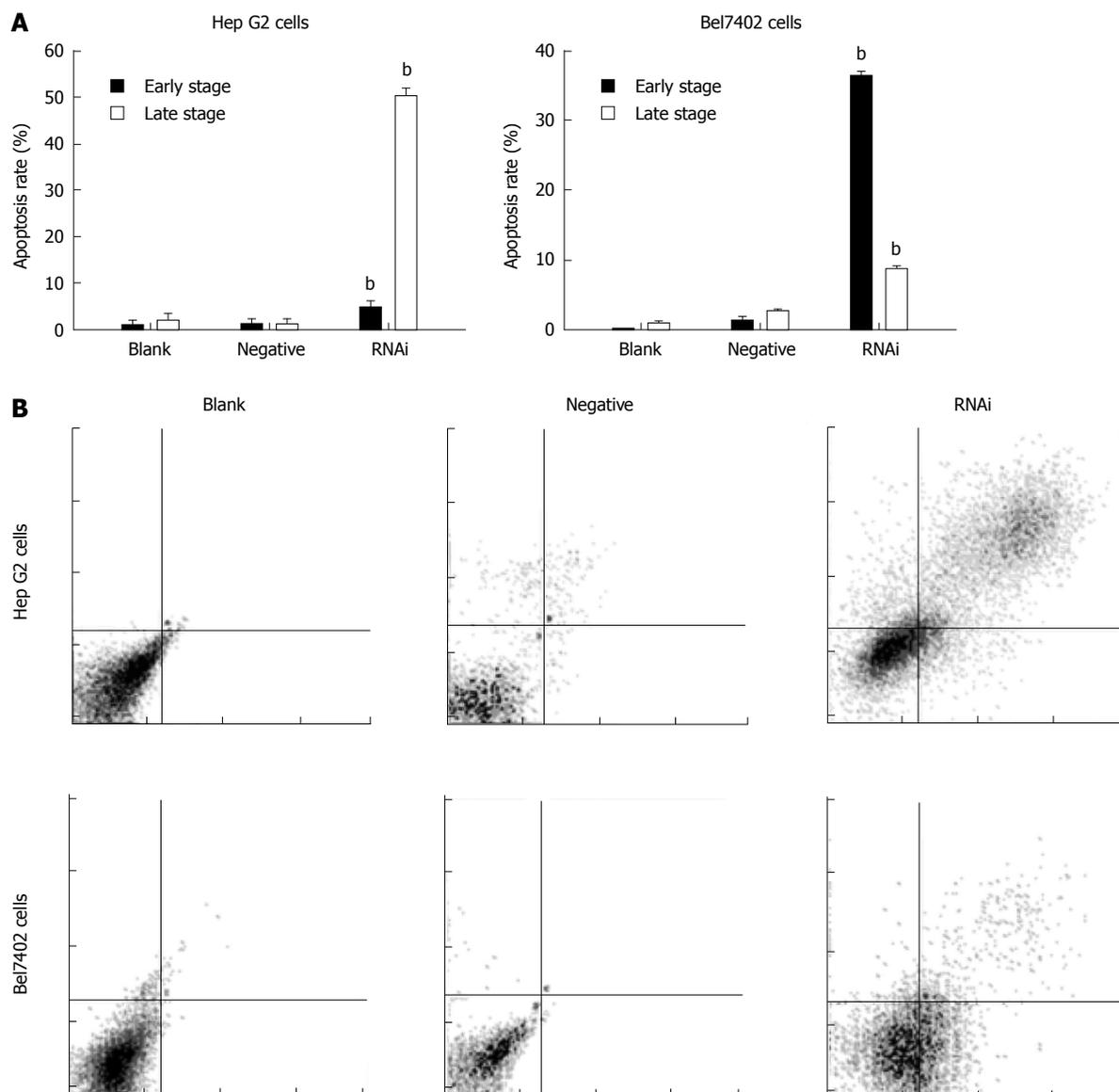


Figure 4 Effect of *GP73*-targeted stealth RNAi transfection on apoptosis of Hep G2 and Bel7402 cells. A: Apoptosis assay was analyzed by flow cytometry; B: Apoptotic effect of Stealth RNAi after 48 h, and later stained with annexin V-FITC and PI. The stained cells were analyzed for apoptosis using a Beckman-Coulter Epics flow cytometer. ^a*P* < 0.01 vs blank group.

be seen in the RNAi group of Bel7402 cells (Figure 5D).

Effects of silencing *GP73* gene on apoptosis-related molecules in Hep G2 cells

Western blotting was performed to analyze the expression of *GP73*, Bax/Bcl-2, cytochrome c and procaspase-3 in Hep G2 cells. The protein levels of *GP73* decreased significantly compared with those in negative and blank control groups at 48 h after transfection in Hep G2 cells. Compared with negative and blank control groups, the protein levels of Bcl-2 were reduced significantly, however, the protein levels of Bax were markedly elevated. Compared to the negative and blank control groups at 48 h after transfection, the protein level of cytochrome c in the cytoplasm was markedly elevated in the RNAi group, and the protein level of procaspase-3 was reduced markedly (Figure 6).

DISCUSSION

There is a growing body of evidence that *GP73* serum levels correlate with the presence of HCC. However, although *GP73* levels in the circulation do not appear to be elevated in healthy subjects, these and other reports suggest elevation of the marker in people with inflammatory liver diseases, in the absence of cancer. Thus, its usefulness in distinguishing cancer from cirrhosis or other liver conditions associated with inflammation is still being evaluated. Work by others has found that *GP73* levels are affected by tumor necrosis factor- α and interferon- γ in tissue culture systems, and there is an association between levels of *GP73* and osmoles in serum from people with liver cirrhosis, but there is no significant correlation in HCC. We do not know the molecular basis for the high expression of *GP73* in these cells, and the effect

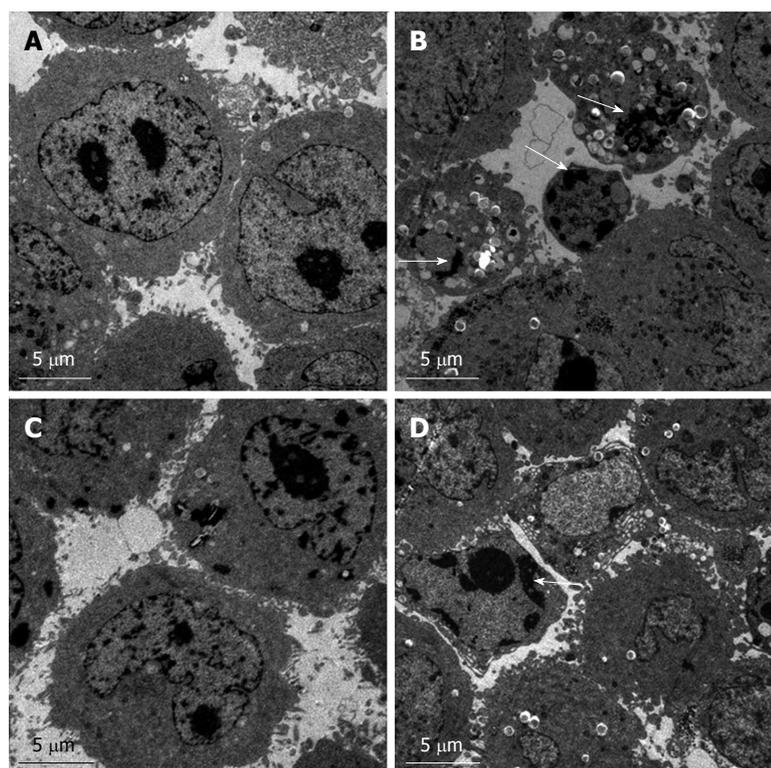


Figure 5 Transmission electron microscopy. The ultrastructural morphology of control (A) and RNAi (B) of Hep G2 cells. The ultrastructural morphology of control (C) and RNAi (D) Bel7402 cells. Nuclear fragmentation and apoptotic bodies are indicated by arrows.

of GP73 in hepatic carcinoma cells is also unclear. We investigated the effect of GP73 in hepatic carcinoma by silencing expression of GP73 through siRNA. In this study, after Stealth RNAi against GP73 was transfected into Hep G2 and Bel7402 cells, the expression of GP73 and mRNA of GP73 decreased significantly in both cells. The decreasing expression of GP73 inhibited cell proliferation and led to apoptosis. Besides, decreasing expression of GP73 also leads to reduction of Bcl-2/Bax ratio, an increase in protein level of cytochrome c in the cytoplasm, and decreasing procaspase-3.

The effects of Stealth RNAi on both GP73 mRNA and protein levels were measured. The results demonstrated that the Stealth RNAi against GP73 (RNAi) can reduce the expression of GP73 effectively at both mRNA and protein levels in Hep G2 and Bel7402 cells after transfection. Western blotting showed that GP73 protein decreased significantly in the RNAi group compared with the other groups. In addition, cell cycle assay confirmed that the proliferation of Bel7402 and Hep G2 cells was reduced significantly after expression of GP73 gene was silenced by RNAi. In Bel7402 and Hep G2 cells, the percentage of cells in G₀/G₁ phase increased significantly, however, the percentage of cells in G₂/M phase decreased significantly. The percentage of cells in S phase decreased in Bel7402 cells after transfection, although the percentage of Hep G2 cells in S phase did not change significantly. However, an apoptotic peak was seen in Hep G2 cells, which indicates that terminal apoptosis had occurred. The proliferation of hepatic cancer

cells is related to the expression of GP73. Decreasing expression of GP73 inhibited hepatic cancer cells in G₀/G₁ phase, and sometimes it also affected hepatic cancer cells in S phase.

Furthermore, fluorescence activated cell sorting methods suggested that Stealth RNAi against GP73 (RNAi) caused accumulation of early-stage apoptotic Bel7402 cells and accumulation of terminal-stage apoptotic Hep G2 cells at 48 h after transfection (GP73 was silenced). The apoptotic peak was also seen in proliferation of Hep G2 cells. The different stage of apoptosis may be related to the different expression of GP73 in Bel7402 and Hep G2 cells. TEM observation agreed with the former results. Dense chromatin appeared near the nuclear membrane, many foaming phenomenon happened in Bel7402 cells, which also indicated that Stealth RNAi against GP73 (RNAi) mainly caused accumulation of early-stage apoptotic Bel7402 cells, however, many vacuoles, nuclear fragmentation and apoptotic bodies appeared in Hep G2 cells, which can also explain why there was an apoptotic peak in the RNAi group. These results also confirmed that Stealth RNAi against GP73 (RNAi) mainly caused accumulation of terminal-stage apoptotic Hep G2 cells. And the difference of its expression in different cells might play different function. Several studies have found that the ratio of Bax/Bcl-2 is important in apoptosis of cancer cells. Bcl-2 is considered as an upstream effector molecule in the apoptotic pathway, but Bax is considered as a downstream effector molecule^[36,37]. The protein levels of Bax and Bcl-2 were measured, and similar results

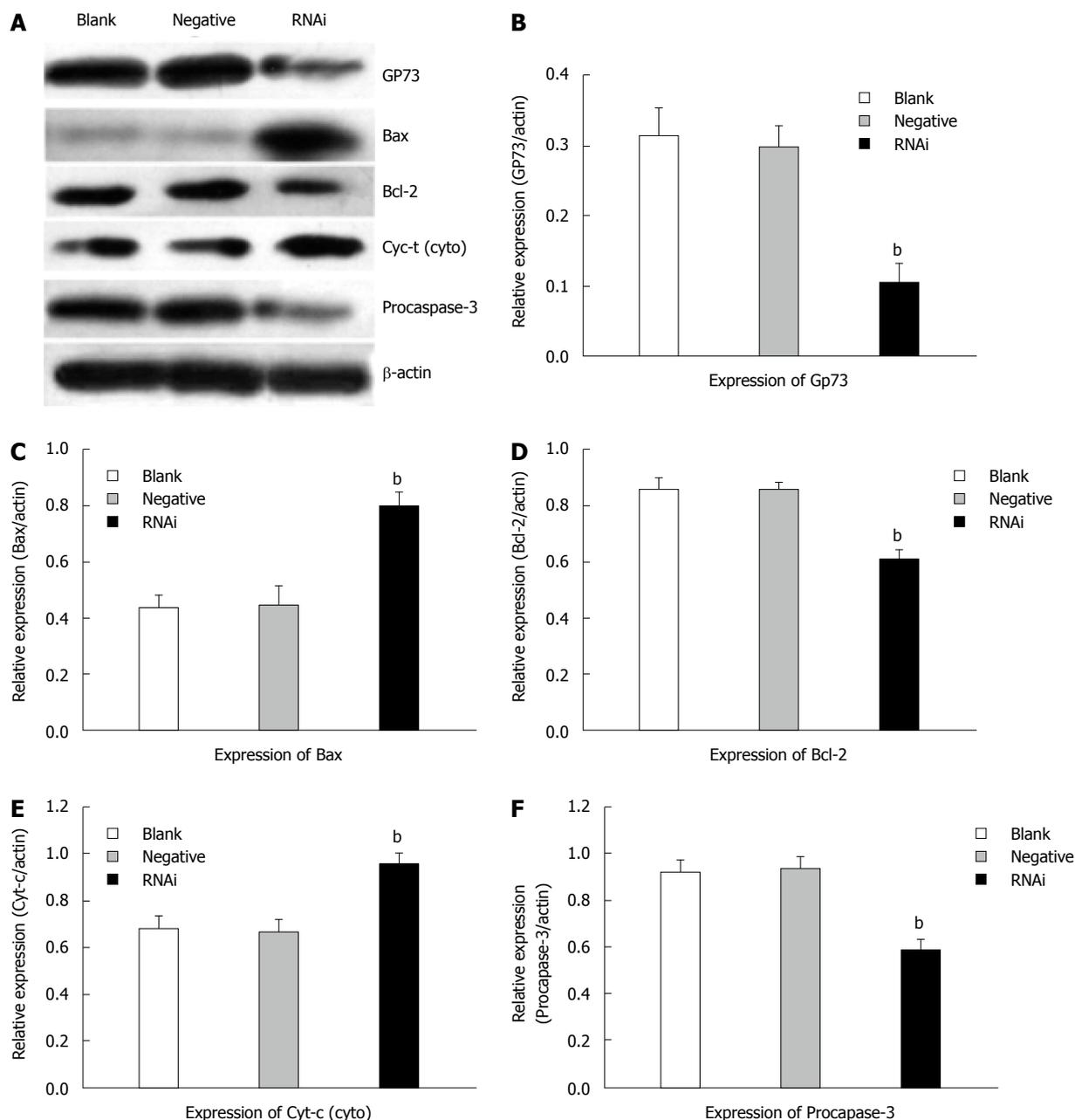


Figure 6 Effects of silencing *GP73* gene on apoptosis-related molecules in Hep G2 cells. A: Western blotting detected expression of GP73, Bcl-2, Bax, cytochrome c and procaspase-3 in Hep G2 cells at 48 h after transfection with Stealth RNAi (HSS181966; Stealth RNAi against GP73, negative control RNAi) or blank control; B-F: Representative immunoblots are shown from three independent experiments. Software Glyko BandsScan 5.0 was used to quantify the bands, and the expression of β -actin was used as a loading control, the protein levels were normalized against β -actin, data were expressed as mean \pm SD (^b*P* < 0.01 vs blank group).

were also found in this study. We demonstrated that the reduction of GP73 led to a decrease in expression of Bcl-2 but an increase in Bax, and the Bcl-2/Bax ratio in Hep G2 cells was reduced, this result was also similar to the other one on the cell apoptotic mechanism^[38]. Besides, in some apoptotic mechanism studies, it was found that Bcl-2 inhibits cytochrome c release from mitochondria, whereas Bax triggers it^[39], and release of cytochrome c activated caspases to induce apoptosis. The expression of cytochrome c and procaspase-3 was also detected by Western blotting to determine their distribution in apoptosis. The level of cytochrome c in the cytoplasm was

markedly elevated in the RNAi group; however, the level of procaspase-3 was reduced significantly. These results proved that GP73 may play a major role in anti-apoptosis in HCC cells, and there is a correlation between it and other apoptosis-related proteins such as Bax, Bcl-2, cytochrome c and procaspase-3.

In conclusion, Stealth RNAi targeting GP73 gene sequence reduced the expression of GP73 markedly. The reduction of GP73 in Hep G2 and Bel7402 cells inhibited cell proliferation and induced apoptosis, however, terminal apoptosis occurred in Hep G2 cells, but early apoptosis occurred in Bel7402 cells. Reduced expression

of *GP73* gene might lead to a reduction in Bcl-2/Bax ratio, an increase in cytochrome c, but a reduction in caspase-3, and the reduction of *GP73* induces apoptosis in Hep G2 cells may through this apoptosis signaling pathway. However, further studies are needed to confirm this conclusion. Consequently, the results imply that *GP73* plays an important role in HCC cells, but unfortunately, expression of *GP73* was not reduced to a low level in this study, therefore, more effective molecular tools are needed to explore the exact function of *GP73* in more HCC cells.

COMMENTS

Background

The physiological and pathological roles of Golgi protein (GP)73 have attracted considerable attention in recent years. However, the function of *GP73* in hepatic carcinoma cells remains obscure.

Research frontiers

This study was performed to explore the function of *GP73* in hepatic carcinoma cells; Stealth RNAi targeting *GP73* gene sequence was used to silence its expression; and cell proliferation and apoptosis were assessed. This study implied that *GP73* plays an important role in hepatocellular carcinoma (HCC) cells.

Innovations and breakthroughs

The results of this study suggested that silencing *GP73* gene in HCC cells inhibits cell proliferation and induces apoptosis.

Applications

The results imply that *GP73* plays an important role in HCC cells, but further research is needed to explore the exact function of *GP73* in more HCC cells.

Peer review

These results are very interesting and may provide important evidence about the mechanism of apoptosis in HCC.

REFERENCES

- 1 Yang JD, Roberts LR. Hepatocellular carcinoma: A global view. *Nat Rev Gastroenterol Hepatol* 2010; **7**: 448-458 [PMID: 20628345 DOI: 10.1038/nrgastro.2010.100]
- 2 Chan AC, Chan SC, Chok KS, Cheung TT, Chiu DW, Poon RT, Fan ST, Lo CM. Treatment strategy for recurrent hepatocellular carcinoma: salvage transplantation, repeated resection, or radiofrequency ablation? *Liver Transpl* 2013; **19**: 411-419 [PMID: 23447460 DOI: 10.1002/lt.23605]
- 3 Shindoh J, Hasegawa K, Inoue Y, Ishizawa T, Nagata R, Aoki T, Sakamoto Y, Sugawara Y, Makuuchi M, Kokudo N. Risk factors of post-operative recurrence and adequate surgical approach to improve long-term outcomes of hepatocellular carcinoma. *HPB (Oxford)* 2013; **15**: 31-39 [PMID: 23216777 DOI: 10.1111/j.1477-2574.2012.00552.x]
- 4 Yau T, Chan P, Epstein R, Poon RT. Management of advanced hepatocellular carcinoma in the era of targeted therapy. *Liver Int* 2009; **29**: 10-17 [PMID: 19120940 DOI: 10.1111/j.1478-3231.2008.01916.x]
- 5 Greten TF, Korangy F, Manns MP, Malek NP. Molecular therapy for the treatment of hepatocellular carcinoma. *Br J Cancer* 2009; **100**: 19-23 [PMID: 19018262 DOI: 10.1038/sj.bjc.6604784]
- 6 Hagymási K, Tulassay Z. [New possibilities of targeted therapy in the treatment of hepatocellular carcinoma with the help of molecular biology]. *Orv Hetil* 2010; **151**: 1763-1768 [PMID: 20940115 DOI: 10.1556/OH.2010.28984]
- 7 Pang RW, Poon RT. From molecular biology to targeted therapies for hepatocellular carcinoma: the future is now. *Oncology* 2007; **72** Suppl 1: 30-44 [PMID: 18087180 DOI: 10.1159/000111705]
- 8 Llovet JM, Bruix J. Novel advancements in the management of hepatocellular carcinoma in 2008. *J Hepatol* 2008; **48** Suppl 1: S20-S37 [PMID: 18304676 DOI: 10.1016/j.jhep.2008.01.022]
- 9 Voiculescu M, Winkler RE, Moscovici M, Neuman MG. Chemotherapies and targeted therapies in advanced hepatocellular carcinoma: from laboratory to clinic. *J Gastrointest Liver Dis* 2008; **17**: 315-322 [PMID: 18836626]
- 10 Kladney RD, Bulla GA, Guo L, Mason AL, Tollefson AE, Simon DJ, Koutoubi Z, Fimmel CJ. *GP73*, a novel Golgi-localized protein upregulated by viral infection. *Gene* 2000; **249**: 53-65 [PMID: 10831838 DOI: 10.1016/S0378-1119(00)00136-0]
- 11 Kladney RD, Tollefson AE, Wold WS, Fimmel CJ. Upregulation of the Golgi protein *GP73* by adenovirus infection requires the E1A CtBP interaction domain. *Virology* 2002; **301**: 236-246 [PMID: 12359426 DOI: 10.1006/viro.2002.1523]
- 12 Block TM, Comunale MA, Lowman M, Steel LF, Romano PR, Fimmel C, Tennant BC, London WT, Evans AA, Blumberg BS, Dwek RA, Mattu TS, Mehta AS. Use of targeted glycoproteomics to identify serum glycoproteins that correlate with liver cancer in woodchucks and humans. *Proc Natl Acad Sci USA* 2005; **102**: 779-784 [PMID: 15642945 DOI: 10.1073/pnas.0408928102]
- 13 Norton PA, Comunale MA, Krakover J, Rodemich L, Pirog N, D'Amelio A, Philip R, Mehta AS, Block TM. N-linked glycosylation of the liver cancer biomarker *GP73*. *J Cell Biochem* 2008; **104**: 136-149 [PMID: 18004786 DOI: 10.1002/jcb.21610]
- 14 Kladney RD, Cui X, Bulla GA, Brunt EM, Fimmel CJ. Expression of *GP73*, a resident Golgi membrane protein, in viral and nonviral liver disease. *Hepatology* 2002; **35**: 1431-1440 [PMID: 12029628 DOI: 10.1053/jhep.2002.32525]
- 15 Malaguarnera G, Giordano M, Paladina I, Berretta M, Cappellani A, Malaguarnera M. Serum markers of hepatocellular carcinoma. *Dig Dis Sci* 2010; **55**: 2744-2755 [PMID: 20339916 DOI: 10.1007/s10620-010-1184-7]
- 16 Shi Y, Chen J, Li L, Sun Z, Zen L, Xu S, Zhang Y, Zhang L. A study of diagnostic value of golgi protein *GP73* and its genetic assay in primary hepatic carcinoma. *Technol Cancer Res Treat* 2011; **10**: 287-294 [PMID: 21517136]
- 17 Hu JS, Wu DW, Liang S, Miao XY. *GP73*, a resident Golgi glycoprotein, is sensibility and specificity for hepatocellular carcinoma of diagnosis in a hepatitis B-endemic Asian population. *Med Oncol* 2010; **27**: 339-345 [PMID: 19399652 DOI: 10.1007/s12032-009-9215-y]
- 18 Puri S, Bachert C, Fimmel CJ, Linstedt AD. Cycling of early Golgi proteins via the cell surface and endosomes upon luminal pH disruption. *Traffic* 2002; **3**: 641-653 [PMID: 12191016]
- 19 Mao Y, Yang H, Xu H, Lu X, Sang X, Du S, Zhao H, Chen W, Xu Y, Chi T, Yang Z, Cai J, Li H, Chen J, Zhong S, Mohanti SR, Lopez-Soler R, Millis JM, Huang J, Zhang H. Golgi protein 73 (GOLPH2) is a valuable serum marker for hepatocellular carcinoma. *Gut* 2010; **59**: 1687-1693 [PMID: 20876776 DOI: 10.1136/gut.2010.214916]
- 20 Marrero JA, Romano PR, Nikolaeva O, Steel L, Mehta A, Fimmel CJ, Comunale MA, D'Amelio A, Lok AS, Block TM. *GP73*, a resident Golgi glycoprotein, is a novel serum marker for hepatocellular carcinoma. *J Hepatol* 2005; **43**: 1007-1012 [PMID: 16137783 DOI: 10.1016/j.jhep.2005.05.028]
- 21 Tian L, Wang Y, Xu D, Gui J, Jia X, Tong H, Wen X, Dong Z, Tian Y. Serological AFP/Golgi protein 73 could be a new diagnostic parameter of hepatic diseases. *Int J Cancer* 2011; **129**: 1923-1931 [PMID: 21140449 DOI: 10.1002/ijc.25838]
- 22 Wang M, Long RE, Comunale MA, Junaidi O, Marrero J, Di Bisceglie AM, Block TM, Mehta AS. Novel fucosylated biomarkers for the early detection of hepatocellular carcinoma. *Cancer Epidemiol Biomarkers Prev* 2009; **18**: 1914-1921 [PMID: 19454616 DOI: 10.1158/1055-9965.EPI-08-0980]
- 23 Hann HW, Wang M, Hafner J, Long RE, Kim SH, Ahn M, Park S, Comunale MA, Block TM, Mehta A. Analysis of *GP73* in patients with HCC as a function of anti-cancer

- treatment. *Cancer Biomark* 2010; **7**: 269-273 [PMID: 21694465]
- 24 **Zhu K**, Dai Z, Zhou J. Biomarkers for hepatocellular carcinoma: progression in early diagnosis, prognosis, and personalized therapy. *Biomark Res* 2013; **1**: 10 [PMID: 24252133 DOI: 10.1186/2050-7771-1-10]
 - 25 **Wang Y**, Yang H, Xu H, Lu X, Sang X, Zhong S, Huang J, Mao Y. Golgi protein 73, not Glypican-3, may be a tumor marker complementary to α -Fetoprotein for hepatocellular carcinoma diagnosis. *J Gastroenterol Hepatol* 2014; **29**: 597-602 [PMID: 24236824 DOI: 10.1111/jgh.12461]
 - 26 **Liu Y**, Zou Z, Zhu B, Hu Z, Zeng P. CXCL10 decreases GP73 expression in hepatoma cells at the early stage of hepatitis C virus (HCV) infection. *Int J Mol Sci* 2013; **14**: 24230-24241 [PMID: 24351813 DOI: 10.3390/ijms141224230]
 - 27 **Ibrahim GH**, Mahmoud MA, Aly NM. Evaluation of circulating Transforming growth factor-beta1, Glypican-3 and Golgi protein-73 mRNAs expression as predictive markers for hepatocellular carcinoma in Egyptian patients. *Mol Biol Rep* 2013; **40**: 7069-7075 [PMID: 24186850 DOI: 10.1007/s11033-013-2829-3]
 - 28 **Hu B**, Tian X, Sun J, Meng X. Evaluation of individual and combined applications of serum biomarkers for diagnosis of hepatocellular carcinoma: a meta-analysis. *Int J Mol Sci* 2013; **14**: 23559-23580 [PMID: 24317431 DOI: 10.3390/ijms141223559]
 - 29 **Chen JG**, Lu WZ, Zhu YR, Zhang YH, Lu JH, Chen TY. [Retrospective observation of dynamic levels of serum Golgi protein 73 in patients prior to the onset of liver cancer]. *Zhonghua Yu Fang Yi Xue Zazhi* 2013; **47**: 731-735 [PMID: 24246080]
 - 30 **Bao YX**, Yang Y, Zhao HR, Mao R, Xiao L, Zhang YF, Aisiker T, Wen H. [Clinical significance and diagnostic value of Golgi-protein 73 in patients with early-stage primary hepatocellular carcinoma]. *Zhonghua Zhong Liu Zazhi* 2013; **35**: 505-508 [PMID: 24257301]
 - 31 **Grishok A**, Tabara H, Mello CC. Genetic requirements for inheritance of RNAi in *C. elegans*. *Science* 2000; **287**: 2494-2497 [PMID: 10741970 DOI: 10.1126/science.287.5462.2494]
 - 32 **Harborth J**, Elbashir SM, Bechert K, Tuschl T, Weber K. Identification of essential genes in cultured mammalian cells using small interfering RNAs. *J Cell Sci* 2001; **114**: 4557-4565 [PMID: 11792820]
 - 33 **Yu JY**, DeRuiter SL, Turner DL. RNA interference by expression of short-interfering RNAs and hairpin RNAs in mammalian cells. *Proc Natl Acad Sci USA* 2002; **99**: 6047-6052 [PMID: 11972060 DOI: 10.1073/pnas.092143499]
 - 34 **Reynolds A**, Leake D, Boese Q, Scaringe S, Marshall WS, Khvorovova A. Rational siRNA design for RNA interference. *Nat Biotechnol* 2004; **22**: 326-330 [PMID: 14758366 DOI: 10.1038/nbt936]
 - 35 **Schwarz DS**, Hutvagner G, Du T, Xu Z, Aronin N, Zamore PD. Asymmetry in the assembly of the RNAi enzyme complex. *Cell* 2003; **115**: 199-208 [PMID: 14567917 DOI: 10.1016/S0092-8674(03)00759-1]
 - 36 **Reed JC**. Bcl-2 and the regulation of programmed cell death. *J Cell Biol* 1994; **124**: 1-6 [PMID: 8294493 DOI: 10.1083/jcb.124.1.1]
 - 37 **Mazel S**, Burtrum D, Petrie HT. Regulation of cell division cycle progression by bcl-2 expression: a potential mechanism for inhibition of programmed cell death. *J Exp Med* 1996; **183**: 2219-2226 [PMID: 8642331 DOI: 10.1084/jem.183.5.2219]
 - 38 **Yasuhara N**, Sahara S, Kamada S, Eguchi Y, Tsujimoto Y. Evidence against a functional site for Bcl-2 downstream of caspase cascade in preventing apoptosis. *Oncogene* 1997; **15**: 1921-1928 [PMID: 9365238 DOI: 10.1038/sj.onc.1201370]
 - 39 **Kuwana T**, Mackey MR, Perkins G, Ellisman MH, Latterich M, Schneider R, Green DR, Newmeyer DD. Bid, Bax, and lipids cooperate to form supramolecular openings in the outer mitochondrial membrane. *Cell* 2002; **111**: 331-342 [PMID: 12419244 DOI: 10.1016/S0092-8674(02)01036-X]

P- Reviewer: Chuma M, Smith RC **S- Editor:** Gou SX
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