

World Journal of *Hepatology*

World J Hepatol 2023 June 27; 15(6): 725-866



REVIEW

- 725 Ductular reaction in non-alcoholic fatty liver disease: When Macbeth is perverted
He YH, Pan JX, Xu LM, Gu T, Chen YW
- 741 Recent advances in pathophysiology, diagnosis and management of hepatorenal syndrome: A review
Kiani C, Zori AG
- 755 Treatment of liver fibrosis: Past, current, and future
Zhang CY, Liu S, Yang M

MINIREVIEWS

- 775 Tumor budding as a potential prognostic marker in determining the behavior of primary liver cancers
Unal B, Celik MY, Gedik EO, Bassorgun CI, Elpek GO
- 786 Role of vascular endothelial growth factor B in nonalcoholic fatty liver disease and its potential value
Li YQ, Xin L, Zhao YC, Li SQ, Li YN

ORIGINAL ARTICLE

Retrospective Study

- 797 Acute pancreatitis in liver transplant hospitalizations: Identifying national trends, clinical outcomes and healthcare burden in the United States
Dahiya DS, Jahagirdar V, Chandan S, Gangwani MK, Merza N, Ali H, Deliwala S, Aziz M, Ramai D, Pinnam BSM, Bapaye J, Cheng CI, Inamdar S, Sharma NR, Al-Haddad M

Observational Study

- 813 Lower alanine aminotransferase levels are associated with increased all-cause and cardiovascular mortality in nonalcoholic fatty liver patients
Zheng JR, Wang ZL, Jiang SZ, Chen HS, Feng B

Randomized Clinical Trial

- 826 Randomized intervention and outpatient follow-up lowers 30-d readmissions for patients with hepatic encephalopathy, decompensated cirrhosis
Pusateri A, Litzenberg K, Griffiths C, Hayes C, Gnyawali B, Manious M, Kelly SG, Conteh LF, Jalil S, Nagaraja HN, Mumtaz K

SYSTEMATIC REVIEWS

- 841 Liver injury from direct oral anticoagulants
Juneja D, Nasa P, Jain R

- 850** Management of sepsis in a cirrhotic patient admitted to the intensive care unit: A systematic literature review

Ndomba N, Soldera J

ABOUT COVER

Editorial Board Member of *World Journal of Hepatology*, Kazuto Tajiri, MD, PhD, Associate Professor, Department of Gastroenterology, Toyama University Hospital, Toyama 930-0194, Japan. tajikazu@med.u-toyama.ac.jp

AIMS AND SCOPE

The primary aim of *World Journal of Hepatology* (*WJH*, *World J Hepatol*) is to provide scholars and readers from various fields of hepatology with a platform to publish high-quality basic and clinical research articles and communicate their research findings online.

WJH mainly publishes articles reporting research results and findings obtained in the field of hepatology and covering a wide range of topics including chronic cholestatic liver diseases, cirrhosis and its complications, clinical alcoholic liver disease, drug induced liver disease autoimmune, fatty liver disease, genetic and pediatric liver diseases, hepatocellular carcinoma, hepatic stellate cells and fibrosis, liver immunology, liver regeneration, hepatic surgery, liver transplantation, biliary tract pathophysiology, non-invasive markers of liver fibrosis, viral hepatitis.

INDEXING/ABSTRACTING

The *WJH* is now abstracted and indexed in PubMed, PubMed Central, Emerging Sources Citation Index (Web of Science), Scopus, Reference Citation Analysis, China National Knowledge Infrastructure, China Science and Technology Journal Database, and Superstar Journals Database. The 2022 edition of Journal Citation Reports® cites the 2021 Journal Citation Indicator (JCI) for *WJH* as 0.52. The *WJH*'s CiteScore for 2021 is 3.6 and Scopus CiteScore rank 2021: Hepatology is 42/70.

RESPONSIBLE EDITORS FOR THIS ISSUE

Production Editor: *Yi-Xuan Cai*, Production Department Director: *Xiang Li*, Editorial Office Director: *Xiang Li*.

NAME OF JOURNAL

World Journal of Hepatology

ISSN

ISSN 1948-5182 (online)

LAUNCH DATE

October 31, 2009

FREQUENCY

Monthly

EDITORS-IN-CHIEF

Nikolaos Pyrsopoulos, Ke-Qin Hu, Koo Jeong Kang

EDITORIAL BOARD MEMBERS

<https://www.wjnet.com/1948-5182/editorialboard.htm>

PUBLICATION DATE

June 27, 2023

COPYRIGHT

© 2023 Baishideng Publishing Group Inc

INSTRUCTIONS TO AUTHORS

<https://www.wjnet.com/bpg/gerinfo/204>

GUIDELINES FOR ETHICS DOCUMENTS

<https://www.wjnet.com/bpg/GerInfo/287>

GUIDELINES FOR NON-NATIVE SPEAKERS OF ENGLISH

<https://www.wjnet.com/bpg/gerinfo/240>

PUBLICATION ETHICS

<https://www.wjnet.com/bpg/GerInfo/288>

PUBLICATION MISCONDUCT

<https://www.wjnet.com/bpg/gerinfo/208>

ARTICLE PROCESSING CHARGE

<https://www.wjnet.com/bpg/gerinfo/242>

STEPS FOR SUBMITTING MANUSCRIPTS

<https://www.wjnet.com/bpg/GerInfo/239>

ONLINE SUBMISSION

<https://www.f6publishing.com>

Role of vascular endothelial growth factor B in nonalcoholic fatty liver disease and its potential value

Yu-Qi Li, Lei Xin, Yu-Chi Zhao, Shang-Qi Li, Ya-Nuo Li

Specialty type: Endocrinology and metabolism

Provenance and peer review:
Invited article; Externally peer reviewed.

Peer-review model: Single blind

Peer-review report's scientific quality classification

Grade A (Excellent): 0
Grade B (Very good): B, B
Grade C (Good): C
Grade D (Fair): 0
Grade E (Poor): 0

P-Reviewer: Fatkhudinov TK, Russia; Mogulkoc R, Turkey; Pham TTT, Viet Nam

Received: March 6, 2023

Peer-review started: March 6, 2023

First decision: April 12, 2023

Revised: April 27, 2023

Accepted: May 9, 2023

Article in press: May 9, 2023

Published online: June 27, 2023



Yu-Qi Li, Ya-Nuo Li, Department of Pathophysiology, School of Basic Medicine, Binzhou Medical University, Yantai 264000, Shandong Province, China

Lei Xin, Department of Gastrointestinal Surgery, Yantaishan Hospital, Yantai 264000, Shandong Province, China

Yu-Chi Zhao, Department of Surgery, Yantaishan Hospital, Yantai 264000, Shandong Province, China

Shang-Qi Li, The First School of Clinical Medicine, Binzhou Medical University, Yantai 264000, Shandong, China, Yantai 264000, Shandong Province, China

Corresponding author: Ya-Nuo Li, PhD, Professor, Department of Pathophysiology, School of Basic Medicine, Binzhou Medical University, No. 346 Guanhai Road, Laishan District, Yantai 264000, Shandong Province, China. liyanuo@bzmc.edu.cn

Abstract

Nonalcoholic fatty liver disease (NAFLD) refers to fatty liver disease caused by liver injury factors other than alcohol. The disease is characterized by diffuse fat infiltration, including simple steatosis (no inflammatory fat deposition), nonalcoholic fatty hepatitis, liver fibrosis, and so on, which may cause liver cirrhosis, liver failure, and even liver cancer in the later stage of disease progression. At present, the pathogenesis of NAFLD is still being studied. The "two-hit" theory, represented by lipid metabolism disorder and inflammatory reactions, is gradually enriched by the "multiple-hit" theory, which includes multiple factors, such as insulin resistance and adipocyte dysfunction. In recent years, vascular endothelial growth factor B (VEGFB) has been reported to have the potential to regulate lipid metabolism and is expected to become a novel target for ameliorating metabolic diseases, such as obesity and type 2 diabetes. This review summarizes the regulatory role of VEGFB in the onset and development of NAFLD and illustrates its underlying molecular mechanism. In conclusion, the signaling pathway mediated by VEGFB in the liver may provide an innovative approach to the diagnosis and treatment of NAFLD.

Key Words: Nonalcoholic fatty liver disease; Vascular endothelial growth factor B; "Two-hit" theory; "Multiple-hit" theory; Obesity

©The Author(s) 2023. Published by Baishideng Publishing Group Inc. All rights reserved.

Core Tip: Nonalcoholic fatty liver disease (NAFLD) is the most common liver disease with lipid accumulation caused by liver injury factors except alcohol. At present, vascular endothelial growth factor B (VEGFB) has been reported to play a special role in regulating lipid metabolism and improving the onset and development of NAFLD. Therefore, the use of VEGFB as a target for treatment has become the focus of current research. This review summarizes the role and potential mechanism of VEGFB in the pathogenesis of NAFLD to provide a theoretical basis for the clinical treatment of NAFLD.

Citation: Li YQ, Xin L, Zhao YC, Li SQ, Li YN. Role of vascular endothelial growth factor B in nonalcoholic fatty liver disease and its potential value. *World J Hepatol* 2023; 15(6): 786-796

URL: <https://www.wjgnet.com/1948-5182/full/v15/i6/786.htm>

DOI: <https://dx.doi.org/10.4254/wjh.v15.i6.786>

INTRODUCTION

With the growth of the economy and the increasing change in people's lifestyles, the prevalence and morbidity of nonalcoholic fatty liver disease (NAFLD) are rising rapidly worldwide. NAFLD occurs in one-fourth of the global population, and the highest incidence rate is in South America (31%) and the Middle East (32%), followed by Asia (27%), the United States (24%), and Europe (23%), while is not common in Africa (14%)[1]. In the United States, the number of NAFLD cases is expected to increase from 83.1 million in 2015 (approximately 24% of the population) to 100.9 million by 2030[2]. Because of its long course and high treatment cost, it has become a global medical and health problem.

NAFLD is an important cause of advanced liver disease, primary liver cancer, and liver transplantation and is also the world's fastest-growing cause of liver-related deaths[3]. In the United States, the burden of NAFLD-related cirrhosis is estimated to be twice that of hepatitis C virus (HCV) related cirrhosis, and it is expected to surpass HCV as the main indication for liver transplantation within 5 years[4]. In Asia, the incidence rate of hepatocellular carcinoma in patients with NAFLD is 1.8/1000 each year, and the total case fatality rate is 5.3/1000 each year[5]. In addition, insulin resistance, upregulation of insulin-like growth factor axis, downregulation of adiponectin expression, and elevated expression of tumor necrosis factor α (TNF α) caused by NAFLD may be potential factors to induce the development of tumors[6]. Meanwhile, NAFLD can also promote coronary atherosclerosis, significantly increase the risk of cardiomyopathy (mainly left ventricular hypertrophy), leading to valvular heart disease (mainly aortic valve and mitral valve), cardiac insufficiency, arrhythmia (mainly atrial fibrillation, prolonged QT interval) and some cardiac conduction system defects (such as an atrioventricular block)[7]. Therefore, more and more research is focusing on exploring the pathogenesis of NAFLD.

The physiological mechanism of NAFLD is very complex. The pathogenesis of early NAFLD is generally believed to be related to lipid metabolism and inflammatory reactions, which could not systematically and comprehensively explain the molecular mechanism and metabolic changes in NAFLD[8,9]. In recent years, studies have confirmed that insulin resistance is closely related to the pathogenesis of NAFLD[10]. In 2019, Lee *et al*[11] reported that NAFLD was related to liver and peripheral insulin resistance, leading to insufficient inhibition of liver insulin resistance, gluconeogenesis, reduced glycogen synthesis, and increased free fatty acid (FFA). Shi *et al*[12] confirmed that insulin resistance can promote the progression of liver fibrosis and NAFLD, and NAFLD can also accelerate insulin resistance in the liver.

With the deepening of research on NAFLD and the increasing understanding of its pathogenesis, it has been found that the onset of NAFLD is also related to "multiple-hit" such as liver insulin resistance, adipocyte dysfunction, gut microbiota imbalance, immune regulation imbalance, and dietary habits besides the "second-hit" caused by lipid metabolism disorder and inflammation reaction. Adolph *et al*[13] found that abnormal adiponectin secretion produced by adipocytes can aggravate high-fat diet (HFD)-induced obesity and related metabolic disorders, and the overexpression of adiponectin can hinder the progression of hepatic microsomal steatosis. Baker *et al*[14] found that the content of enzymes that can metabolize ethanol in the body of patients with NAFLD and intestinal flora imbalance increased significantly, which increased the permeability of the intestinal wall and was conducive to the entry of reactive oxygen species (ROS), bacterial endotoxins, ethanol and other toxic metabolites into the liver, resulting in increased liver damage and accelerating the progression of NAFLD (Figure 1).

Early in 2008, Karpanen *et al*[15] unexpectedly found that vascular endothelial growth factor B (VEGFB) has a weak role in the vascular system but has a significant advantage in regulating lipid metabolism. In 2012, Hagberg *et al*[16] proved that targeting VEGFB as a novel treatment for insulin resistance and type 2 diabetes. In 2016, Robciuc *et al*[17] also found that transferring the VEGFB gene into HFD-induced obese mice can improve lipid metabolism and increase insulin supply and signal transduction. Hu *et al*[18] confirmed that VEGFB recombinant protein can reduce lipid accumulation and improve hyperlipidemia in NAFLD.

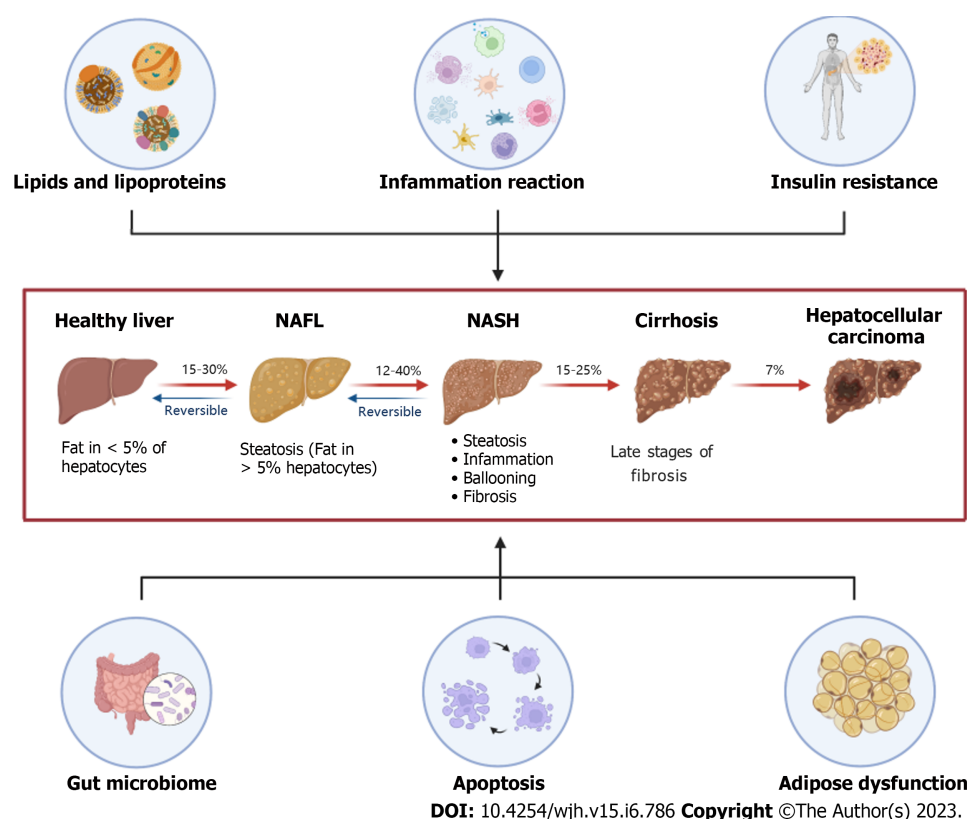


Figure 1 The “multiple-hit” theories are involved in the progress of nonalcoholic fatty liver disease. Lipids and lipoproteins represent the “first-hit”, while the inflammation reaction illustrates the “second-hit” in the development of nonalcoholic fatty liver disease (NAFLD). Six aspects including lipids and lipoproteins, inflammation reaction, insulin resistance, gut microbiome, apoptosis, and adipose dysfunction have a common influence on the pathophysiological mechanism of NAFLD. NAFL: Non-alcoholic fatty liver; NASH: Non-alcoholic steatohepatitis.

The regulatory role of VEGFB in the occurrence and development of metabolic diseases has attracted many scholars' attention. In this review, we mainly focus on the underlying mechanism of VEGFB in the onset of NAFLD and analyze how VEGFB participates in the "multiple-hit" of NAFLD by regulating lipid metabolism, inflammatory reactions, adipocyte dysfunction, and cell apoptosis. First, we introduce the positive regulatory effect of VEGFB on lipid metabolism and discuss how it affects fatty acid oxidation and lipid synthesis under the mediation of Adenosine 5'-monophosphate (AMP)-activated protein kinase (AMPK) signaling. Then, we summarize the role of VEGFB in anti-inflammation in NAFLD and further discuss the current mechanism of VEGFB in insulin resistance and the impact of targeted therapy. Finally, we also explain the controversial role of VEGFB in metabolic diseases and estimate whether VEGFB-mediated signal transduction could provide a theoretical and experimental basis for the pathogenesis of NAFLD and help identify potential treatment targets.

THE NOVEL ROLE OF VEGFB IN NAFLD

VEGFB is a special type of vascular endothelial growth factor. The total length of the VEGFB gene is 1197 bp, with 7 exons, and the total length of the CDS region is 566 bp, with two subtypes, VEGFB¹⁶⁷ and VEGFB¹⁸⁶[19]. The VEGFB¹⁶⁷ homotype has a similar effect to that of VEGFB¹⁸⁶[17]. VEGFB is a glycoprotein that forms a homodimer through the covalent binding of disulfide bonds. It needs to combine with a high-affinity tyrosine kinase receptor to exert biological effects[20]. The VEGF family includes seven members, VEGF-A, VEGF-B, VEGF-C, VEGF-D, VEGF-E, VEGF-F, and placental growth factor[21]. The VEGF receptor family includes VEGFR1, VEGFR2, VEGFR3, and neuropilin 1/2 (NRP1/2). VEGFA can combine with VEGFR1 or VEGFR2 to play a role in promoting angiogenesis[22]. Unlike other members of the VEGF family, the effect of VEGFB on vascular endothelial growth is not obvious. The biological function of VEGFB is exerted by forming a complex with VEGFR1. Moreover, the combination of VEGFB and NRP1 can also induce a series of reactions through a paracrine mechanism [23]. In recent years, studies have shown that the VEGFB/VEGFR1 pathway has therapeutic potential for obesity, type 2 diabetes, and other lipid metabolism disorder-related diseases[17].

VEGFB mainly exists in the heart, skeletal muscle, brown adipose tissue, and other tissues with high metabolic activity and plays a role in regulating blood vessel distribution and lowering blood lipids[24]. The level of VEGFB in the liver is also significantly higher than that in tissues with general metabolic

activity. Shang *et al*[25] found that cardiac-specific overexpression of VEGFB can reduce the activity of lipoprotein lipase and improve the metabolic level of myocardial cells. Wagenmakers *et al*[26] showed that VEGFB can control the expression of fatty acid transport protein (FATP) in the capillary endothelium and connect the uptake of endothelial FFA with the oxidation ability of the skeletal muscle to potentially prevent the accumulation of skeletal muscle lipotoxic FFA. Robciuc *et al*[17] confirmed that the complex of VEGFB and VEGFR1 can reshape the vascular distribution in adipose tissue and improve the insulin function of obese mice.

VEGFB also plays a biological role in the liver by forming a complex with VEGFR1. Cordeiro *et al*[27] showed that targeting VEGFB can effectively prevent lipid deposition in peripheral tissues in animal models. Hu *et al*[18] observed that the complex of VEGFB and VEGFR1 can increase the oxidation level of fatty acids in liver tissue and hepatocytes and reduce obesity-related hyperlipidemia and fatty liver disease in HFD-induced liver. Li *et al*[28] found that inhibiting VEGFB gene expression in liver tissue not only increased the weight and body fat rate of obese mice but also led to pathological changes, such as hepatocyte steatosis and liver fibrosis. These studies suggest that VEGFB is involved in the onset and development of simple steatosis and liver fibrosis in NAFLD (Figure 2).

VEGFB PARTICIPATES IN REGULATING THE "FIRST HIT" IN NAFLD

In 1998, Day *et al*[29] first proposed the "two-hit" theory of the pathogenesis of NAFLD. The "first hit" of NAFLD mainly involves lipid metabolism disorder caused by various factors. Hepatotoxicity is caused by FFA, which leads to an increase in the permeability of the cell membrane, destruction of mitochondrial function, and inhibition of related enzymes to produce genotoxicity. As the disease progresses, excess FFA undergoes β oxidation in mitochondria. When the capacity of the mitochondria to β oxidize FFA is overloaded, excess FFA accumulates in the liver and aggravates the steatosis of hepatocytes. Meanwhile, the triglyceride (TG) synthesized by excess FFA in the liver cannot be converted into very low-density lipoprotein for transport to the peripheral adipose tissue for storage. Therefore, TG can only be stored in the liver and eventually aggravate the onset of liver steatosis. Reducing lipid accumulation and restoring the balance of lipid metabolism are the key methods to improve the "first hit" of NAFLD.

The research findings on the role of VEGFB in improving the lipid disorder of the heart, skeletal muscle, and brown adipose tissue provide the theoretical and experimental basis for VEGFB to participate in the regulation of hepatic lipid metabolism in NAFLD. Shang *et al*[25] observed that rat heart lipoprotein lipase activity and lipid accumulation were decreased and insulin function was improved after cardiac-specific overexpression of VEGFB. Li *et al*[30] found that VEGFB can enhance the expression of FATP1 and FATP4 in C2C12 cells, promote the oxidation of FFA and the decomposition of TG in C2C12 myotubes, and inhibit the re-esterification of FFA to reduce lipid accumulation in myotubes. Chen *et al*[31] found that after inhibition of adipose-specific VEGFB, mice increased in size with more white adipose tissue, and the form and function of fat changed from those of brown adipose tissue to those of white adipose tissue, which indicated that VEGFB was the main regulator of the growth and function of fat.

Some scholars have proposed that the signaling pathway triggered by the combination of VEGFB with its receptor can promote lipid flow in the body, which may become a promising target to prevent the accumulation of ectopic lipids. In 2020, Tong *et al*[32] showed that VEGFB can reduce the levels of TG and FFA in the liver to prevent HFD-induced fatty liver disease by producing *E. coli*-expressed recombinant tPep-VEGFB. In 2021, Hu *et al*[18] found that recombinant VEGFB protein reduced the increase in high-density lipoprotein and low-density lipoprotein in the liver caused by HFD and reduced liver hyperlipidemia.

The mechanism of NAFLD involves multiple signaling pathways, of which the AMPK signaling pathway plays a key role in de novo synthesis and fatty acid oxidation[33]. Harjes *et al*[34] confirmed that the combination of VEGFB with its receptor VEGFR1 can activate AMPK, FATP3, and FATP4 to potentially promote the usage of FFA. AMPK activation is regulated by its upstream molecule Ca^{2+} /Calmodulin-dependent protein kinase β (CaMKK β), which responds to increased intracellular calcium content[35]. Extracellular calcium ions enter the cell through a calcium channel carrier. The elevated intracellular calcium level causes the conformational change of CaMKK β and the phosphorylation of AMPK[36]. Jia *et al*[37] showed that a high concentration of VEGFB recombinant protein can increase the level of calcium ions in MIN6 cells to increase insulin secretion. Li *et al*[28] suggested that inhibiting the expression of VEGFB in the liver can reduce the expression level of CaMKK β and then affect the phosphorylation level of AMPK induced by CaMKK β .

AMPK can control lipid metabolism by regulating its downstream molecules after its phosphorylation[38]. AMPK can directly phosphorylate and inhibit the activation of acetyl coenzyme A carboxylase (ACC), the rate-limiting enzyme that inhibits the synthesis of fatty acids, thereby activating carnitine palmitate transferase (CPT1) and transferring fatty acids into mitochondria for β oxidation[39, 40]. AMPK can also negatively regulate the expression of sterol-regulatory element-binding protein-1c (SREBP1c), downregulate the level of desaturase [stearoyl-CoA desaturase-1 (SCD1)], and inhibit the

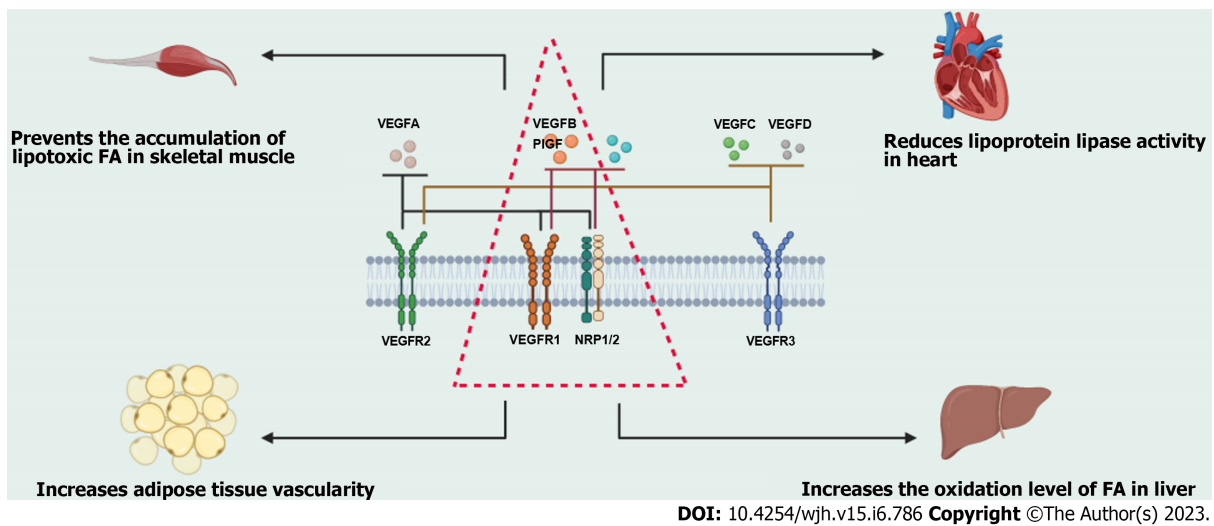


Figure 2 The vascular endothelial growth factor family and its receptors, and the biological function of vascular endothelial growth factor

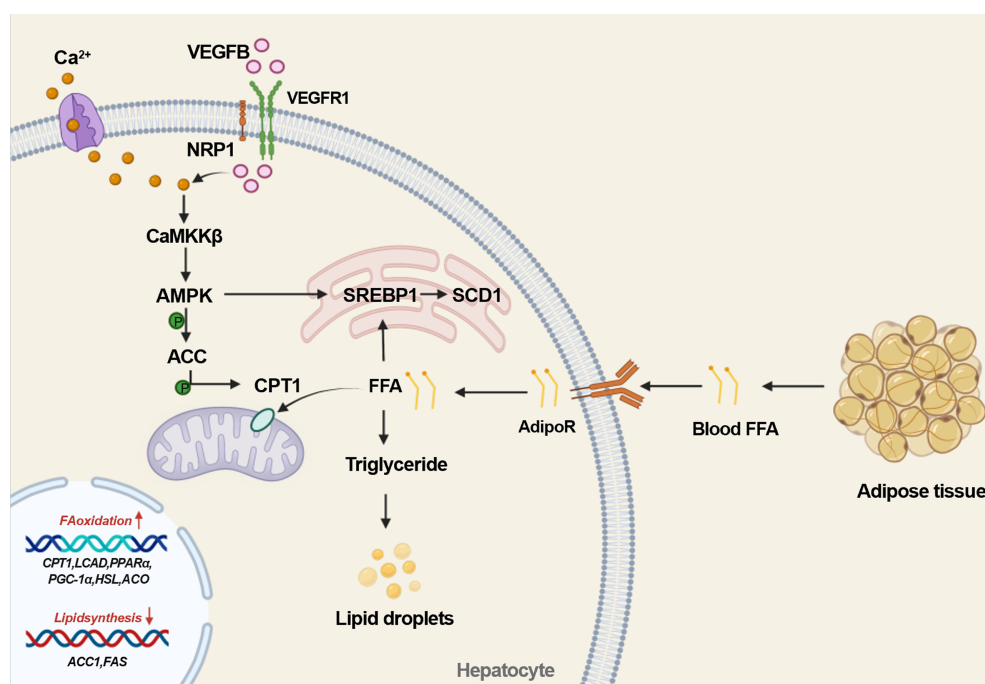
B. The vascular endothelial growth factor (VEGF) family includes VEGFA, vascular endothelial growth factor B (VEGFB), VEGFC, VEGFD, and so on. The VEGF receptor family includes VEGFR1, VEGFR2, VEGFR3, and neuropilin 1/2 (NRP1/2). VEGFA combines with VEGFR1, VEGFR2, or NRP1/2. VEGFB and placental growth factor combine with VEGFR1 or NRP1/2. VEGFC and VEGFD combine with VEGFR2 or VEGFR3 to exert their biological functions. VEGFB can prevent the accumulation of lipotoxic free fatty acid (FFA) in skeletal muscle, reduce lipoprotein activity in the heart, increase adipose tissue vascularity, and increase the oxidation level of FFA in the liver. PIGF: Placental growth factor; FA: Fatty acid; NRP1/2: Neuropilin 1/2; VEGF: Vascular endothelial growth factor; VEGFB: Vascular endothelial growth factor B.

synthesis of fatty acids and TG[41]. Hu *et al*[18] showed that VEGFB recombinant protein can upregulate the AMPK/ACC/CPT1 signaling pathway in the liver by binding to VEGFR1, promoting FFA oxidation and reducing lipid deposition. That study also found that VEGFB can simultaneously upregulate the expression levels of the lipid oxidation-related genes PPAR α , PGC-1 α , HSL, ACO, and CPT1 and that the downregulation of lipid synthesis can inhibit weight gain under HFD conditions and improve obesity-related hyperlipidemia and fatty liver disease[18]. Li *et al*[28] also found that VEGFB knockout can downregulate the CaMKK β -mediated AMPK/ACC/CPT1 signaling pathway to inhibit fatty acid oxidation and activate the AMPK/SREBP1/SCD1 signaling pathway to promote lipid synthesis, thus affecting the level of lipid metabolism (Figure 3).

VEGFB PARTICIPATES IN REGULATING THE "SECOND HIT" IN NAFLD

Liver lipid accumulation induces overloaded lipid catabolism, causing lipid peroxidation. Excessive lipid peroxidation leads to oxidative stress, making it the "second hit" to the progression of NAFLD, which can accelerate inflammation and hepatocyte damage. Nuclear factor-kappa B (NF- κ B) signaling plays an important role in the macrophage-mediated liver inflammatory response[42]. Research has confirmed that NF- κ B can be activated by FFA in patients with NAFLD, and as the severity of NAFLD increases, the activity of NF- κ B increases[43]. Moreover, liver mitochondrial dysfunction also accelerates the occurrence and development of NAFLD. The compensated acceleration of β oxidation in mitochondria can produce a large number of ROS[44]. When the antioxidant system mainly composed of reduced glutathione fails to eliminate ROS in time, oxidative stress develops[45], and a large number of peroxides are generated, which aggravate hepatocyte damage[46]. The apoptotic bodies produced by hepatocyte apoptosis are engulfed by Kupffer cells to decrease the activity of endothelial nitric oxide synthase (eNOS), which affects Mitochondrial function.

VEGFB can induce cell proliferation and differentiation, tumor immunity, and other biological effects through the signaling pathway mediated by the tyrosine-protein kinase receptor[47]. Kusuvara *et al*[48] observed that the VEGFR1 signal in monocytes and macrophages was significantly affected by the upregulation of VEGFB under inflammatory conditions. Akiyoshi U confirmed that VEGFR1 can regulate AKT signaling and affect the activity of NF- κ B and eNOS respectively to regulate macrophage migration and mitochondrial function[49]. Mehlem *et al*[50] found that VEGFB signaling is involved in regulating pathological lipid accumulation in diabetes, obesity, and cardiovascular disease and mainly affects mitochondrial genes related to the regulation of fatty acid intake. Cao *et al*[51] showed that VEGFB/IL-17 inhibits the expression of fatty acid transporters to reduce the accumulation of renal lipids and inhibit renal oxidative stress and mitochondrial dysfunction, thus improving the inflammatory response. Shen *et al*[52] also found that VEGFB/IL-22 can not only regulate glucose and lipid metabolism but also reduce inflammation and ROS accumulation. Robciuc *et al*[17] transduced the



DOI: 10.4254/wjh.v15.i6.786 Copyright ©The Author(s) 2023.

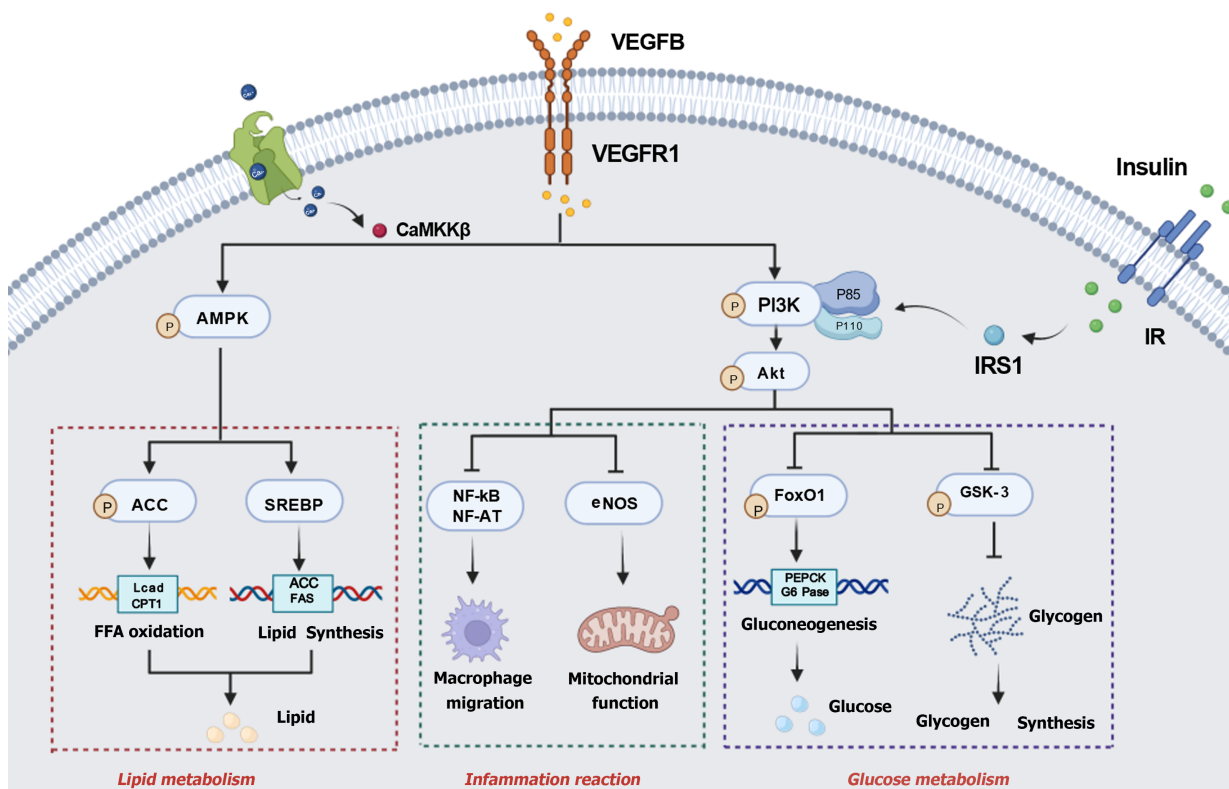
Figure 3 Vascular endothelial growth factor B regulates lipid metabolism in the “first hit” of nonalcoholic fatty liver disease via the activated protein kinase signaling pathway. Vascular endothelial growth factor B (VEGFB) performs its biological function by combining with VEGFR1. Once it enters the cell, it activates Calmodulin-dependent protein kinase β (CaMKK β), which is induced by an increase in intracellular Ca^{2+} content. VEGFB activates the CaMKK β -mediated activated protein kinase (AMPK)/A carboxylase (ACC)/carnitine palmitate transferase (CPT1) signaling pathway and related genes, such as CPT1 and long-chain acyl coenzyme A dehydrogenase, which regulate FFA oxidation in mitochondria. VEGFB activates AMPK/SREBP1/SCD1 and related genes, such as ACC1 and FAS, to inhibit lipid synthesis in the endoplasmic reticulum. CaMKK β : Calmodulin-dependent protein kinase β ; AMPK: Adenosine 5'-monophosphate (AMP)-activated protein kinase; ACC: A carboxylase; CPT1: Carnitine palmitate transferase-1; FFA: Free fatty acid; NRP1/2: Neuropilin 1/2; SCD1: Stearoyl-CoA desaturase-1; SREBP1: Sterol-regulatory element-binding protein-1; VEGFB: Vascular endothelial growth factor B; VEGF: Vascular endothelial growth factor; VEGFB: Vascular endothelial growth factor B; FAS: fatty acid synthase; ACO: Acyl Coenzyme A Oxidase; HSL: hormone-sensitive lipase; LCAD: long-chain acyl-CoA dehydrogenase; PPAR α : proliferator-activated receptor- α ; PGC-1 α : peroxisome proliferator activator receptor γ coactivator 1 α .

VEGFB gene into mice to inhibit obesity-related inflammation and improve metabolic health.

The lipid deposition caused by the "first hit" can lead to an inflammatory cascade, causing the "second hit" to hepatocyte damage and accelerating pathological changes in NAFLD. VEGFB can affect inflammatory response by regulating lipid metabolism in NAFLD, thereby affecting the "first hit" and improving the "second hit" in NAFLD.

VEGFB PARTICIPATES IN REGULATING THE "MULTIPLE-HIT" IN NAFLD

"Multiple-hit" theory believes that the pathological mechanism of NAFLD involves insulin resistance, adipocyte dysfunction, gut microbiota disorder, aggregation of inflammatory factors, mitochondrial dysfunction, lipotoxicity, endoplasmic reticulum stress, and so on[53]. These factors collaborate and overlap with each other, accelerating hepatocyte damage and ultimately developing into cirrhosis, liver cancer, and end-stage liver failure. Insulin resistance is a common metabolic abnormality in patients with NAFLD and is considered the first step in the development of NAFLD[54]. Studies have shown that the activation of insulin receptor substrate 1 (IRS1) protein is downregulated and SREBP-1c is upregulated when insulin resistance occurs, which ultimately increases the expression of de novo synthesis of lipids, thus increasing the transport of FFA to the liver[55]. Meanwhile, hyperinsulinemia can inhibit the β -oxidative of FFA to further promote lipid accumulation in the liver. Excessive lipid accumulation in the liver can disrupt the homeostasis of glucose metabolism[56]. Hepatic insulin resistance participates in the inhibition of forkhead box protein 1 (FOXO1) and serine/threonine kinase (GSK-3) through phosphoinositide 3-kinase (PI3K)/protein kinase B (AKT), reduces Phosphoenolpyruvate carboxykinase (PEPCK) and glucose 6 phosphatase (G6Pase) levels in the liver, which promotes gluconeogenesis and inhibits glycogen synthesis[57] (Figure 4). Recent studies have shown that VEGFB plays an active role in regulating metabolic diseases related to insulin resistance. Robciuc *et al*[17] found that VEGFB can increase the sensitivity of peripheral insulin and improve insulin resistance. Hu *et al*[18] found that VEGFB can reduce insulin resistance by reducing the content of FFA and total cholesterol, thus improving the disorder of lipid metabolism in NAFLD.



DOI: 10.4254/wjh.v15.i6.786 Copyright ©The Author(s) 2023.

Figure 4 Vascular endothelial growth factor B participates in the “multiple-hit” of nonalcoholic fatty liver disease. Vascular endothelial growth factor B (VEGFB) regulates lipid metabolism, inflammation reaction, and glucose metabolism, which co-exist in the nonalcoholic fatty liver disease progression. VEGFB activates the AMPK phosphorylation to regulate free fatty acid oxidation and lipid synthesis. Long-term lipid metabolism disorders will cause inflammatory reactions and glucose metabolism disorders. VEGFB promotes the phosphorylation of protein kinase B (AKT) via combining with the VEGFR1 to affect macrophage migration and mitochondrial inflammation reaction. Meanwhile, VEGFB/VEGFR1 also plays an important role in inhibiting gluconeogenesis and promoting glycogen synthesis by activating the phosphorylation of AKT to regulate glucose metabolism. CaMKKβ: Calmodulin-dependent protein kinase β; VEGFR1: Vascular endothelial growth factor receptor-1; VEGFB: Vascular endothelial growth factor B; IR: Insulin receptor; IRS1: Insulin receptor substrate-1; PI3K: Phosphoinositide 3-kinase; AKT: Protein kinase B; AMPK: Adenosine 5'-monophosphate (AMP)-activated protein kinase; ACC: A carboxylase; CPT1: Carnitine palmitate transferase-1; Lcad: long-chain acyl-CoA dehydrogenase; SREBP: Sterol-regulatory element-binding protein; FAS: fatty acid synthase; NF-κB: Nuclear factor-kappa B; NF-AT: Nuclear factors of activated T; eNOS: endothelial nitric oxide synthase; FoxO1: Forkhead box protein-1; GSK-3: Serine/threonine kinase-3; PEPCK: Phosphoenolpyruvate carboxykinase; G6Pase: Glucose 6 phosphatase;

Insulin resistance has been proven to be an activator of cell apoptosis[58]. Cell apoptosis, an injury factor in the "multiple hits", is a common and important mechanism of NAFLD lesions and liver injury. Li *et al*[59] believe that the decrease in the number of hepatocytes may be due to apoptosis, and excessive apoptosis of hepatocytes is an important sign of NAFLD/NASH patients. As a member of the vascular growth factor family, VEGFB participates in many angiogenesis-dependent diseases, and its pathogenesis is related to cell apoptosis. Williams *et al*[60] showed that inhibition of VEGFB leads to increased apoptosis in cardiomyocytes of patients with diabetes. Dai *et al*[61] also demonstrated that VEGFB plays an antiapoptotic role in the context of tumors.

Adipocyte dysfunction has also been confirmed to be closely related to the pathogenesis of NAFLD [62]. The degree of adipocyte dysfunction is consistent with abnormal metabolism in NAFLD. Martina Rudnicki's study confirms that male mice fed with HFD exhibit adipocyte dysfunction[63]. Robciuc *et al* [17] has shown that the VEGFB/VEGFR1 pathway can be used to enhance vascular distribution in adipose tissue, which improves metabolic health and obesity. The VEGFB gene may affect the occurrence and development of NAFLD by affecting the expansion and loss of adipose tissue.

CONCLUSION

The role of VEGFB in regulating metabolic diseases, such as NAFLD, has attracted increasing attention from scholars. Research has shown that VEGFB can reduce lipid accumulation and restore insulin sensitivity in NAFLD. VEGFB activates the AKT signaling pathway by combining with VEGFR1, inhibits FOXO1 and GSK3 genes, blocks gluconeogenesis, accelerates glycogen synthesis, and improves insulin resistance. VEGFB not only improves liver insulin resistance, but also activates the AMPK signaling pathway, thereby activating the ACC signal to inhibit the expression of SREBP protein,

improving fatty acid oxidation, inhibiting lipid synthesis, and restoring lipid metabolism balance. The activated AKT protein inhibits nuclear factors and proteins such as NF- κ B or eNOS after phosphorylation, regulates inflammatory factors such as macrophages and liver mitochondrial function, reduces the occurrence of inflammatory reactions in hepatocytes, and prevents the progression of NAFLD[43].

Although more and more studies support that VEGFB can be a new target for the treatment of NAFLD and type 2 diabetes, some studies have shown that VEGFB has not played a positive role in regulating lipid metabolism and insulin resistance. Ning *et al*[64] confirmed that the changes in VEGFB did not affect glucose metabolism or lipid uptake. Hagberg *et al*[65] suggested that VEGFB gene deletion can prevent ectopic lipid deposition and ameliorate dyslipidemia. Falkevall *et al*[66] showed that inhibition of VEGFB signaling can target liver steatosis by inhibiting lipolysis and preventing the development of NAFLD.

At present, the understanding of the role of VEGFB in regulating NAFLD and its mechanism remains controversial and is not completely clear. So more research focuses on the mechanism of VEGFB in the occurrence and development of NAFLD, which will provide a new idea for the study of pathological mechanisms and therapeutic targets of NAFLD.

FOOTNOTES

Author contributions: Li YQ prepared and drafted the manuscript; Xin L made the critical revision; Zhao YC edited the manuscript; Li SQ collected the literature review; Li YN approved the final version.

Conflict-of-interest statement: All the authors have declared no conflicts of interest.

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <https://creativecommons.org/licenses/by-nc/4.0/>

Country/Territory of origin: China

ORCID number: Yu-Qi Li 0000-0003-0236-8581; Lei Xin 0000-0003-0084-6498; Yu-Chi Zhao 0000-0002-4776-1419; Shang-Qi Li 0009-0000-1702-9406; Ya-Nuo Li 0000-0002-6441-5024.

S-Editor: Liu JH

L-Editor: A

P-Editor: Cai YX

REFERENCES

- 1 Younossi ZM, Koenig AB, Abdelatif D, Fazel Y, Henry L, Wymer M. Global epidemiology of nonalcoholic fatty liver disease-Meta-analytic assessment of prevalence, incidence, and outcomes. *Hepatology* 2016; **64**: 73-84 [PMID: 26707365 DOI: 10.1002/hep.28431]
- 2 Zhou JH, Cai JJ, She ZG, Li HL. Noninvasive evaluation of nonalcoholic fatty liver disease: Current evidence and practice. *World J Gastroenterol* 2019; **25**: 1307-1326 [PMID: 30918425 DOI: 10.3748/wjg.v25.i11.1307]
- 3 Murphy SK, Yang H, Moylan CA, Pang H, Dellinger A, Abdelmalek MF, Garrett ME, Ashley-Koch A, Suzuki A, Tillmann HL, Hauser MA, Diehl AM. Relationship between methylome and transcriptome in patients with nonalcoholic fatty liver disease. *Gastroenterology* 2013; **145**: 1076-1087 [PMID: 23916847 DOI: 10.1053/j.gastro.2013.07.047]
- 4 Chhatwal J, Samur S, Kues B, Ayer T, Roberts MS, Kanwal F, Hur C, Donnell DM, Chung RT. Optimal timing of hepatitis C treatment for patients on the liver transplant waiting list. *Hepatology* 2017; **65**: 777-788 [PMID: 27906468 DOI: 10.1002/hep.28926]
- 5 Fan JG, Kim SU, Wong VW. New trends on obesity and NAFLD in Asia. *J Hepatol* 2017; **67**: 862-873 [PMID: 28642059 DOI: 10.1016/j.jhep.2017.06.003]
- 6 Meigs JB. Epidemiology of type 2 diabetes and cardiovascular disease: translation from population to prevention: the Kelly West award lecture 2009. *Diabetes Care* 2010; **33**: 1865-1871 [PMID: 20668155 DOI: 10.2337/dc10-0641]
- 7 Pais R, Barritt AS 4th, Calmus Y, Scatton O, Runge T, Lebray P, Poynard T, Ratzu V, Conti F. NAFLD and liver transplantation: Current burden and expected challenges. *J Hepatol* 2016; **65**: 1245-1257 [PMID: 27486010 DOI: 10.1016/j.jhep.2016.07.033]
- 8 Fang YL, Chen H, Wang CL, Liang L. Pathogenesis of non-alcoholic fatty liver disease in children and adolescence: From "two hit theory" to "multiple hit model". *World J Gastroenterol* 2018; **24**: 2974-2983 [PMID: 30038464 DOI: 10.3748/wjg.v24.i27.2974]
- 9 Peverill W, Powell LW, Skoien R. Evolving concepts in the pathogenesis of NASH: beyond steatosis and inflammation. *Int J Mol Sci* 2014; **15**: 8591-8638 [PMID: 24830559 DOI: 10.3390/ijms15058591]

- 10 **Samuel VT**, Shulman GI. Nonalcoholic Fatty Liver Disease as a Nexus of Metabolic and Hepatic Diseases. *Cell Metab* 2018; **27**: 22-41 [PMID: [28867301](#) DOI: [10.1016/j.cmet.2017.08.002](#)]
- 11 **Lee YH**, Cho Y, Lee BW, Park CY, Lee DH, Cha BS, Rhee EJ. Nonalcoholic Fatty Liver Disease in Diabetes. Part I: Epidemiology and Diagnosis. *Diabetes Metab J* 2019; **43**: 31-45 [PMID: [30793550](#) DOI: [10.4093/dmj.2019.0011](#)]
- 12 **Shi H**, Jin J, Hardesty JE, Falkner KC, Prough RA, Balamurugan AN, Mokshagundam SP, Chari ST, Cave MC. Corrigendum to "Polychlorinated biphenyl exposures differentially regulate hepatic metabolism and pancreatic function: Implications for nonalcoholic steatohepatitis and diabetes" [Toxicology & Applied Pharmacology, 363 (2018) 22-33]. *Toxicol Appl Pharmacol* 2020; **404**: 115149 [PMID: [32702359](#) DOI: [10.1016/j.taap.2020.115149](#)]
- 13 **Adolph TE**, Grander C, Grabherr F, Tilg H. Adipokines and Non-Alcoholic Fatty Liver Disease: Multiple Interactions. *Int J Mol Sci* 2017; **18** [PMID: [28758929](#) DOI: [10.3390/ijms18081649](#)]
- 14 **Baker SS**, Baker RD, Liu W, Nowak NJ, Zhu L. Role of alcohol metabolism in non-alcoholic steatohepatitis. *PLoS One* 2010; **5**: e9570 [PMID: [20221393](#) DOI: [10.1371/journal.pone.0009570](#)]
- 15 **Karpanen T**, Bry M, Ollila HM, Seppänen-Laakso T, Liimatta E, Leskinen H, Kivelä R, Helkamaa T, Merentie M, Jeltsch M, Paavonen K, Andersson LC, Mervaala E, Hassinen IE, Ylä-Herttuala S, Oresic M, Alitalo K. Overexpression of vascular endothelial growth factor-B in mouse heart alters cardiac lipid metabolism and induces myocardial hypertrophy. *Circ Res* 2008; **103**: 1018-1026 [PMID: [18757827](#) DOI: [10.1161/CIRCRESAHA.108.178459](#)]
- 16 **Hagberg CE**, Mehlem A, Falkevall A, Muhl L, Fam BC, Ortsäter H, Scotney P, Nyqvist D, Samén E, Lu L, Stone-Elander S, Proietto J, Andrikopoulos S, Sjöholm A, Nash A, Eriksson U. Targeting VEGF-B as a novel treatment for insulin resistance and type 2 diabetes. *Nature* 2012; **490**: 426-430 [PMID: [23023133](#) DOI: [10.1038/nature11464](#)]
- 17 **Robciuc MR**, Kivelä R, Williams IM, de Boer JF, van Dijk TH, Elamaa H, Tigistu-Sahle F, Molotov D, Leppänen VM, Käkälä R, Eklund L, Wasserman DH, Groen AK, Alitalo K. VEGFB/VEGFR1-Induced Expansion of Adipose Vasculature Counteracts Obesity and Related Metabolic Complications. *Cell Metab* 2016; **23**: 712-724 [PMID: [27076080](#) DOI: [10.1016/j.cmet.2016.03.004](#)]
- 18 **Hu L**, Shan Z, Wang F, Gao X, Tong Y. Vascular endothelial growth factor B exerts lipid-lowering effect by activating AMPK via VEGFR1. *Life Sci* 2021; **276**: 119401 [PMID: [33785341](#) DOI: [10.1016/j.lfs.2021.119401](#)]
- 19 **Makinen T**, Olofsson B, Karpanen T, Hellman U, Soker S, Klagsbrun M, Eriksson U, Alitalo K. Differential binding of vascular endothelial growth factor B splice and proteolytic isoforms to neuropilin-1. *J Biol Chem* 1999; **274**: 21217-21222 [PMID: [10409677](#) DOI: [10.1074/jbc.274.30.21217](#)]
- 20 **Lund AW**, Medler TR, Leachman SA, Coussens LM. Lymphatic Vessels, Inflammation, and Immunity in Skin Cancer. *Cancer Discov* 2016; **6**: 22-35 [PMID: [26552413](#) DOI: [10.1158/2159-8290.CD-15-0023](#)]
- 21 **Kivelä R**, Hemanthakumar KA, Vaparanta K, Robciuc M, Izumiya Y, Kidoya H, Takakura N, Peng X, Sawyer DB, Elenius K, Walsh K, Alitalo K. Endothelial Cells Regulate Physiological Cardiomyocyte Growth via VEGFR2-Mediated Paracrine Signaling. *Circulation* 2019; **139**: 2570-2584 [PMID: [30922063](#) DOI: [10.1161/CIRCULATIONAHA.118.036099](#)]
- 22 **Wan A**, Rodrigues B. Endothelial cell-cardiomyocyte crosstalk in diabetic cardiomyopathy. *Cardiovasc Res* 2016; **111**: 172-183 [PMID: [27288009](#) DOI: [10.1093/cvr/cvv159](#)]
- 23 **Wang H**, Yang Y, Yang M, Li X, Tan J, Wu Y, Zhang Y, Li Y, Hu B, Deng S, Yang F, Gao S, Li H, Yang Z, Chen H, Cai W. Pigment Epithelial-Derived Factor Deficiency Accelerates Atherosclerosis Development via Promoting Endothelial Fatty Acid Uptake in Mice With Hyperlipidemia. *J Am Heart Assoc* 2019; **8**: e013028 [PMID: [31711388](#) DOI: [10.1161/JAHA.119.013028](#)]
- 24 **Li X**, Kumar A, Zhang F, Lee C, Tang Z. Complicated life, complicated VEGF-B. *Trends Mol Med* 2012; **18**: 119-127 [PMID: [22178229](#) DOI: [10.1016/j.molmed.2011.11.006](#)]
- 25 **Shang R**, Lal N, Lee CS, Zhai Y, Puri K, Seira O, Boushel RC, Sultan I, Räsänen M, Alitalo K, Hussein B, Rodrigues B. Cardiac-specific VEGFB overexpression reduces lipoprotein lipase activity and improves insulin action in rat heart. *Am J Physiol Endocrinol Metab* 2021; **321**: E753-E765 [PMID: [34747201](#) DOI: [10.1152/ajpendo.00219.2021](#)]
- 26 **Wagenmakers AJ**, Strauss JA, Shepherd SO, Keske MA, Cocks M. Increased muscle blood supply and transendothelial nutrient and insulin transport induced by food intake and exercise: effect of obesity and ageing. *J Physiol* 2016; **594**: 2207-2222 [PMID: [25627798](#) DOI: [10.1113/jphysiol.2014.284513](#)]
- 27 **Cordeiro A**, Costa R, Andrade N, Silva C, Canabrava N, Pena MJ, Rodrigues I, Andrade S, Ramalho A. Does adipose tissue inflammation drive the development of non-alcoholic fatty liver disease in obesity? *Clin Res Hepatol Gastroenterol* 2020; **44**: 394-402 [PMID: [32044284](#) DOI: [10.1016/j.clinre.2019.10.001](#)]
- 28 **Li R**, Li Y, Yang X, Hu Y, Yu H. Reducing VEGFB accelerates NAFLD and insulin resistance in mice via inhibiting AMPK signaling pathway. *J Transl Med* 2022; **20**: 341 [PMID: [35907871](#) DOI: [10.1186/s12967-022-03540-2](#)]
- 29 **Day CP**, James OF. Steatohepatitis: a tale of two "hits"? *Gastroenterology* 1998; **114**: 842-845 [PMID: [9547102](#) DOI: [10.1016/s0016-5085\(98\)70599-2](#)]
- 30 **Li LJ**, Ma J, Li SB, Chen X, Zhang J. Vascular endothelial growth factor B inhibits lipid accumulation in C2C12 myotubes incubated with fatty acids. *Growth Factors* 2019; **37**: 76-84 [PMID: [31215273](#) DOI: [10.1080/08977194.2019.1626851](#)]
- 31 **Chen Y**, Zhao M, Wang C, Wen H, Zhang Y, Lu M, Adlat S, Zheng T, Zhang M, Li D, Lu X, Guo M, Chen H, Zhang L, Feng X, Zheng Y. Adipose vascular endothelial growth factor B is a major regulator of energy metabolism. *J Endocrinol* 2020; **244**: 511-521 [PMID: [31910156](#) DOI: [10.1530/JOE-19-0341](#)]
- 32 **Tong Y**, Zhang Y, Shan Z, Xu Y, Gao X, Yao W. Improving high-fat diet-induced obesity and fatty liver by adipose tissue targeted delivery of vascular endothelial growth factor-B. *Life Sci* 2020; **253**: 117677 [PMID: [32305525](#) DOI: [10.1016/j.lfs.2020.117677](#)]
- 33 **Zhao P**, Sun X, Chagga C, Liao Z, In Wong K, He F, Singh S, Loomba R, Karin M, Witztum JL, Saltiel AR. An AMPK-caspase-6 axis controls liver damage in nonalcoholic steatohepatitis. *Science* 2020; **367**: 652-660 [PMID: [32029622](#) DOI: [10.1126/science.aay0542](#)]
- 34 **Harjes U**, Bensaad K, Harris AL. Endothelial cell metabolism and implications for cancer therapy. *Br J Cancer* 2012; **107**: 1207-1212 [PMID: [23047591](#) DOI: [10.1038/bjc.2012.398](#)]

- 35 **Eichner LJ**, Brun SN, Herzig S, Young NP, Curtis SD, Shackelford DB, Shokhirev MN, Leblanc M, Vera LI, Hutchins A, Ross DS, Shaw RJ, Svensson RU. Genetic Analysis Reveals AMPK Is Required to Support Tumor Growth in Murine Kras-Dependent Lung Cancer Models. *Cell Metab* 2019; **29**: 285-302.e7 [PMID: [30415923](#) DOI: [10.1016/j.cmet.2018.10.005](#)]
- 36 **Jin L**, Chun J, Pan C, Kumar A, Zhang G, Ha Y, Li D, Alesi GN, Kang Y, Zhou L, Yu WM, Magliocca KR, Khuri FR, Qu CK, Metallo C, Owonikoko TK, Kang S. The PLAG1-GDH1 Axis Promotes Anoikis Resistance and Tumor Metastasis through CamKK2-AMPK Signaling in LKB1-Deficient Lung Cancer. *Mol Cell* 2018; **69**: 87-99.e7 [PMID: [29249655](#) DOI: [10.1016/j.molcel.2017.11.025](#)]
- 37 **Jia JD**, Jiang WG, Luo X, Li RR, Zhao YC, Tian G, Li YN. Vascular endothelial growth factor B inhibits insulin secretion in MIN6 cells and reduces Ca(2+) and cyclic adenosine monophosphate levels through PI3K/AKT pathway. *World J Diabetes* 2021; **12**: 480-498 [PMID: [33889292](#) DOI: [10.4239/wjdv12.i4.480](#)]
- 38 **Contreras C**, González-García I, Martínez-Sánchez N, Seoane-Collazo P, Jacas J, Morgan DA, Serra D, Gallego R, Gonzalez F, Casals N, Nogueiras R, Rahmouni K, Diéguez C, López M. Central ceramide-induced hypothalamic lipotoxicity and ER stress regulate energy balance. *Cell Rep* 2014; **9**: 366-377 [PMID: [25284795](#) DOI: [10.1016/j.celrep.2014.08.057](#)]
- 39 **Shimokawa I**, Trindade LS. Dietary restriction and aging in rodents: a current view on its molecular mechanisms. *Aging Dis* 2010; **1**: 89-107 [PMID: [22396859](#)]
- 40 **Herzig S**, Shaw RJ. AMPK: guardian of metabolism and mitochondrial homeostasis. *Nat Rev Mol Cell Biol* 2018; **19**: 121-135 [PMID: [28974774](#) DOI: [10.1038/nrm.2017.95](#)]
- 41 **Gouw AM**, Margulis K, Liu NS, Raman SJ, Mancuso A, Toal GG, Tong L, Mosley A, Hsieh AL, Sullivan DK, Stine ZE, Altman BJ, Schulze A, Dang CV, Zare RN, Felsher DW. The MYC Oncogene Cooperates with Sterol-Regulated Element-Binding Protein to Regulate Lipogenesis Essential for Neoplastic Growth. *Cell Metab* 2019; **30**: 556-572.e5 [PMID: [31447321](#) DOI: [10.1016/j.cmet.2019.07.012](#)]
- 42 **Chalmers SA**, Garcia SJ, Reynolds JA, Herlitz L, Putterman C. NF-κB signaling in myeloid cells mediates the pathogenesis of immune-mediated nephritis. *J Autoimmun* 2019; **98**: 33-43 [PMID: [30612857](#) DOI: [10.1016/j.jaut.2018.11.004](#)]
- 43 **Aqbi HF**, Wallace M, Sappal S, Payne KK, Manjili MH. IFN-γ orchestrates tumor elimination, tumor dormancy, tumor escape, and progression. *J Leukoc Biol* 2018 [PMID: [29469956](#) DOI: [10.1002/JLB.5MIR0917-351R](#)]
- 44 **Carroll EC**, Jin L, Mori A, Muñoz-Wolf N, Oleszycka E, Moran HBT, Mansouri S, McEntee CP, Lambe E, Agger EM, Andersen P, Cunningham C, Hertzog P, Fitzgerald KA, Bowie AG, Lavelle EC. The Vaccine Adjuvant Chitosan Promotes Cellular Immunity via DNA Sensor cGAS-STING-Dependent Induction of Type I Interferons. *Immunity* 2016; **44**: 597-608 [PMID: [26944200](#) DOI: [10.1016/j.immuni.2016.02.004](#)]
- 45 **Gatti M**, Zavatti M, Beretti F, Giuliani D, Vandini E, Ottani A, Bertucci E, Maraldi T. Oxidative Stress in Alzheimer's Disease: In Vitro Therapeutic Effect of Amniotic Fluid Stem Cells Extracellular Vesicles. *Oxid Med Cell Longev* 2020; **2020**: 2785343 [PMID: [33193997](#) DOI: [10.1155/2020/2785343](#)]
- 46 **Vanwong N**, Srisawasdi P, Ngamsamut N, Nuntamool N, Puangpetch A, Chamkrachangpada B, Hongkaew Y, Limsila P, Kittitharaphan W, Sukasem C. Hyperuricemia in Children and Adolescents with Autism Spectrum Disorder Treated with Risperidone: The Risk Factors for Metabolic Adverse Effects. *Front Pharmacol* 2016; **7**: 527 [PMID: [28105014](#) DOI: [10.3389/fphar.2016.00527](#)]
- 47 **Ortega HH**, Veiga-Lopez A, Sreedharan S, del Luján Velázquez MM, Salvetti NR, Padmanabhan V. Developmental Programming: Does Prenatal Steroid Excess Disrupt the Ovarian VEGF System in Sheep? *Biol Reprod* 2015; **93**: 58 [PMID: [26178718](#) DOI: [10.1095/biolreprod.115.131607](#)]
- 48 **Kusuhara S**, Fukushima Y, Ogura S, Inoue N, Uemura A. Pathophysiology of Diabetic Retinopathy: The Old and the New. *Diabetes Metab J* 2018; **42**: 364-376 [PMID: [30362302](#) DOI: [10.4093/dmj.2018.0182](#)]
- 49 **Uemura A**, Fruttiger M, D'Amore PA, De Falco S, Jousen AM, Sennlaub F, Brunck LR, Johnson KT, Lambrou GN, Rittenhouse KD, Langmann T. VEGFR1 signaling in retinal angiogenesis and microinflammation. *Prog Retin Eye Res* 2021; **84**: 100954 [PMID: [33640465](#) DOI: [10.1016/j.preteyeres.2021.100954](#)]
- 50 **Mehlem A**, Palombo I, Wang X, Hagberg CE, Eriksson U, Falkevall A. PGC-1α Coordinates Mitochondrial Respiratory Capacity and Muscular Fatty Acid Uptake via Regulation of VEGF-B. *Diabetes* 2016; **65**: 861-873 [PMID: [26822083](#) DOI: [10.2337/db15-1231](#)]
- 51 **Cao Z**, Zhao H, Fan J, Shen Y, Han L, Jing G, Zeng X, Jin X, Zhu Z, Bian Q, Nan Y, Hu X, Mei X, Ju D, Yang P. Simultaneous blockade of VEGF-B and IL-17A ameliorated diabetic kidney disease by reducing ectopic lipid deposition and alleviating inflammation response. *Cell Death Discov* 2023; **9**: 8 [PMID: [36646672](#) DOI: [10.1038/s41420-023-01304-5](#)]
- 52 **Shen Y**, Chen W, Han L, Bian Q, Fan J, Cao Z, Jin X, Ding T, Xian Z, Guo Z, Zhang W, Ju D, Mei X. VEGF-B antibody and interleukin-22 fusion protein ameliorates diabetic nephropathy through inhibiting lipid accumulation and inflammatory responses. *Acta Pharm Sin B* 2021; **11**: 127-142 [PMID: [33532185](#) DOI: [10.1016/j.apsb.2020.07.002](#)]
- 53 **Tilg H**, Moschen AR. Evolution of inflammation in nonalcoholic fatty liver disease: the multiple parallel hits hypothesis. *Hepatology* 2010; **52**: 1836-1846 [PMID: [21038418](#) DOI: [10.1002/hep.24001](#)]
- 54 **Xu Y**, Zalzal M, Xu J, Li Y, Yin L, Zhang Y. A metabolic stress-inducible miR-34a-HNF4α pathway regulates lipid and lipoprotein metabolism. *Nat Commun* 2015; **6**: 7466 [PMID: [26100857](#) DOI: [10.1038/ncomms8466](#)]
- 55 **Tanaka Y**, Shimanaka Y, Caddeo A, Kubo T, Mao Y, Kubota T, Kubota N, Yamauchi T, Mancina RM, Baselli G, Luukkonen P, Pihlajamäki J, Yki-Järvinen H, Valenti L, Arai H, Romeo S, Kono N. LPIAT1/MBOAT7 depletion increases triglyceride synthesis fueled by high phosphatidylinositol turnover. *Gut* 2021; **70**: 180-193 [PMID: [32253259](#) DOI: [10.1136/gutjnl-2020-320646](#)]
- 56 **Gong Y**, Liu J, Xue Y, Zhuang Z, Qian S, Zhou W, Li X, Qian J, Ding G, Sun Z. Non-monotonic dose-response effects of arsenic on glucose metabolism. *Toxicol Appl Pharmacol* 2019; **377**: 114605 [PMID: [31170414](#) DOI: [10.1016/j.taap.2019.114605](#)]
- 57 **Welty FK**, Alfaddagh A, Elajami TK. Targeting inflammation in metabolic syndrome. *Transl Res* 2016; **167**: 257-280

- [PMID: 26207884 DOI: 10.1016/j.trsl.2015.06.017]
- 58 **Yang S**, Wu M, Li X, Zhao R, Zhao Y, Liu L, Wang S. Role of Endoplasmic Reticulum Stress in Atherosclerosis and Its Potential as a Therapeutic Target. *Oxid Med Cell Longev* 2020; **2020**: 9270107 [PMID: 32963706 DOI: 10.1155/2020/9270107]
 - 59 **Li X**, Wang J, Gong X, Zhang M, Kang S, Shu B, Wei Z, Huang ZS, Li D. Upregulation of BCL-2 by acridone derivative through gene promoter i-motif for alleviating liver damage of NAFLD/NASH. *Nucleic Acids Res* 2020; **48**: 8255-8268 [PMID: 32710621 DOI: 10.1093/nar/gkaa615]
 - 60 **Williams LJ**, Nye BG, Wende AR. Diabetes-Related Cardiac Dysfunction. *Endocrinol Metab (Seoul)* 2017; **32**: 171-179 [PMID: 28685508 DOI: 10.3803/EnM.2017.32.2.171]
 - 61 **Dai W**, Zeller C, Masrour N, Siddiqui N, Paul J, Brown R. Promoter CpG island methylation of genes in key cancer pathways associates with clinical outcome in high-grade serous ovarian cancer. *Clin Cancer Res* 2013; **19**: 5788-5797 [PMID: 23965899 DOI: 10.1158/1078-0432.CCR-13-1217]
 - 62 **Kelley CE**, Brown AJ, Diehl AM, Setji TL. Review of nonalcoholic fatty liver disease in women with polycystic ovary syndrome. *World J Gastroenterol* 2014; **20**: 14172-14184 [PMID: 25339805 DOI: 10.3748/wjg.v20.i39.14172]
 - 63 **Rudnicki M**, Abdifarkosh G, Rezvan O, Nwadozi E, Roudier E, Haas TL. Female Mice Have Higher Angiogenesis in Perigonadal Adipose Tissue Than Males in Response to High-Fat Diet. *Front Physiol* 2018; **9**: 1452 [PMID: 30405427 DOI: 10.3389/fphys.2018.01452]
 - 64 **Ning FC**, Jensen N, Mi J, Lindström W, Balan M, Muhl L, Eriksson U, Nilsson I, Nyqvist D. VEGF-B ablation in pancreatic β -cells upregulates insulin expression without affecting glucose homeostasis or islet lipid uptake. *Sci Rep* 2020; **10**: 923 [PMID: 31969592 DOI: 10.1038/s41598-020-57599-2]
 - 65 **Hagberg CE**, Falkevall A, Wang X, Larsson E, Huusko J, Nilsson I, van Meeteren LA, Samén E, Lu L, Vanwildemeersch M, Klar J, Genove G, Pietras K, Stone-Elander S, Claesson-Welsh L, Ylä-Herttuala S, Lindahl P, Eriksson U. Vascular endothelial growth factor B controls endothelial fatty acid uptake. *Nature* 2010; **464**: 917-921 [PMID: 20228789 DOI: 10.1038/nature08945]
 - 66 **Falkevall A**, Mehlem A, Folestad E, Ning FC, Osorio-Conles Ó, Radmann R, de Hollanda A, Wright SD, Scotney P, Nash A, Eriksson U. Inhibition of VEGF-B signaling prevents non-alcoholic fatty liver disease development by targeting lipolysis in the white adipose tissue. *J Hepatol* 2023; **78**: 901-913 [PMID: 36717026 DOI: 10.1016/j.jhep.2023.01.014]



Published by **Baishideng Publishing Group Inc**
7041 Koll Center Parkway, Suite 160, Pleasanton, CA 94566, USA

Telephone: +1-925-3991568

E-mail: bpgoffice@wjgnet.com

Help Desk: <https://www.f6publishing.com/helpdesk>

<https://www.wjgnet.com>

